1	Cholinergic- α 7 nAChR signaling suppresses SARS-CoV-2 infection and			
2	inflammation in lung epithelial cells			
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4	Jing Wen, Jing Sun, Yanhong Tang, Jincun Zhao, Xiao Su			
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6	Supplementary information			
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8 9	Materials and Methods			
10	Antibodies and compounds			
11	Anti-ACE2 (ab272500, Abcam), anti-FOS (66590-1-Ig, Proteintech), anti-SARS-CoV-2			
12	nucleocapsid (ab271180, Abcam), anti-GAPDH (M20006; Abmart), Rhodamine (TRITC)			
13	AffiniPure Goat Anti-Rabbit IgG (Jackson Immunoresearch). GTS-21 (ab120560,			
14	Abcam), T5224 (T5416, Topscience).			
15				
16	Cells			
17	HEK293T cell line were cultured in Dulbecco's Modified Eagle Medium (DMEM) (Gibco)			
18	supplemented with 10% fetal bovine serum (Gibco), 100 U/ml penicillin, and 100			
19	$\mu g/ml$ of streptomycin (Invitrogen) at 37°C under 5% CO2. Calu3 cell line were cultured			
20	in Minimum Essential Medium (MEM) (Gibco) supplemented with 20% fetal bovine			
21	serum (Gibco), 100 U/ml penicillin, and 100 μ g/ml of streptomycin (Invitrogen) at 37°C			
22	under 5% CO ₂ . Caco2 cell line were cultured in Eagle's minimum essential medium			
23	(MEM) (WISENT) supplemented with 20% fetal bovine serum (Gibco), 100 U/ml			
24	penicillin, and 100 μ g/ml of streptomycin (Invitrogen) at 37°C under 5% CO ₂ .			
25				
26	Animals			
27	lpha7nAChR-deficient mice (C57BL/6 background, B6.129S7-Chrn $lpha$ 7tm1Bay, number			
28	003232) were purchased from the Jackson Laboratory (Bar Harbor, ME, USA. The mice			
29	were housed at 22°C on a 12 h light/dark cycle and with free access to sufficient food			

and water. Anesthesia was conducted with pentobarbital sodium (50 mg/kg) injected
 intraperitoneally (ip). The animal studies were approved by the Committees on Animal
 Research of the Institut Pasteur of Shanghai, Chinese Academy of Sciences. The mice
 were instilled with SARS-CoV-2 (African strain) into the nasal cavity at a dose of 5 × 10⁴
 FFU. All SARS-CoV-2 infection experiments were performed in a biosafety level-3
 laboratory (Guangzhou Institute of Respiratory Health, Guangzhou, Guangdong China).

37 SARS-CoV-2 pseudovirus production and virus entry assay

HEK293T cells were co-transfection with pSPAX2, pLenti-GFP-luc and pCDNA3.1-SARSCoV-2-S by Lipo2000. The supernatants were harvested at 48 h post transfection,
filtered with 0.45 µm filter, and centrifuged at 1000 × g for 5 min to remove cell debris.
To infect cells with pseudovirions, cells were seeded into 24-well plates and inoculated
with 500 µl media containing pseudovirions. After 24 or 48 h, cells were lysed with 100
µl lysis buffer (promega) for 30 min on ice. The entry efficiency was measured by
quantification of the luciferase activity (promega).

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46 Focus forming assay (FFA)

47 Vero E6 cells were seeded in 96-well plates one day before infection. Virus cultures were serially diluted and used to inoculate Vero E6 cells at 37°C for 1 h. Inocula were 48 49 then removed before adding 125 µl 1.6% carboxymethylcellulose warmed to 37°C per well. After 24 h, cells were fixed with 4% paraformaldehyde and permeabilized with 50 51 0.2% Triton X-100. Cells were then incubated with a rabbit anti-SARS-CoV-2 52 nucleocapsid protein polyclonal antibody (Cat. No.: 40143-T62, Sino Biological, Inc. 53 Beijing), followed by an HRP-labeled goat anti-rabbit secondary antibody (Cat. No.: 54 109-035-088, Jackson ImmunoResearch Laboratories, Inc. West Grove, PA). The foci were visualized by TrueBlue Peroxidase Substrate (KPL, Gaithersburg, MD), and 55 56 counted with an ELISPOT reader (Cellular Technology Ltd. Cleveland, OH). Viral titers 57 were calculated as FFU per ml or per gram tissue.

