

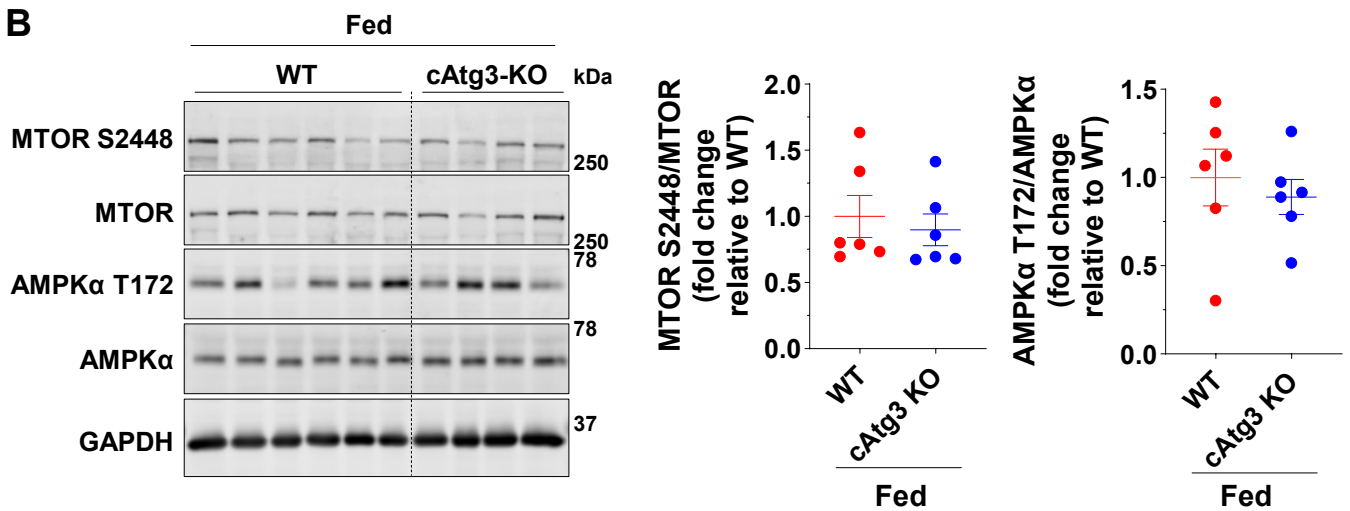
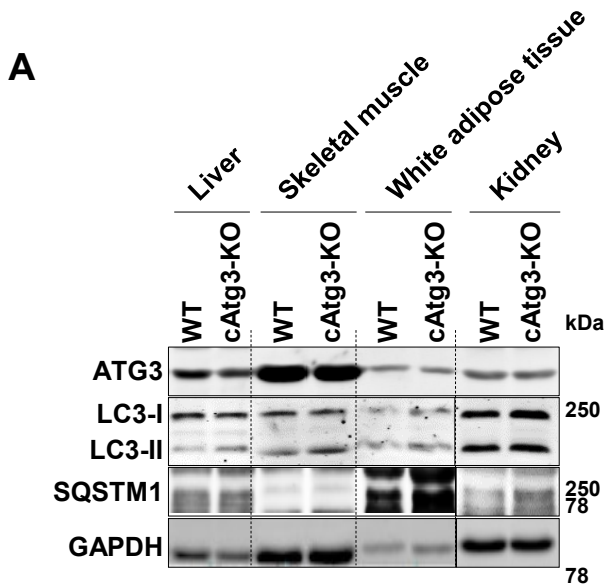
APPENDIX

