

Supplementary Material 1473

S1 Additional Data 1474

In addition to the data utilized in the main body of our paper, we employed several publicly available datasets in the supplementary material. These datasets can be accessed via the following websites or using the provided accession numbers: (1) Stereo-seq data[48]. 1475
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S2 Comparison baselines 1481

Our experimental results demonstrate that our approach achieves very high accuracy. However, it is important to note that previous literature on these methods has some limitations: 1482
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1. Previous spatial transcriptomics analysis methods have not been able to enhance gene expression to single-cell resolution without using single-cell RNA-seq data. 1484
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2. Certain deconvolution methods use public single-cell RNA-seq references, including the Human Cell Atlas, the BRAIN Initiative Cell Census Network (BICCN), and the Human BioMolecular Atlas, to address the problem of low resolution in spatial transcriptomics. However, these methods are prone to incomplete identification of cell types due to batch effects and tissue heterogeneity in samples. Additionally, the accuracy of deconvolution may be impacted by the different perturbations that affect single-cell references and spatial transcriptomics. 1490
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To overcome these limitations, we propose a novel framework called TransformerST, which utilizes a Transformer-based approach to associate the heterogeneity of local gene expression properties and reveal the structural relationships at nearly single-cell resolution. We have also included a table that compares the features of various spatial transcriptomics analysis methods, highlighting their respective strengths and limitations (Supplementary Table 1). 1498
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S3 Tissue identification with Stereo-seq technology 1506

We use the proposed method to a Stereo-seq dataset derived from mouse olfactory bulb tissue. Nowadays, stereo-seq is one of the most promising new spatial holography methods available. DNA nanosphere-patterned array chips provide spatial resolution down to the subcellular level. The information in this study was split down to a cellular level (14 m) for clarity. The rostral migratory stream (RMS), granular cell layer (GCL), endothelial cell layer (GCL), internal plexiform layer (IPL), filamentous cell layer (MCL), exterior plexiform layer (EPL), and olfactory nerve layer were all labeled in DAPI-stained pictures of 1507
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1519 **Supplementary Table 1** Comparison between TransformerST with baselines.

1520	Methods	Objective	Super-resolution	Reference-free	Histology image
1521	TransformerST	Clustering, super-resolution	Single-cell	Yes	Yes
1522	SpaGCN	Clustering	Original	No	Yes
1523	BayesSpace	Clustering, super-resolution	Multi-cellular	Yes	No
1524	CCST	Clustering	Original	No	No
1525	STAGATE	Clustering	Original	No	No
1526	DeepST	Clustering	Original	No	Yes
1527	stLearn	Clustering	Original	No	No
1528	STdeconvolve	Deconvolution	Multi-cellular	Yes	No

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1537 the coronal mouse olfactory bulb by Fu et al.[48]. The results are shown in Sup-
 1538 plementary Supplementary Fig. 1. The results derived from TransformerST
 1539 align with the Allen Reference Atlas, as depicted in Supplementary Supple-
 1540 mentary Fig. 1b, demonstrating the method’s precision and consistency. In
 1541 Supplementary Supplementary Fig. 1c, TransformerST, utilizing the original
 1542 spot resolution, yielded results that closely mirrored those of the Allen Re-
 1543 ference Atlas. This showcases its capacity to capture intricate gene expression
 1544 patterns and cellular structures.

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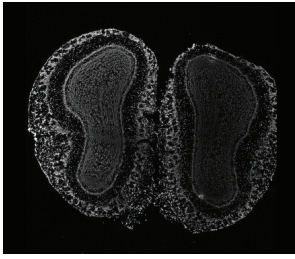
1546 Furthermore, in Supplementary Supplementary Fig. 1d, TransformerST
 1547 effectively reconstructed the original resolution of spatial transcriptomics data
 1548 using downsampled inputs. The outcomes closely resembled those in the Allen
 1549 Reference Atlas, underscoring TransformerST’s robustness and flexibility in
 1550 managing various data resolutions and ensuring accurate results irrespective
 1551 of input quality.

