

## **Supporting Information for**

# Enhancing chimeric antigen receptor T cell therapy by modulating the p53 signaling network with $\Delta$ 133p53 $\alpha$

Christopher Roselle<sup>a,b</sup>, Izumi Horikawa<sup>c</sup>, Linhui Chen<sup>a</sup>, Andre S. Kelly<sup>a</sup>, Donna Gonzales<sup>a</sup>, Tong Da<sup>a</sup>, Nils Wellhausen<sup>a,b</sup>, Philipp C. Rommel<sup>a,d</sup>, Daniel Baker<sup>a,b,e</sup>, Megan Suhoski<sup>a</sup>, John Scholler<sup>a</sup>, Roddy S. O'Connor<sup>a</sup>, Regina M. Young<sup>a</sup>, Curtis C. Harris<sup>c</sup>, and Carl H. June<sup>a,d</sup>

<sup>a</sup>Center for Cellular Immunotherapies, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>b</sup>Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>c</sup> Laboratory of Human Carcinogenesis, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD 20892, USA

<sup>d</sup>Department of Pathology and Laboratory Medicine, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>e</sup>Cardiovascular Institute, Department of Medicine, Perelman School of Medicine, University of Pennsylvania, PA 19104, USA

\*Carl H. June

Email: cjune@upenn.edu

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#### Other supporting materials for this manuscript include the following:

N/A



**Fig. S1.** Evaluation of  $\Delta 133$ -CARs expressing  $\Delta 133p53a$  downstream of P2A linker. (A) Construct maps of  $\Delta 133p53a$ and mCherry2-expressing constructs with hCD19.BBz CAR upstream of P2A linker. (B) Tumor measurements over time from in vitro co-culture assay described in Figure 1D using CARs generated from healthy donor ND410. (C) Tumor measurements over time from in vitro co-culture assay described in Figure 1D using CARs generated from healthy donor ND410. (C) Tumor ND410. Note that aside from the data presented in this figure, all other data in this manuscript was generated using CAR constructs with the CD19-directed CAR downstream of the P2A linker.



**Fig. S2.** Characterization of  $\Delta$ 133-CARs during primary expansion. (A) Representative proliferation curves of total T cells during primary expansion and (B) cell volume for  $\Delta$ 133- and WT CARs. No meaningful or consistent differences were observed during primary expansion with respect to proliferation or cell volume. (C) Surface CAR median fluorescence intensity (MFI) at the end of primary expansion for the six normal donors used in Figure 1E and 1F, as indicated by the header above each graph (i.e. "ND307"). Histograms shown were gated as follows: lymphocytes/single cells/live/CAR+.



**Fig. S3.** Donor-specific tumor clearance curves for in vitro Nalm6 co-culture data shown in Figure 1E. Each row represents a different normal donor, indicated by the header above each graph. Each column represents a different E:T ratio, showing all five E:Ts that were evaluated: 1 to 2.5, 1 to 5, 1 to 7.5, 1 to 10, and 1 to 12.5. Each point represents the mean of two technical replicates. Error bars are not visible due to the size of each data point exceeding the range of the SEM.



**Fig. S4.** Donor-specific proliferation curves from in vitro Nalm6 co-culture data shown in Figure 1F. Each row represents a different normal donor, indicated by the header above each graph. Each column represents a different E:T ratio, showing all five E:Ts that were evaluated: 1 to 2.5, 1 to 5, 1 to 7.5, 1 to 10, and 1 to 12.5. Each point represents the mean of two technical replicates. Error bars are not visible due to the size of each data point exceeding the range of the SEM.