Control of NAD⁺ homeostasis by autophagic flux modulates cardiac function

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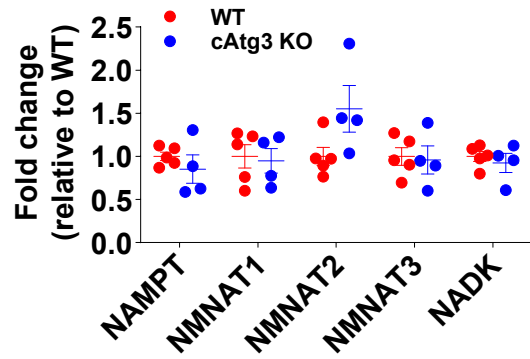
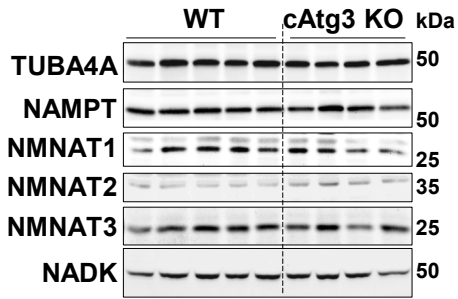
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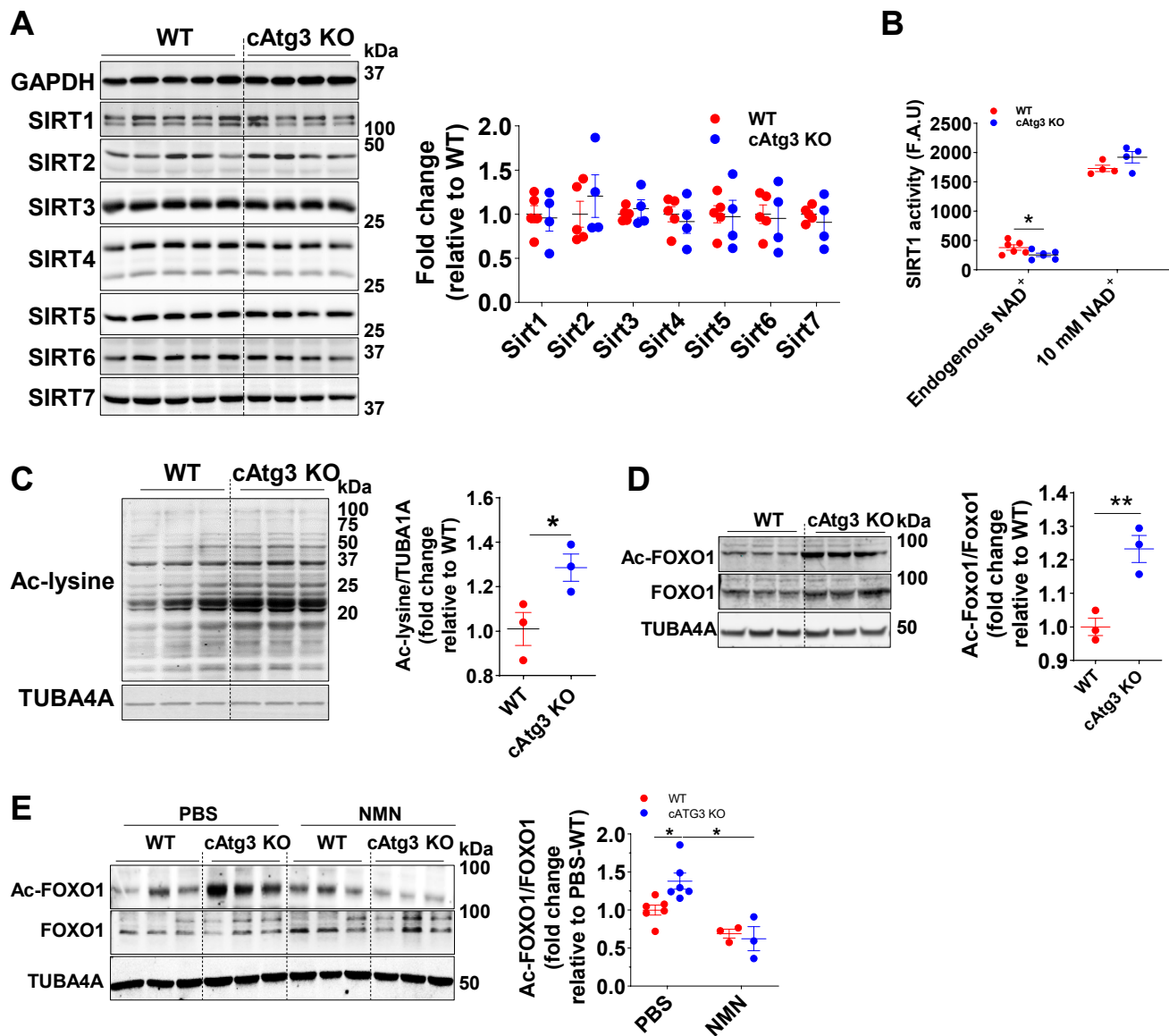
Appendix Figure S1. cAtg3-KO does not alter autophagic flux in non-cardiomyocyte tissues and does not influence MTOR and AMPK signaling in hearts, Related to Figure 1.

