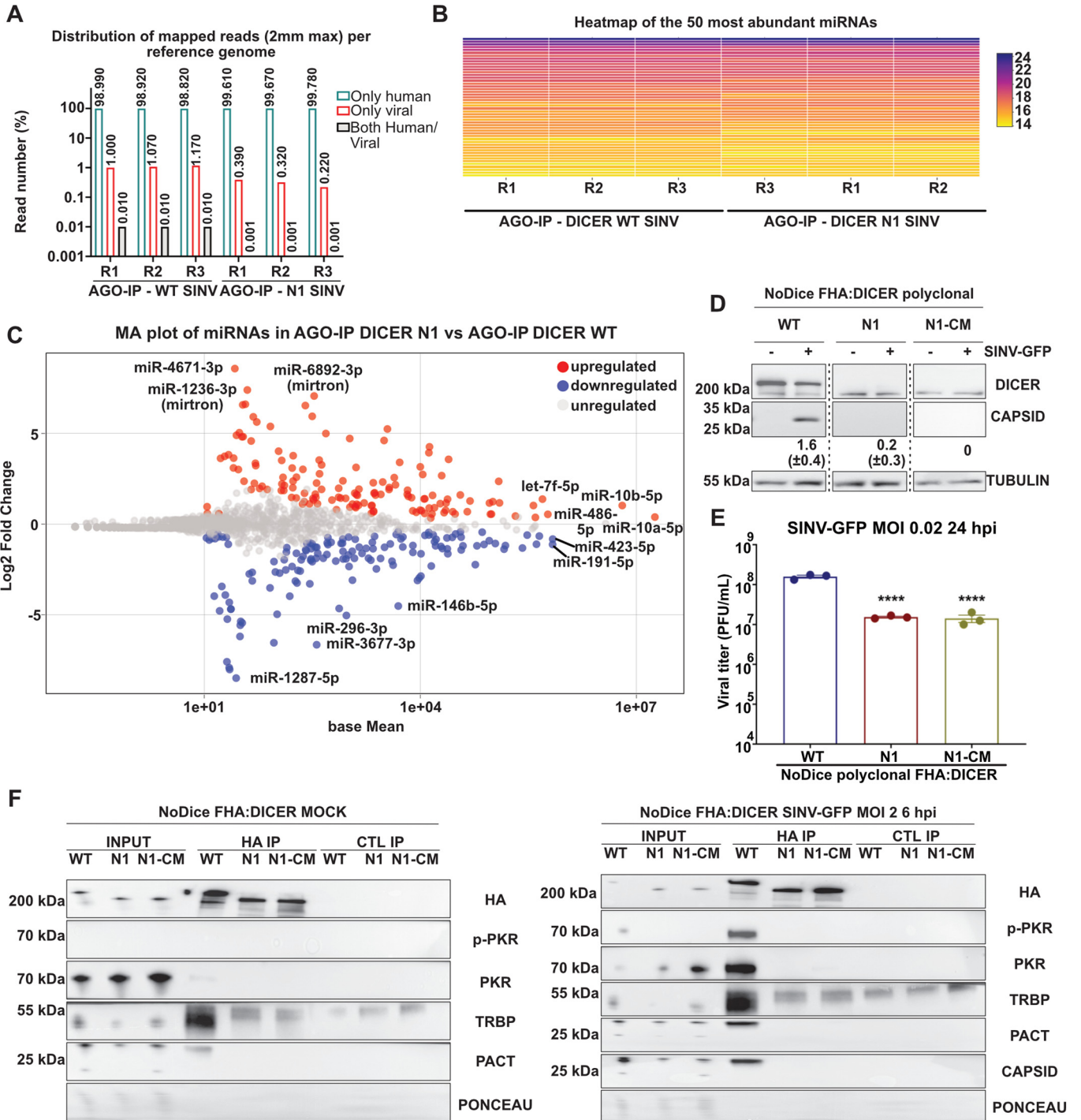


Expanded View Figures

Figure EV1. Dicer N1 retains the same miRNA profile and interacting partners upon infection.

