

Supporting Information for

**The inhibitor protein IF₁ from mammalian mitochondria inhibits
ATP hydrolysis but not ATP synthesis by the ATP synthase
complex**

**Joe Carroll, Ian N. Watt, Charlotte Wright, Shujing Ding, Ian M. Fearnley
and John E. Walker**

*The Medical Research Council Mitochondrial Biology Unit, University of Cambridge,
Cambridge Biomedical Campus, Hills Road, Cambridge CB2 0XY, United Kingdom*

Corresponding author: John E. Walker

Email: walker@mrc-mbu.cam.ac.uk

Supporting information included:

Figures S1-S8

Tables S1-S6

A

Bovine	QKTGTAEVSSILEERILGADTSVDLEETGRVLSIGDGIARVHGLRNVQAEEMVEFSSGLK	60
Human	QKTGTAEVSSILEERILGADTSVDLEETGRVLSIGDGIARVHGLRNVQAEEMVEFSSGLK *****	60
Bovine	GMSLNLPEPDNVGVVVFVFNKDLIKEGDIVKRTGAIVDVVPVGEELLGRVVDALGNAIDGKGP	120
Human	GMSLNLPEPDNVGVVVFVFNKDLIKEGDIVKRTGAIVDVVPVGEELLGRVVDALGNAIDGKGP *****	120
Bovine	IGSKARRRVGLKAPGIIPRISVREPMQTGIKAVDSLVPPIGRGQRELIIGDRQTGKTSIAI	180
Human	IGSKTRRRVGLKAPGIIPRISVREPMQTGIKAVDSLVPPIGRGQRELIIGDRQTGKTSIAI *****	180
Bovine	DTIINQKRFNDGTDEKKKLYCIYVAIGQKRSTVAQLVKRLTDADAMKYTIIVVSATASDAA	240
Human	DTIINQKRFNDGSDKKKLYCIYVAIGQKRSTVAQLVKRLTDADAMKYTIIVVSATASDAA *****	240
Bovine	PLQYLAPYSGCSMGGEYFRDNGKHALIIYDDLKQAVAYRQMSLLLRPPGREAYPGDVFY	300
Human	PLQYLAPYSGCSMGGEYFRDNGKHALIIYDDLKQAVAYRQMSLLLRPPGREAYPGDVFY *****	300
Bovine	LHSRLLERAAMNDAFGGSLTALPVIETQAGDVSAYIPTNVISITDGGIFLETELFYKG	360
Human	LHSRLLERAAMNDAFGGSLTALPVIETQAGDVSAYIPTNVISITDGGIFLETELFYKG *****	360
Bovine	IRPAINVGLSVSRVGSAAQTRAMKQVAGTMKLELAQYREVAFAQFGSDLAATQQLLSR	420
Human	IRPAINVGLSVSRVGSAAQTRAMKQVAGTMKLELAQYREVAFAQFGSDLAATQQLLSR *****	420
Bovine	GVRLTELLKQGQYSPMAIEEQVAVIYAGVRGYLDKLEPSKITKFENAFLSHVVISQHALL	480
Human	GVRLTELLKQGQYSPMAIEEQVAVIYAGVRGYLDKLEPSKITKFENAFLSHVVISQHALL *****	480
Bovine	SKIRTDGKISEESDAKLKEIVTNFLAGFEA	510
Human	GTIRADGKISEQSDAKLKEIVTNFLAGFEA	510

B

Bovine	AAQASPSPKAGATTGRIVAVIGAVVDVQFDEGLPPILNALEVQGRETRLVLEVAQHLGES	60
Human	AAQTSPSPKAGAATGRIVAVIGAVVDVQFDEGLPPILNALEVQGRETRLVLEVAQHLGES *****	60
Bovine	TVRTIAMDGTEGLVRGQKVLDSGAPIRIPVGPETLGRIMNVIGEPIDERGPIKTKQFAAI	120
Human	TVRTIAMDGTEGLVRGQKVLDSGAPIKIPVGPETLGRIMNVIGEPIDERGPIKTKQFAPI *****	120
Bovine	HAEAPEFVEMSVEQEILVTGIKVVDLLAPYAKGGKIGLFGGAGVGKTVLIMELINNVAKA	180
Human	HAEAPEFVEMSVEQEILVTGIKVVDLLAPYAKGGKIGLFGGAGVGKTVLIMELINNVAKA *****	180
Bovine	HGGYSVFAGVGERTREGNDLYHEMIESGVINLKDATSKVALVYQGMNEPPGARARVALTG	240
Human	HGGYSVFAGVGERTREGNDLYHEMIESGVINLKDATSKVALVYQGMNEPPGARARVALTG *****	240
Bovine	LTVAEYFRDQEGQDVLFFIDNIFRFTQAGSEVSALLGRIPSAVGYQPTLATDMGTMQERI	300
Human	LTVAEYFRDQEGQDVLFFIDNIFRFTQAGSEVSALLGRIPSAVGYQPTLATDMGTMQERI *****	300
Bovine	TTTTKGSITSVQAIYVPADDLTPAPATTF AHL DATTVL SRAIAELGIYPAVDPLDSTSR	360
Human	TTTTKGSITSVQAIYVPADDLTPAPATTF AHL DATTVL SRAIAELGIYPAVDPLDSTSR *****	360
Bovine	IMDPNIVGSEHYDVARGVQKILQDYKSLQDIIAILGMDELSEEDKLTVSRARKIQRFLSQ	420
Human	IMDPNIVGSEHYDVARGVQKILQDYKSLQDIIAILGMDELSEEDKLTVSRARKIQRFLSQ *****	420
Bovine	PFQVAEVFTGHLGKLVPLKETIKGFQQILAGEYDHLPEQAFYMGPIEEAVAKADKLAEE	480
Human	PFQVAEVFTGHMGKLVPLKETIKGFQQILAGEYDHLPEQAFYMGPIEEAVAKADKLAEE *****	480
Bovine	HS-	482
Human	HSS	483
	**	