- A Protein levels of ATG3, LC3 (MAP1LC3A), SQSTM1, and GAPDH in liver, skeletal muscle, white adipose tissue, and kidney of WT and cAtg3-KO mice. Representative blots are shown, n=2 per group.
- B Quantification of MTOR S2448 and AMPK α T172 phosphorylation. Mice were randomly fed. n=6 per group. Data are mean \pm SEM.



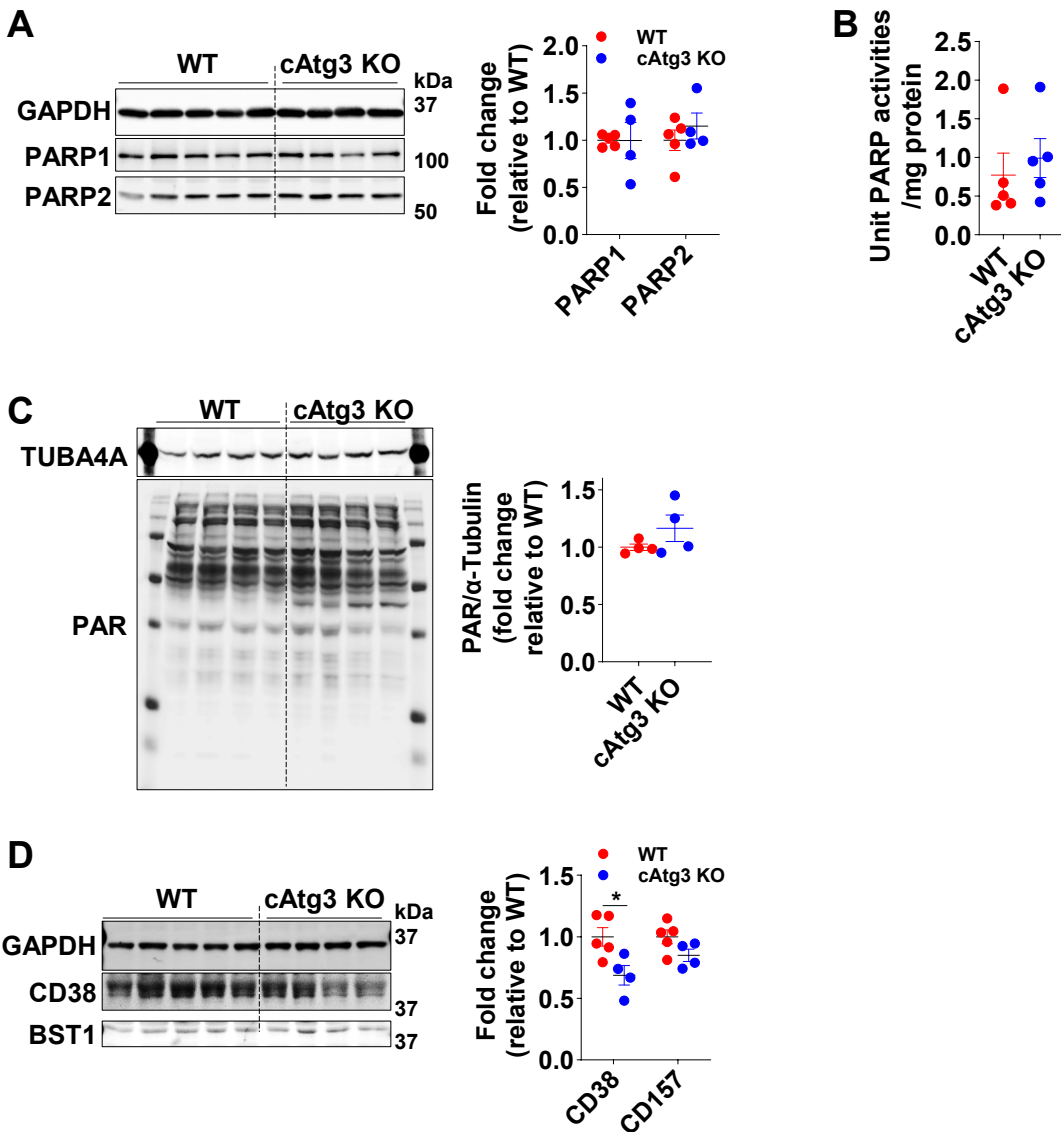
Appendix Figure S2. cAtg3-KO does not alter protein expression of NADK, NAMPT or NAMNAT1-3 in hearts, Related to Figure 5.

