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Deciphering spatial domains from spatially resolved transcriptomics with Siamese Graph Autoencoder --Manuscript Draft--

Manuscript Number:	GIGA-D-23-00329	
Full Title:	Deciphering spatial domains from spatially resolved transcriptomics with Siamese Graph Autoencoder	
Article Type:	Technical Note	
Funding Information:	National Natural Science Foundation of China (32300526)	Dr. Shuangfang Fang
	National Key R&D Program of China (2022YFC3400400)	Pro. Xun Xu
Abstract:	<p>Background</p> <p>Cell clustering is a pivotal aspect of spatial transcriptomics (ST) data analysis as it forms the foundation for subsequent data mining. Recent advances in spatial domain identification have leveraged Graph Neural Network approaches in conjunction with spatial transcriptomics data. However, such GNN-based methods suffer from representation collapse, wherein all spatial spots are projected onto a singular representation. Consequently, the discriminative capability of individual representation feature is limited, leading to suboptimal clustering performance.</p> <p>Results</p> <p>To address this issue, we proposed SGAE, a novel framework for spatial domain identification, incorporating the power of Siamese Graph Autoencoder. SGAE mitigates the information correlation at the both sample and feature level, thus improving the representation discrimination. We adapted this framework to ST analysis by constructing a graph based on both gene expression and spatial information. SGAE outperformed alternative methods by its effectiveness in capturing spatial patterns and generating high-quality clusters, as evaluated by ARI, FMI, and NMI. Moreover, the clustering results derived from SGAE can be further utilized in the identification of 3D Drosophila embryonic structure with enhanced accuracy.</p> <p>Conclusions</p> <p>Benchmarking results from various ST datasets generated by diverse platforms demonstrate compelling evidence for the effectiveness of SGAE against other ST clustering methods. Specifically, SGAE exhibits potential for extension and application on multi-slice 3D reconstruction and tissue structure investigation. The source code and a collection of spatial clustering results can be accessed at https://github.com/STOmics/SGAE/.</p>	
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Question	Response
Are you submitting this manuscript to a special series or article collection?	No
Experimental design and statistics Full details of the experimental design and statistical methods used should be given in the Methods section, as detailed in our Minimum Standards Reporting Checklist . Information essential to interpreting the data presented should be made available in the figure legends. Have you included all the information requested in your manuscript?	Yes
Resources A description of all resources used, including antibodies, cell lines, animals and software tools, with enough information to allow them to be uniquely identified, should be included in the Methods section. Authors are strongly encouraged to cite Research Resource Identifiers (RRIDs) for antibodies, model organisms and tools, where possible.	Yes

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1 **Deciphering spatial domains from spatially resolved transcriptomics**
2 **with Siamese Graph Autoencoder**

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19

20 **Abstract**

21 **Background**

22 Cell clustering is a pivotal aspect of spatial transcriptomics (ST) data analysis as it forms the
23 foundation for subsequent data mining. Recent advances in spatial domain identification have
24 leveraged Graph Neural Network approaches in conjunction with spatial transcriptomics data.
25 However, such GNN-based methods suffer from representation collapse, wherein all spatial spots
26 are projected onto a singular representation. Consequently, the discriminative capability of
27 individual representation feature is limited, leading to suboptimal clustering performance.

28 **Results**

29 To address this issue, we proposed SGAE, a novel framework for spatial domain identification,
30 incorporating the power of Siamese Graph Autoencoder. SGAE mitigates the information
31 correlation at both sample and feature level, thus improving the representation discrimination. We
32 adapted this framework to ST analysis by constructing a graph based on both gene expression and
33 spatial information. SGAE outperformed alternative methods by its effectiveness in capturing
34 spatial patterns and generating high-quality clusters, as evaluated by ARI, FMI, and NMI.
35 Moreover, the clustering results derived from SGAE can be further utilized in the identification of
36 3D Drosophila embryonic structure with enhanced accuracy.

37 **Conclusions**

38 Benchmarking results from various ST datasets generated by diverse platforms demonstrate
39 compelling evidence for the effectiveness of SGAE against other ST clustering methods.
40 Specifically, SGAE exhibits potential for extension and application on multi-slice 3D
41 reconstruction and tissue structure investigation. The source code and a collection of spatial
42 clustering results can be accessed at <https://github.com/STOmics/SGAE/>.

43

44 **Keywords:** Spatial transcriptomics; Spatial clustering; Graph neural networks

45

46 **Background**

47 Spatial transcriptomics (ST) represents a newly emerging technology that revolutionizes the
48 comprehensive characterization of tissue organization and architecture[1, 2]. By profiling the
49 spatially-resolved gene expression patterns, ST technologies allow scientists to delve into the
50 intricate cellular dynamics within tissues. Based on the underlying methodology, these techniques
51 can be categorized into two main categories: (1) imaging-based methods (MERFISH[3] and
52 seqFISH[4]), and (2) sequencing-based methods (Slide-seq[5] and 10x Visium[6]). As the need
53 for higher-resolution analysis to unravel cellular diversity becomes imperative, advancements
54 such as Stereo-seq[7] have been developed to provide improved resolution over the years. The
55 advent of ST technologies holds immense potential to drive biological discoveries in development,
56 physiology and a broad range of diseases[8, 9].

57

58 In parallel with single-cell RNA sequencing (scRNA-Seq) analysis, clustering serves as the initial
59 step in ST data analysis, grouping individual cells based on their gene expression patterns.

60 Similarly, the primary objective for ST data analysis revolves around dissecting tissue into distinct
61 spatial domains. While traditional machine learning approaches have been applied to tackle this
62 task, recent studies have sought to apply deep learning frameworks to learn how to classify spatial
63 spots into specific regions[10-13]. For instance, SpaGCN[12] identifies spatial domains through a
64 graph convolutional network (GCN) framework, while STAGATE[13] deploys a graph attention
65 autoencoder to define spatial clusters. However, such graph neural network (GNN) based methods
66 usually suffer from representations collapse, which tends to map spatial spots into the same
67 representation[14]. Consequently, the discriminative capability of spot representation is limited,
68 leading to inaccurate identification of spatial domains.

69

70 To tackle the aforementioned challenge, we proposed SGAE, which aims to learn discriminative
71 spot representation and accurately decipher spatial domains. This framework is derived from the
72 Dual Correlation Reduction Network (DCRN)[14], which effectively reduces information
73 correlation at the dual level. SGAE adapts this architecture to ST data analysis by constructing a
74 graph that incorporates both gene expression and spatial information. According to benchmarking
75 assessments, SGAE outperforms existing algorithms in the task of domain identification with

76 superior performance. Moreover, SGAE can be extended in the realm of 3D tissue structure
77 identification.

