

b

CLUSTAL O(1.2.4) multiple sequence alignment

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Chymotrypsin      -----WQVSLQDKT--GFHF-----CGGSLINENWVVTAARCGVTTSDVV--      121
FAM111A           VGYLFWDSATTGYATCFVFKGLFILTCRHVIDSIVGDGIEPSKWATIIGQCVRVTFGYEE      416
                   *: : . : * *                               *..: .:*.. .:* . * .

Chymotrypsin      -----VAGEFDQGSSEKIQKLIKAKVFNKSNKYNLITNNRITLLKLSTAASFQ      171
FAM111A           LKDKETNYFFVEPWFEIHNEEL---DYAVLKLKENGQQVPMELYNGITPVPLSG-----      467
                   * *: ... . : * : *.: : : : *..* : **

Chymotrypsin      TVSAVCLPSASDDFAAGTTCVTTGWGLTRYTNANTPDRLLQ--ASLPLLSNTNCKKYWGT      229
FAM111A           LIHIIIGHPYGEKKQI--DACAIVPQGQRA---KKCQERVQSKKAESPEYVHMYTQRSFQK      522
                   : : * ..... :*.. * : : *:*.. *..* : : : .

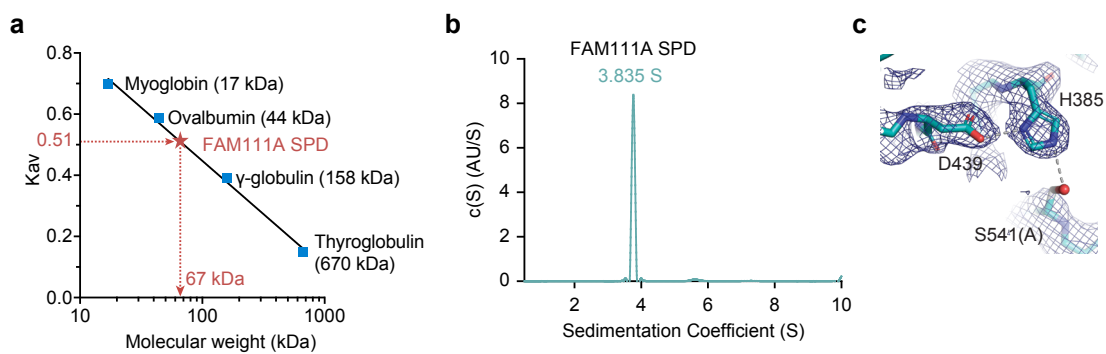
Chymotrypsin      KIKDAMICAGASGVSSCMGDSGGPLVCKKN-----GAWTL-----VGIVSWGSSSTCS      276
FAM111A           IVHNPDVI--TYDTEFFFGASGSPVFDKSGSLVAMHAAGFAYTYQNETRSIIIEFGSTMES      580
                   ::: : : ... :* **.*: .*. .... .*.:**.* *

Chymotrypsin      TSTPGVYARVTALVNWVQQLAAN 300
FAM111A           IL----LDIKQRHKPWYEEVFVN- 599
                   * :...

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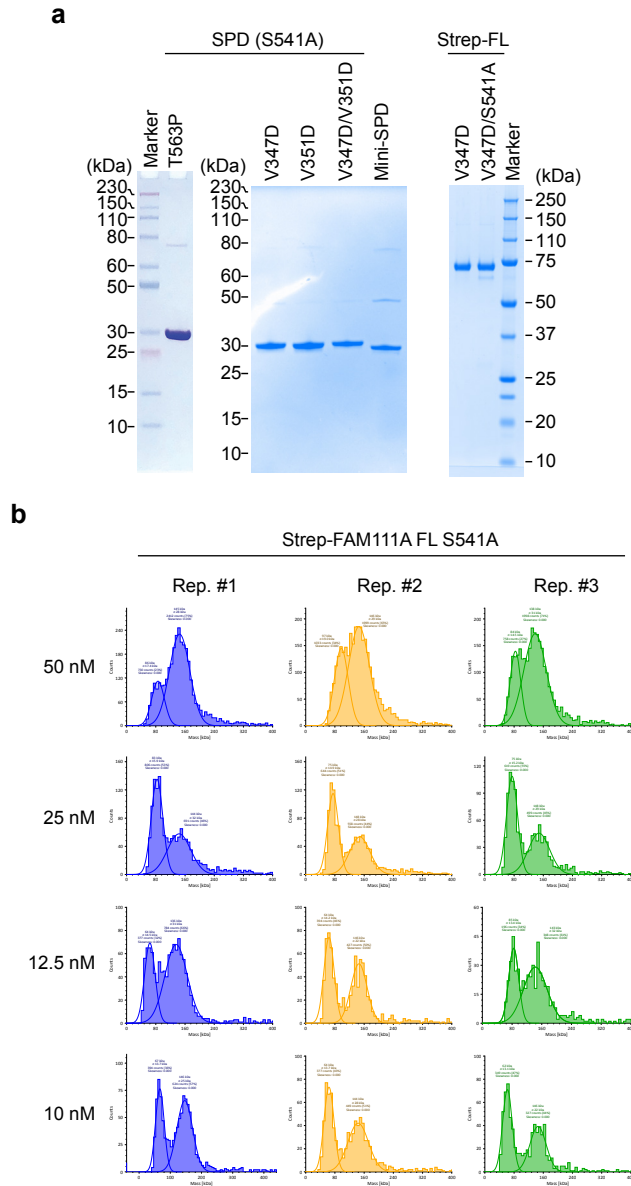
Supplementary Figure 1