Protein levels of NADK, NAMPT, and NAMNAT1-3, TUBA4A (alpha-tubulin) in WT and cAtg3 KO mouse hearts. Mice were randomly fed, n=4 to 5 per group. Data are mean \pm SEM. Representative section images are shown.



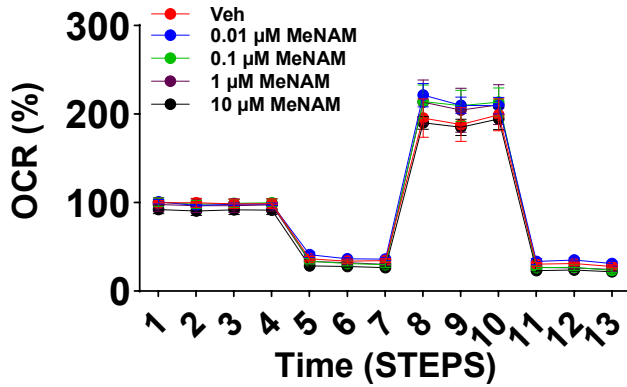
Appendix Figure S3. cAtg3-KO inhibited Sirt1 activities and increased protein acetylation in hearts, Related to Figure 5.

- A** Protein levels of SIRT1 to 7 and GAPDH in WT and cAtg3 KO mouse hearts. Mice were randomly fed, n=4 to 5 per group. Data are mean \pm SEM.
- B** SIRT1 enzymatic activity measurement in 4-week-old WT and Atg3 KO mouse hearts, without or with the addition of 10 mM exogenous NAD⁺, n=4 to 6 per group. Data are mean \pm SEM. Unpaired t-tests were used to determine statistical significance between two groups. *p<0.05.
- C** Levels of Ac-lysine and TUBA4A (alpha-tubulin) in 4-week-old WT and cAtg3 KO mouse hearts. Mice were randomly fed, n=3 per group. Data are mean \pm SEM. An unpaired t-test was used to determine statistical significance between two groups. *p<0.05.
- D** Protein levels of Ac-FOXO1, FOXO1, and TUBA4A in 4-week-old WT and cAtg3 KO mouse hearts. Mice were randomly fed, n=3 per group. Data are mean \pm SEM. An unpaired t-test was used to determine statistical significance between two groups. **p<0.01.
- E** Ac-FOXO1 expression levels in hearts from mice that were injected with PBS or NMN, n=3 to 6 per group. Data are mean \pm SEM. One-way ANOVA followed by Bonferroni's multiple comparison tests was used to determine statistical significance. *p<0.05.



