

Supplementary Materials for
**Activation of CGRP receptor–mediated signaling promotes tendon-bone
healing**

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Supplementary Figure and Table Legends

Fig. S1 Adenovirus vector profile and sequence. (A) Scramble adenovirus. (B) Adenovirus sh*Calcr1* #1. (C) Adenovirus sh*Calcr1* #2. (D) Adenovirus sh*Calcr1* #3. (E) Adenovirus *Calcr1* (NM_018782.2).

Fig. S2 Gene transfection efficiency of BMSCs incubated with different adenoviral plasmids. (A) Adenovirus sh*Calcr1* #1. (B) Adenovirus sh*Calcr1* #2. (C) Adenovirus sh*Calcr1* #3. (D) Adenovirus *Calcr1* (NM_018782.2). Scale bars: 100 μ m.

Fig. S3 The schematic diagram showing the surgical and treatment process (A) and the detailed process conducted in mice (B). I: incisions made around the knee joint; II: isolation of tendon graft from gastrocnemius tendon; III: exposure of the knee joint; IV: fixation of two ends of the tendon graft with sutures; V: transection of ACL; VI: creation of bone tunnel and injection of HMPs; VII: placement of the tendon graft and fixation at the femoral exit and the tibial entrance; VIII: closure of the wound. (C) **The animal number assigned for each testing in each group at different time points.**

Fig. S4 The effect of CGRP on calcium nodule formation in both mouse BMSC (mBMSC) (A) and human BMSC (hBMSC) (B) at 14 days and 21 days after osteogenic differentiation through quantitative analysis of Alizarin Red stained minerals. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$, $n = 3$ independent biological replicates. The Blank indicates BMSCs without incubation in osteogenic induction medium (OIM).

Fig. S5 The effect of CGRP on osteogenic differentiation capability of rat BMSCs and MC3T3-E1. (A) Gene expression levels of *Sp7*, *Runx2*, osteocalcin (*Ocn*), and osteopontin (*Opn*) of rat derived BMSCs after incubation in OIM with or without CGRP. * $P < 0.05$, ** $P < 0.01$, $n = 3$ individual biological replicates. (B) Gene expression levels of *Sp7*, *Runx2*, *Ocn*, and *Opn* of MC3T3-E1 after incubation in OIM with or without CGRP. ns: not significant, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, $n = 3$ individual biological replicates. (C) WB analysis of protein levels of OSX, p-CREB, CREB, PKA, and GAPDH in MC3T3-E1 after incubation in OIM with or without CGRP. The Blank indicates BMSCs without incubation in osteogenic induction medium (OIM).

Fig. S6 The effect of CGRP on osteogenic differentiation capability of human BMSCs. (A) Real-time PCR analysis of *Sp7*, *Runx2*, osteocalcin (*Ocn*), osteopontin (*Opn*) and *Calcr1* mRNA expression in the BMSCs after treatment with CGRP. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$, $n = 3$ independent biological replicates. (B) WB analysis of

protein levels of RUNX2, OSX, p-CREB, CREB, CALCRL and GAPDH in BMSCs with or without treatment of CGRP. (C) Quantification of RUNX2/GAPDH ratio, OSX/GAPDH ratio, and p-CREB/CREB ratio, and CALCRL/GAPDH ratio by WB analysis from (B). (D) WB analysis of protein levels of OSX, JUNB, SHH, p-CREB, CREB, and GAPDH in BMSCs with or without treatment of CGRP or H-89, a selective inhibitor of cyclic AMP-dependent protein kinase (PKA). *P<0.05, **P<0.01 and ***P<0.001, n=3 independent biological replicates. (E) Quantification of OSX/GAPDH ratio, JUNB/GAPDH, p-CREB/CREB ratio, and SHH/GAPDH ratio by WB analysis from (D). The Blank indicates BMSCs without incubation in osteogenic induction medium (OIM).

Fig. S7 Representative immunofluorescence staining of JUNB and NF200 at the tendon-bone interface in mice and the quantitative analysis of relative intensity of JUNB⁺ area at 4 weeks and 6 weeks post-surgery. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface. Scale bar: 200 μ m. *P<0.05, **P<0.01, and ***P<0.001, n=3.

Fig. S8 The fluorescence intensity of CGRP⁺, NF200⁺, CGRP⁺NF200⁺, and SHH⁺ area at the tendon-bone interface in mice. (A) Representative immunofluorescence staining of CGRP, NF200, and their co-immunostaining at the tendon-bone interface in mice at 4 weeks and 6 weeks post-surgery. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface. Scale bar: 200 μ m. (B) Representative immunofluorescence staining of SHH at the tendon-bone interface in mice at 4 weeks and 6 weeks post-surgery. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface. Scale bar: 200 μ m. (C) Quantitative analysis of relative fluorescence intensity of CGRP⁺NF200⁺ and SHH⁺ area at the tendon-bone interface in mice at 4 weeks and 6 weeks post-surgery. *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001, n=3.

Fig. S9 The expression level changes of CGRP and CALCRL at the trabecular bone in mice from pre-surgery to post-surgery. *P < 0.05, n=3. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface.

Fig. S10 Real-time whole-body imaging of mice at different time points after intra-articular injection of the fluorescent dye Cy5 grafted HMPs (HMP@Cy5) and PBS by In Vivo Imaging System (IVIS). The HMPs continued to degrade in the knee joint cavity after injection and a complete degradation was observed after 24 days within a 30-day experimental period.

Fig. S11 The fluorescence intensities of CD31⁺, NF200⁺ and RUNX2⁺ area as well as the ratios of CALCRL overlapping CD31, NF200, or RUNX2 at the tendon-bone interface in mice with or without adv-*Calcr* treatment. (A) Representative immunofluorescence staining of CD31, NF200, RUNX2, and their co-immunostaining with CALCRL at the tendon-bone interface in mice without or with adv-*Calcr* treatment at 1 week, 2 weeks, 4 weeks, and 6 weeks post-surgery. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface. Scale bar: 200 μ m. (B) Quantitative analysis of relative fluorescence intensities of CD31⁺, NF200⁺, and RUNX2⁺ area at the tendon-bone interface in the control group and the adv-*Calcr* group at 1 week, 2 weeks, 4 weeks, and 6 weeks post-surgery. *P<0.05, **P<0.01, ***P<0.001, n=3. (C) The ratios of CALCRL overlapping CD31, NF200, or RUNX2 at the tendon-bone interface in the control group and the adv-*Calcr* group at 1 week, 2 weeks, 4 weeks, and 6 weeks post-surgery. ns: not significant, *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001, n=3.

Fig. S12 Schematic illustration of healing mechanism at the tendon-bone interface modulated by CGRP receptor-mediated signaling in mice after adenoviral transfection through HMP controlled release. The activation of CGRP receptor-mediated signaling promotes the production of osteoblasts, which may be a driving force facilitating innervation and type H vessel formation at the TBI through upregulation of SHH and SLIT3, respectively.

Fig. S13 Representative histologic images showing the tendon-bone interface in both the control group and the adv-*Calcr* group at 6 weeks and 8 weeks post-surgery by Hematoxylin & Eosin (H&E) staining (left) and healing quality assessment using histologic scores. ns: not significant, n=5.

Fig. S14 The effect of SLIT3 treatment on graft healing. (A) Representative immunofluorescence staining of NF200 and CD31 and quantitative analysis of their relative fluorescence intensities at the tendon-bone interface in mice with various treatments at 2 weeks post-surgery. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface. Scale bar: 200 μ m. (B) Quantitative analysis of gait performance and load to failure in mice with various treatments at 6 weeks post-surgery. ns: not significant, *P < 0.05, **P < 0.01, ***P < 0.001, and ****P<0.0001, n=3.

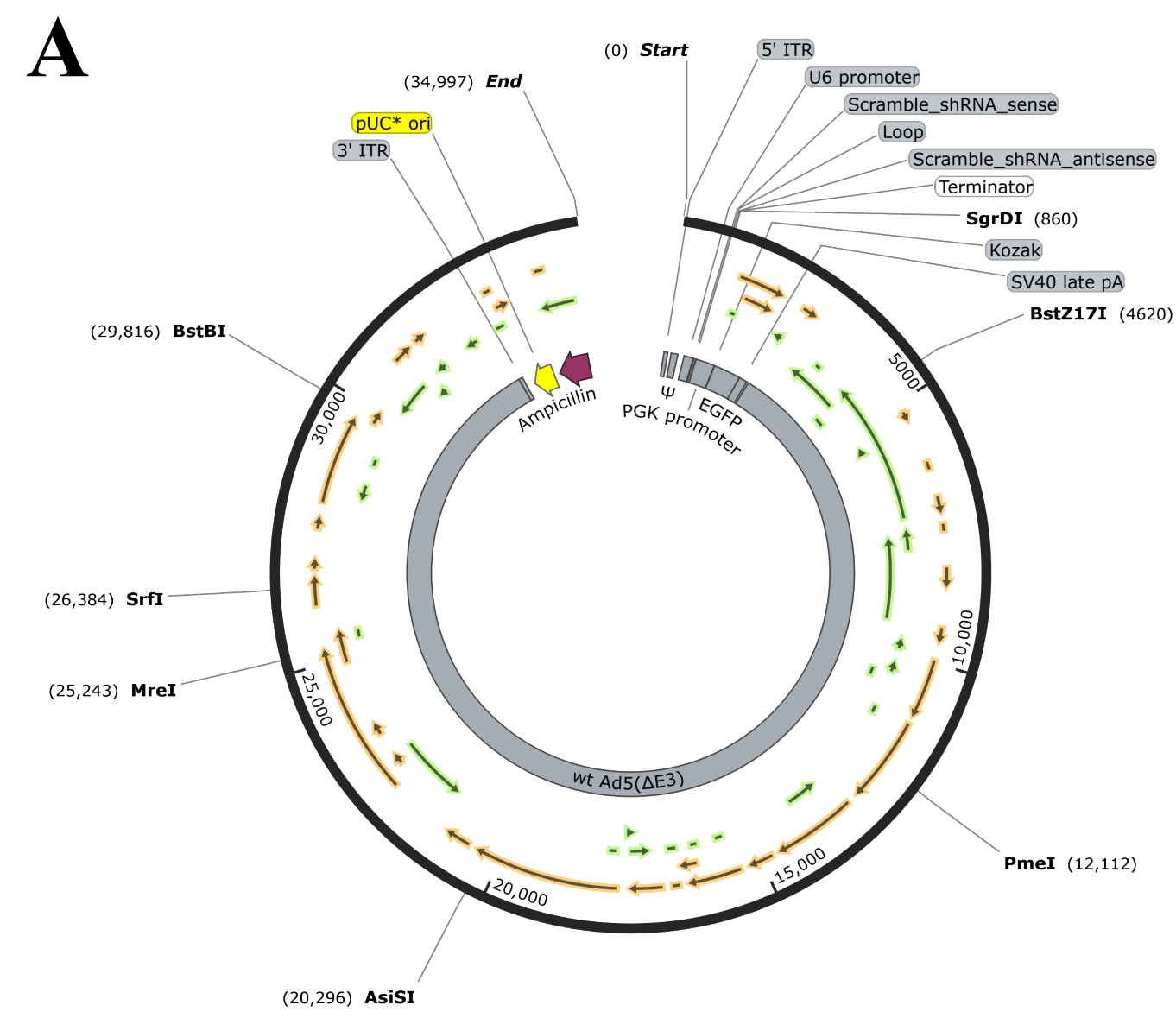
Fig. S15 The adv-*Calcr* and adv-sh*Calcr* injection show no health risks within a short-term period. (A) Hematological characteristics of mice after treatment for 2 days