**Fig. S5.** RNAseq data for additional pairwise comparisons. (A) Volcano plot showing differentially expressed genes in WT and  $\Delta 133p53\alpha$ -expressing CAR T cells isolated prior to co-culture. All genes with an adjusted p-value (padj) less than or equal to 0.05 are highlighted, with genes in red upregulated in the WT CARs (i.e. downregulated in  $\Delta 133$ -CARs) and gene in blue upregulated in  $\Delta 133$ -CARs. Note that TP53 is reflective of ectopic  $\Delta 133p53\alpha$  expression. (B) Gene set enrichment analysis (GSEA) results for p53 Target Genes. Compared with  $\Delta 133$ -CARs pre-coculture,  $\Delta 133$ -CARs mid-coculture have an enrichment in expression of p53 target genes.

A ND307 hCD19.28z - 3x28 Bead Restimulation



Fig. S6.  $\Delta$ 133-CARs have reduced phospho-p53 (serine 15) expression in multiple contexts. (A)  $\Delta$ 133-CARs with a CD28 costimulatory domain have markedly reduced expression of p53-S15 when restimulated post-expansion with magnetic beads coated with anti-CD3 and anti-CD28 antibodies. (B)  $\Delta$ 133-CARs with a 41BB costimulatory domain exhibit reduced p53-S15 expression when co-cultured with Nalm6-GFP leukemia cells and (C) when restimulated post-expansion with with magnetic beads coated with anti-CD3 and anti-CD28 antibodies. For each context in A-C, multiple time points are shown, as indicated by the header above each histogram.



**Fig. S7.** Primary expansion characteristics of CLL-derived CAR T cells. (A) T cell proliferation during primary expansion for responding patient #0047 and non-responding patients #0012 and #0020. (B) Corresponding cell volumes during primary expansion at each time point for data shown in A. (C) Surface CAR expression demonstrates similar transduction efficiencies post-expansion for patient #0047.  $\Delta$ 133-CARs are shown in the top panel, with corresponding WT mCherry-expressing CARs in the bottom panel. The same is shown for patient #0012 in (D) and patient #0020 in (E).



**Fig. S8.** Δ133-CARs generated from CLL patient #0047 have reduced p53-S15 expression during in vitro Nalm6-GFP coculture.



**Fig. S9.** Patient-specific tumor clearance and T cell proliferation curves during in vitro Nalm6-GFP co-culture. In (A) and (B) are the individual tumor clearance curves for the aggregated data shown in Figure 4C for patients #0012 and #0020, respectively, as indicated by the header above each graph. In (C) and (D) are the corresponding T cell proliferation curves for patients #0012 and #0020, respectively, again indicated by the header above each graph.