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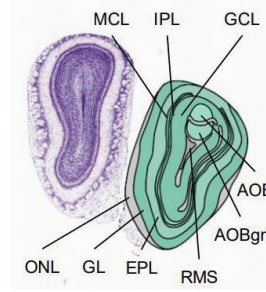
1553 The compatibility of TransformerST with the Allen Reference Atlas can
 1554 also be attributed to its sophisticated algorithms. These enable the precise
 1555 identification of spatial gene expression patterns and facilitate the extraction of
 1556 biologically significant information. Additionally, the method’s robust feature
 1557 extraction and pattern recognition capabilities aid in effectively distinguishing
 1558 between different cell types, tissue structures, and gene expression patterns.
 1559 Moreover, TransformerST’s versatility allows it to be applied across multi-
 1560 ple platforms and datasets, maintaining consistent performance under a wide
 1561 array of experimental conditions. This adaptability ensures the method’s reli-
 1562 ability and relevance, further reinforcing its alignment with the established
 1563 Allen Reference Atlas.

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1563 In conclusion, the concordance between the results obtained from Trans-
 1564 formerST and the Allen Reference Atlas can be attributed to the method’s



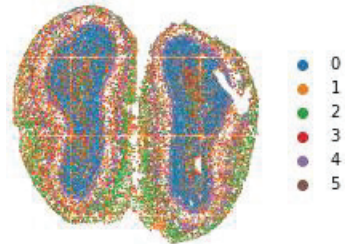
(a) Original Image



(b) Allen Reference Atlas



(c) TransformerST



(d) Super-resolution

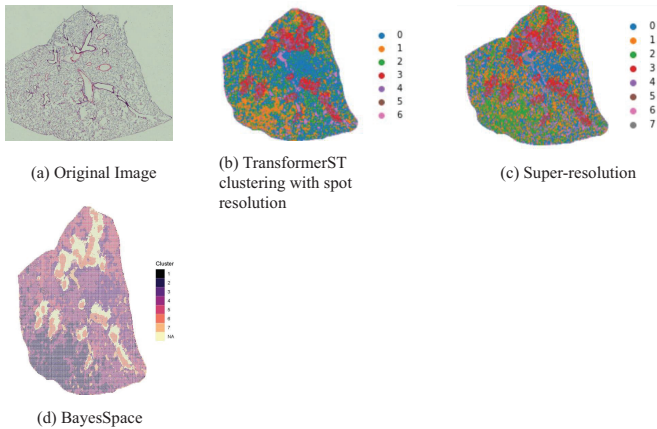
Supplementary Figure 1 TransformerST improves the identification of known tissue structures in the olfactory bulb tissue

precision, consistency, adaptability, and its ability to effectively manage various data resolutions. These attributes position TransformerST as a valuable tool in spatial transcriptomics analysis, offering researchers reliable and biologically pertinent insights.

We applied the proposed methods to a segment of mouse lung tissue profiled by Stereo-seq and found that the spatial regions identified by the method showed strong concordance with the patterns of the original image (Supplementary Supplementary Fig. 2). This underscores the effectiveness of our approach in accurately delineating tissue structures and gene expression patterns, a critical aspect for understanding the complex biological processes within the sample. The method's ability to yield results consistent with expert annotations underscores its potential as a valuable tool for spatial transcriptomics analysis across diverse tissue types and experimental conditions. We also showcase the results of the BayesSpace method in Supplementary Supplementary Fig. 2d, which uses the original data for tissue type clustering. Our simulation outcome aligns with the results of BayesSpace. Importantly, our simulation results for super-resolution with downsampled data are capable of producing more refined patterns that closely mirror the original image.

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1628 **Supplementary Figure 2** TransformerST improves the identification of known tissue
1629 structures in the mouse lung tissue

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1631 This further attests to the effectiveness of our proposed approach in handling
1632 various resolutions and providing detailed spatial transcriptomics analysis.