C

Bovine	ATLKDITRRLKSIKNIQKITKSMKMVAAAKYARAERELKPARVYGVGSLALYEKADIKTP	60
Human	ATLKDITRRLKSIKNIQKITKSMKMVAAAKYARAERELKPARIYGLGSLALYEKADIKGP	60
	*****.*****.*****.*.*:*****.*	
Bovine	EDKKKHLIIGVSSDRGLCGAIHSSVAKQMKSEANLAAAGKEVKIIGVGDKIRSLHRTH	120
Human	EDKKKHLIIGVSSDRGLCGAIHSSIAKQMKSEVATLTAAGKEVMLVGIGDKIRGILYRTH	120
	*****.*****.*****.*.*:*****.*	
Bovine	SDQFLVTFKEVGRPPPTFGDASVIALELLNSGYEFDEGSIIFNFRSVISYKTEEKPIFS	180
Human	SDQFLVAFKEVGRKPPTFGDASVIALELLNSGYEFDEGSIIFNFRSVISYKTEEKPIFS	180
	*****.*****.*****.*****.*****.*****.*****.*****	
Bovine	LDTISSAESMSIYDDIDADVLRNYQEYSLANIIYYSLKESTTSEQSARMTAMDNASKNAS	240
Human	LNTVASADSMSIYDDIDADVLRNYQEYSLANIIYYSLKESTTSEQSARMTAMDNASKNAS	240
	..:**.*:*****.*****.*****.*****.*****.*****.*****	
Bovine	EMIDKLTTLFNRTRQAVITKELIEIISGAAALD	273
Human	EMIDKLTTLFNRTRQAVITKELIEIISGAAALD	273
	*****.*****.*****.*****.*****.*****.*****.*****	

Fig. S1. Comparison of the sequences of the α -, β - and γ -subunits of bovine and human ATP synthase. The mature subunits are numbered from residue 1 to their C-termini. Symbols *, : and . denote identical, strongly conserved and weakly conserved residues respectively. (A) α -subunits (bovine, sp|P19483; human, sp|P25705) identical in 98.4% residues. (B) β -subunits (bovine, sp|P00829; human, sp|P06576), identical in 98.6% residues. (C) γ -subunits (bovine, sp|P05631; human, sp|P36542), identical in 92.3% residues with a further 6.2% residues highly conserved.