78

79 **Results**

80 **Overview of SGAE framework**

81 SGAE is an unsupervised algorithm for ST clustering that leverages a Variational Graph
82 Autoencoder (VGAE)[15] within a Siamese graph neural network to combine gene expression and
83 spatial information (Fig.1). To implement SGAE, the gene expression matrix (X) and adjacency
84 matrix (A) are fed into the encoder, which maps the gene expression data into a lower-dimensional
85 latent space, generating embedding vectors (Z) for individual cells. Pseudo-label is firstly
86 generated by pre-clustering based on gene expression patterns. SGAE adaptively learns the edge
87 weights of the spatial neighbor network (SNN) to capture the similarity between neighboring spots
88 and update the spot representation by aggregating information from neighbors. Finally, the latent
89 embeddings can be visualized using Uniform Manifold Approximation and Projection (UMAP)
90 and various clustering algorithms such as K-means and Louvain can be employed to identify
91 spatial domains for subsequent analysis.

92

93 By calculating K-nearest neighbors (KNN) based on the relative spatial positioning of spots,
94 SGAE can effectively capture the spatial relationships between cells. This is especially essential
95 for spatial transcriptomics (ST) data with low spatial resolutions, such as 10x Visium, where
96 discerning fine-grained spatial details can be challenging. Besides, SGAE introduces the concept
97 of a cell type-aware SNN by pruning the SNN based on the pre-clustering of gene expressions.
98 This preliminary clustering step aids in identifying regions that contain distinct cell types.
99 Through the incorporation of cell type information during the graph construction process, SGAE
100 adeptly captures data heterogeneity and improve the accuracy of the graph representation.

101

102 SGAE uses graph distortion to acquire diverse and informative node representations. This is
103 achieved through the application of two types of perturbation: feature perturbation and graph
104 perturbation. For feature perturbation, a random noise matrix is introduced to the feature matrix
105 using the Hadamard product. On the other hand, graph perturbation involves edge removal and

106 graph diffusion within the Siamese architecture. To implement edge removal, a mask matrix is
107 generated based on the cosine similarity matrix computed through pairwise comparisons in the
108 latent space. The 10% of edges with the lowest values are then removed. Graph diffusion is
109 facilitated using a random walk-based Personalized PageRank algorithm[16], allowing for the
110 passage of messages through higher-order neighborhoods. To optimize the learning process,
111 SGAE employs an objective function inspired by the Barlow Twins approach[17], aiming to
112 minimize the deviation of the cross-correlation matrix from the ideal identity matrix and reduce
113 redundant information among nodes in the latent space, therefore improving the overall accuracy
114 of the learned embedding.

115

116 **SGAE exhibited remarkable effectiveness and robustness in spatial domain exploration**

117 ST datasets generated by different technology platforms possess distinct resolutions and features,
118 making it essential to validate the clustering robustness of SGAE across these platforms. To
119 achieve this, we included ST datasets generated by 10x Visium[18], seqFISH[19], MERFISH[20],
120 and SLIDE-seq v2[21]. Then we comprehensively compare the clustering performance of SGAE
121 against other state-of-the-art spatial clustering methods, including SpaGCN[12], GraphST[10],
122 STAGATE[13] and Leiden[22]. Clustering performance was assessed by spatial visualization
123 combined with Adjusted Rand Index (ARI), Normalized Mutual Information (NMI) and Fowlkes-
124 Mallows Index (FMI).

125

126 **Human dorsolateral prefrontal cortex 10x Visium dataset**

127 We applied SGAE to analyze the 10x Visium ST dataset obtained from the human
128 dorsolateral prefrontal cortex (DLPFC)[18]. The visualization of cell clustering confirmed
129 that SGAE was able to discern the intricate stratified cortex structures with remarkable
130 clarity, surpassing the capabilities of other existing methods (Fig. 2A). Furthermore, our
131 benchmarking results revealed that SGAE outperformed other algorithms in analyzing all 12
132 DLPFC slices (Fig. 2E).

133

134 **Mouse gastrulation seqFISH dataset**

135 The evaluation of SGAE's performance extends to the mouse gastrulation dataset, which was

136 generated through the imaging-based technology seqFISH[19]. The visualization of mouse
137 gastrulation structures derived from different methods demonstrates higher effectiveness of SGAE
138 in accurately discriminating embryo tissue sections (Fig. 2B). In contrast, STAGATE failed to
139 decipher the spatial domain with precision, as it tends to divide the spatial domain into numerous
140 disorder patches. Notably, SGAE reaffirmed its superiority in all benchmark metrics against
141 other methods (Fig. 2F).

142

143 **Mouse cortex MERFISH dataset**

144 Based on the MERFISH dataset of the mouse primary motor cortex[20], we further compare
145 the clustering results obtained by different methods. While all five methods successfully
146 extract the stratified structure of the cortex, SGAE demonstrates a remarkable ability to
147 capture the layered organization of the glutamatergic structures more accurately when
148 compared to the original annotation (Fig. 2C). Furthermore, SGAE achieved the highest
149 performance among all five methods, underscoring its effectiveness in precisely clustering
150 cells and capturing the spatial arrangement of the primary motor cortex (Fig 2G).

151

152 **Mouse olfactory bulb SLIDE-seq v2 dataset**

153 The evaluation also encompasses the SLIDE-seq V2 dataset of the mouse olfactory bulb[21].
154 The spatial domains identified by SGAE exhibited remarkable consistency with the annotation
155 provided by the Allen Reference Atlas, strengthening the confidence in its accuracy and reliability
156 (Fig. 2D). Conversely, the Leiden clustering approach failed to provide a cohesive tissue structure
157 in this dataset, while SpaGCN, GraphST, and STAGATE partially deciphered certain structures
158 within the olfactory bulb.

159

160 Overall, our results unequivocally establish SGAE as a powerful method for analyzing ST data,
161 surpassing other state-of-the-art methods in terms of cell clustering performance and structure
162 exploration of complex tissues.

163

164 **SGAE deciphers spatial domains and provides discriminative representations**

165 Stereo-seq is a groundbreaking ST technology that offers subcellular resolution and has opened up

166 new avenues for investigating the intricate structures within complex tissues[7]. However, the
167 exploitation of its high-resolution capabilities necessitates the utilization of advanced clustering
168 and spatial analysis methods. Therefore, we conducted a meticulous evaluation of SGAE's
169 clustering performance using a Stereo-seq dataset of the mouse adult brain dataset [23].
170 Intriguingly, SGAE showcased exceptional discriminative power in accurately distinguishing
171 mouse brain sections within this dataset, outperforming other methods such as SpaGCN,
172 STAGATE, CCST, and GraphST (Fig. 3A). Subcluster analysis further demonstrated the superior
173 performance of SGAE (Fig. 3B). SGAE accurately delineated distinct subpopulations within the
174 tissue, whereas STAGATE inaccurately divided the DGGRC2 and TEGLU24 regions into two
175 separate clusters, and SpaGCN assigned a larger region for TEGLU24 and HBGLU.