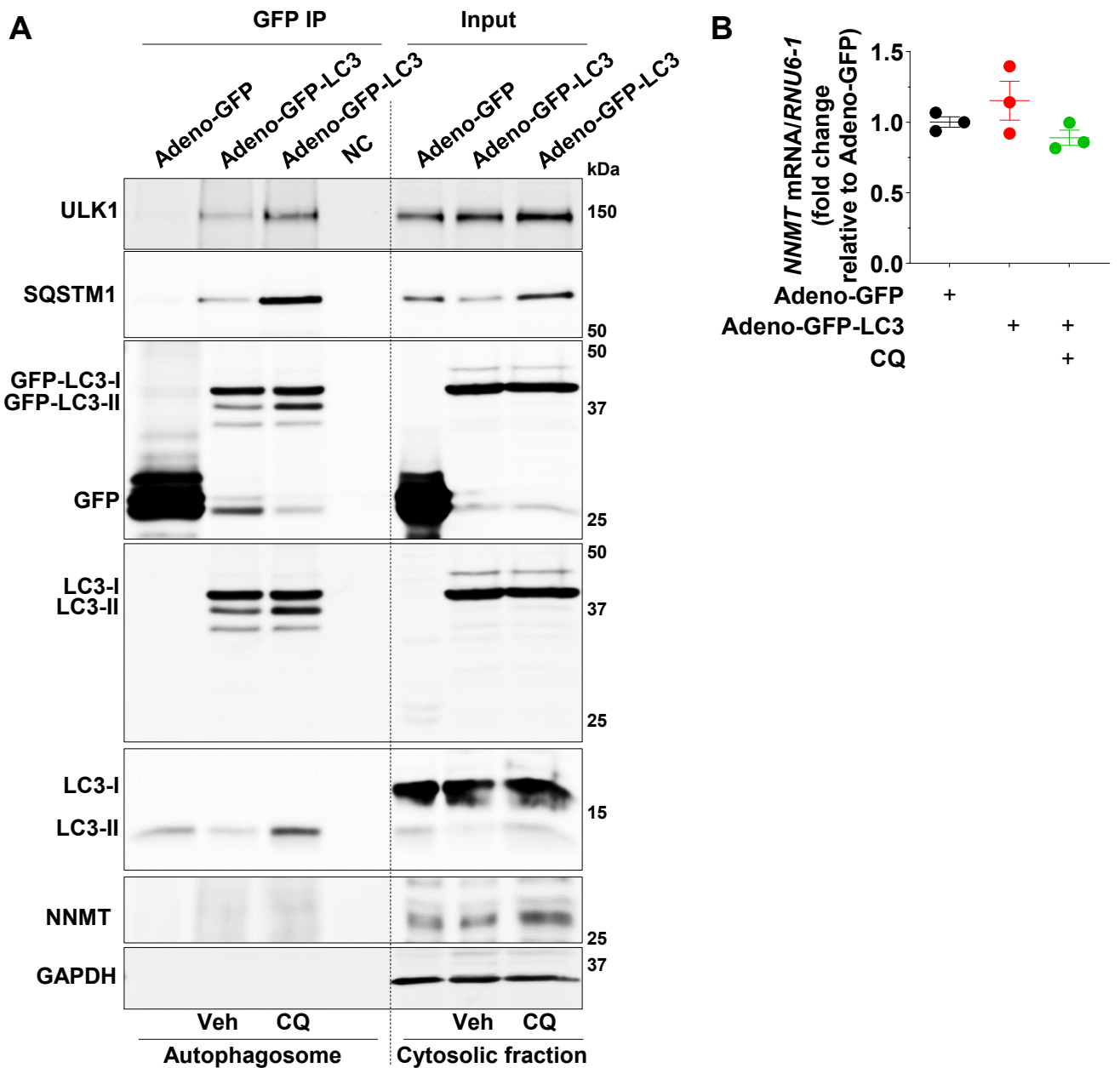
Appendix Figure S4. cAtg3-KO does not alter PARP1/2 protein expression, PARP1/2 activities, or abundance of CD38 and BST1 (CD157) in hearts, Related to Figure 5.