(a) Slopes and specific activities at various enzyme concentrations in the assays shown in Fig. 1d. Slopes represent the change in relative fluorescence units (RFU) over time, and specific activity, representing slope values normalized by enzyme concentrations. (b) Amino acid sequence alignment of the human FAM111A SPD and bovine chymotrypsin generated by Clustal Omega. Asterisks (*): identical residue; colons (:): conserved substitution; period (.): semi-conserved substitution. Catalytic residues are indicated in red. Source data are provided as a Source Data file.



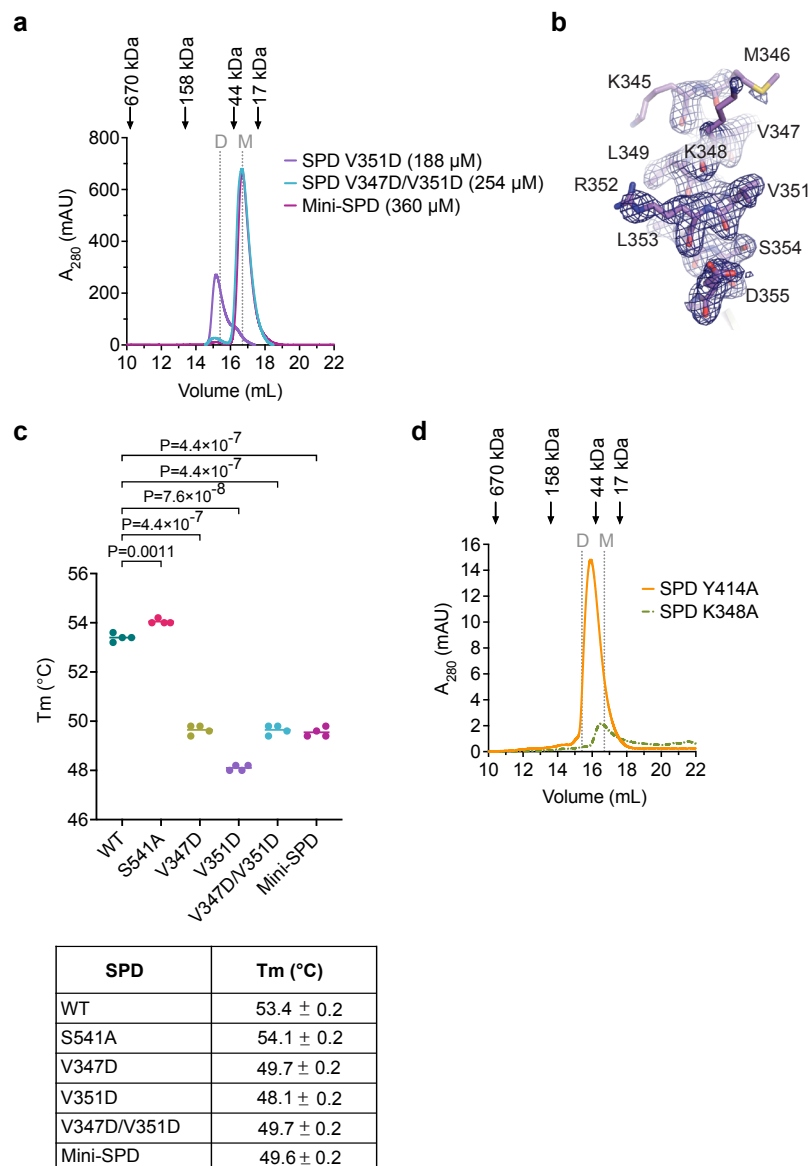
Supplementary Figure 2

(a) Calibration curve for the standard proteins on the Superdex 200 10/300 column. K_{av} values of the indicated standard proteins were plotted against their known molecular masses to generate a standard curve for molecular mass. The molecular mass of FAM111A SPD protein was estimated to be 67 kDa based on its K_{av} value of 0.51. K_{av} is defined as the ratio of the difference between the elution volume (V_e) and the void volume (V_o) to the difference between the total bed volume (V_t) and the void volume: $K_{av} = (V_e - V_o)/(V_t - V_o)$. (b) Analytical ultracentrifugation analysis demonstrates that purified FAM111A SPD is a dimer. The graph represents the sedimentation coefficient distribution $c(s)$ profile attained by sedimentation velocity (SV) data analysis. (c) Electron density map of the SPD active site. A 2mFo-dFc electron density map contoured at 1σ showing density for the catalytic triad of the SPD. A serine residue at position 541 is shown modelled (gray) along with the predicted hydrogen bond to H385. Source data are provided as a Source Data file.