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59 Real-time (RT-) qPCR

Total cellular RNA was extracted by TRIzol reagent (Invitrogen), and reverse transcribed 60 61 into cDNA using a reverse transcriptase kit (Tiangen, Beijing, China). Real-time PCR was 62 performed using the Thunderbird SYBR qPCR Mix (11203ES08; Yeasen) on the ABI QuantStudio 6 flex Real-Time PCR system. The primers were used as follows, 63 GAPDH: forward, 5'-ATCCCATCACCATCTTCCAGG-3' and reverse, 5'-CCTTCTCCATGGTGG 64 65 TGAAGA C-3'; **Human** *ACE2*: forward, 5'-CAAGAGCAAACGGTTGAACAC-3' 5'-66 and reverse, 67 CCAGAGCCTCTCATTGTAGTCT-3'; 5'-Human FOS: forward, 5'- GGGGCAAGGTGGAACAGTTAT-3' 68 and reverse, CCGCTTGGAGTGTATCAGTCA-3'; 69 5'-Mouse Fos: forward 5'-CGGGTTTCAACGCCGACTA-3' 70 and reverse, TTGGCACTAGAGACGGACAGA3'; 71

72 **Mouse** *CXcl2*: forward 5'-GCTGTCAATGCCTGAAG-3' and reverse, 5'-73 GGCGTCACACTCAAGCTCT-3';

- 74 COVID19-N nucleocapsid: forward 5'-CGAACTTCTCCTGCTAGAATGG-3' and reverse,
 75 5'-GTGACAGTTTGGCCTTGTTG-3'.
- 76

77 RNA-Seq and RNA-Seq data analysis

The Calu3 cells (epithelial cells isolated from lung tissue) were pretreated with either
PBS or GTS-21 (40 μM) for 30 min, then infected with or without SARS-CoV-2 virus
(African strain). The cells were harvested at 24 h after infection. Therefore, we had 4
groups: PBS+PBS, GTS-21+PBS, PBS+SARS-CoV-2, and GTS-21+SARS-CoV-2. Each group
had 3 samples.

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The total RNA was extracted using TRIzol reagent (Invitrogen). After quality test, the total RNA of each sample was sequenced using the DNBSEQ platform (BGI BIG DATABASE (https://biosys.bgi.com/#/report/mrna/expression). Data were analyzed in online analysis software Dr. Tom and STRING online (<u>https://string-db.org</u>). GeneCards (the Human Gene Database) (<u>https://www.genecards.org</u>) was used to search genes with known functions.

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91 Lung histology

Hematoxylin and eosin (H&E) staining of lungs was conducted as described previously
in our laboratory.

94

95 **Confocal microscopy**

96 Cells were fixed in 4% formaldehyde and permeabilized with 0.1% Triton-X 100. Then, 97 the cells were blocked with PBS containing 5% BSA for 30 min at room temperature. 98 Further, the cells were stained with ACE2 antibody, followed by secondary Rhodamine 99 (TRITC)-labeled anti-rabbit IgG antibodies (Jackson Immunoresearch) at room 100 temperature. The nucleus was labeled with DAPI. Slides were imaged on a laser-101 scanning confocal microscope (Olympus FV-1200). Images were quantified by Image J 102 Pro.

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104 **Reactive oxygen species (ROS) assay**

105 ROS were detected by using ROS Assay Kit-Highly Sensitive DCFH-DA kit (DOJINDO 106 LABORATORIES). Calu3 cells were washed twice with HBSS buffer. Add DCFH-DA Dye 107 probe to the cells and mixed completely, cells were incubated at 37°C for 30 min. Then 108 wash cells for twice with HBSS buffer and observe ROS by fluorescence microscope.

109

110 Western blotting

111 Cells were lysed for 30 min at 4°C in lysis buffer (Beyotime). After centrifugation for 10 112 min at 12,000 × g, supernatant was boiled in loading buffer and analyzed by SDS-PAGE. 113 Specific primary antibodies were used, followed by horseradish peroxidase-conjugated 114 goat anti-mouse IgG or goat anti-rabbit IgG (Jackson Immunoresearch) as the 115 secondary antibodies.

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117 Immunoprecipitation-Mass Spectrometry

118 A549 cells treated with or without 40 μ M GTS-21 for 48 h, cells were collected and 119 lysed in IP lysis buffer (Beyotime) for 30 min on ice. After centrifugation for 10 min at 120 12,000 × g, the supernatants were incubated with 5 μ g anti- α 7 nAChR (sc-58607, Stan 121 Cruz) antibody at 4°C overnight. 25 μ l Protein G-labeled Dynabeads (Thermo Scientific)

were added into each sample at 4°C for 6 h. Dynabeads were washed 3 times with RIPA buffer. Proteins were eluted by boiling 10 min with loading buffer. The immunoprecipitates were separated by SDS-PAGE, each pull-down sample was run into the separation gel. We cut off the whole band as one sample and subjected it to in-gel trypsin digestion and Mass Spectrometry analysis (Performed by Orbitrap Velos Pro, Shanghai Institute of Nutrition and Health, Chinese Academy of Sciences).