- A PARP1/2 protein expression in 4-week-old WT and cAtg3 KO mouse hearts, n=4 to 5 per group. Data are mean \pm SEM.
- B PARP1/2 enzymatic activity measurement in 4-week-old WT and cAtg3 KO mouse hearts, n=5 per group. Data are mean \pm SEM.
- C Abundance of PAR in 16-week-old WT and cAtg3 KO mouse hearts, n=4 per group. Data are mean \pm SEM.
- D Protein levels of CD38 and BST1, and GAPDH in WT and cAtg3 KO mouse hearts. Mice were randomly fed, n=4 to 5 per group. Data are mean \pm SEM. An unpaired t-test was used to determine statistical significance between two groups. *p<0.05. Representative section images are shown.



Appendix Figure S5. Oxygen consumption in response to increasing concentrations of Me-NAM in H9c2 cardiomyocytes, Related to Figure 6.

Oxygen consumption rate (OCR) in H9c2 cells treated with PBS (Veh) or concentrations of MeNAM as indicated, n=3 to 4 per group. Data are mean \pm SEM. Step 1 to 4: 20 μ M glucose medium, step 5 to 7: oligomycin (1 μ g/ml), step 8 to 10: FCCP (0.5 μ g/ml), step 11 to 13: rotenone (1 μ g/ml).



Appendix Figure S6. Autophagosomes do not engulf NNMT protein or mRNA in H9c2 cardiomyocytes, Related to Figure 7. H9c2 cardiomyocytes were first transfected with adenovirus encoding either GFP or GFP-tagged LC3 (MAP1LC3A), and then cells encoding GFP-LC3 were treated with either vehicle (Veh) or chloroquine (CQ) at 20 μ M for 16 h. under nutrient replete conditions.

A The protein levels of ULK1, SQSTM1, GFP, GFP-LC3, LC3, NNMT, and GAPDH in isolated autophagosomes. GFP-LC3 were probed with antibodies against GFP and LC3, and the images of immunoblots were acquired using LI-COR Odyssey DXL system at two independent laser channels, respectively. Therefore, GFP antibody detects GFP and GFP-LC3, and LC3 antibody detects GFP-LC3 only. The experiments were repeated three times. Representative blots are shown. NC indicates the separation buffer with GFP antibody. Input is cytosolic fractions.

B *NNMT* mRNA levels in isolated autophagosomes. n=3 per group. Data are mean \pm SEM.

Appendix Tables

Table S1. **Percentage of cAtg3 KO mice**

| Sex | Genotype | N. | % |
|-----|----------|----|-------|
| M | WT | 89 | 25.72 |
| | cAtg3 KO | 85 | 24.57 |
| F | WT | 81 | 23.41 |
| | cAtg3 KO | 91 | 26.30 |

