

Supplementary Information

Synthesis and *in vitro* metabolic stability of sterically shielded antimycobacterial phenylalanine amides

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2. Methods & Materials

General

The purchased starting materials were used as received without further purification. Solvents used for either synthetic or purification purposes were distilled and stored over 4 Å-molecular sieves. Glassware was oven-dried at 110 °C prior to use. For the determination of R_f values and other analytical purposes such as qualitative chromatography Merck TLC silica gel 60 on aluminium sheets with fluorescent indicator F254 were used. Flash chromatography was performed with a puriFlash® 430 instrument (Interchim, Montluçon, France). Columns were packed in either 8 g ($v=10$ mL/min), 45 g ($v=30$ mL/min) or 90 g ($v=40$ mL/min) cartridges with 40 - 63 μ m normal phase silica gel produced by Carl Roth. Column loading was performed with the dry load method. NMR spectra were recorded on an Agilent Technologies VNMRS 400 MHz spectrometer. Chemical shifts are reported relative to the residual solvent signal (CDCl_3 : $\delta_{\text{H}} = 7.26$ ppm; $\delta_{\text{C}} = 77.36$ ppm; CD_3OD $\delta_{\text{H}} = 3.31$ ppm). ^{13}C NMR spectral data were determined as attached-proton-test spectra (APT) and plain ^{13}C spectra. Spectra have been cut, baseline and phase corrected and analyzed utilizing MestreNova 11.0 software (Mestrelab Research, S.L., Spain). APCI-MS (atmospheric pressure chemical ionization) was performed using an expression CMS mass spectrometer (Advion Inc., Ithaca, NY, USA), with both ASAP (atmospheric solids analysis probe) sampling and with the help of the Plate Express TLC-plate extractor. ESI measurements have been conducted on the same expression CMS mass spectrometer with an ESI ionization module and direct injection sampling. HRMS was carried out using a LTQ Orbitrap XL mass spectrometer (Thermo Fisher Scientific Inc., Waltham, MA, USA).

X-ray crystallography

Crystals of **2** suitable for single-crystal X-ray diffraction were obtained by slow evaporation from a solution in ethanol. The X-ray intensity data collection was carried out at 100 K on a Bruker AXS D8 Venture diffractometer, using Mo- K_{α} radiation from an Incoatec $1\mu\text{S}$ Diamond microfocus X-ray source with Montel multilayer optics. A crystal was mounted on a MiTeGen cryo loop using perfluoropolyether PFO-XR75. The programs APEX4 [35] and SAINT [36] were used to control the data collection and to process the raw diffraction data. Scaling and an absorption corrections based on indexed crystal faces was performed with APEX5 [37]. The crystal structure was solved with SHELXT [38] and initially refined with SHELXL-2019/3 [39]. The final structure refinement was carried out by Hirshfeld atom refinement with aspherical scattering factors using NoSpherA2 [30, 31] partitioning in Olex2 1.5 [40] based on electron density from iterative single determinant SCF single point DFT calculations using ORCA v.4.1.1 [41] with a B3LYP functional [42, 43] and a def2-TZVPP basis set. Positions and U_{iso} values of all hydrogen atoms were refined freely. The R configuration at the α -carbon atoms was inferred from the known absolute configuration of the starting material and the absolute structure was confirmed by a Flack x parameter [44] and a Hooft y parameter [45] close to zero (**Table S1**). Structure pictures were drawn with Mercury [46]. Crystal data and refinement details are given in **Table S1**. **Figure S1** shows a displacement ellipsoid plot and **Table S2** lists the corresponding hydrogen bond parameters. CCDC 2293688 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre and Fachinformationszentrum Karlsruhe via www.ccdc.cam.ac.uk/structures.

Table S1 Crystal data and refinement details for **2**.

Empirical formula	C ₂₆ H ₂₆ FN ₃ O ₄ S
<i>M</i> _r	495.578
<i>T</i> (K)	100(2)
λ (Å)	0.71073
Crystal system, space group	Triclinic, <i>P</i> 1
<i>a</i> (Å)	9.5510(8)
<i>b</i> (Å)	10.4458(9)
<i>c</i> (Å)	12.6068(10)
α (°)	73.785(4)
β (°)	78.752(4)
γ (°)	85.654(4)
<i>V</i> (Å ³)	1184.22(17)
<i>Z</i> , ρ_{calc} (mg m ⁻³)	2,
μ (mm ⁻¹)	0.184
<i>F</i> (000)	520.570
Crystal size (mm)	0.197 × 0.165 × 0.118
θ range (°)	2.03 - 36.38
Reflections collected / unique	551394
<i>R</i> _{int}	0.062
Completeness to θ_{full} (%)	99.67
Data / restraints / parameters	23006 / 3 / 839
Observed data [<i>I</i> > 2 σ (<i>I</i>)]	21992
Goodness-of-fit on <i>F</i> ²	1.2264
<i>R</i> 1 [<i>I</i> > 2 σ (<i>I</i>)]	0.0179
<i>wR</i> 2 (all data)	0.0374
Flack <i>x</i> parameter	−0.002(6) ^a
Hooft <i>y</i> parameter	−0.009(6) ^b
$\Delta\rho_{\text{max}}$, $\Delta\rho_{\text{min}}$ (e Å ⁻³)	0.24, −0.11
CCDC No.	2293688

^a Calculated with Olex2.^b Calculated with PLATON [47].

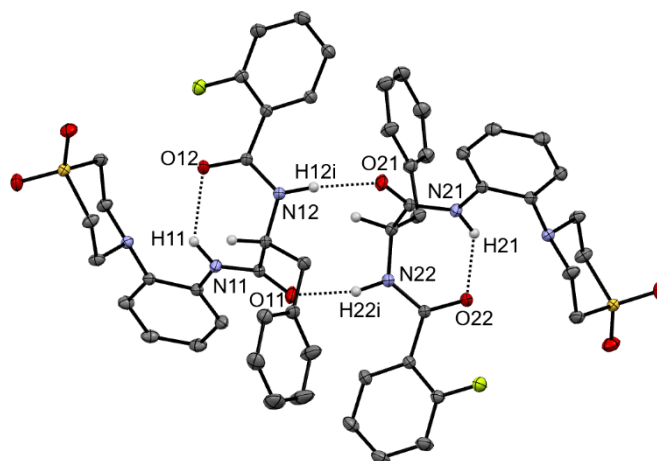


Figure S1 Asymmetric unit of **2**. Displacement ellipsoids are drawn at the 50 % probability level. Nitrogen-bound hydrogen atoms and the carbon-bound hydrogen atoms attached to centres of chirality are represented by small spheres of arbitrary radius, otherwise omitted for clarity. Colour scheme: C, grey; H, white; N, blue; O, red; F, light green; S, yellow.

Table S2 Hydrogen bond parameters in **2**.

<i>D</i> –H··· <i>A</i>	<i>d</i> (<i>D</i> –H) (Å)	<i>d</i> (H··· <i>A</i>) (Å)	<i>d</i> (<i>D</i> ··· <i>A</i>) (Å)	∠(<i>DHA</i>) (°)
N11 H11 O12	1.006(7)	2.287(7)	3.0836(6)	135.2(6)
N12 H12i O21	0.988(7)	1.913(7)	2.8929(5)	170.9(6)
N21 H21 O22	0.991(8)	2.134(8)	2.9709(6)	141.1(6)
N22 H22i O11	0.984(8)	1.843(8)	2.7998(5)	163.3(6)

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Plasma stability

Plasma stability in human and murine plasma was determined at five different time points over 120 min using HPLC-MS/MS analysis. Pooled human plasma, anti-coagulated with trisodium citrate, and non-sterile murine plasma, anti-coagulated with Li-heparin (Bienta/Enamine Ltd., batch #M190406), were used. The human plasma was used by a third-party lab and the authors were not involved in the extraction and preparation of the samples. The source of the samples was the Kyiv city blood center, Kiev, Ukraine. The plasma stability is given as the percentage of substance remaining in plasma over time. All measurements were performed using the Shimadzu Prominence HPLC system including vacuum degasser, gradient pumps, reverse phase column (ZORBAX Extend-C18 column, 2.1x50 mm, 5 μ m), column oven and autosampler. The HPLC system was coupled with tandem mass API 3000 (AB Sciex). Both positive and negative ion modes of the TurbolonSpray ion source were used. Acquisition and analysis of the data were performed using Analyst 1.6.3 software (PE Sciex).

Incubations of every compound + references (Verapamil and Propantheline) were carried out in 5 aliquots of 60 μ L each (one for each time point), in duplicates. Plasma was spiked with test compounds from a 10 mM DMSO stock solution to yield a test concentration of 1 μ M, final DMSO concentration 1%. The aliquots were incubated at 37 $^{\circ}$ C with shaking at 100 rpm. Five time points over 120 min have been analyzed. The reactions were stopped by adding 300 μ L of methanol containing internal standard with subsequent plasma proteins sedimentation by centrifuging at 6000 rpm for 4 minutes. Supernatants were analyzed by the HPLC system that was coupled with a tandem mass spectrometer. The percentage of the test compounds remaining after incubation were calculated.

Microsomal stability

Microsomal stability in human and murine microsomal suspensions was determined at five different time points over 40 min using HPLC-MS/MS analytic. The microsomes were purchased commercially from XenoTech: human liver microsomes (pooled, mixed gender, H0630/lot N#1210097) and mouse liver microsomes (pooled, male BALB/c, M3000/lot #2010026). All measurements were performed using the Shimadzu Prominence HPLC system including vacuum degasser, gradient pumps, reverse phase column (Phenomenex Luna[®]-C18 column, 2.1x50 mm, 5 μ m), column oven and autosampler. The HPLC system was coupled with tandem mass API 3000 (AB Sciex). Both positive and negative ion

modes of the TurbolonSpray ion source were used. Acquisition and analysis of the data were performed using Analyst 1.6.3 software (PE Sciex).

Incubations of every compound + references (Diclofenac and Propranolol for human microsomes, Propranolol and Imipramin for murine microsomes) were carried out in 5 aliquots of 30 μL each (one for each time point), in duplicates. Liver microsomal incubation medium contained 0.42 mg of liver microsomal protein as well as phosphate buffer (100 mM, pH 7.4), MgCl_2 (3.3 mM), NADPH (3 mM), glucose-6-phosphate (5.3 mM) and glucose-6-phosphate dehydrogenase (0.67 units/ml). In the control reactions, the NADPH-cofactor system was substituted with phosphate buffer. Microsomal suspensions were spiked with test compounds from a 10 mM DMSO stock solution to yield a test concentration of 2 μM , final DMSO concentration 1.6 %. The aliquots were incubated at 37 $^\circ\text{C}$ with shaking at 100 rpm. Five time points over 40 min have been analyzed. The reactions were stopped by adding 5 volumes of 90 % acetonitrile containing internal standard with subsequent protein sedimentation by centrifuging at 5500 rpm for 5 minutes. Supernatants were analyzed by the HPLC system that was coupled with a tandem mass spectrometer. The percentage of the test compounds remaining after incubation were calculated. The elimination constants (k_{el}) and half-lives ($t_{1/2}$) of the compounds were determined in a plot of $\ln(\text{AUC})$ versus time, using linear regression analysis:

$$k_{el} = -slope$$

$$t_{1/2} = \frac{0.693}{k_{el}}$$

HPLC

Analytical HPLC

All described final compounds described were confirmed to be of >95 % purity. The used Shimadzu HPLC system (Shimadzu, Kyoto, Japan) consists of a CBM-40 control unit, a DGU-403 degassing unit, two LC-40D chromatography pumps, a SIL-40C autosampler unit, a CTO-40C column oven and an SPD-M40 PDA UV detector. The standard method for purity determination utilized an Agilent Poroshell 120, EC-C18, 3,0x50mm, 2,7 μm analytical column at a flow rate of 1.2 mL/min at room temperature. The 6 min gradient started at 15 % and increased to 85 % acetonitrile in water and was used in all cases. All solvents used are HPLC grade purity. UV absorbance at 254 nm was measured and the purity was derived from the integrated intensity signal.

Preparative HPLC

If necessary, final compounds were purified by preparative HPLC. The Shimadzu HPLC system (Shimadzu, Kyoto, Japan) used consists of a CBM-40 control unit, two LC-20AP chromatography pumps, a SIL-40C autosampler unit, a CTO-40C column oven, an SPD-M40 PDA UV detector and an FRC-10A fraction collector. A preparative column manufactured by Agilent (Polaris C18-A, 5 μm , 250 X 21.2mm) was used at a flow rate of 15 mL/min with a standard gradient of 50 % to 95% acetonitrile in water over 20 min at room temperature.

Microbiological assays

MIC determination against *M. intracellulare* ATCC 35761 pTEC27 and *M. abscessus* ATCC 19977 pTEC27

M. intracellulare ATCC 35761 pTEC27 and *M. abscessus* ATCC 19977 pTEC27 expressing tomato RFP were used for the activity assays. Cryo-stocks of the bacteria grown in 7H9 broth supplemented with 10 % ADS and 0.05 % Tween 80 were stored with approximately 15 % glycerol at -80°C. Using an inoculation loop, bacteria were streaked on 7H10 plates supplemented with 0.5 % glycerol, 10 % ADS containing and 400 µg/mL hygromycin and grown for 5 days in an incubator at 37°C. A single colony was picked from the 7H10 plates and the liquid culture was grown in 7H9 broth supplemented with 10 % ADS, 0.05 % Tween 80 and 400 µg/mL hygromycin. The culture volume was 10 mL in a 50 mL Falcon tube. The tubes were shaken in an incubator at 37°C.

MIC values were determined by the broth microdilution method. 96-well flat bottom tissue culture plates (Sarstedt, 83.3924.500) were used³¹. In the third well of each row two times the desired highest concentration of each compound was added in 7H9 medium supplemented with 10 % ADS and 0.05 % Tween 80. Each compound was diluted twofold in a nine-point serial dilution in 50 µL medium. (All drugs were prepared as 10 mM stocks in dimethyl sulfoxide (DMSO).)

The starting inoculum was diluted from a liquid culture described above at the mid-log phase (OD₆₀₀ 0.3 to 0.7) and an OD₆₀₀ of 0.1 was correlated to 1 x 10⁸ CFU/mL. A starting inoculum of 5 x 10⁵ cells/mL was used and 50 µL were added to each well. The plates were sealed with parafilm, placed in a container with moist tissue and incubated for three days at 37 °C (*M. abscessus*) or five days (*M. intracellulare*). After incubation the plates were monitored by OD measurement at 550 nm (BMG labtech Fluostar Optima) and by measurement of fluorescence (λ_{ex} = 544 nm λ_{em} = 590 nm). The assay was performed in duplicate, and the results were averaged.

Data analysis:

Every assay plate contained eight wells with dimethyl sulfoxide (1 %) as negative control, which corresponds to 100 % bacterial growth and eight wells with amikacin (100 µM) as positive control in which 100 % inhibition of bacterial growth was reached. Controls were used to monitor the assay quality through determination of the Z' score. The Z' factor was calculated as follows:

$$Z' = 1 - \frac{3(SD_{\text{amikacin}} + SD_{\text{DMSO}})}{M_{\text{amikacin}} - M_{\text{DMSO}}}$$

(SD = standard deviation, M = mean)

The percentage of growth inhibition was calculated by the equation:

$$\% \text{ growth inhibition} = 100 \% \times \frac{\text{signal(DMSO)} - \text{signal(sample)}}{\text{signal(DMSO)} - \text{signal(amikacin)}}$$

MIC determination against *M. avium ssp. hominissuis strain 109 (MAC109)*

MIC value determination by optical density at 600 nm [OD₆₀₀] was carried out in 96-well plate format. 96-well plates were initially set up with 100 µl of 7H9 per well. For each compound, a 10-point twofold dilution series starting at twice the desired highest concentration was dispensed onto the 96-well plates using a Tecan D300e Digital Dispenser, with the DMSO concentration normalized to 2 % v/v. A bacteria culture grown to mid-log-phase (OD₆₀₀, 0.4 to 0.6) was diluted to OD₆₀₀ = 0.1 (1*10⁷ CFU/mL). 100 µl of the resulting bacteria suspension was dispensed onto the 96-well plates containing the sample compounds to give a final volume of 200 µl per well with an initial OD₆₀₀ = 0.05 (5*10⁶ CFU/mL) and a final DMSO concentration of 1 % v/v. Final compound concentration ranges were typically 50 to 0.098 µM or 6.25 to 0.012 µM. Untreated control wells, which contained bacteria suspension and 1 % v/v DMSO, were included on each plate. Plates were sealed with parafilm, stored in boxes with wet paper towels, and incubated at 37°C with shaking (110 rpm) and were incubated for 5 days.

To determine growth, OD₆₀₀ was measured using a Tecan Infinite M200 plate reader on day 0 and day 3. Two biological replicates were performed. Clarithromycin was included in each experiment as a positive control. For each well on the 96-well plate, bacterial growth was calculated by subtracting the day 0 OD₆₀₀ value from the day 3 OD₆₀₀ value. For each compound series, the bacterial growth values for the untreated control wells were averaged to give the average drug-free bacterial growth. For compound-containing wells, percentage growth was calculated by dividing their growth values by the average drug-free bacterial growth for the compound series and multiplying by 100. For each compound series, we plotted percentage growth versus compound concentration. By visual inspection of the dose-response curve, we determined the MIC of a compound as the compound concentrations that would result in 90 % growth inhibition. The MIC determination was performed two times with different starter cultures. The MIC values shown in the script are the averaged results of biological duplicates.

MIC determination against *M. tuberculosis H37Rv*

MICs against Mtb H47Rv were determined as described in [48]: Yang, X.; Wedajo, W.; Yamada, Y.; Dahlroth, S. L.; Neo, J. J. L.; Dick, T.; Chui, W. K. 1,3,5-Triazaspiro[5.5]Undeca-2,4-Dienes as Selective Mycobacterium Tuberculosis Dihydrofolate Reductase Inhibitors with Potent Whole Cell Activity. *Eur J Med Chem* **2018**, *144*, 262–276. <https://doi.org/10.1016/J.EJMECH.2017.12.017>. Briefly, compounds were serially diluted in flat-bottom 96-well plates, and a mid-log-phase culture was mixed with the compound-containing broth (final OD₆₀₀ = 0.05). Plates were sealed with Breathe-Easy sealing membrane (Sigma), placed in humidified plastic boxes and incubated at 37°C for 7 days, shaking at 80 rpm. Growth was monitored by measuring turbidity at 600 nm using a Tecan Infinite 200 Pro microplate reader (Tecan). MIC₉₀ values were deduced from the generated dose–response curves. The MIC values shown in the script are the averaged results of the two biological replicates.

Cytotoxicity against immortalized human kidney epithelial

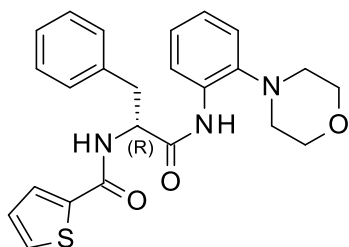
For evaluation of possible cytotoxic effects on mammalian cells, 20.000 HEK293 cells (immortalized human kidney epithelial cells; DSMZ, Braunschweig, Germany) were seeded out per well of a 96-well plate (Sarstedt, Germany) in DMEM supplemented with 10% FCS (both Gibco) and cultured in a humidified incubator at 37°C and 5% CO₂. After 4 hours and attaching cells to the bottom, all compounds in DMSO were added to a final concentration of 50 µM. A sample containing only 0.5%

DMSO was used as control. Cells were incubated for 24 h under the above-described conditions before AlamarBlue reagent (Thermo Fisher Scientific, Germany) was added. Samples were incubated again for 4 h before measurement was performed. The plates were analyzed in a microtiter plate reader (BMG, Germany) and the forming resorufin was quantified in fluorescence channel with Ex:550/Em:590 nm. The data are means of triplicates with standard deviation.

3. Structures and synthetic protocols

MMV

(2R)-N-[2-(morpholin-4-yl)phenyl]-3-phenyl-2-[(thiophen-2-yl)formamido]propanamide



Synthesis & Purification:

Described as compound **2B-(R)** in [18]: Lang, M., Ganapathy, U. S., Mann, L., Abdelaziz, R., Seidel, R. W., Goddard, R., Sequenzia, I., Hoenke, S., Schulze, P., Aragaw, W. W., Csuk, R., Dick, T., & Richter, A. (2023). Synthesis and Characterization of Phenylalanine Amides Active against Mycobacterium abscessus and Other Mycobacteria. *Journal of Medicinal Chemistry*. <https://doi.org/10.1021/ACS.JMEDCHEM.3C00009>

Analyses:

R_f value: 0.37 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 1**) (400 MHz, Chloroform-*d*) δ 8.83 (s, 1H), 8.40 (dd, $J = 8.0, 1.5$ Hz, 1H), 7.52 (dd, $J = 3.7, 1.2$ Hz, 1H), 7.47 (dd, $J = 5.0, 1.1$ Hz, 1H), 7.32 – 7.25 (m, 4H), 7.23 – 7.17 (m, 1H), 7.16 – 7.05 (m, 3H), 7.03 (dd, $J = 5.0, 3.7$ Hz, 1H), 6.95 (d, $J = 7.8$ Hz, 1H), 4.95 (td, $J = 8.0, 5.6$ Hz, 1H), 3.68 – 3.53 (m, 4H), 3.39 (dd, $J = 13.6, 5.6$ Hz, 1H), 3.25 (dd, $J = 13.6, 8.1$ Hz, 1H), 2.69 – 2.50 (m, 4H).

^{13}C NMR APT (**Spectrum 2**) (101 MHz, Chloroform-*d*) δ 168.77, 161.56, 141.19, 137.97, 136.50, 132.91, 130.75, 129.38, 128.86, 128.66, 127.81, 127.12, 125.69, 124.33, 120.80, 119.52, 67.25, 56.19, 52.46, 38.47.

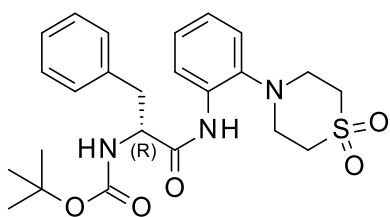
Mass: Calculated m/z for $\text{C}_{24}\text{H}_{26}\text{N}_3\text{O}_3\text{S}^+ [\text{M}+\text{H}]^+ = 436.1689$; $\text{C}_{24}\text{H}_{25}\text{N}_3\text{O}_3\text{SNa}^+ [\text{M}+\text{Na}]^+ = 458.1509$; found APCI: 436.1; found HRMS: $[\text{M}+\text{H}]^+ 436.1691$; $[\text{M}+\text{Na}]^+ 458.1506$

HPLC purity: 98.9 % $t_R = 4.526$ min (**HPLC-Trace 1**); (50 mm Eclipse Plus C18 1.8 μm , 4.6 mm, acetonitrile/ water 45:55, $v = 1.0$ mL/min, $\lambda = 220$ nm)

Specific optical rotation: Boc-protected intermediate (**2-D**) $[\alpha]^{22.7}_D 46.01$ (c 0.119, MeOH); final compound (**3-D**) $[\alpha]^{22.7}_D 56.74$ (c 0.1375, MeOH)

1

tert-butyl N-[(1R)-1-benzyl-2-[2-(1,1-dioxo-1,4-thiazinan-4-yl)anilino]-2-oxo-ethyl]carbamate



Synthesis & Purification:

Described as compound **23** in: Lang, M., Ganapathy, U. S., Mann, L., Abdelaziz, R., Seidel, R. W., Goddard, R., Sequenzia, I., Hoenke, S., Schulze, P., Aragaw, W. W., Csuk, R., Dick, T., & Richter, A. (2023). Synthesis and Characterization of Phenylalanine Amides Active against Mycobacterium abscessus and Other Mycobacteria. *Journal of Medicinal Chemistry*. <https://doi.org/10.1021/ACS.JMEDCHEM.3C00009>

Analyses:

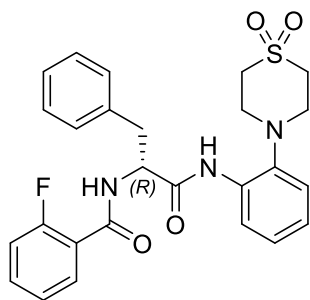
R_f value: 0.31 ethyl acetate/heptane 1:1

^1H NMR (δ) (400 MHz, Chloroform- d) δ 8.89 (s, 1H), 8.38 (d, J = 8.1 Hz, 1H), 7.31 – 7.11 (m, 7H), 7.08 – 7.01 (m, 1H), 5.40 – 5.13 (m, 1H), 4.47 (q, J = 7.3 Hz, 1H), 3.33 – 3.00 (m, 10H), 1.37 (s, 9H).

Mass: Calculated m/z for $\text{C}_{24}\text{H}_{32}\text{N}_3\text{O}_5\text{S}^+$ $[\text{M}+\text{H}]^+ = 474.2057$; found APCI: 474.2

2

(2R)-2-[(2-fluorophenyl)formamido]-N-[2-(1,1-dioxo-1λ⁶-thiomorpholin-4-yl)phenyl]-3-phenylpropanamide



Synthesis & Purification:

Described as compound **24** in: Lang, M., Ganapathy, U. S., Mann, L., Abdelaziz, R., Seidel, R. W., Goddard, R., Sequenzia, I., Hoenke, S., Schulze, P., Aragaw, W. W., Csuk, R., Dick, T., & Richter, A. (2023). Synthesis and Characterization of Phenylalanine Amides Active against Mycobacterium abscessus and Other Mycobacteria. *Journal of Medicinal Chemistry*. <https://doi.org/10.1021/ACS.JMEDCHEM.3C00009>

Analyses:

R_f value: 0.30 ethyl acetate

^1H NMR (**Spectrum 4**) (500 MHz, Chloroform-*d*) δ 9.16 (s, 1H), 8.44 (dd, $J = 8.2, 1.4$ Hz, 1H), 8.08 (td, $J = 7.9, 1.9$ Hz, 1H), 7.52 (dddd, $J = 8.3, 7.2, 5.3, 1.9$ Hz, 1H), 7.36 – 7.22 (m, 7H), 7.20 (ddd, $J = 8.4, 7.5, 1.5$ Hz, 1H), 7.18 – 7.13 (m, 2H), 7.06 (td, $J = 7.7, 1.5$ Hz, 1H), 5.04 (qd, $J = 6.9, 2.2$ Hz, 1H), 3.40 – 3.30 (m, 2H), 3.21 (t, $J = 5.3$ Hz, 4H), 3.12 (t, $J = 5.1$ Hz, 4H).

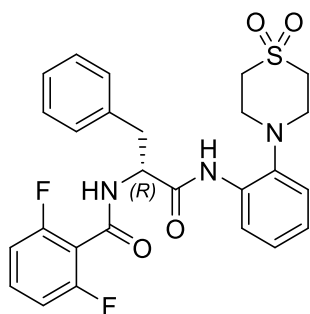
^{13}C NMR (**Spectrum 5**) (126 MHz, Chloroform-*d*) δ 168.70, 163.62 (d, $J = 3.3$ Hz), 160.82 (d, $J = 248.0$ Hz), 139.97, 136.41, 134.36 (d, $J = 9.5$ Hz), 133.12, 131.91 (d, $J = 1.8$ Hz), 129.41, 128.89, 127.19, 126.84, 125.29 (d, $J = 3.2$ Hz), 124.25, 121.51, 120.06, 119.43 (d, $J = 10.8$ Hz), 116.47 (d, $J = 24.7$ Hz), 56.07, 51.92, 51.27, 36.92.

