

Supporting Information

for Adv. Sci., DOI 10.1002/advs.202303366

Mosaic RBD Nanoparticles Elicit Protective Immunity Against Multiple Human Coronaviruses in Animal Models

Yanjun Zhang, Jing Sun, Jian Zheng, Suxiang Li, Haiyue Rao, Jun Dai, Zhaoyong Zhang, Yanqun Wang, Donglan Liu, Zhao Chen, Wei Ran, Airu Zhu, Fang Li, Qihong Yan, Yiliang Wang, Kuai Yu, Shengnan Zhang, Dong Wang, Yanhong Tang, Banghui Liu, Linling Cheng, Jiandong Huo*, Stanley Perlman*, Jingxian Zhao* and Jincun Zhao*

Supporting Information

Mosaic RBD nanoparticles elicit protective immunity against multiple human coronaviruses in animal models

Yanjun Zhang ^{1#}, Jing Sun ^{1#}, Jian Zheng ^{2#}, Suxiang Li ^{1#}, Haiyue Rao ^{1#}, Jun Dai ^{3#}, Zhaoyong Zhang ¹, Yanqun Wang ¹, Donglan Liu ¹, Zhao Chen ¹, Wei Ran ¹, Airu Zhu ¹, Fang Li ¹, Qihong Yan ¹, Yiliang Wang ¹, Kuai Yu ¹, Shengnan Zhang ¹, Dong Wang ¹, Yanhong Tang ¹, Banghui Liu ⁴, Linling Cheng ¹, Jiandong Huo ^{1,5*}, Stanley Perlman ^{2*}, Jingxian Zhao ^{1*}, Jincun Zhao ^{1,5,6,7,8*}

¹State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, Guangzhou Institute of Respiratory Health, the First Affiliated Hospital of Guangzhou Medical University, Guangzhou, China.

²Department of Microbiology and Immunology, University of Iowa, Iowa City, Iowa 52242, USA.

³Guangzhou Customs District Technology Center, Guangzhou 510700, China.

⁴State Key Laboratory of Respiratory Disease, Guangdong Laboratory of Computational Biomedicine, Guangzhou Institutes of Biomedicine and Health, Chinese Academy of Sciences, Guangzhou, People's Republic of China.

⁵Guangzhou laboratory, Bio-island, Guangzhou, China.

⁶Institute of Infectious disease, Guangzhou Eighth People's Hospital of Guangzhou Medical University, Guangzhou, China.

⁷Institute for Hepatology, National Clinical Research Center for Infectious Disease, Shenzhen Third People's Hospital, the Second Affiliated Hospital, School of Medicine, Southern University of Science and Technology, Shenzhen, China. ⁸Shanghai Institute for Advanced Immunochemical Studies, School of Life Science and Technology, ShanghaiTech University, Shanghai, 201210, China.

[#]Co-first author, these authors contributed equally to this work.

*Corresponding authors.

Jincun Zhao, State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, Guangzhou Institute of Respiratory Health, the First Affiliated Hospital of Guangzhou Medical University, Guangzhou, China, email: zhaojincun@gird.cn

Jingxian Zhao, State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, Guangzhou Institute of Respiratory Health, the First Affiliated Hospital of Guangzhou Medical University, Guangzhou, China, email: zhaojingxian@gird.cn

Stanley Perlman Department of Microbiology and Immunology, University of Iowa, Iowa City, Iowa 52242, USA; Department of Pediatrics, University of Iowa, Iowa City, Iowa 52242, USA. Electronic address: <u>stanley-perlman@uiowa.edu</u>

Jiandong Huo, State Key Laboratory of Respiratory Disease, National Clinical Research Center for Respiratory Disease, Guangzhou Institute of Respiratory Health, the First Affiliated Hospital of Guangzhou Medical University, Guangzhou, China, email: huojiandong@gird.cn



Fig. S1. Related to Fig.1. Construction and characterization of mosaic-RBD nanoparticles. (A) DLS analysis in intensity for the Mosaic RBD-np. (B) the binding assay of Mosaic-RBD nanoparticles with the SARS-CoV-2, SARS-CoV and MERS-CoV RBD-specific monoclonal antibodies by BLI.

SARS-CoV-2 RBD-Ferritin>

SFTVEKGIYQTSNFR VQPTESIVRFPNITNLCPFGE VFNATRFAS VYAWNRKR ISNCVAD YSVLY NSASFSTFKC YGVSPTKLNDLCFTNVYADSF VIR GDE VRQIAP GQTGKIAD YNYKLPDDFTGC VI AWNSNNLDSK VGGNYNYLYR LFRKSNLKPFERD ISTE IYQ AGSTPCNG VE GFNCYFPLQS YGF QPTNG VGYQP YRVVVLSFELLHAPAT VCGPKKSSGGGSGGGESQ VRQQFSKD IIKLLNEQ VNK EMQSSNLYMSMSSW CYTHSLDGAGLF LFDHAAEE YE HAKKLIIFLNE NNVP VQLTSISAPE HKF EGLTQIFQKAYE HEQHISES INNIVD HAIKSKD HATFNFLQWYVAEQHEEE VLFKD ILDKIELIGNE NHGLYLADQYVKGIAKSRKS