176

177 To provide a systematic comparison, we conducted an extensive evaluation of SGAE's clustering
178 results using multiple benchmark metrics, including ARI, NMI, and FMI. Remarkably, SGAE
179 outperformed all other existing methods across all benchmark metrics (Fig. 3C). Besides, we
180 utilized Moran's Index (MI) to assess the spatial autocorrelation of each cluster. Although
181 SpaGCN and STAGATE achieved higher MI scores, SGAE exhibited a distribution most closely
182 aligned with the ground truth in terms of MI (Fig. 3D). It is suggested that SGAE effectively
183 utilizes spatial information in a more reasonable and appropriate manner.

184

185 Furthermore, we evaluated the representative embedding provided by SGAE, CCST[11],
186 STAGATE, and GraphST through UMAP visualization (Fig. 3E). The results showed that SGAE
187 exhibited a remarkable proficiency in extracting the embedding of the mouse brain Stereo-seq
188 data, while GraphST struggled to distinguish different cell groups. To further evaluate the
189 capability of SGAE to characterize biological representation, we performed pseudotime analysis
190 and calculated the F-score for each cell type (Fig. 3F). Surprisingly, SGAE achieved the highest F-
191 score, highlighting the discriminative capability of SGAE's embedding in accurately
192 distinguishing between different cell types.

193

194 Taken together, these findings provide compelling evidence that SGAE not only surpasses other
195 methods in terms of clustering accuracy, but also excels in providing superior embedding

196 representation for the datasets.

197

198 **SGAE enhanced complex spatial domain dissection in 3D Drosophila**

199 The advanced use of ST clustering involves integrating 3D reconstruction technology to gain a
200 comprehensive understanding of the spatial organization and gene expression patterns within
201 complex tissues. The fundamental topic of 3D tissue structure dissection is to identify shared and
202 specific spatial domains across multiple slices of ST datasets. Our investigation sought to
203 determine whether SGAE could effectively accomplish this challenging multi-slice clustering
204 task, especially for the datasets with less batch effect. Notably, we found that SGAE surpassed
205 Leiden and STAligner[24] in accurately dissecting the spatial domains of Drosophila embryos at
206 different stages (E14-16, E16-18 and L1)[25], as evidenced by its higher similarity to the ground
207 truth (Fig4. A, B). These findings highlighted the effectiveness of SGAE in achieving reliable
208 multi-slice clustering for ST analysis.

209

210 After obtaining the clustering results from SGAE, STAligner and Leiden, we proceeded with the
211 crucial step of stack slice registration to enable 3D tissue reconstruction. This involved aligning
212 consecutive tissue slices to generate a complete and accurate 3D representation of the tissue. We
213 observed that the 3D meshes generated from SGAE results exhibited exceptional accuracy in
214 dividing the tissue into correct structures, aligning perfectly with the corresponding marker genes
215 (Fig. 4C). It indicated that the spatial domains generated by SGAE are highly effective in
216 achieving promising 3D tissue reconstruction. In contrast, STAligner and Leiden faltered in
217 accurately dividing the tissue into correct structures in certain cases. This suggests the robustness
218 and reliability of the spatial domains generated by SGAE.

219

220 **Discussion**

221 Spatial transcriptomics is a cutting-edge technology that allows us to simultaneously capture gene
222 expression while retain spatial information of the tissue. The emergence of large-scale ST data has
223 increased the demand for effective algorithms capable of dissecting spatial domains. To achieve
224 this, we proposed SGAE, a framework composed of two identical encoders based on a Siamese
225 network, which enabled us to encode cell features. Additionally, SGAE employs a graph neural

226 network that facilitates the learning of informative representations of both gene expression and
227 spatial locations. To fully leverage the spatial information provided by ST, we constructed a graph
228 based on the spatial information of each cell and pre-clustered gene expression. We then used a
229 linear combination operation to merge the decorrelated latent embeddings, enhancing the
230 discriminative power of the resulting embedding and clustering accuracy, thus facilitating
231 comprehensive analysis to provide profound insights into complex biological systems.

232

233 Our study demonstrates the effectiveness and robustness of SGAE in capturing tissue structures
234 across different ST technology platforms. This superiority over other methods indicates the
235 immense potential of SGAE as a reliable tool for analyzing ST datasets. Another notable strength
236 of SGAE lies in its ability to accurately capture the heterogeneity present within ST datasets. The
237 complexity and diversity of cell types within tissues pose significant challenges in accurately
238 characterizing gene expression patterns. Notably, SGAE's embedding successfully captures the
239 heterogenic information, enabling a more comprehensive understanding of the spatial organization
240 of gene expression patterns within tissues. While SGAE has demonstrated its advantages in ST
241 clustering, further validation across a wider range of ST datasets and biological systems is
242 necessary to fully assess the generalizability of SGAE's performance.

243

244 In this study, we also applied SGAE to analyze the Drosophila 3D dataset and unravel the spatial
245 domains during the E14-16, E16-18, and Larva L1 stages. We further compared the performance
246 of SGAE with that of STAligner, a commonly used method developed for multi-slice ST analysis.
247 By evaluating benchmark metrics, we consistently observed that SGAE outperformed STAligner
248 in effectively grouping cells into biologically meaningful clusters. The superior clustering results
249 of SGAE carry significant implications for the analysis of 3D tissue structure reconstruction. In
250 conclusion, SGAE demonstrates its proficiency in spatial domain identification on spatial
251 transcriptomics with moderate batch effect. For datasets with a high batch effect, it is
252 recommended to integrate batch removal methods upstream of SGAE to effectively mitigate this
253 issue. By accurately categorizing cells into reasonable groups, SGAE could contribute to a more
254 precise characterization of the spatial organization of gene expression patterns. This is particularly
255 important for understanding the complex processes underlying biological development and

256 differentiation.