Appendix Tables

Table S2. **Primer Sequences**

| Gene | Sequences |
|-------------------|--|
| <i>GAPDH</i> (M) | GCAACAATCTCCACTTTGCCAC AATGGTGAAGGTCGGTGTGAAC |
| <i>GAPDH</i> (R) | GCTCTCTGCTCCTCCCTGTTT GAGGCTGGCACTGCACAA |
| <i>RP16S</i> | GATTTGCTGGTGTGGATATC TCTTTGATCTCCTTCTTAGA |
| <i>ACTB</i> (M) | CGATGCCCTGAGGCTCTT T TGGATGCCACAGGATTCCA |
| <i>ACTB</i> (R) | GAGACCTTCAACACCCCAGCC TCGGGGCATCGGAACCGCTCA |
| <i>RPL13A</i> (M) | CTCTGGCCTTTTTCTTTTTG CCGAAGAAGGGAGACAGTTC |
| <i>RPL13A</i> (R) | CCACCCTATGACAAGAAAAGC ACATTCTTTTCTGCCTGTTTCC |
| <i>18S</i> (R) | GCCGCTAGAGGTGAAATTCTTA CTTTCGCTCTGGTCCGTCTT |
| <i>RNU6-1</i> (R) | GCAAATTCGTGAAGCGTTCC |
| <i>NPPA</i> (M) | ATGGGCTCCTTCTCCATCA CCTGCTTCTCAGTCTGCTC |
| <i>NPPB</i> (M) | GGATCTCCTGAAGGTGCTGT TTCTTTTGTGAGGCCTTGGT |
| <i>MYH7</i> (M) | GCCATCATGCACTTTGGAAAC CCCATGAGGTAGGCTGATTTGT |
| <i>PLN</i> (M) | AACAGGCAGCCAAATGTGA CCCAGCTAAGCTCCCATAAG |
| <i>NNMT</i> (R) | CAGAGCTGAGACACGATGGA GCAGGCAGAGAGAAGCTGAT |
| <i>NNMT</i> (M) | GATTGCACGCCTCAACTTCT GAACCAGGAGCCTTTGACTG |
| <i>NRF1</i> (M) | CTTCAGAACTGCCAACCACA GCTTCTGCCAGTGATGCTAC |
| <i>NRF2</i> (M) | AGTCTTCACTGCCCTCATC TCTGTCAGTGTGGCTTCTGG |
| <i>TFAM</i> (M) | GCAAAGGATGATTCCGGCTC TCTGCTCTTCCCAAGACTTCA |
| <i>NMNAT1</i> (M) | TGAGTCCATGGGGAGAAGTT AGGACTAGGGCCGTTTGG |
| <i>NMNAT2</i> (M) | ATCCCGCCAATCACAATAAA GCAGCTTCAATCCCATCACT |
| <i>NMNAT3</i> (M) | CAGAAGCACCACAGGGATTG CCTGCAGCACGTTTACAGTC |
| <i>NMAPT</i> (M) | TCACGGCATTCAAAGTAGGA GCAGAAGCCGAGTTCAACAT |
| <i>NMRK1</i> (M) | CTTGAAGCTTGCTCTGCGAC CTCCGTTTGTACACCACCA |
| <i>NMRK2</i> (M) | AAGCCCCAGGACCAAATAGC GCGTGCAAACCTTGTGTGGAT |

| | |
|--------------------------------|--|
| <i>PARP1</i> (M) | CACCTTCCAGAAGCAGGAGA GCAGCGAGAGTATTCCCAAG |
| <i>PARP2</i> (M) | GCAACAGAAGACGACTCTCCT CAGCCATAGGCCCTTTTCTCT |
| <i>RPL32</i> | ACATCGGTTATGGGAGCAAC GGGATTGGTGACTCTGATGG |
| <i>mtDNA</i> (M) | CCTATCACCCCTTGCCATCAT GAGGCTGTTGCTTGTGTGAC |
| <i>CHROMOSOME 6</i> (M) | ATGGAAAGCCTGCCATCATG TCCTTGTTGTTTCAGCATCAC |
| <i>PPARα</i> | GAGAATCCACGAAGCCTACC AATCGGACCTCTGCCTCTTT |
| <i>VLCAD</i> | AGGCAGTTCTGGACAAGCCA TTCCTCAAAGAACCGGGCCA |
| <i>LCAD</i> | GCATTGGTGGGGACTTGCTC TGTCATGGCTATGGCACCGA |
| <i>MCAD</i> | ACTGACGCCGTGCAGATTTT GCTTAGTTACACGAGGGTGATG |

