

**Cell Reports, Volume 43**

**Supplemental information**

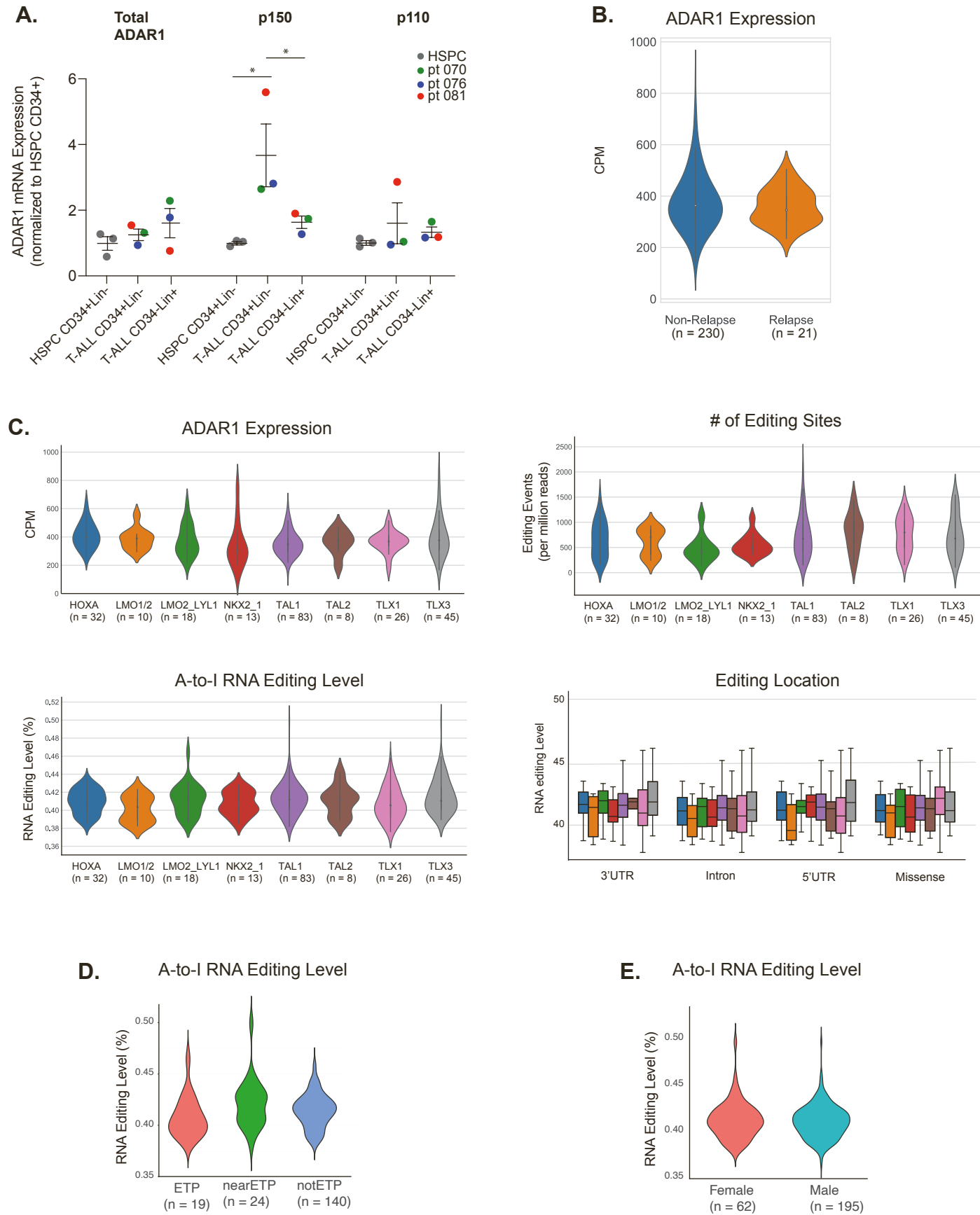
**Malignant A-to-I RNA editing by ADAR1 drives**