257

258 **Methods**

259 **Notations and Problem Definition**

260 An undirected graph is usually represented by $G = \{V, E\}$, where $V = \{v_1, v_2, \dots, v_N\}$ and

261 E are the node and edge respectively. Each node v_i is characterized by a vector $x_i \in \mathbb{R}^D$,

262 where D is the dimension of the attribute. Then the graph can be characterized by the feature

263 matrix $X \in \mathbb{R}^{N \times D}$. The relation between each node is characterized by the adjacency matrix

264 $A = (a_{ij})_{N \times N}$, where $a_{ij} = 1$ if v_i and v_j are connected by an edge, otherwise $a_{ij} = 0$.

265 A degree matrix describes the number of edges connected to each node and can be

266 expressed in a diagonal matrix $D = \text{diag}(d_1, d_2, \dots, d_N) \in \mathbb{R}^{N \times N}$, d_i is the degree of

267 node v_i and calculated by $d_i = \sum_{(v_i, v_j) \in E} a_{ij}$. We normalize the adjacency matrix as $\tilde{A} =$

268 $D^{-1}(A + I)$ where $I \in \mathbb{R}^{N \times N}$ is the identity matrix.

269 In this paper, we aim to train a Siamese graph encoder that embeds all nodes into the low-

270 dimension latent space in an unsupervised manner. The resultant latent embedding can then

271 be directly utilized to perform node clustering by clustering metrics such as K-means and

272 Leiden.

273

274 **The Overall Architecture of SGAE**

275 The overall architecture of SGAE consists of Graph Distortion, Siamese Encoders, Siamese

276 Decoders, and a reconstruction loss function.

277

278 **Graph Distortion**

279 We utilized two types of graph distortion including feature corruption and edge perturbation.

280 For feature corruption, which is the feature-level distortion, we apply Hadamard product to feature

281 matrix and a random noise matrix generated from a Gaussian distribution, i.e., $\tilde{X} = X \odot N$,

282 where \odot means the Hadamard product and $N \sim N(1, 0.1)$.

283 For edge perturbation, which is the structure-level distortion, we adopt two types of distortion, i.e.,

284 edge-removing and graph diffusion. For the edge removal, we generated a mask matrix M

285 according to the similarity matrix by calculating the pair-wise cosine similarity in the latent space,
 286 where the 10% of the lowest edges will be removed. The final adjacency matrix after edge
 287 removal is

$$288 \quad A^m = D^{-\frac{1}{2}}((A \odot M) + I)D^{-\frac{1}{2}}$$

289

290 In the graph diffusion treatment, we used Personalized PageRank to calculate the normalized
 291 adjacency matrix into a graph diffusion matrix by following MVGRL method[26]:

$$292 \quad A^d = \alpha \left(I - (1 - \alpha) \left(D^{-\frac{1}{2}}(A + I)D^{-\frac{1}{2}} \right) \right)^{-1}$$

293 where $\alpha = 0.2$ as the teleport probability in a random walk.

294

295 Siamese Encoders

296 In order to reduce the utilization of space while learning richer cell representations, we
 297 constructed two same encoders based on Siamese network structure to encode cell features.
 298 The inputs of Siamese Encoders are graph $G_1 = (X_1, A_m)$ and graph $G_2 = (X_2, A_d)$.
 299 And the output is the embedding matrix H . First, we use two parameter-shared encoders
 300 to encode graph G_1 and graph G_2 respectively, and generate embedding matrices H_1 and
 301 H_2 . The encoder in the l -th layer can be formulated as:

$$302 \quad H_1^{(l)} = \sigma \left(\widehat{A}_m H_1^{(l-1)} W_1^{(l)} \right) + \sigma \left(H_1^{(l-1)} W_2^{(l)} + b^{(l)} \right)$$

$$303 \quad H_2^{(l)} = \sigma \left(\widehat{A}_d H_2^{(l-1)} W_1^{(l)} \right) + \sigma \left(H_2^{(l-1)} W_2^{(l)} + b^{(l)} \right)$$

304 where, $\widehat{A}_m = D_m^{-\frac{1}{2}}(A_m + I)D_m^{-\frac{1}{2}}$, $\widehat{A}_d = D_d^{-\frac{1}{2}}(A_d + I)D_d^{-\frac{1}{2}}$, D_m and D_d are degree matrices
 305 of A_m and A_d , I is the identity matrix, $W_1^{(l)}$ and $W_2^{(l)}$ are weight matrices of encoders in
 306 the l -th layer, $b^{(l)}$ is the bias vector of encoder in the l -th layer, σ is the non-linear
 307 activate function, such as ReLU and Tanh. When layer $l = 1$, $H_1^{(l-1)} = X_1$.

308 Ultimately, the decorrelated latent embeddings derived from two different views, namely
 309 H_1 and H_2 , are merged using a linear combination operation. This amalgamation
 310 produces clustering-focused latent embeddings that can be effectively employed for
 311 clustering purposes, particularly through the utilization of the K-means algorithm.

312

313 **Siamese Decoders**

314 For SGAE, we construct a decoder based on graph convolutional neural networks, while
315 reconstructing feature embeddings and adjacency matrices. The input is the embedding matrix
316 H , and the output is the original feature matrix X and the adjacency matrix A . Firstly,
317 we use the graph convolutional neural network to decode the embedding H to generate a
318 feature matrix \hat{H} , and the calculation formula of the k layer decoder is as follows:

$$319 \quad H^{(k)} = \sigma \left(D^{-\frac{1}{2}}(A + I)D^{-\frac{1}{2}}H^{(k-1)}W^{(k)} \right)$$

320 where D is the degree matrix of the matrix A , and $W^{(k)}$ is the parameter matrix of the
321 k -th layer of the decoder. Then, taking inner product computation between the
322 embedding matrix H and its transpose to generate the adjacency matrix \hat{A} .

323

324 **Reconstruction Loss Function**

325 Finally, we calculate the feature matrix reconstruction loss L_{REC-F} , the calculation
326 formula is as follows:

$$327 \quad L_{REC-F} = \frac{1}{2N} \left\| AX - \hat{H} \right\|_F^2$$

328 Calculate the adjacency matrix reconstruction loss L_{REC-A} , the calculation formula is as
329 follows:

$$330 \quad L_{REC-A} = \frac{1}{2N} \left\| A - \hat{A} \right\|_F^2$$

331

332 The reconstruction loss L_{REC} is the sum of the feature matrix reconstruction loss and the
333 adjacency matrix reconstruction loss, and the calculation formula is as follows:

$$334 \quad L_{REC} = L_{REC-F} + L_{REC-A}$$

335

336 **Redundant Reduction Module**

337 In order to eliminate redundant information in node embedding and generate distinguishable
338 embeddings for each node, the present invention designs a de-redundancy module, which
339 eliminates redundant information from two levels: node level and feature level:

$$340 \quad S_N = \frac{H_1 H_2^T}{\|H_1\| \|H_2\|}$$

341
$$S_F = \frac{Z_1 Z_2^T}{\|Z_1\| \|Z_2\|}$$

342
$$L_{RR} = L_{RR-N} + L_{RR-F}$$

343

344 **Clustering Guidance Module**

345 In order to effectively learn the feature embedding related to the clustering task, the
 346 present invention designs a clustering guidance module. Firstly, we pre-train the model, and
 347 use K-means to cluster the generated node embeddings. Secondly, we construct a
 348 clustering guidance loss L_C according to the node embedding matrix and the clustering
 349 result of the previous step: a) Compute the soft assignment matrix Q for all nodes and pre-
 350 trained cluster centers using the Student's t distribution. b) Generate the target distribution
 351 matrix P according to the soft allocation matrix Q , the element p_{ij} of the i row j
 352 column is calculated by the following formula:

