

# Studies on Molybdenum Absorption and Transport in Bean and Rice

Received for publication December 9, 1977 and in revised form March 10, 1978

SESHADRI KANNAN AND SARADHA RAMANI

Biology and Agriculture Division, Bhabha Atomic Research Centre, Bombay 400 085, India

## ABSTRACT

The patterns of molybdenum ( $\text{MoO}_4^{2-}$ ) absorption and transport were investigated in intact bean (*Phaseolus vulgaris* L.) and rice (*Oryza sativa* L. cv. I.R.8) plants. The mobility of  $\text{MoO}_4^{2-}$  absorbed by roots and by leaves was compared with that of a freely mobile element,  $\text{Rb}^+$ . Although  $\text{MoO}_4^{2-}$  absorption by bean roots was nearly as high as that of  $\text{Rb}^+$ , its transport to the shoot was considerably less. When  $\text{MoO}_4^{2-}$  was fed to one of the primary leaves, most of it was transported to the stem and root. Evidence obtained here showed that  $\text{MoO}_4^{2-}$  was mobile. Experiments with intact rice seedlings revealed large differences in the absorption and transport of  $\text{MoO}_4^{2-}$  between the plants grown in  $\text{CaSO}_4$  and those in Hoagland solution. Molybdate uptake by excised rice roots was suggested to be an active process since it was greatly inhibited by a metabolic inhibitor. The presence of  $\text{Mn}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Cl}^-$ , or  $\text{SO}_4^{2-}$  in the absorption medium reduced  $\text{MoO}_4^{2-}$  uptake which was markedly enhanced by the presence of  $\text{Fe}^{2+}$ .

Molybdenum is one of the micronutrient elements essential for plants. Following the early work of Arnon and Stout (1) and Piper (16), the occurrence of its deficiency has been recorded in many soils and crop plants, and remedial measures like soil, foliar, or seed treatment have been widely advocated (7, 17).

Several studies have been made in the past years relating to the mechanisms of absorption and transport of major nutrients and also some micronutrient elements (5, 12). However, information on the mechanisms of micronutrient absorption is still inadequate, and Moore (13) has rightly emphasized the need for a systematic investigation to understand the manner in which the micronutrients, especially boron and molybdenum, are absorbed by plant systems and also the factors which control their absorption and transport. Molybdate absorption and transport have been studied in intact plants and excised roots, and the results are reported here.

## MATERIALS AND METHODS

Experiments on the absorption and transport were carried out with bean plants and excised roots or intact seedlings of rice. Beans (*Phaseolus vulgaris* L.) were germinated in sand for 5 days, transferred to 0.5 Hoagland solution No. 1 (8) minus  $\text{MoO}_4^{2-}$  and grown for 7 to 9 days under a 12-hr photoperiod (500 ft-c) at 25 C. Rice (*Oryza sativa* L. cv. I.R.8) seeds were germinated in trays over 0.5 Hoagland solution minus  $\text{MoO}_4^{2-}$  for 7 days, and the roots were excised for use.

The absorption medium consisted of  $^{86}\text{Rb}$ - or  $^{99}\text{Mo}$ -labeled 0.01 mM  $\text{RbCl}$  or ammonium molybdate  $[(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}]$  adjusted and maintained at pH 5.5 with  $\text{KOH}$  twice during the absorption. Unless otherwise stated, the same but nonradioactive cold (5 C) solutions were used for desorption of the roots at the

end of the experimental period. In one experiment, the absorption and transport of  $\text{MoO}_4^{2-}$  were compared with those for  $\text{Rb}^+$ . The roots of bean plants were exposed to the absorption medium and removed at different times. Foliar absorption and transport of  $\text{MoO}_4^{2-}$  and  $\text{Rb}^+$  were also measured in another set of bean plants. The isotope solutions (0.2 ml of 0.01 mM  $\text{RbCl}$  or ammonium molybdate) were placed in a plastic ring 1 cm in diameter (15) which was fixed in the intercostal region on the upper surface of a primary leaf. The transport to different parts was assayed after punching out and discarding the leaf area below the plastic ring.

The patterns of absorption of  $\text{MoO}_4^{2-}$  by root and transport to shoot were examined in rice seedlings grown in three different media, viz. 0.1 mM  $\text{CaSO}_4$ , 0.5 Hoagland or full Hoagland solutions without  $\text{MoO}_4^{2-}$  for 15 days. For measuring the absorption and transport, sets of 10 seedlings were placed in flasks containing 200 ml of the absorption medium, and removed at different time intervals during a 6-hr period.