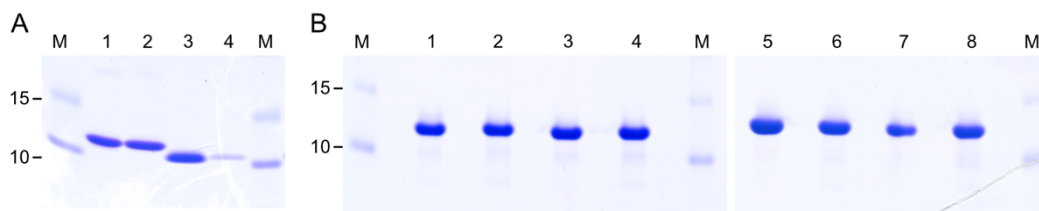
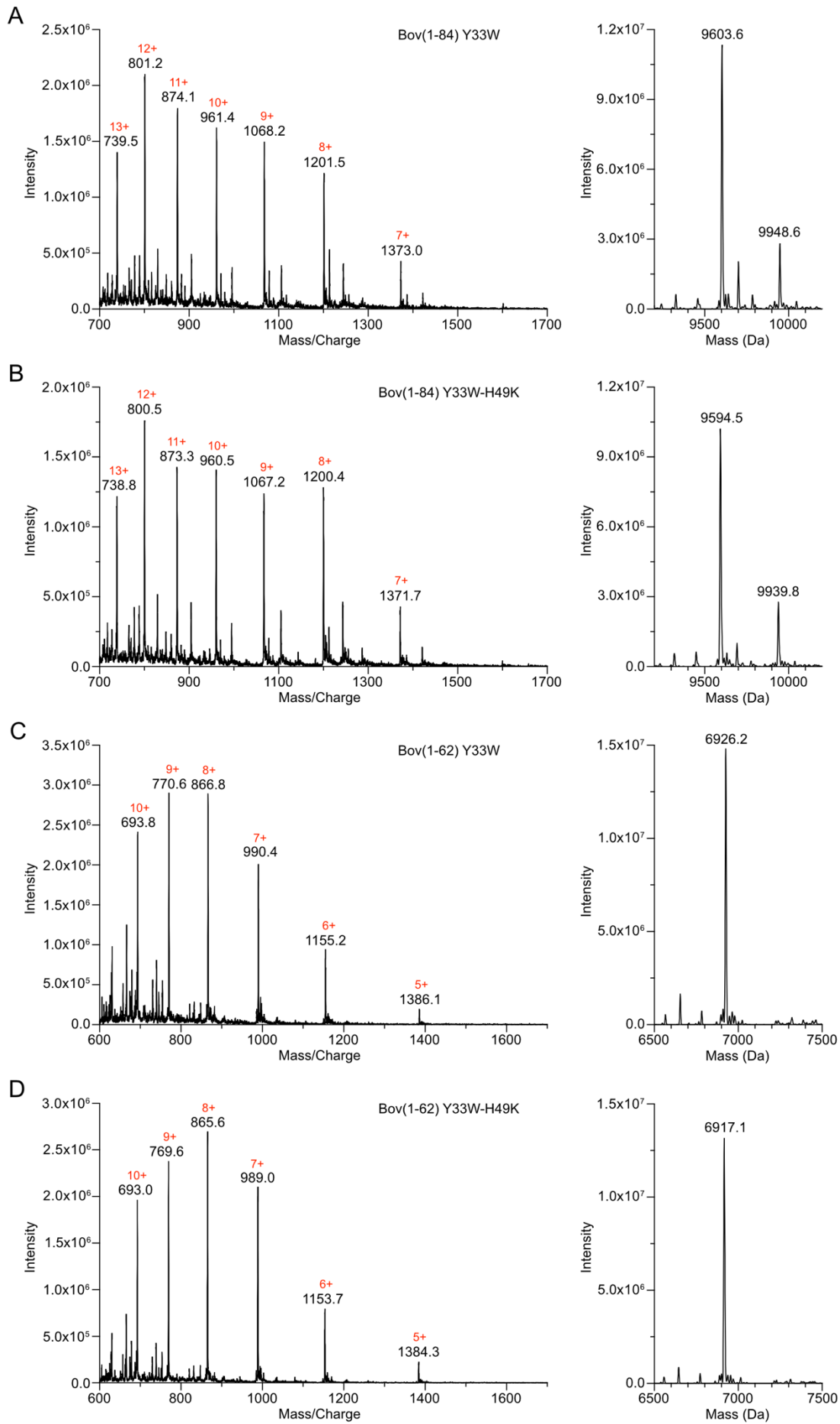
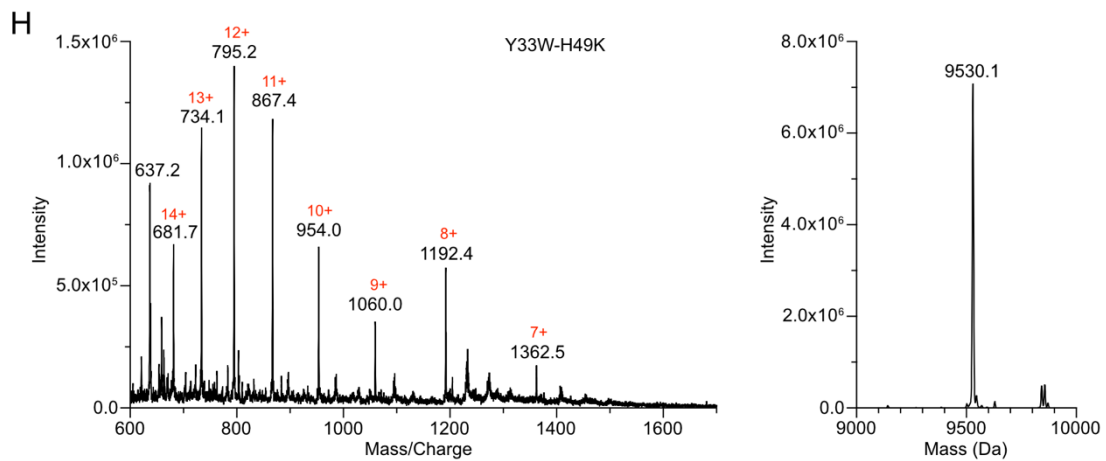