353
$$p_{ij} = \frac{q_{ij}^2 / \sum_i q_{ij}}{\sum_{j'} (q_{ij'}^2 / \sum_i q_{ij'})}$$

354 Then compute the clustering guidance loss L_C using the KL divergence from the soft
 355 assignment, the target distribution and the pretrained soft assignment.

356 During training, the model is optimized by minimizing the loss function:

357
$$L = L_{REC} + L_C + L_{RR}$$

358 After the model training is completed, the main flow of data in the model inference
 359 process is as follows: firstly, the model is used to obtain the low-dimensional feature
 360 embedding H of cells, and then based on the learned embedding, K-means is used for
 361 clustering, and finally the cluster labels of all cells are obtained.

362

363 **Clustering Refinement**

364 SGAE also incorporates an optional clustering refinement step. During this step, SGAE analyzes
 365 the domain assignment of each spot and its neighboring spots. Specifically, for a given spot, the
 366 label that appears most frequently among its surrounding spots is assigned to that spot. The
 367 clustering refinement step was exclusively performed for the human DLPFC 10x Visium data.

368

369 **Performance Evaluation**

370 We use five indices to evaluate the quality of the clustering results: Adjusted Rand Index (ARI),
371 Normalized Mutual Information (NMI), Fowlkes-Mallows Index (FMI), FM Index, and Moran
372 Index. These indices provide different perspectives on the clustering performance. ARI measures
373 the similarity of predicted types in the clusters, with a range from -1 to 1. NMI measures the
374 relationship between variables and is normalized to a range of [0,1]. FMI calculates the geometric
375 mean of pairwise precision and recall, also ranging from 0 to 1. FM Index evaluates the similarity
376 between predicted and true binary rankings. The Moran Index is used to assess spatial
377 autocorrelation in the clustering results. Together, these indices offer a comprehensive evaluation
378 of the clustering quality across various aspects.

379

380 **Data Preprocessing**

381 SGAE utilizes transcriptome-wide gene expression profiles with spatial coordinates as
382 input. The raw gene counts per spot are first normalized to the total counts per cell and
383 then scaled through log-transformation. In the case of 3D Drosophila datasets, we did not
384 employ any multi-slice integration method as there was little batch effect observed from
385 the UMAP result. Principal component analysis (PCA) is then conducted on the gene
386 expression data using the *sc.pp.pca()* function, and the top 50 principal components per
387 spot are subsequently utilized as the default expression feature.

388

389 **Identifying differentially expressed genes.** Wilcoxon test implemented in SCANPY [27] is
390 used to calculate differentially expressed genes for each spatial domain Benjamin-Hochberg
391 adjustment correlation via *sc.tl.rank_genes_groups()*.

392

393 **Spatial trajectory inference**

394 We employed the PAGA algorithm [28] implemented in the SCANPY package to depict spatial
395 trajectory. The PAGA trajectory and PAGA tree were inferred by the *scanpy.tl.paga()* function
396 based on cell embedding generated by SGAE. Furthermore, *scanpy.tl.dpt()* was applied to
397 estimate the pseudo time as well. To compare the performance of each clustering method with
398 embedding, we calculate trajectory and pseudo time using methods above with same parameters

399 settings.

400

401 **Availability of supporting source code and requirements**

402 Project name: SGAE

403 Project home page: <https://github.com/STOmics/SGAE/>

404 Operating system: Linux

405 Programming language: Python

406 License: MIT license

407

408 **Data Availability**

409 Supporting data sets for this article are available via spatialLIBD

410 (<https://research.libd.org/spatialLIBD>), Brain Image Library (<https://doi.brainimagelibrary.org/>),

411 SpatialMouseAtlas (<https://crukci.shinyapps.io/SpatialMouseAtlas/>), Single Cell PORTAL

412 (https://singlecell.broadinstitute.org/single_cell/study/SCP815/highly-sensitive-spatial-

413 [transcriptomics-at-near-cellular-resolution-with-slide-seqv2#study-summary](https://singlecell.broadinstitute.org/single_cell/study/SCP815/highly-sensitive-spatial-transcriptomics-at-near-cellular-resolution-with-slide-seqv2#study-summary)), CNGBdb

414 (<https://db.cngb.org/stomics/flysta3d/spatial>), and Zenodo (<https://zenodo.org/record/7340795>).

415

416 **Abbreviations**

417 ST: spatial transcriptomics, MERFISH: Multiplexed Error-Robust Fluorescence in situ

418 Hybridization, seqFISH: sequential fluorescence in situ hybridization, scRNA-Seq: single-cell

419 RNA sequencing, GCN: graph convolution network, GNN: graph neural network, DCRN: Dual

420 Correlation Reduction Network, VGAE: Variational Graph Autoencoder, SNN: spatial neighbor

421 network, UMAP: Uniform Manifold Approximation and Projection, KNN: K-nearest neighbors,

422 ARI: Adjusted Rand Index, NMI: Normalized Mutual Information, FMI: Fowlkes-Mallows Index,

423 DLPFC: dorsolateral prefrontal cortex, MI: Moran's Index.

424

425 **Funding**

426 This work is supported by National Natural Science Foundation for Young Scholars of China

427 (32300526) and National Key R&D Program of China (2022YFC3400400)

428

429 **Authors' Contributions**

430 S.F. and Y.Z. conceived and designed the study. W.J., L.C., C.Y. and Y.R. proposed the SGAE
431 model. L.C., L.H., C.Y., and Y.J. performed the data analysis. T.X. helped with the 3D
432 reconstruction analysis. M.L. X.X, and Y.L. participated in the study discussions. L.C., L.H.,
433 C.Y., and S.F. wrote the manuscript.