Mass: Calculated m/z for $\text{C}_{26}\text{H}_{27}\text{FN}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 496.1701$; found APCI: 496.6; found HRMS: $[\text{M}+\text{H}]^+ 496.1698$

HPLC purity: 98.86 %; $t_R = 3.0$ min (**HPLC-Trace 2**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

3

(2R)-2-[(2,6-difluorophenyl)formamido]-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)phenyl]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product from **2**: 156 mg (0.42 mmol, 1 eq.); **THF** (10 ml); **2,6-difluorobenzoic acid** (80 mg, 0.50 mmol, 1.2 eq., BLDpharm); **PyBOP** (328 mg, 0.63 mmol, 1.5 eq., Carbolution); **DIPEA** (214 μL , 0.60 mmol, 3 eq., Aldrich)

Procedure: Boc-protected crude product from **2**, **2,6-difluorobenzoic acid** and **PyBOP** were dissolved in **THF**. Subsequently, **DIPEA** was added, and the mixture was stirred at room temperature overnight.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 80 % heptane to 20 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μm particle size. Yield: 194 mg; 90 %

Analyses:

R_F value: 0.30 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 6**) (500 MHz, Chloroform-*d*) δ 9.07 (s, 1H), 8.35 (d, $J = 8.2$ Hz, 1H), 7.40 – 7.29 (m, 5H), 7.27 – 7.22 (m, 1H), 7.21 – 7.14 (m, 2H), 7.08 (td, $J = 7.7, 1.4$ Hz, 1H), 6.91 (tt, $J = 8.5, 2.1$ Hz, 3H), 5.03 (q, $J = 7.4$ Hz, 1H), 3.40 – 3.11 (m, 10H).

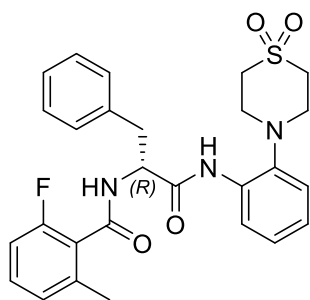
^{13}C NMR (**Spectrum 7**) (126 MHz, Chloroform-*d*) δ 168.61, 160.78, 160.01 (d, $J = 253.2$ Hz), 159.96 (d, $J = 252.7$ Hz), 140.24, 136.32, 132.99, 132.40 (t, $J = 10.2$ Hz), 129.41, 128.79, 127.13, 126.68, 124.44, 121.57, 120.20, 112.93 (d, $J = 18.7$ Hz), 112.25 (d, $J = 4.3$ Hz), 112.09 (d, $J = 4.1$ Hz), 56.10, 51.87, 51.27, 37.08.

Mass: Calculated m/z for $\text{C}_{26}\text{H}_{26}\text{F}_2\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 514.1607$; found APCI: 514.6; found HRMS: $[\text{M}+\text{H}]^+ 514.1602$

HPLC purity: 98.8 %; $t_R = 4.7$ min (**HPLC-Trace 3**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 10 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

4

(2R)-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)phenyl]-2-[(2-fluoro-6-methylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product from **2**: 183 mg (0.49 mmol, 1 eq.); **THF** (10 ml); **2-fluoro-6-methylbenzoic acid** (91 mg, 0.59 mmol, 1.2 eq., BLDpharm); **PyBOP** (382 mg, 0.74 mmol, 1.5 eq., Carbolution); **DIPEA** (250 μL , 1.47 mmol, 3 eq., Aldrich)

Procedure: Boc-protected crude product from **2**, **2-fluoro-6-methylbenzoic acid** and **PyBOP** were dissolved in **THF**. Subsequently, **DIPEA** was added, and the mixture was stirred at room temperature overnight.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 75 % heptane to 25 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μm particle size. Preparative HPLC afterwards. Yield: 203 mg; 81 %

Analyses:

R_f value: 0.31 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 8**) (400 MHz, Chloroform-*d*) δ 9.15 (s, 1H), 8.35 (dd, $J = 8.2, 1.5$ Hz, 1H), 7.32 – 7.13 (m, 8H), 7.07 (td, $J = 7.7, 1.5$ Hz, 1H), 6.93 (d, $J = 7.6$ Hz, 1H), 6.84 (t, $J = 8.9$ Hz, 1H), 6.68 – 6.59 (m, 1H), 5.07 (q, $J = 7.8$ Hz, 1H), 3.35 (dd, $J = 14.2, 6.9$ Hz, 1H), 3.31 – 3.12 (m, 9H), 2.06 (s, 3H).

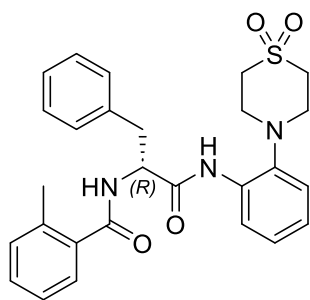
^{13}C NMR (**Spectrum 9**) (101 MHz, Chloroform-*d*) δ 168.88, 165.77, 159.04 (d, $J = 246.0$ Hz), 140.19, 138.37 (d, $J = 2.8$ Hz), 136.52, 133.07, 130.91 (d, $J = 8.9$ Hz), 129.29, 128.82, 127.11, 126.72, 126.34 (d, $J = 2.9$ Hz), 124.40, 123.71 (d, $J = 17.0$ Hz), 121.58, 120.27, 112.90 (d, $J = 21.8$ Hz), 55.66, 51.76, 51.29, 36.82, 18.84.

Mass: Calculated m/z for $\text{C}_{27}\text{H}_{29}\text{FN}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 510.1858$; found APCI: 510.3; found HRMS: $[\text{M}+\text{H}]^+ 510.1852$

HPLC purity: 98.8 %; $t_R = 3.0$ min (**HPLC-Trace 4**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

5

(2R)-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)phenyl]-2-[(2-methylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product from **2**: 42 mg (0.11 mmol, 1 eq.); **THF** (5 ml); **2-methylbenzoic acid** (18 mg, 0.13 mmol, 1.2 eq., TCI); **PyBOP** (88 mg, 0.17 mmol, 1.5 eq., Carbolution); **DIPEA** (44 μL , 0.34 mmol, 3 eq., Aldrich)

Procedure: Boc-protected crude product from **2**, **2-methylbenzoic acid** and **PyBOP** were dissolved in **THF**. Subsequently, **DIPEA** was added, and the mixture was stirred at room temperature overnight.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 75 % heptane to 25 % over 10 CV on 8 g normal phase silica gel 0.63 – 0.2 μm particle size. Afterwards preparative HPLC. Yield: 29 mg; 54 %

Analyses:

R_f value: 0.42 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 10**) (400 MHz, Chloroform-*d*) δ 9.18 (s, 1H), 8.39 (d, $J = 8.6, 1.5$ Hz, 1H), 7.32 (d, $J = 4.3$ Hz, 4H), 7.30 (dd, $J = 7.3, 1.7$ Hz, 1H), 7.28 – 7.23 (m, 1H), 7.22 – 7.17 (m, 4H), 7.14 (td, $J = 7.4,$

1.2 Hz, 1H), 7.11 – 7.05 (m, 1H), 6.39 (d, $J = 7.7$ Hz, 1H), 4.96 (q, $J = 7.5$ Hz, 1H), 3.35 (dd, $J = 14.1, 7.4$ Hz, 1H), 3.31 – 3.14 (m, 9H), 2.30 (s, 3H).

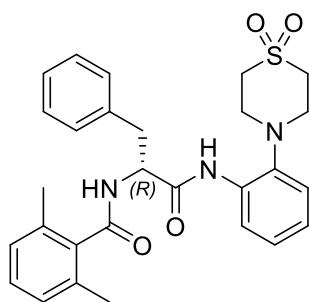
^{13}C NMR (**Spectrum 11**) (101 MHz, Chloroform- d) δ 170.23, 169.04, 140.01, 136.52, 136.43, 134.69, 133.09, 131.33, 130.56, 129.27, 128.88, 127.17, 126.80, 126.63, 125.86, 124.36, 121.53, 120.17, 55.92, 51.86, 51.30, 36.97, 19.79.

Mass: Calculated m/z for $\text{C}_{27}\text{H}_{29}\text{FN}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 492.1952$; $[\text{M}+\text{Na}]^+ = 514.1771$; found APCI: 492.2; found HRMS: $[\text{M}+\text{H}]^+ 492.1956$; $[\text{M}+\text{Na}]^+ 514.1775$

HPLC purity: 97.2 %; $t_R = 3.0$ min (**HPLC-Trace 5**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

6

(2R)-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)phenyl]-2-[(2,6-dimethylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product from **2**: 42 mg (0.11 mmol, 1 eq.); **DMF** (10 ml); **2,6-dimethylbenzoic acid** (21 mg, 0.14 mmol, 1.2 eq., TCI); **PyBOP** (117 mg, 0.23 mmol, 2 eq., Carbolution); **DIPEA** (56 μL , 0.33 mmol, 3 eq., Aldrich)

Procedure: Boc-protected crude product from **2**, **2,6-dimethylbenzoic acid** and **PyBOP** were dissolved in **THF**. Subsequently, **DIPEA** was added, and the mixture was stirred at room temperature for two days.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 80 % heptane to 30 % over 10 CV on 8 g normal phase silica gel 0.63 – 0.2 μm particle size. Afterwards preparative HPLC. Yield: 21 mg; 38 %

Analyses:

R_f value: 0.39 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 12**) (400 MHz, Chloroform- d) δ 9.30 (s, 1H), 8.40 (dd, $J = 8.6, 1.5$ Hz, 1H), 7.35 – 7.28 (m, 4H), 7.28 – 7.19 (m, 3H), 7.13 (t, $J = 7.6$ Hz, 1H), 7.12 – 7.07 (m, 1H), 6.94 (d, $J = 7.6$ Hz, 2H),

6.16 (d, $J = 7.8$ Hz, 1H), 4.99 (ddd, $J = 8.8, 7.8, 6.8$ Hz, 1H), 3.44 – 3.19 (m, 9H), 3.13 (dd, $J = 14.2, 8.8$ Hz, 1H), 2.00 (s, 6H).

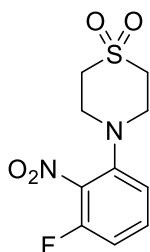
^{13}C NMR (**Spectrum 13**) (101 MHz, Chloroform- d) δ 170.84, 168.91, 139.99, 136.42, 136.22, 134.09, 133.11, 129.23, 129.09, 128.92, 127.51, 127.21, 126.86, 124.34, 121.55, 120.14, 55.55, 51.88, 51.39, 36.60, 18.68.

Mass: Calculated m/z for $\text{C}_{28}\text{H}_{31}\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 506.2108$; found APCI: 506.4; found HRMS: $[\text{M}+\text{H}]^+ 506.2101$

HPLC purity: 99.0 %; $t_R = 3.1$ min (**HPLC-Trace 6**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

7

4-(3-fluoro-2-nitrophenyl)-1 λ^6 -thiomorpholine-1,1-dioxide



Synthesis:

Quantities: **1,3-Difluoro-2-nitrobenzene** (1114 mg, 7 mmol, 1 eq., BLDpharm); **Thiomorpholinedioxide** (946 mg, 7 mmol, 1 eq., BLDpharm); **DIPEA** (2382 μL , 14 mmol, 2 eq., Aldrich)

Procedure: **1,3-Difluoro-2-nitrobenzene** and **thiomorpholinedioxide** were dissolved in **DIPEA** and stirred at 50 $^{\circ}\text{C}$ for 3 days.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/chloroform gradient 40 % heptane to 0 % over 10 CV on 90 g normal phase silica gel 0.63 – 0.2 μm particle size. Yield: 1261 mg; 95 %

Analyses:

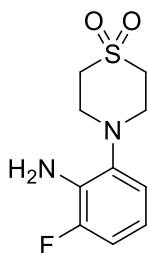
R_f value: 0.76 dichloromethane/ethyl acetate 1:1

^1H NMR (**Spectrum 14**) (400 MHz, Chloroform- d) δ 7.47 (td, $J = 8.4, 6.0$ Hz, 1H), 7.15 – 7.02 (m, 2H), 3.65 – 3.43 (m, 4H), 3.27 – 3.03 (m, 4H).

Mass: Calculated m/z for $\text{C}_{10}\text{H}_{12}\text{FN}_2\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 275.0497$; found APCI: 275.0

8

4-(2-amino-3-fluorophenyl)-1λ⁶-thiomorpholine-1,1-dioxide



Synthesis:

Quantities: **7** (1261 mg, 4.60 mmol, 1 eq.); **H₂** (Balloon); **Pd(OH)₂/C 20 %** (252 mg, 20 m %, Aldrich); **EtOH 96 % v/v** (90 mL)

Procedure: **7** was dissolved in **EtOH**, add catalyst; close the flask with a septum and purge the mixture with hydrogen for 30 min under stirring; add a hydrogen balloon; check for full conversion with TLC; after completion remove the catalyst with paper filter and wash the filter thoroughly with ethyl acetate; evaporate the solvents under reduced pressure

Purification:

Product was used without further purification. Yield: 955 mg; 85 %

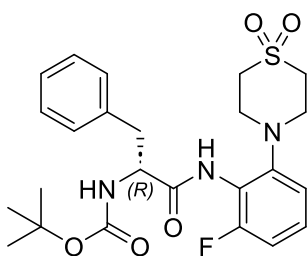
Analyses:

R_F value: 0.25 ethyl acetate/heptane 1:1

Mass: Calculated m/z for C₁₀H₁₄FN₂O₂S⁺ [M+H]⁺ = 245.0755; found APCI: 245.2

9

tert-butyl N-[(1R)-1-[[2-(1,1-dioxo-1λ⁶-thiomorpholin-4-yl)-6-fluorophenyl]carbonyl]-2-phenylethyl]carbamate



Synthesis:

Quantities: **Pyridine** (20 mL); **EtOAc** (40 mL); **8** (955 mg, 3.91 mmol, 1 eq.); **N-Boc-(R)-phenylalanine** (1141 mg, 4.30 mmol, 1.1 eq., TCI); **T3P 50 % m/V in EtOAc** (4655 μL, 7.82 mmol, 2 eq., Aldrich)

Procedure: **8** and **N-Boc-(R)-phenylalanine** were dissolved in a 1:2 mixture of distilled **pyridine** and **EtOAc**. The mixture was cooled to -20 °C with an isopropanol dry-ice bath before **T3P 50 % m/V in EtOAc** was added. After the addition, the cooling bath was removed, and the mixture was stirred for

20 h at room temperature. Afterwards the mixture was washed with 0.25 M KH_2PO_4 solution three times before the organic phase was evaporated on silica gel under reduced pressure for purification.

Purification:

Flash chromatography heptane/methyl tert-butyl ether gradient 50 % heptane to 0 % over 10 CV on 90 g normal phase silica gel 0.63 – 0.2 μm particle size. Yield: 1769 mg; 92 %

Analyses:

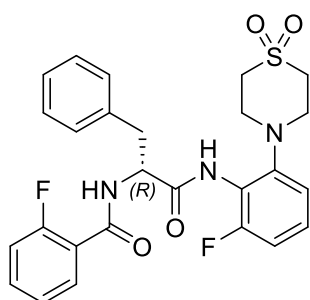
R_f value: 0.42 heptane/ethyl acetate 1:1

^1H NMR (**Spectrum 15****Spectrum 14**) (400 MHz, cdCl_3) δ 7.78 (s, 1H), 7.34 – 7.21 (m, 5H), 7.17 (td, $J = 8.2, 5.9$ Hz, 1H), 6.92 – 6.81 (m, 2H), 5.15 (d, $J = 7.6$ Hz, 1H), 4.54 (q, $J = 7.2$ Hz, 1H), 3.45 – 3.00 (m, 10H), 1.39 (s, 9H).

Mass: Calculated m/z for $\text{C}_{10}\text{H}_{12}\text{FN}_2\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 492.1963$; found APCI: 492.6

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((2R)-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)-6-fluorophenyl]-2-[(2-fluorophenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: **DCM** (10 mL); **TFA** (10 mL); **9** (1769 mg, 3.60 mmol) Boc-protected crude product isolation: 1019 mg; Used for next step: 201 mg (0.51 mmol, 1 eq.); **THF** (10 ml); **2-fluorobenzoic acid** (86 mg, 0.62 mmol, 1.2 eq., TCI); **DEPBT** (169 mg, 0.57 mmol, 1.1 eq., BLDpharm); **DIPEA** (174 μL , 1.02 mmol, 2 eq., Aldrich)

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 70 % heptane to 20 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μm particle size. Yield: 238 mg; 91 %

Analyses:

R_f value: 0.15 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 16**) (400 MHz, Chloroform-*d*) δ 8.08 (s, 1H), 7.97 (t, $J = 7.8$ Hz, 1H), 7.50 – 7.43 (m, 1H), 7.36 – 7.20 (m, 7H), 7.16 (td, $J = 8.2, 5.8$ Hz, 1H), 7.09 (dd, $J = 12.1, 8.3$ Hz, 1H), 6.94 – 6.83 (m, 2H), 5.07 (qd, $J = 6.9, 1.7$ Hz, 1H), 3.42 – 3.23 (m, 6H), 3.23 – 3.02 (m, 4H).

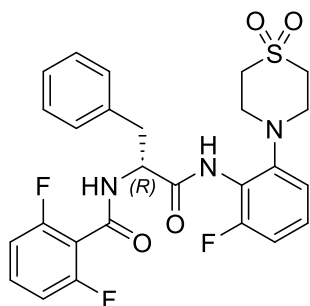
^{13}C NMR (**Spectrum 17**) (101 MHz, Chloroform-*d*) δ 169.65, 163.93 (d, $J = 3.0$ Hz), 160.73 (d, $J = 248.7$ Hz), 157.63 (d, $J = 252.4$ Hz), 147.82, 136.03, 134.12 (d, $J = 9.3$ Hz), 131.77 (d, $J = 1.8$ Hz), 129.44, 128.83, 127.72 (d, $J = 9.3$ Hz), 127.29, 125.06 (d, $J = 3.2$ Hz), 119.81 (d, $J = 10.7$ Hz), 119.47 (d, $J = 13.5$ Hz), 116.36 (d, $J = 3.4$ Hz), 116.26 (d, $J = 24.4$ Hz), 112.90 (d, $J = 20.7$ Hz), 55.62, 51.96, 50.66, 37.36.

Mass: Calculated m/z for $\text{C}_{26}\text{H}_{26}\text{F}_2\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 514.1607$; found APCI: 514.5; found HRMS: $[\text{M}+\text{H}]^+ 514.1601$

HPLC purity: 97.6 %; $t_R = 2.7$ min (**HPLC-Trace 7**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

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(2R)-2-[(2,6-difluorophenyl)formamido]-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)-6-fluorophenyl]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product of **10** (184 mg, 0.47 mmol, 1 eq.); **THF** (10 ml); **2,6-difluorobenzoic acid** (89 mg, 0.56 mmol, 1.2 eq., TCI); **DEPBT** (295 mg, 0.99 mmol, 2.1 eq., BLDpharm); **DIPEA** (160 μL , 1.41 mmol, 3 eq., Aldrich)

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 70 % heptane to 25 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μm particle size. Afterwards preparative HPLC. Yield: 170 mg; 68 %

Analyses:

R_F value: 0.75 ethyl acetate

^1H NMR (**Spectrum 18**) (400 MHz, Acetone-*d*₆) δ 8.85 (s, 1H), 8.31 (d, $J = 8.3$ Hz, 1H), 7.52 – 7.37 (m, 3H), 7.36 – 7.21 (m, 4H), 7.09 – 6.91 (m, 4H), 5.19 (q, $J = 8.3$ Hz, 1H), 3.50 – 3.23 (m, 7H), 3.22 – 3.10 (m, 3H).

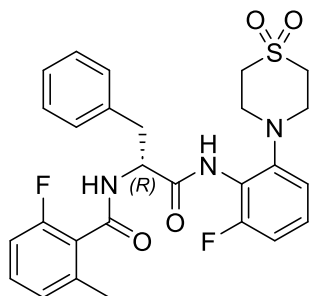
^{13}C NMR (**Spectrum 19**) (101 MHz, Acetone- d_6) δ 169.76 (d, $J = 8.5$ Hz), 160.34 (d, $J = 8.2$ Hz), 159.63 (d, $J = 250.2$ Hz), 159.56 (d, $J = 250.3$ Hz), 158.16 (d, $J = 250.0$ Hz), 149.09, 137.28, 131.76 (t, $J = 10.0$ Hz), 129.35, 128.29, 127.71 (d, $J = 9.5$ Hz), 126.64, 119.99 – 119.35 (m), 116.33 (d, $J = 3.0$ Hz), 115.25 – 114.38 (m), 111.82 – 111.41 (m), 55.35, 52.00, 50.39, 37.74.

Mass: Calculated m/z for $\text{C}_{26}\text{H}_{25}\text{F}_3\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 532.1513$; found APCI: 532.5; found HRMS: $[\text{M}+\text{H}]^+ 532.1507$

HPLC purity: 98.9 %; $t_R = 2.6$ min (**HPLC-Trace 8**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

12

(2R)-N-[2-(1,1-dioxo--thiomorpholin-4-yl)-6-fluorophenyl]-2-[(2-fluoro-6-methylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product of **10** (193 mg, 0.49 mmol, 1 eq.); **THF** (10 ml); **2-fluoro-6-methylbenzoic acid** (91 mg, 0.59 mmol, 1.2 eq., TCI); **DEPBT** (308 mg, 1.03 mmol, 2.1 eq., BLDpharm); **DIPEA** (251 μL , 1.47 mmol, 3 eq., Aldrich)

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 70 % heptane to 25 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μm particle size. Yield: 245 mg; 95 %

Analyses:

R_f value: 0.25 heptane/ethyl acetate 1:1

^1H NMR (**Spectrum 20**) (400 MHz, Chloroform- d) δ 8.09 (d, $J = 10.3$ Hz, 1H), 7.37 – 7.22 (m, 5H), 7.22 – 7.12 (m, 2H), 6.93 – 6.79 (m, 4H), 6.70 (q, $J = 7.9$ Hz, 1H), 5.16 (tdd, $J = 8.0, 6.9, 6.4, 2.6$ Hz, 1H), 3.45 – 3.23 (m, 5H), 3.20 – 3.03 (m, 5H), 2.08 (s, 3H).

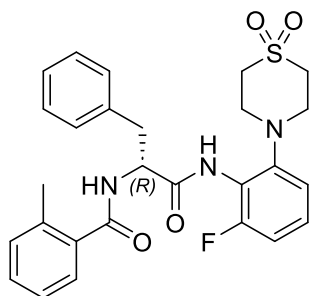
^{13}C NMR (**Spectrum 21**) (101 MHz, Chloroform- d) δ 169.61, 165.82, 159.08 (d, $J = 245.5$ Hz), 157.78 (d, $J = 251.1$ Hz), 148.01, 138.83 – 137.77 (m), 136.13, 130.84 (d, $J = 8.8$ Hz), 129.37, 128.81, 127.91 (d, $J = 9.7$ Hz), 127.25, 126.27 (d, $J = 2.6$ Hz), 123.77 (d, $J = 16.6$ Hz), 119.17 (d, $J = 13.4$ Hz), 116.25 (d, $J = 3.0$ Hz), 112.86 (d, $J = 21.7$ Hz), 112.70 (d, $J = 20.4$ Hz), 54.93, 51.84, 50.54, 37.40, 18.79.

Mass: Calculated m/z for C₂₇H₂₈F₂N₃O₄S⁺ [M+H]⁺ = 528.1763; found APCI: 528.5; found HRMS: [M+H]⁺ 528.1758

HPLC purity: 99.3 %; t_R=2.7 min (**HPLC-Trace 9**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm, acetonitrile/ water 15:85, 6 min, v=1.2 mL/min, λ=254 nm)

13

(2R)-N-[2-(1,1-dioxo-1λ⁶-thiomorpholin-4-yl)-6-fluorophenyl]-2-[(2-methylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product of **10** (306 mg, 0.82 mmol, 1 eq.); **DMF** (10 ml); **2-methylbenzoic acid** (134 mg, 0.98 mmol, 1.2 eq., TCI); **PyBOP** (640 mg, 1.23 mmol, 1.5 eq., BLDpharm); **DIPEA** (418 μL, 2.46 mmol, 3 eq., Aldrich)

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 70 % heptane to 30 % over 10 CV on 8 g normal phase silica gel 0.63 – 0.2 μm particle size. Afterwards preparative HPLC. Yield: 311 mg; 74 %

Analyses:

R_F value: 0.20 heptane/ethyl acetate 1:1

¹H NMR (**Spectrum 22**) (400 MHz, Chloroform-*d*) δ 8.11 (s, 1H), 7.41 – 7.24 (m, 6H), 7.22 – 7.09 (m, 4H), 6.93 – 6.81 (m, 2H), 6.48 (d, *J* = 7.6 Hz, 1H), 5.05 (q, *J* = 7.4 Hz, 1H), 3.46 – 3.23 (m, 5H), 3.18 (dd, *J* = 13.8, 7.2 Hz, 1H), 3.13 – 3.02 (m, 4H), 2.29 (s, 3H).

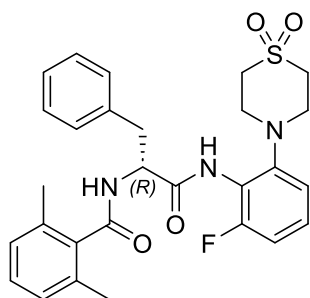
¹³C NMR (**Spectrum 23**) (101 MHz, Chloroform-*d*) δ 170.66, 169.92, 157.74 (d, *J* = 251.5 Hz), 147.83, 136.37, 136.20, 134.69, 131.21, 130.54, 129.39, 128.89, 127.85 (d, *J* = 9.4 Hz), 127.35, 126.84, 125.86, 119.25 (d, *J* = 13.4 Hz), 116.20 (d, *J* = 2.7 Hz), 112.76 (d, *J* = 20.7 Hz), 55.25, 51.96, 50.58, 37.19, 19.72.

Mass: Calculated m/z for C₂₇H₂₉FN₃O₄S⁺ [M+H]⁺ = 510.1858⁺; [M+Na]⁺ = 532.1677; found APCI: 528.5; found HRMS: [M+H]⁺ 510.1864; [M+Na]⁺ 532.1679

HPLC purity: 99.5 %; t_R=2.7 min (**HPLC-Trace 10**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm, acetonitrile/ water 15:85, 6 min, v=1.2 mL/min, λ=254 nm)

14

(2R)-2-[(2,6-dimethylphenyl)formamido]-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)-6-fluorophenyl]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product from **10**: 195 mg (0.50 mmol, 1 eq.); **THF** (10 ml); **2,6-dimethylbenzoic acid** (90 mg, 0.60 mmol, 1.2 eq., TCI); **PyBOP** (313 mg, 1.05 mmol, 2.1 eq., Carbolution); **DIPEA** (255 μ L, 1.5 mmol, 3 eq., Aldrich)

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 60 % heptane to 15 % over 10 CV on 8 g normal phase silica gel 0.63 – 0.2 μ m particle size. Afterwards preparative HPLC. Yield: 99 mg; 38 %

Analyses:

R_f value: 0.25 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 24**) (500 MHz, Chloroform-*d*) δ 8.25 – 8.15 (m, 1H), 7.32 (dd, J = 23.3, 4.4 Hz, 5H), 7.24 – 7.16 (m, 1H), 7.12 (t, J = 7.6 Hz, 1H), 6.94 (d, J = 7.7 Hz, 2H), 6.89 (dd, J = 12.0, 8.3 Hz, 2H), 6.33 (d, J = 7.9 Hz, 1H), 5.17 (q, J = 7.2, 6.2 Hz, 1H), 3.45 – 3.27 (m, 5H), 3.16 – 3.03 (m, 5H), 2.07 (s, 6H).