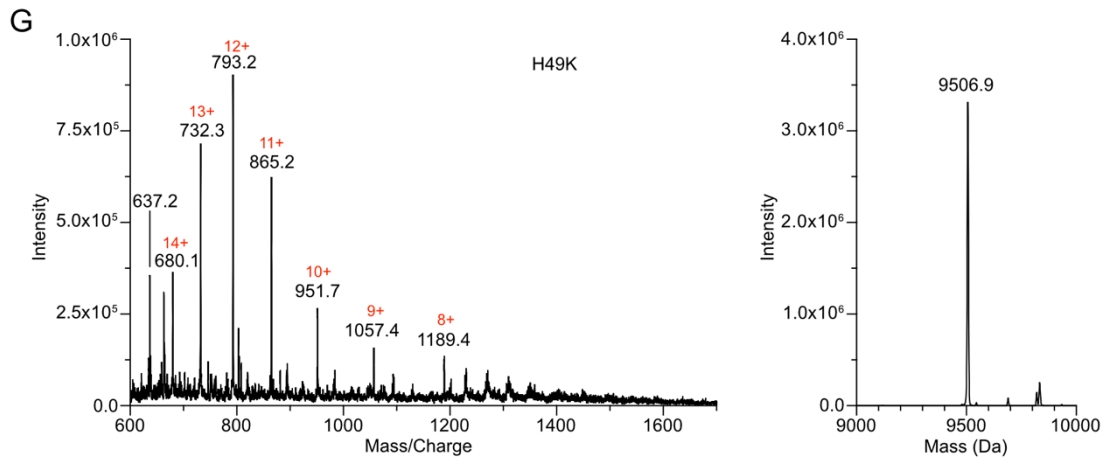
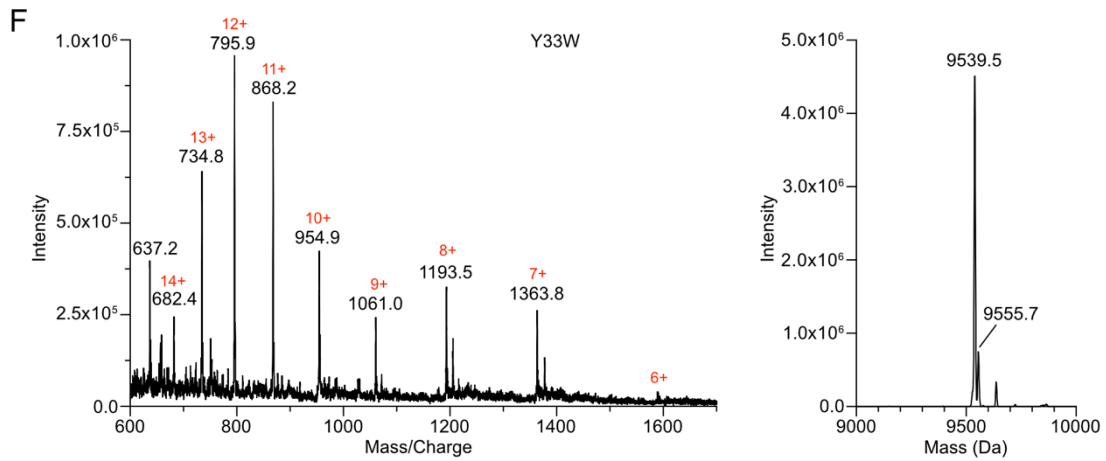
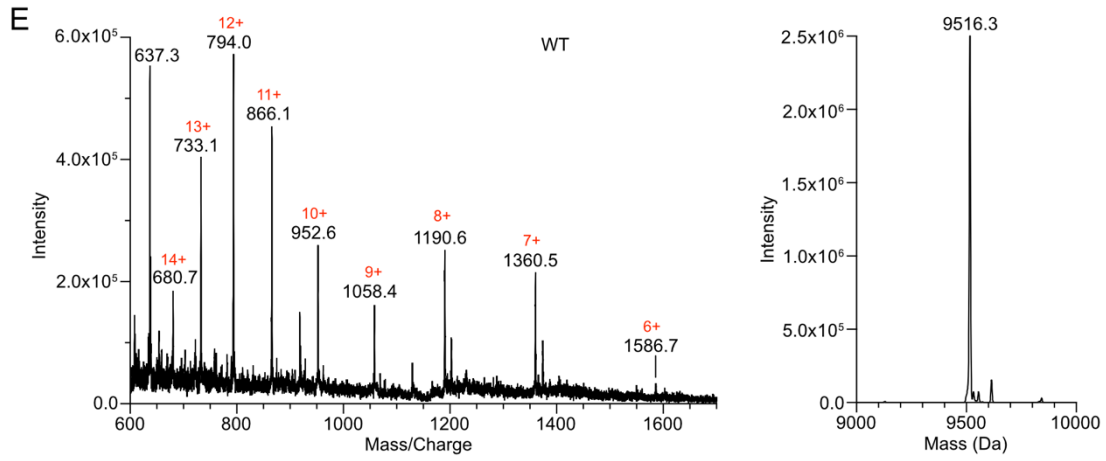