(A) Representation of small RNA reads distribution (in percent) upon AGO-IP in the three replicates for NoDice FHA:DICER WT and N1. Green: reads mapping to human RNAs only; Red: reads mapping to viral RNAs only; gray: reads mapping to both human and viral RNAs. (B) Heatmap representing the relative expression levels of the 50 most abundant human miRNAs in all three replicates upon AGO-IP followed by small RNA sequencing in SINV-GFP infected NoDice FHA:DICER WT and N1 cells at an MOI of 0.02 for 24 h. (C) MA plot for miRNA enrichment upon AGO-IP followed by small RNA sequencing in SINV-GFP infected NoDice FHA:DICER N1 vs WT cells at an MOI of 0.02 for 24 h. Each dot represents a miRNA either up- (red), down- (blue) or un-regulated (gray). $n = 3$ biological replicates (Wald test, DESeq2 package). (D) Western blot analysis of DICER and CAPSID expression in SINV-GFP infected polyclonal NoDice FHA:DICER WT, N1 and N1-CM cells at an MOI of 0.02 for 24 h. Alpha-Tubulin was used as loading control. Band intensity was quantified and normalized to Tubulin for three independent biological replicates, then represented as mean ($+/-$ SD) under the corresponding lane. (E) Mean ($+/-$ SEM) of SINV-GFP viral titers in the same samples as in (D), infected at an MOI of 0.02 for 24 h ($n = 3$ biological replicates) from plaque assay quantification. Ordinary one-way ANOVA test with Dunnett's correction. **** $p < 0.0001$. (F) Western blot analysis of Dicer interacting partners upon HA-IP in NoDice FHA:DICER WT, N1 and N1-CM cells, in mock (left) or SINV-GFP infected (right) conditions at an MOI of 2 for 6 h ($n = 3$ biological replicates). Anti-HA antibodies were used to validate the immunoprecipitation and Ponceau was used as a loading control. Source data are available online for this figure.



