SUPPLEMENTARY INFORMATION

A.								
	Chr1	Chr2	Chr3	Chr4	Chr5	Chr6	Chr7	ChrR
Evolved 3.2	2 2 1	na na na mana na	on in a superior of the constituents of a spine	www.arminianianianianianianianianianianianiania			apterporter	ndunanan andar da andar da ana A
AMS5615	4 2 1 1		a way and a set of the		(lenne ser les	analan ^y aka per	
AMS5617	4321 A	× + + + + + + + + + + + + + + + + + + +	V management of the state of the second	androne straphic frances	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~			(a.t.,a
AMS5618	4 3 2 1	(n pyring an article at a	and a shear of particle spectrum.	(l umangang kana dipu V		
AMS5622	4 2 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	- 	-will be a second s	andre al and the second state of the second st	y Angeler and an and a state of the state of the A		and the state of the	with the state of
AMS5623		V 			(и 		
AMS5624			(magazar alar angazar ang		V 	- approximation of the second		Land and the second
AMS5625	V	×						- he was a first of the second s
AMS5626	å()		(managera and a subsedurated	(× • • • • • • • • • • • • • • • • • • •			(
AMS4130	4 2 1 2 1 2		V Angle Angle Angl	(manune lugar and larger	nation of the state of the stat			(
В.	Ch-4	Ob - D	01-2	Chad	Ch.r.F	Ob -C	017	0 k - D
ERG251/ERG251 ^{L113}	Chr1	Chr2	Chr3	Chr4	Chr5	Chr6	Chr7	ChrR
ERG251 ^{W265G} /ERG25	2 VI		V V			N N		
					(×)	N N		
ERG251 ^{E273*} /ERG251	ч))	A 1	have a second					
ERG251/ERG251*321Y	- 3 2	and a second s	••••••••••••••••••••••••••••••••••••••		Managana Man Managana Managana Mana		ملغانيت وليريب م مصد م	
erg251∆/ERG251		(w yearsy after the second and a					
ERG251/erg251∆	4 3 1 1		V name of the state of the stat			hangson an shift	-lalaria di secondaria di s	
erg251∆/ERG251 +ERG251-A	4 2 1 1	(~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	V mayering an angle ang A		~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	wayaa ahaa haa haa haa haa haa haa haa ha	alad waigh a grad	hard a star a star and a star a st
ERG251/erg251∆ +ERG251-B	4 2 2 4 - 2 - 4 - 2 - 4 - 4 - 4 - 4 - 4 - 4 -	, V 	Lunghan again mile an ann an daong da				-	(m. down to be seen with a new states and
+ERG251-B		ι <u>΄</u> Λ.)				·/		
+ERG251-B erg251∆/∆ -d51	1 4 3 2 1 1 1 1 1 1 1 1 1 1 1 1 1	hand the system of the second state of the sec	(marked a second		Land and the second sec	langer er senseligt		

Fig S1. Whole genome sequencing analysis of FLC-evolved and engineered strains.

A. *De novo* point mutations in *ERG251* often occur together with other aneuploidies. Representative whole genome sequencing (WGS) data of the FLC-evolved strains from Table 1: Evolved 3.2, AMS5615, AMS5617, AMS5618, AMS5622, AMS5623, AMS5624, AMS5625, AMS5626 and AMS4130 which acquired point mutations on *ERG251* during FLC evolution. **B.** The engineered *ERG251* mutants remain euploid. WGS data for all *ERG251* mutations engineered into the euploid SC5314 genetic background: the *ERG251* heterozygous point mutants (L113*, W265G, E273*, and *321Y), both heterozygous deletion strains of *ERG251*, two strains with complementation of the heterozygous deletion, and two independent homozygous deletions of *ERG251* (d51 and d70). **A&B** WGS data are plotted as the log2 ratio and converted to chromosome copy number (y-axis, 1-4 copies) as a function of chromosome

position (x-axis, Chr1-ChrR). Haplotypes are indicated by color: gray is heterozygous (AB), magenta is homozygous B, and cyan is homozygous A. The baseline ploidy was determined by propidium iodide staining (S1 Table).

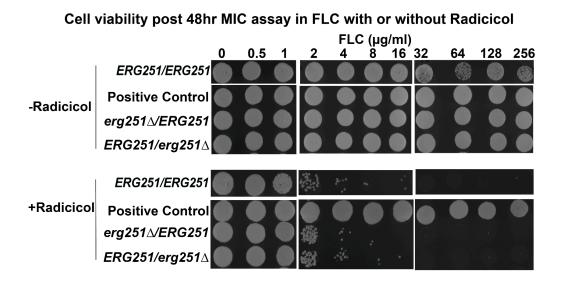


Fig S2. Radicicol, an Hsp90 inhibitor, blocks Erg251-driven tolerance and makes fluconazole fungicidal. Cells from the MIC assay at 48 hr in Fig 1D, with or without radicicol, were plated for viability on YPAD agar plates and imaged after 24 hr incubation. Wildtype SC5314 (*ERG251/ERG251*), a positive control strain known to be resistant to fluconazole (FLC), and both heterozygous deletion mutants of *ERG251* were tested. At least three biological replicates were performed.