Rank	ID	Gene.name	Up/Down	log2FoldChange	padj	Rank	ID	Gene.name	Up/Down
1	ENSG0000016845 3	HR	DOWN	-3.65	1.69E-10	51	ENSG0000013020 2	NECTIN 2	DOW N
2	ENSG0000020486 6	IGFL2	DOW N	-3.62	9.81E-04	52	ENSG000000480 9	SLC22A16	DOW N
3	ENSG0000022201 2	AC005481.1	DOWN	-3.37	2.48E-0 3	53	ENSG0000010293 5	ZNF42 3	DOW N
4	ENSG0000028096 9	RPS4Y 2	DOW N	-3.22	2.46E-0 3	54	ENSG0000022494 0	PRRT4	DOW N
5	ENSG0000026038 6	LINC0122 5	DOWN	-3.11	2.23E-04	55	ENSG0000016769 3	NXN	DOW N
6	ENSG0000020496 0	BLACE	DOWN	-2.78	8.54E-04	56	ENSG0000026706 0	PTGES3 L	DOW N
7	ENSG0000009986 4	PALM	DOWN	-2.56	3.17E-15	57	ENSG0000026143 4	AC021087.4	DOW N
8	ENSG000001541 3	DPEP 1	DOWN	-2.56	1.69E-10	58	ENSG0000016689 7	ELFN 2	DOW N
9	ENSG0000022941 7	NPM1P2 5	DOWN	-2.54	1.04E-0 2	59	ENSG0000017653 3	GNG7	DOW N
10	ENSG0000009986 6	MADCAM 1	DOWN	-2.46	1.04E-0 2	60	ENSG0000017528 7	PHYHD 1	DOW N
11	ENSG0000012732 4	TSPAN 8	DOWN	-2.45	1.73E-05	61	ENSG000000274 5	WNT16	DOW N
12	ENSG0000024521 3	AC105285. 1	DOW N	-2.44	3.44E-0 3	62	ENSG0000018217 5	RGMA	DOW N
13	ENSG0000014519 8	VWA5B 2	DOWN	-2.33	1.79E-0 2	63	ENSG0000027273 3	AP000345. 2	DOW N
14	ENSG000006981 2	HES 2	DOWN	-2.28	9.10E-0 3	64	ENSG000009593 2	SMIM2 4	DOW N
15	ENSG0000025710 8	NHLRC 4	DOWN	-2.25	3.74E-0 2	65	ENSG000006571 7	TLE2	DOW N
16	ENSG0000017322 7	SYT12	DOWN	-2.23	2.52E-04	66	ENSG000003586 2	TIMP 2	DOW N
17	ENSG0000018363 8	RP1L1	DOWN	-2.23	2.23E-04	67	ENSG0000017299 5	ARPP21	DOW N
18	ENSG0000010133 1	CCM2 L	DOWN	-2.23	4.63E-0 2	68	ENSG0000022597 8	HAR1A	DOW N
19	ENSG0000010119 8	NKAIN 4	DOWN	-2.21	2.15E-09	69	ENSG0000011255 9	MDFI	DOW N
20	ENSG0000015189 2	GFRA1	DOWN	-2.18	1.38E-06	70	ENSG0000026792 4	AC139769. 2	DOW N
21	ENSG0000016924 2	EFNA 1	DOWN	-2.17	8.55E-0 3	71	ENSG0000026773 1	AC005332. 3	DOW N
22	ENSG0000023562 1	LINC0049 4	DOWN	-2.15	4.81E-0 2	72	ENSG0000018470 2	SEPTIN 5	DOW N
23	ENSG0000027902 4	AC112255. 1	DOWN	-2.12	2.79E-0 2	73	ENSG000000653 4	ALDH3B 1	DOW N
24	ENSG0000010547 2	CLEC11 A	DOWN	-2.10	9.19E-11	74	ENSG0000013511 6	HRK	DOW N
25	ENSG0000017688 4	GRIN 1	DOWN	-2.08	6.25E-04	75	ENSG0000017480 7	CD248	DOW N
