

Supplementary Materials for
**Engineered MgO nanoparticles for cartilage-bone synergistic
therapy**

Liming Zheng *et al.*

Corresponding author: Yifeng Zhang, zhangyf3@shanghaitech.edu.cn; Hui Wei,
weihui@nju.edu.cn; Qing Jiang, qingj@nju.edu.cn

Sci. Adv. **10**, eadk6084 (2024)
DOI: 10.1126/sciadv.adk6084

This PDF file includes:

Figs. S1 to S11
Tables S1 to S5

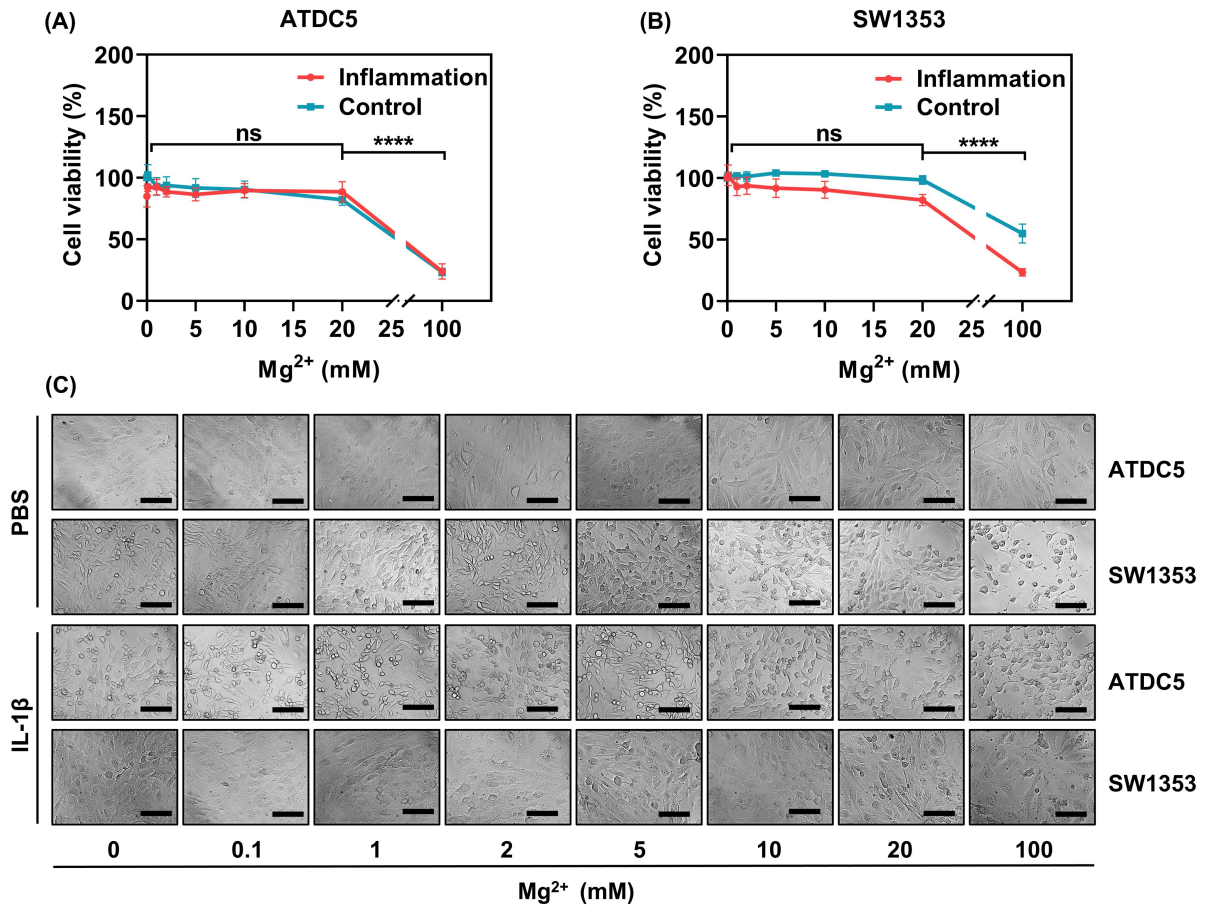


Fig. S1. Cell viability of Mg²⁺ in different kinds of cell lines.

Cell viability of ATDC5 (A) and SW1353 (B) cells after treatment with gradient concentrations of Mg²⁺ with/without inflammatory stimulation. (C) Representative microscopy images of ATDC5 and SW1353 cells after treatment with gradient concentrations of Mg²⁺ with/without inflammatory stimulation.

Notes: Scale Bar: 100 μm. All data are the mean ± s.d. Statistical differences between groups were determined by one-way ANOVA analysis. ****P<0.0001, ns P>0.05. n=3.

