Expanded View Figures

Figure EV1. Ubiquitylation of mitochondria is dependent on MOMP by BAX/BAK pores, but independent of caspase activity.

(A) U2OS EMPTY^{CRISPR} and BAX/BAK^{CRISPR} cells were treated with 10 µM ABT-737, 2 µM S63845 with or without 20 µM Q-VD-OPh. Cell death was determined using Sytox Green inclusion normalised to starting confluence. Graph is representative of three independent experiments and displays mean values ± s.e.m. (error bars) of technical triplicates. (B) Lysates from U2OS EMPTY^{CRISPR} and BAX/BAK^{CRISPR} cells were blotted for BAX, BAK and Actin. (C) SVEC4-10 EMPTY^{CRISPR} and BAX/BAK^{CRISPR} cells were treated for 1 h with 10 µM ABT-737, 10 µM S63845 and 30 µM Q-VD-OPh. Mitochondria were isolated using digitonin fractionation buffer and blotted for ubiquitin (UBCJ2), BAX, BAK, HSP60 and Actin. (D) SVEC4-10 cells were treated for 1 h with 10 µM ABT-737, 10 µM S63845 with or without 30 µM Q-VD-OPh. Mitochondria were isolated using digitonin fractionation buffer and lysates were blotted for ubiquitin (UBCJ2), HSP60 and actin. (E) U2OS cells were treated for 7 h with 10 µM ABT-737, 2 µM S63845 and 20 µM Q-VD-OPh with or without the addition of 10 µM MG-132. Lysates were blotted for ubiquitin (UBCJ2) and GAPDH. (F) U2OS cells were treated for 24 h with 10 µM ABT-737, 2 µM S63845 and 20 µM Q-VD-OPh with or without the addition of 10 µM MG-132. Mitochondrial depletion was assessed by blotting for several mitochondrial proteins and GAPDH. (G) U2OS cells were treated for 7 h with 10 µM ABT-737, 2 µM S63845 and 20 µM Q-VD-OPh with or without the addition of 1 µM TAK-243. Lysates were blotted for ubiquitin (UBCJ2) and GAPDH. (H) U2OS cells were treated for 24 h with 10 µM ABT-737, 2 µM S63845 and 20 µM Q-VD-OPh with or without the addition of 1 µM TAK-243. Lysates were blotted for z h with 10 µM ABT-737, 2 µM S63845 and 20 µM Q-VD-OPh with or without the addition of 1 µM TAK-243. Mitochondrial depletion was assessed by blotting for several mitochondrial proteins and GAPDH. Data information: (C, D, E, F, G, H) blots are representative of three independent experiments.







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Figure EV2. Mitochondrial ubiquitylation and GFP-NEMO translocation can be blocked by E1 inhibition and is independent of neddylation.

(A) SVEC4-10 cells treated for 3 h with 10 μ M ABT-737, 10 μ M S63845 and 30 μ M Q-VD-OPh. Mitochondria were isolated using Dounce homogeniser and cellular fractions were probed with relevant antibodies. Mitochondrial localised NEMO was quantified normalising to mitochondrial content defined by HSP60 signal. (B) SVEC4-10 cells pre-treated with 2 μ M TAK-243 for 1 h followed by additional 1 h treatment with 10 μ M ABT-737, 10 μ M S63845 and 30 μ M Q-VD-OPh with or without 2 μ M TAK-243. Blots are representative for four independent experiments. (C) Upper: SVEC4-10 cells expressing GFP-NEMO were pre-treated for 1 h with 2 μ M TAK-243 followed by 1 h treatment of 10 μ M ABT-737, 10 μ M S63845, 30 μ M Q-VD-OPh with or without 2 μ M TAK-243. Cells were immunostained for TOM20 and ubiquitin (FK2). Scale bar is 50 μ m and images are representative for three independent experiments. Lower: quantification showing the percentage of cells with mitochondrial localised GFP-NEMO and ubiquitin puncta. (D) SVEC4-10 cells pre-treated with 1 μ M MLN4924 (NAE inhibitor) for 1 h followed by additional 1 h treatment with 10 μ M ABT-737, 10 μ M S63845 and 30 μ M Q-VD-OPh with or without 1 μ M MLN4924. Blots are representative for two independent experiments. (E) SVEC4-10 cells expressing GFP-NEMO were immunostained for 1 h with 2 μ M TAK-243. Cells were immunostained for 1 h followed by additional 1 h treatment with 10 μ M S63845 and 30 μ M Q-VD-OPh with or without 1 μ M MLN4924. Blots are representative for two independent experiments. (E) SVEC4-10 cells expressing GFP-NEMO pre-treated with 1 μ M MLN4924 for 1 h followed by additional 1 h treatment with 10 μ M ABT-737, 10 μ M S63845 and 30 μ M Q-VD-OPh with or without 1 μ M MLN4924. Cells were immunostained for mitochondrial TOM20 and DAPI. Images are representative for three independent experiments and are shown with a 50 μ m scale bar. (F) Quantification of (E) showing the percentage of cells with mitochondrial translocation of GFP-NEMO. Data informa