**T cell acute lymphoblastic leukemia relapse**

**via attenuating dsRNA sensing**

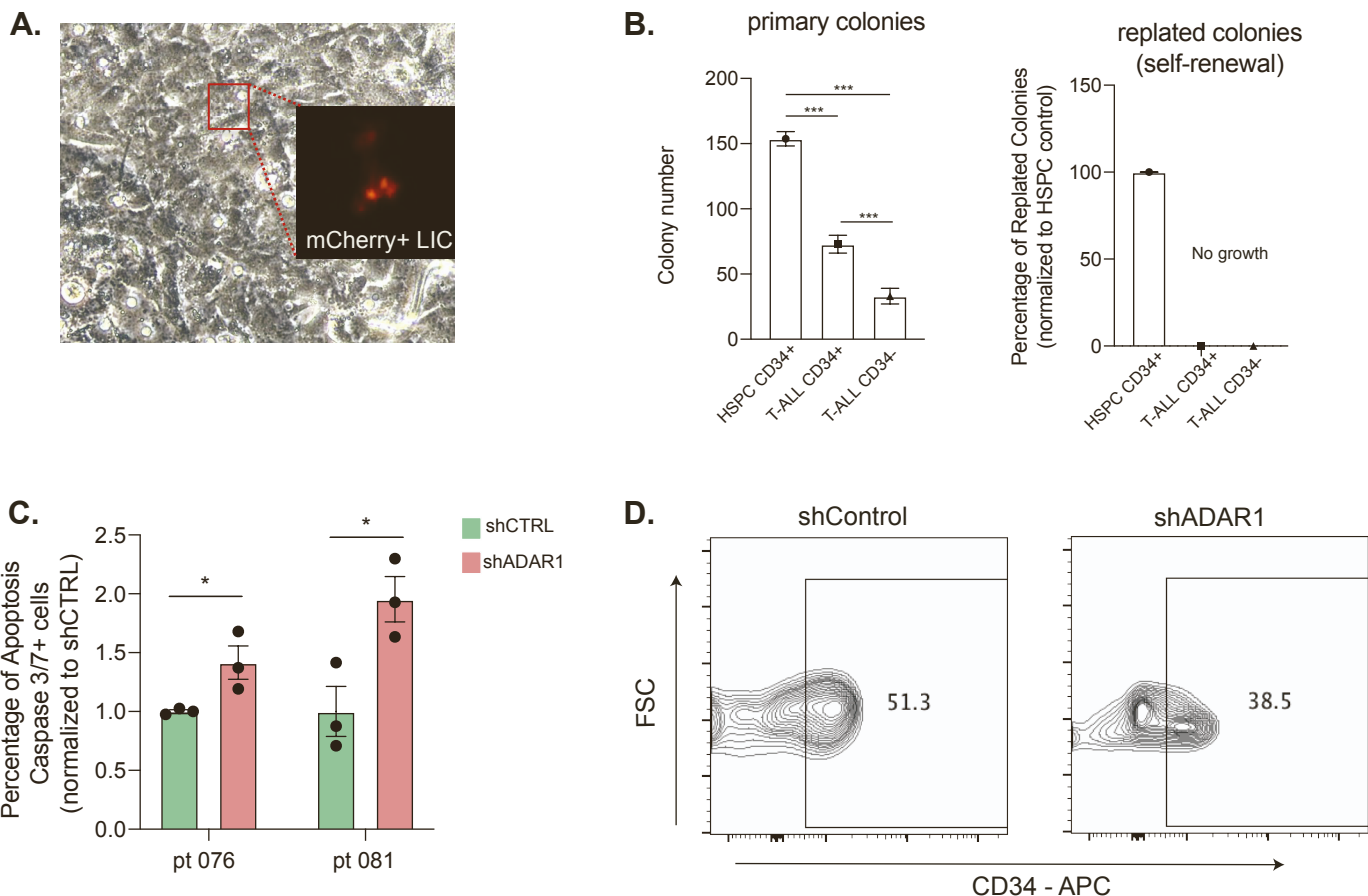
**Maria Rivera, Haoran Zhang, Jessica Pham, Jane Isquith, Qingchen Jenny Zhou, Larisa Balaian, Roman Sasik, Sabina Enlund, Adam Mark, Wenxue Ma, Frida Holm, Kathleen M. Fisch, Dennis John Kuo, Catriona Jamieson, and Qingfei Jiang**