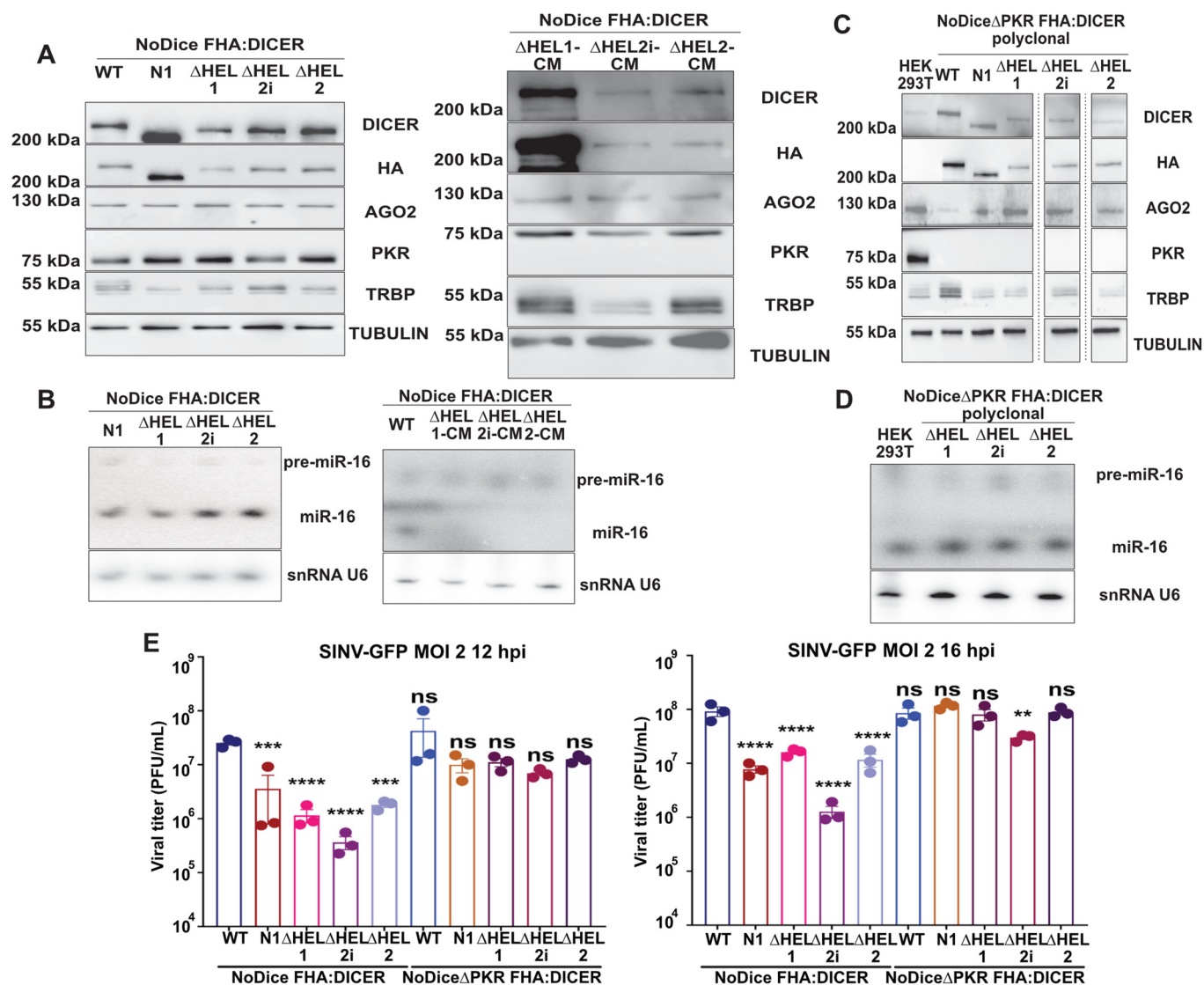


Figure EV2. The helicase sub-domains as well as PKR are important for Dicer antiviral activity.

(A) Western blot analysis of DICER, HA, AGO2, PKR and TRBP expression in the monoclonal cell lines NoDice FHA:DICER WT, N1, Δ HEL1, Δ HEL2i and Δ HEL2 (left, $n = 3$ biological replicates) and Δ HEL1-CM, Δ HEL2i-CM and Δ HEL2-CM (right, $n = 2$ biological replicates). Alpha-Tubulin was used as a loading control. (B) Northern blot analysis of miR-16 expression in the same samples as in (A) ($n = 3$ biological replicates for Δ HEL1, 2i and 2; $n = 2$ biological replicates for Δ HEL1-CM, 2i-CM and 2-CM). Expression of snRNA U6 was used as a loading control. (C) Western blot analysis of DICER, HA, AGO2, PKR and TRBP expression in the polyclonal cell lines NoDice Δ PKR FHA:DICER WT, N1, Δ HEL1, Δ HEL2i and Δ HEL2 ($n = 1$ biological replicate). Alpha-Tubulin was used as a loading control. (D) Northern blot analysis of miR-16 expression in the same samples as in (C) ($n = 2$ biological replicates). Expression of snRNA U6 was used as a loading control. (E) Mean (\pm SEM) of SINV-GFP viral titers in the polyclonal cell lines NoDice and NoDice Δ PKR FHA:DICER WT, N1, Δ HEL1, Δ HEL2i and Δ HEL2, infected at an MOI of 2 for 12 h (left) or 16 h (right) from plaque assay quantification ($n = 3$ biological replicates). Ordinary one-way ANOVA test with Dunnett's correction. ** $p < 0.01$; *** $p < 0.001$; ***** $p < 0.0001$; ns: non-significant. Source data are available online for this figure.