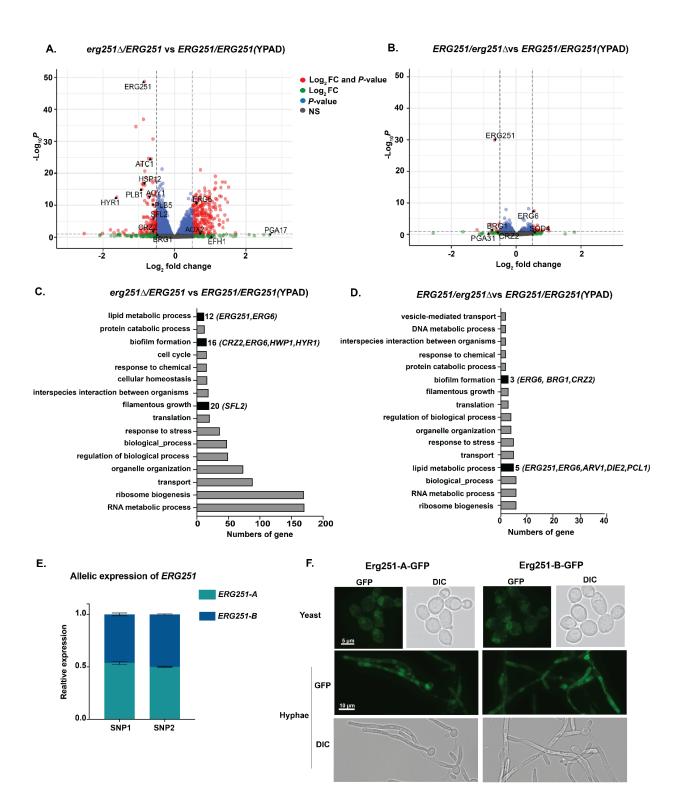


Fig S3. Heterozygous deletion of *ERG251-A* leads to a transcriptional response in filamentation regulation. Volcano plots for differentially expressed genes (log2 fold change ≥ 0.5 or ≤ -0.5 and adjusted *p*-value < 0.1) in the heterozygous mutants (A) *erg251* Δ /*ERG251* and (B) *ERG251*/*erg251* Δ in YPAD compared to the wildtype *ERG251*/*ERG251* in YPAD. Both the fold change and *p*-value are indicated. **C&D.** Gene

Ontology (GO) terms for genes differentially expressed in (C, S7 Table) *erg251* Δ /*ERG251* in YPAD and (D, S8 Table) *ERG251*/*erg251* Δ in YPAD compared to *ERG251*/*ERG251* in YPAD. **E.** Relative expression of *ERG251-A* and *ERG251-B* in the SC5314 background in YPAD. Relative expression was estimated using allelic RNA reads compared to overall reads at the two loci with polymorphisms in the *ERG251* gene (indicated as SNP1 and SNP2 above). Values are mean ± SEM calculated from three biological replicates. **F.** Subcellular localization of Erg251-A-GFP and Erg251-B-GFP in yeast and hyphal inducing conditions in SC5314 background. Yeast: scale bar, 5 µm; hyphae: scale bar, 10 µm.

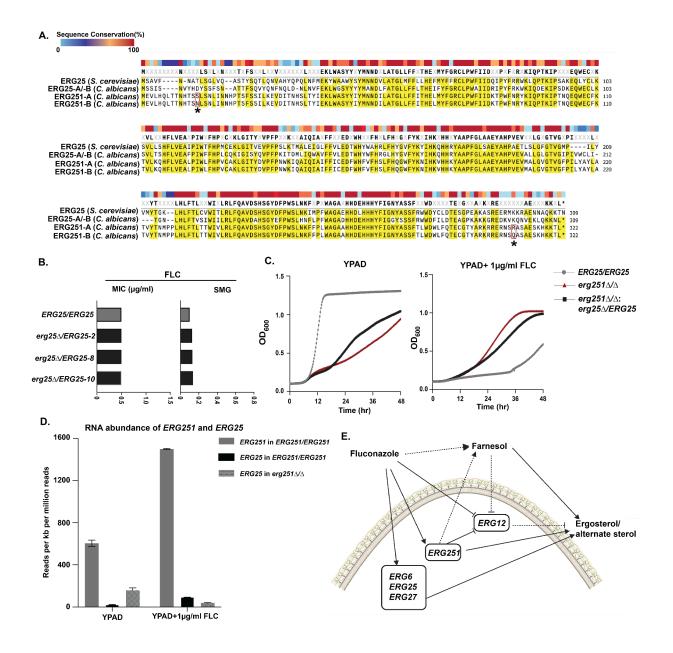


Fig S4. Erg251 is the major methyl sterol oxidase controlling drug susceptibility