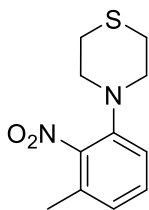
^{13}C NMR (**Spectrum 25**) (126 MHz, Chloroform-*d*) δ 171.19, 169.73, 157.72 (d, J = 251.8 Hz), 147.79, 136.19, 136.15, 134.32, 129.31, 129.13, 128.93, 127.83 (d, J = 9.9 Hz), 127.53, 127.38, 119.14 (d, J = 14.3 Hz), 116.14, 112.83 (d, J = 20.6 Hz), 54.80, 52.01, 50.57, 37.37, 18.76.

Mass: Calculated m/z for $\text{C}_{28}\text{H}_{31}\text{FN}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+$ = 524.2014; found APCI: 524.5; found HRMS: $[\text{M}+\text{H}]^+$ 524.2010

HPLC purity: 98.4 %; t_R = 2.9 min (**HPLC-Trace 11**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μ m, acetonitrile/ water 15:85, 6 min, v = 1.2 mL/min, λ = 254 nm)

15

4-(3-methyl-2-nitrophenyl)thiomorpholine



Synthesis:

Quantities: **1-Fluoro-3-methyl-2-nitrobenzene** (1000 mg, 6.45 mmol, 1 eq., BLDpharm); **thiomorpholine** (731 mg, 7.09 mmol, 1.1 eq., abcr GmbH); **DIPEA** (2194 μ L, 12.90 mmol, 2 eq., Aldrich)

Procedure: Mix **1-fluoro-3-methyl-2-nitrobenzene** and **thiomorpholine**; add **DIPEA**; stir for 24 h at 50 $^{\circ}$ C.

After the reaction time the crude reaction mixture was directly evaporated on silica gel.

Purification:

Flash chromatography heptane/dichloromethane gradient 95 % heptane to 50 % over 10 CV on 90 g normal phase silica gel 0.63 – 0.2 μ m particle size. Yield: 313 mg; 18 %

Analyses:

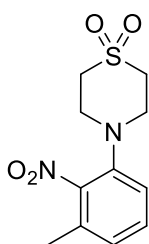
R_f value: 0.30 dichloromethane/heptane 1:1

^1H NMR (**Spectrum 26**) (400 MHz, Chloroform-*d*) δ 7.30 (t, J = 7.9 Hz, 1H), 7.09 (dd, J = 8.0, 1.2 Hz, 1H), 7.03 (dt, J = 7.6, 1.0 Hz, 1H), 3.20 – 3.13 (m, 4H), 2.74 – 2.65 (m, 4H), 2.25 (s, 3H).

Mass: Calculated m/z for $\text{C}_7\text{H}_7\text{FNO}_2^+$ $[\text{M}+\text{H}]^+$ = 239.0849; found APCI: 239.1

16

4-(3-methyl-2-nitrophenyl)-thiomorpholine-1,1-dioxide



Synthesis:

Quantities: **15** (331 mg, 1.39 mmol, 1 eq.); **mCPBA 77 % m/m** (747 mg, 3.33 mmol, 2.4 eq., Aldrich); **DCM** (15 + 15 mL)

Procedure: Dissolve **15** in 15 mL **DCM**; cool the solution to -20 $^{\circ}$ C (iso-propanol bath); dissolve mCPBA in 15 mL **DCM** and add it to the **15** solution over 30 min; stir for 4 h at 0 $^{\circ}$ C; wash the mixture with saturated NaHCO_3 solution; concentrate under reduced pressure

Purification:

Was used without further purification. Crude product yield: 376 mg; 100 %

Analyses:

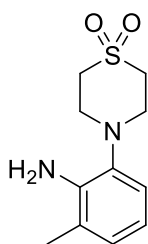
R_f value: 0.7 ethyl acetate

^1H NMR (**Spectrum 27**) (400 MHz, Chloroform- d) δ 7.36 (t, $J = 7.9$ Hz, 1H), 7.17 (d, $J = 8.0$ Hz, 1H), 7.13 (d, $J = 7.7$ Hz, 1H), 3.49 – 3.41 (m, 4H), 3.16 – 3.07 (m, 4H), 2.28 (s, 3H).

Mass: Calculated m/z for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 271.3103$; found APCI: 271.2

17

4-(2-amino-3-methylphenyl)-1 λ^6 -thiomorpholine-1,1-dioxide



Synthesis:

Quantities: **16** (376 mg, 1.39 mmol, 1 eq.); **PdOH/C 20 % m/m** (75 mg (20 % m of **16**), Aldrich); **methanol** (40 mL)

Procedure: Dissolve **15** in 40 mL **methanol**; add catalyst; close the flask with a septum and purge the mixture with hydrogen for 30 min under stirring; add a hydrogen balloon; check for full conversion with TLC; after completion remove the catalyst with paper filter and wash the filter thoroughly with ethyl acetate; evaporate the solvents under reduced pressure

Purification:

Was used without further purification. Crude product yield: 196 mg; 58 %

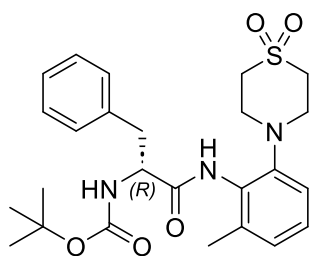
Analyses:

R_f value: 0.25 ethyl acetate/heptane 1:1

Mass: Calculated m/z for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 241.1005$; found APCI: 241.1

18

tert-butyl-N-[(1R)-1-[[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)-6-methylphenyl]carbamoyl]-2-phenylethyl]carbamate



Synthesis:

Quantities: Pyridine (15 mL); EtOAc (30 mL); **17** (196 mg, 0.82 mmol, 1 eq.); **N-Boc-D-phenylalanine** (238 mg, 0.90 mmol, 1.1 eq., TCI); **T3P 50 % m/V in EtOAc** (976 μ L, 1.64 mmol, 2 eq., Aldrich)

Procedure: Dissolve **17** and **(N)-Boc-(R)-phenylalanine** in pyridine and ethyl acetate; cool the mixture to -20 °C before adding **T3P**; keep the temperature at 0 °C for 20 h; wash with 0.25 M KH₂PO₄ solution 3x; evaporate the solvents under reduced pressure

Purification:

Flash chromatography heptane/methyl tert-butyl ether gradient 50 % heptane to 10 % over 8 CV on 90 g normal phase silica gel 0.63 – 0.2 μ m particle size. Yield: 222 mg; 56 %

Analyses:

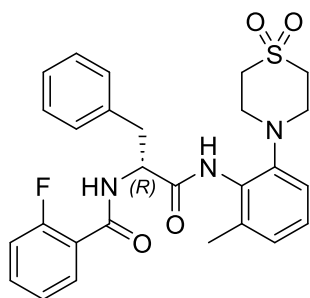
R_f value: 0.35 ethyl acetate/heptane 1:1

¹H NMR (**Spectrum 28**) (400 MHz, Chloroform-*d*) δ 7.88 (s, 1H), 7.35 – 7.23 (m, 5H), 7.14 (t, *J* = 7.8 Hz, 1H), 7.01 (d, *J* = 7.6 Hz, 1H), 6.94 (d, *J* = 7.8 Hz, 1H), 5.23 – 5.10 (m, 1H), 4.58 (q, *J* = 7.5 Hz, 1H), 3.34 – 3.17 (m, 5H), 3.15 – 3.00 (m, 5H), 2.09 (s, 3H), 1.41 (s, 9H).

Mass: Calculated *m/z* for C₂₅H₃₄N₃O₅S⁺ [M+H]⁺ = 488.2214; found APCI: 488.2

19

(2R)-N-[2-(1,1-dioxo-1 λ ⁶-thiomorpholin-4-yl)-6-methylphenyl]-2-[(2-fluorophenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: DCM (10 mL); TFA (10 mL); **18** (222 mg, 0.46 mmol) Boc-protected crude product isolation: 145 mg; Used for next step: 59 mg (0.15 mmol, 1 eq.); THF (10 ml); **2-fluorobenzoic acid** (26 mg, 0.18 mmol, 1.2 eq., TCI); **DEPBT** (68 mg, 0.23 mmol, 1.5 eq., BLDpharm); **DIPEA** (78 μ L, 0.47 mmol, 3 eq., Aldrich)

Procedure: Dissolve **18** in 10 mL **DCM** and add 10 mL **TFA** under stirring; stir for 1 h and check for full conversion by TLC; after conversion evaporate the solvents under reduced pressure; resuspend the residue in ethyl acetate and wash with saturated NaHCO₃ 3x; dry the organic phase with NaSO₄ and concentrate under reduced pressure; dissolve the Boc-protected reactant, the **2-fluorobenzoic acid** and **DEPBT** in 10 mL **THF**; add **DIPEA**; stir over night

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 80 % heptane to 40 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 µm particle size. Preparative HPLC afterwards. Yield: 48 mg; 62 %

Analyses:

R_F value: 0.30 ethyl acetate/heptane 1:1

¹H NMR (**Spectrum 29**) (400 MHz, Chloroform-*d*) δ 8.10 (s, 1H), 8.05 (td, *J* = 7.9, 1.9 Hz, 1H), 7.55 – 7.47 (m, 1H), 7.40 – 7.24 (m, 7H), 7.17 – 7.10 (m, 2H), 7.02 (d, *J* = 7.5 Hz, 1H), 6.96 (dd, *J* = 7.9, 1.4 Hz, 1H), 5.14 (qd, *J* = 7.1, 2.0 Hz, 1H), 3.39 (dd, *J* = 13.8, 7.0 Hz, 1H), 3.34 – 3.19 (m, 5H), 3.18 – 3.01 (m, 4H), 2.10 (s, 3H).

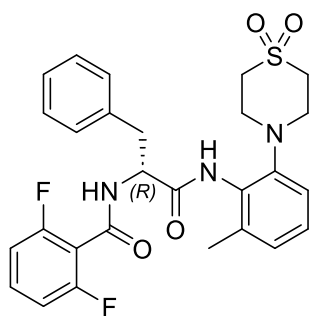
¹³C NMR (**Spectrum 30**) (101 MHz, Chloroform-*d*) δ 169.15, 163.77 (d, *J* = 3.3 Hz), 160.76 (d, *J* = 248.3 Hz), 146.54, 136.36, 136.20, 134.21 (d, *J* = 9.5 Hz), 131.99, 129.97, 129.50, 128.92, 127.82, 127.37, 127.28, 125.17 (d, *J* = 3.2 Hz), 119.68 (d, *J* = 11.0 Hz), 118.46, 116.27 (d, *J* = 24.7 Hz), 55.67, 52.00, 51.07, 37.30, 18.98.

Mass: Calculated m/z for C₂₇H₂₉FN₃O₄S⁺ [M+H]⁺ = 510.1858; found APCI: 510.3; found HRMS: [M+H]⁺ 510.1852

HPLC purity: 98.6 %; t_R=2.8 min (**HPLC-Trace 12**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 µm, acetonitrile/water 15:85, 6 min, v=1.2 mL/min, λ=254 nm)

20

(2R)-2-[(2,6-difluorophenyl)formamido]-N-[2-(1,1-dioxo-1λ⁶-thiomorpholin-4-yl)-6-methylphenyl]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product of **19** (61 mg, 0.16 mmol, 1 eq.); **THF** (10 ml); **2,6-difluorobenzoic acid** (30 mg, 0.19 mmol, 1.2 eq., TCI); **DEPBT** (71 mg, 0.24 mmol, 1.5 eq., BLDpharm); **DIPEA** (80 μ L, 0.47 mmol, 3 eq., Aldrich)

Procedure: Dissolve the Boc-protected reactant of **19**, the **2,6-difluorobenzoic acid** and **DEPBT** in 10 mL **THF**; add **DIPEA**; stir over night

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 70 % heptane to 30 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μ m particle size. Preparative HPLC afterwards. Yield: 67 mg; 81 %

Analyses:

R_f value: 0.50 heptane/ethyl acetate 1:1

^1H NMR (**Spectrum 31**) (400 MHz, Chloroform-*d*) δ 8.01 (s, 1H), 7.40 – 7.23 (m, 6H), 7.14 (t, J = 7.8 Hz, 1H), 7.00 (d, J = 7.6 Hz, 1H), 6.95 (d, J = 7.9 Hz, 1H), 6.90 (t, J = 8.2 Hz, 2H), 6.81 (d, J = 8.0 Hz, 1H), 5.15 (q, J = 7.4 Hz, 1H), 3.36 (dd, J = 13.9, 7.4 Hz, 1H), 3.32 – 3.18 (m, 5H), 3.14 – 3.03 (m, 4H), 2.07 (s, 3H).

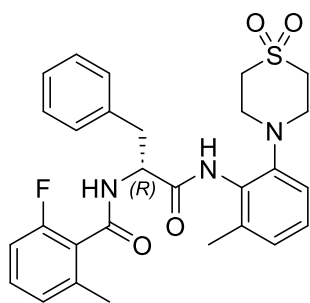
^{13}C NMR (**Spectrum 32**) (101 MHz, Chloroform-*d*) δ 168.88, 160.79, 159.94 (d, J = 252.9 Hz), 159.87 (d, J = 252.8 Hz), 146.72, 136.52, 135.98, 132.33 (t, J = 10.4 Hz), 129.71, 129.50, 128.88, 127.62, 127.44, 127.38, 118.40, 113.12 (t, J = 19.6 Hz), 112.35 – 111.90 (m), 55.32, 51.92, 50.95, 37.48, 18.82.

Mass: Calculated m/z for $\text{C}_{27}\text{H}_{28}\text{F}_2\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+$ = 528.1763; found APCI: 528.3; found HRMS: $[\text{M}+\text{H}]^+$ 528.1760

HPLC purity: 99.0 %; t_R = 2.7 min (**HPLC-Trace 13**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μ m, acetonitrile/ water 15:85, 6 min, v = 1.2 mL/min, λ = 254 nm)

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(2R)-N-[2-(1,1-dioxo-1 λ ⁶-thiomorpholin-4-yl)-6-methylphenyl]-2-[(2-fluoro-6-methylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product of **19** (64 mg, 0.17 mmol, 1 eq.); **THF** (10 mL); **2-fluoro-6-methylbenzoic acid** (31 mg, 0.20 mmol, 1.2 eq., TCI); **DEPBT** (74 mg, 0.25 mmol, 1.5 eq., BLDpharm); **DIPEA** (84 μ L, 0.50 mmol, 3 eq., Aldrich)

Procedure: Dissolve the Boc-protected reactant of **19**, the **2-fluoro-6-methylbenzoic acid** and **DEPBT** in 10 mL **THF**; add **DIPEA**; stir over night

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 60 % heptane to 20 % over 10 CV on 45 g normal phase silica gel 0.63 – 0.2 μ m particle size. Preparative HPLC afterwards. Yield: 50 mg; 58 %

Analyses:

R_f value: 0.55 heptane/ethyl acetate 1:1

^1H NMR (**Spectrum 33**) (400 MHz, Chloroform-*d*) δ 8.10 (s, 1H), 7.36 – 7.12 (m, 7H), 7.03 (d, $J = 7.7$ Hz, 1H), 6.95 (dd, $J = 16.5, 7.8$ Hz, 2H), 6.86 (t, $J = 9.0$ Hz, 1H), 6.57 (t, $J = 9.4$ Hz, 1H), 5.18 (td, $J = 8.4, 6.3$ Hz, 1H), 3.44 (dd, $J = 14.1, 6.2$ Hz, 1H), 3.29 (q, $J = 6.1, 5.1$ Hz, 4H), 3.14 (tdd, $J = 15.4, 10.6, 5.3$ Hz, 5H), 2.14 (s, 3H), 2.09 (s, 3H).

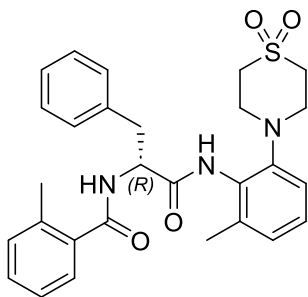
^{13}C NMR (**Spectrum 34**) (101 MHz, Chloroform-*d*) δ 169.24, 165.63, 159.14 (d, $J = 245.3$ Hz), 146.70, 138.56 (d, $J = 2.8$ Hz), 136.54, 136.25, 130.90 (d, $J = 9.0$ Hz), 129.81, 129.31, 128.89, 127.70, 127.43, 127.31, 126.39 (d, $J = 2.8$ Hz), 123.73 (d, $J = 17.0$ Hz), 118.43, 112.90 (d, $J = 22.0$ Hz), 55.01, 51.97, 50.99, 37.55, 18.88, 18.85.

Mass: Calculated m/z for $\text{C}_{28}\text{H}_{31}\text{FN}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 524.2014$; found APCI: 524.4; found HRMS: $[\text{M}+\text{H}]^+ 524.2010$

HPLC purity: 98.0 %; $t_R = 2.8$ min (**HPLC-Trace 14**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μ m, acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

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(2R)-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)-6-methylphenyl]-2-[(2-methylphenyl)formamido]-3-phenylpropanamide



Synthesis:

Quantities: Boc-deprotected crude product of **19** (67 mg, 0.17 mmol, 1 eq.); **THF** (10 mL); **2-methylbenzoic acid** (28 mg, 0.21 mmol, 1.2 eq., TCI); **PyBOP** (135 mg, 0.26 mmol, 1.5 eq., Carbolution); **DIPEA** (88 μ L, 0.52 mmol, 3 eq., Aldrich)

Procedure: Dissolve the Boc-deprotected reactant of **19**, the **2-methylbenzoic acid** and **PyBOP** in 10 mL **THF**; add **DIPEA**; stir over night

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 75 % heptane to 30 % over 8 CV on 8 g normal phase silica gel 0.63 – 0.2 μ m particle size. Preparative HPLC afterwards. Yield: 14 mg; 16 %

Analyses:

R_F value: 0.20 heptane/ethyl acetate 1:1

^1H NMR (**Spectrum 35**) (400 MHz, Chloroform-*d*) δ 8.11 (s, 1H), 7.41 – 7.26 (m, 6H), 7.23 – 7.12 (m, 4H), 7.04 (d, $J = 7.6$ Hz, 1H), 6.98 (dd, $J = 7.9, 1.4$ Hz, 1H), 6.31 (d, $J = 7.6$ Hz, 1H), 5.03 (q, $J = 7.5$ Hz, 1H), 3.42 (dd, $J = 14.0, 7.1$ Hz, 1H), 3.39 – 3.24 (m, 4H), 3.20 (dd, $J = 14.0, 7.9$ Hz, 1H), 3.18 – 3.09 (m, 4H), 2.32 (s, 3H), 2.15 (s, 3H).

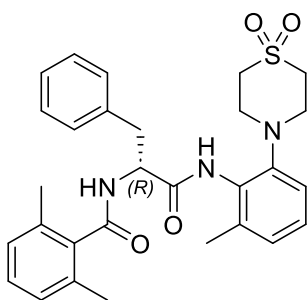
^{13}C NMR (**Spectrum 36**) (101 MHz, Chloroform-*d*) δ 170.40, 169.57, 146.31, 136.49, 136.32, 136.25, 134.64, 131.33, 130.60, 129.81, 129.34, 128.99, 127.83, 127.44, 127.36, 126.72, 125.92, 118.33, 55.38, 52.06, 51.08, 37.37, 19.84, 18.99.

Mass: Calculated m/z for $\text{C}_{28}\text{H}_{32}\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 506.2108$; $[\text{M}+\text{Na}]^+ = 528.1927$; found APCI: 506.1; found HRMS: $[\text{M}+\text{H}]^+ 506.2112$; $[\text{M}+\text{Na}]^+ 528.1932$

HPLC purity: 97.3 %; $t_R = 2.8$ min (**HPLC-Trace 15**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μ m, acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

23

(2R)-2-[(2,6-dimethylphenyl)formamido]-N-[2-(1,1-dioxo-1 λ^6 -thiomorpholin-4-yl)-6-methylphenyl]-3-phenylpropanamide



Synthesis:

Quantities: Boc-protected crude product of **19** (74 mg, 0.19 mmol, 1 eq.); **DMF** (10 mL); **2,6-dimethylbenzoic acid** (35 mg, 0.23 mmol, 1.2 eq., TCI); **PyBOP** (198 mg, 0.38 mmol, 2 eq., Carbolution); **DIPEA** (97 μ L, 0.57 mmol, 3 eq., Aldrich)

Procedure: Dissolve the Boc-protected reactant of **19**, the **2,6-dimethylbenzoic acid** and **PyBOP** in 10 mL **DMF**; add **DIPEA**; stir over night

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography heptane/ethyl acetate gradient 70 % heptane to 30 % over 8 CV on 8 g normal phase silica gel 0.63 – 0.2 μ m particle size. Preparative HPLC afterwards. Yield: 76 mg; 77 %

Analyses:

R_f value: 0.30 heptane/ethyl acetate 1:1

^1H NMR (**Spectrum 37**) (400 MHz, Chloroform-*d*) δ 8.26 (s, 1H), 7.38 – 7.31 (m, 4H), 7.31 – 7.26 (m, 1H), 7.17 (t, $J = 7.8$ Hz, 1H), 7.12 (t, $J = 7.6$ Hz, 1H), 7.06 (d, $J = 7.6$ Hz, 1H), 7.00 (d, $J = 7.8$ Hz, 1H), 6.94 (d, $J = 7.6$ Hz, 2H), 6.20 (d, $J = 7.6$ Hz, 1H), 5.13 (ddd, $J = 9.6, 7.6, 5.8$ Hz, 1H), 3.44 (dd, $J = 14.3, 5.8$ Hz, 1H), 3.41 – 3.25 (m, 4H), 3.20 (s, 4H), 3.07 (dd, $J = 14.3, 9.6$ Hz, 1H), 2.18 (s, 3H), 2.03 (s, 6H).

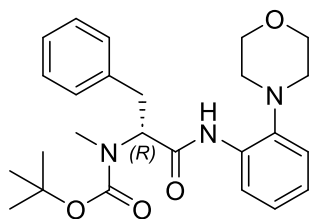
^{13}C NMR (**Spectrum 38**) (126 MHz, Chloroform-*d*) δ 170.97, 169.82, 146.18, 136.33, 136.25, 136.15, 134.27, 129.80, 129.17, 129.07, 129.01, 127.88, 127.52, 127.42, 127.28, 118.29, 54.89, 52.07, 51.08, 37.76, 19.13, 18.81.

Mass: Calculated m/z for $\text{C}_{29}\text{H}_{34}\text{N}_3\text{O}_4\text{S}^+$ $[\text{M}+\text{H}]^+ = 520.2265$; $[\text{M}+\text{Na}]^+ = 542.2084$; found APCI: 520.5; found HRMS: $[\text{M}+\text{H}]^+ 520.2271$; $[\text{M}+\text{Na}]^+ 542.2088$

HPLC purity: 99.3 %; $t_R = 3.0$ min (**HPLC-Trace 16**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μ m, acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

24

tert-butyl (*R*)-methyl(1-((2-morpholinophenyl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate



Synthesis:

Quantities: Pyridine (5 mL); EtOAc (10 mL); **2-morpholinoaniline** (166 mg, 0.94 mmol, 1 eq.); **N-Boc-N-methyl-(R)-phenylalanine** (289 mg, 1.1 mmol, 1.1 eq.); **T3P 50 % m/v in EtOAc** (559 μ L, 1.88 mmol, 2.0 eq.)

Procedure: **2-Morpholinoaniline** and **N-Boc-N-methyl-(R)-phenylalanine** were dissolved in a 1:2 mixture of distilled pyridine and EtOAc. The mixture was cooled to -20 °C with an isopropanol dry-ice bath before **T3P 50 % m/V in EtOAc** was added. After the addition, the cooling bath was removed, and the mixture was stirred for 20 h at room temperature. Afterwards the mixture was washed with 0.25 M KH₂PO₄ solution three times before the organic phase was evaporated on silica gel under reduced pressure for purification.

Purification:

Flash chromatography ethyl acetate/heptane gradient 80 % heptane to 40 % over 8 CV on 45 g. Yield: 162 mg; 39 %

Analyses:

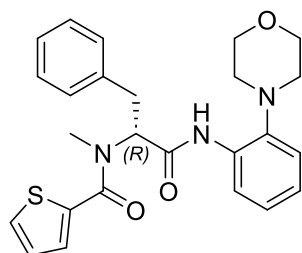
R_F value: 0.25 ethyl acetate/heptane 1:1

¹H NMR (**Spectrum 39**) (400 MHz, Chloroform-*d*) δ 9.25 – 9.14 (m, 1H), 8.51 – 8.41 (m, 1H), 7.32 – 7.05 (m, 8H), 5.27 – 5.20 (m, 1H), 4.86 – 4.78 (m, 0H), 3.89 – 3.75 (m, 4H), 3.59 – 3.47 (m, 1H), 3.01 – 2.93 (m, 1H), 2.91 – 2.66 (m, 7H), 1.42 – 1.10 (m, 9H).

Mass: Calculated m/z for C₂₅H₃₄N₃O₄⁺ [M+H]⁺ = 440.2544; found APCI: [M+H]⁺ 440.1

25

(*R*)-*N*-methyl-*N*-(1-((2-morpholinophenyl)amino)-1-oxo-3-phenylpropan-2-yl)thiophene-2-carboxamide



Synthesis:

Quantities: DCM (2 mL); TFA (2 mL); **24** (100 mg, 0.23 mmol, 1.0 eq.); **thiophene-2-carboxylic acid** (30 mg, 0.23 mmol, 1.0 eq.); **PyBOP** (132 mg, 0.25 mmol, 1.1 eq.); **DMF** (5 mL), **DIPEA** (120 μ L, 0.69 mmol, 3.0 eq.)

Procedure: Dissolve **24** in **DCM** and add **TFA** under stirring; stir for 1 h and check for full conversion by TLC; after conversion evaporate the solvents under reduced pressure; followed by coevaporation with toluene 3x, DCM 3x; dissolve the Boc-protected reactant, the **thiophene-2-carboxylic acid** and **PyBOP** in **DMF**; add **DIPEA**; stir over night at 65 °C under argon and light protection.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography ethyl acetate/heptane gradient 95 % heptane to 50 % over 10 CV on 45 g. Yield: 38 mg; 37 %

Analyses:

R_f value: 0.25 ethyl acetate/heptane 1:1

^1H NMR (**Spectrum 40**) (400 MHz, Chloroform-*d*) δ 9.44 (s, 1H), 8.46 (d, $J = 8.1$ Hz, 1H), 7.46 (d, $J = 5.0$ Hz, 1H), 7.36 – 7.26 (m, 4H), 7.24 – 7.05 (m, 5H), 7.03 – 6.97 (m, 1H), 5.63 (s, 1H), 3.93 (s, 4H), 3.51 (dd, $J = 14.8, 6.7$ Hz, 1H), 3.23 (s, 1H), 3.19 (s, 3H), 2.88 – 2.75 (m, 4H).

^{13}C NMR (**Spectrum 41**) (126 MHz, Chloroform-*d*) δ 168.1, 165.6, 141.4, 137.1, 134.0, 130.3, 129.0, 128.8, 127.1, 126.9, 125.8, 124.2, 121.2, 119.7, 67.4, 59.5, 52.9, 33.9.

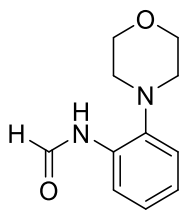
Additional HSQC is provided because of superimposed signals (**Spectrum 42**).