Fig. S2. Analysis of purified bovine and human IF₁ by SDS-PAGE. (A) Bovine IF₁. Lanes 1 and 2, bovine IF₁(1-84) with the mutations Y33W and Y33W-H49K, respectively; lanes 3 and 4, bovine IF₁(1-62) with the mutations Y33W and Y33W-H49K, respectively. (B) Human IF₁ (1-81). Lane 1, wild type (residues 1-81); 2-8, forms containing, respectively, the following mutations: 2, H49K; 3, Y33W; 4, Y33W-H49K; 5, S14D-Y33W; 6, S14D-Y33W-H49K; 7, S14E-Y33W; 8, S14E-Y33W-H49K. M, molecular weight markers (kDa) are indicated.





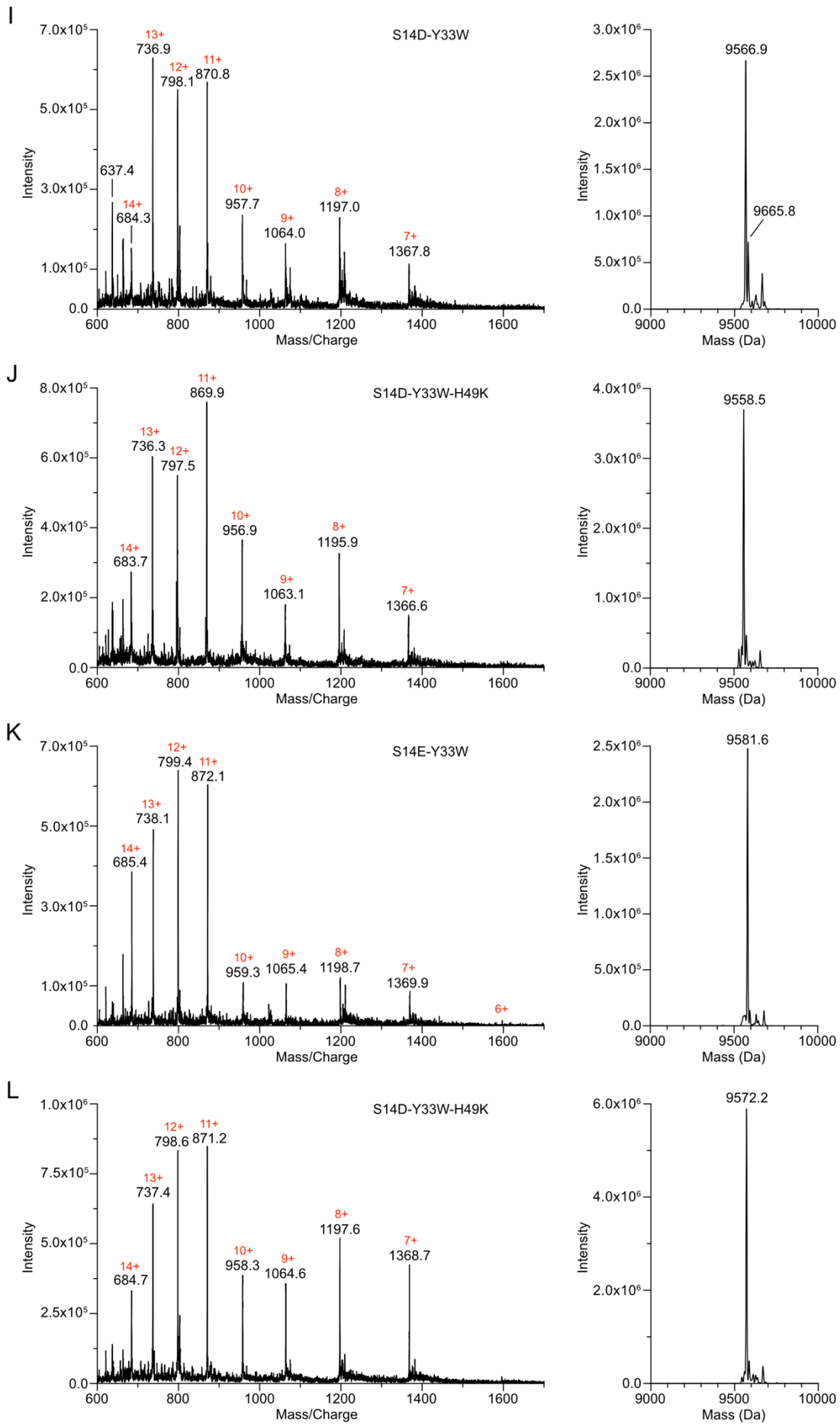


Fig. S3. ESI-MS spectra of recombinant bovine and human IF₁ proteins. Each row shows the raw spectra and the transformations of these data onto a molecular mass scale. (A) BovIF₁(1-84)-Y33W; (B) BovIF₁(1-84)-Y33W-H49K; (C) BovIF₁(1-62)-Y33W; (D) BovIF₁(1-62)-Y33W-H49K; (E) HumIF₁(1-81)-wild-type; (F) HumIF₁(1-81)-Y33W; (G) HumIF₁(1-81)-H49K; (H) HumIF₁(1-81)-Y33W-H49K; (I) HumIF₁(1-81)-S14D-Y33W; (J) HumIF₁(1-81)-S14D-Y33W-H49K; (K) HumIF₁(1-81)-S14E-Y33W; (L) HumIF₁(1-81)-S14E-Y33W-H49K.

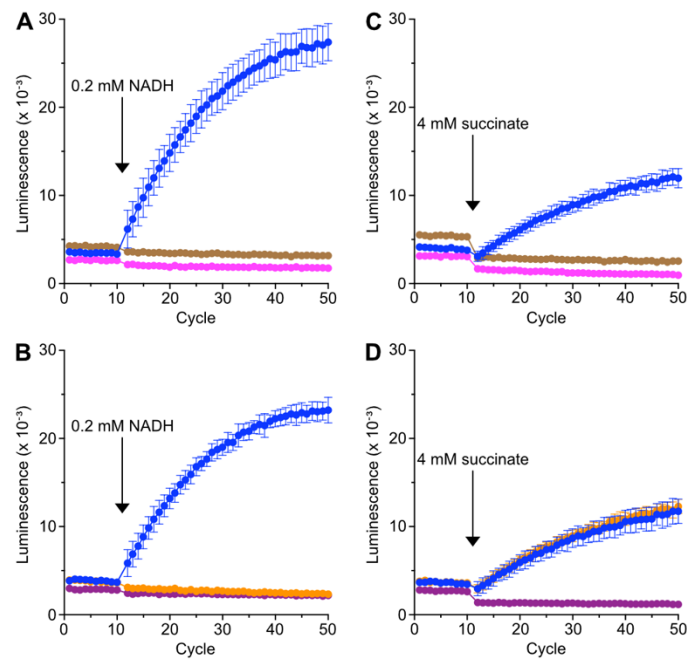


Fig. S4. Inhibition of ATP synthesis in bovine SMPs by respiratory inhibitors. ATP synthesis coupled to oxidation of (A and B), NADH (0.2 mM) or (C and D), succinate (4 mM). ATP synthesis in the presence of (●), DMSO vehicle only; (●), 1 μ M oligomycin; (●), 0.5 μ M FCCP; (●), 1 μ M rotenone; (●), 1 μ M antimycin A. Background luminescence levels were established for 10 measurement cycles before addition of substrate and generation of ATP. Measurements of luminescence were made in quadruplicate and average values \pm SD are shown.

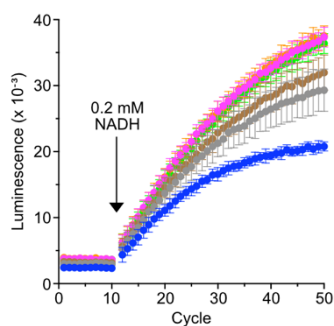


Fig. S5. Effect of the concentration of BovIF₁(1-84)-Y33W on the synthetic activity of ATP synthase. (●), no inhibitor; (●), 1.35 μM, (●), 6.75 μM; (●), 33.8 μM; (●), 67.5 μM; (●), 135 μM. At a concentration of 135 μM inhibitor protein, the molar ratio with respect to the ATP synthase is ca. 11000:1. ATP synthesis was initiated by the addition of NADH, and monitored by a luminescence continuous real-time assay with a luciferase-luciferin reagent. Background luminescence levels were established for 10 measurement cycles. N=4 wells, and data points show the average signal ± SD.