1633 For both datasets, our proposed method produced visually superior tissue
1634 identification and super-resolution results when compared to other methods.
1635 In the two figures, 'TransformerST' denotes the clustering result at the origi-
1636 nal resolution, while 'Super-resolution' refers to the super-resolution outcome
1637 achieved by TransformerST. BayesSpace denotes the result achieved when data
1638 at the original resolution is employed for tissue type clustering.

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1640 S4 Enhanced Gene Expression Prediction at 1641 Sub-cellular Resolution 1642

1643 **Enhanced Gene expression prediction at sub-cellular resolution in**
1644 **breast cancer data HER2+.** To evaluate the tissue identification and super-
1645 resolution performance in predicting gene expression at sub-cellular resolution
1646 using histology images, we employed the leave-one-out method (36 fold) using
1647 HER2+ breast cancer data, which includes 36 tissue sections from 8 patients.
1648 This approach underscores the effectiveness in gene expression prediction and
1649 super-resolution. For the leave-one-out evaluation, 32 sections were utilized to
1650 train the tissue identification and super-resolution model, with the remaining
1651 sections reserved for evaluation. The results of this approach are represented
1652 as TransformerST. We also assessed the super-resolution performance at the
1653 sub-cellular level, referred to as Super-resolution.

1654 Manual annotation of three tissue sections was included for evaluating
1655 clustering accuracy [49]. We compared our proposed method, TransformerST,
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Supplementary Table 2 Average correlation of predicted expression for top 50 most highly variable genes (HVG) compared to ground truth expressions on held out dataset

Methods	HVG
TransformerST	0.226
BLEEP	0.168
TCGN	0.151

with TCGN [37] and BLEEP [36] for gene expression prediction using three tissue sections, as detailed in Supplementary Fig. 3. TCGN and BLEEP, in comparison, demonstrated less effective gene prediction performance. The leave-one-out evaluation showed a higher correlation with biological interpretation.

TransformerST markedly improved clustering accuracy (ARI) for the evaluated sections out of the 32 samples, significantly outperforming TCGN and BLEEP. This advancement in clustering accuracy by TransformerST is evident in the results for sample B1 (Supplementary Fig. 3a), where TransformerST achieved the highest clustering accuracy (ARI of 0.308 for TransformerST compared to lower ARIs for TCGN and BLEEP). Similarly, Supplementary Fig. 3b displays the superiority of TransformerST (ARI=0.325) over TCGN and BLEEP. These results might be due to substantial gene expression differences among patients, which led to the leave-one-out evaluation by TransformerST attaining superior tissue identification performance, as shown in Supplementary Fig. 3). Supplementary Table 2 displays the performance comparison of BLEEP, TCGN, and TransformerST in predicting the top 50 most highly variable genes (HVG). In this assessment, the expression profiles predicted by TransformerST showed the strongest correlation with the actual data among all gene sets.

The enhanced single-cell resolution results further demonstrate TransformerST could predict the biological meaningful patterns as in the manual annotations. While it is hard to estimate the ARI for the super-resolution result, the study is visually consistent with the manual annotations by pathologists in the spatial domain (Supplementary Fig. 3).

S5 Meta Gene and SVGs Analysis with DLPFC and IDC Samples

To further demonstrate that TransformerST could explore the biological relevance, we detected the spatial variable genes and meta genes for LIBD human dorsolateral prefrontal cortex (DLPFC) data and IDC sample. As shown in Supplementary Fig. 4a, SVGs and their corresponding meta gene show similar spatial patterns for human DLPFC samples at spot resolution. For example, TMSB10 is enriched in cluster 0 of tissue sample 151508. The combination of meta genes (TMSB10+MBP-MT-CO2) shows the strengthened spatial patterns in the neighboring regions. GFAP is enriched in cluster

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1703 2 of tissue sample 151508, its corresponding meta gene is GFAP+SNORC-
1704 TMSB10+CDT3-MBP, which is also spatially correlated with the SVGs of
1705 cluster 2 in the histology image.