Mass: Calculated m/z for $\text{C}_{25}\text{H}_{28}\text{N}_3\text{O}_3\text{S}^+$ $[\text{M}+\text{H}]^+ = 450.1846$; $[\text{M}+\text{Na}]^+ = 472.1665$; found APCI: $[\text{M}+\text{H}]^+ 450.1$; found HRMS: $[\text{M}+\text{H}]^+ 450.1850$; $[\text{M}+\text{Na}]^+ 472.1668$

HPLC purity: 98.4 %; $t_R = 3.3$ min (**HPLC-Trace 17**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

26

N-(2-morpholinophenyl)formamide



Synthesis:

Quantities: **Formic acid** (422 μ L, 11.2 mmol, 4.0 eq.); **2-morpholinoaniline** (500 mg, 2.8 mmol, 1.0 eq.); **sodium formate** (38 mg, 0.56 mmol, 0.2 eq.)

Procedure: **2-morpholinoaniline** and **sodium formate** were suspended in **formic acid** and stirred at room temperature overnight.

Purification:

The reaction mixture was poured in ethyl acetate and washed with saturated NaHCO_3 solution three times. Followed by flash chromatography ethyl acetate/DCM gradient 70 % ethyl acetate to 30 % over 13 CV on 45 g. Yield: 456 mg; 79 %

Analyses:

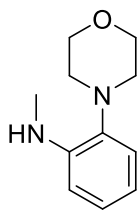
R_f value: 0.15 ethyl acetate/DCM 9:1

^1H NMR (**Spectrum 43**) (400 MHz, Chloroform- d) δ 8.87 (d, $J = 11.9$ Hz, 0.4H), 8.51 (s, 1H), 8.39 (dd, $J = 8.0, 1.5$ Hz, 0.6H), 8.14 (s, 1H), 7.25 – 7.08 (m, 3H), 3.93 – 3.78 (m, 4H), 2.93 – 2.79 (m, 4H).

Mass: Calculated m/z for $\text{C}_{11}\text{H}_{15}\text{N}_2\text{O}_2^+$ $[\text{M}+\text{H}]^+ = 207.1128$; found APCI: $[\text{M}+\text{H}]^+ 207.0$

27

N-methyl-2-morpholinoaniline



Synthesis:

Quantities: **26** (457 mg, 2.21 mmol, 1.0 eq.); **LiAlH₄ 1 M in THF** (4.4 mL, 4.42 mmol, 2.0 eq.); **THF** (10 mL)

Procedure: **26** was dissolved in **THF** and **LiAlH₄ 1 M in THF** was added dropwise over 30 minutes at 0 °C under argon atmosphere. The reaction mixture was warmed to room temperature and stirred overnight.

Purification:

The reaction mixture was quenched by addition of water and the aqueous phase was extracted with ethyl acetate 3x. Followed by flash chromatography ethyl acetate/heptane gradient 95 % ethyl acetate to 50 % over 8 CV on 45 g. Yield: 219 mg; 51 %

Analyses:

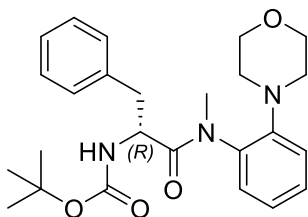
R_f value: 0.45 ethyl acetate/heptane 1:1

¹H NMR (**Spectrum 44**) (400 MHz, Chloroform-*d*) δ 7.08 (td, *J* = 7.7, 1.5 Hz, 1H), 7.02 (dd, *J* = 7.8, 1.5 Hz, 1H), 6.71 (td, *J* = 7.6, 1.4 Hz, 1H), 6.64 (dd, *J* = 8.0, 1.4 Hz, 1H), 4.73 (s, 1H), 3.85 (t, *J* = 4.5 Hz, 4H), 2.92 – 2.88 (m, 4H), 2.87 (s, 3H).

Mass: Calculated *m/z* for C₁₁H₁₇N₂O⁺ [M+H]⁺ = 193.1335; found APCI: [M+H]⁺ 193.0

28

tert-butyl (*R*)-(1-(methyl(2-morpholinophenyl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate



Synthesis:

Quantities: **Pyridine** (5 mL); **EtOAc** (10 mL); **27** (125 mg, 0.65 mmol, 1.0 eq.); **N-Boc-(R)-phenylalanine** (189 µg, 0.72 mmol, 1.1 eq.); **T3P 50 % m/v in EtOAc** (774 µL, 1.30 mmol, 2.0 eq.)

Procedure: **27** and **N-Boc-(R)-phenylalanine** were dissolved in a 1:2 mixture of distilled **pyridine** and **EtOAc**. The mixture was cooled to -20 °C with an isopropanol dry-ice bath before **T3P 50 % m/v in EtOAc** was added. After the addition, the cooling bath was removed, and the mixture was stirred for 20 h at room temperature. Afterwards the mixture was washed with 0.25 M KH₂PO₄ solution three times before the organic phase was evaporated on silica gel under reduced pressure for purification.

Purification:

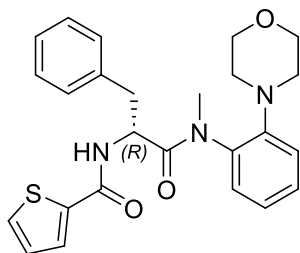
Flash chromatography ethyl acetate/heptane gradient 80 % ethyl acetate to 50 % over 9 CV on 45 g. Yield: 95 mg; 33 %

Analyses:

Mass: Calculated m/z for C₂₅H₃₄N₃O₄⁺ [M+H]⁺ = 440.2544; found APCI: [M+H]⁺ 440.2

29

(R)-N-(1-(methyl(2-morpholinophenyl)amino)-1-oxo-3-phenylpropan-2-yl)thiophene-2-carboxamide



Synthesis:

Quantities: **DCM** (2 mL); **TFA** (2 mL); **28** (95 mg, 0.22 mmol, 1.0 eq.); **thiophene-2-carboxylic acid** (28 mg, 0.22 mmol, 1.0 eq.); **PyBOP** (125 mg, 0.24 mmol, 1.1 eq.); **DCM** (2 mL), **DIPEA** (114 µL, 0.66 mmol, 3.0 eq.)

Procedure: Dissolve **28** in **DCM** and add **TFA** under stirring; stir for 1 h and check for full conversion by TLC; after conversion evaporate the solvents under reduced pressure; followed by coevaporation with toluene 3x, DCM 3x; dissolve the Boc-protected reactant, the **thiophene-2-carboxylic acid** and **PyBOP** in **DMF**; add **DIPEA**; stir over night at 65 °C under argon and light protection.

After the reaction time the crude reaction mixture was directly evaporated on silica gel without previous washing.

Purification:

Flash chromatography ethyl acetate/heptane gradient 80 % heptane to 40 % over 7 CV on 45 g. Yield: 76 mg; 77 %

Analyses:

R_f value: 0.45 ethyl acetate/heptane 4:1

^1H NMR (**Spectrum 45**) (400 MHz, Chloroform-*d*) δ 7.52 (dd, $J = 3.7, 1.0$ Hz, 1H), 7.47 (dd, $J = 5.0, 1.1$ Hz, 1H), 7.24 – 7.18 (m, 4H), 7.11 – 7.02 (m, 2H), 7.02 – 6.93 (m, 3H), 6.86 (td, $J = 7.6, 1.4$ Hz, 1H), 6.18 (dd, $J = 7.8, 1.5$ Hz, 1H), 4.95 – 4.87 (m, 1H), 3.65 – 3.57 (m, 2H), 3.55 – 3.45 (m, 2H), 3.21 (s, 3H), 2.93 – 2.83 (m, 4H), 2.78 – 2.70 (m, 2H).

^{13}C NMR (**Spectrum 46**) (126 MHz, Chloroform-*d*) δ 172.7, 160.4, 148.4, 139.1, 136.5, 136.5, 130.3, 129.7, 128.9, 128.5, 128.1, 128.1, 127.9, 126.9, 124.2, 120.4, 67.1, 52.5, 51.7, 41.0, 37.1.

Mass: Calculated m/z for $\text{C}_{25}\text{H}_{28}\text{N}_3\text{O}_3\text{S}^+$ $[\text{M}+\text{H}]^+ = 450.1846$; $[\text{M}+\text{Na}]^+ = 472.1665$; found APCI: $[\text{M}+\text{H}]^+ 450.1$; found HRMS: $[\text{M}+\text{H}]^+ 450.1850$; $[\text{M}+\text{Na}]^+ 472.1670$

HPLC purity: 96.3 %; $t_{\text{R}} = 2.9$ min (**HPLC-Trace 18**); (Agilent Poroshell 120, EC-C18, 3,0x50 mm, 2,7 μm , acetonitrile/ water 15:85, 6 min, $v = 1.2$ mL/min, $\lambda = 254$ nm)

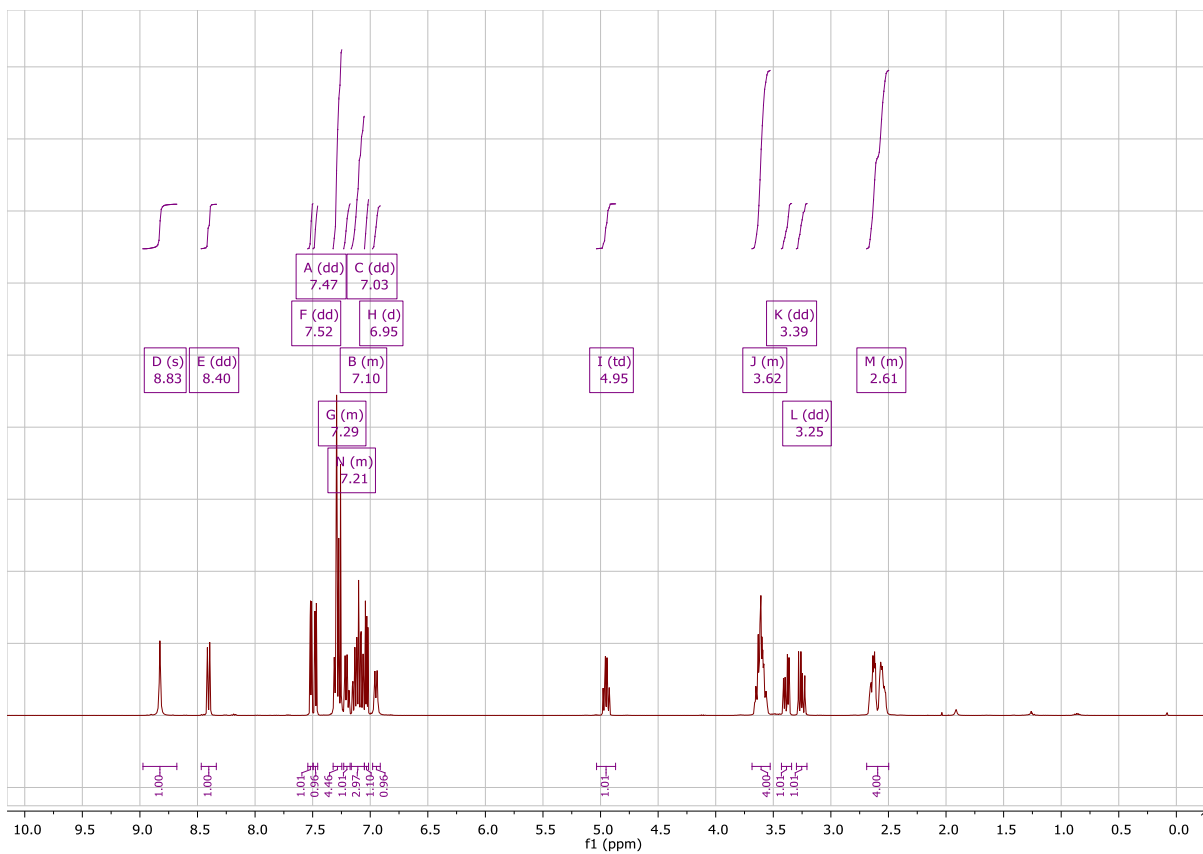
4. NMR spectra and HPLC traces

Table of NMR spectra:

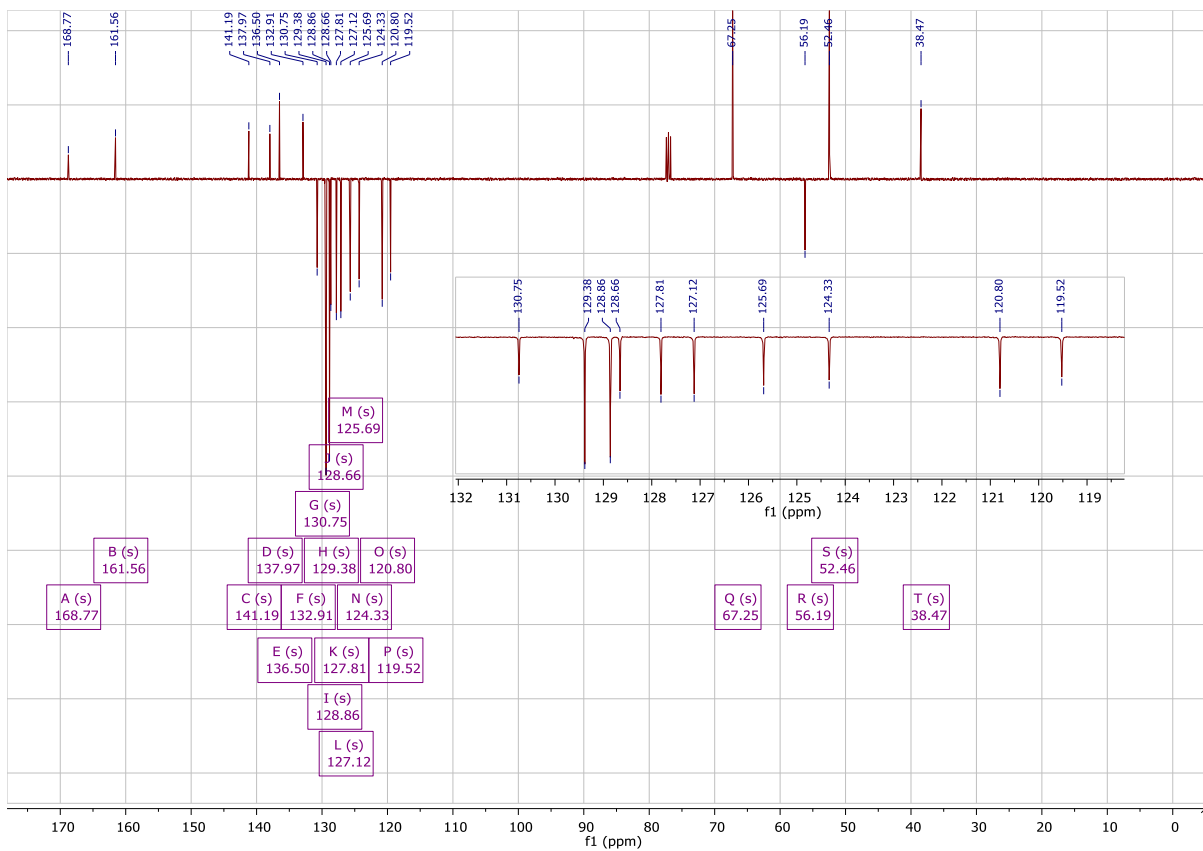
Spectrum 1 ^1H -NMR MMV	41
Spectrum 2 APT ^{13}C -NMR MMV	41
Spectrum 3 ^1H -NMR 1	42
Spectrum 4 ^1H -NMR 2	43
Spectrum 5 APT ^{13}C -NMR 2	43
Spectrum 6 ^1H -NMR 3	44
Spectrum 7 APT ^{13}C -NMR 3	45
Spectrum 8 ^1H -NMR 4	46
Spectrum 9 APT ^{13}C -NMR 4	46
Spectrum 10 ^1H -NMR 5	47
Spectrum 11 APT ^{13}C -NMR 5	48
Spectrum 12 ^1H -NMR 6	49
Spectrum 13 APT ^{13}C -NMR 6	49
Spectrum 14 ^1H -NMR 7	50
Spectrum 15 ^1H -NMR 10	51
Spectrum 16 APT ^{13}C -NMR 10	52
Spectrum 17 ^1H -NMR 11	53
Spectrum 18 APT ^{13}C -NMR 11	53
Spectrum 19 ^1H -NMR 12	54
Spectrum 20 APT ^{13}C -NMR 12	55
Spectrum 21 ^1H -NMR 13	56
Spectrum 22 APT ^{13}C -NMR 13	56
Spectrum 23 ^1H -NMR 14	57
Spectrum 24 APT ^{13}C -NMR 14	58
Spectrum 25 ^1H -NMR 15	59
Spectrum 26 ^1H -NMR 16	59
Spectrum 27 ^1H -NMR 18	60
Spectrum 28 ^1H -NMR 19	60
Spectrum 29 APT ^{13}C -NMR 19	61
Spectrum 30 ^1H -NMR 20	62
Spectrum 31 APT ^{13}C -NMR 20	62
Spectrum 32 ^1H -NMR 21	63
Spectrum 33 APT ^{13}C -NMR 21	64
Spectrum 34 ^1H -NMR 22	65
Spectrum 35 ^{13}C -NMR 22	65
Spectrum 36 ^1H -NMR 23	66
Spectrum 37 ^{13}C -NMR 23	67
Spectrum 38 ^1H -NMR 24	68
Spectrum 39 ^1H -NMR 25	68
Spectrum 40 ^{13}C -NMR 25	69
Spectrum 41 HSQC 25	69
Spectrum 42 ^1H -NMR 26	70
Spectrum 43 ^1H -NMR 27	71
Spectrum 44 ^1H -NMR 29	71
Spectrum 45 APT ^{13}C -NMR 29	72

Table of HPLC-Traces

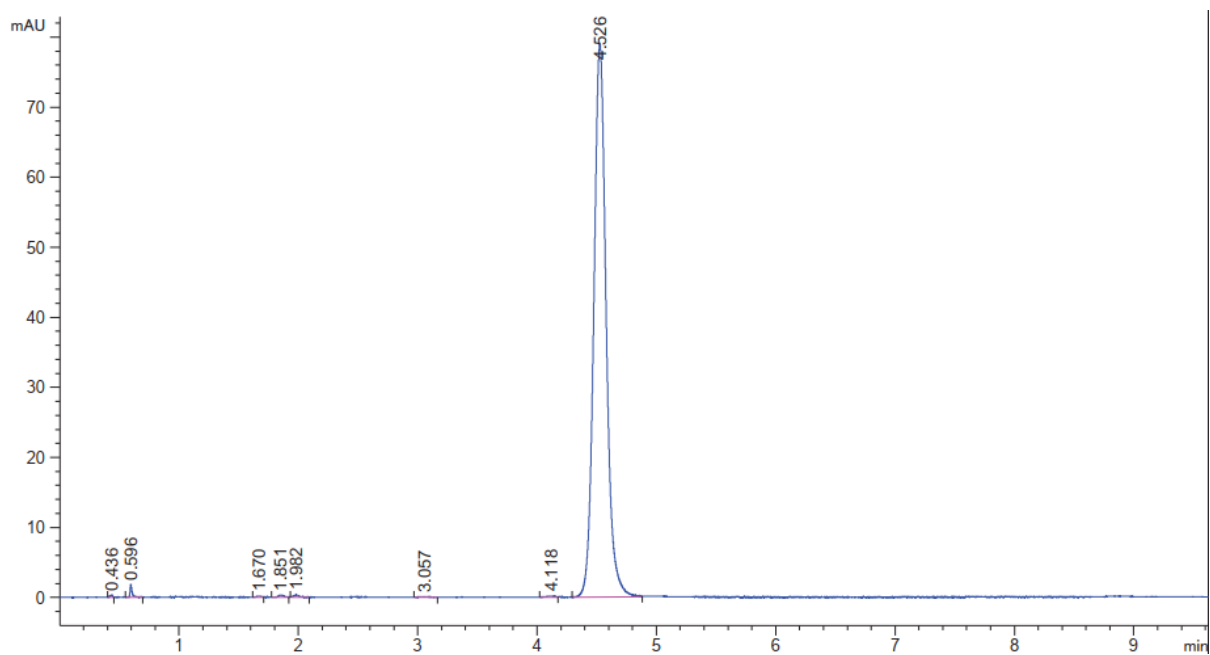
HPLC-Trace 1 HPLC-purity MMV	42
HPLC-Trace 2 HPLC-purity 2	44
HPLC-Trace 3 HPLC-purity 3	45
HPLC-Trace 4 HPLC-purity 4	47
HPLC-Trace 5 HPLC-purity 5	48
HPLC-Trace 6 HPLC-purity 6	50
HPLC-Trace 7 HPLC-purity 10	52
HPLC-Trace 8 HPLC-purity 11	54
HPLC-Trace 9 HPLC-purity 12	55
HPLC-Trace 10 HPLC-purity 13	57
HPLC-Trace 11 HPLC-purity 14	58
HPLC-Trace 12 HPLC-purity 19	61
HPLC-Trace 13 HPLC-purity 20	63
HPLC-Trace 14 HPLC-purity 21	64
HPLC-Trace 15 HPLC-purity 22	66
HPLC-Trace 16 HPLC-purity 23	67
HPLC-Trace 17 HPLC-purity 25	70
HPLC-Trace 18 HPLC-purity 29	72