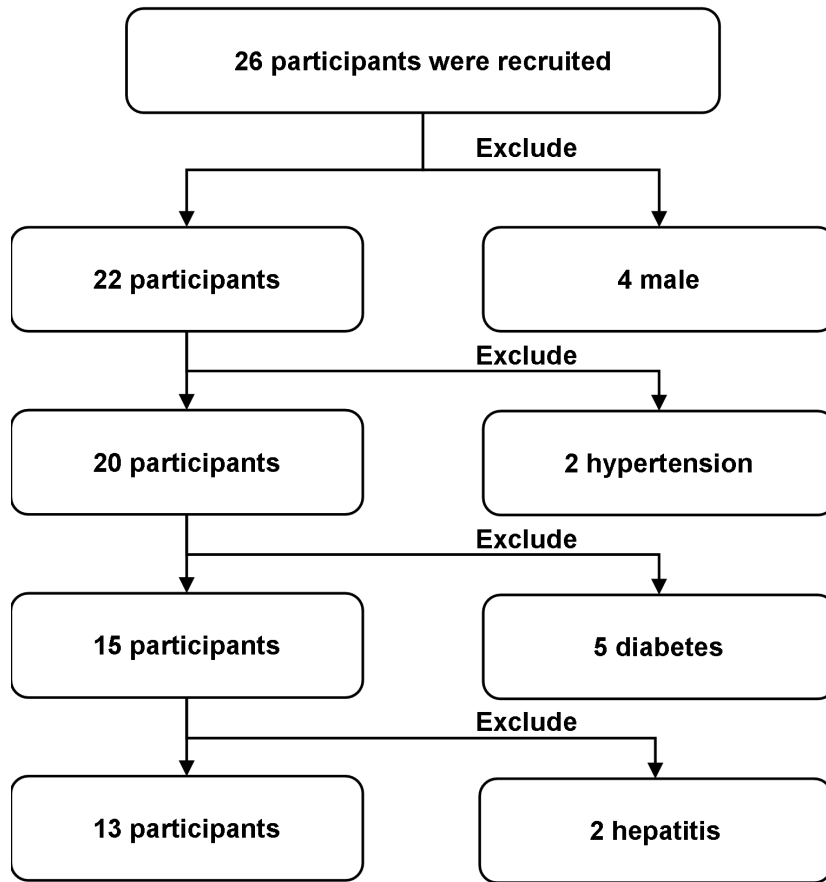


Fig. S2. Flow chart of the inclusion and exclusion criteria.

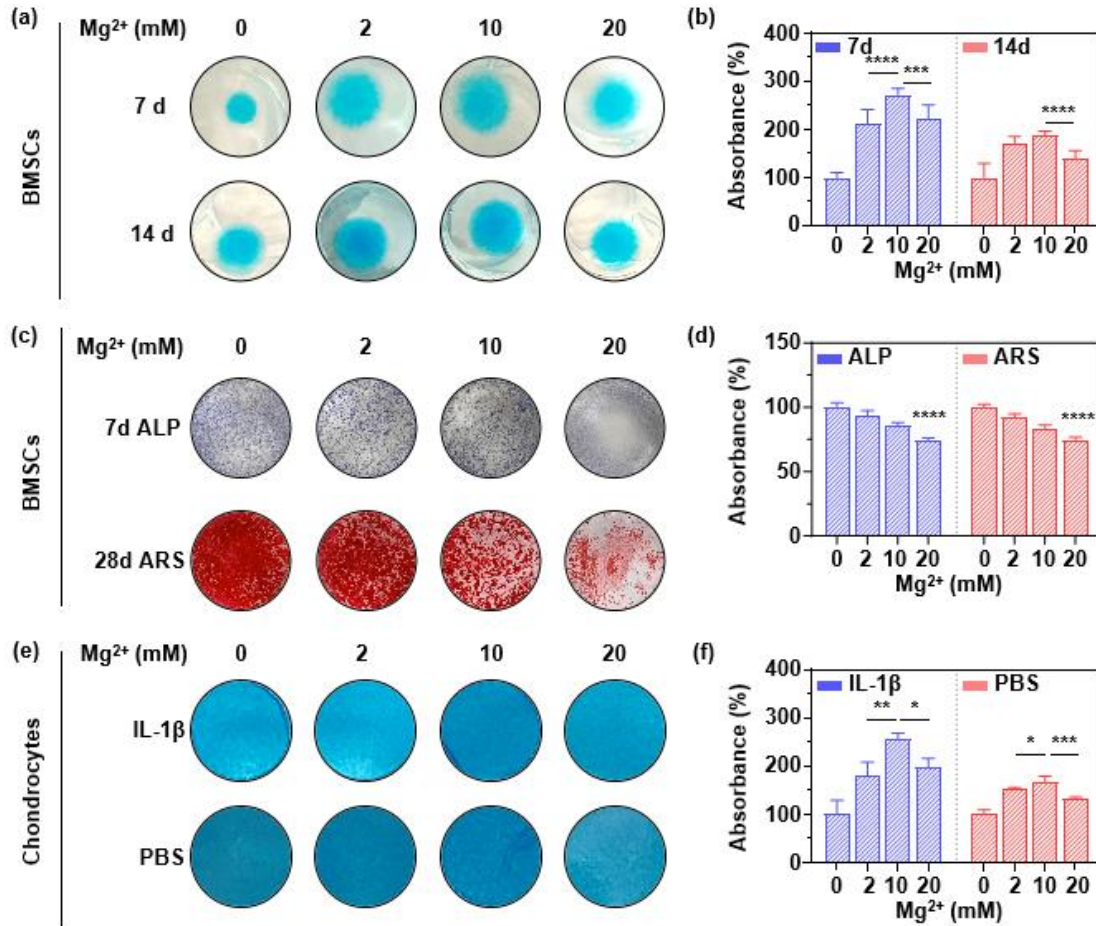


Fig. S3. Gradient Mg²⁺ regulated chondrogenic and osteogenic differentiation of BMSCs and protected inflammatory damaged chondrocytes *in vitro*.