# Supplemental Figure 1



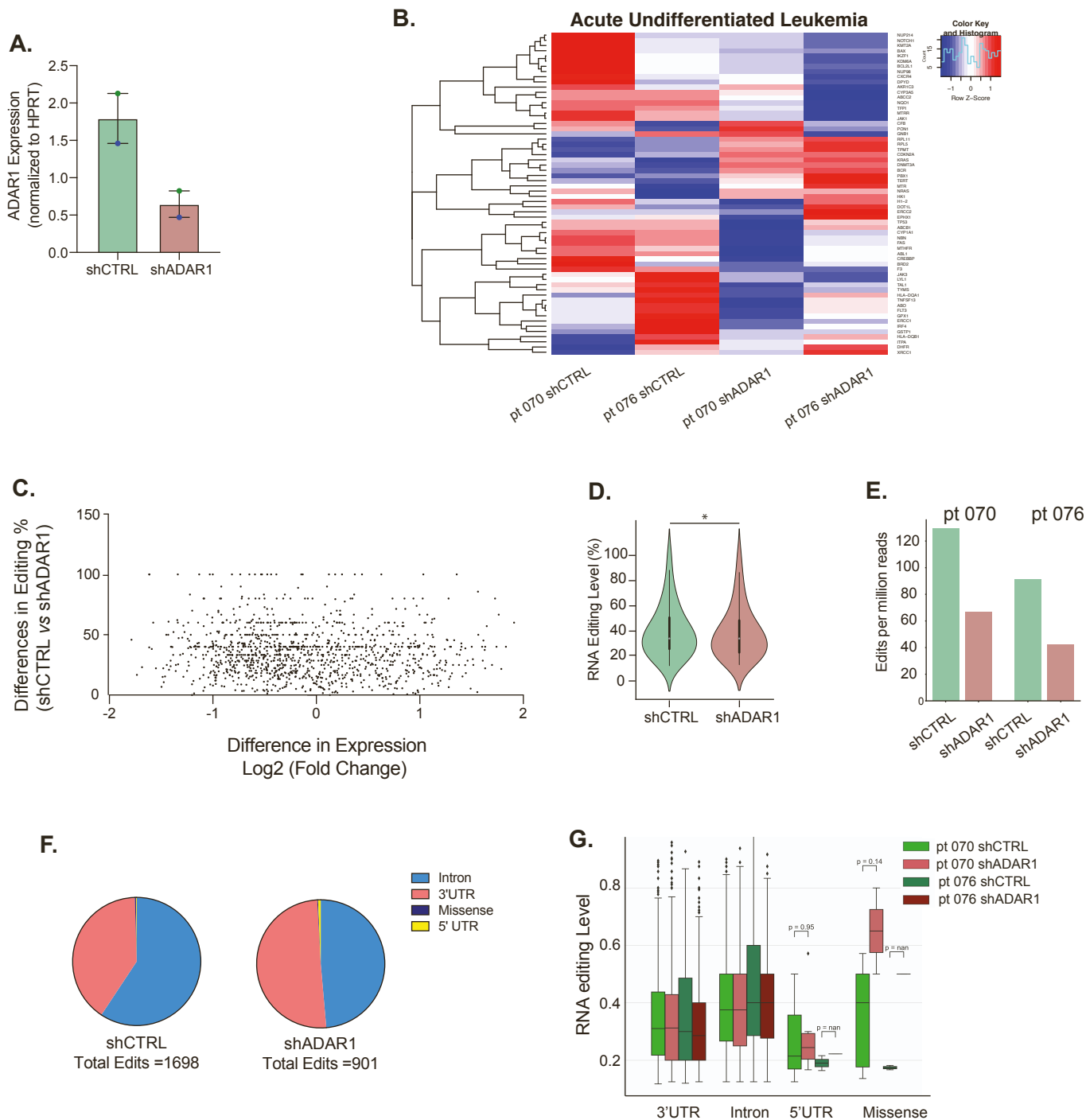
**Supplemental Figure 1, related to Figure 1. ADAR1 expression and the A-to-I Editing landscape in T-ALL.** **A.** Expression levels of total ADAR1, ADAR1 p150 and ADAR1 p110 were quantified by RT-qPCR in cord blood derived CD34+Lin- HSPC (n = 3 cord blood) and CD34+Lin- or CD34-Lin+ T-ALL LIC (n = 3 patients, pt 070, pt 076 and pt 081). Error bars represents mean with SEM. \* p < 0.05, unpaired student t-test. **B.** Expression of ADAR1 in non-relapsed and relapsed T-ALL cohort. **C.** Violin plots depicting ADAR1 expression, RNA editing level, number of editing events, and editing location were quantified based on patient molecular characteristics of NCI T-ALL TARGET dataset. **D-E.** RNA editing level based on ETP status (D) or gender (E). No significance by t-test (B and E) or one-way ANOVA (C-D).