and 4 days, respectively. The treatment indicates injection of HMPs loading PBS, scramble shRNA, *adv-Calcr1*, or *adv-shCalcr1*. The reference range was indicated by the dotted line. n=3. (B) Representative immunofluorescence staining of F4/80 assigned for macrophage at the tendon-bone interface in mice with different treatments at 2 weeks post-surgery. Dotted line: tendon graft boundary; B: bone; T: tendon; IF: interface.

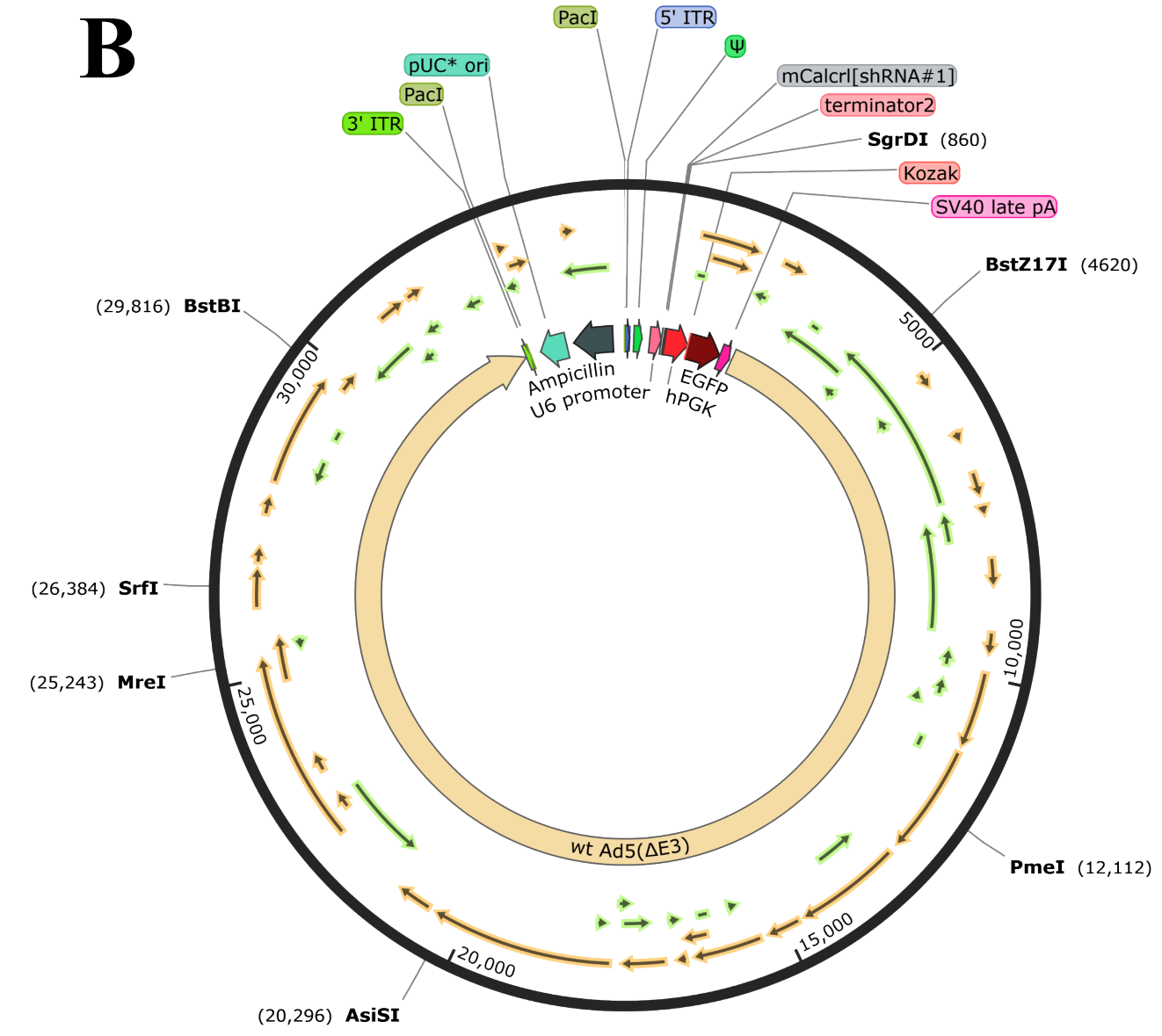
Table S1 Primer sequences used for real-time quantitative reverse transcriptase polymerase chain reaction (qRT-PCR)

Table S2 Histologic Scoring Criteria for semi-quantitative analysis of the tendon-bone interface healing.