Alcian blue staining of BMSCs treated with gradient Mg²⁺ (A) and quantitative results (B). ALP and ARS staining of BMSCs treated with gradient Mg²⁺ (C) and quantitative results (D). Alcian blue staining of chondrocytes treated with gradient Mg²⁺ with/without supplementation with 10 ng/mL IL-1β (E) and quantitative results (F).

Notes: All data are the mean ± s.d. Statistical differences between groups were determined by one-way ANOVA analysis. *P<0.05, **P<0.01, ***P<0.001, ****P<0.0001. n=3.

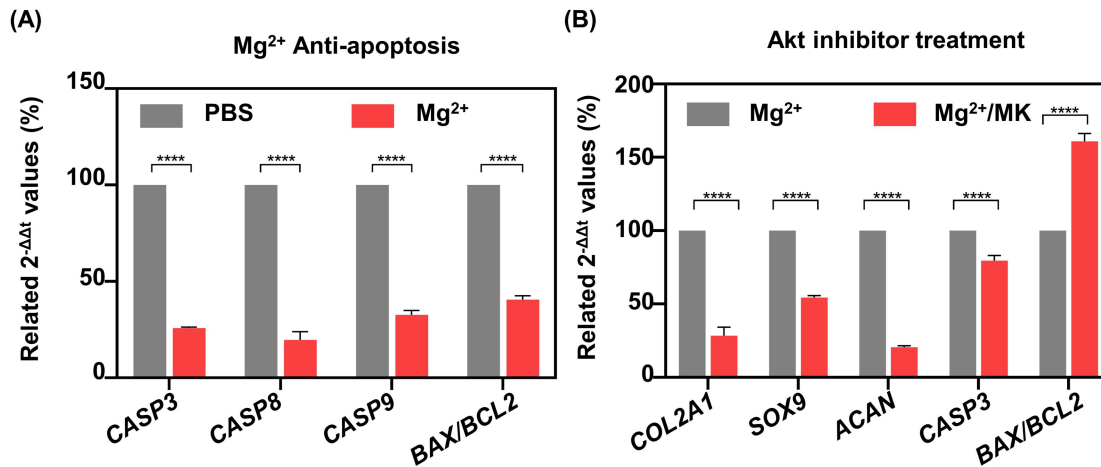


Fig. S4. Mg²⁺ inhibited apoptosis of chondrocytes via phosphorylation of Akt.

(A) Relative mRNA levels of apoptosis-related genes (*CASP3*, *CASP8*, *CASP9*, and *BAX/BCL2*) in SW1353 cells after treatment with 10 mM Mg²⁺ for 24 hours. (B) Relative mRNA levels (*COL2A1*, *SOX9*, *ACAN*, *CASP3*, *BAX/BCL2*) of apoptosis-related genes in SW1353 cells after treatment with 10 mM Mg²⁺ and MK2206 for 24 hours.

Notes: All data are the mean \pm s.d. Statistical differences between groups were determined by t test. ****P<0.0001. n = 4.