Figure EV3. Loss of NEMO cannot be rescued during CICD by expressing non-ubiquitin-binding mutants of NEMO.

(A) MEF Tnf^{-/}/Hoip^{+/+} and Tnf^{-/}/Hoip^{-/-} expressing GFP-NEMO were treated for 3 h with 10 μ M ABT-737, 5 μ M S63845 and 30 μ M Q-VD-OPh. Cells were immunostained for mitochondrial TOM20 and DAPI. Images are representative of three independent experiments. (B) Quantification of A showing the percentage of cells with mitochondrial translocation of GFP-NEMO. Graph displays mean values ± s.e.m. (error bars) of n = 3 independent experiments. Statistical analysis was performed using two-way ANOVA with Tukey correction. (C) Validation of SVEC4-10, SVEC4-10 GFP-NEMO and SVEC4-10 GFP-D311N cells transfected with NTC or siNEMO. Lysates were blotted for NEMO and α -tubulin. (D) SVEC4-10, SVEC4-10 GFP-NEMO and SVEC4-10 GFP-D311N cells transfected with NTC or siNEMO were treated for 1 h with 10 μ M ABT-737, 10 μ M S63845 and 30 μ M Q-VD-OPh. Cells were immunostained for p65 and DAPI. Images are representative of two independent experiments. (E) Quantification of (D) showing the percentage of cells with nuclear translocation of p65. Graph displays mean values ± s.e.m. (error bars) of n = 2 independent experiments. ****P < 0.0001.

Α	PINK1	
/EC		R ² = 0.95 Edit eff = 92
Ś		KO-score = 72
SVEC GFP-NEMO		R ² = 0.91 Edit eff = 91 KO-score = 68





0.

. Tnf

Кc

Ccl5

Actin

Figure EV4. Validation of PINK1^{CRISPR}, NIK^{CRISPR}, MUL1/MARCH5^{CRISPR} and XIAP^{CRISPR} knockout cell lines.

(A) Validation of PINK1 knockout in SVEC4-10 cells with or without GFP-NEMO expression using genomic PCR and ICE (inference of CRISPR edits) analysis. (B) SVEC4-10 EMPTY^{CRISPR} and PINK1^{CRISPR} cells expressing YFP-Parkin were treated for 1 h with 10 μM ABT-737, 10 μM S63845 and 30 μM Q-VD-OPh or for 3 h with 10 μM CCCP. Mitochondria were immunostained with HSP60 and DAPI. Images are representative of two independent experiments and displayed with 50 μm scale bar. (C) Validation of MUL1 knockout in SVEC4-10 MUL1/MARCH5^{CRISPR} cells using genomic PCR and ICE analysis. (D) Validation of MARCH5 knockout in SVEC4-10 MUL1/MARCH5^{CRISPR} cells using genomic PCR and ICE analysis. (D) Validation of MARCH5 knockout in SVEC4-10 MUL1/MARCH5^{CRISPR} cells using genomic PCR and ICE analysis. (E) Validation of SVEC4-10 XIAP^{CRISPR} cells with and without GFP-NEMO expression using western blot. Lysates were blotted for XIAP and Actin. (F) *Tnf, Kc*, and *CclS* expression of SVEC4-10 EMPTY^{CRISPR} and XIAP^{CRISPR} cells treated with 10 μM ABT-737, 10 μM S63845 and 30 μM Q-VD for 3 h. Graph is representative of three independent experiments. (G) Validation of NIK knockout in GFP-NEMO expressing SVEC4-10 EMPTY^{CRISPR} cells. Cells were treated for 2 h with 10 μM MG-132. Lysates were blotted for NIK and actin.