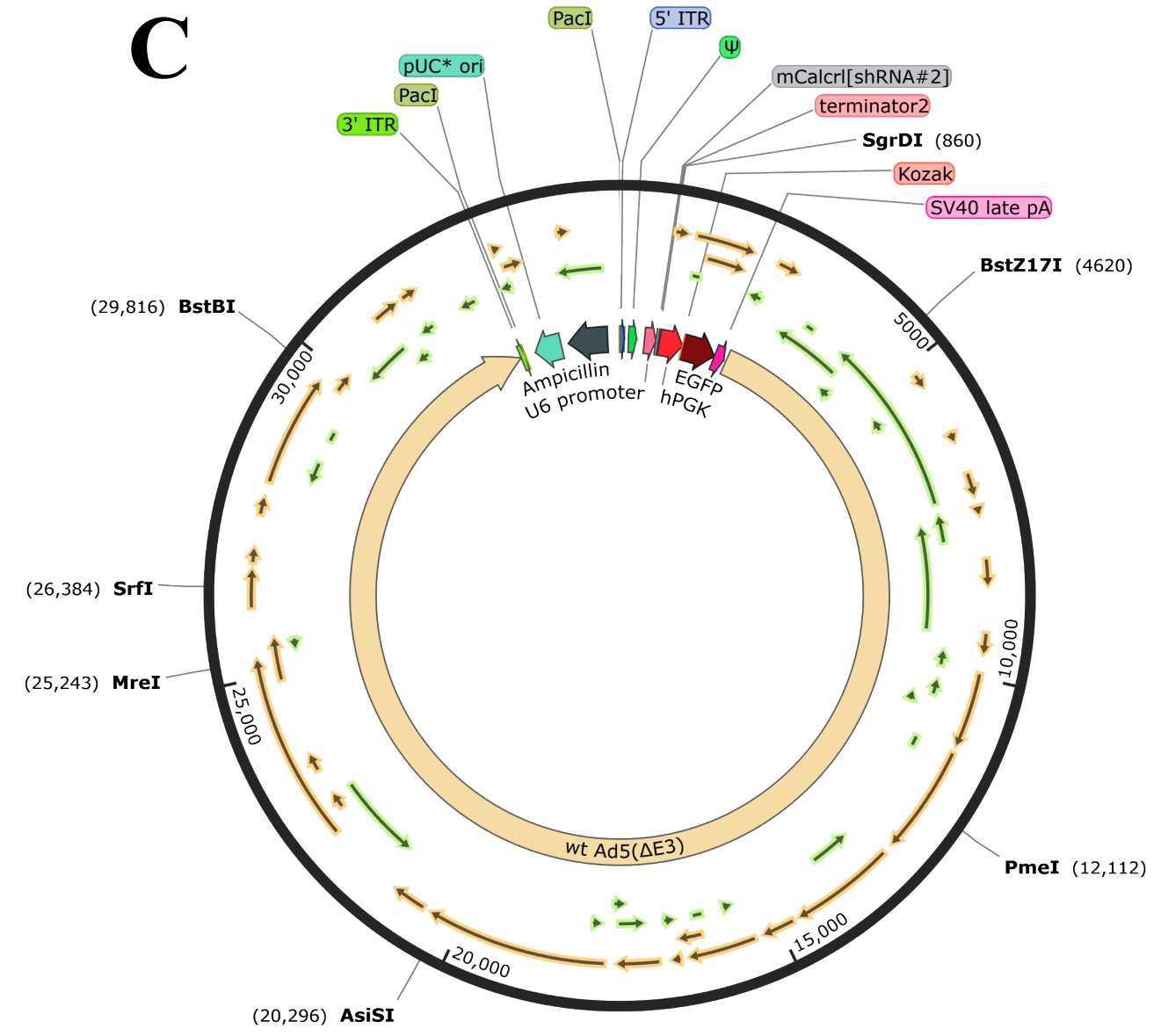
Fig. S1



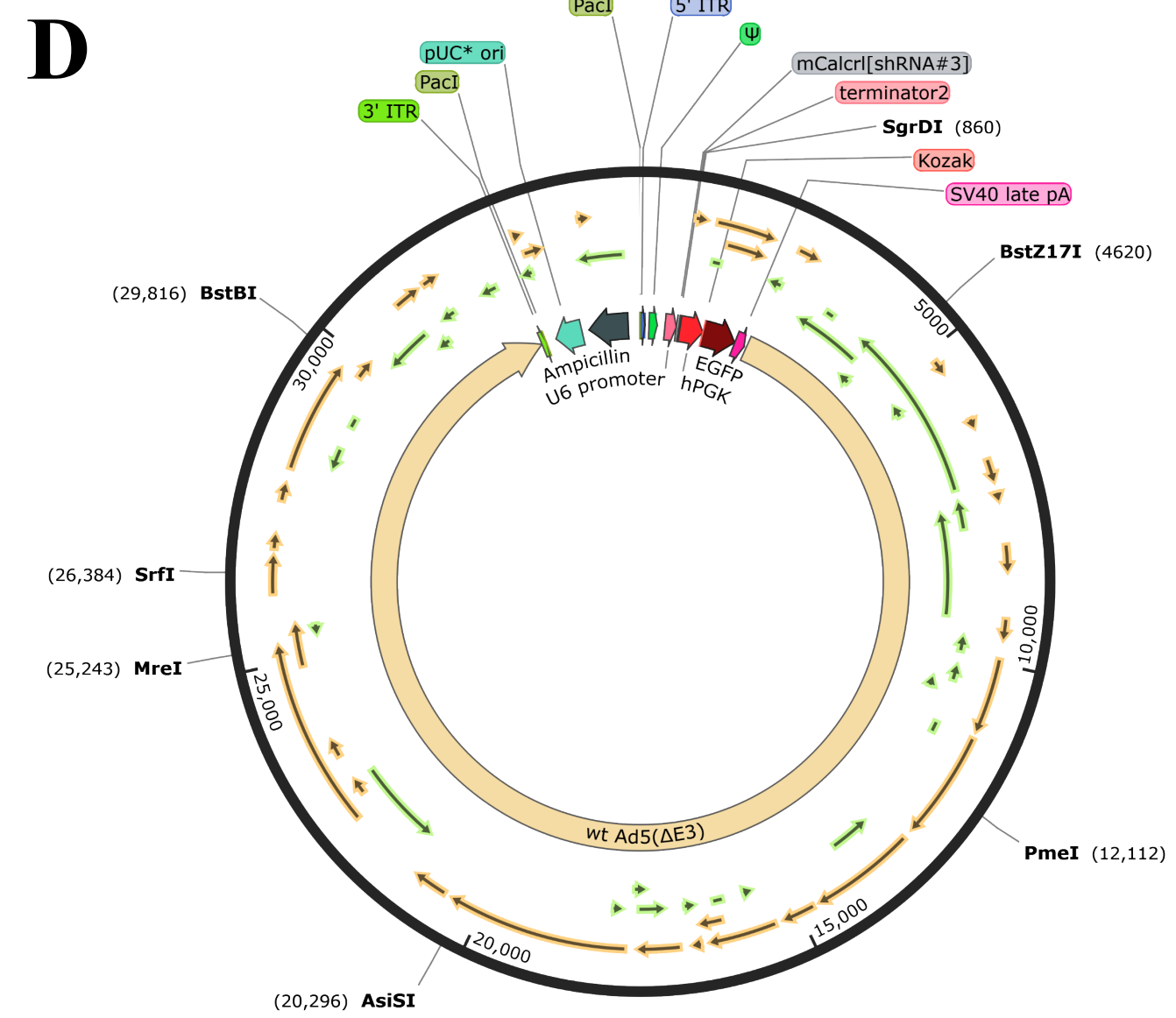
Mammalian knockdown vector; Adenovirus shRNA knockdown vector Scramble.
34,997 bp



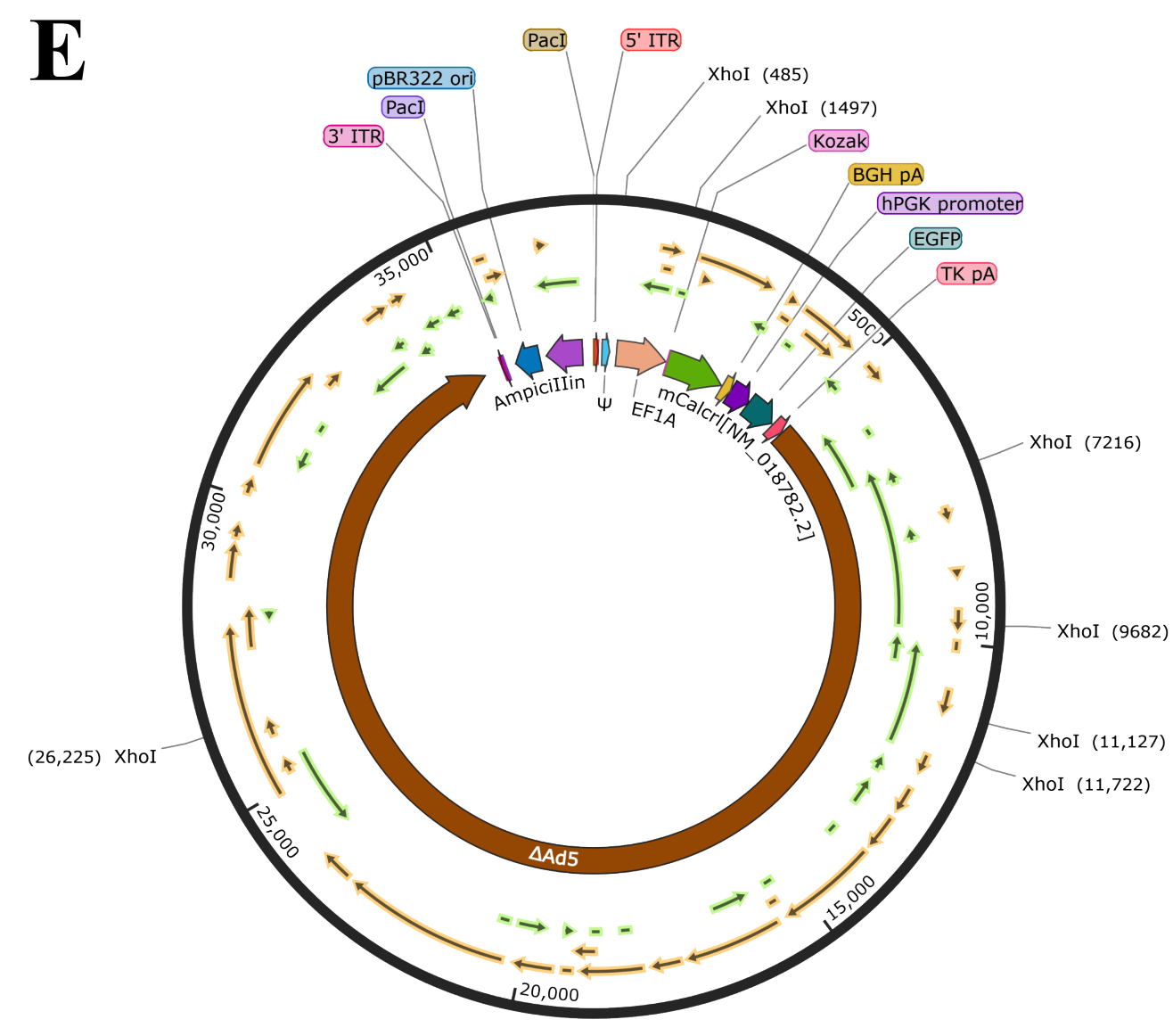
Mammalian knockdown vector; Adenovirus shRNA knockdown vector#1.
34,997 bp



Mammalian knockdown vector; Adenovirus shRNA knockdown vector#2.
34,997 bp



Mammalian knockdown vector; Adenovirus shRNA knockdown vector#3.
34,997 bp



Mammalian overexpression vector; Adenovirus mCalcrI[NM_018782.2] vector.
37,570 bp