## Supplemental Figure 2



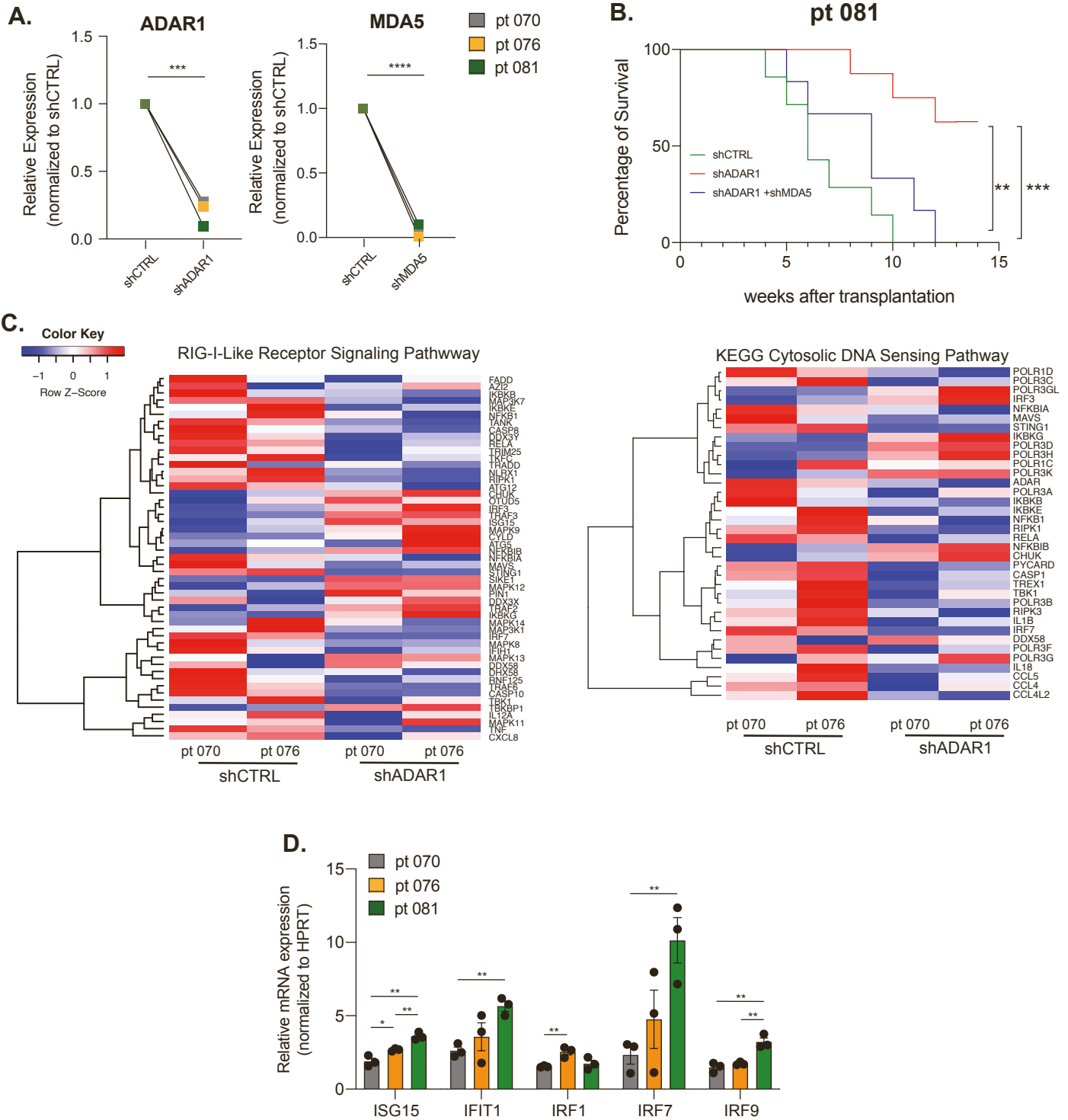
**Supplemental Figure 2, related to Figure 2. Functional characterization of T-ALL LICs in ATO co-culture and PDX systems. A.** Images of mCherry-labeled T-ALL LICs in the Artificial Organoid System (ATO). **B.** Bar plot showing number of primary colonies per 10,000 cells and percentage of replated secondary colonies. CD34<sup>+</sup>Lin<sup>-</sup> cord blood HSPC (n = 3), CD34<sup>+</sup>Lin<sup>-</sup> T-ALL LIC, and CD34<sup>+</sup>Lin<sup>+</sup> T-ALL cells were sorted from PDX mouse bone marrow (n = 3, pt 070, pt 076, and pt 081). **C.** Percentage of apoptosis after ADAR1 knockdown in T-ALL LIC (pt 076 and pt 081, n = 3 experiments). **D.** Representative flow of CD45<sup>+</sup>Lin<sup>-</sup>CD34<sup>+</sup> LICs cultured in ATOs. Cells were gated on CD45<sup>+</sup>Lin<sup>-</sup> population. T-ALL LICs were transduced with control or shRNA lentiviral vectors targeting ADAR1 for 3 days and cultured on MS5-DLL4 stroma. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001, unpaired student t-test.