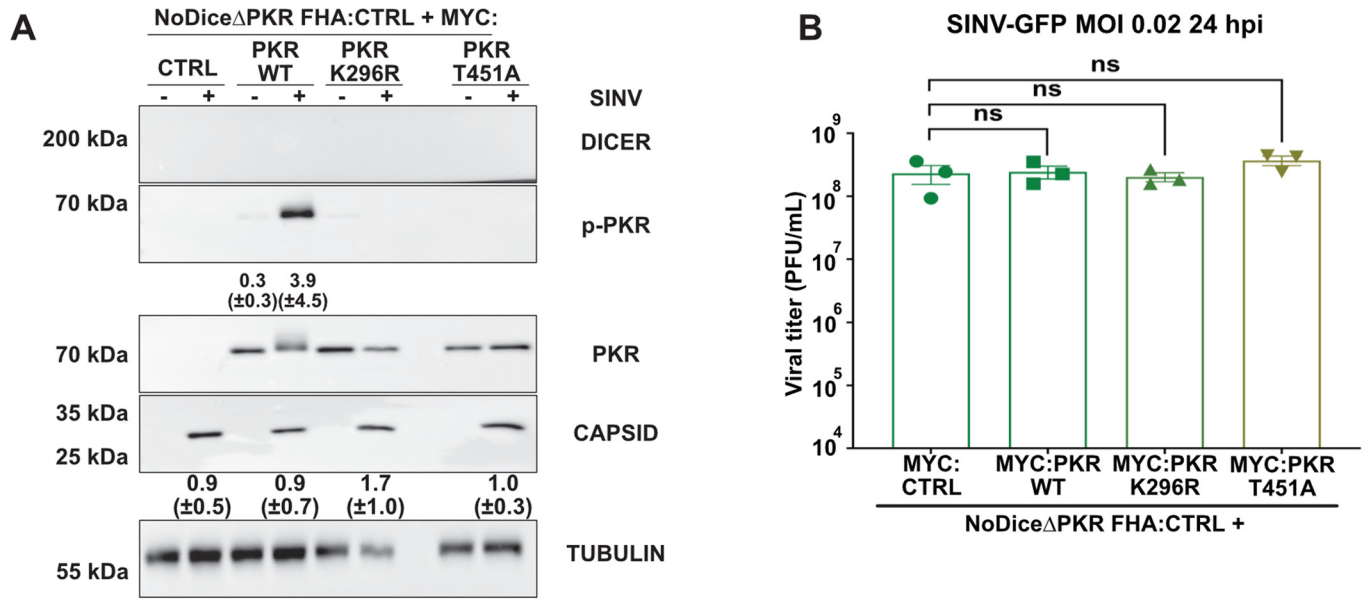


Figure EV3. PKR dimerization and/or catalytic activities do not change the infection outcome in NoDiceΔPKR FHA:CTRL cells.

(A) Western blot analysis of DICER, p-PKR, PKR and CAPSID expression in SINV-GFP infected NoDiceΔPKR FHA:CTRL cells expressing MYC:EMPTY CTRL vector, MYC:PKR, MYC K296R or MYC:T451A at an MOI of 0.02 for 24 h. Alpha-Tubulin was used as loading control. Band intensity was quantified and normalized to Tubulin for three independent biological replicates, then represented as mean (+/- SD) under the corresponding lane; p-PKR = p-PKR/PKR quantification. (B) Mean (+/- SEM) of SINV-GFP viral titers in the same samples as in (A), infected at an MOI of 0.02 for 24 h (n = 3 biological replicates) from plaque assay quantification. Ordinary one-way ANOVA test with Dunnett's correction. ns: non-significant. Source data are available online for this figure.