compared to its paralog Erg25. (**A**) Multiple sequence alignment for *ERG251-A*, *ERG251-B*, and *ERG25-A*/-B (no SNPs between A and B) from *C. albicans* and *ERG25* from *S. cerevisiae*, with yellow highlighting similarity among all four proteins. Colored blocks on the top indicate the sequence conservation. Asterisks (*) and red boxes indicate the locus of non-synonymous variation between *ERG251-A* and *ERG251-B* in *C. albicans*. **B.** FLC susceptibility determined by liquid microbroth dilution at 24hr MIC (left, µg/mI) and 48hr SMG (right, tolerance) in FLC for three *ERG25* heterozygous deletion mutants (*ERG25/erg25A-2, -8* and *-10*) in the SC5314 background with SC5314 (*ERG25/ERG25*) as the control. **C.** 48hr growth curve analysis of *erg25* heterozygous deletion strain in *erg251* Δ/Δ background (*erg251* Δ/Δ : *ERG25/erg25* Δ) in

YPAD (left) and YPAD+1µg/ml FLC (right) with SC5314 (*ERG25/ERG25*) and *erg251* Δ/Δ as the controls. The initial cell densities were OD₆₀₀ of 0.001. MIC and SMG are not measurable for *erg251* Δ/Δ or *erg251* Δ/Δ : *ERG25/erg25* Δ given growth defects in YPAD. **B&C**: Minimum of three biological replicates were performed. **D.** RNA abundance of *ERG251* and *ERG25* in SC5314 (*ERG251/ERG251*), and *ERG25* in *erg251* Δ/Δ . RNA reads were normalised to transcript length and total RNA reads. Values are mean ± SEM calculated from three biology replicates. **E.** Predicted model for how FLC and farnesol impact the expression of *ERG* genes. In the wildtype, low concentrations of FLC promote the expression of most *ERG* genes, including *ERG6*, *ERG251*, *ERG25*, *ERG11* and *ERG27*, leading to the upregulation of ergosterol or/and alternate sterol biosynthesis. However, both low concentrations of FLC and Erg251 pose a negative regulation on Erg12, which may be achieved via farnesol which we predict inhibits *ERG12* [107]. Dashed lines indicate predicted relationships. Figure created in BioRender.com.

SUPPLEMENTARY TABLES

S1 Table. Strains used in this study.

S2 Table. Differentially expressed genes in *erg251* Δ/Δ in YPAD compared to wildtype in YPAD.

S3 Table. GO term analysis for differentially expressed genes in $erg251\Delta/\Delta$ in YPAD compared to wildtype in YPAD.

S4 Table. Differentially expressed GPI genes in $erg251\Delta/\Delta$ in YPAD compared to wildtype in YPAD.

S5 Table. Differentially expressed genes in *erg251∆/ERG251* in YPAD compared to wildtype in YPAD.

S6 Table. Differentially expressed genes in *ERG251/erg251*∆ in YPAD compared to wildtype in YPAD.

S7 Table. GO term for differentially expressed genes in *erg251∆/ERG251* in YPAD compared to wildtype in YPAD.

S8 Table. GO term for differentially expressed genes in *ERG251/erg251*∆ in YPAD compared to wildtype in YPAD.

S9 Table. Differentially expressed genes in *erg251∆/ERG251* in FLC compared to wildtype in FLC.

S10 Table. Differentially expressed genes in *ERG251/erg251*∆ in FLC compared to wildtype in FLC.

S11 Table. Differentially expressed genes in *erg251* Δ/Δ in FLC compared to wildtype in FLC.

S12 Table. Primers used in this study.

S13 Table. ERG251 SNPs from all FLC-evovled strains.