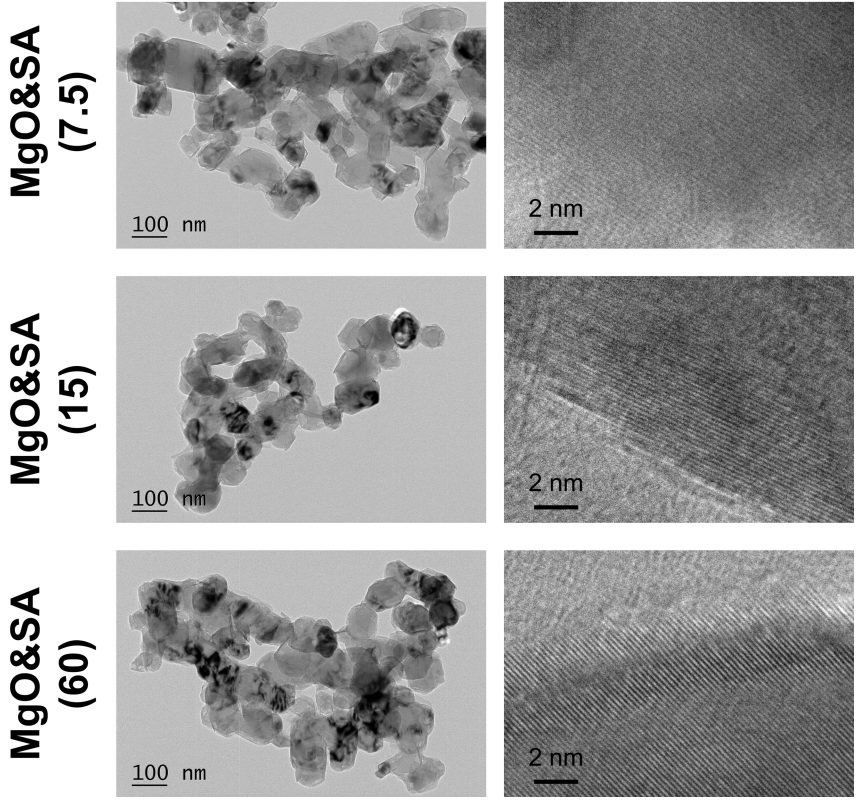


Fig. S5. TEM images of MgO nanoparticles modified with different concentrations of SA.

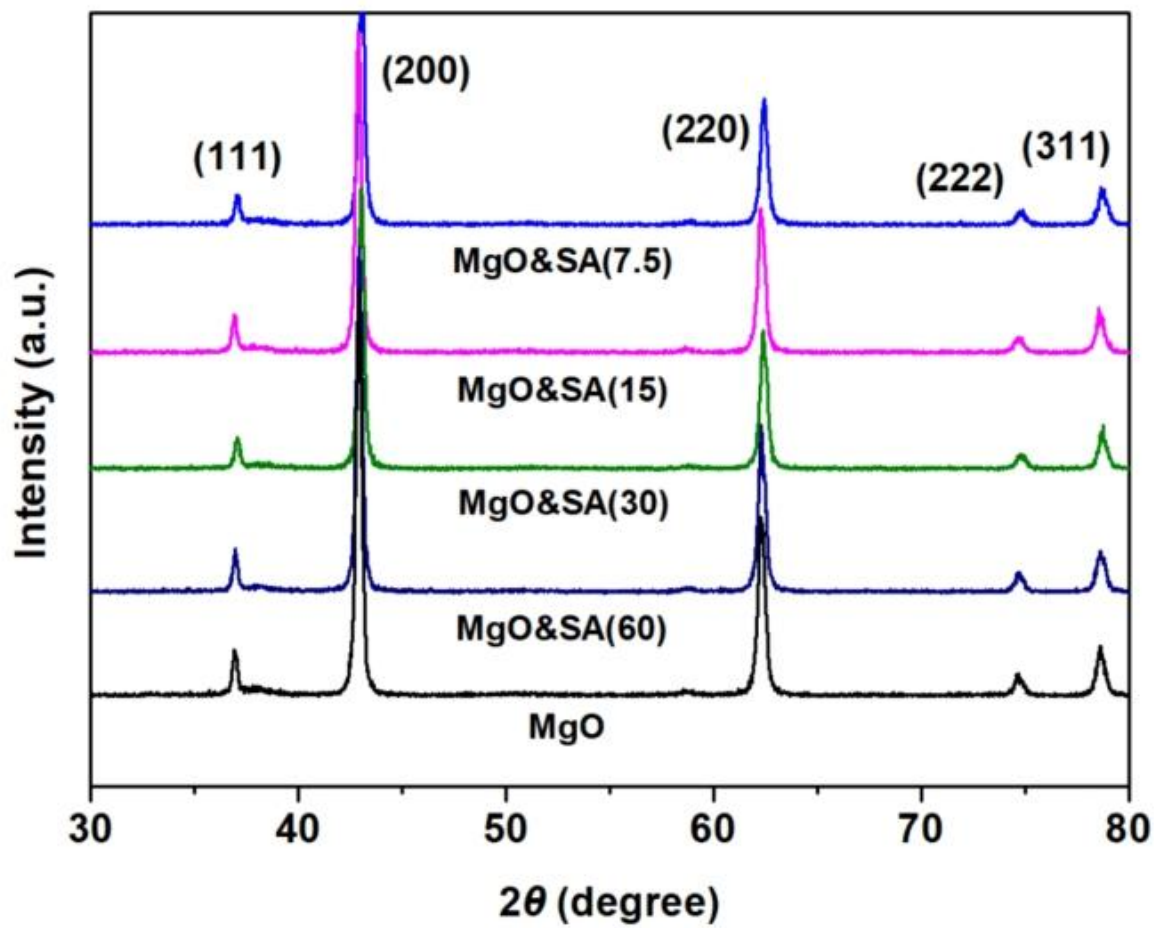


Fig. S6. XRD patterns of MgO nanoparticles modified with different concentrations of SA.