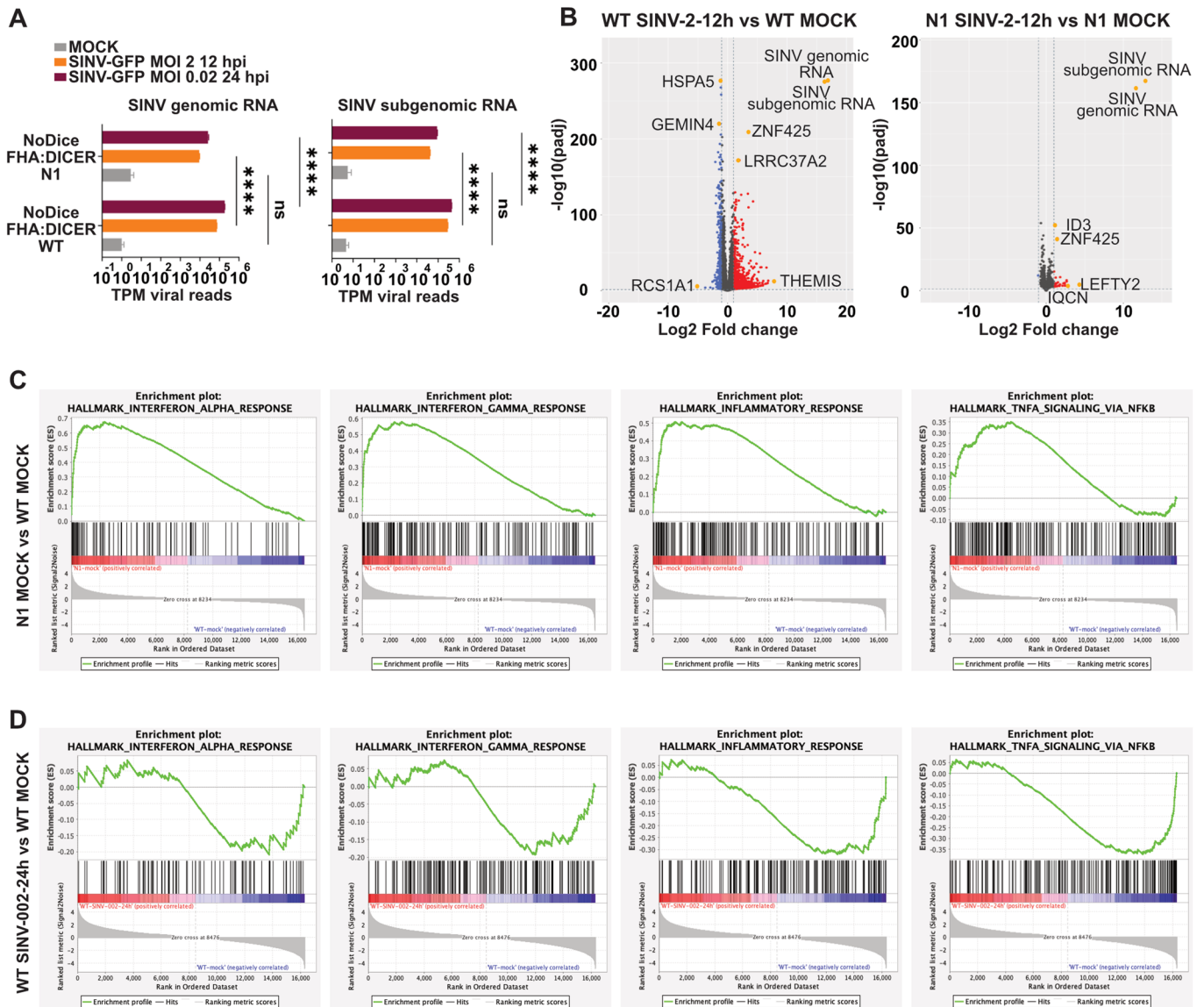


Figure EV4. Dicer N1 cells express a different gene set from Dicer WT cells.

(A) SINV-GFP genomic and subgenomic read distribution detected by RNA-sequencing analysis in NoDice FHA:DICER WT or N1 cells uninfected (Mock, gray) or infected at an MOI of 2 for 12 h (orange) or 0.02 for 24 h (purple). The viral reads number (mean \pm SEM) is normalized to the total mapped reads in each condition. TPM: transcripts per million. Ordinary two-way ANOVA test with Sidak's correction. **** $p < 0.0001$; ns: non-significant. $n = 3$ biological replicates, error bars represent SEM. (B) Volcano plots showing for each gene the log₂ fold change and adjusted p value (Wald test, DESeq2 package) between SINV-infected (MOI of 2 for 12 h) and mock NoDice FHA:DICER WT cells (left), or SINV-infected and mock NoDice FHA:DICER N1 cells (right). Each gene is marked as a dot (red: upregulated, blue: downregulated, gray: unchanged). The horizontal line denotes an adjusted p -value of 0.05 and the vertical ones the Log₂ fold change cut-offs (-1 and 1) ($n = 3$ biological replicates). (C,D) GSEA enrichment plots for selected biological states and processes linked to inflammatory and antiviral pathways for mock NoDice FHA:DICER N1 vs mock NoDice FHA:DICER WT (C), or SINV-infected (MOI of 0.02 for 24 h) vs. mock NoDice FHA:DICER WT cells (D). Source data are available online for this figure.