REFERENCES

- 1. Pfaller MA, Diekema DJ, Turnidge JD, Castanheira M, Jones RN. Twenty Years of the SENTRY Antifungal Surveillance Program: Results for Candida Species From 1997–2016. Open Forum Infect Dis. 2019;6: S79–S94.
- 2. Pfaller MA. Antifungal drug resistance: mechanisms, epidemiology, and consequences for treatment. Am J Med. 2012;125: S3–13.
- Perea S, López-Ribot JL, Kirkpatrick WR, McAtee RK, Santillán RA, Martínez M, et al. Prevalence of molecular mechanisms of resistance to azole antifungal agents in Candida albicans strains displaying high-level fluconazole resistance isolated from human immunodeficiency virus-infected patients. Antimicrob Agents Chemother. 2001;45: 2676–2684.
- 4. Pappas PG, Kauffman CA, Andes DR, Clancy CJ, Marr KA, Ostrosky-Zeichner L, et al. Clinical Practice Guideline for the Management of Candidiasis: 2016 Update by the Infectious Diseases Society of America. Clin Infect Dis. 2015;62: e1–e50.
- 5. Andes DR, Safdar N, Baddley JW, Playford G, Reboli AC, Rex JH, et al. Impact of treatment strategy on outcomes in patients with candidemia and other forms of invasive candidiasis: a patient-level quantitative review of randomized trials. Clin Infect Dis. 2012;54: 1110–1122.
- 6. Cowen LE. The evolution of fungal drug resistance: modulating the trajectory from genotype to phenotype. Nat Rev Microbiol. 2008;6: 187–198.
- 7. Berman J, Krysan DJ. Drug resistance and tolerance in fungi. Nat Rev Microbiol. 2020;18: 319–331.
- 8. Sanglard D. Emerging Threats in Antifungal-Resistant Fungal Pathogens. Front Med. 2016;3: 11.
- 9. Rosenberg A, Ene IV, Bibi M, Zakin S, Segal ES, Ziv N, et al. Antifungal tolerance is a subpopulation effect distinct from resistance and is associated with persistent candidemia. Nat Commun. 2018;9: 2470.
- 10. Todd RT, Soisangwan N, Peters S, Kemp B, Crooks T, Gerstein A, et al. Antifungal Drug Concentration Impacts the Spectrum of Adaptive Mutations in Candida albicans. Mol Biol Evol. 2023;40: msad009.
- 11. White TC, Marr KA, Bowden RA. Clinical, cellular, and molecular factors that contribute to antifungal drug resistance. Clin Microbiol Rev. 1998;11: 382–402.
- 12. Shapiro RS, Robbins N, Cowen LE. Regulatory circuitry governing fungal development, drug resistance, and disease. Microbiol Mol Biol Rev. 2011;75: 213–267.
- 13. Cowen LE, Steinbach WJ. Stress, drugs, and evolution: the role of cellular signaling in fungal drug resistance. Eukaryot Cell. 2008;7: 747–764.
- 14. Kelly SL, Lamb DC, Kelly DE, Manning NJ, Loeffler J, Hebart H, et al. Resistance to

fluconazole and cross-resistance to amphotericin B in Candida albicans from AIDS patients caused by defective sterol delta5,6-desaturation. FEBS Lett. 1997;400: 80–82.

- 15. Lupetti A, Danesi R, Campa M, Del Tacca M, Kelly S. Molecular basis of resistance to azole antifungals. Trends Mol Med. 2002;8: 76–81.
- Sanglard D, Kuchler K, Ischer F, Pagani JL, Monod M, Bille J. Mechanisms of resistance to azole antifungal agents in Candida albicans isolates from AIDS patients involve specific multidrug transporters. Antimicrob Agents Chemother. 1995;39: 2378–2386.
- 17. Revie NM, Iyer KR, Robbins N, Cowen LE. Antifungal drug resistance: evolution, mechanisms and impact. Curr Opin Microbiol. 2018;45: 70–76.
- 18. Shrivastava Manjari, Kouyoumdjian Gaëlle S., Kirbizakis Eftyhios, Ruiz Daniel, Henry Manon, Vincent Antony T., et al. The Adr1 transcription factor directs regulation of the ergosterol pathway and azole resistance in Candida albicans. MBio. 2023;14: e01807–23.
- 19. Blosser SJ, Merriman B, Grahl N, Chung D, Cramer RA. Two C4-sterol methyl oxidases (Erg25) catalyse ergosterol intermediate demethylation and impact environmental stress adaptation in Aspergillus fumigatus. Microbiology. 2014;160: 2492–2506.
- Lu H, Li W, Whiteway M, Wang H, Zhu S, Ji Z, et al. A Small Molecule Inhibitor of Erg251 Makes Fluconazole Fungicidal by Inhibiting the Synthesis of the 14α-Methylsterols. MBio. 2023;14: e0263922.
- Bhattacharya Somanon, Esquivel Brooke D., White Theodore C. Overexpression or Deletion of Ergosterol Biosynthesis Genes Alters Doubling Time, Response to Stress Agents, and Drug Susceptibility in Saccharomyces cerevisiae. MBio. 2018;9: 10.1128/mbio.01291–18.
- 22. Martel CM, Parker JE, Bader O, Weig M, Gross U, Warrilow AGS, et al. Identification and characterization of four azole-resistant erg3 mutants of Candida albicans. Antimicrob Agents Chemother. 2010;54: 4527–4533.
- 23. Jordá T, Puig S. Regulation of Ergosterol Biosynthesis in Saccharomyces cerevisiae. Genes . 2020;11. doi:10.3390/genes11070795
- 24. Hornby JM, Jensen EC, Lisec AD, Tasto JJ, Jahnke B, Shoemaker R, et al. Quorum sensing in the dimorphic fungus Candida albicans is mediated by farnesol. Appl Environ Microbiol. 2001;67: 2982–2992.
- Hornby JM. Quorum sensing and the regulation of morphology in the dimorphic fungus Candida albicans. 2003. Available: https://search.proquest.com/openview/0b2bfd0efe29f67bc0222719435cd8b2/1?pq-origsite =gscholar&cbl=18750&diss=y
- Ramage G, Saville SP, Wickes BL, López-Ribot JL. Inhibition of Candida albicans biofilm formation by farnesol, a quorum-sensing molecule. Appl Environ Microbiol. 2002;68: 5459–5463.
- 27. Sanglard D, Ischer F, Parkinson T, Falconer D, Bille J. Candida albicans mutations in the ergosterol biosynthetic pathway and resistance to several antifungal agents. Antimicrob

Agents Chemother. 2003;47: 2404–2412.