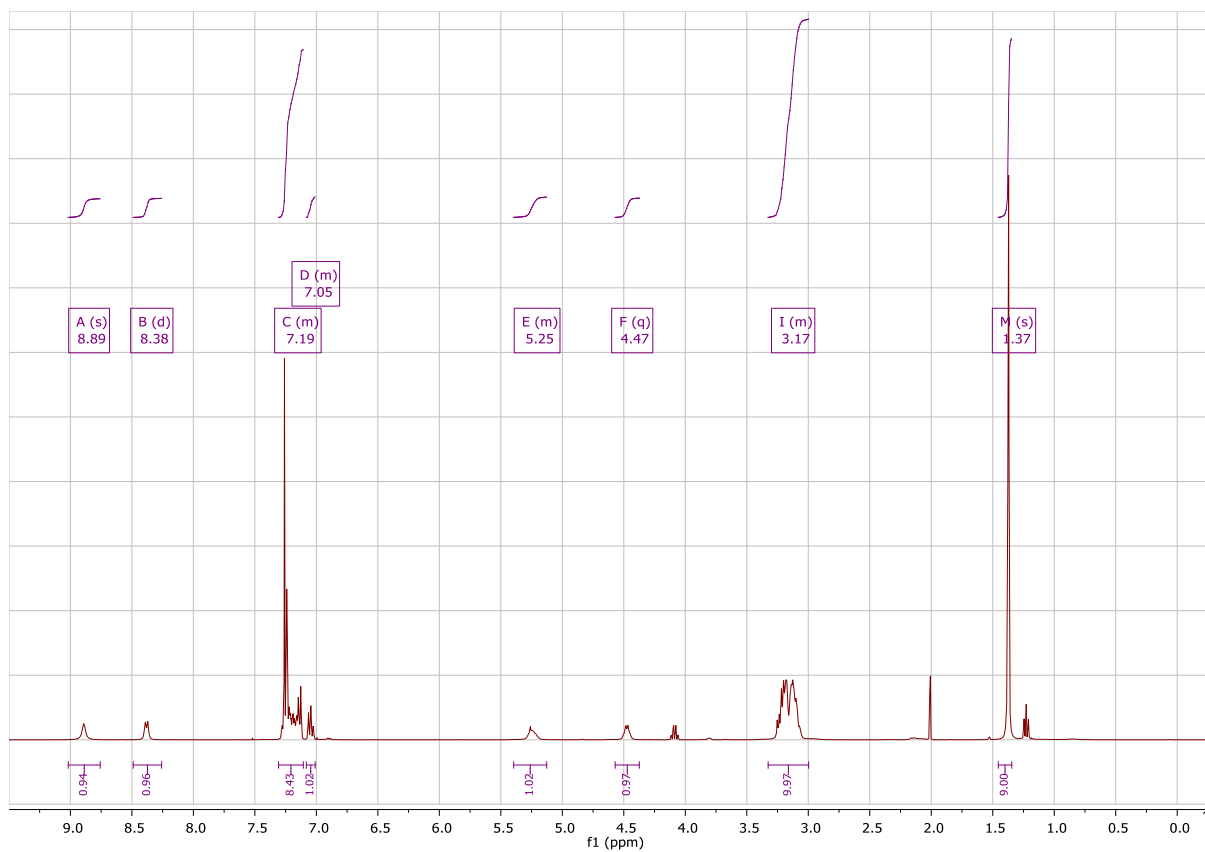
Spectrum 1 ¹H-NMR MMV



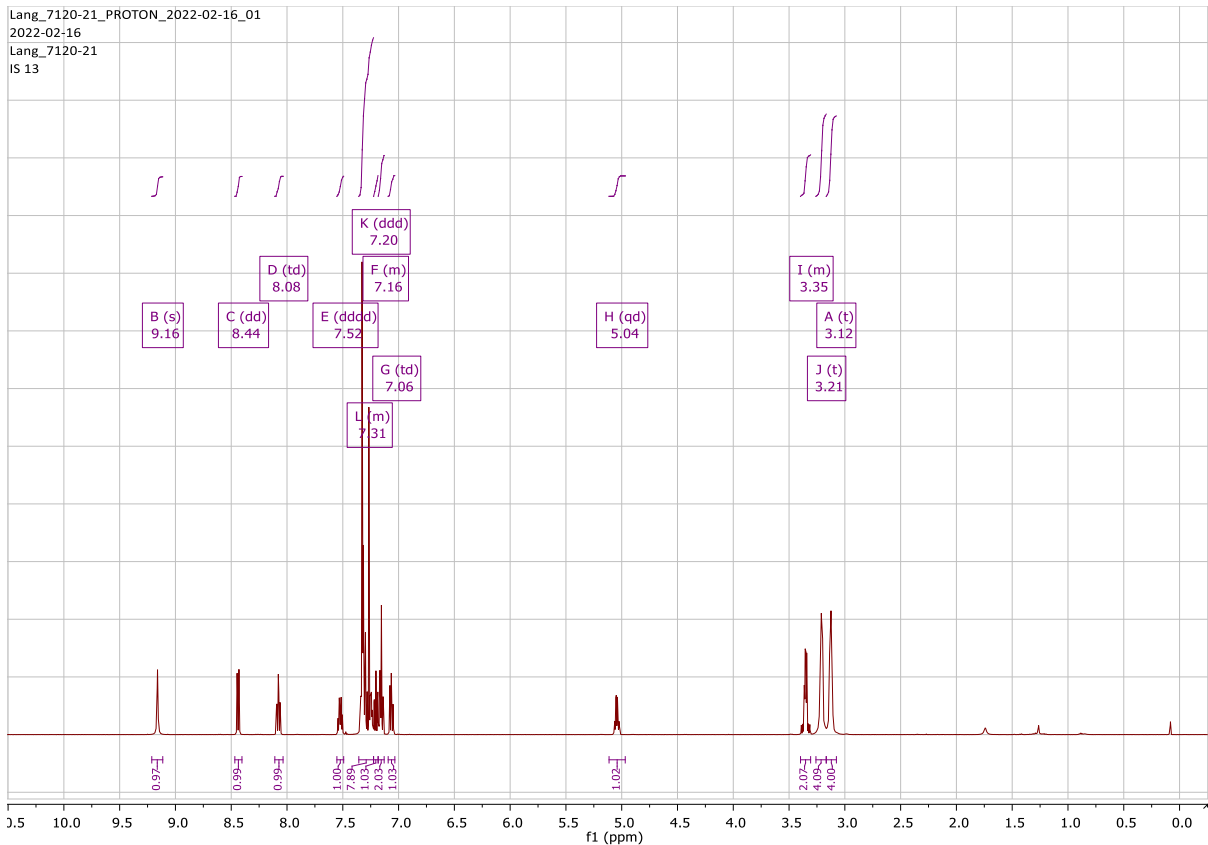
Spectrum 2 APT ¹³C-NMR MMV



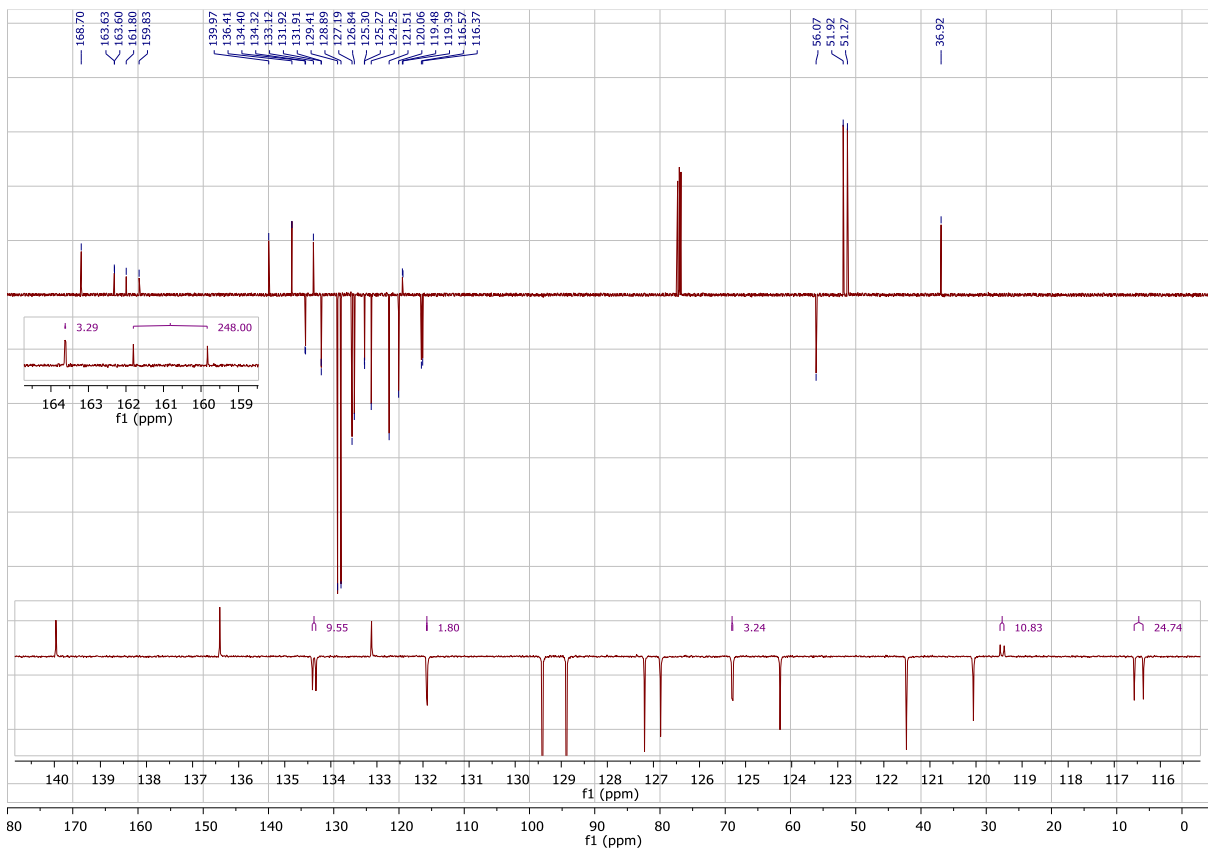
HPLC-Trace 1 HPLC-purity MMV



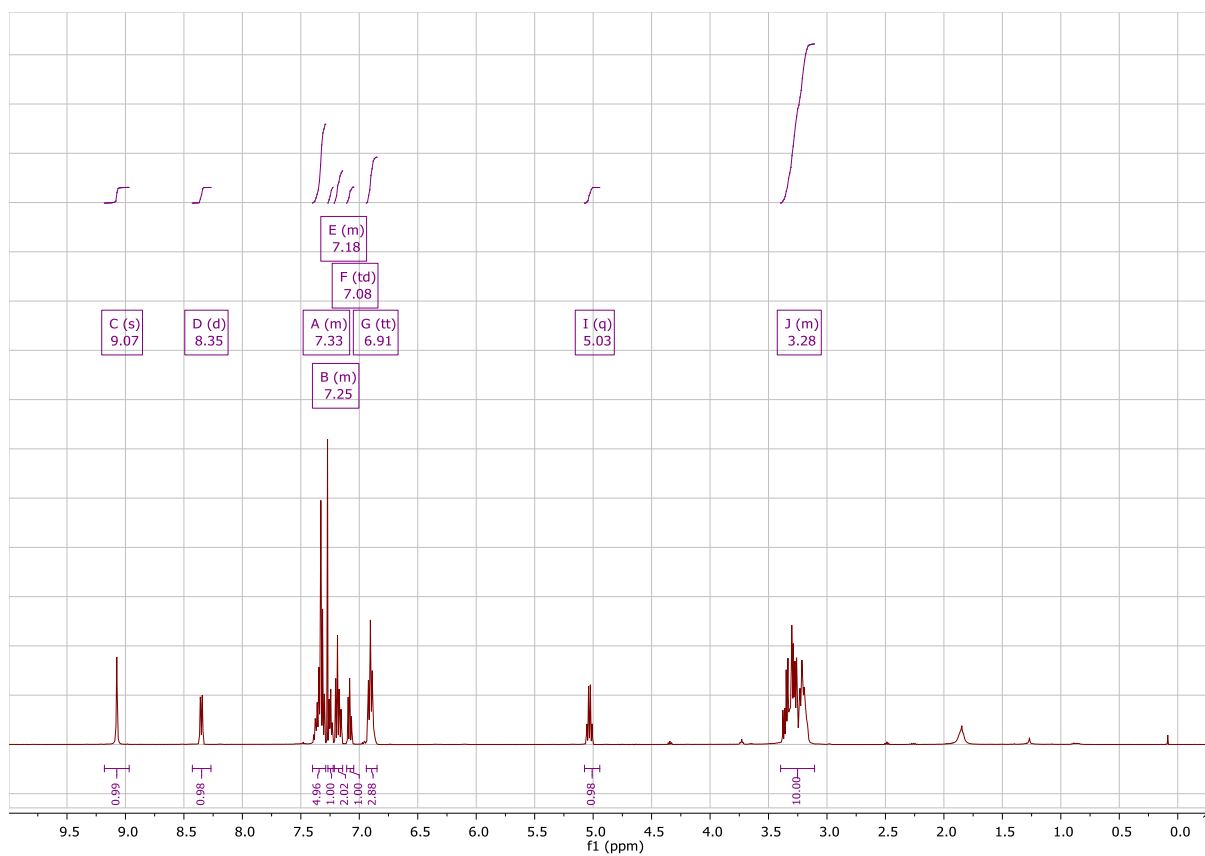
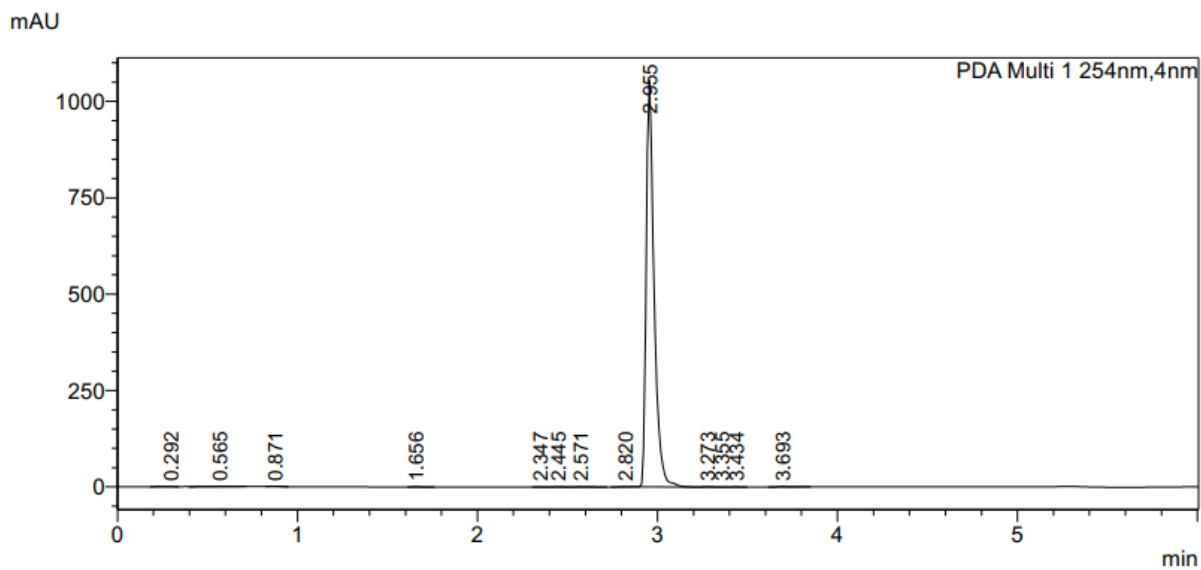
Spectrum 3 ¹H-NMR 1

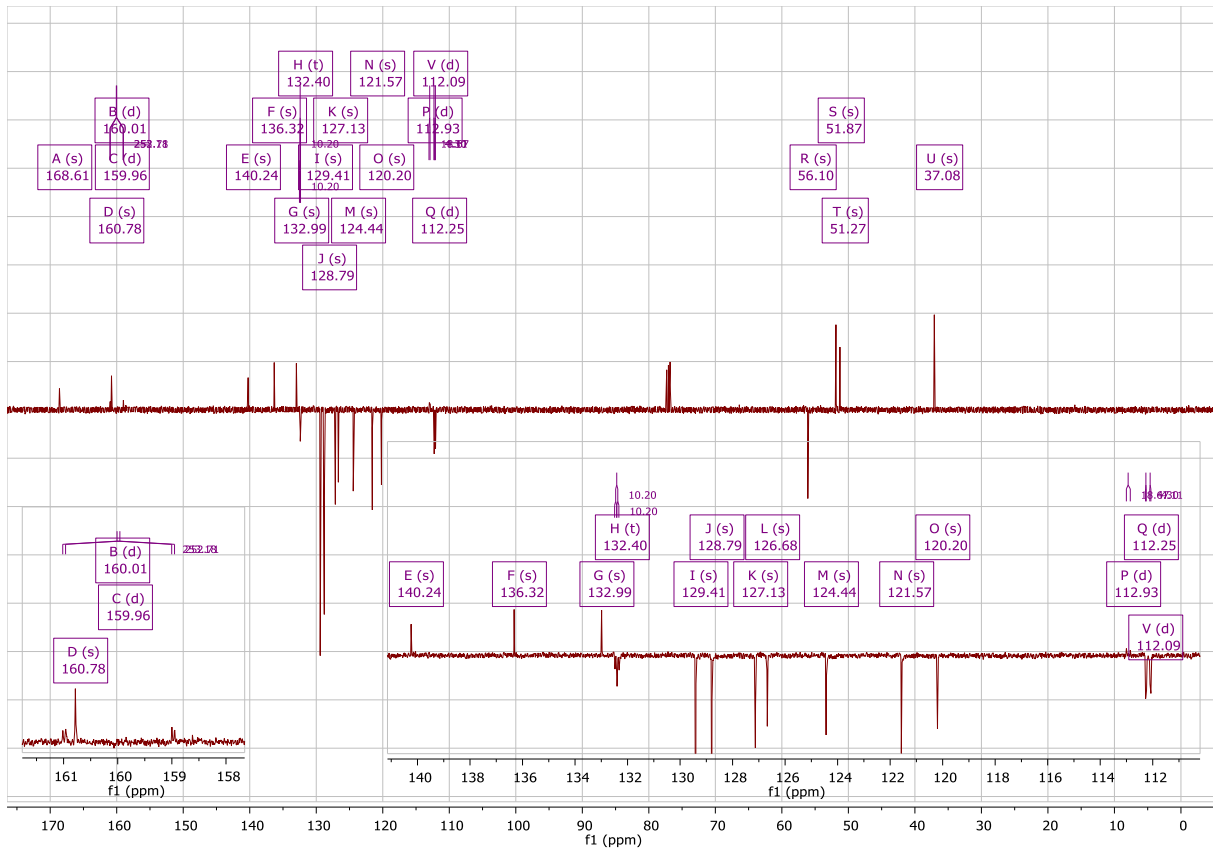


Spectrum 4 ¹H-NMR 2



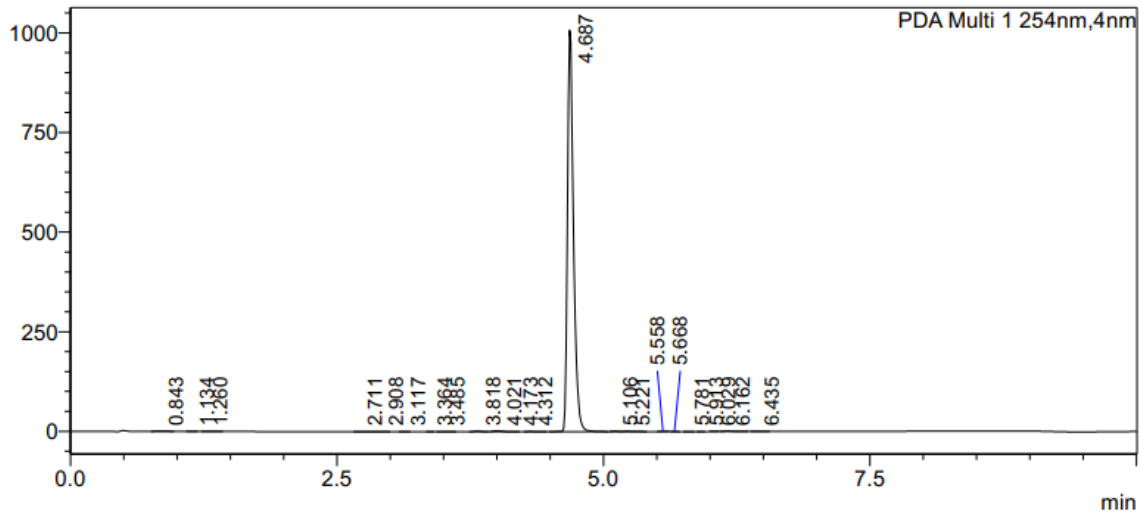
Spectrum 5 APT ¹³C-NMR 2



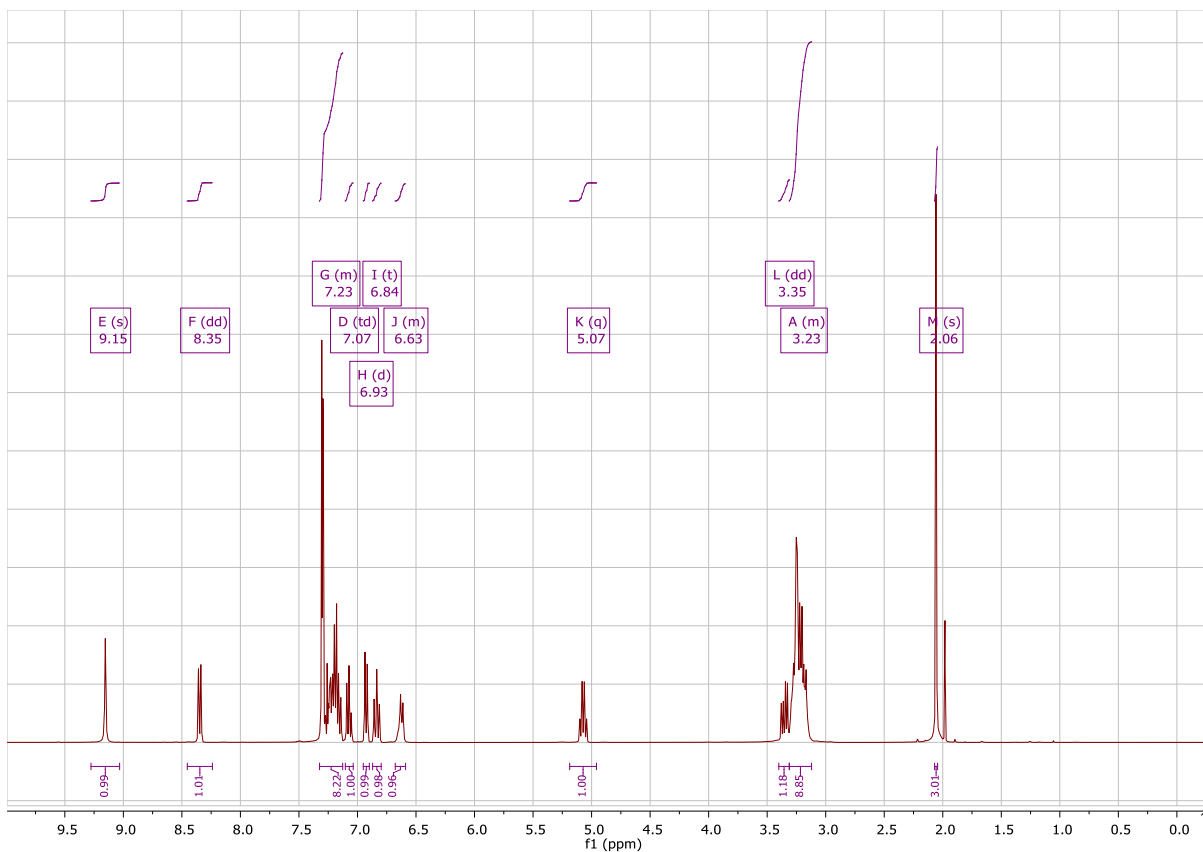


Spectrum 7 APT ¹³C-NMR 3

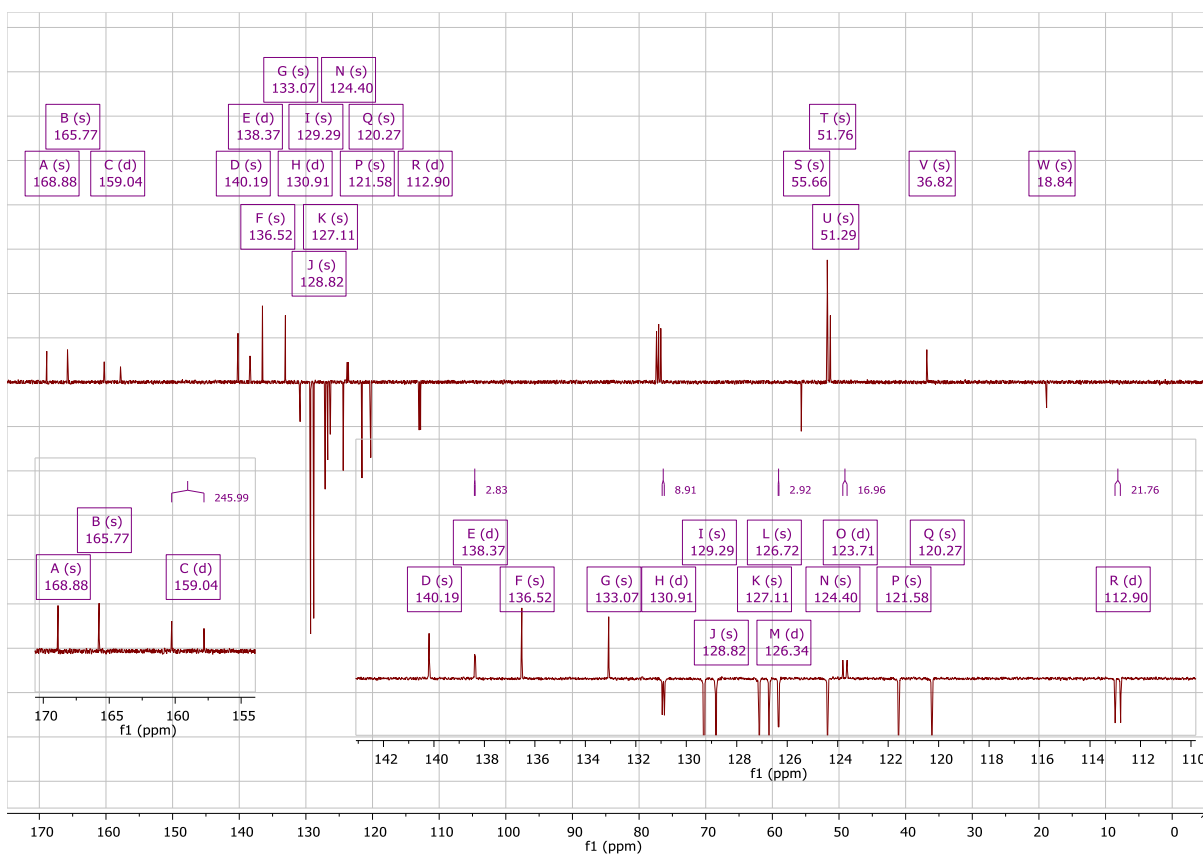
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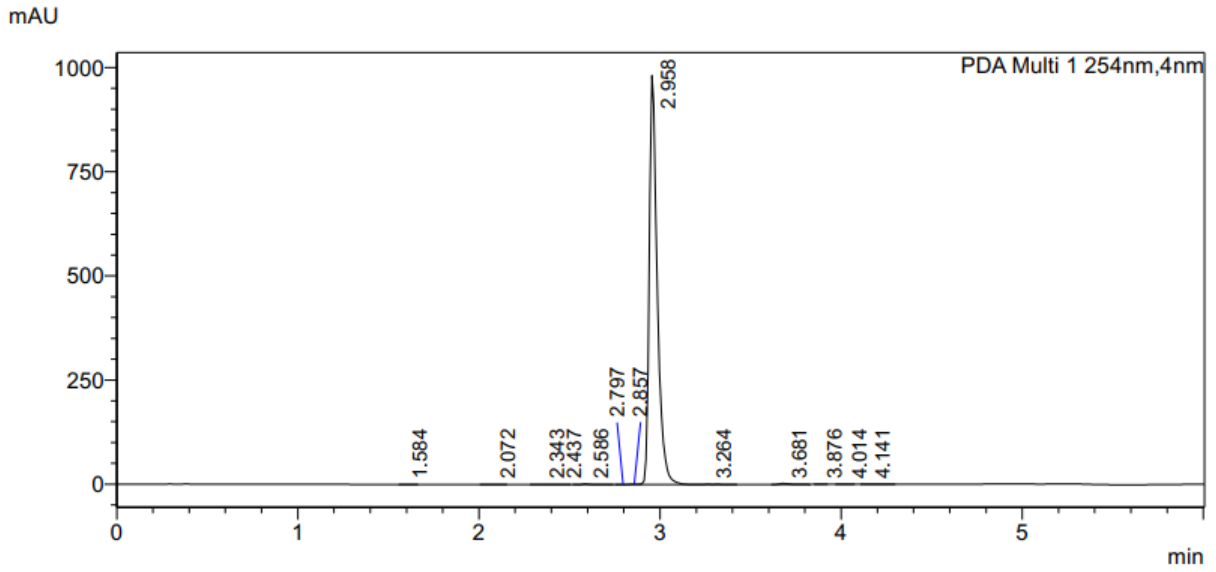
HPLC-Trace 3 HPLC-purity 3