Fig. S2

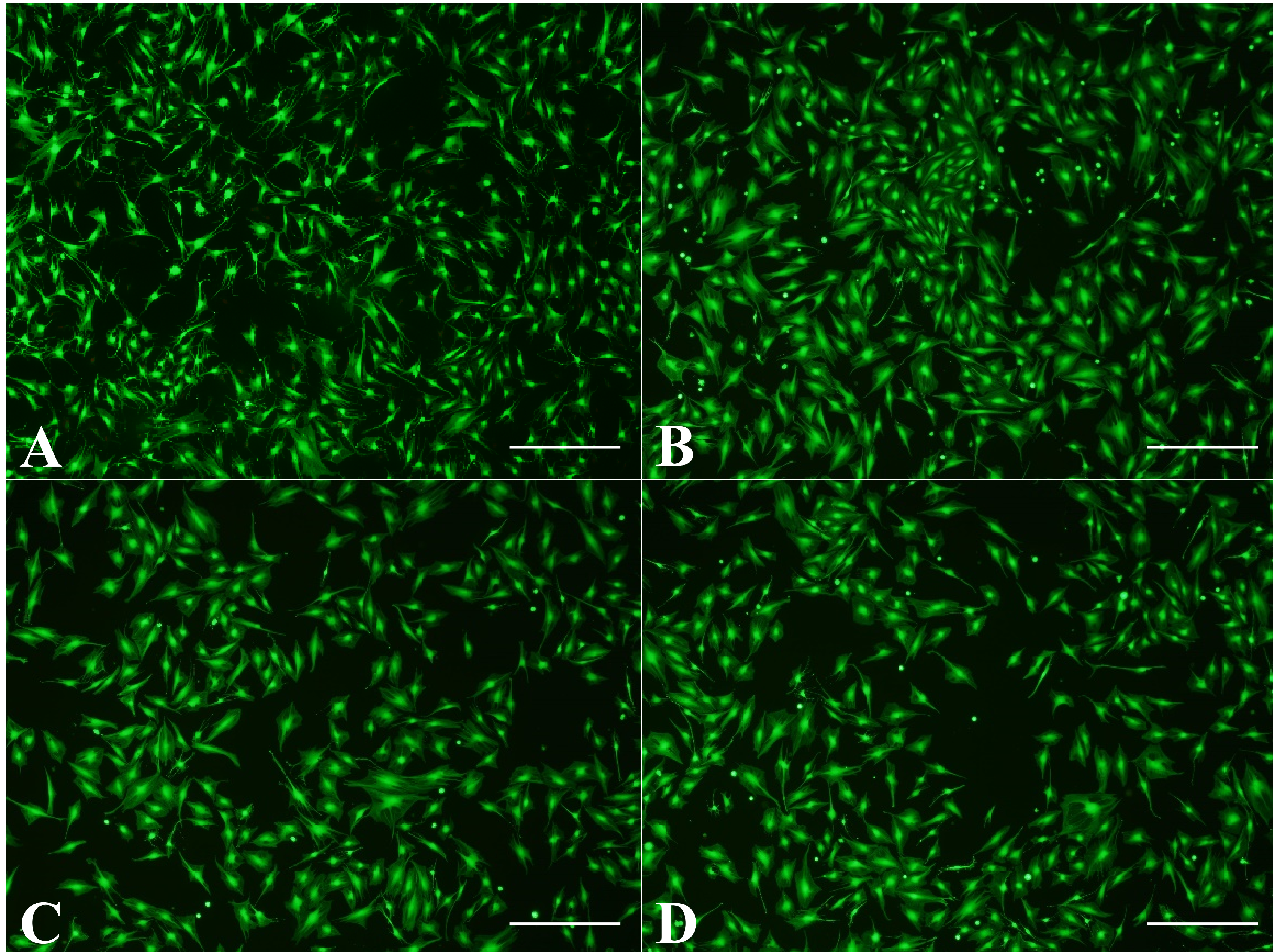
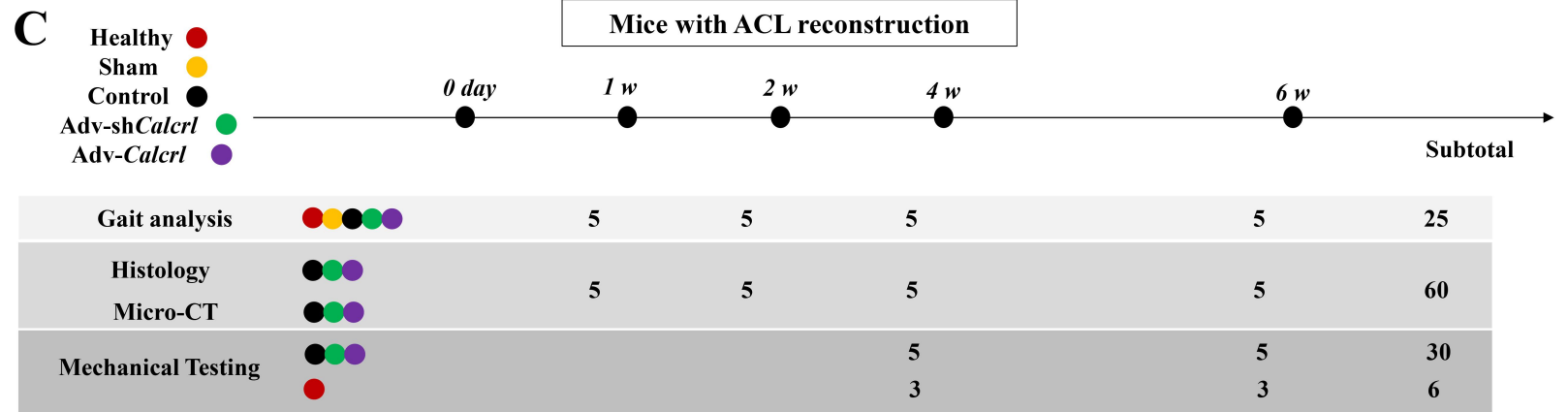
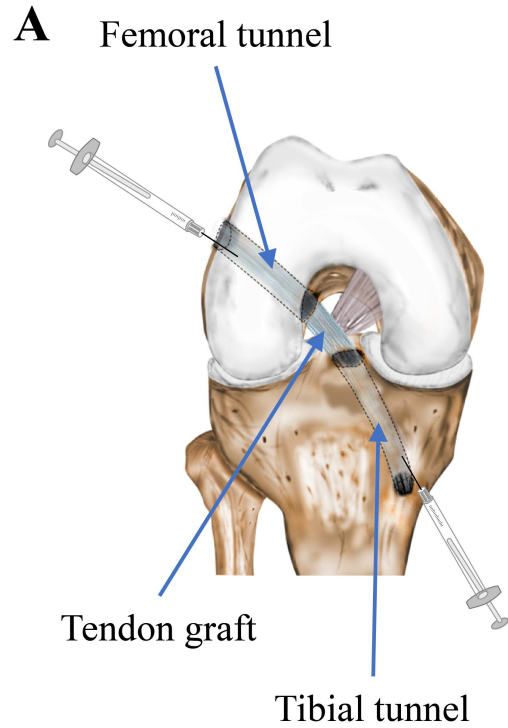


Fig. S3



Note: n=5 for H&E and Safranin&O staining, n=4-5 for micro-CT analysis while n=3 for immunofluorescence staining.