Figure EV5. Raptinal induces cell death independent of mitochondrial permeabilization by BAX and BAK.

(A) SVEC4-10 cells were treated for 2 h with 1 µM rotenone, 1 µM oligomycin, 5 µM antimycin A, 3 µM erastin or the combination 10 µM ABT-737, 10 µM S63845 and 30 µM Q-VD-OPh. Mitochondria were isolated using digitonin fractionation buffer and antibodies against ubiquitin (UBCJ2), HSP60 and GAPDH were used. (B) SVEC4-10 cells expressing GFP-NEMO were treated for 2 h with 1 µM rotenone, 1 µM oligomycin, 5 µM antimycin A, 3 µM erastin or the combination 10 µM ABT-737, 10 µM S63845 and 30 µM Q-VD-OPh. Cells were immunostained with mitochondrial TOM20 and DAPI for confocal microscopy. Graph shows the quantification of three independent experiments in which the percentage of cells with mitochondrial localisation of GFP-NEMO was analysed. (C) SVEC4-10 cells were treated for 2 h with 1 µM rotenone, 1 and 5 µM antimycin A or 3 µM erastin. ROS levels were determined using MitoSOX Red via flow cytometry. Graph displays mean values ± s.e.m. (error bars) of n = 3 independent experiments. (D) SVEC4-10 cells were treated with 0.5, 1, 5, 10 and 20 µM erastin. Mitochondrial calcium was measured using Rhod2-AM via flow cytometry. Graph displays mean values \pm s.e.m. (error bars) of n = 3 independent experiments. (E) SVEC4-10 cells were treated for 2 h with 10 μ M ABT-737, 10 μ M S63845 and 30 µM Q-VD-OPh with or without 25 µM cyclosporin A. Mitochondria were isolated using digitonin fractionation buffer and antibodies against ubiquitin (UBCJ2), HSP60 and GADPH were used. (F) SVEC4-10 cells expressing GFP-NEMO were treated for 2 h with 10 µM ABT-737, 10 µM S63845 and 30 µM Q-VD-OPh with or without 25 µM cyclosporin A. Cells were immunostained with mitochondrial TOM20 and DAPI for confocal microscopy. Graph shows the quantification of three independent experiments showing the percentage of cells with mitochondrial localisation of GFP-NEMO, error bars represent s.e.m. (G) EMPTY^{CRISPR} and BAX/BAK^{CRISPR} validation of SVEC4-10 cells and SVEC4-10 cells expressing GFP-NEMO. Lysates for blotted for BAX, BAK and α-tubulin. (H) SVEC4-10 EMPTY^{CRISPR} and SVEC4-10 BAX/BAK^{CRISPR} cells treated with 10 µM ABT-737 and 10 µM S63845 or treated with 2.5 or 10 µM raptinal. Caspase-dependency of death was assessed using 30 µM Q-VD-OPh. Cell viability was measured using Sytox Green exclusion. Graphs are representative of two independent experiments and display the mean and s.e.m. of two replicates. (I) SVEC4-10 cells treated for 3 h with 10 µM ABT-737, 10 µM S63856 and 30 µM Q-VD-OPh or 2.5 µM raptinal and 30 µM Q-VD-OPh. Expression of Kc, Tnf and Actin were validated using RT-qPCR. Two repeats of Fig. 6E. Data information: (A, E) blots are representative for three independent experiments. Statistics performed using two-way ANOVA with Tukey correction. ***P < 0.001, ****P < 0.0001.