Appendix Tables

Table S3. **Antibodies**

| Name | Company | Catalog Number |
|------------------------------|-------------------------|-----------------------|
| ATG3 | Sigma-Aldrich | A3231 |
| LC3 | Sigma-Aldrich | L8918 |
| TUBA4A | Sigma-Aldrich | T5168 |
| ACTB | Cell Signaling | 3700 |
| SQSTM1 | Cell Signaling | 5114 |
| Acetylated-Lysine | Cell Signaling | 9441 |
| ATG7 | Cell Signaling | 8858 |
| FOXO1 | Cell Signaling | 2880 |
| RELA S536 | Cell Signaling | 3036 |
| RELA | Cell Signaling | 6956 |
| MTOR S2448 | Cell Signaling | 4517 |
| MTOR | Cell Signaling | 2971 |
| P70S6K T389 | Cell Signaling | 9206 |
| p70S6K | Cell Signaling | 2708 |
| AMPK α T172 | Cell Signaling | 2531 |
| AMPK α | Cell Signaling | 2793 |
| ULK1 | Cell Signaling | 8054 |
| GFP | Cell Signaling | 55494 |
| Sirtuin antibody sampler kit | Cell Signaling | 9787 |
| SIRT4 | Cell Signaling | 69786 |
| PARP1 | Cell Signaling | 9532 |
| NADK | Cell Signaling | 89833 |
| NRF1 | Cell Signaling | 69432 |
| GAPDH | Cell Signaling | 2118 |
| PRKN | Cell Signaling | 4211 |
| CASP3 | Cell Signaling | 9665 |
| CD38 | Proteintech | 60006-1-Ig |
| BST1 | Proteintech | 16337-1-AP |
| NMNAT1 | Proteintech | 11399-1-AP |
| NMNAT3 | Proteintech | 13236-1-AP |
| PARP2 | Proteintech | 20555-1-AP |
| NMNAT2 | ThermoFisher Scientific | PA5-115662 |
| NNMT | Proteintech | 15123-1-AP |
| PINK1 | Proteintech | 23274-1-AP |
| NAMPT | Proteintech | 11776-1-AP |
| GAPDH | SCBT | SC-32233 |
| TFAM | SCBT | SC-166965 |
| NRF2 | SCBT | SC-722 |
| Ac-FKHR | SCBT | SC-49437 |
| PARGC1A | SCBT | SC-13067 |
| SIRT1 | SCBT | SC-74504 |
| TOMM20 | SCBT | SC-17764 |
| SDHA | abcam | ab14715 |

Appendix Tables

Table S4. **siRNA sequence**

| Name | Sequences/Company with catalog number |
|--|--|
| Control siRNA | 5'-UAAGGCUAUGAAGAGAUAC-3' 5'-GUAUCUCUUCAUAGCCUUA-3' |
| <i>ATG3</i> siRNA | 5'-CCCAGAAGAGUUUGUGGCAGCUGGA-3' 5'-UCCAGCUGCCACAAACUCUUCUGGG-3' |
| ON-TARGETplus SMARTpool <i>NNMT</i> siRNA | GE Dharmacon (L-101014-02-0005) |
| SignalSilence <i>SQSTM1/p62</i> siRNA II | Cell Signaling Technology (6399) |
| <i>SQSTM1/p62</i> | 5'-CCUGUGGUGGGAACUCGCUAUAAGU-3' 5'-ACUUAUAGCGAGUUCCCACCACAGG-3' |
| SignalSilence® <i>NF-κB p65</i> (<i>RELA</i>) siRNA I | Cell Signaling Technology (6261S) |

Appendix Tables

Table S5. **Plasmids and adenovirus**

| | Name | Company/Institute | Catalog Number/Provider |
|------------|------------------|--|-------------------------|
| Plasmids | <i>HA-FLAG</i> | Addgene | 10792 |
| | <i>HA-SQSTM1</i> | Addgene | 28027 |
| | <i>ATG7</i> | Addgene | 24921 |
| Adenovirus | <i>GFP</i> | Beth Israel Deaconess Medical Center, Boston, MA | Dr. Pavlos Pissios |
| | <i>GFP-LC3</i> | Cedars Sinai, Los Angeles, CA | Dr. Roberta A. Gottlieb |
| | <i>NNMT</i> | Beth Israel Deaconess Medical Center, Boston, MA | Dr. Pavlos Pissios |