434

435 **Competing Interests**

436 All authors declare no conflicts of interest in regard to this manuscript.

437

438 **ACKNOWLEDGEMENTS**

439 We thank Lidong Guo, Xiaobin Liu for their help to the manuscript. This work is part of the
440 “SpatioTemporal Omics Consortium” (STOC) paper package. A list of STOC members is
441 available at: <http://sto-consortium.org>. We acknowledge the Stomics Cloud platform to provide
442 convenient ways for analyzing spatial omics datasets. We acknowledge the CNGB Nucleotide
443 Sequence Archive (CNSA) of China National GeneBank DataBase (CNGBdb) for maintaining the
444 MOSTA and Flysta3D database.

445

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523

524

525 **Figure legends**

526 **Figure 1. An overview of SGAE framework.** SGAE algorithm consists of three key
527 modules. Firstly, the graph distortion module generates two distorted graphs by introducing
528 both attribute and graph disturbances. Secondly, the encoder module generates two sets of
529 representations for each sample. Thirdly, the redundant reduction module ensures that the
530 same sample within the two distorted graphs has identical representations at both the feature
531 and sample levels. Lastly, the discriminative representations are applied to clustering
532 algorithms such as k-means to decipher spatial domains.

533 **Figure 2. SGAE exhibited high effectiveness and robustness in spatial domain**
534 **exploration.** (A-D) Visualization of clustering results from SGAE, SpaGCN, GraphST,
535 STAGATE, Leiden and annotation. (A) Human dorsolateral prefrontal cortex (DLPFC) 10x
536 Visium dataset, (B) Mouse gastrulation seqFISH dataset, (C) Mouse cortex MERFISH
537 dataset, (D) Mouse olfactory bulb SLIDE-seq v2 dataset. (E-G) Benchmark metrics
538 comparison of SGAE against SpaGCN, GraphST, STAGATE and Leiden. (E) 12 DLPFC 10x
539 Visium datasets, (F) Mouse gastrulation seqFISH dataset, (G) Mouse cortex MERFISH
540 dataset.

541 **Figure 3. SGAE unraveled spatial domains and provided discriminative representations.** (A)
542 Visualization of human adult brain clustering results from SGAE, SpaGCN, STAGATE, CCST,
543 and GraphST. (B) Subclustering results of DGGRC2, TEGLU24 and HBGLU from SGAE, SpaGCN,
544 STAGATE, CCST, and GraphST. (C) Benchmark metrics comparison of SGAE against SpaGCN,
545 STAGATE, CCST, and GraphST. (D) Moran's Index comparison of SGAE against SpaGCN,
546 STAGATE, CCST, and GraphST. (E) UMAP visualization of embedding from SGAE, SpaGCN,
547 STAGATE and GraphST. (F) F score of pseudo-time calculated from embedding provided by PCA,
548 CCST, STAGATE and GraphST.

549 **Figure 4. SGAE enhanced complex spatial domain dissection in 3D Drosophila Embryo.** (A) 2D
550 visualization of Drosophila Embryo clustering results at different stages (E14-16, E16-18, and
551 L1) from SGAE and STAligner. (B) Benchmark metrics comparison of SGAE, Leiden and
552 STAligner. (C) 3D visualization of Drosophila Embryo. The first row showed the marker
553 genes of Drosophila Embryo at different stages, while the last three rows displayed the meshes
554 generated by SGAE, STAligner and Leiden respectively.