Total 121

Fig. S4

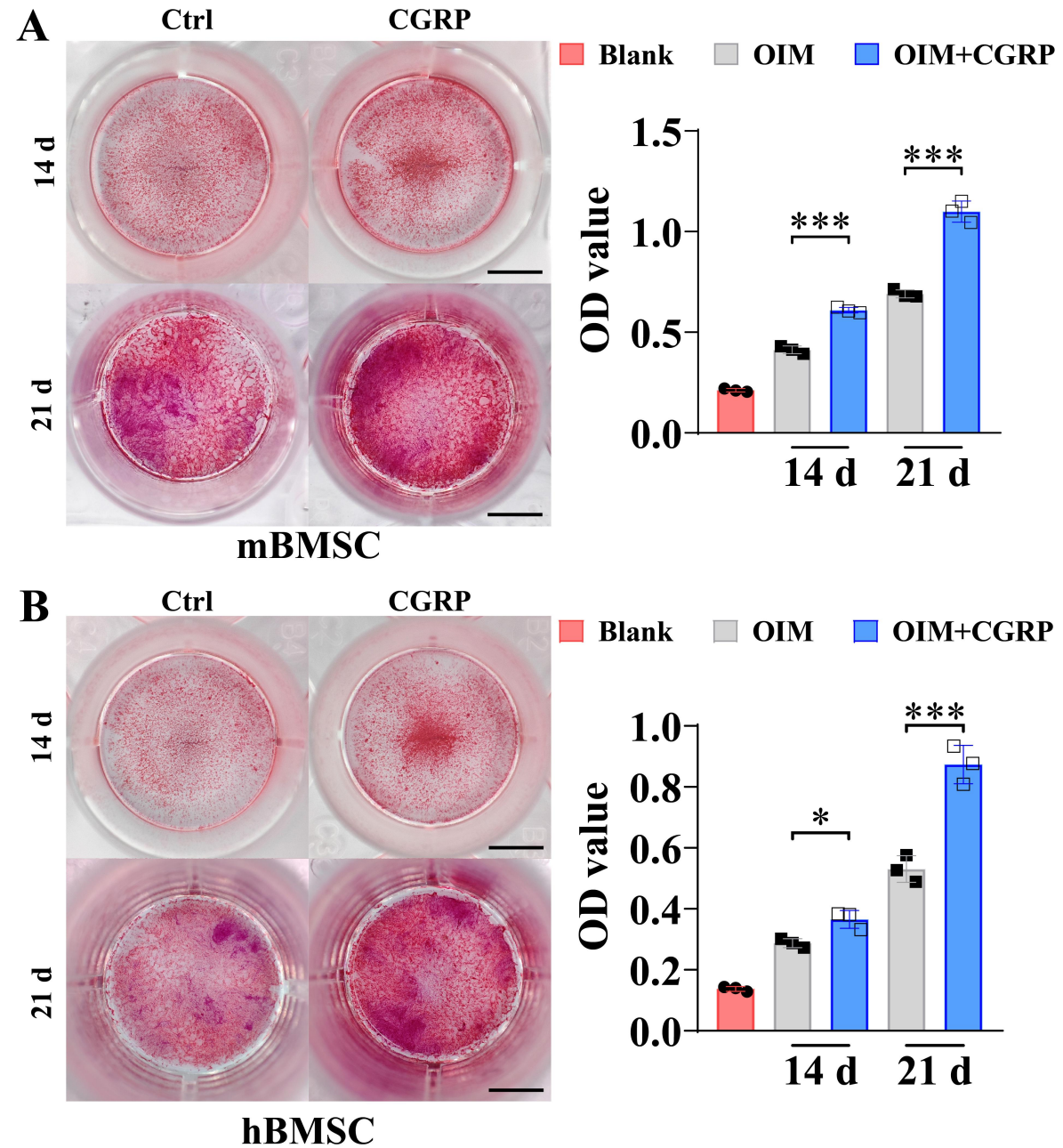


Fig. S6

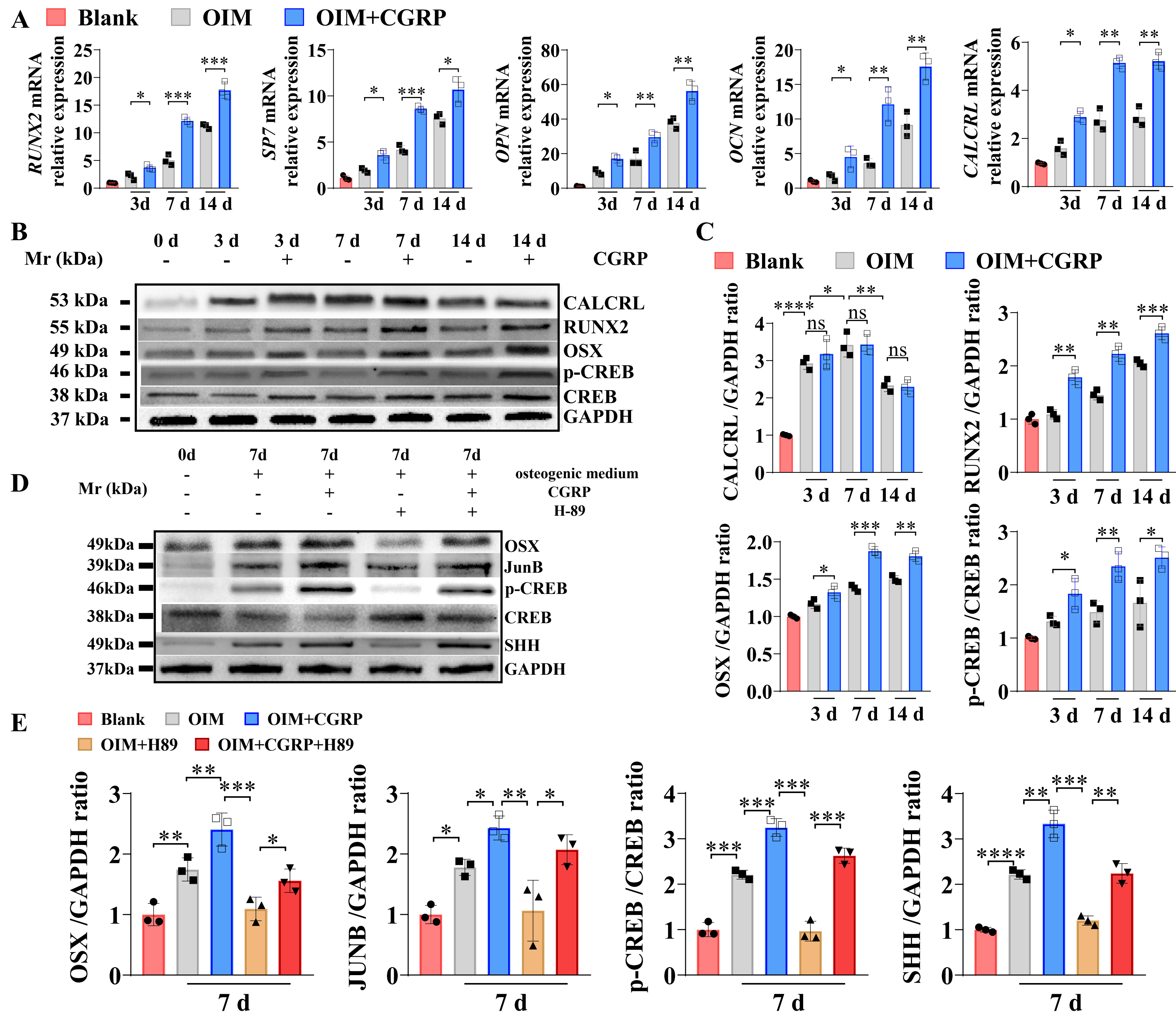


Fig. S7

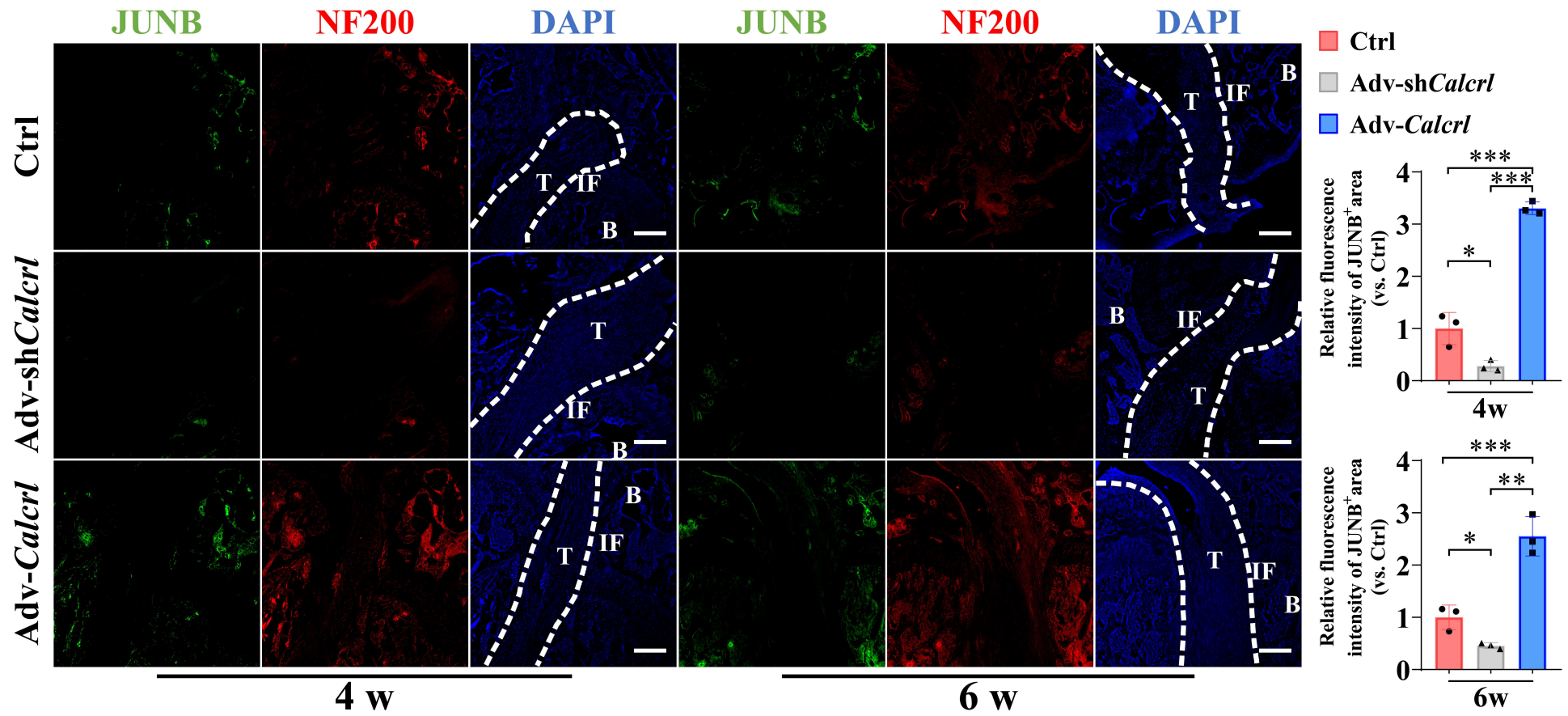


Fig. S8