1706 As illustrated in Supplementary Fig. 4b, boxplots display Moran's I and
1707 Geary's C values for spatially variable genes (SVGs) identified by Transform-
1708 erST, SpatialPCA [38], and Vesalius[39], offering a comparative analysis of
1709 the performance of these methods in detecting spatial patterns in brain tissue
1710 slice 151508. Notably, the Moran's I and Geary's C values for SVGs detected
1711 by SpatialPCA and Vesalius are lower than those identified by Transform-
1712 erST, indicating a weaker presence of spatial patterns in their results. The
1713 experimental results with different tissue samples and different cluster domains
1714 demonstrate TransformerST could mark specific gene-expressed regions for
1715 different cluster domains.

1716 To illustrate how TransformerST works for different tissue samples, We
1717 detected the spatial variable genes and meta gene for IDC samples at nearly
1718 single-cell resolution. As shown in Supplementary Fig. 4c, TransformerST
1719 detected single SVGs (ACADSB) for cluster 2. Its corresponding meta gene was
1720 defined as ACADSB+NME2-MUC1+ATP5MPL-CD74+LAPTM4B-CRIP1.
1721 TransformerST detected DEGS1 SVG for cluster 3, which accords with
1722 its meta gene DEGS1+RPS18-CXCL14+AGR2-MGP+CSTA-NEAT1 visu-
1723 ally. TTLL12 is enriched in cluster 4 with its corresponding meta gene as
1724 TTLL12+HMGN2-MALAT1+KRT8-SLC9A3R1.

1725 The detection outcomes for metagenes and SVGs demonstrate that Trans-
1726 formerST can effectively identify heterogeneity among spatial domains and
1727 predict boundaries not recognized by current state-of-the-art methods. These
1728 findings highlight TransformerST's ability to better uncover spatial patterns
1729 utilizing a graph transformer network. The detection outcomes for metagenes
1730 and SVGs showcase that TransformerST not only effectively identifies het-
1731 erogeneity among spatial domains, but also accurately predicts boundaries
1732 that remain undetected by current state-of-the-art methods. These findings
1733 not only highlight TransformerST's superior performance in uncovering spa-
1734 tial patterns but also emphasize its ability to leverage a graph transformer
1735 network for a more comprehensive understanding of spatial transcriptomics
1736 data. Consequently, TransformerST emerges as a valuable tool for researchers
1737 aiming to analyze complex spatial relationships within biological samples.

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1739 **S6 Tissue identification and super-resolution in** 1740 **breast cancer block at nearly single-cell** 1741 **resolution with Xenium technology.**

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1743 We introduce the cutting-edge Xenium In Situ technology, which has a wide
1744 field of view and the unique ability to combine gene expression and histo-
1745 logical pictures (H&E and IF staining) in a single tissue segment. Using this
1746 method, Chromium scFFPE-seq data were generated from 2 x 25 μ m FFPE
1747 sections obtained from a breast cancer block (stage II-B, ER+/PR-/HER2+).
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These sections subjected to scFFPE-seq were situated across the tissue sections 1749
 employed for Visium and Xenium procedures. 1750

The Visium CytAssist and Xenium platforms were employed to analyze 1751
 5 μm tissue sections adjacent to those used for scFFPE-seq. Prior to imag- 1752
 ing, sections subjected to Visium were deparaffinized and stained with H&E. 1753
 The glass slides containing tissue sections were then processed using a Visium 1754
 CytAssist instrument to transfer analytes to Visium slides. Using the same 1755
 probe set as in scFFPE-seq, 18,536 genes targeted by 54,018 probes were quan- 1756
 tified for sections subjected to Visium. The median number of genes in the 1757
 Visium data set was 5,712, and dimensionality reduction revealed 17 spatial 1758
 clusters (the same number of clusters as the scFFPE-seq data). The molecular 1759
 subtypes of ductal carcinoma in situ (DCIS) (hence referred to as DCIS #1 1760
 and DCIS #2) and invasive tumors were each identified as separate clusters 1761
 by scFFPE-seq, and Visium was able to determine their exact locations. 1762