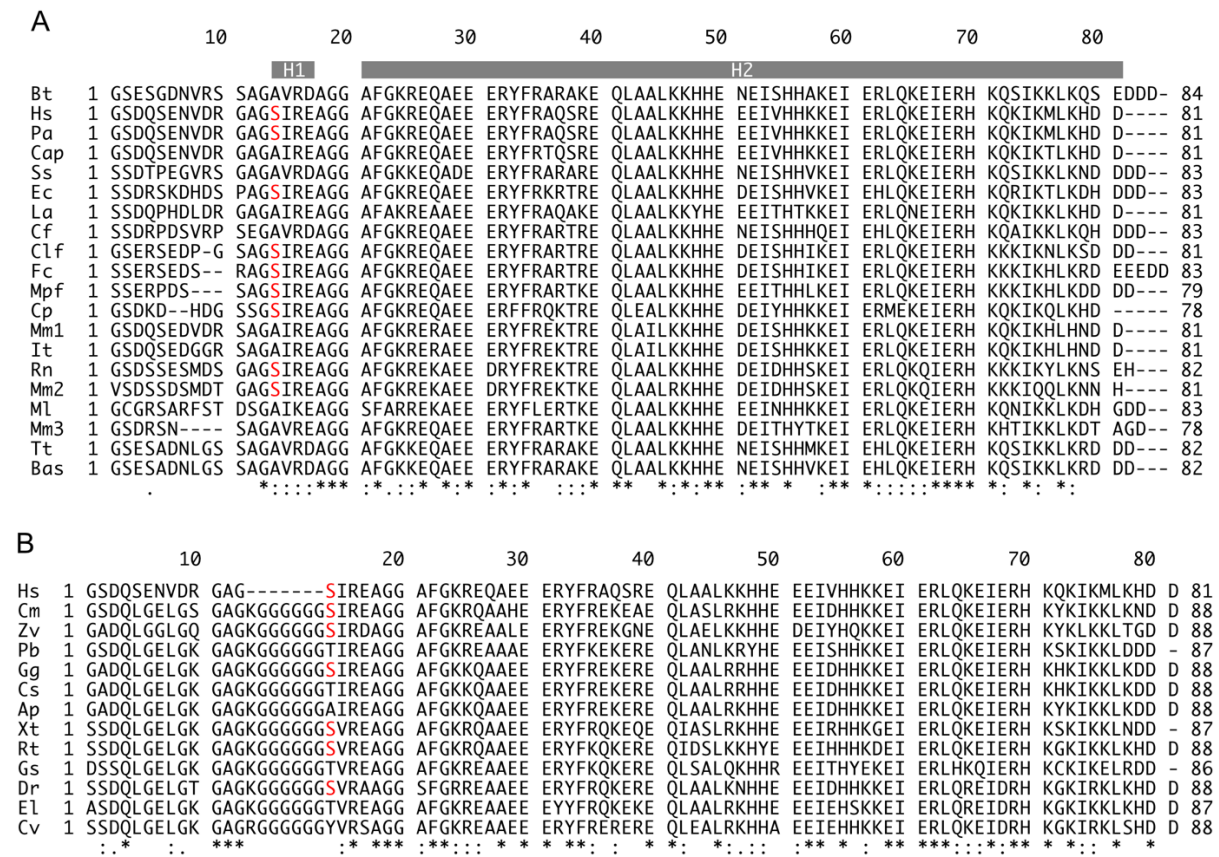


Fig. S6. Comparison of sequences of IF₁ proteins from various vertebrates. The N-terminal residues of only the mature bovine and human proteins have been identified experimentally. The mitochondrial import precursors of the inhibitors from other species are assumed to be processed at the same site. Residue Ser-14 in the human protein has been proposed to be phosphorylated reversibly. This residue and conserved serine residues in the same position in other species are shown in red. The symbols *, : and . denote identical, strongly conserved and weakly conserved residues respectively. (A) Sequences of mammalian IF₁ molecules. The grey bars H1 and H2 denote the positions of α -helices in the structure of the bovine protein. Bt, *Bos taurus* (cattle, sp|P01096); Hs, *Homo sapiens* (human, sp|Q9UII2); Pa, *Pongo abelii* (Sumatran orangutan, sp|Q5RFJ9); Cap, *Colobus angolensis palliatus* (Angolan colobus, tr|A0A2K5KBI5); Ss, *Sus scrofa* (pig, sp|Q29307); Ec, *Equus caballus* (horse, tr|F6ZXT0); La, *Loxodonta africana* (African bush elephant, tr|G3SWQ8); Cf, *Camelus ferus* (wild Bactrian camel, tr|S9XNE5); Clf, *Canis lupus familiaris* (domestic dog,

tr|E2QYN4); Fc, *Felis catus* (cat, tr|M3WIS8); Mpf, *Mustela putorius furo* (ferret, tr|M3YVR5); Cp, *Cavia porcellus* (guinea pig, tr|A0A286Y431); Mm1, *Marmota monax* (groundhog, tr|A0A5E4AD98); It, *Ictidomys tridecemlineatus* (thirteen-lined ground squirrel, tr|I3N8E6); Rn, *Rattus norvegicus* (brown rat, sp|Q03344); Mm2, *Mus musculus* (mouse, sp|O35143); Ml, *Myotis lucifugus* (little brown bat, tr|G1NSN7); Mm3, *Molossus molossus* (Pallas's mastiff bat, tr|A0A7J8F6B1); Tt, *Tursiops truncatus* (common bottlenose dolphin, tr|A0A2U3V0R3); Bas, *Balaenoptera acutorostrata scammoni* (minke whale, tr|A0A383Z6R7). (B) The sequence of human IF₁ compared with examples of orthologs from reptiles, birds, amphibians and fish. Hs, *Homo sapiens* (human, sp|Q9UII2); Cm, *Chelonia mydas* (green sea turtle, ncbi|XP_037737775); Zv, *Zootoca vivipara* (European common lizard, ncbi|XP_034976781); Pb, *Python bivittatus* (Burmese python, ncbi|XP_007434913); Gg, *Gallus gallus* (chicken, ncbi|XP_015153068); Cs, *Callipepla squamata* (scaled quail, ncbi|OXB61861); Ap, *Anas platyrhynchos* (mallard duck, ncbi|XP_027299537); Xt, *Xenopus tropicalis* (western clawed frog, sp|F7BK26); Rt, *Rana temporaria* (European common frog, ncbi|XP_040193223); Gs, *Geotrypetes seraphini* (Gaboon caecilian, ncbi|XP_033813241); Dr, *Danio rerio* (zebrafish, sp|A3KNL5); El, *Esox lucius* (northern pike, tr|C1BWJ3); Cv, *Cyprinodon variegatus* (sheepshead minnow, tr|A0A3Q2D9N0). Sequences were taken from the Swiss-Prot (sp), TrEMBL (tr) and NCBI (ncbi) databases. The alignments were made with Clustal Omega (1.2.4). In (B) the alignment was adjusted manually post Clustal Omega, placing the gap in the human sequence with the run of Gly residues found in the other proteins.