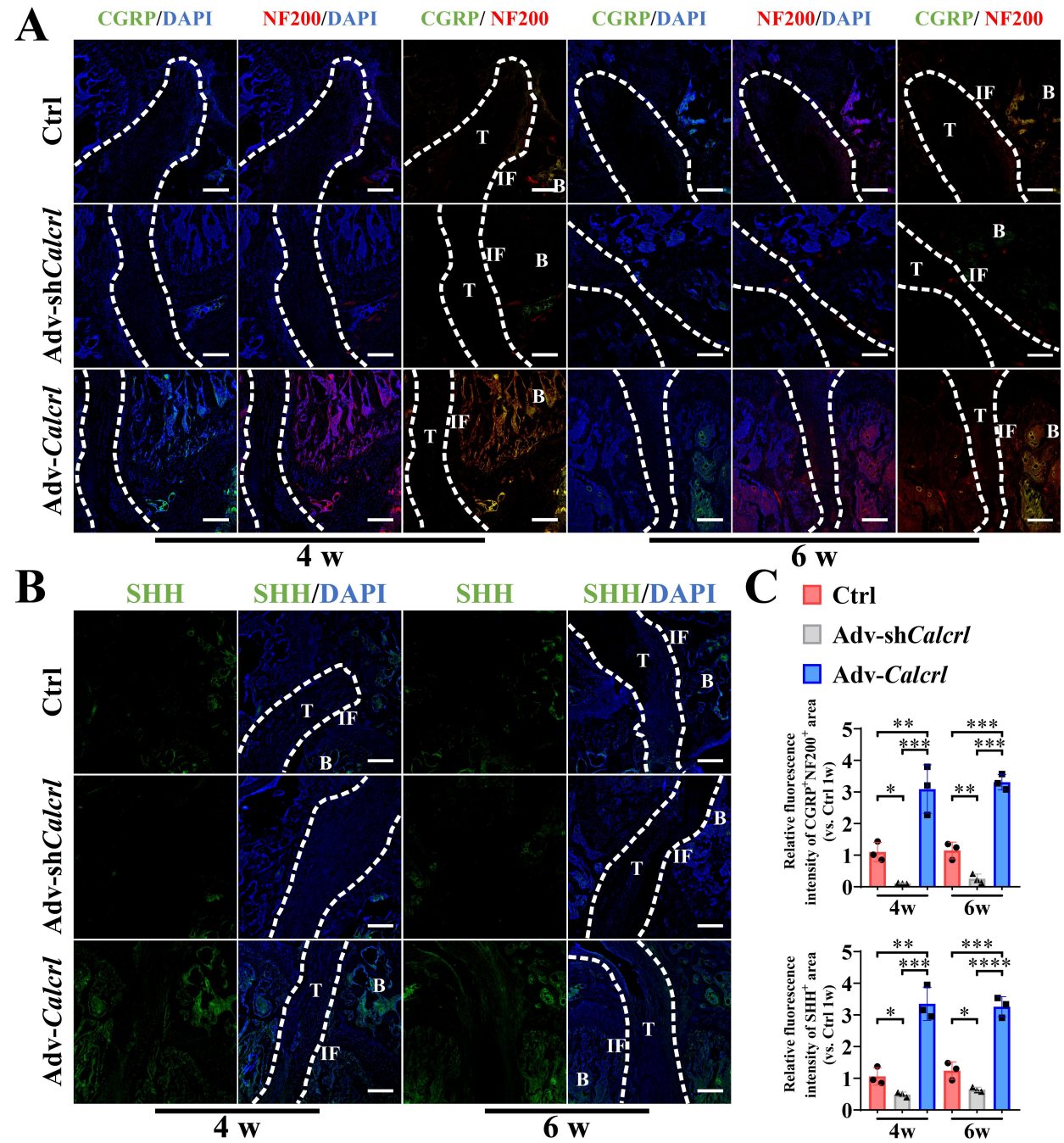


Fig. S9

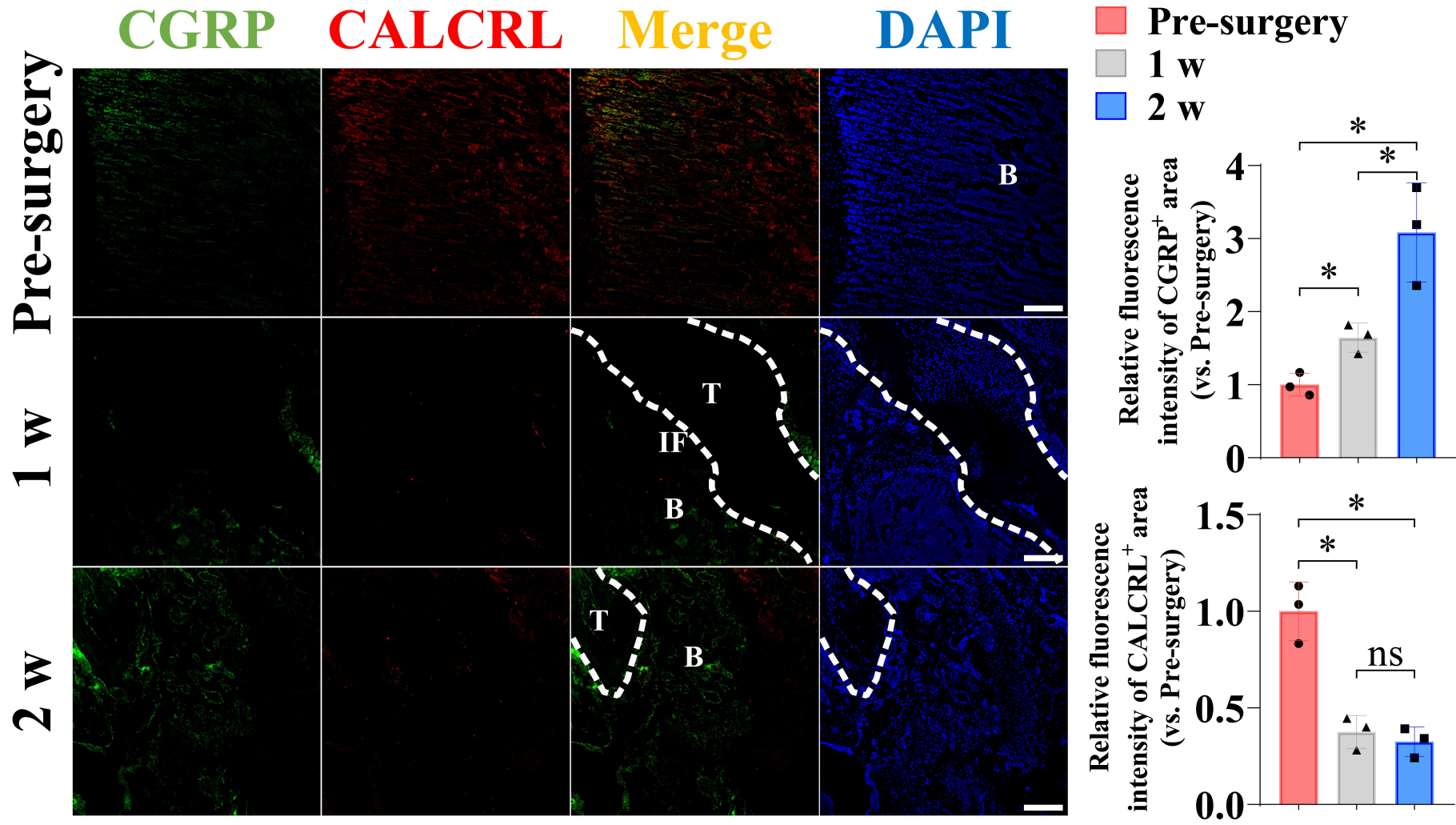


Fig. S10

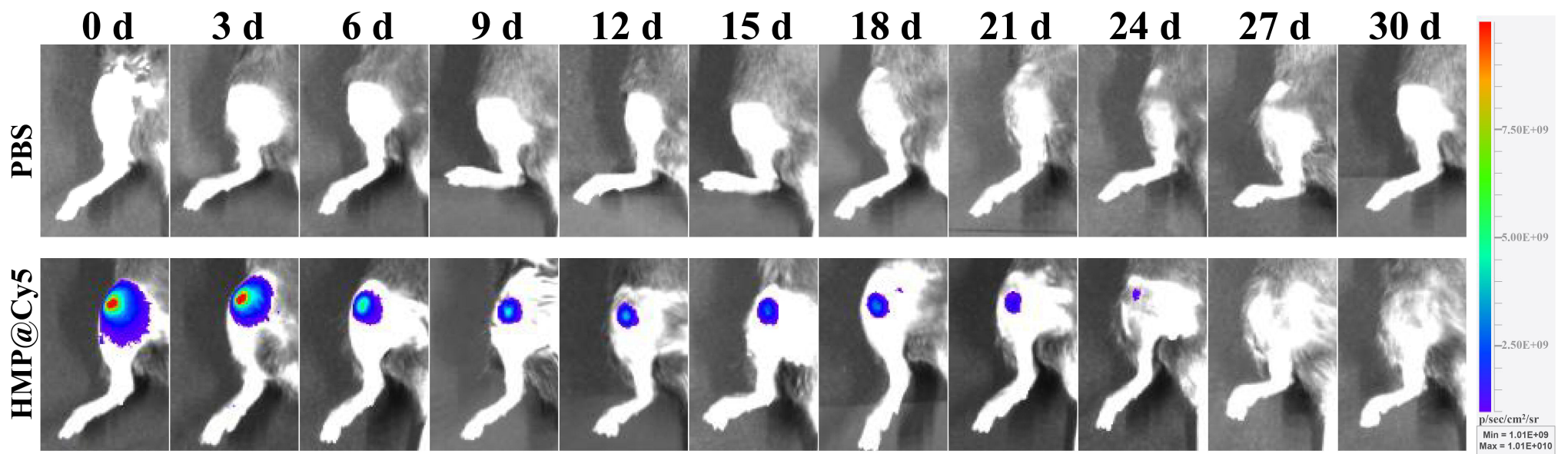


Fig. S11

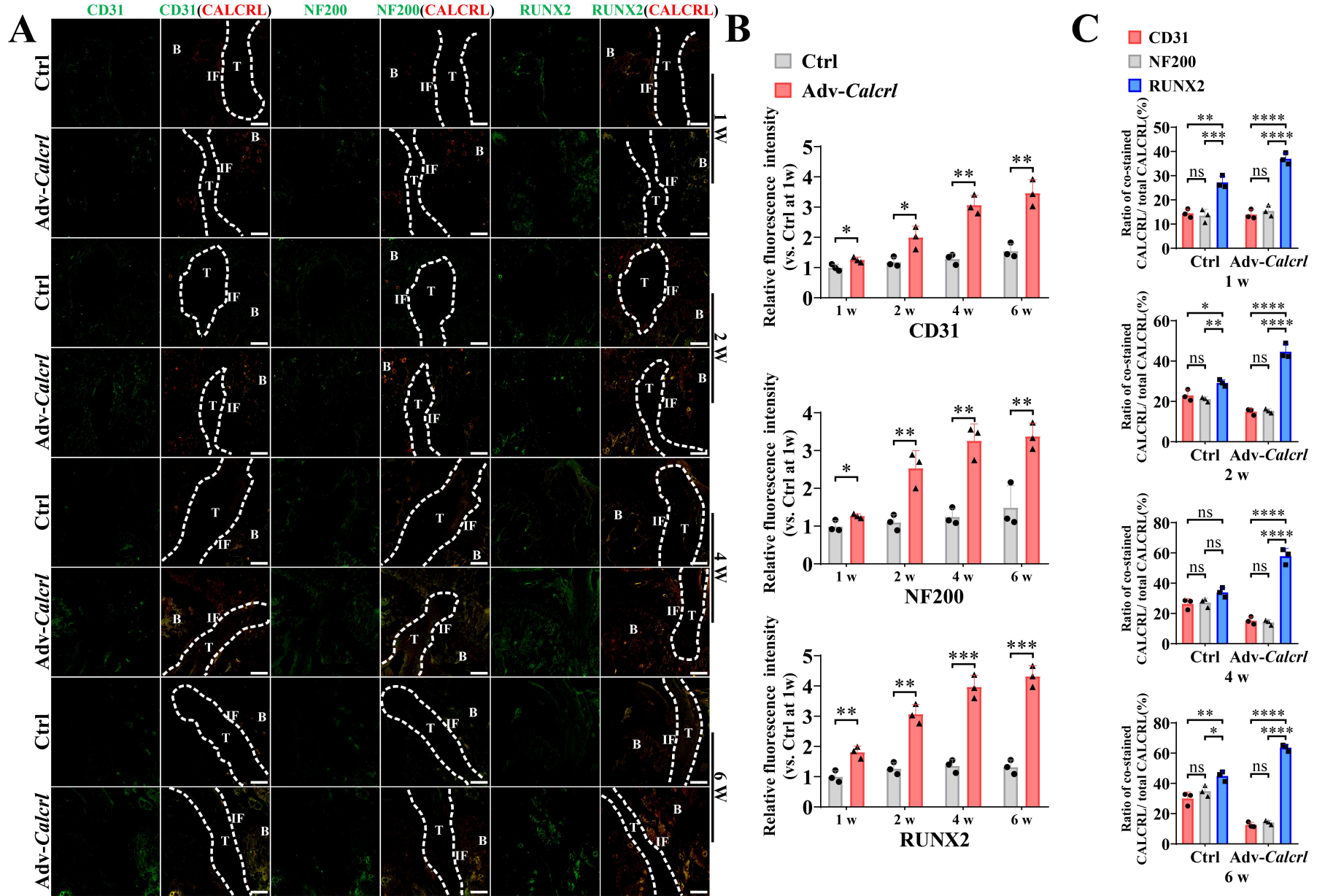


Fig. S12