129 Statistical analysis

130 Statistics were calculated using GraphPad Prism software (GraphPad, San Diego, CA).

131 An unpaired t-test was used unless there were multiple comparisons, in which case we

132 used one-way ANOVA with a post hoc Bonferroni test (with a significance level of p <

- 133 0.05). The results are shown as mean ± SD.



/	Biological Process (Gene Ontology)				
GO-term	description	count in network	strength	false discovery rate	
GO:0051607	Defense response to virus	39 of 170	2.11	3.51e-77 🔘	
GO:0045087	Innate immune response	35 of 558	1.55	7.29e-48	
GO:0006955	Immune response	37 of 979	1.33	1.40e-44	
GO:0045071	Negative regulation of viral genome replication	15 of 53	2.2	8.22e-26	
GO:0048525	Negative regulation of viral process	16 of 96	1.97	1.73e-24	

Supplementary Figure S1. Analysis of downregulated viral defense response genes 154 155 in RNAseq data from GTS-21 treated SARS-CoV-2 infected Calu3 cells. A Hotmap of enriched genes related to viral defense responses in PBS or GTS-21 treated Calu3 cells 156 157 infected with or without wild SARS-CoV-2. N = 3 in each group, P values are indicated between PBS + SARS-CoV-2 and GTS-21 + SARS-CoV-2 groups. Unpaired t test. B-C 158 Protein interaction network and KEGG analysis of enriched genes related to viral 159 defense responses in PBS or GTS-21 treated Calu3 cells infected with or without wild 160 SARS-CoV-2. 161

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Supplementary Figure S2. Activation of α 7 nAChR suppresses ACE2 expression 163 depending on FOS. A Hotmap of enriched genes of FOS family in PBS or GTS-21 164 treated Calu3 cells infected with or without wild SARS-CoV-2 N = 3 in each group, P 165 values are indicated between PBS + SARS-CoV-2 and GTS-21 + SARS-CoV-2 groups. 166 167 Unpaired t test. B-C Protein interaction network and GO analysis of enriched FOS family genes in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-168 CoV-2. D Calu3 cells were pretreated with GTS-21 for 24 h and infected with 169 pseudovirus for another 24 h, the expression levels of FOS and ACE2 were detected by 170 western blotting. E-F In Caco2 cells, the endogenous FOS was knocked down with 171 lentiviruses containing specific shRNA. The mRNA and protein expression levels of FOS 172 and ACE2 were detected by RT-qPCR and western blotting. N = 3 in each group, ***P < 173 0.01 unpaired-t test. G the negative control and FOS knockdown cells were infected 174

- 175 with S pseudovirus, the infectivity was analyzed by luciferase assay. N = 3 in each group,
- 176 ******P < **0.01** unpaired-t test.



Supplementary Figure S3. Activation of α7 nAChR suppresses ROS and SARS-CoV-2 178 S protein entry. A-B Calu3 cells were infected with pseudovirus and treated with GTS-179 180 21 for 6 h, intracellular ROS was labeled by DCFH-DA fluorescent probes, ROS level was observed by fluorescence microscope. Green positive areas were analyzed by Image-181 182 pro Plus software. Experiments were repeated 3 times. P values are indicated between compared groups. Wilcoxson test. C Caco2 cells were pretreated with H₂O₂ for 4 h and 183 184 then infected with pseudovirus for 24 h, virus infection level was analyzed by luciferase assay. N = 3 in each group, *P < 0.05, unpaired-t test. 185

178 proteins interacted with α 7 nAChR were detected by Co-IP and Mass Spectrometry. Among them, 145 protein coding genes were found in our RNAseq database



\sim	Biological Process (Gene Ontology)			
GO-term	description	count in network	strength	false discovery rate
GO:0019083	Viral transcription	11 of 115	1.11	9.52e-07 🔘
GO:0061621	Canonical glycolysis	6 of 27	1.48	2.78e-05 🥘
GO:0012501	Programmed cell death	24 of 1054	0.49	0.00013 🔘
GO:0000398	mRNA splicing, via spliceosome	9 of 294	0.62	0.0212 🔘
GO:0019322	Pentose biosynthetic process	2 of 4	1.83	0.0352

187 Supplementary Figure S4. Analysis of 145 α7 nAChR-binding proteins incorporated

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188 with RNAseq data from GTS-21 treated SARS-CoV-2 infected Calu3 cells. A Protein

189 interaction network in 145 α 7 nAChR-binding proteins. **B** GO analysis of genes of 145

190 α7 nAChR-binding proteins. N = 3 in each group, P < 0.05 is considered as a significant
 191 difference in compared groups when identified the differentially expressed genes.
 192 Unpaired-t test.