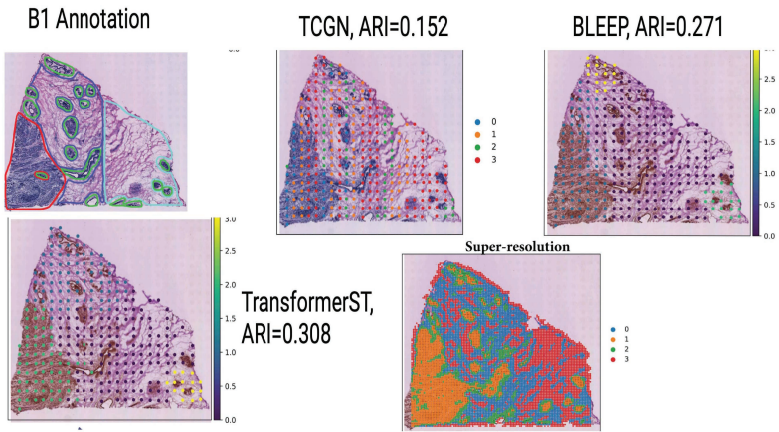
We use the following pipeline to show the performance of the proposed 1763
 method. Firstly, we choose the region of interest using the Visium image. 1764
 Then we obtain the corresponding higher-resolution image and gene expression 1765
 using Xenium technology. We perform cell segmentation. To allocate mRNAs 1766
 to cells, facilitating subsequent analysis and integration with Chromium and 1767
 Visium data, the spatial boundaries between cells and mRNA transcripts need 1768
 to be determined. Initially, DAPI images were employed to identify nuclei 1769
 using a neural network. Subsequently, each nucleus was expanded outward up 1770
 to a maximum distance of 15 μm or until encountering the border of another 1771
 cell. With the original Visium image, we perform cell-type clustering. Next, 1772
 we employ the proposed method to perform super-resolution, utilizing the 1773
 clustering and cell segmentation outcomes. The processed steps are shown in 1774
 Supplementary Fig. 5a. 1775

As shown in Supplementary Fig. 5b, similar to Xenium, the proposed 1776
 method is able to pinpoint precisely where adipocyte markers are located, 1777
 but it shows much more detail in cases when adipocyte transcripts escape the 1778
 boundaries of the cells. The examination of TransformerST and BayesSpace 1779
 demonstrates that the methods can be replicated with high precision. Notably, 1780
 strong correlations were observed between the quantities of transcripts and the 1781
 proportions of cell types in these replicates, further validating the effectiveness 1782
 and reproducibility of the approaches. 1783