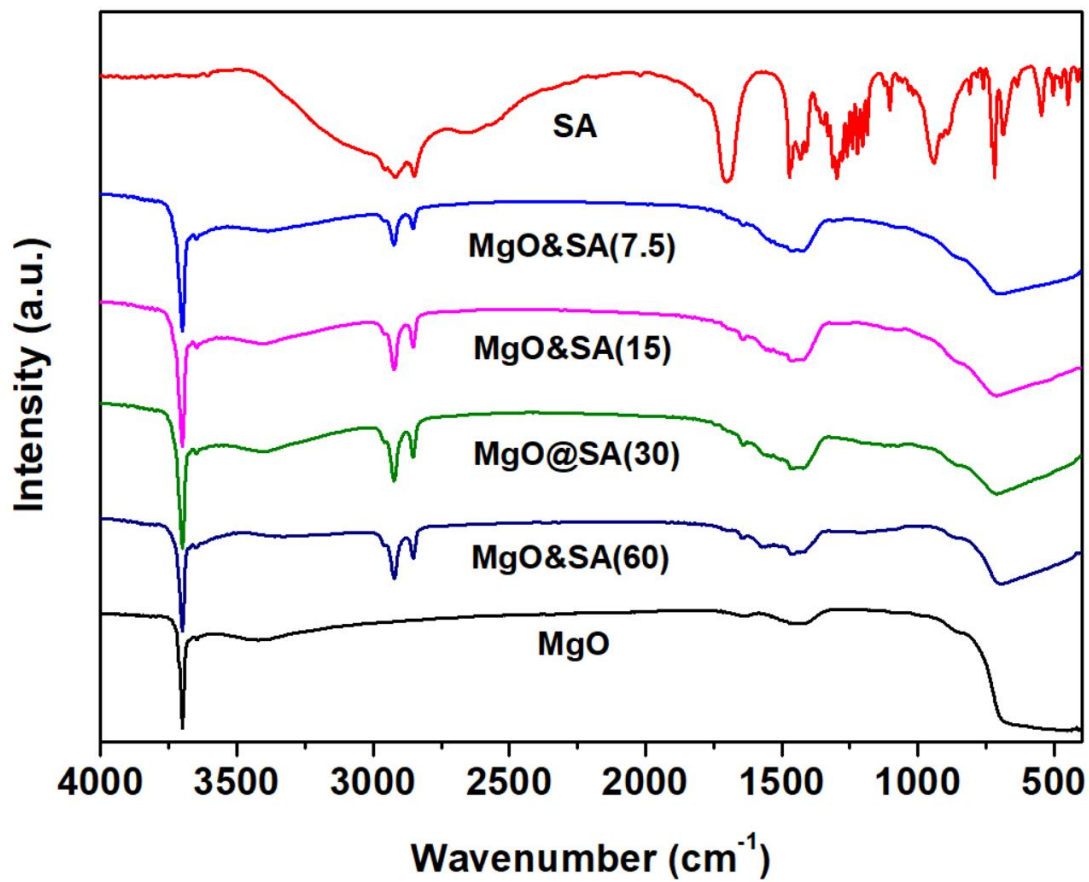


Fig. S7. FTIR spectra of MgO nanoparticles modified with different concentrations of SA.