SARS-CoV RBD-Ferritin>

NITNLCPFGE VFNATKFPS VYAWERKKISNCVAD YS VLYNSTFFSTFKC YG VSATKLNDLCFSNV YADSF VVKGDD VRQIAP GQTG VIAD YN YKLPDDFMGC VLAW NTR NID ATST GNYN YK YR YLR H GKLRPFERD ISN VPFSPDGKPCTPPALNC YW PLND YGFYTTTGIG YQ P YR VVVLSFELLNAPAT VSGGGSGGGESQ VRQQFSKDIIKLLNEQ VNKE MQSSNLYMSMSSW CYTHSLDGAGLFLFD HA AEE YE HAKKLIIFLNE NN VP VQLTSISAPE HKFEGLTQIFQKAYE HEQ HISESINNI VDHAIKSKDH ATFNFLQW YVAEQHEEE VLFKDILDKIELIGNEN HGLYLADQ YVKGIAKSRKS

MERS-CoV RBD-Ferritin>

QAEGVECDFSPLLSGTPPQVYNFKRLVFTNCNYNLTKLLSLFSVNDFTCSQISPAAIASNCYSSLI LDYFSYPLSMKSDLSVSSAGPISQFNYKQSFSNPTCLILATVPHNLTTITKPLKYSYINKCSRLLS DDRTEVPQLVNANQYSPCVSIVPSTVWEDGDYYRKQLSPLEGGGWLVASGSTVAMTEQLQMG FGITVQYGTDTNSVCPKLSGGGSGGGESQVRQQFSKDIIKLLNEQVNKEMQSSNLYMSMSSW CYTHSLDGAGLFLFDHAAEEYEHAKKLIIFLNENNVPVQLTSISAPEHKFEGLTQIFQKAYEHEQH ISESINNIVDHAIKSKDHATFNFLQWYVAEQHEEEVLFKDILDKIELIGNENHGLYLADQYVKGIAK SRKS

Table S1. Amino acid sequences for nanoparticles used in this study.



Fig. S2. Comparison of the neutralization titres against SARS-CoV-2 WT strain and its

variants. (A) Red and dark line represent the day 7 and day 21 after the boost immunization. All results are expressed as mean ± SEM. Statistical analyses were performed using two-way ANOVA. (B) Realted information about Fig.2.D-F (C) Divergence amino acids in the RBD domain of MERS-CoV Strains used in this study.

А

	Mean Neutralization titer (boost+7d)	Neutralization assay	Number of mice in this experiment
MERS-CoV EMC	3.34 (Log10)	Focus formation assay	3
MERS-CoV GD01	3.39 (Log ₁₀)	Pseudovirus assay	3
MERS-CoV Nigeria	3.30 (Log ₁₀)	Pseudovirus assay	3
SARS-CoV	3.80 (Log ₁₀)	Plaque formation assay	5
SARS-CoV-2 WT	4.52 (Log ₁₀)	Focus formation assay	3
SARS-CoV-2 Alpha	4.43 (Log ₁₀)	Focus formation assay	4
SARS-CoV-2 Beta	4.63 (Log ₁₀)	Focus formation assay	4
SARS-CoV-2 Delta	4.13 (Log ₁₀)	Focus formation assay	4
SARS-CoV-2 BA.1	3.83 (Log ₁₀)	Focus formation assay	4
SARS-CoV-2 BA.2	3.70 (Log ₁₀)	Focus formation assay	5
SARS-CoV-2 BA.5	3.32 (Log ₁₀)	Focus formation assay	6

MERS- <u>CoV</u> strains ↩	Divergence in the RBD	
MERS-CoV EMC/2012	L495, L588↩	
MERS- <u>CoV</u> ChinaGD01↩	L495, L588↩	
MERS- <u>CoV</u> Nigeria (Nig1675)⊲	F495, F588⇔	

Table S2. (A) relevant information about Fig. 2. (B) the RBD sequence divergence ofMERS CoV EMC, GD01 and Nigeria.



Fig. S3. Tfh and B cell gating strategy. Cells were gated as singlets and live cells on forward and side scatter and a live/dead FVS440 stain. (**A**) CD16/32 negative, CD4 positive cells were then gated on the expression of CXCR5 and PD-1. (**B**) CD45 and CD19 positive cells were gated, and then GC B cells were selected based on lower expression of CD38 and positive expression of GL7.



Fig. S4. Comparison of neutralization titres. (**A**) Comparison of neutralization titres between SARS-CoV-2 WT strain and different variants. (**B**) Comparison of neutralization titres between 7 days and 5 months after booster immunization for different strains. All results are expressed as mean ± SEM. Statistical analyses were performed using unpaired t test.



Fig. S5. Temperature change after immunization of RBD-np vaccine in cynomolgus monkeys and comparison of neutralization titres induced by immunization. (A) Temperature was monitored within 96 hours of primary and booster immunizations. (B) Comparison of the neutralization titres against SARS -CoV-2 WT strain and its variants. All results are expressed as mean ± SEM. Statistical analyses were performed using unpaired t test.



Fig. S6. Complete flow cytometry analysis for cross reactive-RBD⁺ GC B cells isolated from mice immunized with heterotypic mosaic-RBD nanoparticles or homotypic RBD nanoparticles (SARS-CoV RBD-np and SARS-CoV-2 RBD-np).