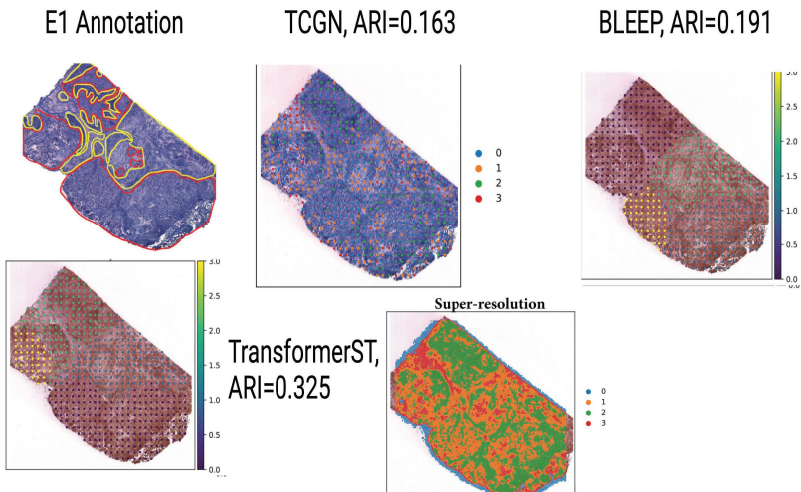
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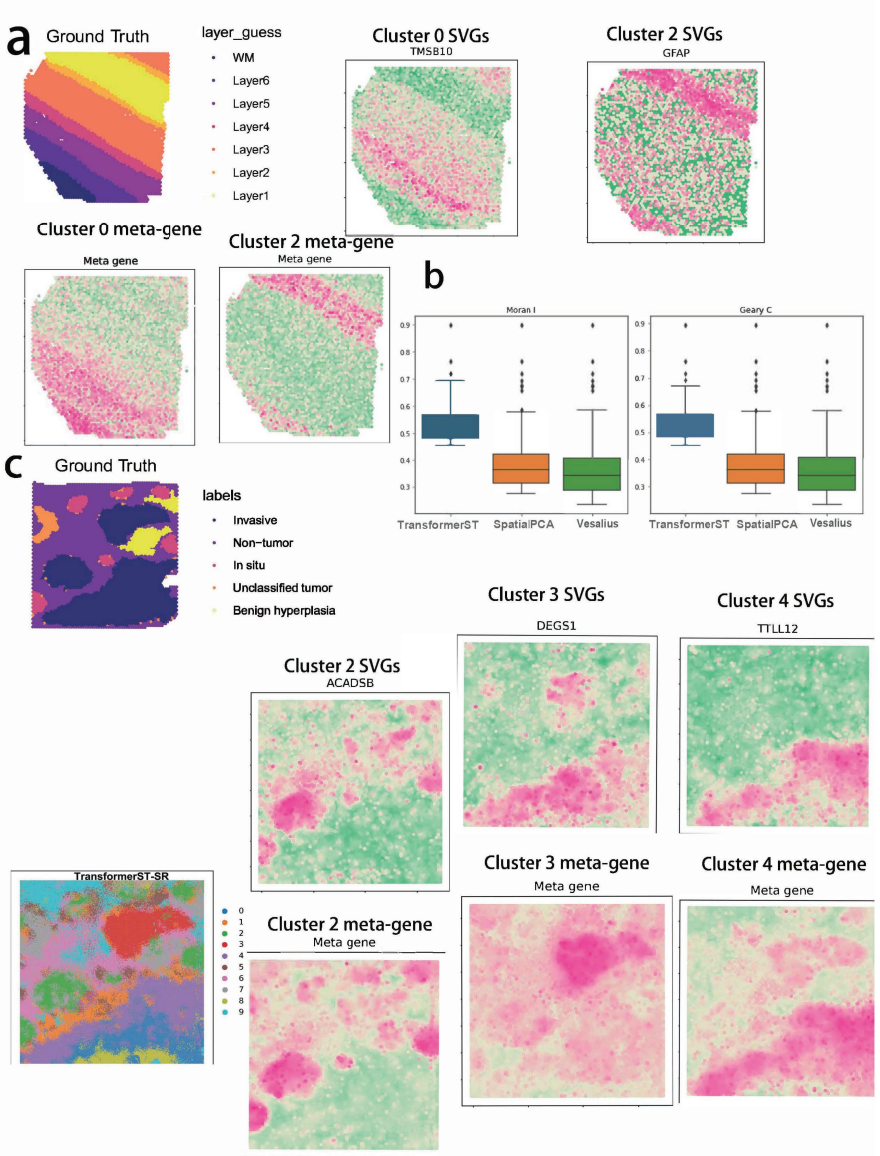
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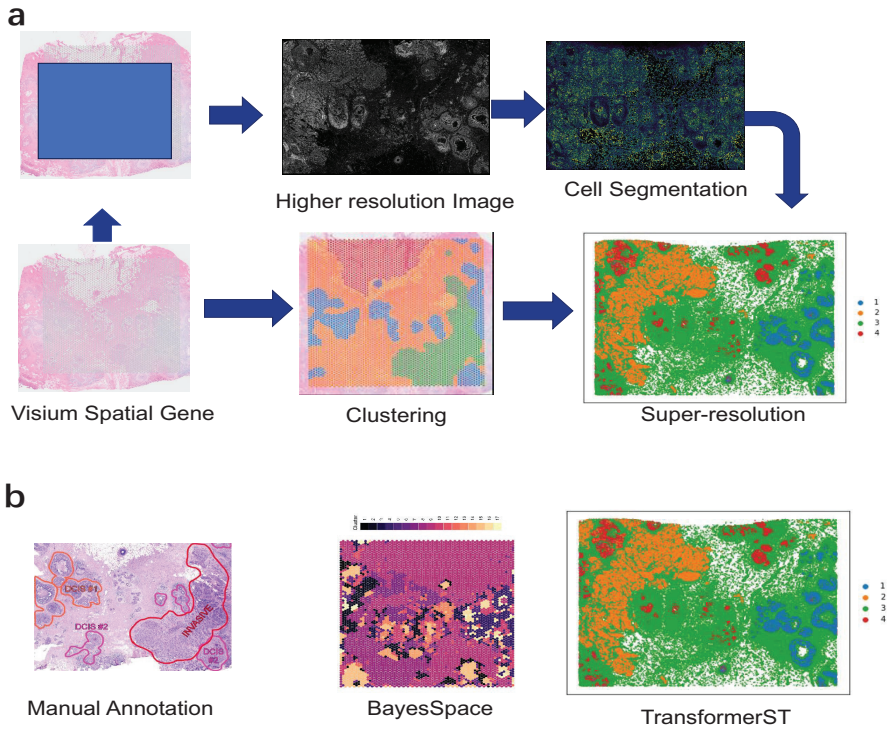
Supplementary Figure 3 Super-resolved gene expression prediction with breast cancer data. The first column displays the histology image accompanied by pathologists' annotations, in which the red lines represent invasive cancer, green lines signify breast glands, yellow lines indicate immune infiltration and blue lines denote connective tissue. a, Tissue type assignments and nearly single cell super-resolution using B1 section. b, Tissue type assignments and nearly single cell super-resolution using E1 section. *TransformerST* refers to the clustering results obtained from *TransformerST*. Super-resolution, on the other hand, represents the ability of the method to achieve sub-cellular resolution performance in enhancing the spatial transcriptomics data.