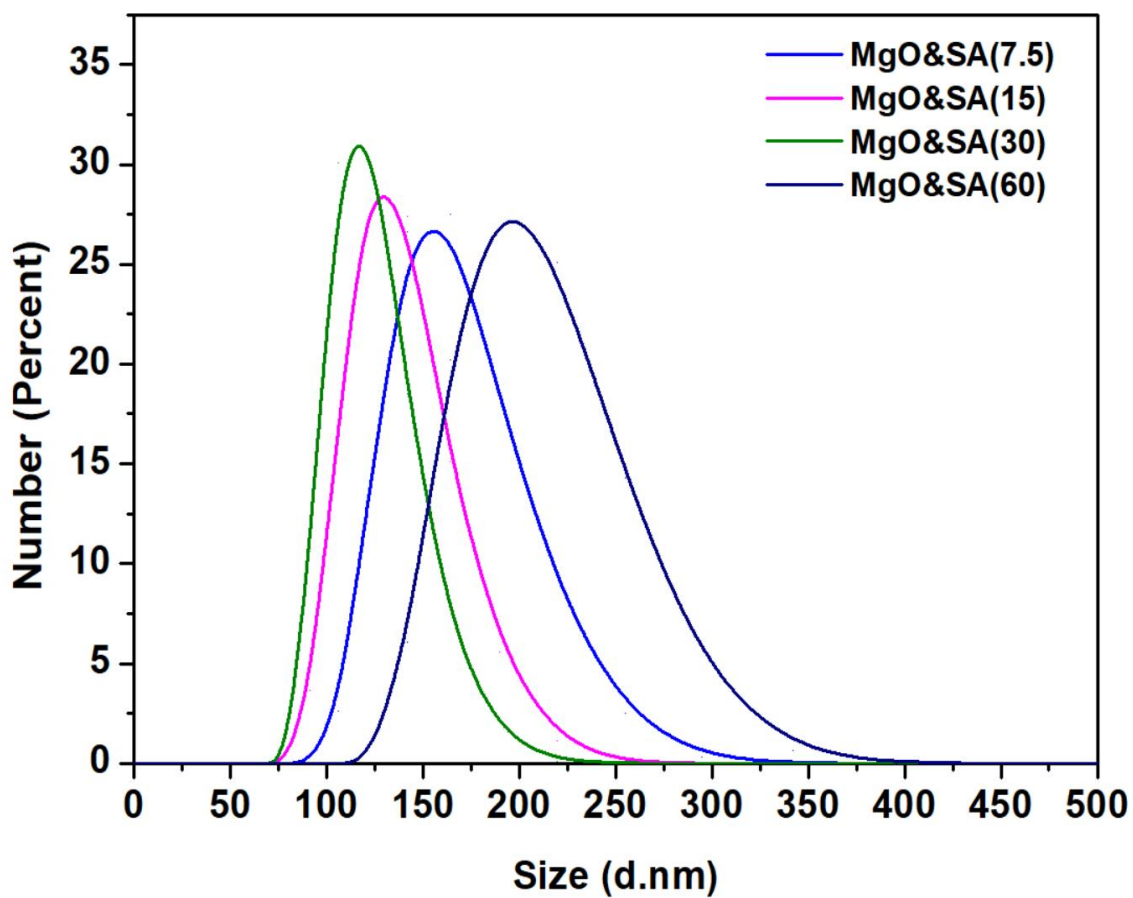


Fig. S8. DLS results of MgO nanoparticles modified with different concentrations of SA in DCM.

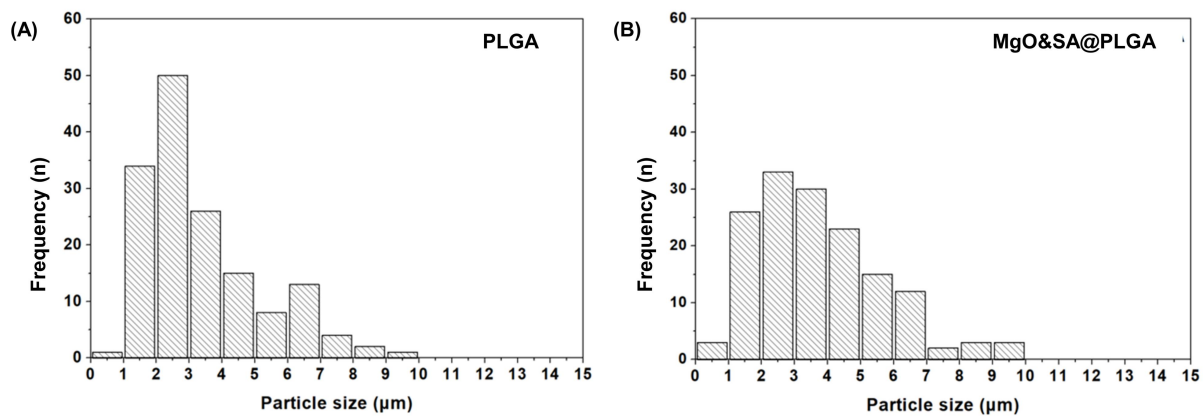


Fig. S9. Size distribution of PLGA (A) and MgO&SA@PLGA microspheres (B).

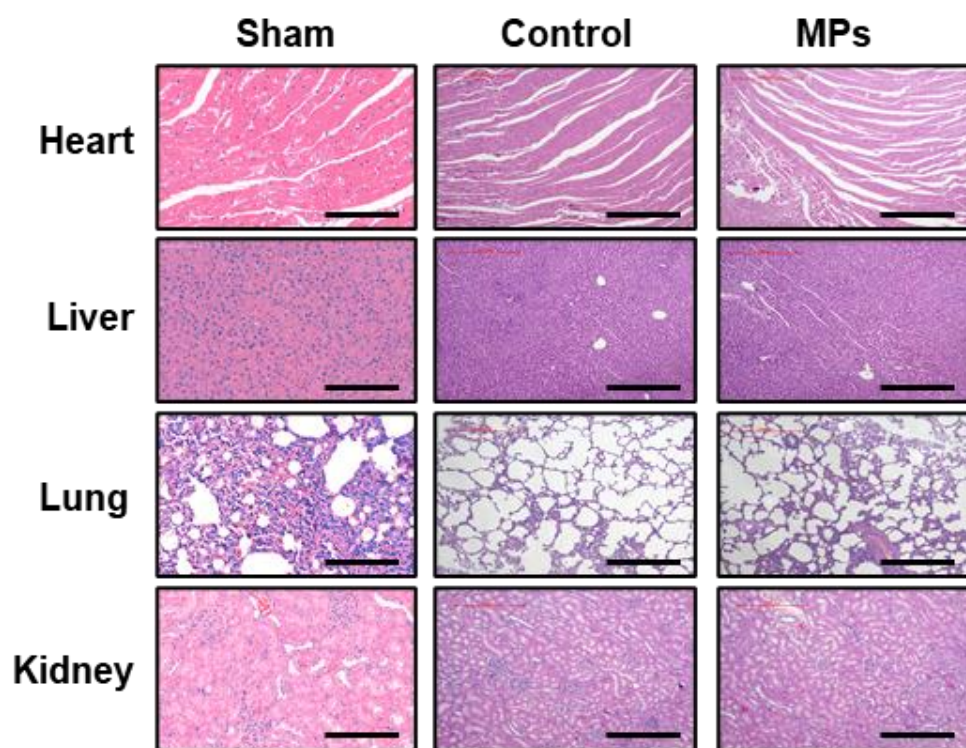


Fig. S10. Mg²⁺-releasing MPs showed no toxicity *in vivo*.