# Supplemental Figure 3



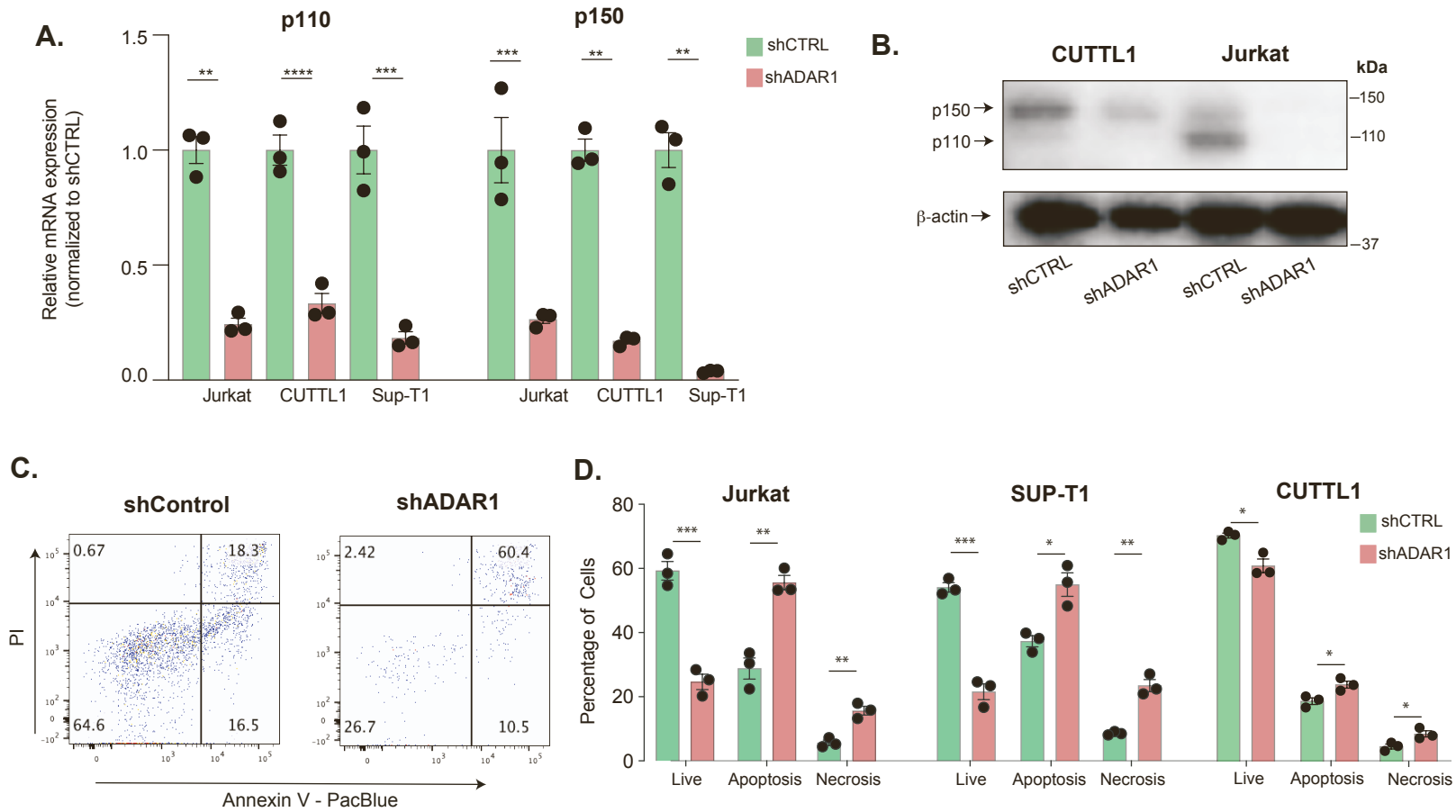
**Supplemental Figure. 3, related to Figure 3. ADAR1 knockdown reduces A-to-I RNA editing and leads to differential gene expression in T-ALL LICs. A.** ADAR1 expression in LICs after transduction with shADAR1 by RT-qPCR (n = 2 patient samples). **B.** Heatmap depicting genes in Acute Undifferentiated Leukemia pathway in T-ALL LICs with ADAR1 knockdown. **C.** Correlation of differential expression and differential RNA editing level per gene in shCTRL cells compared with shADAR1 T-ALL LICs (n = 2 samples). **D.** Overall RNA editing level was quantified by RNA-seq (n = 2 samples). \*p < 0.05, unpaired student t-test. **E.** Number of edits per million reads in each patient. **F.** Pie chart showing RNA editing locations in scramble control or shADAR1 in T-ALL LICs (n = 2 samples). **G.** Level of RNA editing (A-to-G ratio) in RNA locations.

## Supplemental Figure 4



**Supplemental Figure 4, related to Figure 4. Cell intrinsic differences of dsRNA sensing in T-ALL LICs. A.** Expression of ADAR1 and MDA5 was quantified in double-knockdown in FACS-sorted mCherry+EGFP+ T-ALL LICs ( $n = 3$ , pt 070, pt 076, and pt 081). **B.** Survival curve of mice serially transplanted with pt 081 T-ALL LICs isolated from scramble control shRNA (shCTRL), shADAR1, or co-knockdown of shADAR1 and shMDA5 primary xenografts ( $n = 6-8$  mice per group).  $**p < 0.01$ ,  $***p < 0.001$ ,  $p$  value determined by the Mantel–Cox log-rank test. **C.** Heatmap shows the upregulation (red) and downregulation (blue) of genes that were differentially expressed in RIG-I signaling pathway and Cytosolic DNA sensing pathways during comparison of T-ALL LICs (CD34+Lin-) transduced with a scramble shRNA lentivirus or ADAR1 targeting shRNA ( $n = 2$  patient samples, patient 070 and patient 076). **D.** Expression of ISGs and IFN signaling components were quantified by RT-qPCR in T-ALL LICs.  $n = 3$  independent experiments. **A** and **D.** Error bars represent mean with SEM.  $*p < 0.05$ ,  $**p < 0.01$ ,  $***p < 0.001$ , unpaired student t-test.

