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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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n/a	a Confirmed					
	The exact	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement				
	A stateme	ement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly				
	The statis Only comm	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.				
\boxtimes	A description of all covariates tested					
\boxtimes	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons					
	A full deso	cription of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) ition (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)				
\boxtimes		For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>				
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings					
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes					
\boxtimes	\square Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated					
	•	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.				
Software and code						
Policy information about <u>availability of computer code</u>						
Da	ata collection	Data collection was adquired and stored using microsoft excel. Clinical signs were recorded in a in house developed form and subsequentenly transferred to Microsoft excel.				

Data

Data analysis

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

- Accession codes, unique identifiers, or web links for publicly available datasets

All data analysis was performed using Prism v9.3.1.

- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our <u>policy</u>

The protein microarray data discussed in this publication have been deposited in NCBI's Gene Expression Omnibus 48 and are accessible through GEO Series accession number GSE180642 and GSE205172.

Research involving human participants, their data, or biological material

Policy information and sexual orientat		vith <u>human participants or human data</u> . See also policy information about <u>sex, gender (identity/presentation),</u> thnicity and racism.		
Reporting on sex	and gender	Not applicable		
Reporting on race other socially rele groupings		Not applicable		
Population chara	cteristics	Not applicable		
Recruitment		Not applicable		
Ethics oversight		Not applicable		
Note that full informa	tion on the appro	oval of the study protocol must also be provided in the manuscript.		
Field-spe	cific re	porting		
Please select the or	ne below that is	s the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
Life sciences	В	ehavioural & social sciences		
For a reference copy of t	he document with a	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>		
Life scier	ices stu	udy design		
All studies must dis	close on these	points even when the disclosure is negative.		
Sample size	No sample size calculation was performed. Number of animals/group (n=12) meets and exceeds the standards for these types of studies in the field.			
Data exclusions	No data was excluded in the analyses.			
Replication	Virus growth kinetics were analyzed using the Gompertz growth non-linear regression model. Differences in growth rate between viruses were calculated by AUC analysis, followed by Brown-Forsythe and Welch ANOVA plus Dunnett's T3 post hock analysis. Two-way ANOVA was employed to determine virus growth differences by timepoint. Statistically significant differences between viruses' growth rates are denoted. Western blot analyses were repeated at least twice starting from repeated infections/transfections.			
Randomization	Mice were randomly assigned to different group. Dfferent analyses were performed based on vaccine backbone, presence/absence of IGIP, and impact of biological sex as extensively detailed in the manuscript.			
Blinding	The study design, particularly during the animal study, required that personnel be fully aware of the groups in question to avoid errors in the allocation of data points during data collection. Personnel working and generating protein microarray data were blind to sample type and group allocation.			
We require information	on from authors a	Decific materials, systems and methods about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & exp	perimental sv	vstems Methods		
n/a Involved in th	· · · · · · · · · · · · · · · · · · ·			
Antibodies		ChIP-seq		
Eukaryotic	karyotic cell lines			
Palaeontol	ogy and archaeol	ogy MRI-based neuroimaging		
Animals an	d other organism	is and the second secon		
Clinical data				
Dual use re	Dual use research of concern			
Plants				

Antibodies

Antibodies used

The following primary antibodies were used in this study: 1) Influenza B (B/Massachusetts/03/2010) Hemagglutinin / HA1 Antibody, Rabbit PAb, Antigen Affinity Purified, Sino Biological, catalog number #40191-T56. 2) Influenza B NP (B/Florida/4/2006) Polyclonal Antibody, Invitrogen, catalog number #PA5-81758, lot number XE3571622A. 3) IGIP Polyclonal Antibody, Invitrogen, catalog number #PA5-65316, lot number YD3882996. 4) GAPDH Monoclonal Antibody (6C5), Invitrogen, catalog number #AM4300, lot number 2673398. 5) Recombinant Anti-GFP antibody [EPR14104], abcam, catalog number #ab183734, lot number 1033451-2. 5) Polyclonal anti-influenza B raised in mice, developed in house. The following seconday antibodies were used:1) Goat anti-Mouse IgG (H+L) Secondary Antibody, HRP, invitrogen, catalog number #31430, lot number YD375335. 2) Goat Anti-Rabbit IgG H&L (HRP), abcam, catalog number #ab6721, lot number GR3251481-1. The molecular marker was detected using Precision Protein™ StrepTactin-HRP Conjugate, BioRad, catalog number #1610380, lot number 64333457

Validation

Influenza B (B/Massachusetts/03/2010) Hemagglutinin / HA1 Antibody, Rabbit PAb, Antigen Affinity Purified, Sino Biological, catalog number #40191 has been valudated for western blot (1:1000-1:5000) and ELISA (1:5000-1:10000). Influenza B NP (B/Florida/4/2006) Polyclonal Antibody, Invitrogen, catalog number #PA5-81758 has been validated for Western blot (1:1000-1:5000) and ELISA (1:5000-1:10000). IGIP Polyclonal Antibody, Invitrogen, catalog number #PA5-65316 has been validated for ICC/IFF (0.25ug/mL-2ug/mL). Validation for western blot was performed in house. Polyclonal anti-influenza B raised in mice, developed in house was validated for western blot in house. GAPDH Monoclonal Antibody (6C5), Invitrogen, catalog number #AM4300 has been validated for western blot (3ug/mL), IHC, ICC/IF (1ug/mL), IP and dot blot. Recombinant Anti-GFP antibody [EPR14104], abcam, catalog number #ab183734 has been validated for Flow citometry, western blot, IHC-P and ICC/IF.

Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

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Cell line source(s) Madin-Darby canine kidney (MDCK) and human embryonic kidney 293T cells (HEK293T) were a kind gift from Robert Webster, St Jude Children's Research Hospital, Memphis, TN, USA.

Authentication Cell lines were not authenticated

Mycoplasma contamination Mycoplasma testing is performed routinely in the lab using Fisher Scientific Universal Mycoplasma Detection Kit (catalog # 50189644FP)

Commonly misidentified lines

Not applicable.

Animals and other research organisms

Policy information about <u>studies involving animals</u>; <u>ARRIVE guidelines</u> recommended for reporting animal research, and <u>Sex and Gender in Research</u>

Laboratory animals

(See ICLAC register)

Male and female DBA/2J mice (Strain #000671) were purchased from The Jackson Laboratory (Bar Harbor, ME) at 5 weeks old and raised until 7 weeks of age. DBA/2J mice are also known as DBA2, D2J, D2, DBA or DBA/2

Wild animals

Not applicable

Reporting on sex

Biological sex as a variable of immune responses is at the core of the manuscript. It was taken into account and reported throughout the manuscript.

Field-collected samples

Not applicable

Ethics oversight

Animal studies were approved and conducted in compliance with all the regulations stated by the Institutional Animal Care and Use Committee (IACUC) of the University of Georgia (UGA; under AUP A2019 01-004-A2). Vaccination and challenge studies were conducted under ABSL-2 conditions at the Davison Life Sciences Complex, UGA. Animal studies and procedures were performed according to the Institutional Animal Care and Use Committee Guidebook of the Office of Laboratory Animal Welfare and PHS policy on Humane Care and Use of Laboratory Animals. Animal studies were carried out in compliance with the ARRIVE guidelines (https://arriveguidelines.org).

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Plants

Seed stocks	Not applicable
Novel plant genotypes	Not applicable
Authentication	Not applicable