H&E staining of sections of heart, liver, kidney, and lung of rats conditionally treated for 4 weeks.

Scale bar: 100 μ m.

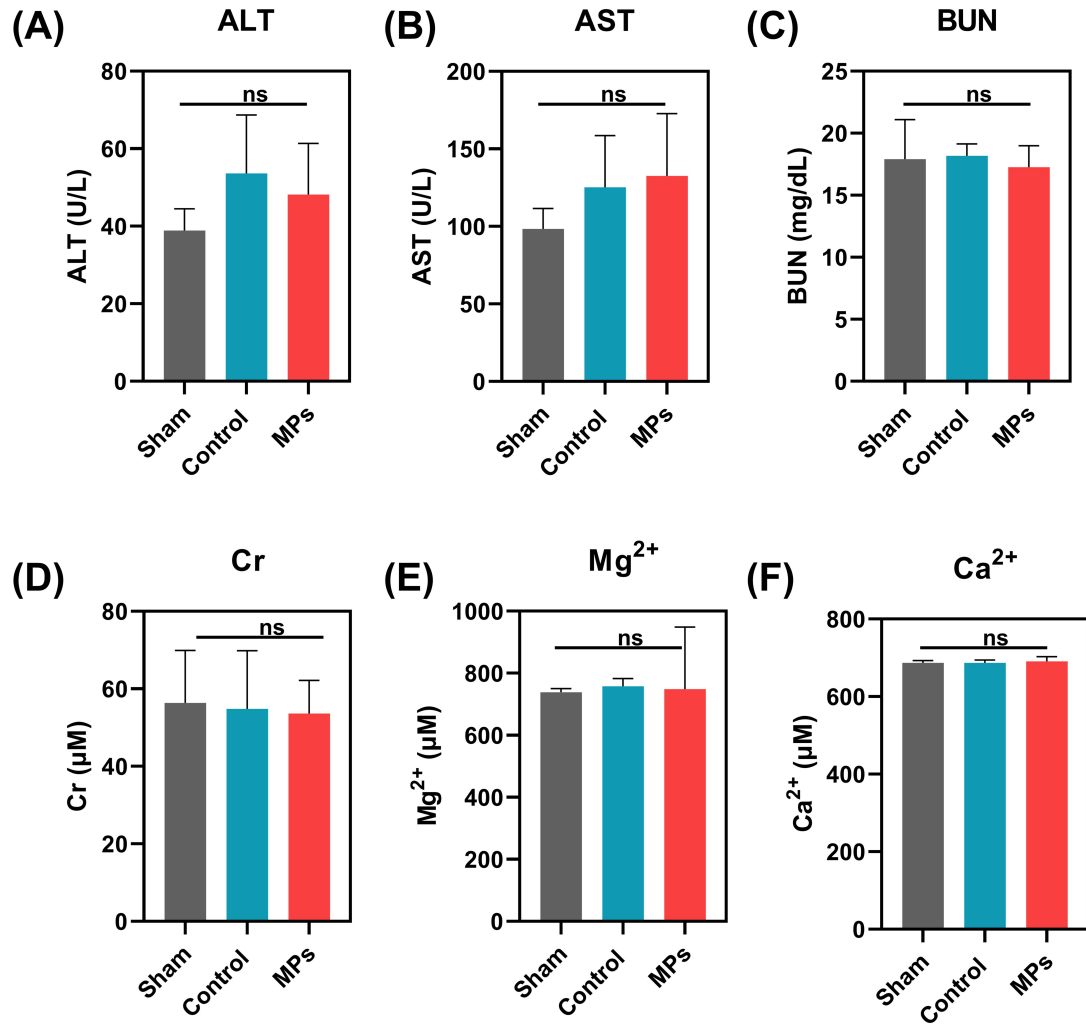


Fig. S11. Mg²⁺-releasing MPs showed no toxicity *in vivo*.

Serum indexes, including ALT (A), AST (B), BUN (C), Cr (D), Mg²⁺ (E), and Ca²⁺ (F), were tested.

Note: All data are the mean ± s.d. Statistical differences between groups were determined by one-way ANOVA analysis. ns, P>0.05. n=6.

Table S2. Bone morphological parameters of subchondral bone at 2 and 4 weeks after the OA model was established.

Indexes	Sham	Control	MPs	P-value
BMC (mg/cm ³)	571.3 ± 149.1	421.1 ± 35.7	512.4 ± 39.0	0.0048
BMD (mg/cm ³)	929.1 ± 37.3	953.1 ± 18.7	957.2 ± 21.9	0.7596
BV/TV (A.U.)	0.61 ± 0.19	0.43 ± 0.05	0.53 ± 0.05	0.0099
BS/BV (1/mm)	14.78 ± 4.79	19.32 ± 1.95	16.50 ± 1.75	0.0423
Conn.D (1/mm ³)	84.46 ± 29.21	80.92 ± 20.8	93.44 ± 14.89	0.3054
SMI (A.U.)	-3.41 ± 3.89	-0.38 ± 0.58	-1.16 ± 0.78	0.1113
Tb.N (1/mm)	4.05 ± 0.40	3.96 ± 0.36	4.20 ± 0.36	0.3258
Tb.Th (µm)	0.15 ± 0.06	0.10 ± 0.01	0.12 ± 0.01	0.0344
Tb.Sp (µm)	0.10 ± 0.05	0.15 ± 0.02	0.12 ± 0.02	0.0267

Note: P value showed the difference between Control and MPs groups and the P value was bold when P < 0.05. A.U. Arbitrary Units.