Supplementary Figure 4 Spatial variable genes (SVGs) and meta gene detection. a, SVGs and corresponding meta genes for cluster 0 (TMSB10, TMSB10+MBP-MT-CO2), cluster 2 (GFAP, GFAP+SNORC-TMSB10+CDT3-MBP) in brain tissue slice 151508 at spot resolution. b, Boxplots of Moran's I and Geary's C values for SVGs detected by TransformerST, SpatialPCA, and Vesalius using brain tissue slice 151508. c, SVGs and corresponding meta gene for cluster 2 (ACADSB, ACADSB+NME2-MUC1+ATP5MPL-CD74+LAPTM4B-CRIP1), cluster 3 (DEGS1, DEGS1+RPS18-CXCL14+AGR2-MGP+CSTA-NEAT1), and cluster 4 (TTLL12, TTLL12+HMG2-MALAT1+KRT8-SLC9A3R1) in IDC sample at nearly single cell resolution. Pathologist annotated different regions in different colors (IC outlined in red, carcinoma in yellow, benign hyperplasia in green, unclassified tumor in grey).

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Supplementary Figure 5 Tissue identification and super-resolution in breast cancer block at nearly single-cell resolution with Xenium technology. a, Schematic illustration of TransformerST with Xenium data. b, Tissue identification in breast cancer tissue with Xenium and Visium data.