Supplementary Figure S5. Analysis of glutathione metabolism genes in RNAseq data from GTS-21 treated SARS-CoV-2 infected Calu3 cells. A Hotmap of enriched genes related to glutathione metabolism in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2. N = 3 in each group, P values are indicated between PBS + SARS-CoV-2 and GTS-21 + SARS-CoV-2 groups. Unpaired t test. B-C Protein interaction network and KEGG analysis of enriched genes related to glutathione metabolism in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2.



Supplementary Figure S6. Analysis of cytokine storm genes in RNAseq data from GTS-21 treated SARS-CoV-2 infected Calu3 cells. A Hotmap of enriched genes related to cytokine storm in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2. N = 3 in each group, P values are indicated between PBS + SARS-CoV-2 and GTS-21 + SARS-CoV-2 groups. Unpaired t test. B-C Protein interaction network and KEGG analysis of enriched genes related to cytokine and chemokine in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2.





Supplementary Figure S7. Activation of α 7 nAChR, which regulates SARS-CoV-2 infection, involves upregulation of α 7 nAChR, acetylcholine hydrolysis, HMGB and NF-kB activation. A-B. α 7 nAChR expression at mRNA levels in SARS-CoV-2-infected mouse lungs and human peripheral mononuclear cells isolated from SRAS patients. We analyzed GEO DataSets in Pubmed

215 (https://www.ncbi.nlm.nih.gov/sites/GDSbrowser?acc=GDS1028). Expression profiling of peripheral blood mononuclear cells (PBMC) from 10 adult patients with 216 217 severe acute respiratory syndrome (SARS) was presented. C Hotmap of CHRNA7 and ACHE in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2. N 218 = 3 in each group, *P < 0.05 between PBS + SARS-CoV-2 and GTS-21 + SARS-CoV-2 219 220 groups. Unpaired t test. D Gene expression of Hmgb-related genes in in SARS-CoV-2infected mouse lungs. N=3 in PBS and N = 6 in SARS-CoV-2 infected mice. Unpaired t 221 222 test. E Hotmap of HMGB1, HMGB2 and HMGB3 in PBS or GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2. N = 3 in each group, *P < 0.05 between PBS 223 + SARS-CoV-2 and GTS-21 + SARS-CoV-2 groups. Unpaired t test. F Gene expression of 224 225 *Nfkb*-related genes in in SARS-CoV-2-infected mouse lungs. N = 3 in PBS and N = 6 in SARS-CoV-2 infected mice. Unpaired t test. G Hotmap of NFKB related genes in PBS or 226 227 GTS-21 treated Calu3 cells infected with or without wild SARS-CoV-2. N = 3 in each group, *P < 0.05 between PBS + SARS-CoV-2 and GTS-21 + SARS-CoV-2 groups. 228 229 Unpaired t test.

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232 Supplementary Figure S8. Deficiency of *Chrna7* affects lung inflammation in SARS-

233 **CoV-2 infected mice.** The mice were infected with SARS-CoV-2 for 4 days and sacrificed.

The lungs were removed to extract RNA for qPCR or histology. **A** Changes of lung SRARs-CoV-2 *N protein gene, Fos,* and *Cxcl2*. N = 5 in each group. *P < 0.05, unpaired t test. **B** Histology of SARS-CoV-2-infected lungs from both *Chrna7* wildtype and knockout mice. (H&E staining, objective magnification × 10, scale bar 500 μ m); N = 5 in each group.

239

240 Repeated experiment G

As stated in our last response to a reviewer request, we are unable to replicate SARS-Cov-2 infection experiments due to the temporary unavailability of the P3 facility. However, in the pseudo-SARS-Cov-2-infected Calu3 cells, we found that both pretreatment and treatment with GTS-21 reduced ACE2 and FOS expression (A and B).



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246 Repeated experiment H

T-5224 is a transcription factor c-Fos/AP-1 inhibitor, which specifically inhibits the DNA binding activity of c-Fos/c-Jun without affecting other transcription factors. As below figure demonstrated that treatment with T5224 in Caco2 cells (A) reduced ACE2

250 expression at both mRNA and protein levels (B-C).