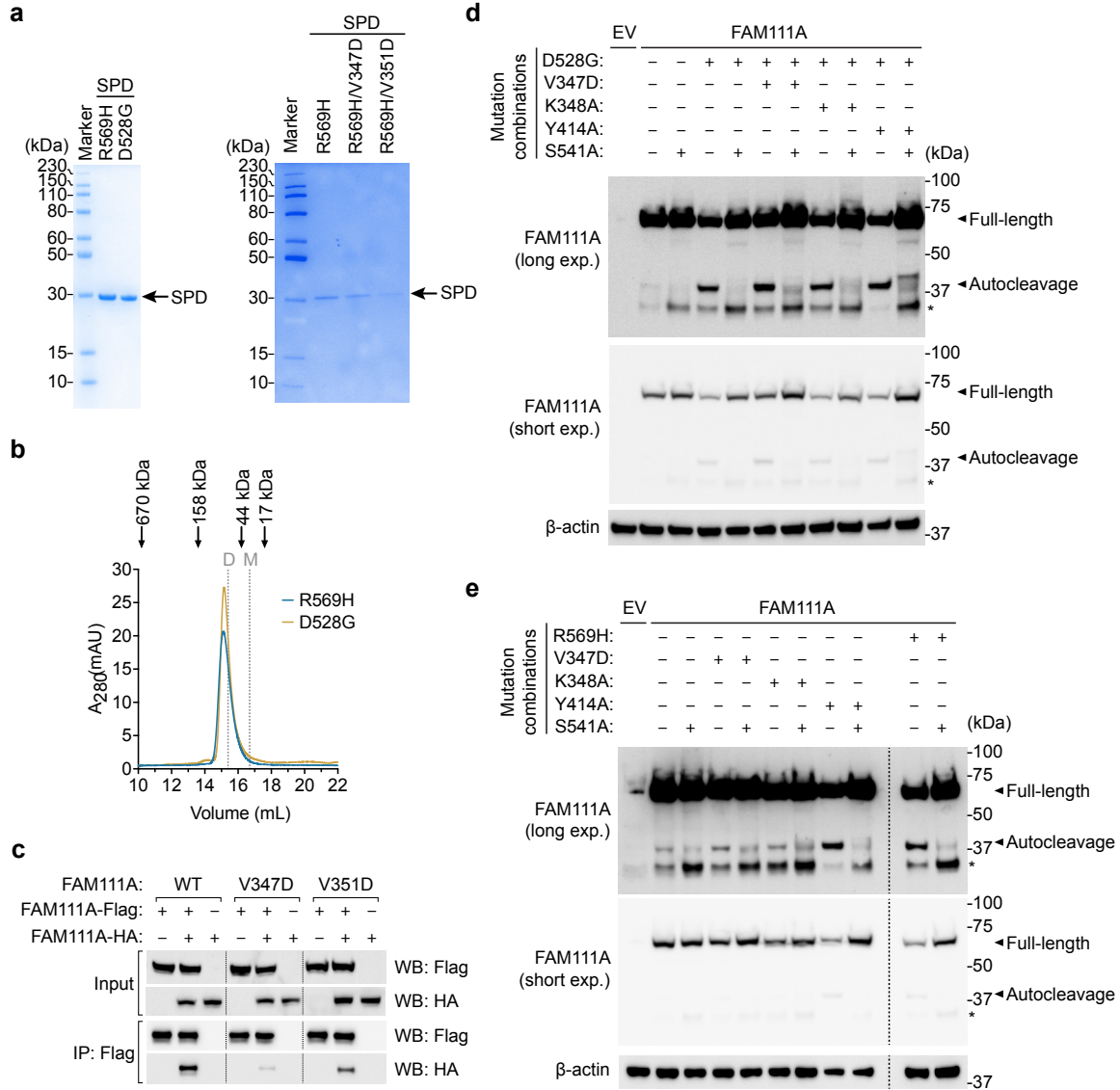
Supplementary Figure 4

(a) Purified recombinant proteins with various mutations analyzed by SDS-PAGE. The gel was stained with Coomassie Brilliant Blue. Positions of molecular weight marker proteins are indicated. (b) Mass distribution for Strep-FAM111A S541A obtained by mass photometry. Measurements were taken at indicated concentrations in three technical replicates (Rep. #1-3). Source data are provided as a Source Data file.



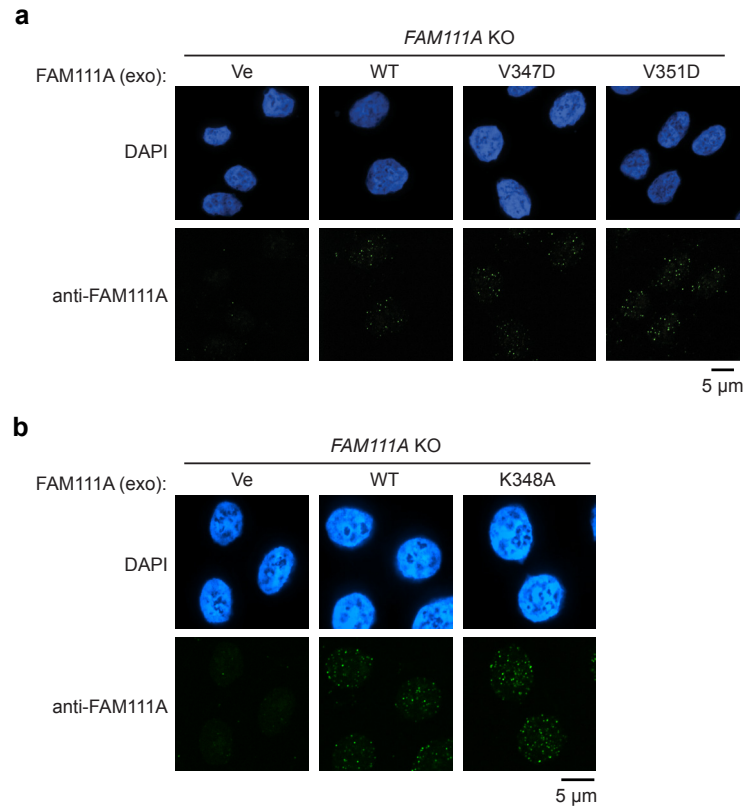
Supplementary Figure 5

(a) Size-exclusion chromatogram of high concentration monomeric mutants on Superdex 200 Increase 10/300 GL. Absorbance at 280 nm (A₂₈₀) is shown. Concentrations of injected proteins are shown in the legends. Elution volumes for SEC standard proteins are indicated with arrows on top. Expected theoretical elution volumes for an SPD monomer (31.5 kDa, indicated with a letter M) and an SPD dimer (63.0 kDa, indicated with a letter D) are shown with dotted lines. (b) Electron density map of helix α1 of mini-SPD. A 2mFo-dFc electron density map contoured at 1σ shows density for helix α1 backbone and most sidechains are visible. (c) Thermal stability of purified SPD and monomeric mutant proteins. The melting temperatures (T_m) of the indicated proteins were measured by thermal shift assays. Top: a graph showing the T_m values of individual measurements (line: mean). Significance of differences between mutants and WT was determined by two-tailed unpaired t-test. Bottom: Mean ± error of four replicates. Error is the higher value of the Instrument Limit of Error (ILE)=0.2°C and s.d. (d) Size-exclusion chromatogram of the Y414A and K348A mutants. Absorbance at 280 nm (A₂₈₀) is shown. Elution volumes for SEC standard proteins are indicated with arrows on top. Expected theoretical elution volumes for an SPD monomer (31.5 kDa, indicated with a letter M) and an SPD dimer (63.0 kDa, indicated with a letter D) are shown with dotted lines. Source data are provided as a Source Data file.