- 28. Jensen-Pergakes KL, Kennedy MA, Lees ND, Barbuch R, Koegel C, Bard M. Sequencing, disruption, and characterization of the Candida albicans sterol methyltransferase (ERG6) gene: drug susceptibility studies in erg6 mutants. Antimicrob Agents Chemother. 1998;42: 1160–1167.
- Vale-Silva LA, Coste AT, Ischer F, Parker JE, Kelly SL, Pinto E, et al. Azole resistance by loss of function of the sterol Δ^{5,6}-desaturase gene (ERG3) in Candida albicans does not necessarily decrease virulence. Antimicrob Agents Chemother. 2012;56: 1960–1968.
- 30. Young LY, Hull CM, Heitman J. Disruption of ergosterol biosynthesis confers resistance to amphotericin B in Candida Iusitaniae. Antimicrob Agents Chemother. 2003;47: 2717–2724.
- Pinjon E, Moran GP, Jackson CJ, Kelly SL, Sanglard D, Coleman DC, et al. Molecular mechanisms of itraconazole resistance in Candida dubliniensis. Antimicrob Agents Chemother. 2003;47: 2424–2437.
- Morio F, Pagniez F, Lacroix C, Miegeville M, Le Pape P. Amino acid substitutions in the Candida albicans sterol Δ5,6-desaturase (Erg3p) confer azole resistance: characterization of two novel mutants with impaired virulence. J Antimicrob Chemother. 2012;67: 2131–2138.
- Ksiezopolska E, Schikora-Tamarit MÀ, Beyer R, Nunez-Rodriguez JC, Schüller C, Gabaldón T. Narrow mutational signatures drive acquisition of multidrug resistance in the fungal pathogen Candida glabrata. Curr Biol. 2021;31: 5314–5326.e10.
- 34. Vandeputte P, Tronchin G, Larcher G, Ernoult E, Bergès T, Chabasse D, et al. A nonsense mutation in the ERG6 gene leads to reduced susceptibility to polyenes in a clinical isolate of Candida glabrata. Antimicrob Agents Chemother. 2008;52: 3701–3709.
- 35. Carolus H, Sofras D, Boccarella G, Sephton-Clark P, Romero CL, Vergauwen R, et al. Acquired amphotericin B resistance and fitness trade-off compensation in Candida auris. Research Square. 2023. doi:10.21203/rs.3.rs-3621420/v1
- 36. Gao J, Wang H, Li Z, Wong AH-H, Wang Y-Z, Guo Y, et al. Candida albicans gains azole resistance by altering sphingolipid composition. Nat Commun. 2018;9: 1–14.
- Rybak JM, Dickens CM, Parker JE, Caudle KE, Manigaba K, Whaley SG, et al. Loss of C-5 Sterol Desaturase Activity Results in Increased Resistance to Azole and Echinocandin Antifungals in a Clinical Isolate of Candida parapsilosis. Antimicrob Agents Chemother. 2017;61. doi:10.1128/AAC.00651-17
- Kennedy MA, Johnson TA, Lees ND, Barbuch R, Eckstein JA, Bard M. Cloning and sequencing of the Candida albicans C-4 sterol methyl oxidase gene (ERG25) and expression of an ERG25 conditional lethal mutation in Saccharomyces cerevisiae. Lipids. 2000;35: 257–262.
- 39. Kim SH, Steere L, Zhang Y-K, McGregor C, Hahne C, Zhou Y, et al. Inhibiting C-4 Methyl Sterol Oxidase with Novel Diazaborines to Target Fungal Plant Pathogens. ACS Chem Biol. 2022;17: 1343–1350.