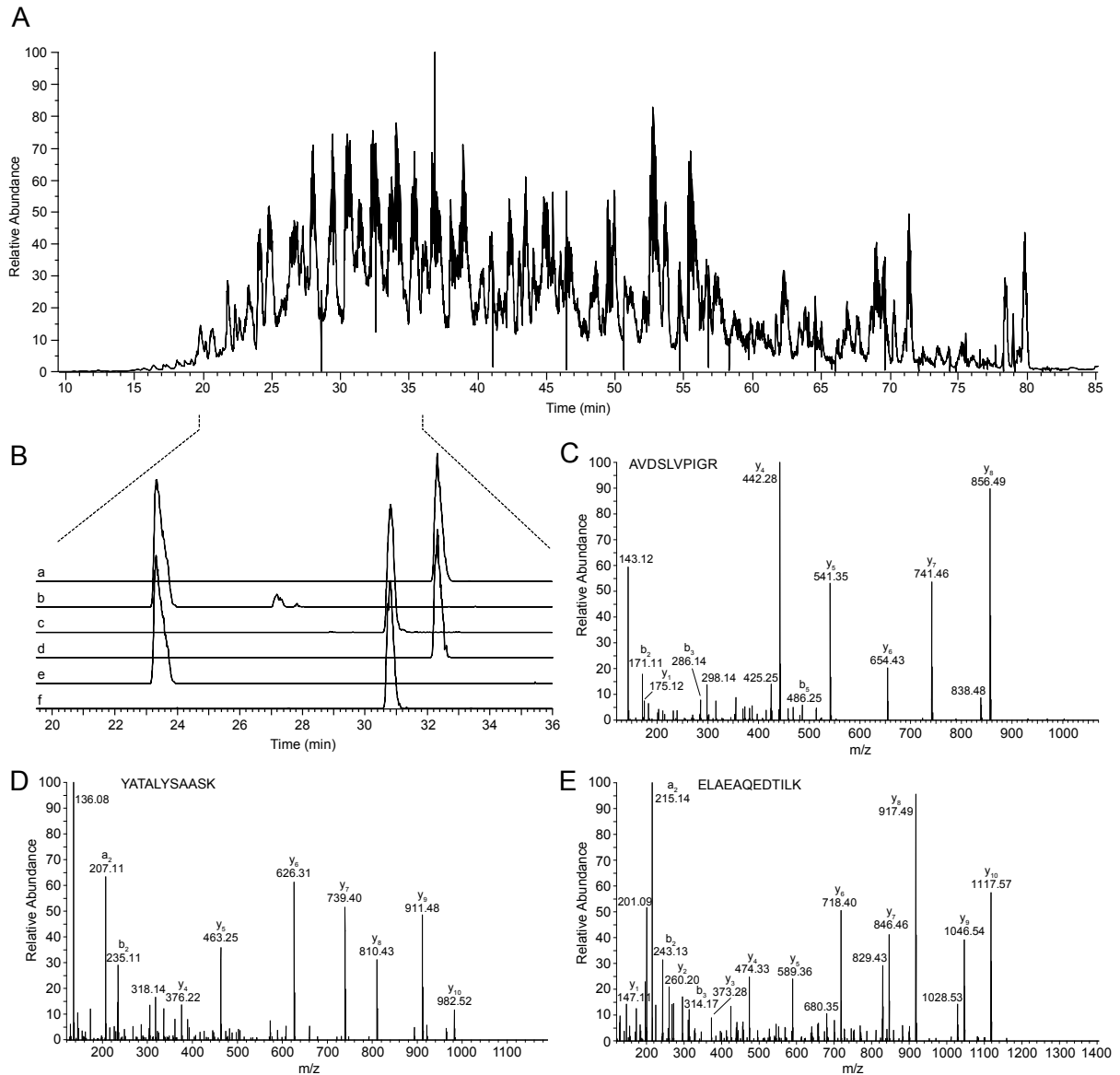


Fig. S7. The abundance of ATP synthase in bovine submitochondrial particles (SMPs) determined by quantitative mass spectrometry. Examples of: (A) LC-MSMS total ion current from an analysis of a tryptic digest of SMPs, spiked with heavy labelled subunit specific peptides (see Table S5). (B) Extracted ion chromatograms showing co-eluting peaks for the subunit specific labelled and endogenous peptides (charge state 2⁺) of subunit- α (a and d), OSCP (b and e) and subunit-e (c and f). (C-E) Tandem-MSMS spectra providing identification of the endogenous peptides.

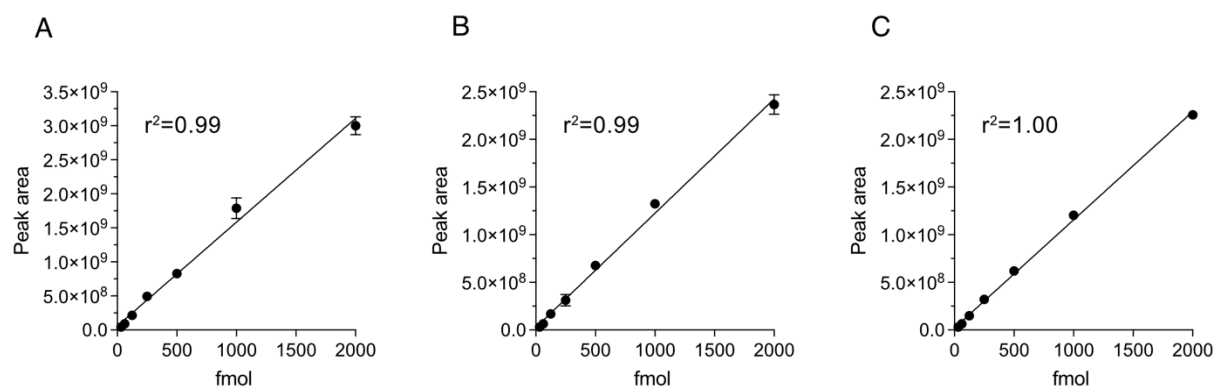


Fig. S8. Peak areas versus standard peptide concentration. A serial dilution of a mixture of three heavy labelled reference tryptic peptides (see Table S5) for ATP synthase were analyzed using an Orbitrap QE mass spectrometer (ThermoFisher). (A) α -subunit, (B) OSCP and (C) α -subunit-e with charge state 2^+ were obtained from the extracted ion chromatograms of the parent ions with Xcalibur (ThermoFisher) and an m/z tolerance of 5 ppm. $N=2$, \pm SD.