26	ENSG0000018573 7	NRG 3	DOWN	-2.08	3.80E-0 3	76	ENSG0000018762 1	TCL6	DOW N
27	ENSG0000018502 8	LRRC14 B	DOWN	-2.08	4.87E-07	77	ENSG0000017769 4	NAALADL 2	UP
28	ENSG0000021435 7	NEURL1 B	DOWN	-2.07	2.44E-09	78	ENSG0000016891 6	ZNF60 8	DOW N
29	ENSG0000009237 7	TBL1 Y	DOWN	-2.05	1.33E-0 5	79	ENSG0000016799 2	VWCE	DOW N
30	ENSG0000022685 9	LINC0205 1	DOWN	-2.04	2.39E-03	80	ENSG0000017667 8	FOXL 1	DOW N
31	ENSG0000017083 5	CEL	DOW N	-2.03	2.87E-0 3	81	ENSG000006760 6	PRKC Z	DOW N
32	ENSG0000017643 5	CLEC14 A	DOWN	-2.00	1.61E-07	82	ENSG0000019678 1	TLE1	DOW N
33	ENSG0000023588 8	AF064858.1	DOWN	-1.98	4.62E-0 5	83	ENSG0000018933 7	KAZN	DOW N
34	ENSG0000026115 9	AC112484. 3	DOWN	-1.97	3.13E-08	84	ENSG0000022513 8	SLC9A3-AS 1	DOW N
35	ENSG0000025431 9	AC246817.2	DOWN	-1.95	1.20E-0 3	85	ENSG0000016273 8	VANGL2	DOW N
36	ENSG0000023141 9	LINC0068 9	DOWN	-1.93	2.77E-03	86	ENSG0000010096 8	NFATC4	DOW N
37	ENSG0000016259 1	MEGF 6	DOWN	-1.93	9.60E-0 3	87	ENSG0000017105 6	SOX 7	DOW N
38	ENSG0000026366 7	AL035696. 3	DOWN	-1.88	9.37E-04	88	ENSG0000018077 7	ANKRD30 B	DOW N
39	ENSG0000010855 1	RASD 1	DOWN	-1.88	6.36E-0 6	89	ENSG0000016067 8	S100A 1	DOW N
40	ENSG0000016436 2	TERT	DOWN	-1.87	1.02E-04	90	ENSG0000009988 9	ARVCF	DOW N
41	ENSG0000017220 1	ID4	DOWN	-1.86	4.11E-07	91	ENSG000008707 6	HSD17B1 4	DOW N
42	ENSG0000004060 8	RTN4R	DOWN	-1.84	1.46E-04	92	ENSG0000024195 6	AC109466. 1	DOW N
43	ENSG0000022401 6	AC092681.1	DOWN	-1.83	3.15E-0 2	93	ENSG0000021197 8	IGHV5-78	DOW N
44	ENSG0000019842 9	ZNF69	DOWN	-1.82	2.99E-0 2	94	ENSG0000022705 1	C14orf13 2	DOW N
45	ENSG0000020597 8	NYNRIN	DOWN	-1.80	3.37E-0 6	95	ENSG000000439 9	PLXND 1	DOW N
46	ENSG0000010002 7	YPEL1	DOWN	-1.79	2.46E-04	96	ENSG0000010840 5	P2RX1	DOW N
47	ENSG0000025727 5	AL139020. 1	DOWN	-1.79	2.13E-0 3	97	ENSG0000015798 5	AGAP1	DOW N
48	ENSG0000016581 0	BTNL9	DOWN	-1.79	2.69E-0 2	98	ENSG0000013165 0	KREMEN 2	DOW N
49	ENSG0000013063 5	COL5A 1	DOWN	-1.78	2.65E-18	99	ENSG0000023812 1	LINC0042 6	DOW N
50	ENSG0000018086 1	LINC0155 9	DOWN	-1.76	1.04E-0 3	100	ENSG0000016957 5	VPREB 1	DOW N

