

**Supplementary Figure 1.** Raw minute volume (mL.min<sup>-1</sup>) values for morphine, tianeptine, oliceridine, SR-17018, methadone and oxycodone dose response curves shown as percent of baseline MV in Figure 1 and Figure 2. N=6 for all groups.



**Supplementary Figure 2.** (A) Alternative regressions fit using AUC instead of maximum respiratory depression shown in Fig 3A. (B) Sigmoidal 3-parameter fit of Fig 3A. N=6 for each data point.

![](_page_2_Figure_1.jpeg)

**Supplementary Figure 3.** Raw minute volume (mL.min<sup>-1</sup>) **(A)** and raw hot plate latency (seconds) **(B)** values for morphine, tianeptine, oliceridine, SR-17018, methadone and oxycodone dose response curves shown as percent of baseline MV in Figure 3 or as %MPE in Figure 4, respectively. N=6 and N=10 for plethysmography and hot plate, respectively.

![](_page_3_Figure_1.jpeg)

**Supplementary Figure 4. (A)** Raw minute volume (mL.min<sup>-1</sup>) and percent of baseline **(B)** values for mice administered SR-17018 0.66 mg/kg by oral or i.p administration and respective vehicles. **(C)** AUC analysis of (B) shows significant depression of respiration by oral but not i.p administered SR-17018. **(D)** Raw hot plat latency (sec) and % maximum possible effect (%MPE) **(E)** for mice administered SR-17018 0.66 mg/kg by oral or i.p administration or vehicle. **(F)** AUC analysis of **(E)** shows significant antinociception by oral but not i.p. SR-17018 Comparisons in **(C-E)** made by Two-way ANOVA with Tukey's comparison. Comparison in **(F)** made by One-way ANOVA with Tukey's comparison). \* indicates p<0.05). N=6 for **(A-C)** and N=10 **(D-E)**. Statistical test details are provided in Supplementary Table 2.

![](_page_4_Figure_1.jpeg)

**Supplementary Figure 5. Induction of opioid respiratory depression in mice breathing 5% CO**<sub>2</sub>. Raw minute volume (ml.min<sup>-1</sup>) (**A**), percent of baseline MV (**B**) and AUC (**C**) values for calculated equi-effective respiratory doses for morphine (3.44 mg.kg<sup>-1</sup>), tianeptine (12.74 mg.kg<sup>-1</sup>), oliceridine (0.745 mg.kg<sup>-1</sup>), oxycodone (0.71 mg.kg<sup>-1</sup>), methadone (2.5 mg.kg<sup>-1</sup>) and SR-17018 (0.66 mg.kg<sup>-1</sup>). Comparisons in **B** were made by One-way ANOVA using Tukey's comparison. \* indicates significance to vehicle control, p<0.05. N=6 for all groups. Statistical test details are provided in Supplementary Table 2.

![](_page_5_Figure_1.jpeg)

Supplementary Figure 6. Effect of different buffers on live cell BRET measurements. Cells were treated with 10% DMSO, 4% DMSO-16% Tween (in vivo vehicle), 0.1% DMSO (in vitro vehicle) or 10  $\mu$ M DAMGO. The effect of each addition on the raw intensity counts for both bioluminescence (475 nm) and fluorescence (535 nm) emission wavelengths was measured over time. Data show mean  $\pm$  SEM of N=6 experiments performed in duplicate.

![](_page_6_Figure_1.jpeg)

Supplementary Figure 7. Standard calibration curves for SR-17018 in HPLC LCMS analysis (See Materials and Methods). SR-17018 concentration range  $0.61 - 620.86 \mu$ M; area under the curve (AUC) axis has arbitrary units.

![](_page_7_Figure_1.jpeg)

**Supplementary Figure 8. Biased agonism quantification.** Biased agonism was quantified as previously described (Kenakin et al., 2012) using predefined equations in GraphPad Prism 9.5.0 to determine transduction coefficients ( $Log[\tau/K_A]$ ) for each agonist at each pathway. **A-G** Normalised transduction coefficients ( $\Delta Log[\tau/K_A]$ ) for each agonist in each assay were determined by subtracting the transduction coefficient for DAMGO on the corresponding plate from the transduction coefficient of each agonist.  $\Delta Log[\tau/K_A]$  values for each agonist at each assay point were determined from 5 separate experimental repeats. These values were used to determine the mean  $\Delta Log[\tau/K_A]$  values for each agonist at each pathway. The mean  $\Delta Log[\tau/K_A]$  values for a particular agonist were compared between Nb33 and arrestin-3 recruitment using an unpaired two-tailed Students t-test, \* indicates significant difference p < 0.05. A significant difference in  $\Delta Log[\tau/K_A]$  values for a particular agonist for a restin-3 recruitment relative to Nb33 recruitment is indicative of bias. **H.** The LogBias factor ( $\Delta \Delta Log[\tau/K_A]$ )) was determined by calculating the difference between the  $\Delta Log[\tau/K_A]$  values for each agonist between the two signalling assays.