Table S1. Residues involved in binding bovine IF₁ to the β -, α - and γ -subunits of bovine F₁-ATPase and the identities of the equivalent amino acids in the human orthologs

IF ₁		β -subunit		α -subunit		γ -subunit	
bovine	human	bovine	human	bovine	human	bovine	human
S11	G11					N15	N15
A12	A12			E353	E353		
G13	G13	D386	D386				
V15	I15	D386	D386				
D17	E17	D386	D386				
F22	F22	D386	D386			I16	I16
		I390	I390				
		L391	L391				
R25	R25					E241	E241
E30	E30	R408	R408				
E31	E31	R408	R408				
Y33	Y33	K401	K401				
		M393	M393				
		D394	D394				
F34	F34	V404	V404				
		S405	S405				
		R408	R408				
		E454	E454				
R35	R35			E399	E399		
Q41	Q41	D450	D450				
L42	L42	P453	P453				
		L473	L473				
		A474	A474				
		H477	H477				
L45	L45	A470	A470				
		D471	D471				
		A474	A474				

The bovine data are taken from Bason *et al* (2014) (1).

Table S2. Content of ATP synthase in bovine SMPs

Experiment ¹	ATP synthase		Subunit ratio α : OSCP : e
	nmol/mg SMP \pm SD	mg/mg SMP \pm SD	
A	0.49 \pm 0.09	0.28 \pm 0.05	3.0 : 0.9 : 1.4
B	0.47 \pm 0.05	0.27 \pm 0.03	3.0 : 0.9 : 1.1
Average	0.48	0.28	3.0 : 0.9 : 1.2

¹ For experiment A, two samples of the same preparation of extracted SMPs were digested separately with trypsin and each analyzed twice by MS. The ratio is an average of the four calculated values. For experiment B, a single digest was performed, analyzed twice by MS, with the ratio an average of the two calculated values. Data file S27.

Table S3. Molar ratios of IF₁ and ATPase in various animals and tissues estimated by quantitative Western blotting

Molar ratio	Sample	Method	Reference
2.5:1	Goat heart mitochondria	Western blotting	(2)
4-6:1	Human heart and brain tissue	Western blotting	(3)
0.5-1:1	Human liver and kidney tissue	Western blotting	(3)
4-9:1	Mouse kidney, brain and colon tissue	Western blotting	(3)
0-1:1	Mouse heart and liver tissue	Western blotting	(3)

Table S4. Molecular masses of bovine and human IF₁ and mutant forms determined by electrospray mass spectrometry

Protein	Mass (Da)		Mass Difference (Da)
	Expected	Observed	
BovIF ₁ (1-84)-Y33W	9604.5	9603.6	-0.9
BovIF ₁ (1-84)-Y33W-H49K	9595.3	9594.5	-0.8
BovIF ₁ (1-62)-Y33W	6926.5	6926.2	-0.3
BovIF ₁ (1-62)-Y33W-H49K	6917.6	6917.1	-0.5
HumIF ₁ (1-81)-wild-type	9516.6	9516.3	-0.3
HumIF ₁ (1-81)-Y33W	9539.6	9539.5	-0.1
HumIF ₁ (1-81)-H49K	9507.6	9506.9	-0.7
HumIF ₁ (1-81)-Y33W-H49K	9530.6	9530.1	-0.5
HumIF ₁ (1-81)-S14D-Y33W	9567.6	9566.9	-0.7
HumIF ₁ (1-81)-S14D-Y33W-H49K	9558.6	9558.5	-0.1
HumIF ₁ (1-81)-S14E-Y33W	9581.6	9581.6	0
HumIF ₁ (1-81)-S14E-Y33W-H49K	9572.7	9572.2	-0.5

Table S5. Reference tryptic peptides for subunits of bovine ATP synthase

Subunit	Sequence (residue numbers)	Mass	
		Unlabelled	Heavy Labelled
α	AVDSLVPPIGR (152-161) ¹	1025.5869	1032.6041
OSCP	YATALYSAASK (18-28) ¹	1144.5764	1151.5936
e	ELAEAQEDTILK (59-70) ¹	1358.6929	1365.7101

¹With ¹³C and ¹⁵N at leucine residues 156, 22 and 69, respectively.

Table S6. Determination of subunit abundance and ratio data

Sequence	Labelled peptide		Endogenous peptide			Subunit ratio	F ₁ F ₀ nmol/mg
	MH ²⁺	Area	MH ²⁺	Area	fmol		
AVDSLVPPIGR	517.3093	2550550439	513.8007	21750821234	1731.2	3.00	0.451
YATALYSAASK	576.8041	5329160543	573.2955	13880112315	528.7	0.92	0.413
ELAEAQEDTILK	683.8623	2921375675	680.3537	10941705354	760.3	1.32	0.594

The Table contains a representative example of data obtained from an LC-MSMS analysis of a tryptic digest of SMPs (1.28 μ g), spiked with heavy labelled subunit specific peptides (203 fmol; see Table S5). Peak area ratios were obtained from the extracted ion chromatograms of the co-eluting peaks for the subunit specific peptides (charge state 2⁺) and the amount of endogenous peptide was calculated from the known quantity of labelled peptide. The subunit ratio was calculated based on the presence of three α -subunits in each enzyme complex.

References

1. Bason, J. V., Montgomery, M. G., Leslie, A. G. W. and Walker, J. E. (2014) Pathway of binding of the intrinsically disordered mitochondrial inhibitor protein to F₁-ATPase. *Proc. Natl. Acad. Sci. U S A* **111**, 11305-11310
2. Di Pancrazio, F., Mavelli, I., Isola, M., Losano, G., Pagliaro, P., Harris, D. A. and Lippe, G. (2004) In vitro and in vivo studies of F₀F₁ ATP synthase regulation by inhibitor protein IF₁ in goat heart. *Biochim. Biophys. Acta* **1659**, 52-62
3. Esparza-Moltó, P. B., Nuevo-Tapióles, C., Chamorro, M., Nájera, L., Torresano, L., Santacatterina, F. and Cuezva, J. M. (2019) Tissue-specific expression and post-transcriptional regulation of the ATPase inhibitory factor 1 (IF1) in human and mouse tissues. *FASEB J.* **33**, 1836-1851