A series of three experiments was carried out with excised rice roots. For studying the action of a metabolic inhibitor on  $\text{MoO}_4^{2-}$  absorption, excised roots were suspended in the absorption medium at pH 7.4, with or without  $10^{-6}$  M  $p\text{CF}_3\text{O-CCP}$ ,<sup>1</sup> and roots (500 mg fresh wt) were removed for assay. In another set, the influence of other ions on  $\text{MoO}_4^{2-}$  absorption was examined by suspending the rice roots in the absorption medium containing 0.01 or 0.1 mM  $\text{FeSO}_4$ ,  $\text{MnSO}_4$ ,  $\text{CuSO}_4$ ,  $\text{ZnSO}_4$ ,  $\text{NH}_4\text{Cl}$ , or  $(\text{NH}_4)_2\text{SO}_4$  for 4 hr. The absorption of  $\text{MoO}_4^{2-}$  in the presence of 0.01 or 0.1 mM  $\text{FeSO}_4$  was further investigated with respect to different washing medium. The samples were allowed to absorb  $\text{MoO}_4^{2-}$  for 4 hr, and then desorbed for 30 min in solutions containing 0.1 mM ammonium molybdate alone, or in combination with either 0.1 mM  $\text{FeSO}_4$  or  $\text{FeEDDHA}$ .

In general, the roots were desorbed for 30 min. Low amounts of 0.1 mM  $\text{CaSO}_4$  or  $\text{CaCl}_2$  were routinely included in all of the solutions. The plant parts were dried and radioassayed in a gamma ray spectrometer and the results presented are the means of values obtained for five bean plants, three sets of rice seedlings, and five sets of excised rice roots on a fresh wt basis.

## RESULTS

Molybdate absorption by roots of intact bean seedlings and transport to the stem, primary and trifoliate leaves were measured during a 24-hr period and compared with those for  $\text{Rb}^+$  (Fig. 1). Molybdate absorption by root is slightly less than that of  $\text{Rb}^+$ , although its transport is considerably less than that for  $\text{Rb}^+$ . The transport to the trifoliate is the largest, followed by stem and primary leaves in both  $\text{Rb}^+$  and  $\text{MoO}_4^{2-}$ . Furthermore, the absorption and transport of both  $\text{Rb}^+$  and  $\text{MoO}_4^{2-}$  follow a biphasic pattern, with a second phase beginning approximately at 6 hr. The rate of transport of  $\text{Rb}^+$  to the trifoliate nearly runs parallel to its

<sup>1</sup> Abbreviations:  $p\text{-CF}_3\text{O-CCP}$ :  $p$ -trifluoro-methoxyphenylhydrazone;  $\text{FeEDDHA}$ : ferric ethylenediaminedi( $\alpha$ -hydroxyphenyl acetate).

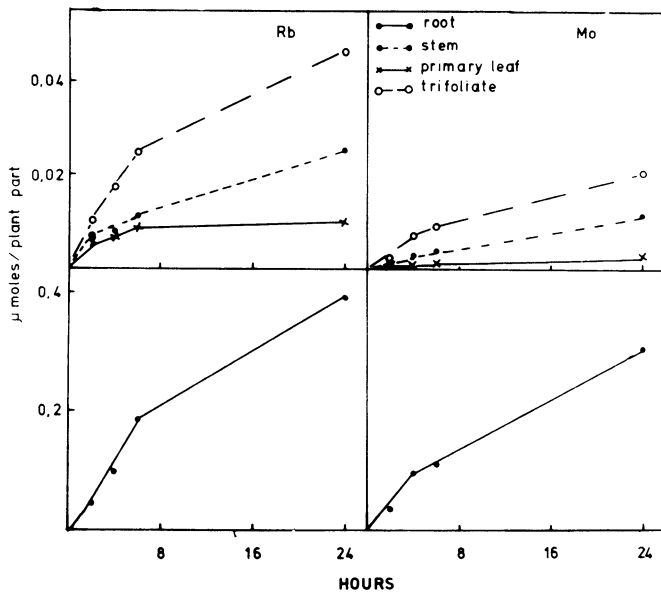


FIG. 1. Course of  $\text{MoO}_4^{2-}$  and  $\text{Rb}^+$  absorption by roots and transport to different parts of intact bean seedlings during 24 hr. The absorption medium contained 0.01 mM  $\text{RbCl}$  or ammonium molybdate.