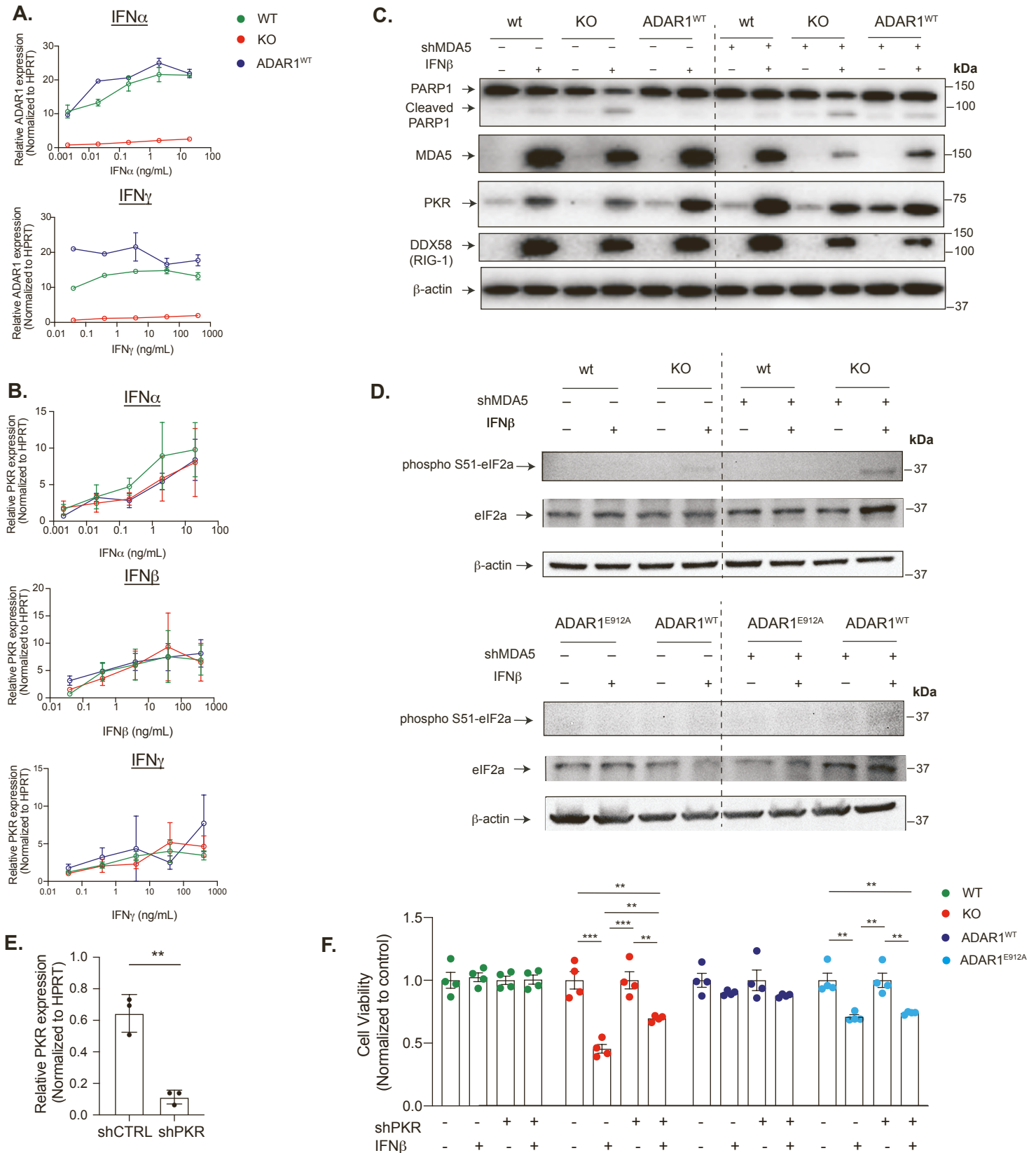
## Supplemental Figure 5



### Supplemental Figure 5, related to Figure 5. ADAR1 knockdown induces apoptosis in T-ALL cell lines.

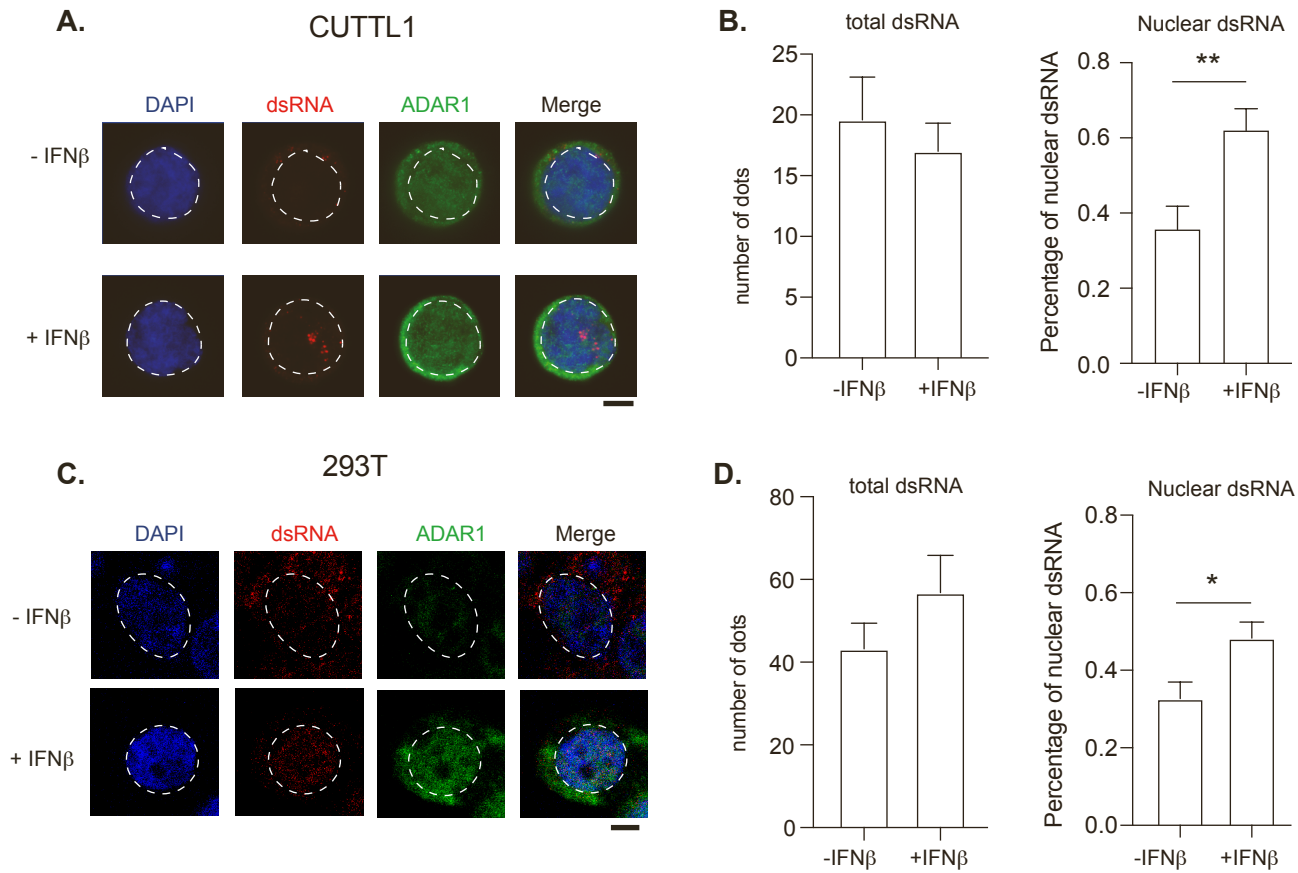
**A-B.** Expression of ADAR1 was quantified by RT-qPCR (A, 3 independent experiments) or western blot (B) in T-ALL cell lines. **C.** Representative flow showing increased Annexin V and PI staining in SUP-T1 cells after ADAR1 knockdown. **D.** Quantification of live, apoptosis, and necrosis populations in control and ADAR1 knock-down conditions (3 independent experiments). \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , unpaired student t-test.

## Supplemental Figure 6



**Supplemental Figure 6, related to Figure 6. ADAR1 suppresses ISG activation during interferon stimulation.** **A-B.** Jurkat cells were stimulated with IFN $\alpha$ , IFN $\beta$ , or IFN $\gamma$  and the gene expression of ADAR1 (A) and PRK (B) was determined ( $n = 2$  independent experiments). **C.** Western blot showing levels of dsRNA sensor (PRK, MDA5, DDX58) and apoptosis markers (PARP1 and cleaved PARP1) in cells stimulated with IFN $\beta$  (1 ng/mL).  $\beta$ -actin is used as loading control. **D.** Western blot of total eIF2 $\alpha$  and phosphorylated eIF2 $\alpha$  in wildtype, ADAR1 KO, ADAR1<sup>WT</sup>, and ADAR1<sup>E912A</sup> Jurkat cells. Cells were stimulated with IFN $\beta$  (1 ng/mL) for 24 hours. **E.** PKR expression after shRNA knockdown ( $n = 3$  experiment). **F.** Effect of PKR knockdown on cell viability after IFN $\beta$  (1 ng/mL, 24 hours) stimulation ( $n = 4$  experiments). Error bars represent mean with SEM. \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , \*\*\*\* $p < 0.0001$ , unpaired student t-test.