Supplementary Figure 6

(a) SDS-PAGE analysis of purified recombinant SPD proteins. The gel was stained with Coomassie Brilliant Blue. Positions of molecular weight marker proteins are indicated. (b) Size-exclusion chromatogram of the SPD R569H and D528G mutants. Absorbance at 280 nm (A₂₈₀) is shown. Elution volumes for SEC standard proteins are indicated with arrows on top. Expected theoretical elution volumes for an SPD monomer (31.5 kDa, indicated with a letter M) and an SPD dimer (63.0 kDa, indicated with a letter D) are shown with dotted lines. (c) Intermolecular self-interactions of FAM111A (WT, V347D, or V351D) assessed by co-immunoprecipitation. FAM111A proteins with either a Flag or an HA tag were coexpressed in 293T cells. Inputs (top) and anti-Flag precipitation samples (bottom) are analyzed by Western blotting (WB) using indicated antibodies. (d,e) FAM111A autocleavage. FAM111A proteins with indicated combinations of mutations were transiently expressed in 293T cells (which lacks endogenous FAM111A expression) and examined by Western blotting. Long (top) and short (middle) exposures of anti-FAM111A blots are shown. β -actin (bottom) is shown as a loading control. The positions of full-length and autocleavage bands are indicated by black arrowheads. *Nonspecific degradation bands. Source data are provided as a Source Data file.



Supplementary Figure 7

(a,b) Immunofluorescence microscope images of *FAM111A* foci on chromatin. The indicated *FAM111A* proteins were stably expressed exogenously (exo) by lentiviral vectors in *FAM111A* KO HAP1 cells. Cells were pre-extracted with 0.5% TX-100 before fixation with methanol:acetone (3:1) and costaining with DAPI (blue) and anti-*FAM111A* antibody (green, pseudo-colored from orange). Ve: empty vector. Images are single z slices. Scale bar, 5 μ m.

P1 residue:	FAM111A (20 μ M) Activity	FAM111A Rate Relative to P1=F	Chymotrypsin (1.7 μ M) Activity	Chymotrypsin Rate Relative to P1=F
R	-1.9 \pm 1.3	0		
V	0.0 \pm 0.2	0		
F	3990 \pm 30	1	5700 \pm 500	1
Y	2.6 \pm 0.5	6.5 $\times 10^{-4}$	90 \pm 10	1.6 $\times 10^{-2}$
W	10.2 \pm 0.9	2.6 $\times 10^{-3}$	190 \pm 30	3.3 $\times 10^{-2}$

Supplementary Table 1. Protease Specificity for the P1 Residue. Chymotrypsin or FAM111A were incubated with AMC-peptide substrates with the indicated residue in the P1 position and protease activity was monitored as an increase in AMC fluorescence. The rate from reactions containing catalytically inactive (S541A) SPD were used to correct for background in FAM111A assays, and buffer-only reactions were used to correct for background for reactions containing chymotrypsin. Substrate specificity defined as the rate relative to phenylalanine are shown. N=4, error bars \pm s.d. Experiments were repeated twice and similar results were obtained.

Supplementary Table 2. X-ray crystallographic data collection and refinement statistics

Structure	SPD S541A	Mini SPD
PDB Entry ID:	8S9K	8S9L
Data collection		
Space group	P 2 ₁ 2 ₁ 2 ₁	P 2 ₁ 2 ₁ 2 ₁
Cell dimensions		
<i>a</i> , <i>b</i> , <i>c</i> (Å)	74.81, 83.76, 191.12	68.46,70.03,128.02
α , β , γ (°)	90, 90, 90	90, 90, 90
Resolution (Å)	50-2.7 (2.8-2.7)	70-1.85 (1.92-1.85)
<i>R</i> _{sym} or <i>R</i> _{merge}	0.026 (1.37)	0.061 (1.79)
<i>I</i> / σ <i>I</i>	8.3 (1.4)	32.2 (1.3)
Completeness (%)	99.8 (100)	99.8 (100)
Redundancy	6.8 (7.1)	6.6 (6.9)
Refinement		
Resolution (Å)	50-2.7	50-1.85
No. reflections	32707	53161
<i>R</i> _{work} / <i>R</i> _{free}	0.196/ 0.249	0.218/0.252
No. atoms		
Protein	8279	3799
Ligand/ion	6	5
Water	21	90
<i>B</i> -factors (Å ²)		
Protein	65.2	67.8
Ligand/ion	60.9	134
Water	50.8	56.4
R.m.s. deviations		
Bond lengths (Å)	0.002	0.012
Bond angles (°)	0.52	1.10

*Values in parentheses are for the highest-resolution shell (10% of reflections).