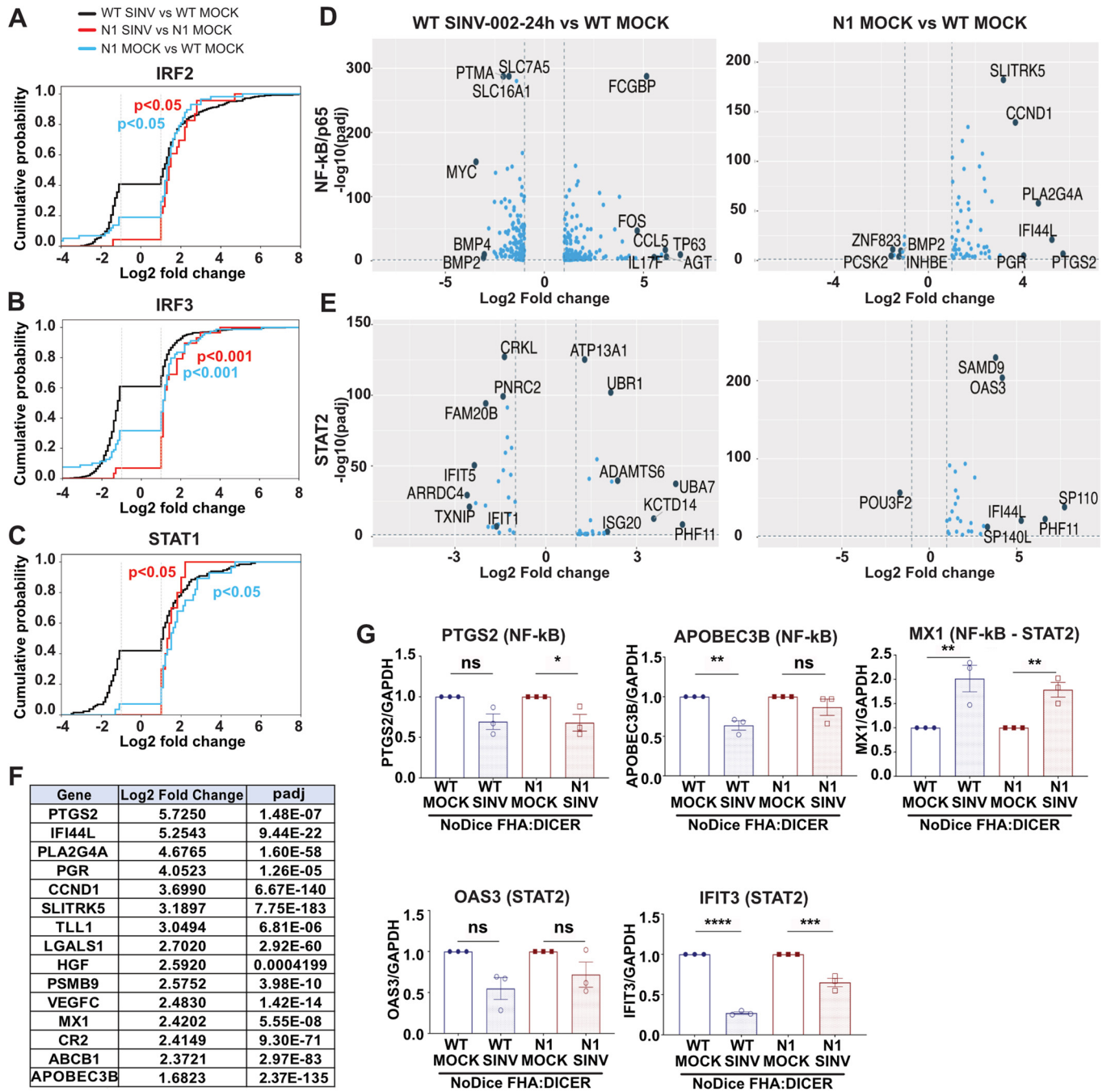


Figure EV5. Immune-related transcription factors activation is involved in the deregulation of Dicer N1 cells mRNAs.

(A-C) Histograms representing the cumulative probability of differentially expressed genes controlled by the transcription factors IRF2 (A), IRF3 (B) or STAT1 (C), plotted according to their Log2 fold change. The vertical lines stand for the Log2 fold change cut-offs (-1 and 1). The two-sample Kolmogorov-Smirnov test was used to assess whether each distribution was statistically different from the distribution of NoDice FHA:DICER WT cells infected with SINV vs. mock. *p*-values are indicated on each histogram. Black: WT SINV-002-24h vs WT MOCK; red: N1 SINV-002-24h vs N1 MOCK; blue: N1 MOCK vs WT MOCK. (D,E) Volcano plots for differentially expressed genes (DEGs) under the control of NF-κB/p65 (D) or STAT2 (E). Each gene is marked as a dot and plotted based on its log2 fold change and adjusted *p* values (Wald test, DESeq2 package) comparing SINV-infected (MOI of 0.02 for 24 h) vs mock NoDice FHA:DICER WT cells (left), or mock NoDice FHA:DICER WT vs mock NoDice FHA:DICER N1 cells (right). The horizontal line denotes an adjusted *p*-value of 0.05 and the vertical ones the Log2 fold change cut-offs (-1 and 1). *n* = 3 biological replicates. (F) Table of 15 representative upregulated NF-κB/p65 targets from the DEGs in the comparison mock NoDice FHA:DICER N1 vs mock NoDice FHA:DICER WT. Classification was made according to their Log2 fold change values. padj: adjusted *p*-value (Wald test, DESeq2 package). (G) RT-qPCR on selected DEGs controlled by either NF-κB/p65 or STAT2 in NoDice FHA:DICER WT and N1 cells infected or not with SINV-GFP at an MOI of 0.02 for 24 h. Mean (+/- SEM); *n* = 3 biological replicates. One-way ANOVA with Sidak's correction. **p* < 0.05; ***p* < 0.01; ****p* < 0.001; *****p* < 0.0001; ns: non-significant. Source data are available online for this figure.