#### **Table S1.** Top 100 differentially expressed genes with p-adjusted ≤ 0.05 sorted by the magnitude of log2 fold change.

log2FoldChange padj

7.31E-03

3.03E-02

2.46E-02

2.50E-03

1.61E-07

3.65E-04

7.70E-03

6.71E-09

9.78E-12

8.05E-06

2.28E-03

2.07E-02

4.07E-06

4.59E-06

7.92E-04

1.01E-15

5.80E-10

4.78E-04

7.70E-09

6.36E-06

2.60E-04

1.82E-04

4.25E-03

2.50E-03

8.42E-06

4.25E-03

2.71E-0 2

8.74E-11

1.74E-04

2.83E-07

8.73E-05

2.67E-03

1.60E-05

3.35E-02

9.08E-04

4.28E-06

1.37E-05

6.25E-04

1.63E-02

2.23E-03

6.40E-03

3.13E-05

4.89E-07

2.08E-06

1.69E-10

1.83E-03

1.00E-06

1.68E-03

3.49E-05

1.49E-05

-1.75

-1.75

-1.75

-1.74

-1.73

-1.73

-1.73

-1.72

-1.72

-1.70

-1.67

-1.67

-1.67

-1.66

-1.65

-1.65

-1.64

-1.63

-1.63

-1.62

-1.61

-1.61

-1.61

-1.61

-1.60

-1.60

1.59

-1.59

-1.57

-1.57 -1.57

-1.56

-1.56

-1.56

-1.55

-1.55

-1.54

-1.54

-1.54

-1.53

-1.53

-1.52

-1.52

-1.52

-1.51

-1.51

-1.51

-1.51

-1.50

-1.49

### Table S2. Gene sets enriched in $\Delta$ 133-CARs and WT CARs with False discovery rate (FDR) < 0.1

Gene Set	NES	NOM p-va l	FDR q-val
GOBP_RIBOSOME_BIOGENESIS	2.08	0	0
HALLMARK_MYC_TARGETS_V2	2.07	0	0
GOBP_RNA_MODIFICATION	2.01	0	0
	1.99	0	0
GOBP CELULAR RESPONSE TO HEAT	1.95	0	0
GOBP_REGULATION_OF_TELOMERE_MAINTENANCE	1.93	0	0
GOBP_RNA_SPLICING_VIA_TRANSESTERIFICATION_REACTIONS	1.91	0	0
GOBP_POSITIVE_REGULATION_OF_TELOMERE_MAINTENANCE	1.88	0	0.001
	1.87	0	0.001
	1.85	0	0.001
GOBP TINA PROCESSING	1.85	0	0.001
GOBP_RNA_METHYLATION	1.85	0	0.001
GOBP_RRNA_METABOLIC_PROCESS	1.83	0	0.002
GOBP_MITOCHONDRIAL_TRANSLATION	1.82	0	0.003
GOBP_REGULATION_OF_MRNA_PROCESSING	1.81	0	0.003
GOBP_POSITIVE_PEGULATION_OF_URROWDSUME_URGANIZATION	1.81	0	0.003
	1.79	0	0.004
GOBP REGULATION OF TRANSCRIPTION BY RNA POLYMERASE III	1.78	0	0.005
GOBP_MITOCHONDRIAL_RNA_METABOLIC_PROCESS	1.75	0	0.011
GOBP_REGULATION_OF_HETEROTYPIC_CELL_CELL_ADHESION	1.74	0	0.016
GOBP_REGULATION_OF_TRANSLATIONAL_INITIATION	1.74	0.005	0.015
GOBP_MATURATION_OF_5_85_RRNA	1.74	0.007	0.015
GOBP_REGULATION_OF_RIVE_SFLCING	1.73	0	0.015
	1.73	0	0.015
GOBP_REGULATION_OF_TELOMERE_MAINTENANCE_VIA_TELOMERE_LENGTHENING	1.73	0.003	0.016
GOBP_TYROSINE_PHOSPHORYLATION_OF_STAT_PROTEIN	1.72	0	0.017
GOBP_LYMPHOCYTE_CHEMOTAXIS	1.72	0	0.016
GOBP_MATURATION_OF_SSU_RRNA	1.72	0.002	0.017
	1./1	0.002	0.021
GOBP POSITIVE REGULATION OF TELOMERE MAINTENANCE VIA TELOMERE LENGTHENING	1.71	0.004	0.021