- 40. Kodedová M, Sychrová H. Changes in the Sterol Composition of the Plasma Membrane Affect Membrane Potential, Salt Tolerance and the Activity of Multidrug Resistance Pumps in Saccharomyces cerevisiae. PLoS One. 2015;10: e0139306.
- 41. Li Y, Dai M, Zhang Y, Lu L. The sterol C-14 reductase Erg24 is responsible for ergosterol biosynthesis and ion homeostasis in Aspergillus fumigatus. Appl Microbiol Biotechnol. 2021;105: 1253–1268.
- 42. Gupta SS, Ton V-K, Beaudry V, Rulli S, Cunningham K, Rao R. Antifungal activity of amiodarone is mediated by disruption of calcium homeostasis. J Biol Chem. 2003;278: 28831–28839.
- 43. Barreto L, Canadell D, Petrezsélyová S, Navarrete C, Maresová L, Peréz-Valle J, et al. A genomewide screen for tolerance to cationic drugs reveals genes important for potassium homeostasis in Saccharomyces cerevisiae. Eukaryot Cell. 2011;10: 1241–1250.
- 44. Luna-Tapia A, Peters BM, Eberle KE, Kerns ME, Foster TP, Marrero L, et al. ERG2 and ERG24 Are Required for Normal Vacuolar Physiology as Well as Candida albicans Pathogenicity in a Murine Model of Disseminated but Not Vaginal Candidiasis. Eukaryot Cell. 2015;14: 1006–1016.
- 45. Kim SH, Iyer KR, Pardeshi L, Muñoz JF, Robbins N, Cuomo CA, et al. Genetic Analysis of Candida auris Implicates Hsp90 in Morphogenesis and Azole Tolerance and Cdr1 in Azole Resistance. MBio. 2019;10. doi:10.1128/mBio.02529-18
- 46. Robbins N, Cowen LE. Roles of Hsp90 in Candida albicans morphogenesis and virulence. Curr Opin Microbiol. 2023;75: 102351.
- 47. Maesaki S, Marichal P, Vanden Bossche H, Sanglard D, Kohno S. Rhodamine 6G efflux for the detection of CDR1-overexpressing azole-resistant Candida albicans strains. J Antimicrob Chemother. 1999;44: 27–31.
- 48. Mehmood A, Liu G, Wang X, Meng G, Wang C, Liu Y. Fungal Quorum-Sensing Molecules and Inhibitors with Potential Antifungal Activity: A Review. Molecules. 2019;24. doi:10.3390/molecules24101950
- 49. Nickerson KW, Atkin AL, Hornby JM. Quorum sensing in dimorphic fungi: farnesol and beyond. Appl Environ Microbiol. 2006;72: 3805–3813.
- 50. Yu L-H, Wei X, Ma M, Chen X-J, Xu S-B. Possible inhibitory molecular mechanism of farnesol on the development of fluconazole resistance in Candida albicans biofilm. Antimicrob Agents Chemother. 2012;56: 770–775.
- 51. Song J, Liu X, Li R. Sphingolipids: Regulators of azole drug resistance and fungal pathogenicity. Mol Microbiol. 2020;114: 891–905.
- 52. Kadosh D, Johnson AD. Induction of the Candida albicans filamentous growth program by relief of transcriptional repression: a genome-wide analysis. Mol Biol Cell. 2005;16: 2903–2912.
- 53. Villa S, Hamideh M, Weinstock A, Qasim MN, Hazbun TR, Sellam A, et al. Transcriptional control of hyphal morphogenesis in Candida albicans. FEMS Yeast Res. 2020;20.

doi:10.1093/femsyr/foaa005

- 54. Znaidi S, Nesseir A, Chauvel M, Rossignol T, d'Enfert C. A comprehensive functional portrait of two heat shock factor-type transcriptional regulators involved in Candida albicans morphogenesis and virulence. PLoS Pathog. 2013;9: e1003519.
- 55. Carbrey JM, Cormack BP, Agre P. Aquaporin in Candida: characterization of a functional water channel protein. Yeast. 2001;18: 1391–1396.
- 56. Gong Y, Li T, Yu C, Sun S. Candida albicans Heat Shock Proteins and Hsps-Associated Signaling Pathways as Potential Antifungal Targets. Front Cell Infect Microbiol. 2017;7: 520.
- 57. Yan L, Li M, Cao Y, Gao P, Cao Y, Wang Y, et al. The alternative oxidase of Candida albicans causes reduced fluconazole susceptibility. J Antimicrob Chemother. 2009;64: 764–773.
- 58. Martchenko M, Alarco A-M, Harcus D, Whiteway M. Superoxide dismutases in Candida albicans: transcriptional regulation and functional characterization of the hyphal-induced SOD5 gene. Mol Biol Cell. 2004;15: 456–467.
- 59. Burgain A, Tebbji F, Khemiri I, Sellam A. Metabolic Reprogramming in the Opportunistic Yeast Candida albicans in Response to Hypoxia. mSphere. 2020;5. doi:10.1128/mSphere.00913-19
- 60. Plaine A, Walker L, Da Costa G, Mora-Montes HM, McKinnon A, Gow NAR, et al. Functional analysis of Candida albicans GPI-anchored proteins: roles in cell wall integrity and caspofungin sensitivity. Fungal Genet Biol. 2008;45: 1404–1414.
- Richard M, de Groot P, Courtin O, Poulain D, Klis F, Gaillardin C. GPI7 affects cell-wall protein anchorage in Saccharomyces cerevisiae and Candida albicans. Microbiology. 2002;148: 2125–2133.
- 62. Victoria GS, Yadav B, Hauhnar L, Jain P, Bhatnagar S, Komath SS. Mutual co-regulation between GPI-N-acetylglucosaminyltransferase and ergosterol biosynthesis in Candida albicans. Biochem J. 2012;443: 619–625.
- 63. Yadav B, Bhatnagar S, Ahmad MF, Jain P, Pratyusha VA, Kumar P, et al. First step of glycosylphosphatidylinositol (GPI) biosynthesis cross-talks with ergosterol biosynthesis and Ras signaling in Candida albicans. J Biol Chem. 2014;289: 3365–3382.
- 64. Doedt T, Krishnamurthy S, Bockmühl DP, Tebarth B, Stempel C, Russell CL, et al. APSES proteins regulate morphogenesis and metabolism in Candida albicans. Mol Biol Cell. 2004;15: 3167–3180.
- 65. White SJ, Rosenbach A, Lephart P, Nguyen D, Benjamin A, Tzipori S, et al. Self-regulation of Candida albicans population size during GI colonization. PLoS Pathog. 2007;3: e184.
- MacPherson S, Akache B, Weber S, De Deken X, Raymond M, Turcotte B. Candida albicans zinc cluster protein Upc2p confers resistance to antifungal drugs and is an activator of ergosterol biosynthetic genes. Antimicrob Agents Chemother. 2005;49: 1745–1752.
- 67. Silver PM, Oliver BG, White TC. Role of Candida albicans transcription factor Upc2p in drug