Table I. Foliar Absorption and Translocation of  $\text{MoO}_4^{2-}$  and  $\text{Rb}^+$  in Bean Plants

The isotope solution, 0.2 ml of 0.01 mM Ammonium molybdate or  $\text{RbCl}$  was supplied to one of the primary leaves by means of a plastic ring and after 24 hr the leaf area with the ring was punched out and discarded. The amount transported to the remaining part of the applied leaf and other parts is estimated and given as absolute amounts per plant part.

Plant Parts	$\text{MoO}_4^{2-}$		$\text{Rb}^+$	
	nmoles $\times 10^2$ $\pm$ SE	Distribution as % of total	nmoles $\times 10^2$ $\pm$ SE	Distribution as % of total
Applied Primary Leaf	6.04 $\pm$ 1.4	17.16	3.83 $\pm$ 0.1	13.03
Opposite Primary Leaf	6.65 $\pm$ 1.1	18.89	2.99 $\pm$ 1.2	10.16
Trifoliolate Leaf	2.76 $\pm$ 0.7	7.84	5.90 $\pm$ 0.1	20.05
Stem	10.80 $\pm$ 0.7	30.68	4.32 $\pm$ 1.2	14.68
Root	8.95 $\pm$ 0.1	25.43	12.38 $\pm$ 3.1	42.08

absorption by root while this is not so for  $\text{MoO}_4^{2-}$ . Transport of  $\text{Rb}^+$  and  $\text{MoO}_4^{2-}$  to the primary leaves reached a near maximum at 6 hr and remained at the same level up to 24 hr.

The data on foliar absorption and transport reveal that a considerable amount of  $\text{MoO}_4^{2-}$  is absorbed and transported within the plant. The total amount absorbed after discarding the site of isotope application is 35.23 and 22.90 nmol/plant for  $\text{MoO}_4^{2-}$  and  $\text{Rb}^+$ , respectively (Table I). The pattern of translocation of  $\text{MoO}_4^{2-}$  is different from that of  $\text{Rb}^+$ . While  $\text{MoO}_4^{2-}$  and  $\text{Rb}^+$  are nearly equally distributed between the applied and opposite primary leaves, the transport of  $\text{MoO}_4^{2-}$  is largely downward to the stem and root, and  $\text{Rb}^+$  is transported equally to the growing regions, namely trifoliolate leaves and the root. Transport of  $\text{MoO}_4^{2-}$  to shoot is much less than to other parts.

The absorption and transport of  $\text{MoO}_4^{2-}$  in intact rice seedlings are influenced by the nutrient media used for their culture. The absorption is the lowest in the plants grown in  $\text{CaSO}_4$  while it is higher in those raised in Hoagland solution (Fig. 2). In contrast, the transport in plants grown in  $\text{CaSO}_4$  was higher than in those raised in Hoagland solution. The absorption and transport in plants grown in 0.5 and full Hoagland solution did not differ very much.

Studies with excised rice roots show that  $\text{MoO}_4^{2-}$  absorption is sensitive to the metabolic inhibitor pCF<sub>3</sub> O-CCP (Fig. 3). Furthermore, the presence of  $\text{Cu}^{2+}$ , chloride, and sulfate inhibited the

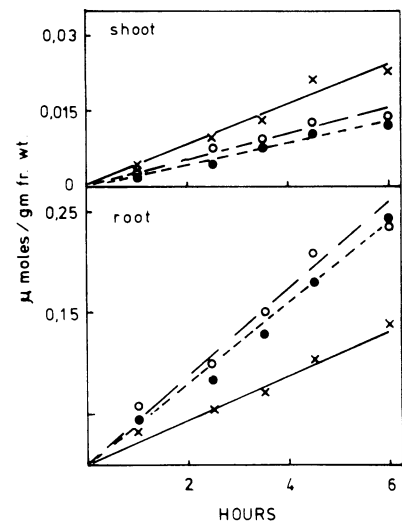


FIG. 2. Absorption and transport of  $\text{MoO}_4^{2-}$  from 0.01 mM ammonium molybdate in rice seedlings grown in 0.1 mM  $\text{CaSO}_4$  (x—x), 0.5 Hoagland (o—o), and full Hoagland (●—●) nutrient medium. The values are the means of three sets of 10 seedlings each.