Table S3. Culture medium used to induce differentiation and indexes evaluated in different cells.

Cells	Medium for differentiation	Indexes evaluated
ATDC5	BM + ITS	Expression of Sox9 and Col2 α 1 separately at 7 and 14 days.
BMSCs (chondrogenic differentiation)	BM + Insulin (6.25 μ g/mL) + TGF- β 1 (10 ng/mL) + ascorbic acid (50 μ g/mL)	Alcian blue staining at 7 and 14 days.
hBMSCs (osteogenic differentiation)	BM + β -sodium glycerophosphate (10 mM) + dexamethasone (0.1 μ M) + ascorbic acid (50 μ g/mL)	ALP and ARS staining at 7 and 28 days. Expression of osteogenic related genes at specific period.
Mice monocytes	BM + MCSF (30 ng/mL) + RANKL (50 ng/mL)	The number and area of osteoclasts at 7 days.

Table S4. List of sequences of primers used in this study.

Gene	Primer sequences (5'-3')	Annealing temperature	Source	Species
<i>ACTB</i>	F: TCACCCACACTGTGCCCATCTACGA	64.1	Thermo	Human
	R: CAGCGGAACCGCTCATTGCCAATGG	70.1	Fisher	
<i>RUNX2</i>	F: TGGTTACTGTCATGGCGGGTA	62.9	Thermo	Human
	R: TCTCAGATCGTTGAACCTTGCTA	61.1	Fisher	
<i>OPN</i>	F: CTCCATTGACTCGAACGACTC	60.2	Thermo	Human
	R: CAGGTCTGCGAACTTCTTAGAT	60.0	Fisher	
<i>OCN</i>	F: CACTCCTCGCCCTATTGGC	62.2	Thermo	Human
	R: CCCTCCTGCTTGGACACAAAG	62.9	Fisher	
<i>COL1A2</i>	F: GAGGGCCAAGACGAAGACATC	55.0	Thermo	Human
	R: CAGATCACGTCATCGCACAAAC	53.8	Fisher	
<i>CASP3</i>	F: CATGGAAGCGAATCAATGGACT	60.7	Thermo	Human
	R: CTGTACCAGACCGAGATGTCA	60.6	Fisher	
<i>CASP8</i>	F: AGAGTCTGTGCCCAAATCAAC	51.3	Thermo	Human
	R: GCTGCTTCTCTTTGCTGAA	52.6	Fisher	
<i>CASP9</i>	F: CTGTCTACGGCACAGATGGAT	61.3	Thermo	Human
	R: GGGACTCGTCTTCAGGGGAA	62.8	Fisher	
<i>BAX</i>	F: CCCGAGAGGTCTTTTCCGAG	62.1	Thermo	Human
	R: CCAGCCCATGATGGTTCTGAT	61.9	Fisher	
<i>BCL2</i>	F: GGTGGGGTCATGTGTGTGG	62.6	Thermo	Human
	R: CGGTTCAGGTA CT CAGTCATCC	61.8	Fisher	
<i>SOX9</i>	F: AGCGAACGCACATCAAGAC	50.9	Thermo	Human
	R: CTGTAGGCGATCTGTTGGGG	54.4	Fisher	
<i>ACAN</i>	F: ACTCTGGGTTTTCGTGACTCT	49.5	Thermo	Human
	R: ACACTCAGCGAGTTGTCATGG	52.5	Fisher	
<i>COL2A1</i>	F: TGGACGATCAGGCGAAACC	55.6	Thermo	Human
	R: GCTGCGGATGCTCTCAATCT	54.4	Fisher	

Table S5. List of antibodies used in this study.

Name	Source	Cat.NO	Species
Anti-GAPDH	Abcam	ab8245	Mouse
Anti-CASPASE-3	Abcam	ab184787	Rabbit
Anti-B-ACTIN	Abcam	Ab115777	Mouse
Anti-SOX9	Abcam	ab185966	Rabbit
Anti-AKT1(phospho S473)	Abcam	ab81283	Rabbit
Anti-AKT3+AKT2+AKT1	Abcam	ab32505	Rabbit
Anti-COL2A1	Abcam	Ab34712	Rabbit
Anti-COL1A2	Abcam	AB308455	Rabbit
Anti-COL10A1	Abcam	AB182563	Rabbit
Anti-PI3KCA	CST	4249S	Rabbit
Anti-BAX	Abcam	Ab32503	Rabbit
Anti-BCL2	Abcam	Ab182858	Rabbit
Anti-INTEGRIN-B1	CST	4706	Rabbit
Anti-ERK1/2	CST	4695	Rabbit
Anti-ERK1/2(phosphor Thr202/Tyr204)	CST	4370	Rabbit