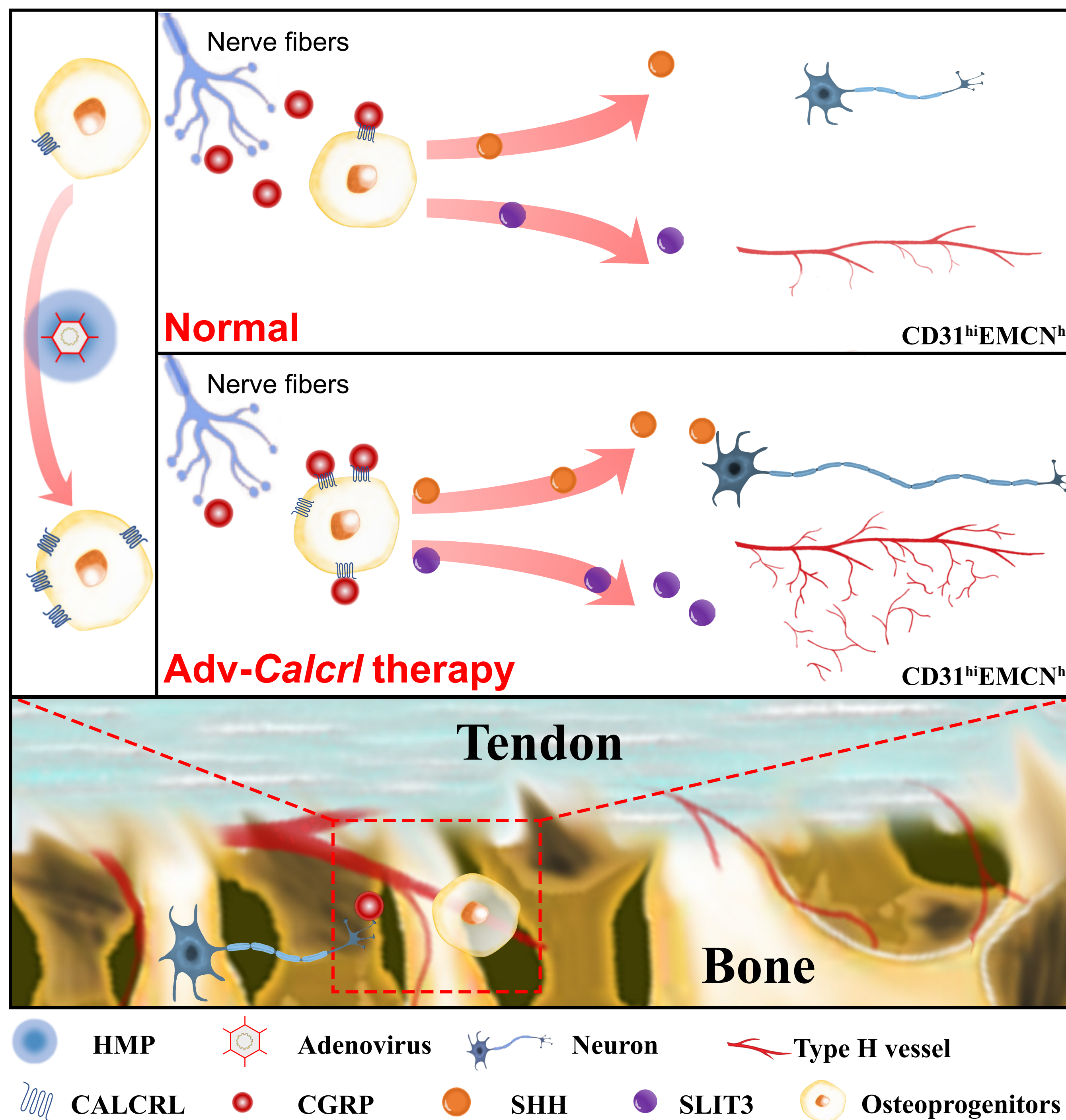


Fig. S13

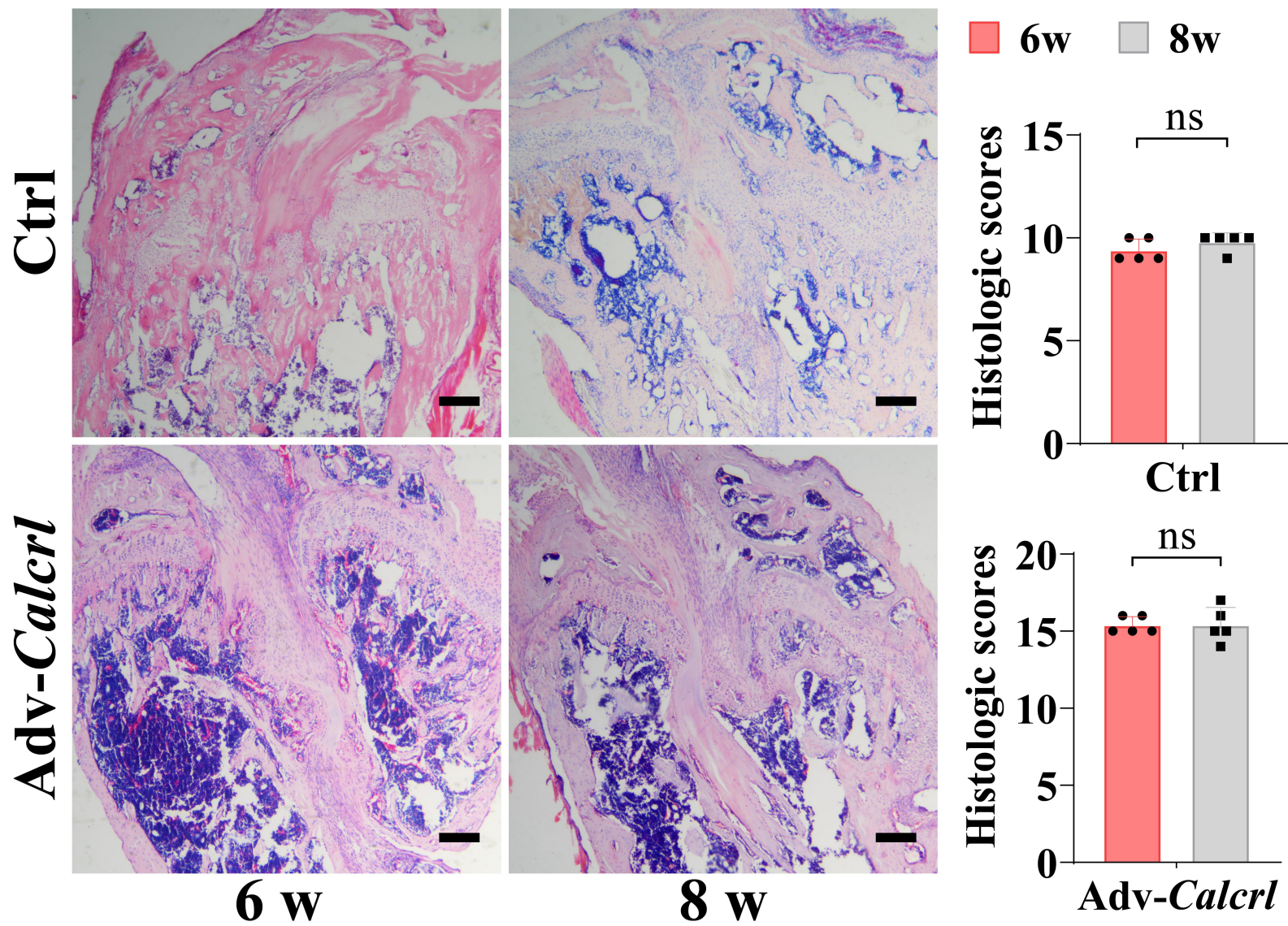


Fig. S14

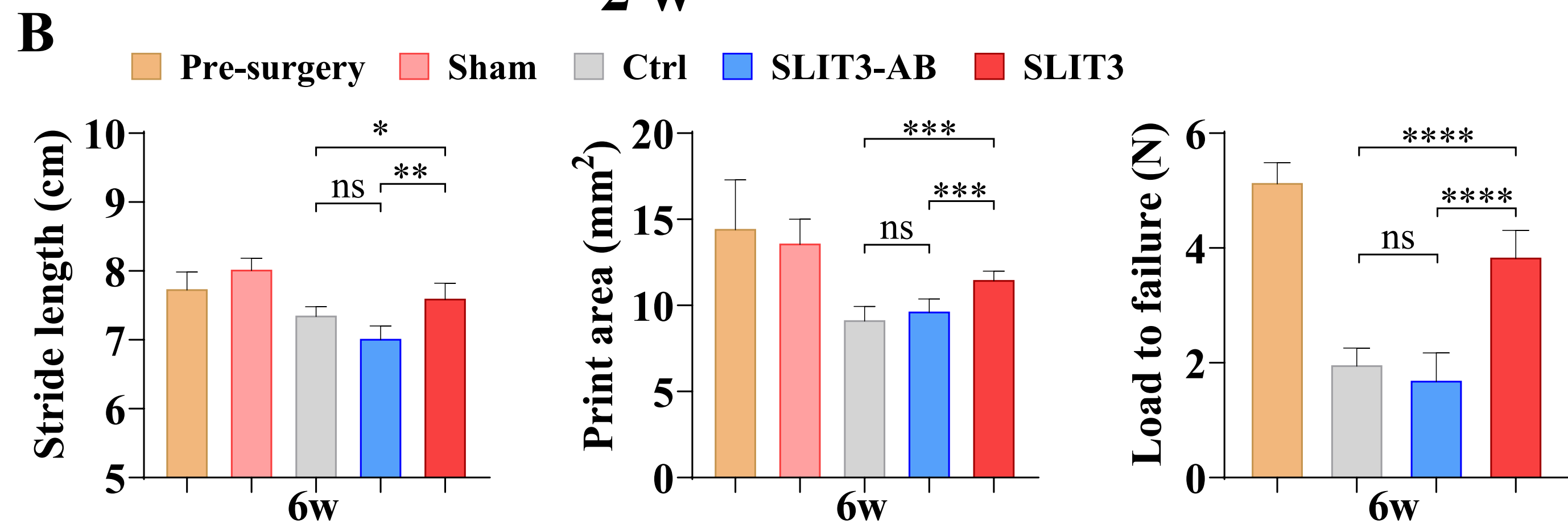
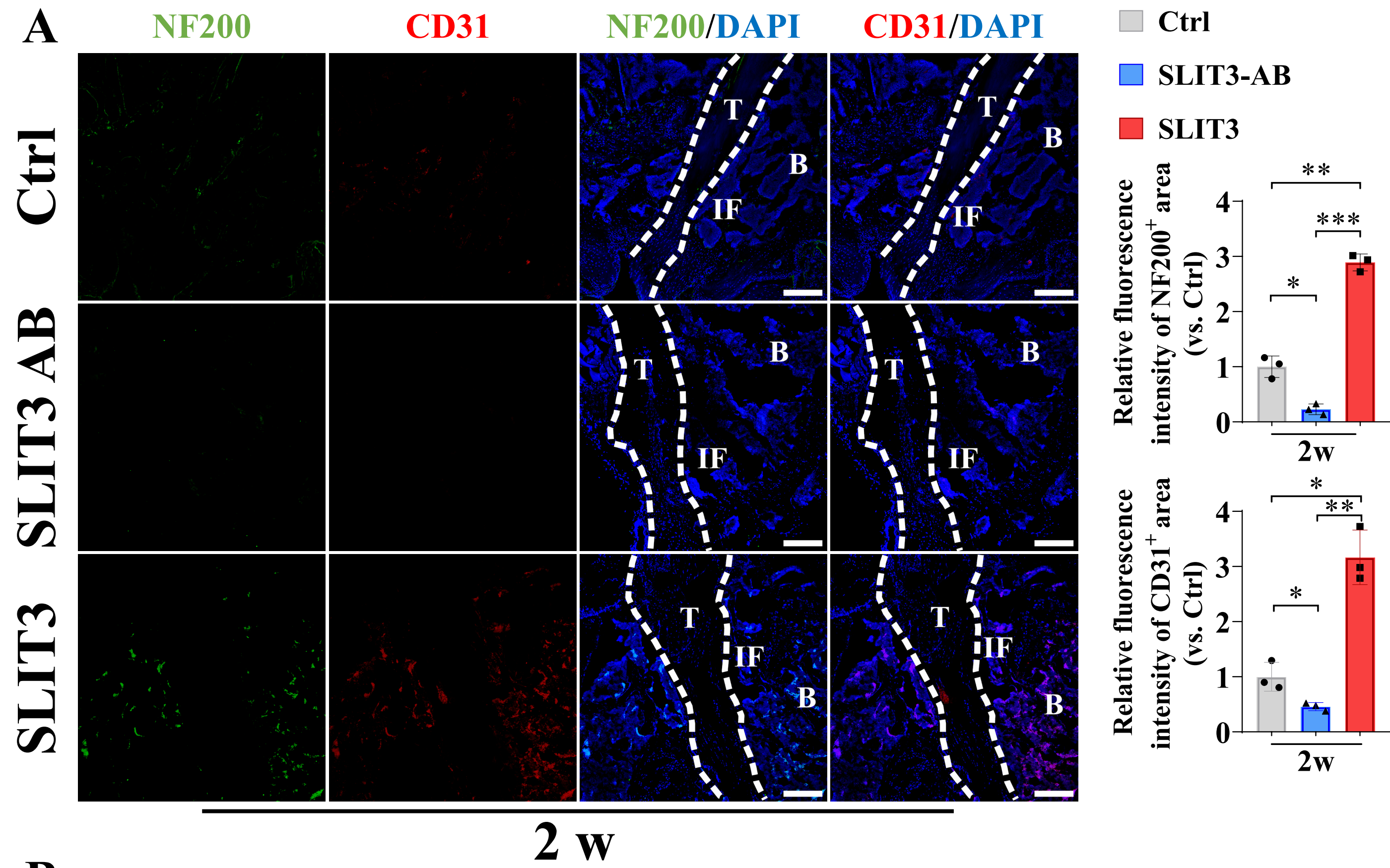