resistance and sterol metabolism. Eukaryot Cell. 2004;3: 1391–1397.

- 68. Lv Q-Z, Yan L, Jiang Y-Y. The synthesis, regulation, and functions of sterols in Candida albicans: Well-known but still lots to learn. Virulence. 2016;7: 649–659.
- 69. Revie NM, Iyer KR, Maxson ME, Zhang J, Yan S, Fernandes CM, et al. Targeting fungal membrane homeostasis with imidazopyrazoindoles impairs azole resistance and biofilm formation. Nat Commun. 2022;13: 1–20.
- 70. Pierson CA, Eckstein J, Barbuch R, Bard M. Ergosterol gene expression in wild-type and ergosterol-deficient mutants of Candidaalbicans. Med Mycol. 2004;42: 385–389.
- Veen M, Stahl U, Lang C. Combined overexpression of genes of the ergosterol biosynthetic pathway leads to accumulation of sterols in Saccharomyces cerevisiae. FEMS Yeast Res. 2003;4: 87–95.
- 72. Crawford AC, Lehtovirta-Morley LE, Alamir O, Niemiec MJ, Alawfi B, Alsarraf M, et al. Biphasic zinc compartmentalisation in a human fungal pathogen. PLoS Pathog. 2018;14: e1007013.
- 73. Sanchez AA, Johnston DA, Myers C, Edwards JE Jr, Mitchell AP, Filler SG. Relationship between Candida albicans virulence during experimental hematogenously disseminated infection and endothelial cell damage in vitro. Infect Immun. 2004;72: 598–601.
- 74. Kukurudz RJ, Chapel M, Wonitowy Q, Adamu Bukari A-R, Sidney B, Sierhuis R, et al. Acquisition of cross-azole tolerance and aneuploidy in Candida albicans strains evolved to posaconazole. G3 . 2022;12. doi:10.1093/g3journal/jkac156
- 75. Yang Feng, Scopel Eduardo F. C., Li Hao, Sun Liu-liu, Kawar Nora, Cao Yong-bing, et al. Antifungal Tolerance and Resistance Emerge at Distinct Drug Concentrations and Rely upon Different Aneuploid Chromosomes. MBio. 2023;14: e00227–23.
- 76. Pompei S, Lagomarsino MC. A fitness trade-off explains the early fate of yeast aneuploids with chromosome gains. Proceedings of the National Academy of Sciences. 2023;120: e2211687120.
- 77. Taylor AM, Shih J, Ha G, Gao GF, Zhang X, Berger AC, et al. Genomic and Functional Approaches to Understanding Cancer Aneuploidy. Cancer Cell. 2018;33: 676–689.e3.
- 78. Kolodner RD, Cleveland DW, Putnam CD. Cancer. Aneuploidy drives a mutator phenotype in cancer. Science. 2011. pp. 942–943.
- Yona AH, Manor YS, Herbst RH, Romano GH, Mitchell A, Kupiec M, et al. Chromosomal duplication is a transient evolutionary solution to stress. Proc Natl Acad Sci U S A. 2012;109: 21010–21015.
- Hirakawa MP, Martinez DA, Sakthikumar S, Anderson MZ, Berlin A, Gujja S, et al. Genetic and phenotypic intra-species variation in Candida albicans. Genome Res. 2015;25: 413–425.
- Flowers SA, Barker KS, Berkow EL, Toner G, Chadwick SG, Gygax SE, et al. Gain-of-function mutations in UPC2 are a frequent cause of ERG11 upregulation in azole-resistant clinical isolates of Candida albicans. Eukaryot Cell. 2012;11: 1289–1299.