## Supplemental Figure 7



**Supplemental Figure 7, related to Figure 6. Increased nuclear dsRNA upon IFN $\beta$  stimulation. A and C.** Immunofluorescent staining to detect the localization of dsRNA in CUTTL1 (A) and 293T (C) cells stimulated with IFN $\beta$  (1ng/mL, 24 hrs). Scale bars represent 5 $\mu$ m. **B and D.** Quantification of total dsRNA dots and percentage of nuclear dsRNA from CUTTL1 (B) and 293T (D) cells shown in panel C (n = 10 cells/condition). Error bars represent mean with SEM. \*p<0.05, \*\*p<0.01, unpaired student t-test.



**Table S1. Cell surface markers used for FACS sorting, related to STAR Methods.**

<b>Name</b>	<b>Phenotype</b>
Lin-	CD8- CD56- CD4- CD3- CD19- CD2- CD14-
Stem	hCD45+ Lin- CD34+ CD38-
Progenitor	hCD45+ Lin- CD34+ CD38+

**Table S2. Patient characteristics, related to STAR Methods.**

Sample ID	Diagnosis	Collection Timepoint	Sample Type	Age (years)	Karyotype at diagnosis	cytogenetic abnormalities at diagnosis	Experiments
070	T-Lymphoblastic Lymphoma	Diagnostic	Bone Marrow	13	46,XY[20]	No abnormalities detected	RNA-seq, PDX, ATO
076	T-Lymphoblastic Lymphoma	Diagnostic	Blood	17	46,XY,add(9)(p13),t(11;14)(p13;q11.2)[11]/46,XY[9]	CDKN1a (9p21) deletion	RNA-seq, PDX, ATO
081	T-Lymphoblastic Lymphoma	Diagnostic	Blood	18 month	t(5;14)	TLX3-BCL11B fusion or NKX2-5-BCL11B fusion, CDKN1a (9p21) deletion	PDX, ATO

**Table S6. Primers for RT-qPCR used in this study, related to STAR Methods**

<b>RT-qCPR Primers</b>	<b>Sequence (5' to 3')</b>
ADAR1 fw (total ADAR1)	TGC TGC TGA ATT CAA GTT GG
ADAR1 rev (total ADAR1)	TCG TTC TCC CCA ATC AAG AC
ADAR1 fw (ADAR1p150)	AAC GAA AGC GAA ATT GAA CC
ADAR1 rev (ADAR1p150)	GGG TGT AGT ATC CGC TGA GG
ADAR1 fw (ADAR1p110)	GAC TGA AGG TAG AGA AGG CTA CG
ADAR1 rev (ADAR1p110)	TGC ACT TCC TCG GGA CAC
PKR fw	TGG AAA GCG AAC AAG GAG TAAG
PKR rev	CCA TCC CGT AGG TCT GTG AA
MDA5 fw	GCC TAA TTT ACA GCA ACC ATG A
MDA5 rev	TCA TCA ATG GAT AAC TCC CAT GT
ISG15 fw	GGC TGG GAG CTG ACG GTG AAG
ISG15 rev	GCT CCG CCC GCC AGG CTC TGT
IFIT1 fw	AGG CAA TTA GCC CAG AAG GT
IFIT1 rev	CTC CTC TCT CGG GAA GGT CT
IRF1 fw	CAG CCC AAG AAA GGT CCT C
IRF1 rev	TTG AAC GGT ACA GAC AGA GCA
IRF7 fw	AGC TGT GCT GGC GAG AAG
IRF7 rev	CAT GTG TGT GTG CCA GGA A
IRF9 fw	AGC CTG GAC AGC AAC TCA G
IRF9 rev	GAA ACT GCC CAC TCT CCA C
CD34 fw	TCT GGA TCA AAG TAG GCAG GA
CD34 rev	GAT CCAG CCTC AGA GGA AGA
CD44 fw	ATG AGC ACT AGT GCT ACA GC
CD44 rev	AGACGTACCAGCCATTTGTGT
LMO2 fw	AGG CAA TTA GCC CAG AAG GT
LMO2 rev	CTC CTC TCT CGG GAA GGT CT
hHPRT fw	TCA GGG ATT TGA ATC ATG TTT GTG
hHPRT rev	CGA TGT CAA TAG GAC TCC AGA TG