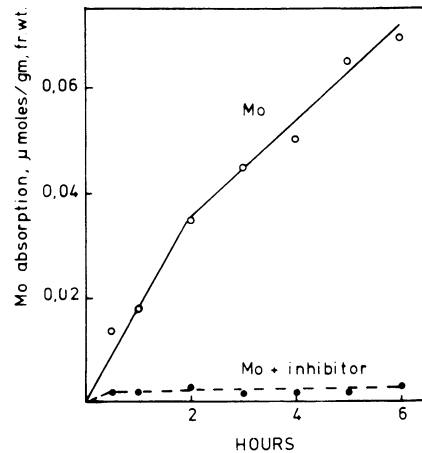


FIG. 3. Effect of  $10^{-6}$  M pCF<sub>3</sub> O-CCP on the absorption of  $\text{MoO}_4^{2-}$  from 0.01 mM ammonium molybdate by excised rice roots.

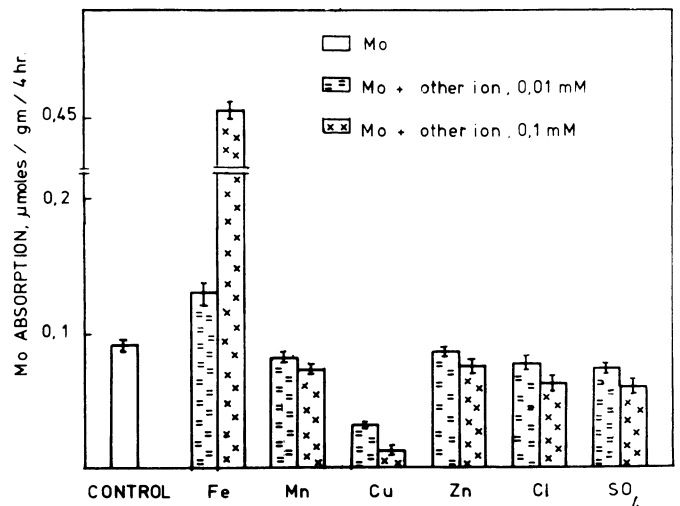


FIG. 4. Absorption of  $\text{MoO}_4^{2-}$  from 0.01 mM ammonium molybdate by excised rice roots, in the absence and presence of 0.01 or 0.1 mM  $\text{FeSO}_4$ ,  $\text{MnSO}_4$ ,  $\text{CuSO}_4$ ,  $\text{ZnSO}_4$ ,  $\text{NH}_4\text{Cl}$ , and  $(\text{NH}_4)_2\text{SO}_4$ .

Table II. Effects of FeSO<sub>4</sub> and FeEDDHA on the Absorption of MoO<sub>4</sub><sup>2-</sup> from 0.01 mM Molybdate by Excised Rice Roots

Excised roots were suspended in <sup>99</sup>Mo labelled 0.01 mM Ammonium molybdate in the absence and presence of FeSO<sub>4</sub> or FeEDDHA for 4 hr, and samples were allowed to desorb for 30 min in solutions containing molybdate alone, or in combination with FeSO<sub>4</sub> or FeEDDHA.

Absorption Medium	Desorption Medium		
	0.1 mM Molybdate nmoles/g ± SE	0.1 mM Molybdate + 0.1 mM FeSO <sub>4</sub> nmoles/g ± SE	0.1 mM Molybdate + 0.1 mM FeEDDHA nmoles/g ± SE
0.01 mM Molybdate	54.91 ± 4.08	32.07 ± 1.92	49.98 ± 3.49
0.01 mM Molybdate + 0.01 mM FeSO <sub>4</sub>	51.26 ± 3.16	53.98 ± 1.80	48.83 ± 0.99
0.01 mM Molybdate + 0.1 mM FeSO <sub>4</sub>	337.45 ± 22.55	334.45 ± 11.68	292.27 ± 25.14
0.01 mM Molybdate + 0.01 mM FeEDDHA	58.10 ± 5.03	50.20 ± 4.63	50.96 ± 2.72
0.01 mM Molybdate + 0.1 mM FeEDDHA	58.32 ± 4.09	46.89 ± 3.86	59.52 ± 3.30