- 82. Flowers SA, Colón B, Whaley SG, Schuler MA, Rogers PD. Contribution of clinically derived mutations in ERG11 to azole resistance in Candida albicans. Antimicrob Agents Chemother. 2015;59: 450–460.
- Rybak Jeffrey M., Sharma Cheshta, Doorley Laura A., Barker Katherine S., Palmer Glen E., Rogers P. David. Delineation of the Direct Contribution of Candida auris ERG11 Mutations to Clinical Triazole Resistance. Microbiology Spectrum. 2021;9: e01585–21.
- 84. Burrack LS, Todd RT, Soisangwan N, Wiederhold NP, Selmecki A. Genomic Diversity across Candida auris Clinical Isolates Shapes Rapid Development of Antifungal Resistance In Vitro and In Vivo. MBio. 2022;13: e0084222.
- 85. Li J, Aubry L, Brandalise D, Coste AT, Sanglard D, Lamoth F. Upc2-mediated mechanisms of azole resistance in Candida auris. Microbiol Spectr. 2024; e0352623.
- 86. Glazier Virginia E., Kramara Juraj, Ollinger Tomye, Solis Norma V., Zarnowski Robert, Wakade Rohan S., et al. The Candida albicans reference strain SC5314 contains a rare, dominant allele of the transcription factor Rob1 that modulates filamentation, biofilm formation, and oral commensalism. MBio. 2023;14: e01521–23.
- Vande Zande P, Siddiq MA, Hodgins-Davis A, Kim L, Wittkopp PJ. Active compensation for changes in TDH3 expression mediated by direct regulators of TDH3 in Saccharomyces cerevisiae. PLoS Genet. 2023;19: e1011078.
- Shen J, Guo W, Köhler JR. CaNAT1, a heterologous dominant selectable marker for transformation of Candida albicans and other pathogenic Candida species. Infect Immun. 2005;73: 1239–1242.
- 89. Veri AO, Miao Z, Shapiro RS, Tebbji F, O'Meara TR, Kim SH, et al. Tuning Hsf1 levels drives distinct fungal morphogenetic programs with depletion impairing Hsp90 function and overexpression expanding the target space. PLoS Genet. 2018;14: e1007270.
- 90. Gerami-Nejad M, Forche A, McClellan M, Berman J. Analysis of protein function in clinical C. albicans isolates. Yeast. 2012;29: 303–309.
- 91. Patro R, Duggal G, Love MI, Irizarry RA, Kingsford C. Salmon provides fast and bias-aware quantification of transcript expression. Nat Methods. 2017;14: 417–419.
- 92. Soneson C, Love MI, Robinson MD. Differential analyses for RNA-seq: transcript-level estimates improve gene-level inferences. F1000Res. 2015;4: 1521.
- 93. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. Genome Biol. 2014;15: 550.
- 94. Skrzypek MS, Binkley J, Binkley G, Miyasato SR, Simison M, Sherlock G. The Candida Genome Database (CGD): incorporation of Assembly 22, systematic identifiers and visualization of high throughput sequencing data. Nucleic Acids Res. 2017;45: D592–D596.
- 95. Todd RT, Braverman AL, Selmecki A. Flow Cytometry Analysis of Fungal Ploidy. Curr Protoc Microbiol. 2018;50: e58.
- 96. Selmecki A, Forche A, Berman J. Aneuploidy and isochromosome formation in drug-resistant Candida albicans. Science. 2006;313: 367–370.

- 97. Bushnell B. BBTools software package. 2014. Available online: http://sourceforgenet/projects/bbmap.
- 98. Li H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv [q-bio.GN]. 2013. Available: http://arxiv.org/abs/1303.3997
- 99. Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, et al. Twelve years of SAMtools and BCFtools. Gigascience. 2021;10. doi:10.1093/gigascience/giab008
- 100. Andrews S. FastQC: A quality control analysis tool for high throughput sequencing data. Github; Available: https://github.com/s-andrews/FastQC
- 101. Okonechnikov K, Conesa A, García-Alcalde F. Qualimap 2: advanced multi-sample quality control for high-throughput sequencing data. Bioinformatics. 2016;32: 292–294.
- 102. Ewels P, Magnusson M, Lundin S, Käller M. MultiQC: summarize analysis results for multiple tools and samples in a single report. Bioinformatics. 2016;32: 3047–3048.
- 103. Abbey DA, Funt J, Lurie-Weinberger MN, Thompson DA, Regev A, Myers CL, et al. YMAP: a pipeline for visualization of copy number variation and loss of heterozygosity in eukaryotic pathogens. Genome Med. 2014;6: 100.
- 104. Van der Auwera GA, O'Connor BD. Genomics in the Cloud: Using Docker, GATK, and WDL in Terra. "O'Reilly Media, Inc."; 2020.
- 105. Cingolani P, Platts A, Wang LL, Coon M, Nguyen T, Wang L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of Drosophila melanogaster strain w1118; iso-2; iso-3. Fly . 2012;6: 80–92.
- 106. Robinson JT, Thorvaldsdóttir H, Wenger AM, Zehir A, Mesirov JP. Variant Review with the Integrative Genomics Viewer. Cancer Res. 2017;77: e31–e34.
- 107. Hornby JM, Nickerson KW. Enhanced production of farnesol by Candida albicans treated with four azoles. Antimicrob Agents Chemother. 2004;48: 2305–2307.