

Expanded View Figures

Figure EV1. The mouse model of CCl₄-induced liver fibrosis.

(A, B) Immunofluorescence images (A) and quantification (B) of the α -SMA positive areas in the liver of mice treated with CCl₄ for 0-4 months (n = 6 mice). Scale bar, 20 µm. (C, D) Representative images of Sirius red staining and H&E staining in the liver of C57BL6 mice treated with CCl₄ for 2 months (C). The Image J software was used to quantify the collagen-positive areas (D), n = 6 mice). Scale bars for Sirius red staining and H&E staining, 200 µm. Scale bar for liver, 1 cm. (E, F) The activities of AST (E) and ALT (F) in the serum were analyzed in CCl₄-treated mice for 2 months (n = 4 mice). Data information: Data are presented as mean ± SD. Statistical significance was determined by unpaired two-tailed Student's *t* test. ns not significant; **P < 0.001. Related to Fig. 1. Source data are available online for this figure.





Figure EV2. Persistent presence of primary cilia on BECs and PVCs during CCl₄-induced liver fibrosis.

(A-C) Immunofluorescence images of primary cilia on hepatocytes (A), BECs (B), and PVCs (C) from CCI_4 -treated mice (n = 10 mice). Scale bars, 10 µm. (D) Quantification of the percentage of ciliated cells in hepatocytes, PVCs, and BECs from mice described in (A-C) (n = 10 mice). To quantify the percentage of ciliated cells (D), >200 cells from six fields were analyzed for each mouse. The same group of mice (n = 10 mice for each time point) were used for Fig. 1A-D and Fig. EV2. Data information: Data are presented as mean ± SD. Statistical significance was determined by one-way ANOVA. ns not significant. Related to Fig. 1. Source data are available online for this figure.



Figure EV3. Whole-body deficiency in IFT88 exacerbates CCl_a-induced liver fibrosis over a 2-month period.

(A-C) Immunofluorescence images (A) and quantification of the levels of α -SMA (B) and vimentin (C) in the liver of *Ift88^{II/II}*; *Ubc-CreER* mice treated with CCl₄ or corn oil (vehicle) for 2 months (n = 6 mice). Nuclei were stained with DAPI (blue). Scale bar, 50 µm. (D, E) CCl₄-induced liver fibrosis in *Ift88^{II/II}*; *Ubc-CreER* mice treated with CCl₄ mice was examined with Sirius red staining and H&E staining (D), and the percentage of collagen-positive areas was quantified (E) (n = 6 mice). Scale bars for Sirius red staining and H&E staining, 200 µm. Scale bar for liver, 1 cm. (F, G) Examination of the activities of AST (F) and ALT (G) in the serum of *Ift88^{II/II}*; *Ubc-CreER* mice (n = 4 mice). (H-L) The mRNA levels of IL-6 (H), IFNAR2 (I), TNF- α (J), COL1A1 (K), and α -SMA (L) in liver tissues were measured by quantitative RT-PCR (n = 4 mice). (M, N) Immunoblotting (M) and quantification (N) of the levels of IFT88 and α -SMA in the liver of *Ift88^{II/II}*; *Ubc-CreER* mice treated with CCl₄ or vehicle (n = 4 mice). Data information: Data are presented as mean ± SD. Statistical significance was determined by two-way ANOVA with post hoc tests. ns not significant; **P* < 0.05, ***P* < 0.001. Related to Fig. 2. Source data are available online for this figure.



Figure EV4. Whole-body deficiency in IFT88 exacerbates CCI₄-induced liver fibrosis over a 4-month period.

(A-C) Immunofluorescence images (A) and quantification of α -SMA (B) and vimentin (C) in the liver of *Ift88^{II/II}* and *Ift88^{II/II};Ubc-CreER* mice treated with CCl₄ or corn oil (vehicle) for 4 months (n = 6 mice). Scale bar, 50 µm. (D, E) CCl₄-induced liver fibrosis in *Ift88^{II/II}* and *Ift88^{II/II};Ubc-CreER* mice for 4 months was evaluated with Sirius red staining and H&E staining (D). The Image J software was used for the quantification of collagen-positive areas (E) (n = 6 mice). Scale bars for Sirius red staining and H&E staining, 200 µm. Scale bar for liver, 1 cm. (F, G) The activities of AST (F) and ALT (G) in the serum were analyzed in *Ift88^{II/II}* and *Ift88^{II/II}* (*IICC)* (n = 4 mice). (H–L) The mRNA levels of IL-6 (H), IFNAR2 (I), TNF- α (J), COL1A1 (K), and α -SMA (L) in liver tissues were measured by quantitative RT-PCR (n = 4 mice). Data information: Data are presented as mean ± SD. Statistical significance was determined by two-way ANOVA with post hoc tests. ns, not significant; **P* < 0.05, ***P* < 0.01, ****P* < 0.001. Related to Fig. 2. Source data are available online for this figure.

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Figure EV5. TGF- β promotes IFT88 degradation during LX-2 cell activation.

(A, B) Immunofluorescence images (A) and quantification of the density of cilia (B) in LX-2 cells treated with TGF- β for 24 h (n = 6 independent experiments). To quantify the percentage of ciliated cells (B), >140 cells were analyzed for each experiment. Scale bar, 10 µm. (C) IFT88 mRNA expression was measured by quantitative RT-PCR after TGF- β treatment for 24 h in LX-2 cells (n = 3 independent experiments). (D-F) Immunoblotting (D) of IFT88 and α -SMA in LX-2 cells treated with TGF- β , and the levels of IFT88 (E) and α -SMA (F) were quantified by densitometry (n = 3 independent experiments). (G, H) The effect of TGF- β on the half-life of IFT88 in LX-2 cells treated with CHX (20 mg/mL) was examined by immunoblotting (G), and the protein half-life curves were obtained (H) (n = 3 independent experiments). (I, J) LX-2 cells were treated with TGF- β for 24 h and then treated with MGI32 (5 mM) for 12 h. The levels of IFT88 and GAPDH were examined by immunoblotting (I), and the level of IFT88 was determined by densitometry (J) (n = 3 independent experiments). Data information: Data are presented as mean ± SD. Statistical significance was determined by one-way ANOVA with post hoc tests (E, F) or unpaired two-tailed Student's *t* test (B, C, H, J). ns not significant; **P* < 0.01, ****P* < 0.001. Related to Fig. 3. Source data are available online for this figure.