absorption (Fig. 4). There is also a less marked reduction in the uptake in the presence of Mn<sup>2+</sup> and Zn<sup>2+</sup>. The presence of low concentrations of FeSO<sub>4</sub> (0.01 mM) enhanced MoO<sub>4</sub><sup>2-</sup> uptake and this enhancement was even greater at high concentrations (0.1 mM). Additional experiments revealed that this enhancement occurred in the presence of 0.1 mM FeSO<sub>4</sub> and not FeEDDHA (Table II). When the roots treated with 0.01 mM FeSO<sub>4</sub> were desorbed with high concentrations of ammonium molybdate (0.1 mM) no enhancement of MoO<sub>4</sub><sup>2-</sup> absorption was observed. However, FeSO<sub>4</sub> increased the uptake and this is also observed when the desorption medium contained molybdate and FeSO<sub>4</sub> (0.1 mM). High concentrations of FeSO<sub>4</sub> (0.1 mM) enhanced MoO<sub>4</sub><sup>2-</sup> absorption and there was no effect of the presence of FeSO<sub>4</sub> or FeEDDHA in the desorption medium.

## DISCUSSION

Bukovac and Wittwer (4) investigated the mobility of several nutrient elements by leaf disc removal technique and placed Fe<sup>2+</sup>, Mn<sup>2+</sup>, and MoO<sub>4</sub><sup>2-</sup> in the partially mobile group. However, others reported that Fe<sup>2+</sup> (3) and Mn<sup>2+</sup> (10) were absorbed by roots and readily translocated to the shoot. We have examined the mobility of molybdenum as an element in comparison with a freely mobile element, rubidium, and it is not our interest to study the mechanisms of uptake of molybdate which enters the plant as anion. Molybdate transport from the root to shoot was less than that of Rb<sup>+</sup>, despite the fact their absorption by the roots was nearly the same (Fig. 1). Molybdate supplied to the primary leaf was translocated to other parts, although most of it was in the stem and root. Differences occurred between MoO<sub>4</sub><sup>2-</sup> and Rb<sup>+</sup> only in the per cent transport to the trifoliolate leaves (Table I). The absorption and transport of a moderately mobile element Fe<sup>2+</sup> were studied earlier (11), and although the experimental techniques differed, some meaningful comparison could still be made between the mobility of Fe<sup>2+</sup> and MoO<sub>4</sub><sup>2-</sup>. Only 0.06 μmol out of 4.2 μmol of Fe<sup>2+</sup> absorbed by bean roots was translocated to shoot, while 0.002 μmol was transported to other parts out of 0.3 μmol of Fe<sup>2+</sup> absorbed by the primary leaf. In comparison with Fe<sup>2+</sup>, MoO<sub>4</sub><sup>2-</sup> appears to be more mobile.

The patterns of absorption and transport of nutrient elements were much altered by the medium of growth of the plants (9, 10). Molybdate absorption by rice roots grown in Hoagland solution was greater than in those raised in CaSO<sub>4</sub> (Fig. 2). It is possible that MoO<sub>4</sub><sup>2-</sup> absorption in seedlings grown in Hoagland solution

relates to the entry into vacuole, on the basis of the argument that cytoplasmic sites get saturated during growth in nutrient medium (9). However, more precise experiments are needed to confirm this explanation.

