- **1** Supplementary Information
- 2 Supplemental Figures and Legends



Figure S1. Pull-down assays confirmed a direct protein-protein interaction between
FKBP5 and SEC22B. Purified FLAG-FKBP5-HIS was immobilized on anti-FLAGconjugated magnetic beads and incubated with purified SEC22B-HIS. After washing, bound
proteins were eluted first in sodium dodecyl sulfate–polyacrylamide gel electrophoresis (SDSPAGE) loading buffer at room temperature, and then at 95 °C, also in SDS-PAGE loading
buffer (replicated in 3 independent *in vitro* experiments).





Figure S2. SKA1, SKA2 and SKA3 co-immunoprecipitation (co-IP) experiments (SNAP29-IP) in tissue homogenates from the mouse hippocampus. Co-IP experiments did not reveal any associations of SKA2's partner proteins SKA1 and SKA3 of the SKA complex with SNAP29 (n=3 mice).





Figure S3. VPS34 inhibitor (VPS34i; SAR405) and ULK1 inhibitor (ULK1i; MRT68921) 18 attenuate LLOMe-induced secretory autophagy and autophagic flux in SIM-A9 cells. (A) 19 Cathepsin D secretion (FC relative to veh) after a 4-hour treatment of SIM-A9 cells with 20 LLOMe (0.25 mM) alone to induce secretory autophagy or with VPS34i at 1 and 10 µM 21 (PIK3C3/Vps34 inhibition) in combination with 0.25 mM LLOMe (one-way ANOVA, F<sub>3,8</sub> 22 = 11.74, p = 0.0027; Dunnett's multiple comparisons post hoc test: vehicle vs LLOMe, p =23

24	0.002, vehicle vs LLOMe-VPS34i-1 $\mu$ M p = 0.0036, vehicle vs LLOMe-VPS34i-10 $\mu$ M p =
25	0.0068; n = mean derived from 3 independent <i>in vitro</i> experiments). (B) Cathepsin D secretion
26	(FC relative to veh) after a 4-hour treatment of SIM-A9 cells with LLOMe (0.25 mM) alone to
27	induce secretory autophagy or with ULK1i at 1 and 10 $\mu M$ in combination with 0.25 mM
28	LLOMe (one-way ANOVA, $F_{3, 8} = 3.665$ , $p = 0.0630$ ; Dunnett's multiple comparisons post
29	hoc test: vehicle vs LLOMe, $p = 0.0483$ , vehicle vs LLOMe- ULK1i-1µM $p = 0.3198$ , vehicle
30	vs LLOMe- ULK1i-10 $\mu$ M p = 0.9940; n = mean derived from 3 independent <i>in vitro</i>
31	experiments). (C) LC3B-II/ GAPDH levels determined in autophagic flux assays using
32	bafilomycin A1 (BafA1, 0.1 $\mu$ M) in SIM-A9 cells subjected to increasing concentrations (0.1,
33	1 and 10 $\mu$ M) of VPS34i (one-way ANOVA, F <sub>7, 16</sub> = 16.69, p < 0.0001; Tukey's multiple
34	comparisons post hoc test: vehicle vs BafA1, $p < 0.0001$ , VPS34i-0.1 $\mu$ M vs VPS34i-0.1 $\mu$ M-
35	BafA1 $p = 0.0100$ ; $n =$ mean derived from 3 independent <i>in vitro</i> experiments). (D) LC3B-II/
36	GAPDH levels determined in autophagic flux assays using bafilomycin A1 (0.1 $\mu$ M) in SIM-
37	A9 cells subjected to increasing concentrations (0.1, 1 and 10 $\mu M)$ of ULK1i (one-way
38	ANOVA, $F_{7, 16} = 8.408$ , p = 0.0002; Tukey's multiple comparisons post hoc test: vehicle vs
39	BafA1, p = 0.0160, ULK1i -0.1 $\mu$ M vs ULK1i -0.1 $\mu$ M-BafA1, p = 0.0051, ULK1i -1 $\mu$ M vs
40	ULK1i-1 $\mu$ M-BafA1, p = 0.0530; n = mean derived from 3 independent <i>in vitro</i> experiments).
41	Data are presented as mean + SEM. $* = p < 0.05$ ; $** = p < 0.01$ ; $*** = p < 0.001$ . Source data
42	are provided as a Source Data file.



Figure S4. Viral-mediated knockdown of Ska2 (Ska2-shRNA-1-AAV) in mouse the 44 45 hippocampus. (A) Viral-mediated knockdown of Ska2 (Ska2-shRNA-1-AAV) in the hippocampus leads to significantly decreased SKA2 expression (unpaired, two tailed t-test: t<sub>6</sub> 46 = 3.13, p = 0.0230; n = 4 mice per group). (B) Ska2 mRNA expression is significantly 47 decreased following transfection with Ska2-shRNA-1 or Ska2-shRNA-2 in Neuro2a cells (one-48 way analysis of variance (ANOVA),  $F_{2,29} = 19.60$ , p < 0.0001; Tukey's multiple comparisons 49 post hoc test: scr-shRNA vs Ska2-shRNA-1, p < 0.0001, scr-shRNA vs Ska2-shRNA-2, p = 50 0.0002; scr-shRNA: n = 9, Ska2-shRNA-1: n = 11, Ska2-shRNA-2: n = 12). (C) Viral-51 mediated knockdown of Ska2 (Ska2-shRNA-2-AAV) leads to increased IBA1 expression 4 52 53 weeks after viral injection (paired t-test:  $t_2 = 5.981$ , p = 0.0268; n = 3 mice. Data are presented as mean + SEM. \* = p < 0.05; \*\*\* = p < 0.001; \*\*\*\* = p < 0.0001. Scale bar represents 250 54 µm. Source data are provided as a Source Data file. 55



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Figure S5. Correlation analysis of technical and biological variables with principal 58 components. Depicted are technical and biological variables in the data, with the color, shape, 59 and direction of the ovals in the top right side and numerical values on the bottom left side 60 indicating the correlation value. The top 5 principal components explain >90% of the variance 61 in the gene expression data. The total RNA concentration denoted by rnaconc and totalng is 62 the amount of RNA that is used as input for RNAseq. Libbatch represents the library batches. 63 Num condition denotes the different conditions of the Ska2 knockdown at 2 and 4 weeks, 64 including the controls. Prop [celltypes] are the proportions of cell types obtained from the 65 deconvolution analysis. PC1 is highly correlated with the groups and proportion cell types; this 66 indicated the strong effect of Ska2 knockdown. Additionally, libbatch is correlated with PC3 67 until PC5 and therefore added as a covariate in the differential expression analysis. The 68

- 69 proportion microglia is strongly correlated with other cell types and is therefore representative
- 70 for cell type composition.
- 71
- 72



Figure S6. Gene ontology (GO) enrichment analysis with differentially expressed genes
at 4 weeks. Enriched GO terms. Circle size is proportional to the number of genes. BP,
biological process, CC, cellular component. Source data are provided as a Source Data file.



Figure S7. PheWAS plots of phenotypes associated with the *FKBP5* (A) and *SKA2* (B) gene. The x axes represent phenotypes, and the y axes represent the  $-\log_{10}$  of uncorrected p values. The dashed lines indicate the experiment-wide threshold to survive Bonferroni correction (*FKBP5*:  $p_{-Log10} < 4.016$  and *SKA2*:  $p_{-Log10} < 3.876$ ). Each dot represents one phenotype, and the colors indicate their according traits. Representative top findings are annotated in the figure.

S1