GOBP_SPLICEOSOMAL_COMPLEX_ASSEMBLY	1.71	0	0.021
GOBP_ALTERNATIVE_MRNA_SPLICING_VIA_SPLICEOSOME	1.7	0.006	0.023
GOBP_RNA_S_END_PROCESSING	1.7	0.001	0.023
GOBP_T_HELPER_17_CELL_DIFFERENTIATION	1.68	0.004	0.031
GOBP_POSITIVE_REGULATION_OF_MRNA_METABOLIC_PROCESS	1.68	0	0.033
	1.07	0.002	0.041
GoBP MRIA MODIFICATION	1.66	0.005	0.042
GOBP_POSITIVE_REGULATION_OF_TRANSCRIPTION_BY_RNA_POLYMERASE_III	1.66	0.002	0.041
GOBP_POSITIVE_REGULATION_OF_RNA_SPLICING	1.66	0.007	0.042
GOBP_C04_POSITIVE_OR_C08_POSITIVE_ALPHA_BETA_T_CELL_LINEAGE_COMMITMENT	1.66	0.002	0.045
GOBP_NEGATIVE_REGULATION_OF_SIGNALING_RECEPTOR_ACTIVITY	1.64	0.004	0.057
	1.04	0.002	0.037
GOBP POSITIVE REGULATION OF TELOMERASE ACTIVITY	1.64	0.005	0.059
GOBP_RNA_CAPPING	1.63	0.006	0.06
GOBP_CLEAVAGE_INVOLVED_IN_RRNA_PROCESSING	1.63	0.005	0.061
GOBP_TELOMERASE_RNA_LOCALIZATION	1.63	0.008	0.061
GOBP_POSITIVE_REGULATION_OF_TYROSINE_PHOSPHORYLATION_OF_STAT_PROTEIN	1.63	0.005	0.063
GOBY REGULATION OF KRNA-PROLESSING	1.62	0.004	0.066
GOBP MATURATION OF LSU RINA	1.62	0.005	0.07
GOBP_CHRONIC_INFLAMMATORY_RESPONSE	1.62	0.001	0.069
GOBP_NEGATIVE_REGULATION_OF_CYTOKINE_PRODUCTION_INVOLVED_IN_IMMUNE_RESPONSE	1.62	0.014	0.068
GOBP_TELOMERE_MAINTENANCE_IN_RESPONSE_TO_DNA_DAMAGE	1.62	0.013	0.067
GODE_TRANSLATIONAL_TERMINATION	1.62	0.004	0.066
GUBP_MATUKATUON_OF_S_85_KKINA_FKOM_TKICISTKUNIL_KKINA_TKANSCKIPI_SSU_KKINA_S_85_KKINA_LSU_KKINA GORD_PEGILIATION_OF_HILMORALI IMMILINE PESOPONSE	1.61	0.011	0.071
	1.6	0.015	0.077
GOBP_GPI_ANCHOR_METABOLIC_PROCESS	1.6	0.011	0.078
GOBP_POSITIVE_REGULATION_OF_TRANSLATIONAL_INITIATION	1.6	0.007	0.078
GOBP_RNA_PHOSPHODIESTER_BOND_HYDROLYSIS_EXONUCLEOLYTIC	1.6	0.015	0.077
GOBP_IRON_SULFUR_CLUSTER_ASSEMBLY	1.59	0.009	0.084
GORP ISOTIFE_SWITCHING_TO_IGG_SUITES	1.59	0.008	0.093
GOBP T HELPER CELL LINEAGE COMMITMENT	1.59	0.006	0.092
GOBP_MONOUBIQUITINATED_PROTEIN_DEUBIQUITINATION	1.58	0.017	0.091
GOBP_MITOCHONDRIAL_RNA_PROCESSING	1.58	0.015	0.093
GOBP_DNA_TEMPLATED_DNA_REPLICATION_MAINTENANCE_OF_FIDELITY	1.58	0.011	0.093
GOBP_REGULATION_OF_TELOMERE_MAINTENANCE_IN_RESPONSE_TO_DNA_DAMAGE	1.58	0.013	0.093
UUDY_KINA_3_END_YKULESSING GORP REGULATION OF T HEIPER 17 TYPE IMMUNE RESPONSE	1.58	0.01.2	0.092
GOBP_REGULATION_OF_MITOCHONDRIAL_GENE_EXPRESSION	1.58	0.012	0.094
GOBP_ENDONUCLEOLYTIC_CLEAVAGE_INVOLVED_IN_RRNA_PROCESSING	1.58	0.011	0.095
GOBP_REGULATION_OF_MITOCHONDRIAL_TRANSLATION	1.57	0.013	0.097
GOBP_ENDODERMAL_CELL_DIFFERENTIATION	-1.44	0	0.091