Fig. S15

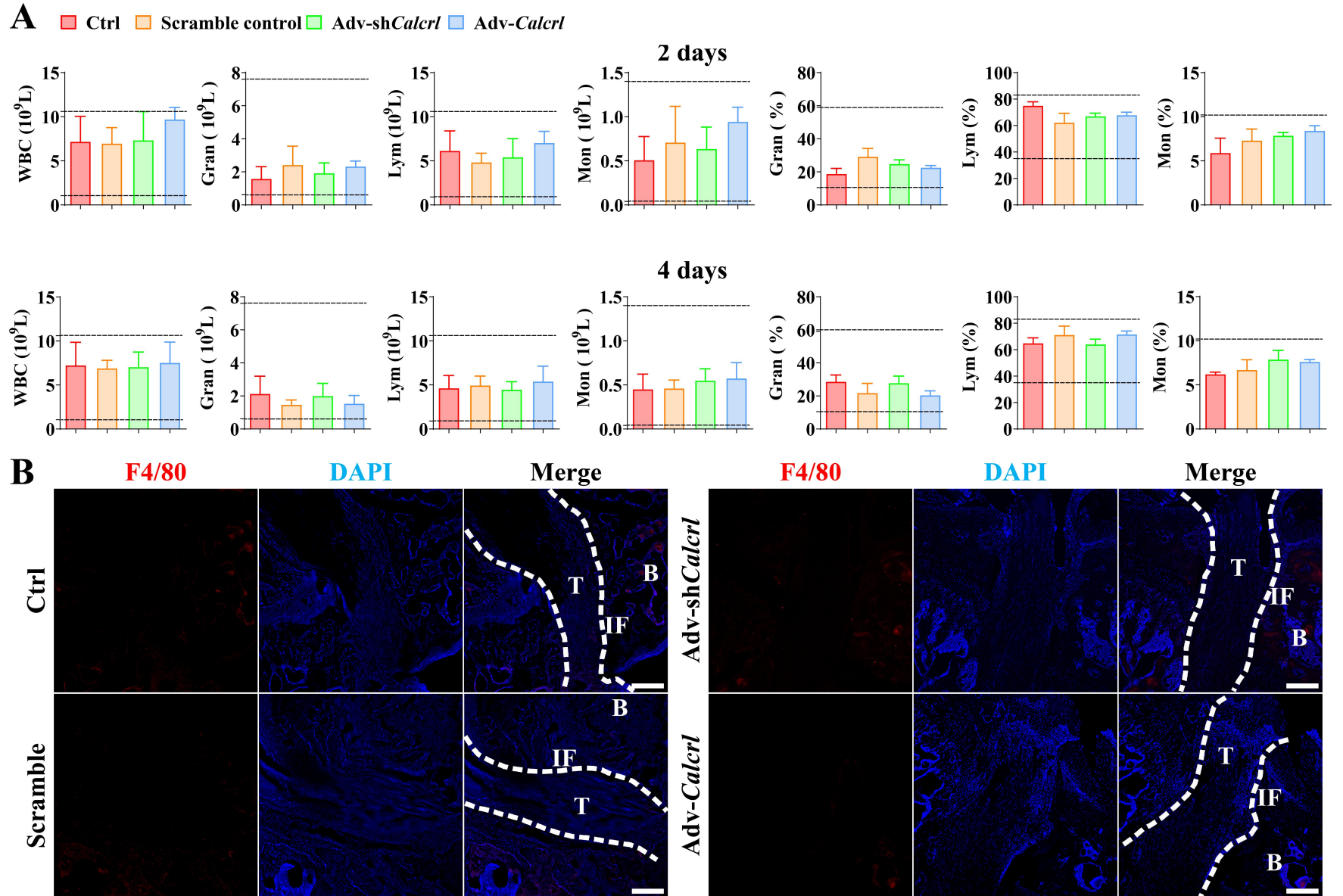


Table S1

Table S1. Primer sequences used for real-time quantitative reverse transcriptase polymerase chain reaction (qRT-PCR)

Gene name (ID)	Forward primer sequence (5'>3')	Reverse primer sequence (5'>3')
<i>Runx2</i> (ID: 12393)	CTGGCGGTGCAACAAG	CAGCGGAGGCATTTTCG
<i>Sp7</i> (ID: 170574)	CACCAGGTCCAGGCAACA	GAGCAAAGTCAGATGGGTAAGT
<i>Ocn</i> (ID: 12096)	GAGGGCAATAAGGTAGTGAA	CATAGATGCGTTTGTAGGC
<i>Opn</i> (ID: 20750)	ATGAGACCGTCACTGCTA	CTCAGTCCATAAGCCAAG
<i>Gapdh</i> (ID: 14433)	AAATGGTGAAGGTCGGTGTGAAC	AACAATCTCCACTTTGCCACTG
<i>Calcr1</i> (ID: 54598)	AATCATCCACCTCACGG	AAGTAGTTACAGCCCATCA
<i>RUNX2</i> (ID: 860)	CTACTATGGCACTTCGTCAGG	TCCATCAGCGTCAACACC
<i>SP7</i> (ID: 121340)	CCTCCTGCGACTGCCCTAA	GCGAAGCCTTGCCATACA
<i>OCN</i> (ID: 632)	GGCAGCGAGGTAGTGAAGA	CCTGAAAGCCGATGTGGT
<i>OPN</i> (ID: 6696)	GTTTCGCAGACCTGACATCC	TTCAACTCCTCGCTTTCCAT
<i>GAPDH</i> (ID: 2597)	CAGGAGGCATTGCTGATGAT	GAAGGCTGGGGCTCATTT
<i>CALCRL</i> (ID: 10203)	ATTGTGGTGGCCGTGTTT	TGGATAATGTAGAGGAGATGGGT
<i>Runx2</i> (ID: 367218)	CCCAACTTCCTGTGCTCC	AGTGAAACTCTTGCCTCGTC
<i>Sp7</i> (ID: 300260)	CCCTCTGCGAGACTCAACA	TGCCACCCACCTAACCAA
<i>Ocn</i> (ID: 25295)	GGGCAGTAAGGTGGTGAA	GTCCTGGAAGCCAATGTG
<i>Opn</i> (ID: 25353)	AGGAGTTTCCCTGTTTCTG	GTCTTCCCGTTGCTGTC
<i>Gapdh</i> (ID: 24383)	GACATGCCGCCTGGAGAAAC	AGCCCAGGATGCCCTTTAGT

Table S2

Table S2. Histologic Scoring Criteria for semi-quantitative analysis of the tendon-bone interface healing.

		0	1	2	3	4
Cellularity	Separation		Marked	Moderate	Mild	Minimal
Cellular morphology of interface tissue	Only fibrovascular tissue		Fibrous tissue with Sharpey-like fibers <50%	Fibrous tissue with Sharpey-like fibers $\geq 50\%$	Fibrocartilage with mature cartilage cells <50%	Fibrocartilage with mature cartilage cells $\geq 50\%$
Extent of fibrocartilage tissue surrounding tendon	Not visible		Slightly surrounded (<25%)	Partially surrounded (25%-50%)	Moderately surrounded (50%-75%)	Mostly surrounded ($\geq 75\%$)
Interface tissue transition from bone to tendon	Discontinuous		Continuous but slightly indistinct (<25%)	Partially indistinct (25-50%)	Moderately indistinct (50%-75%)	Mostly indistinct ($\geq 75\%$)
Tidemark	No visible		<25%	25-50%	50-75%	$\geq 75\%$