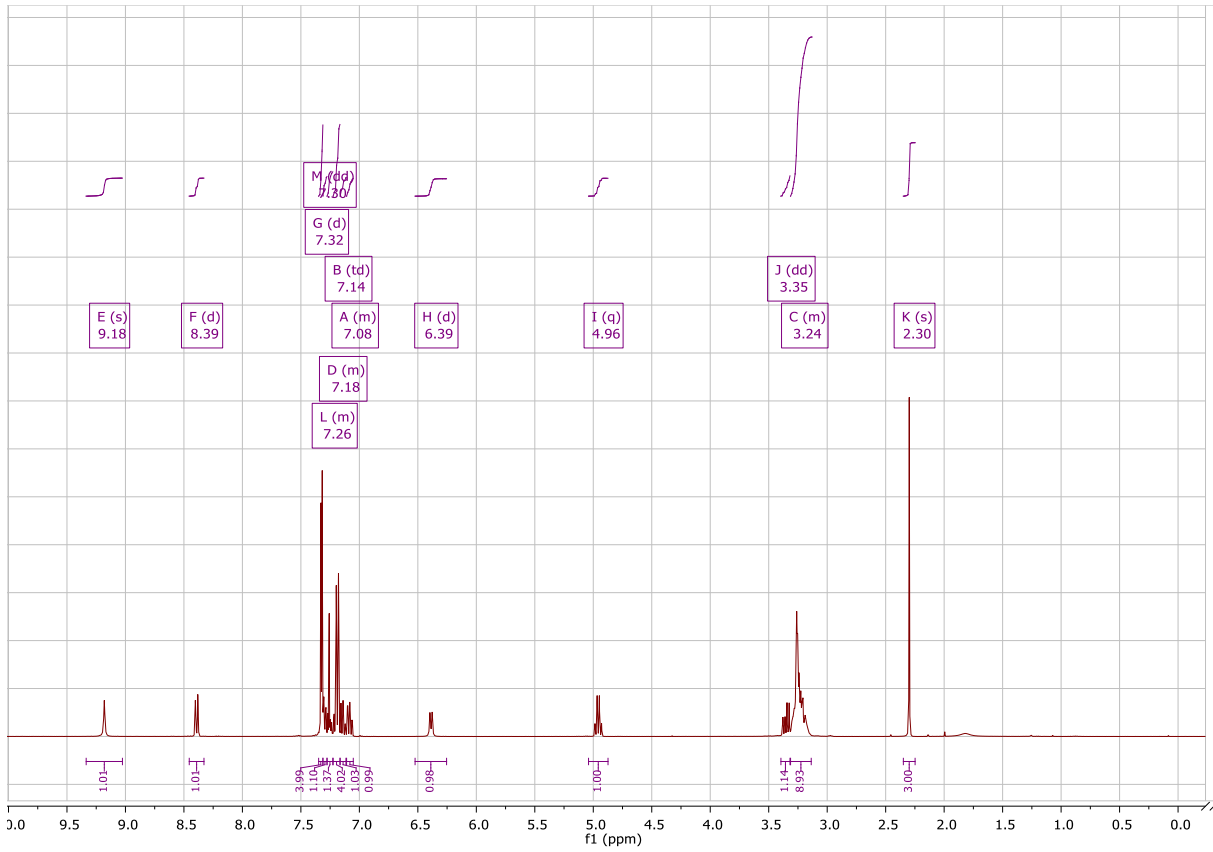
Spectrum 8 ¹H-NMR 4



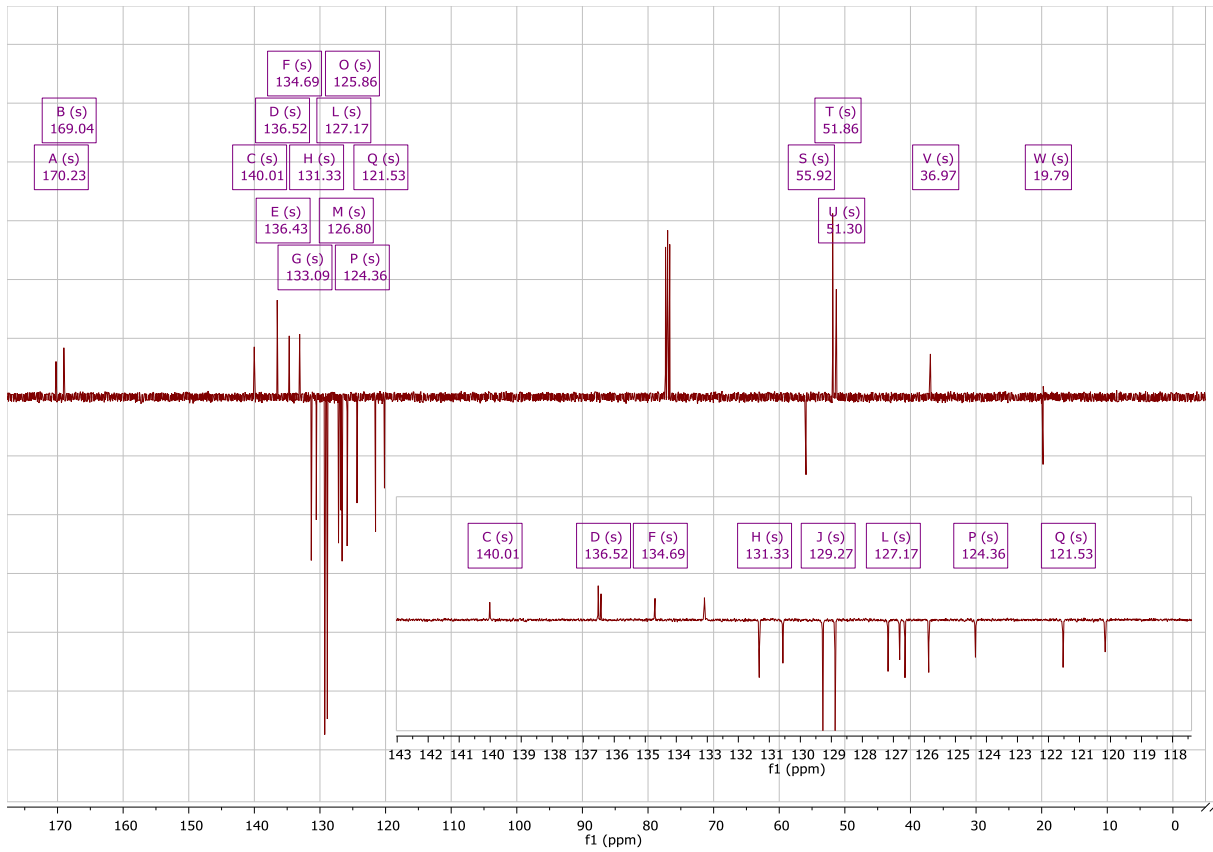
Spectrum 9 APT ¹³C-NMR 4



HPLC-Trace 4 HPLC-purity 4

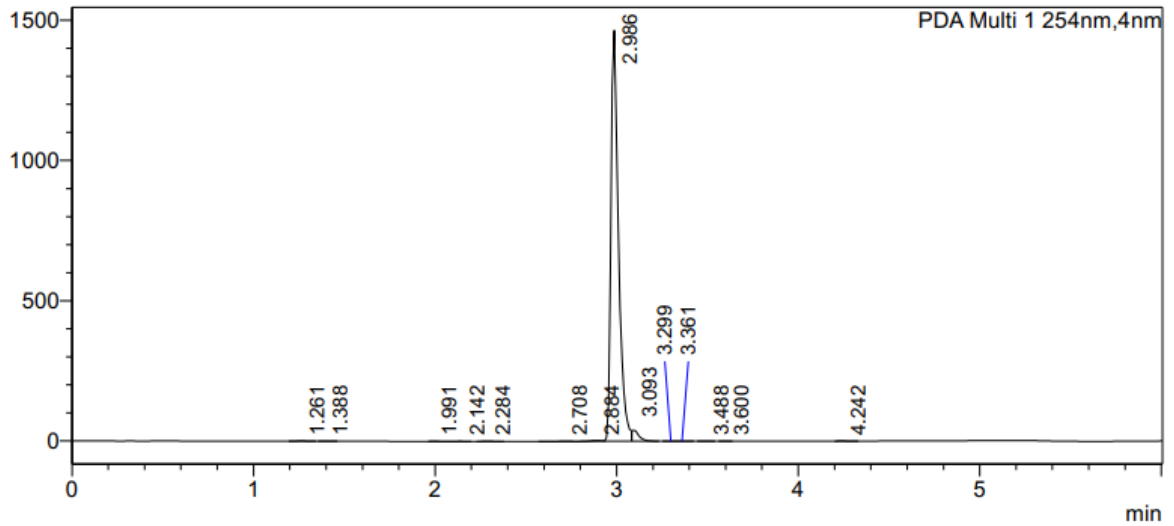


Spectrum 10 ¹H-NMR 5

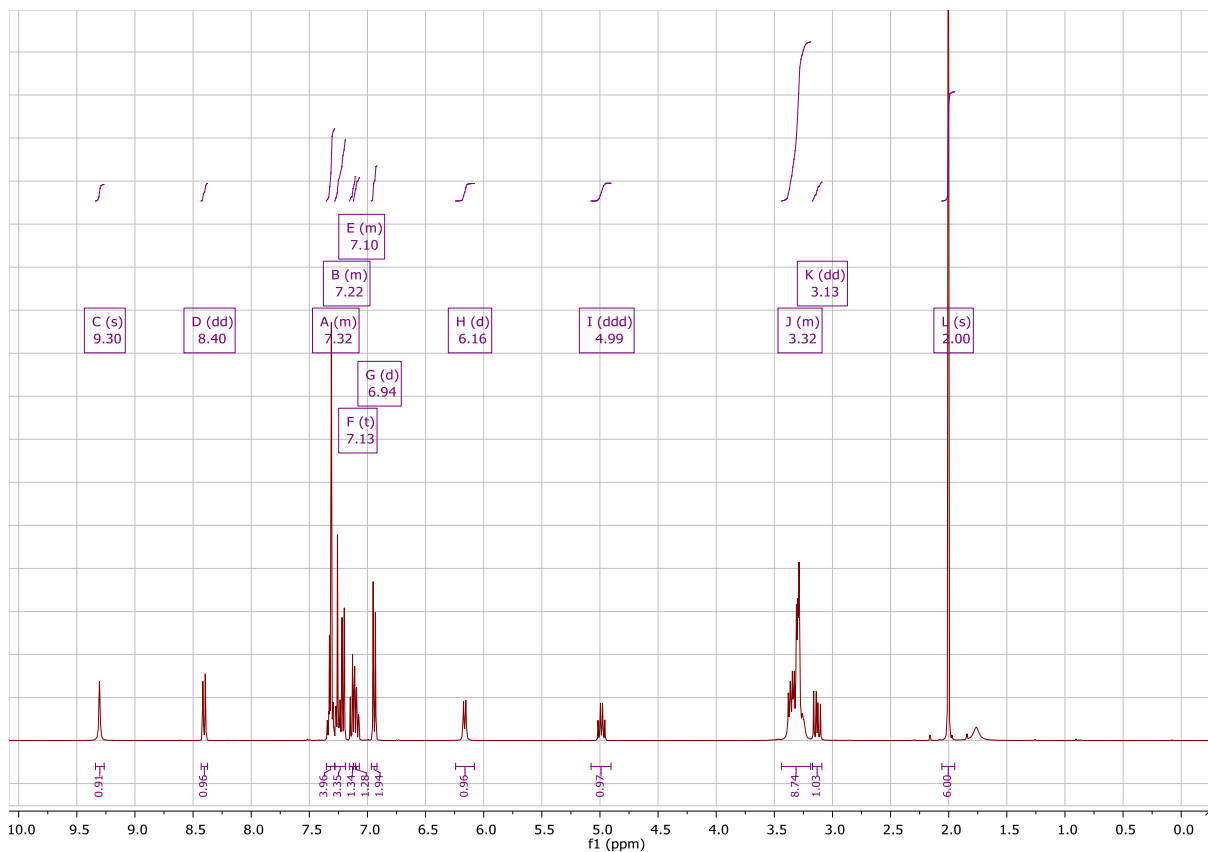


Spectrum 11 APT ¹³C-NMR 5

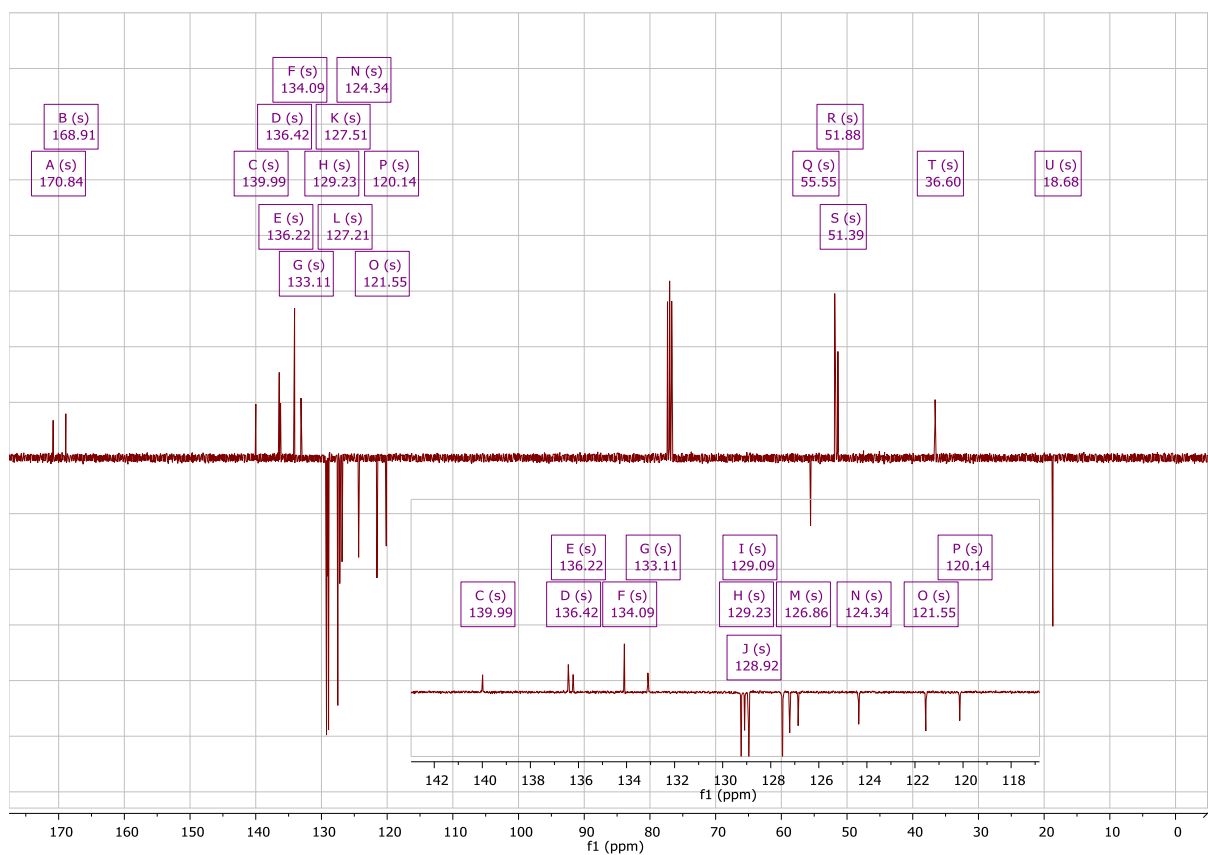
mAU



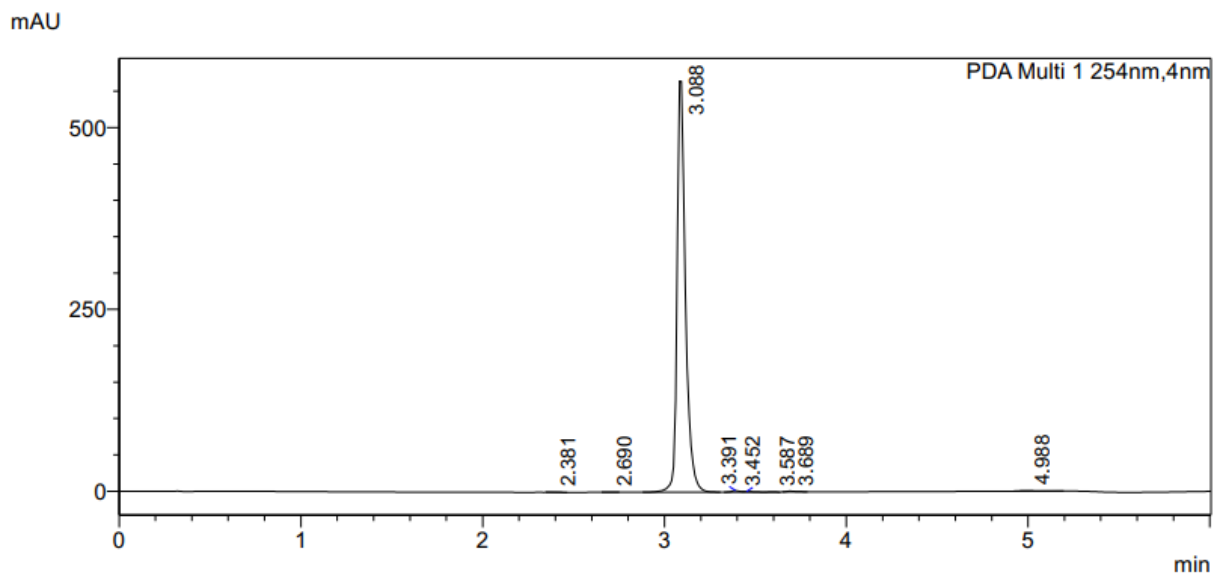
HPLC-Trace 5 HPLC-purity 5



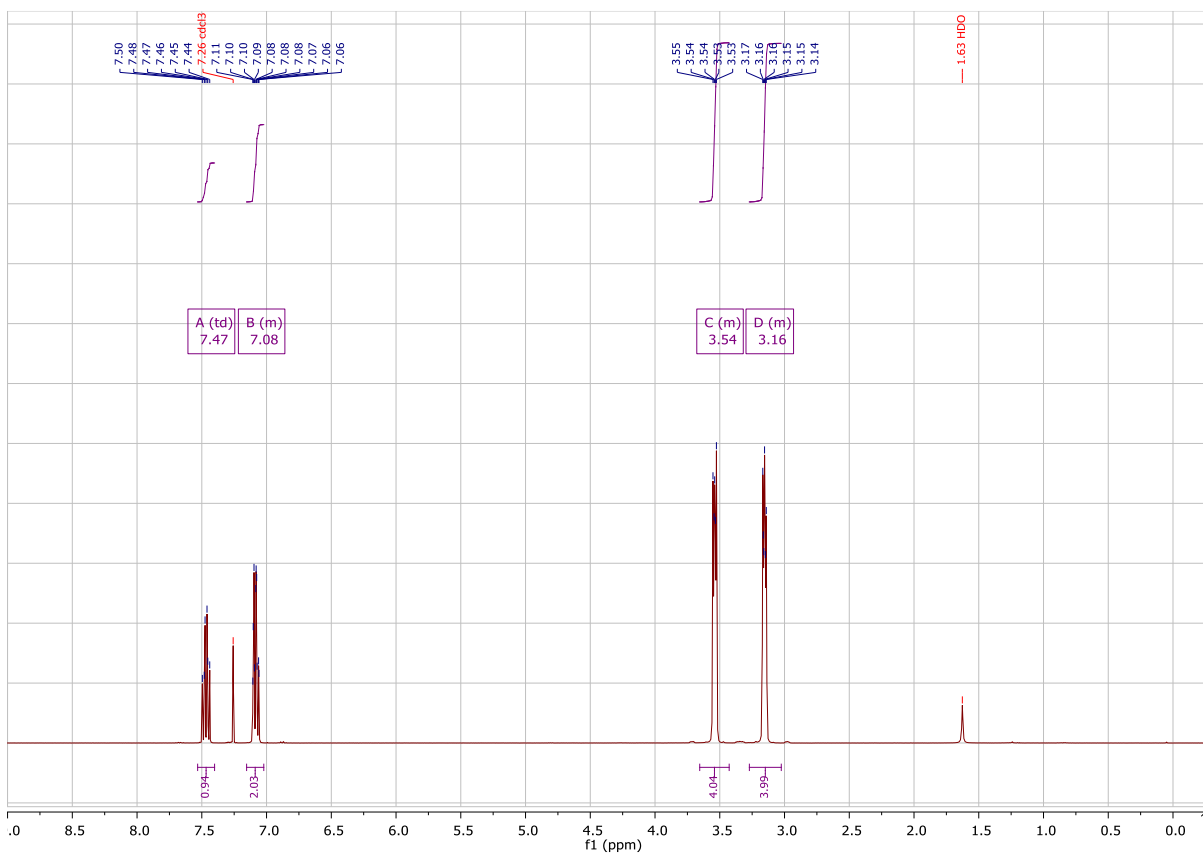
Spectrum 12 $^1\text{H-NMR}$ 6



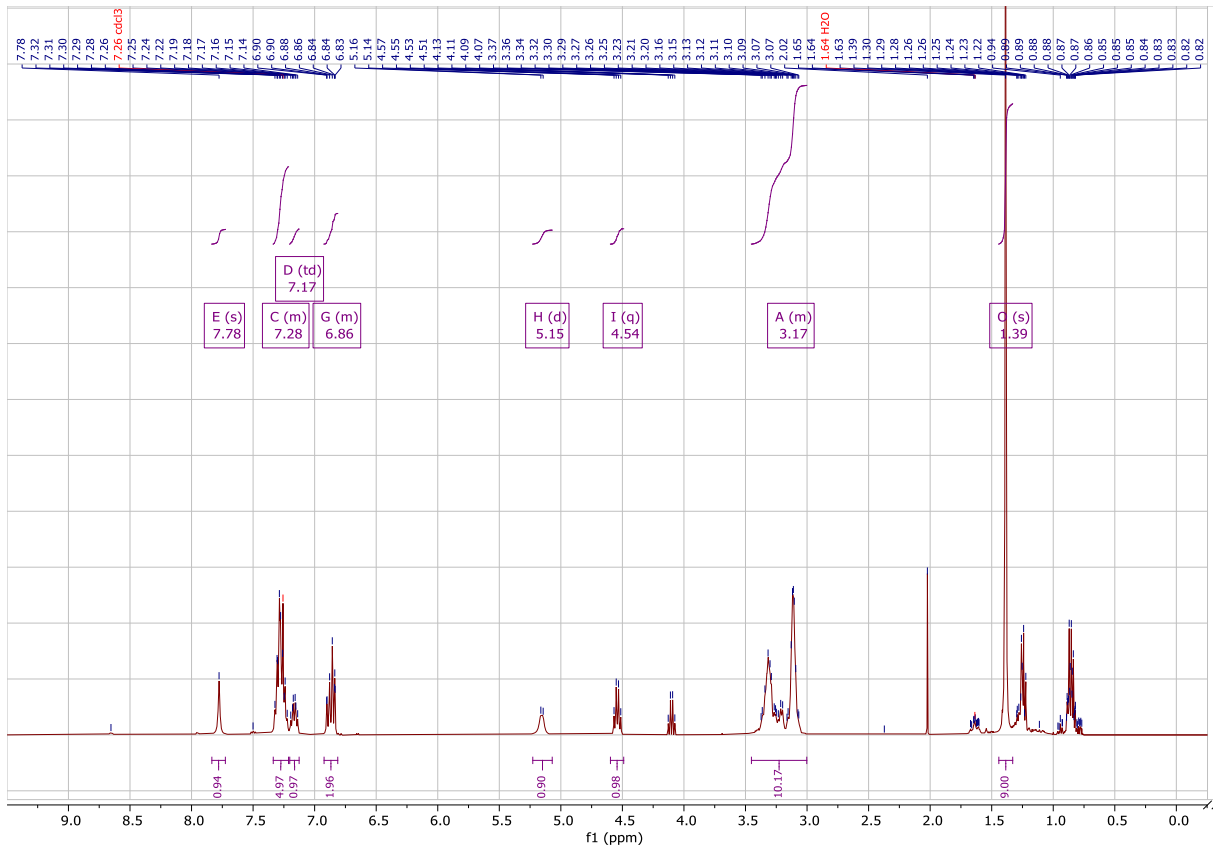
Spectrum 13 APT $^{13}\text{C-NMR}$ 6



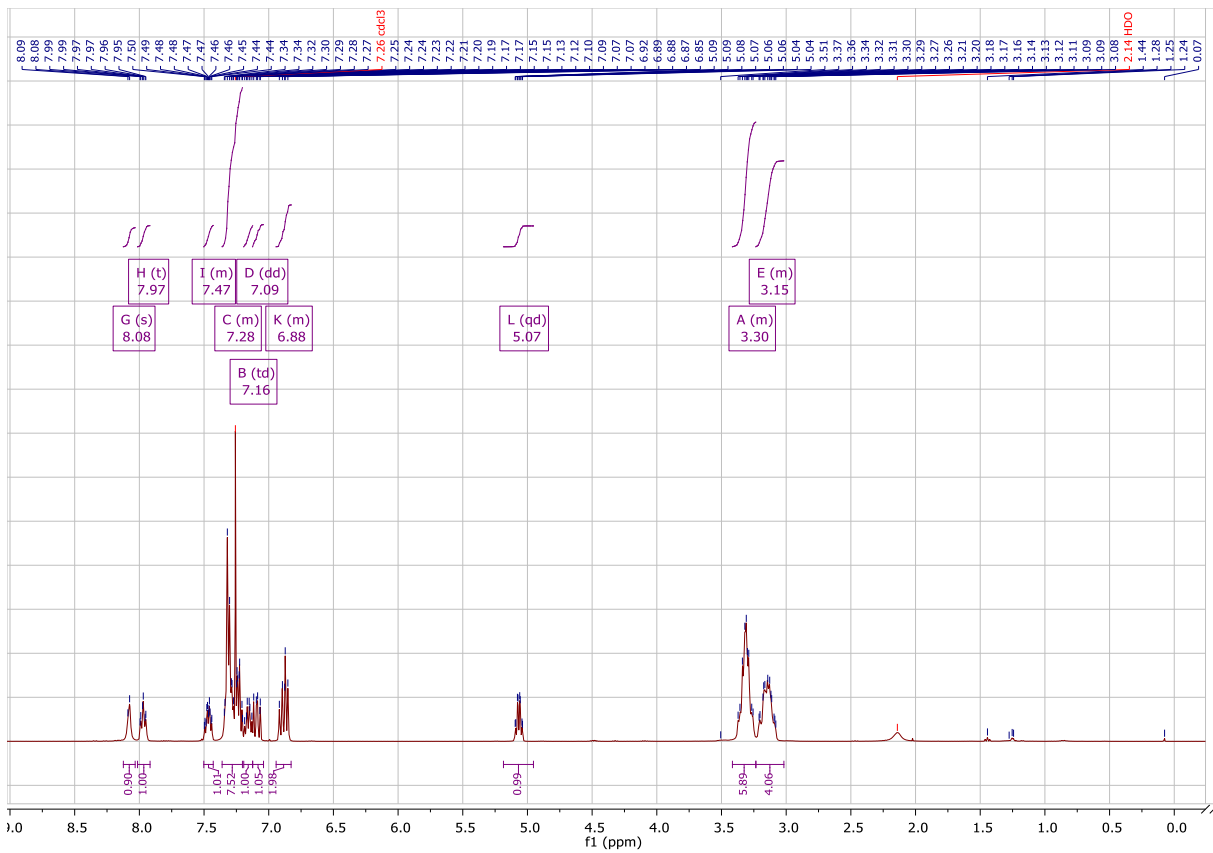
HPLC-Trace 6 HPLC-purity 6



Spectrum 14 ¹H-NMR 7



Spectrum 15 $^1\text{H-NMR}$ 9

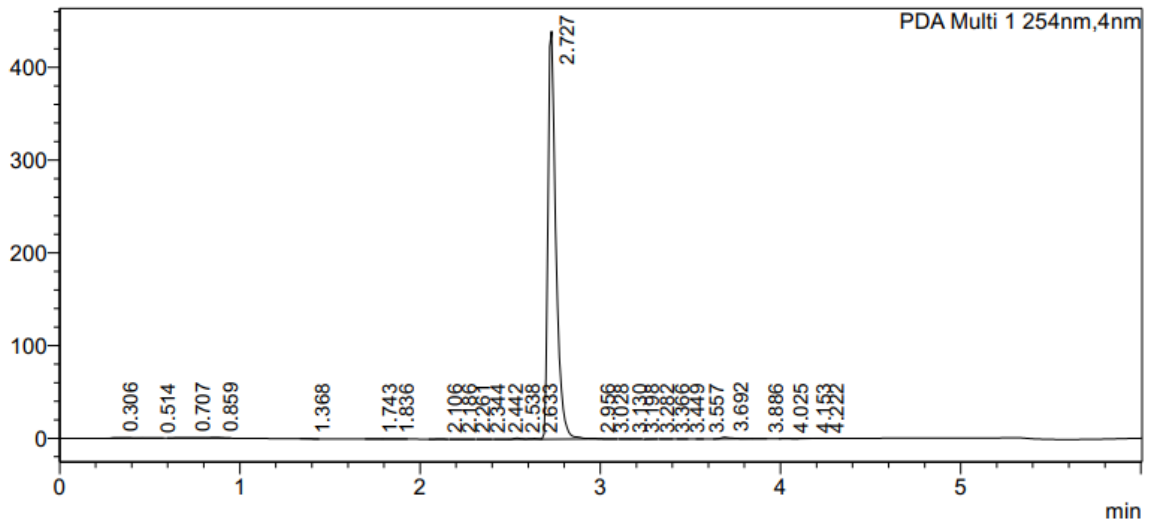


Spectrum 16 $^1\text{H-NMR}$ 10