The absorption of MoO<sub>4</sub><sup>2-</sup> is influenced by the presence of both cations and anions, which are all inhibitory, with the exception of Fe<sup>2+</sup> (Fig. 4). It is rather surprising that Fe<sup>2+</sup> is found to enhance the absorption, and this effect is great when FeSO<sub>4</sub> is 0.1 mM. It is equally interesting that this enhancement is consistently absent when FeEDDHA is used instead of FeSO<sub>4</sub>. Molybdate may possibly combine with Fe<sup>2+</sup> dissociated from FeSO<sub>4</sub> and enter as iron molybdate. This does not, however, offer an explanation of why more MoO<sub>4</sub><sup>2-</sup> is absorbed. The effect of Fe<sup>2+</sup> on MoO<sub>4</sub><sup>2-</sup> absorption appears to be a direct one. Berry and Reisenauer (2) studied the influence of MoO<sub>4</sub><sup>2-</sup> on the uptake and distribution of Fe<sup>2+</sup> by tomato, and found that MoO<sub>4</sub><sup>2-</sup> enhanced the plant's ability to absorb Fe<sup>2+</sup>. Gerloff *et al* (6) reported that MoO<sub>4</sub><sup>2-</sup> and Mn<sup>2+</sup> were capable of affecting Fe<sup>2+</sup> availability in tomato. These authors have examined the effects of MoO<sub>4</sub><sup>2-</sup> on the absorption of cations and anions. Stout and Meagher (18) obtained evidence for increased translocation of MoO<sub>4</sub><sup>2-</sup> by phosphate in culture solution. Furthermore, they found from radioautographic studies that the distribution of molybdenum in the leaves was distinctly different from that of potassium. In the review of interactions between MoO<sub>4</sub><sup>2-</sup> and other cations and anions (14) mention has been made of the enhancement of MoO<sub>4</sub><sup>2-</sup> deficiency by Cu<sup>2+</sup> and the inhibition of MoO<sub>4</sub><sup>2-</sup> uptake by sulfate. Our studies, on the other hand, reveal the effects of both cations Mn<sup>2+</sup>, Zn<sup>2+</sup>, and Cu<sup>2+</sup> and anions Cl<sup>-</sup> and SO<sub>4</sub><sup>2-</sup> on the absorption of MoO<sub>4</sub><sup>2-</sup>.

## LITERATURE CITED

- ARNON DI, PR STOUT 1939 Molybdenum as an essential element for higher plants. *Plant Physiol* 14: 599-602
- BERRY JA, HM REISENAUER 1967 The influence of molybdenum on iron nutrition of tomato. *Plant Soil* 27: 303-313
- BROWN, AL, S YAMAGUCHI, J LEAL-DIAZ 1965 Evidence for translocation of iron in plants. *Plant Physiol* 40: 35-38
- BUKOVAC MJ, SH WITTWER 1957 Absorption and mobility of foliar applied nutrients. *Plant Physiol* 32: 428-435
- EPSTEIN E 1972 *Mineral Nutrition of Plants: Principles and Perspectives*. John Wiley & Sons, New York
- GERLOFF GC, PR STOUT, LHP JONES 1959 The micronutrient cations, iron, manganese, zinc and copper: their uptake by plants from the absorbed state. *Soil Sci* 72: 47-65
- HAGSTROM GR, KC BERGER 1963 Molybdenum status of three Wisconsin soils and its effect on four legume crops. *Agron J* 55: 399-401
- HOAGLAND DR, DI ARNON 1950 The water-culture method for growing plants without soil. *Calif Agric Exp Sta Circ* 347, 32 pp
- HOORMANS JJM 1974 Role of cell compartments in the redistribution of K<sup>+</sup> and Na<sup>+</sup> ions absorbed by the roots of intact barley plants. *Z Pflanzenphysiol* 73: 234-242
- KANNAN S, H KEPPEL 1976 Intracellular regulation of absorption and transport of Fe and Mn in wheat seedlings, cultivated in low and high salt media. *Z Pflanzenphysiol* 79: 132-142
- KANNAN S, T MATHEW 1970 Effects of growth substances on the absorption and transport of iron in plants. *Plant Physiol* 45: 206-209
- MAAS EV, DP MOORE, BJ MASON 1968 Manganese absorption by excised barley roots. *Plant Physiol* 43: 527-530
- MOORE DP 1971 Mechanisms of micronutrient uptake by plants. In JJ Mortvedt, PM Giordana, WL Lindsay, eds, *Micronutrients in Agriculture*. Soil Sci Soc America, Madison, Wisconsin, pp 171-192
- OLSEN SR 1971 Micronutrient interactions. In JJ Mortvedt, PM Giordano, WL Lindsay, eds, *Micronutrients in Agriculture*. Soil Sci Soc America, Madison, Wisconsin, pp 243-264
- PANDEY DP, S KANNAN 1976 Action of ABA and some cytokinins on the transport of foliar and root absorbed Rb<sup>+</sup> and Fe<sup>2+</sup> in bean plants. *Z Pflanzenphysiol* 78: 95-102
- PIPER CS 1940 Molybdenum as an essential element for plant growth. *J Aust Inst Agric Sci* 6: 112-114
- REISENAUER HM 1963 Relative efficiency of seed and soil applied molybdenum fertilizer. *Agron J* 55: 459-460
- STOUT PR, WR MEAGHER 1948 Studies of the molybdenum nutrition of plants with radioactive molybdenum. *Science* 108: 471-473