Figure 1

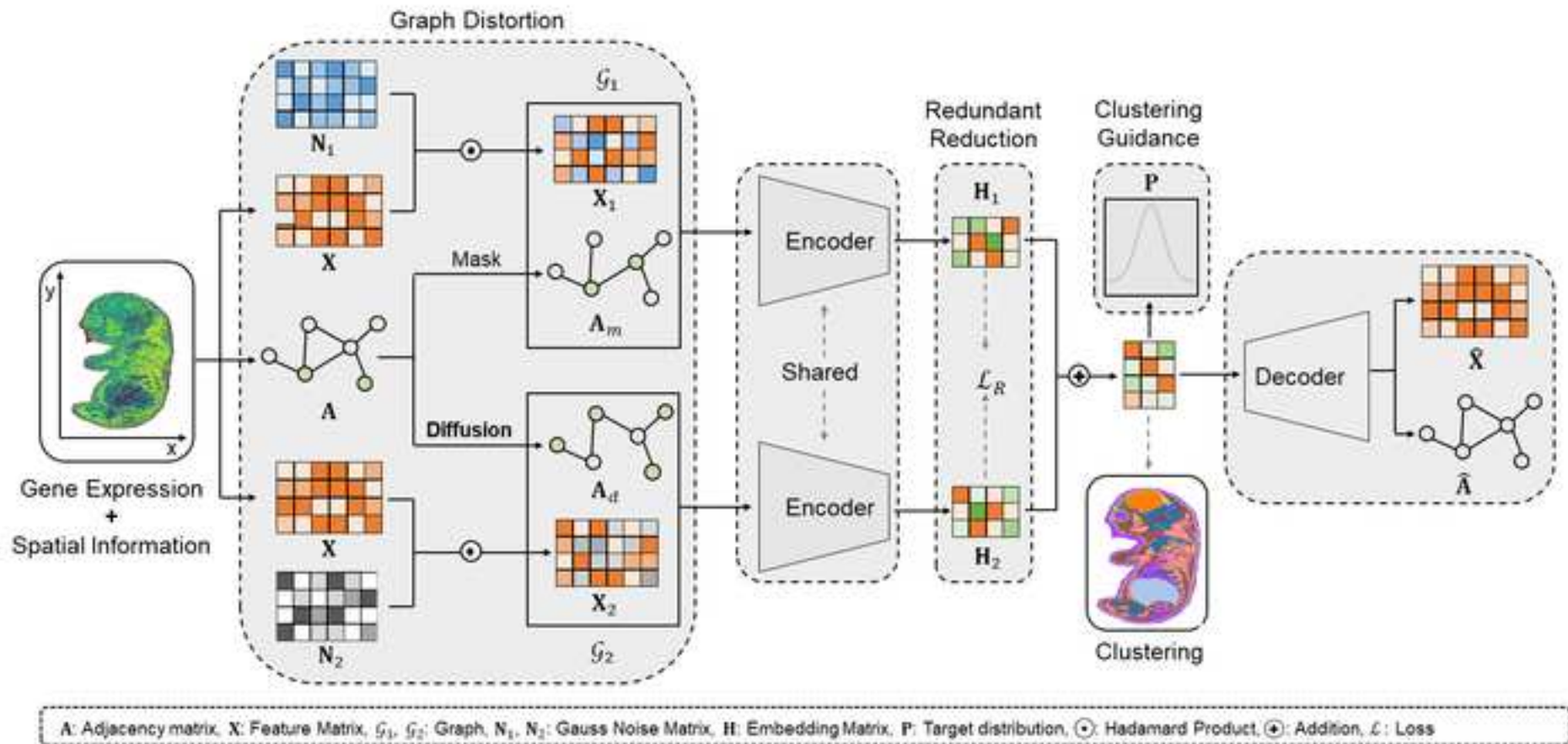


Figure 2

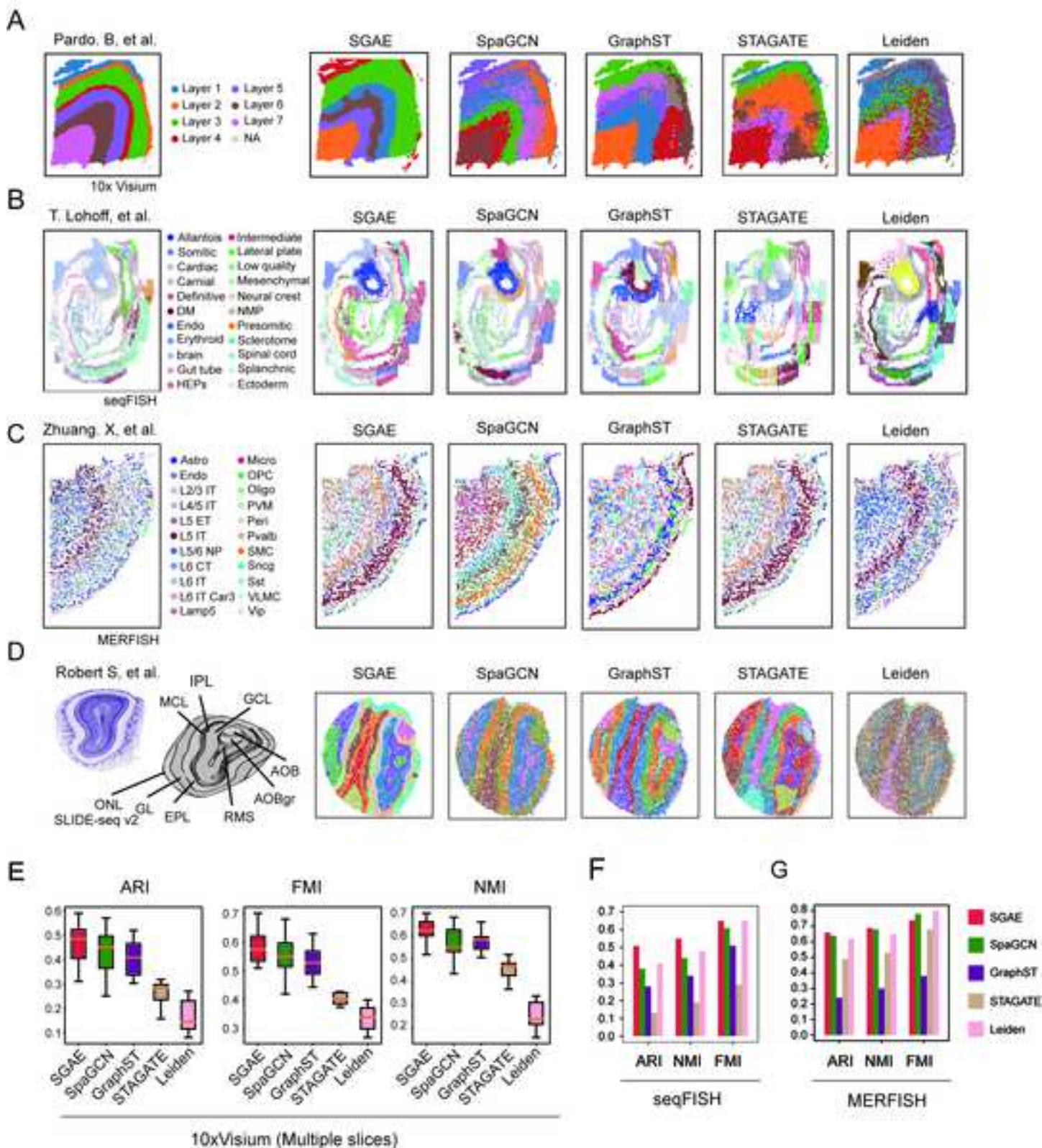


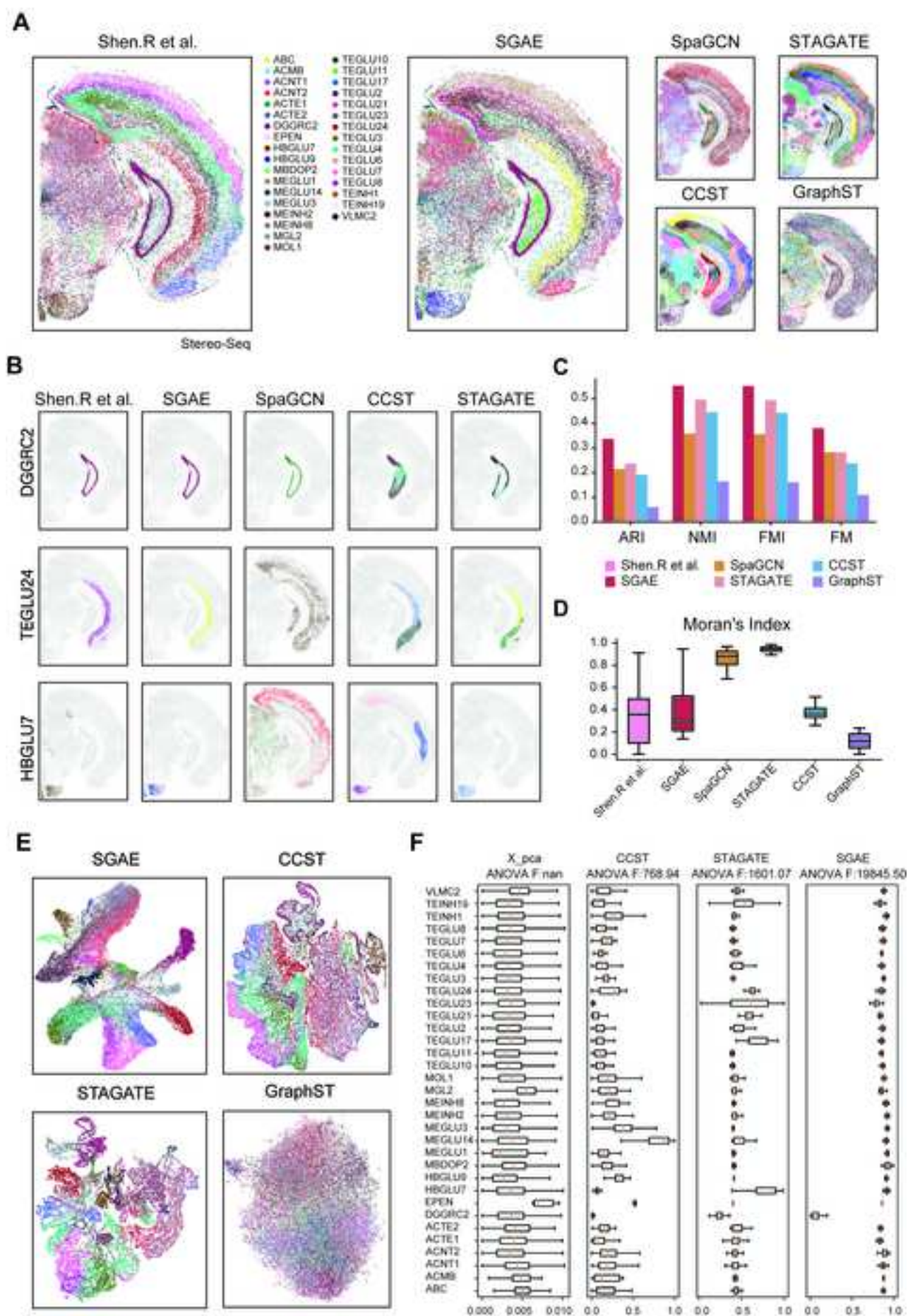
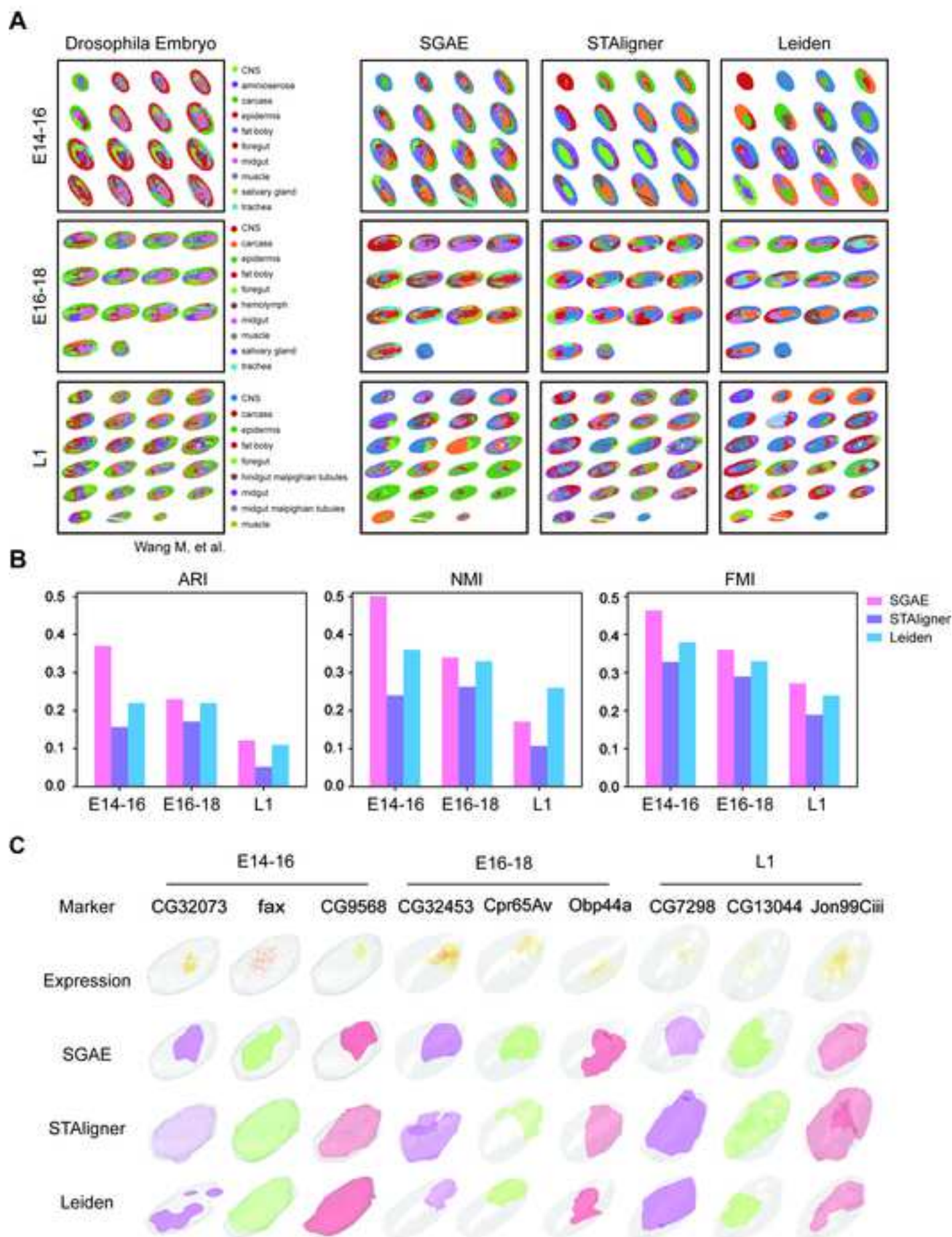
Figure 3

Figure 4

October 31, 2023

Dear editors,

We hereby submit our original manuscript entitled “Deciphering spatial domains from spatially resolved transcriptomics with Siamese Graph Autoencoder” for consideration by GigaScience. We confirm that this work is original and has not been previously published, nor is currently being reviewed for publication elsewhere.

In this paper, we describe a Siamese Graph Autoencoder (SGAE) spatial domain identification method for spatial transcriptomics. SGAE mitigates the information correlation at both sample and feature level, thus improving the representation discrimination. With an enhanced cellular representation, SGAE enables improved spatial domain identification and facilitates downstream analyses, such as trajectory inference. Entire computational framework of SGAE has been validated in multiple tissue samples from various technology platforms. All of which achieved state-of-the-art performance on the evaluation of clustering index including ARI, FMI and NMI. We believe this work effectively addresses the spatial domain identification for spatial transcriptomics with improved representation discrimination.

This work belongs to the thematic series of Spatio-temporal omics algorithms.

We have no conflicts of interest to disclose.

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Thank you for your consideration of our manuscript.

Sincerely,
Shuangfang Fang

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