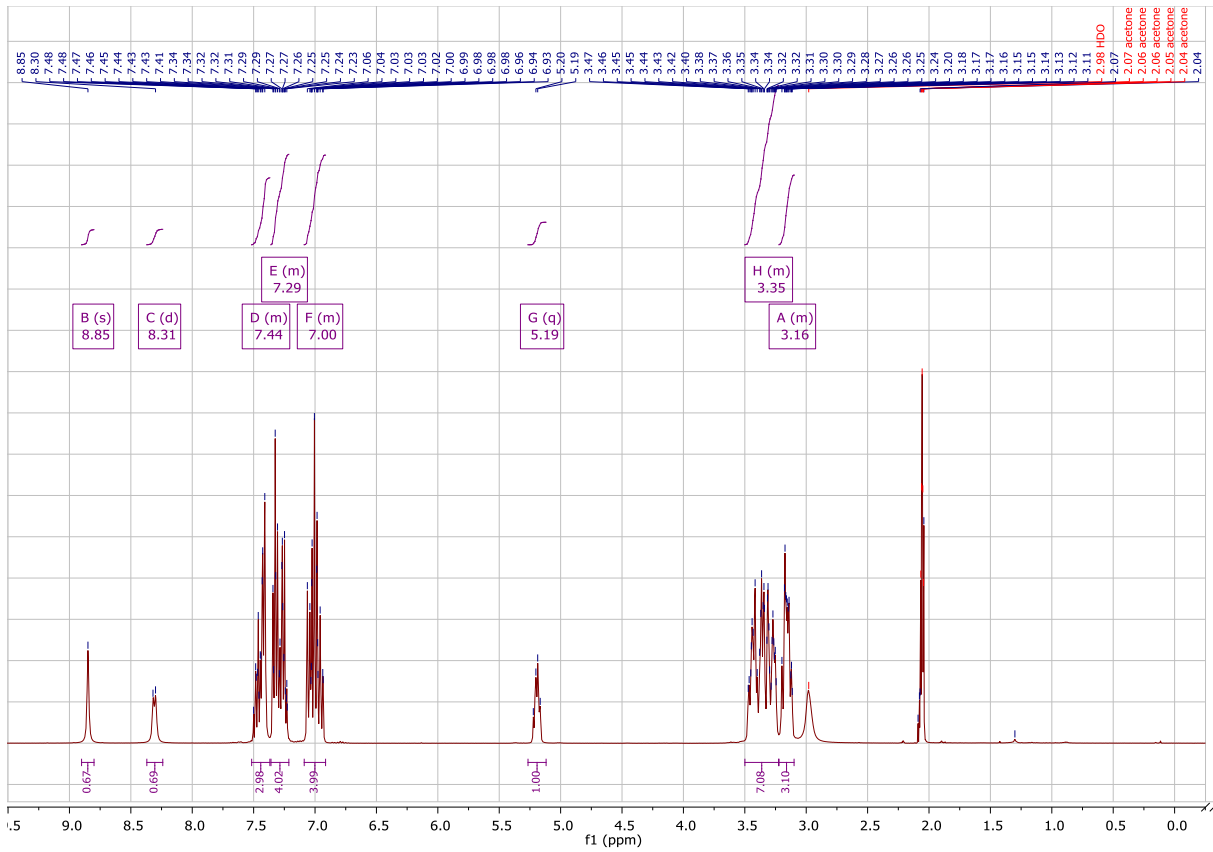


Spectrum 17 APT ¹³C-NMR 10

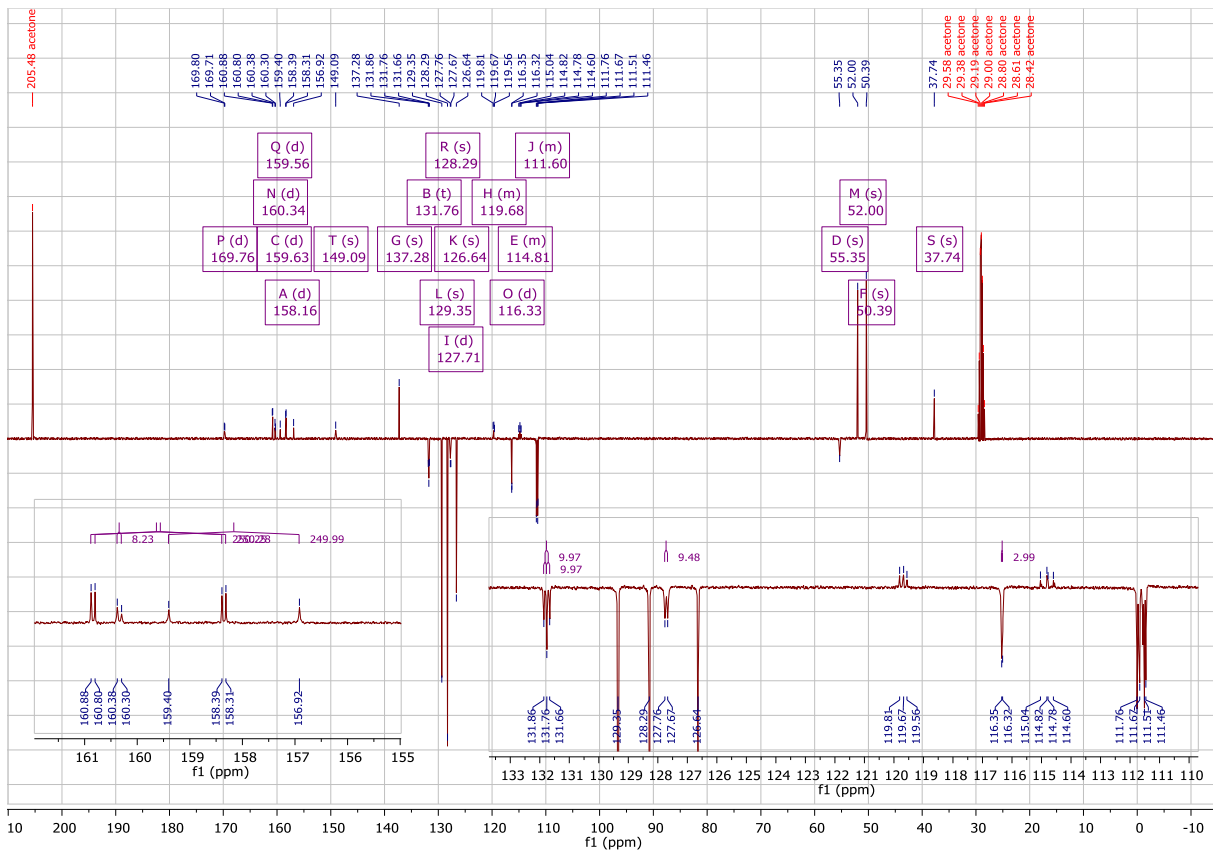
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HPLC-Trace 7 HPLC-purity 10

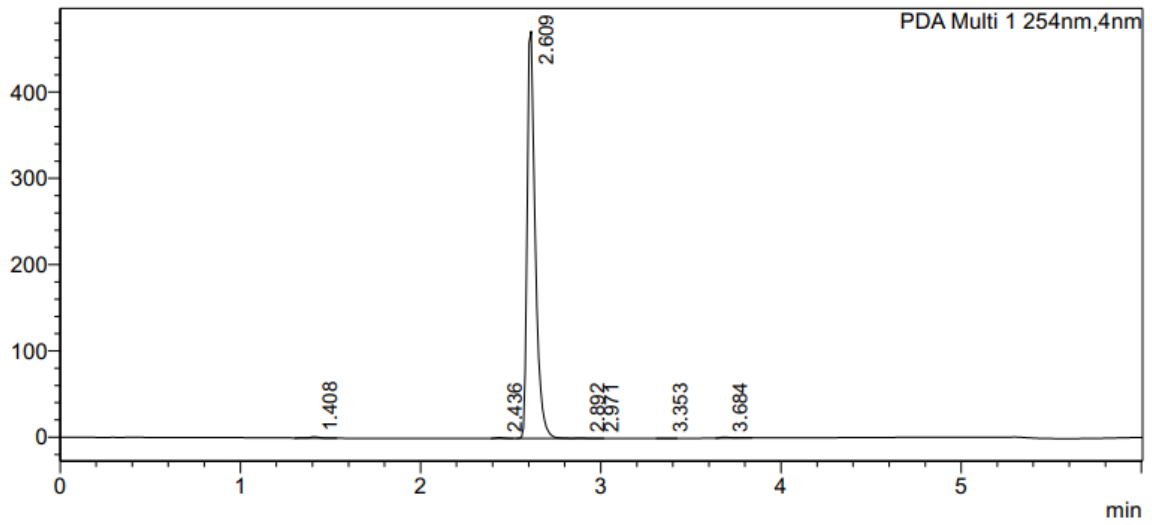


Spectrum 18 ¹H-NMR 11

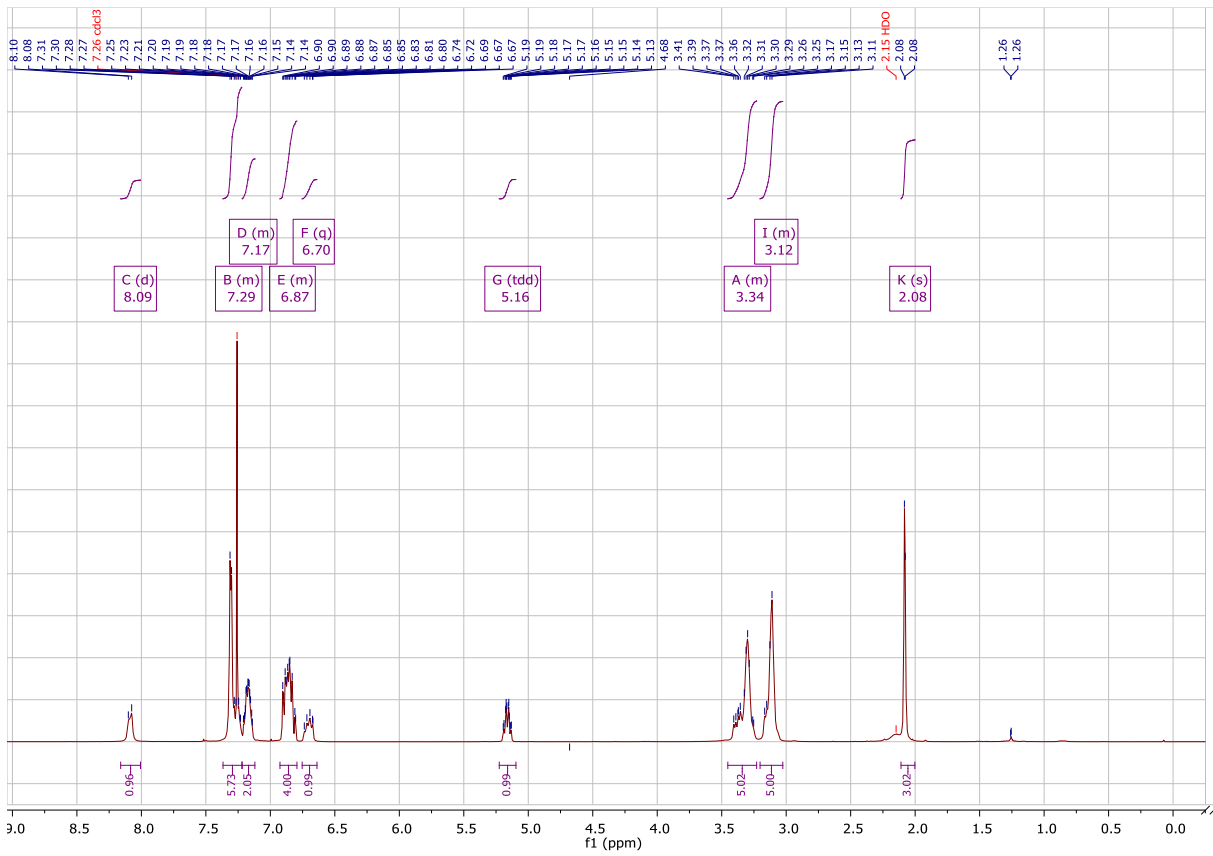


Spectrum 19 APT ¹³C-NMR 11

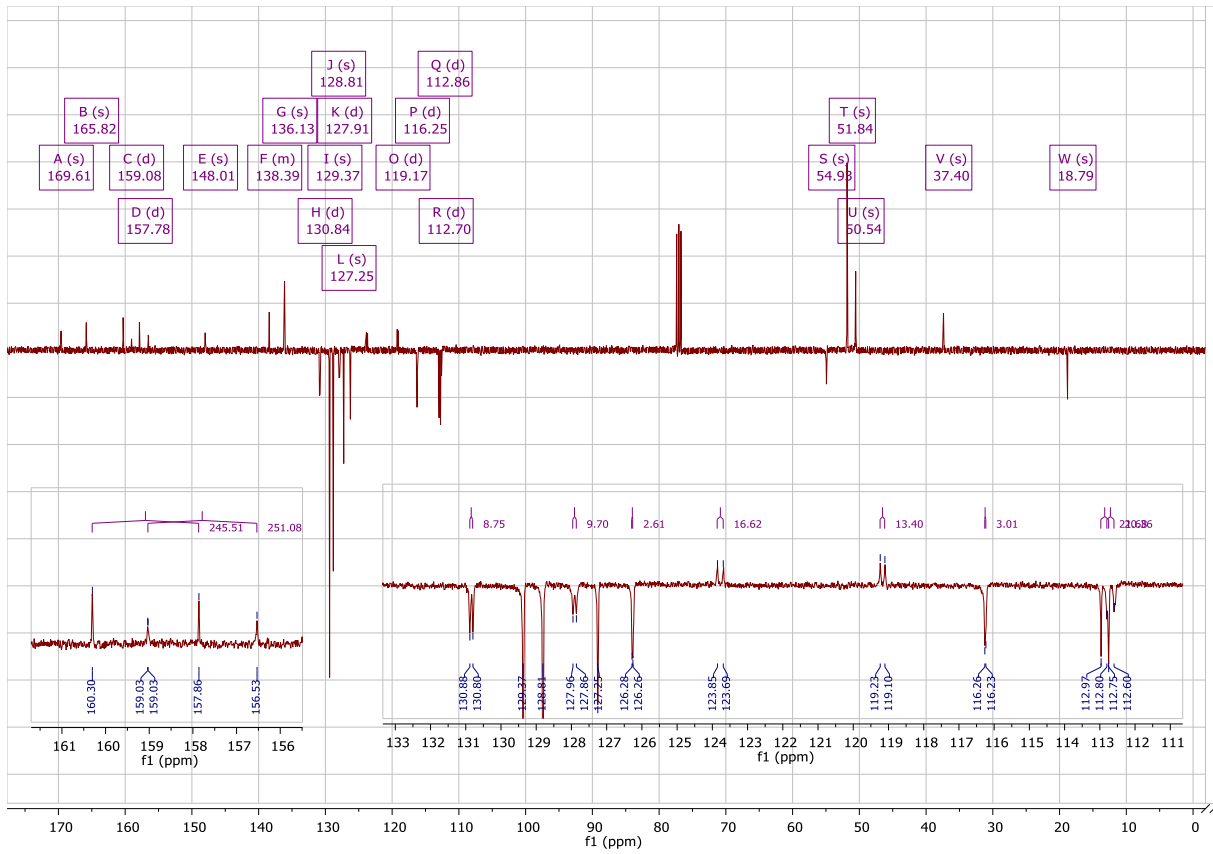
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HPLC-Trace 8 HPLC-purity 11

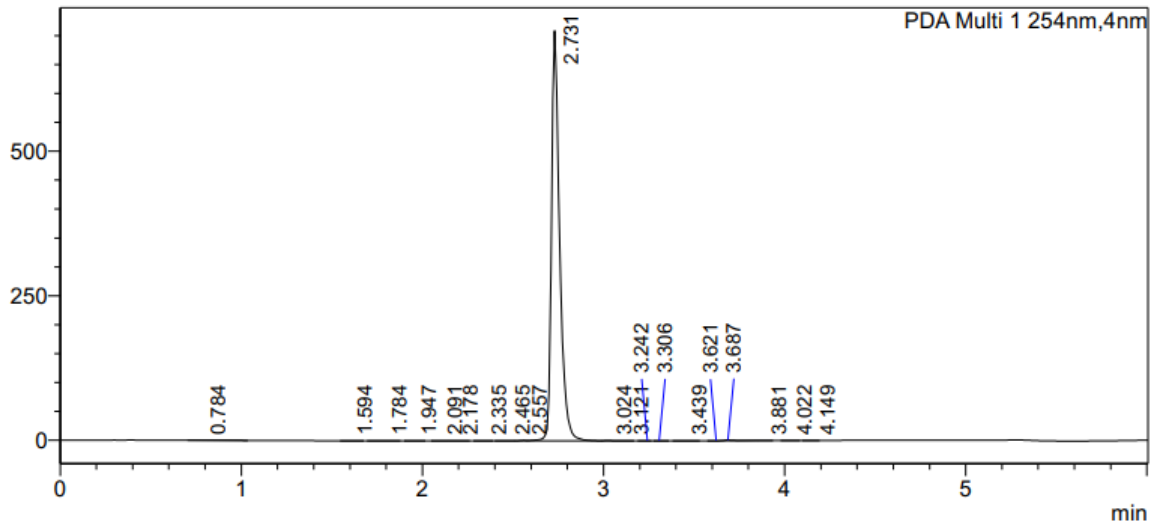


Spectrum 20 ¹H-NMR 12

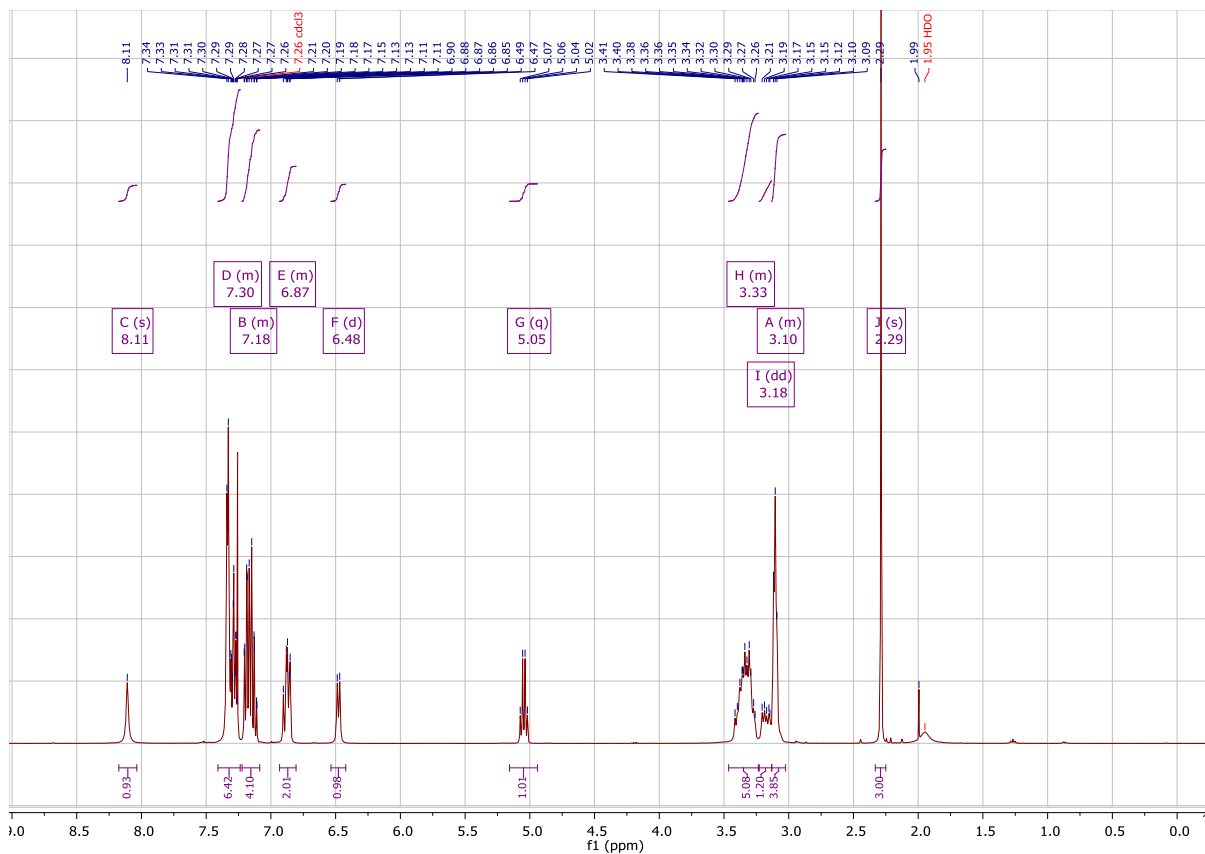


Spectrum 21 APT ¹³C-NMR 12

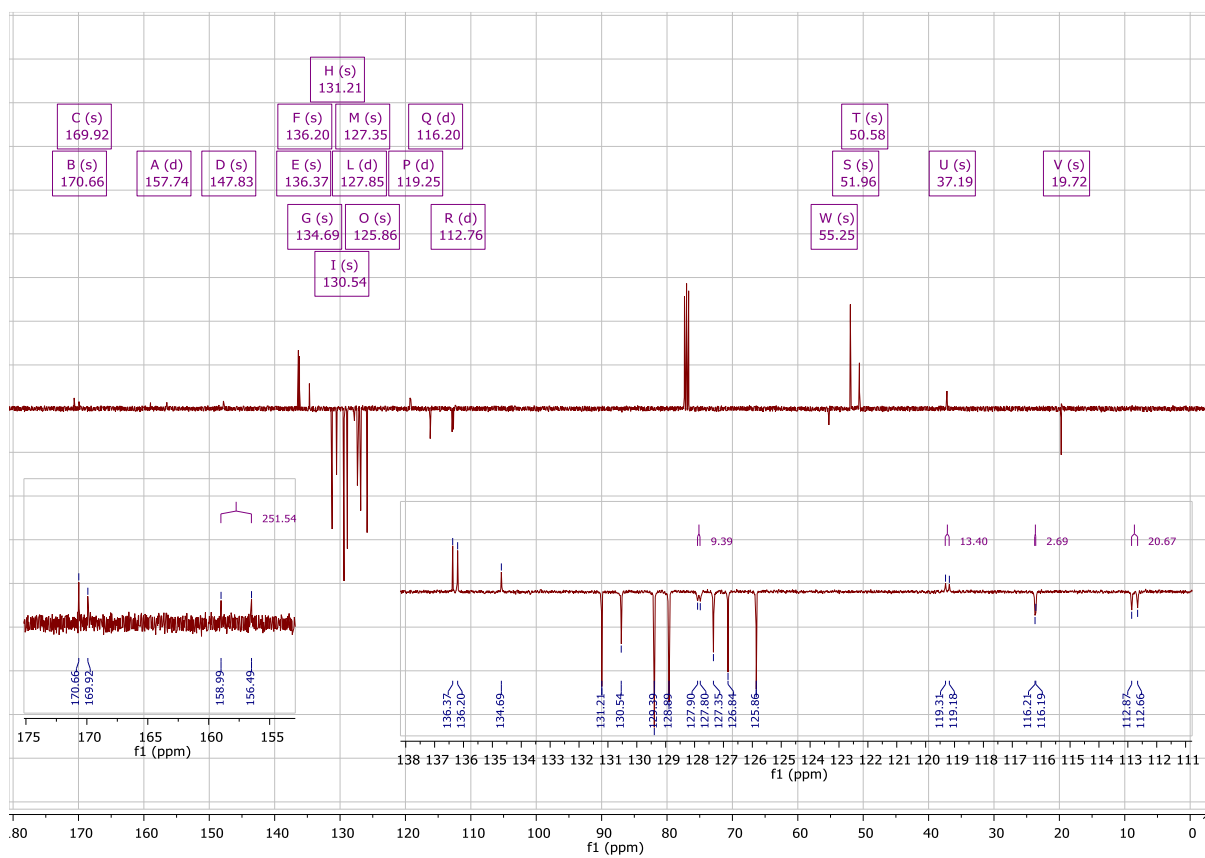
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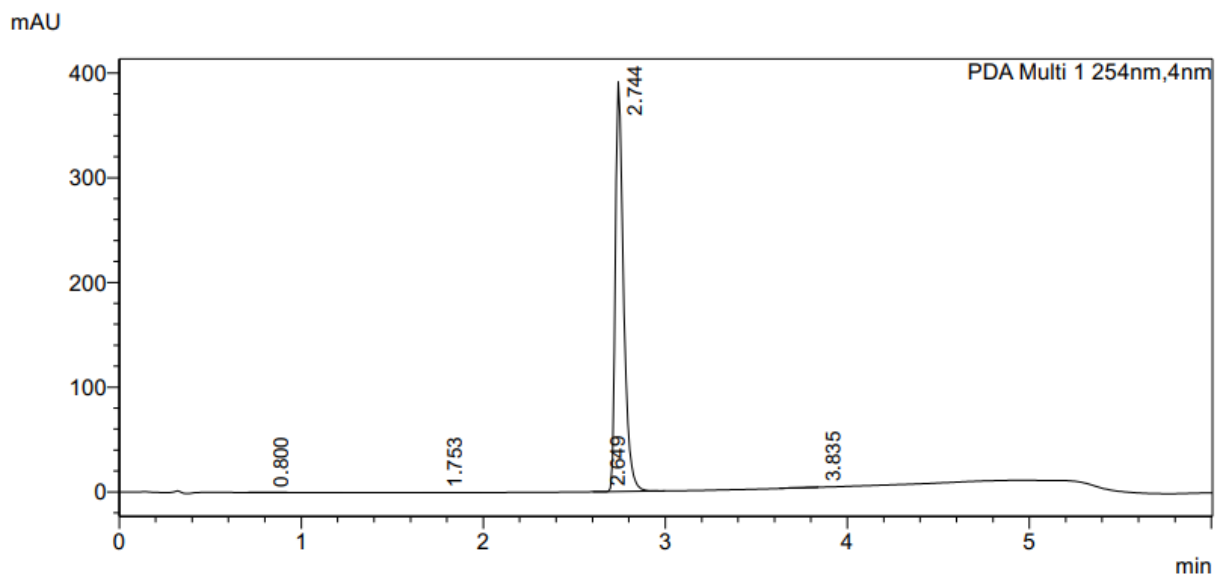
HPLC-Trace 9 HPLC-purity 12



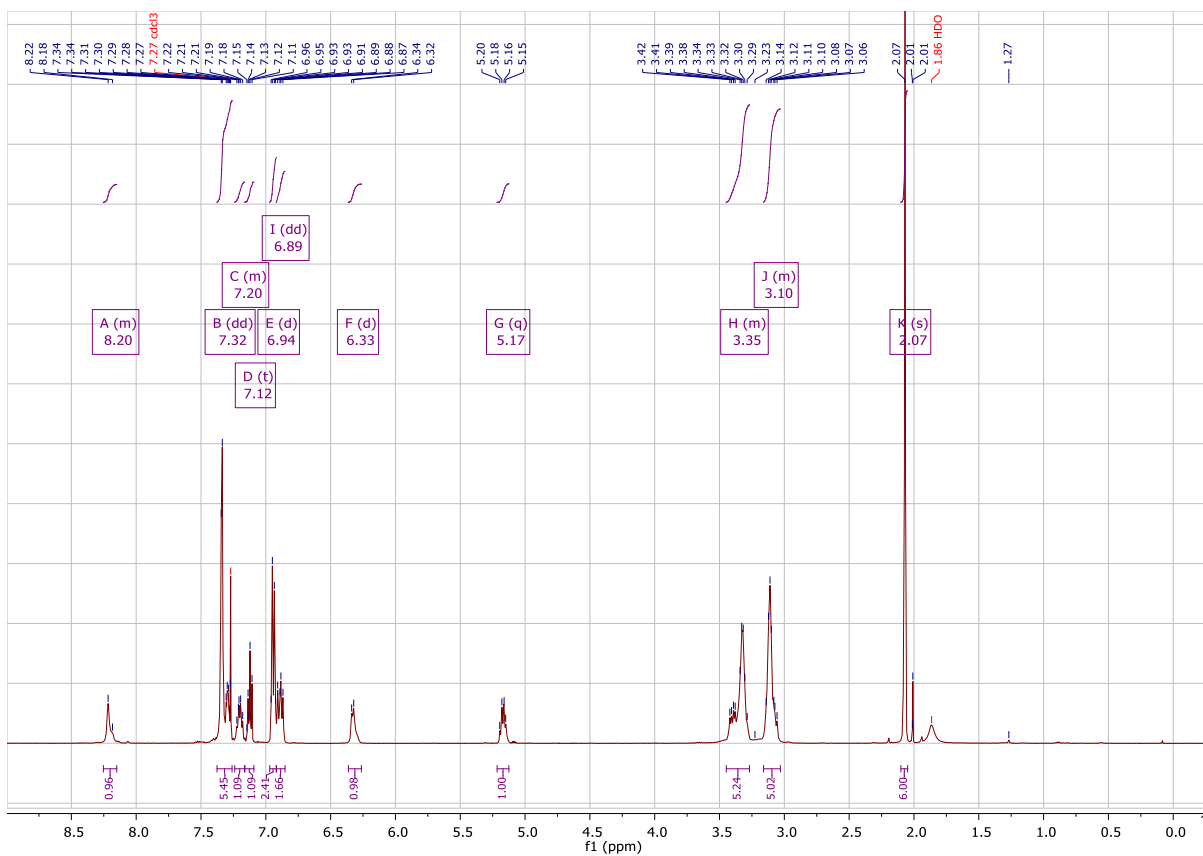
Spectrum 22 ¹H-NMR 13



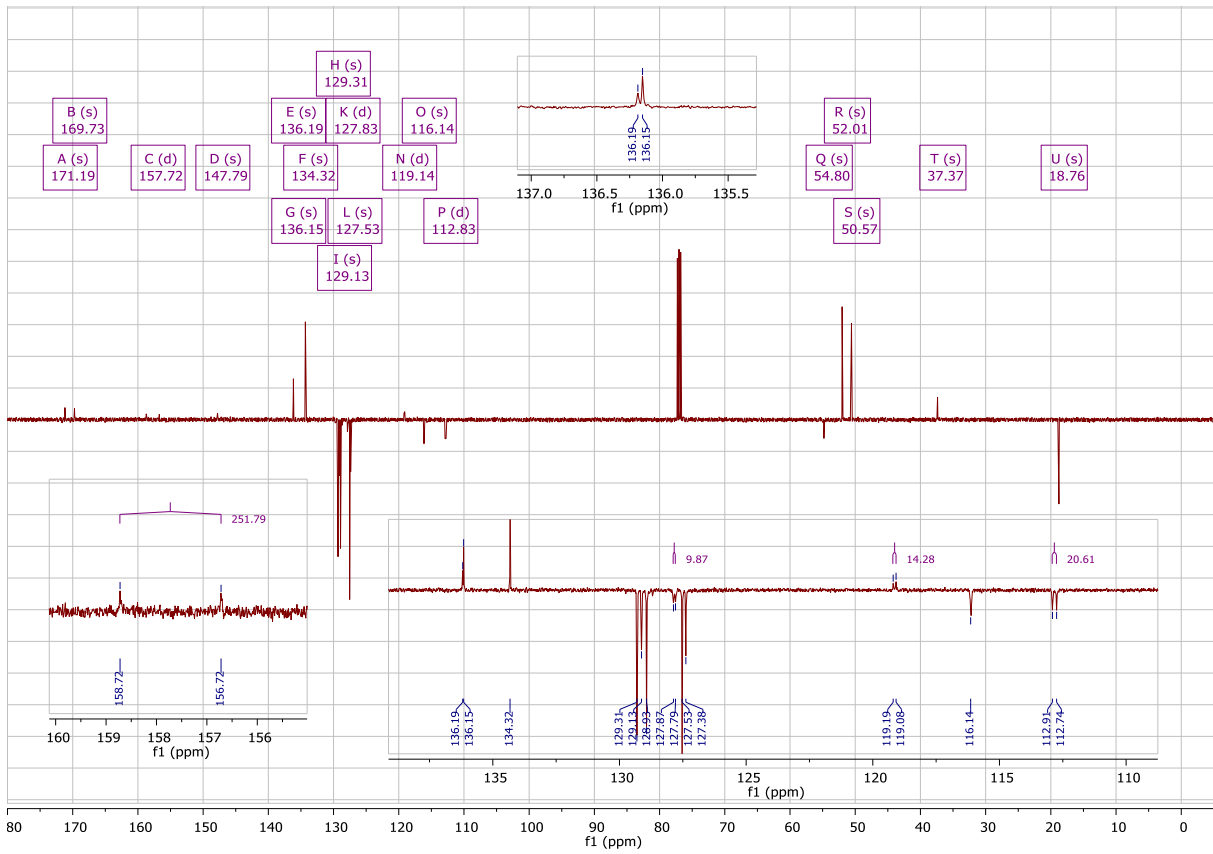
Spectrum 23 APT ¹³C-NMR 13



HPLC-Trace 10 HPLC-purity 13

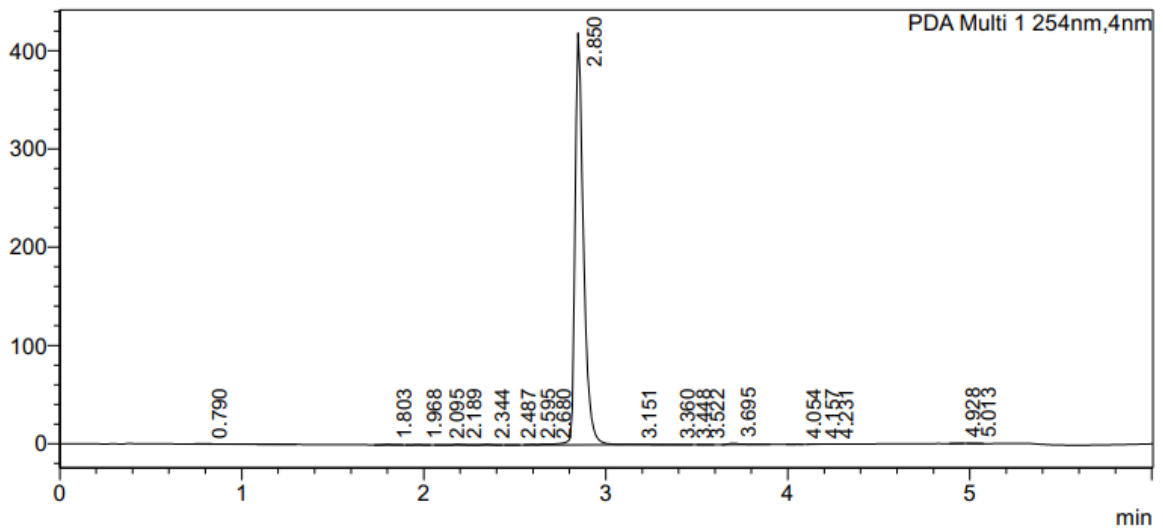


Spectrum 24 ¹H-NMR 14

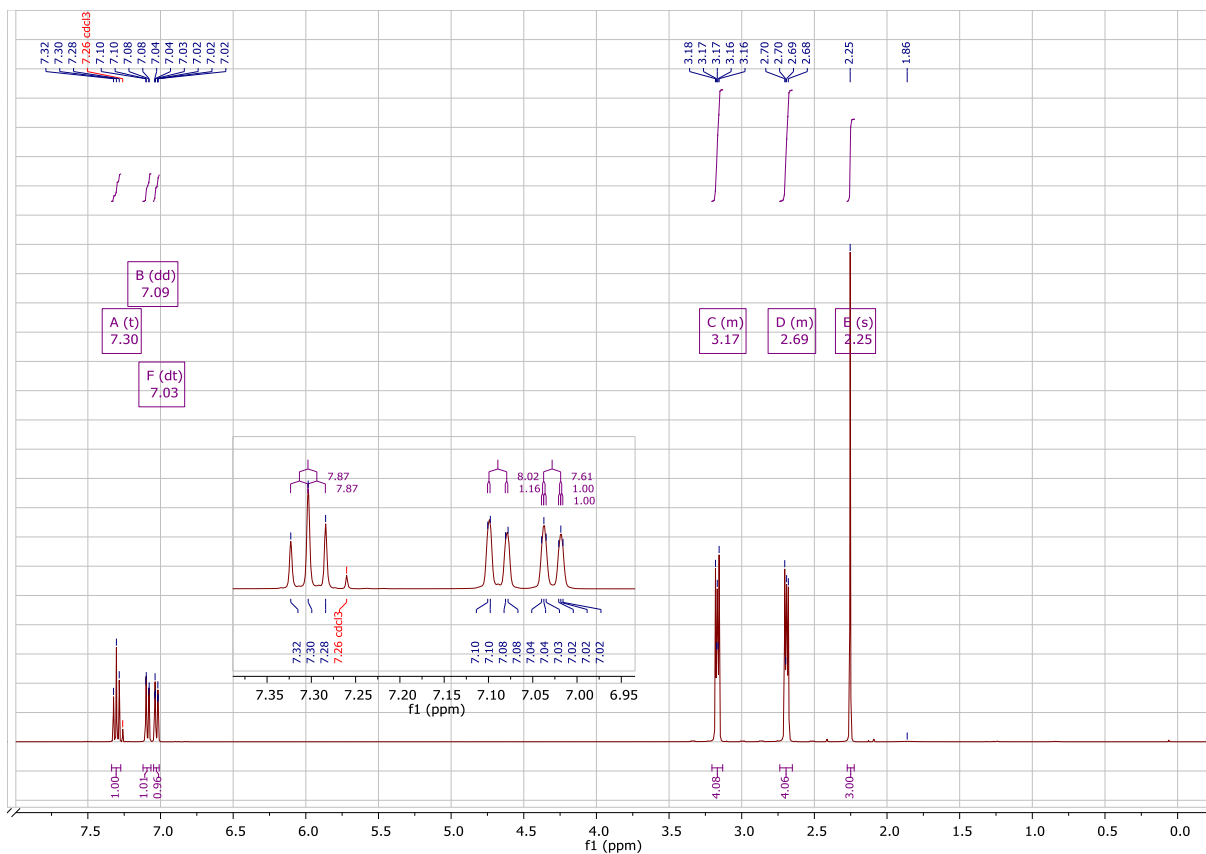


Spectrum 25 APT ¹³C-NMR 14

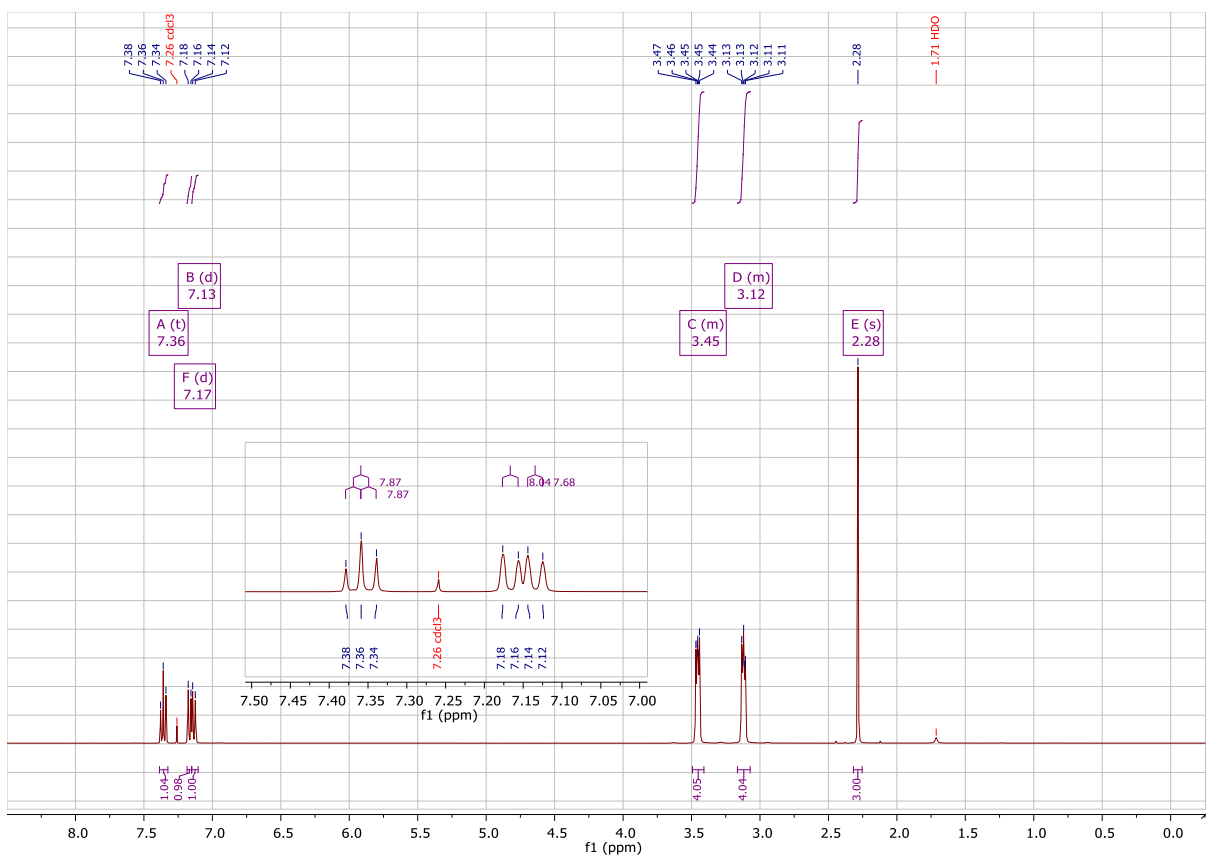
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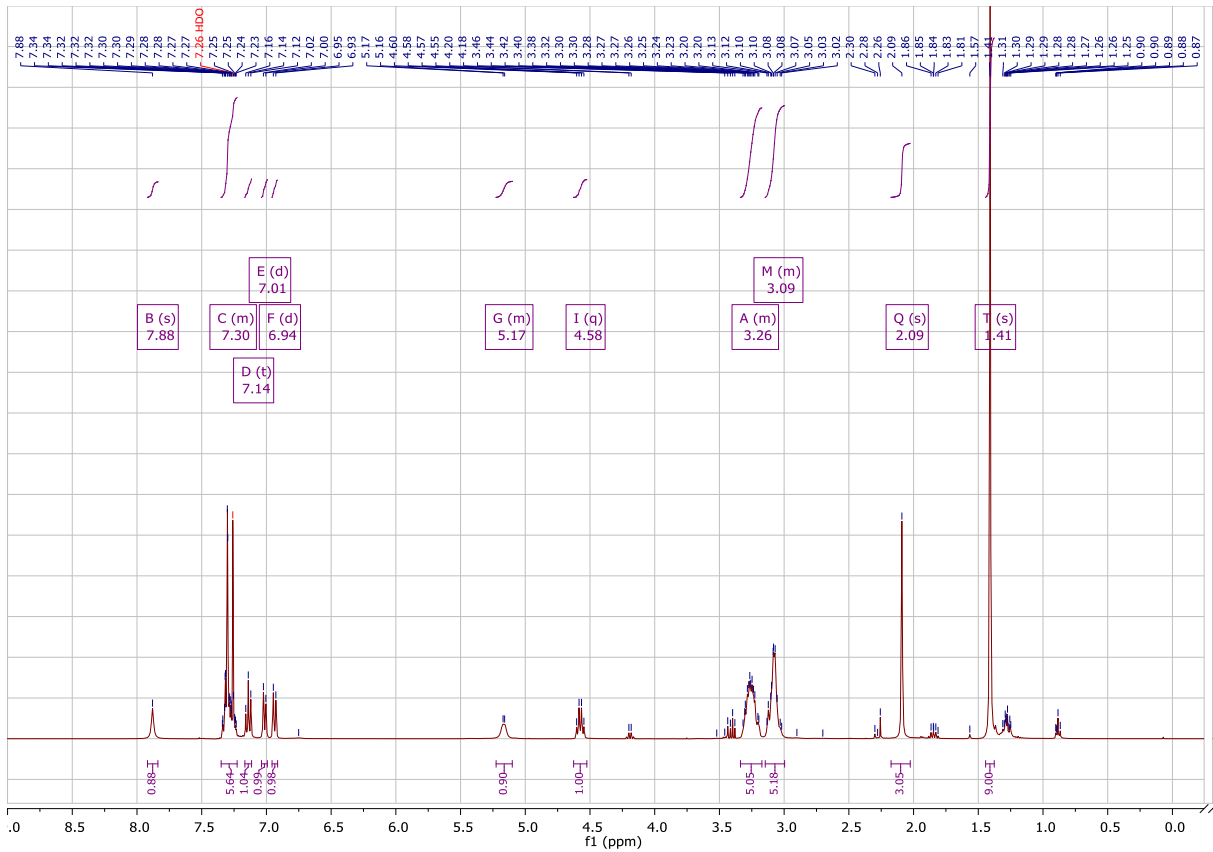
HPLC-Trace 11 HPLC-purity 14



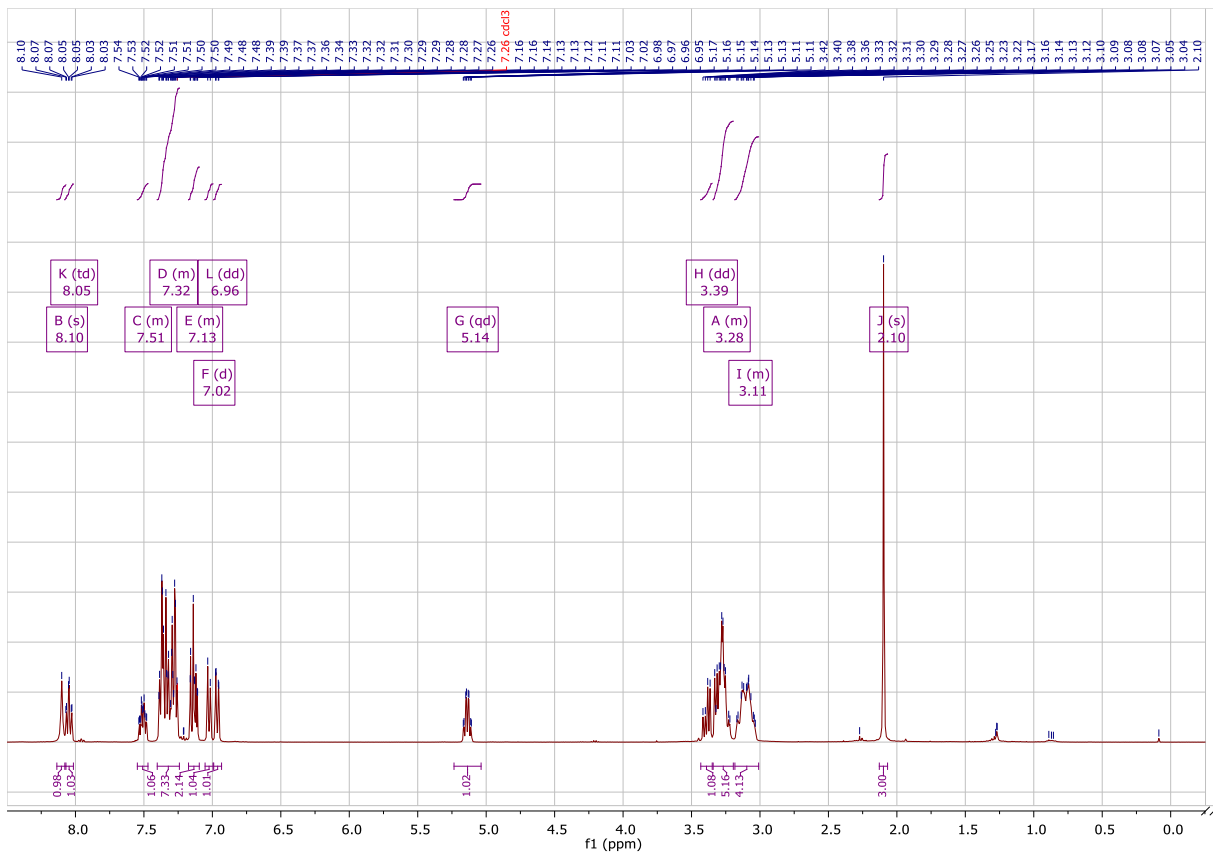
Spectrum 26 $^1\text{H-NMR}$ 15



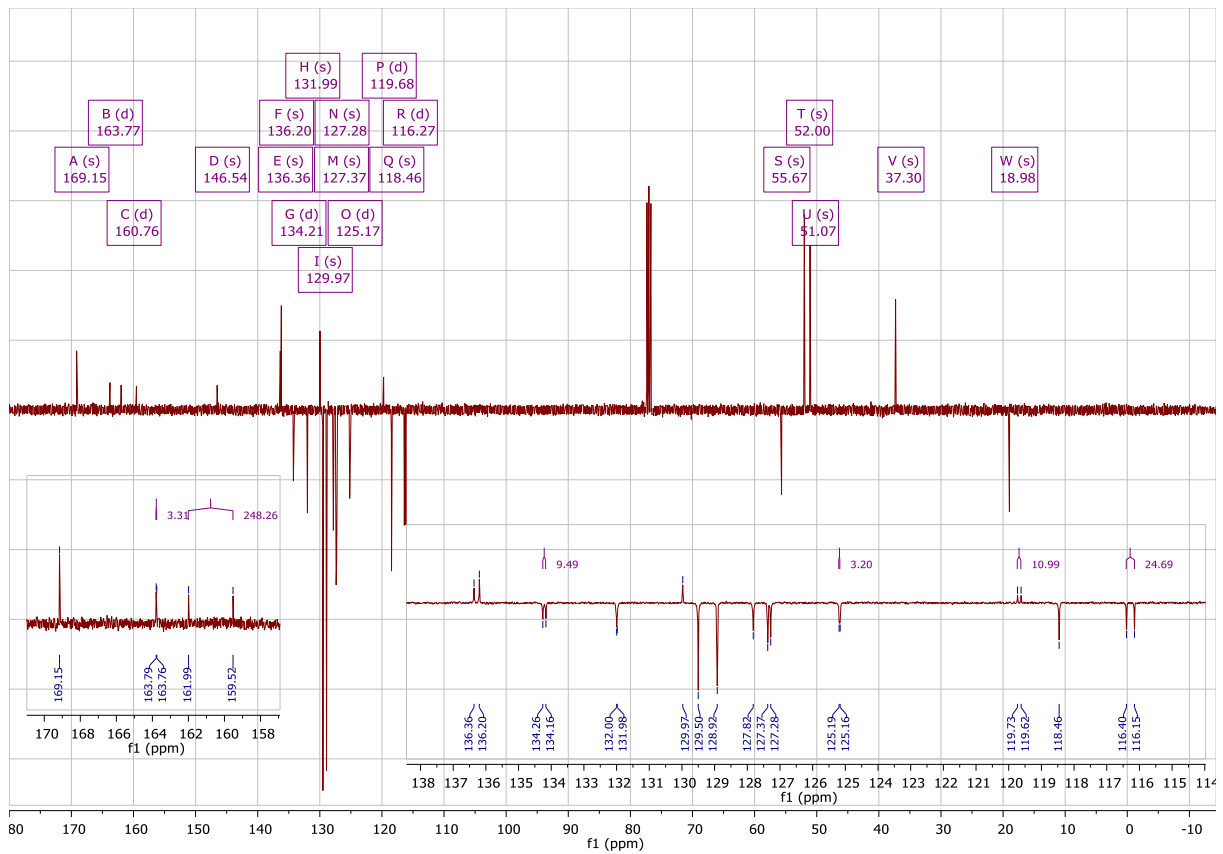
Spectrum 27 $^1\text{H-NMR}$ 16



Spectrum 28 $^1\text{H-NMR}$ 18

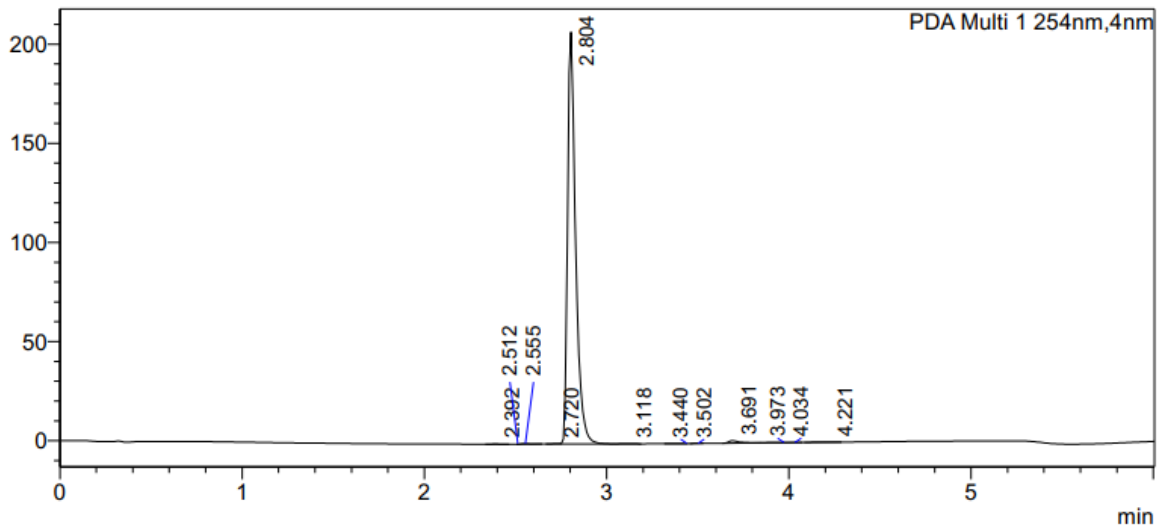


Spectrum 29 $^1\text{H-NMR}$ 19

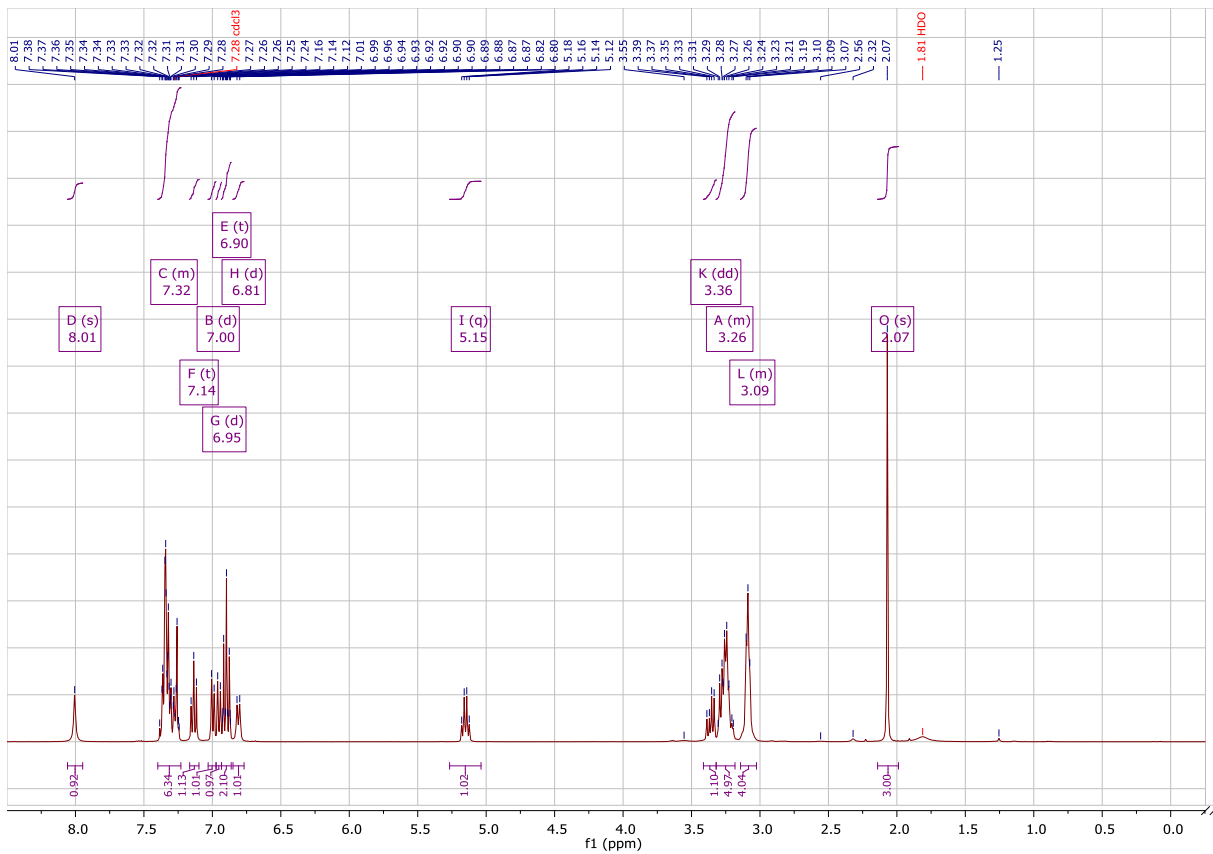


Spectrum 30 APT ¹³C-NMR 19

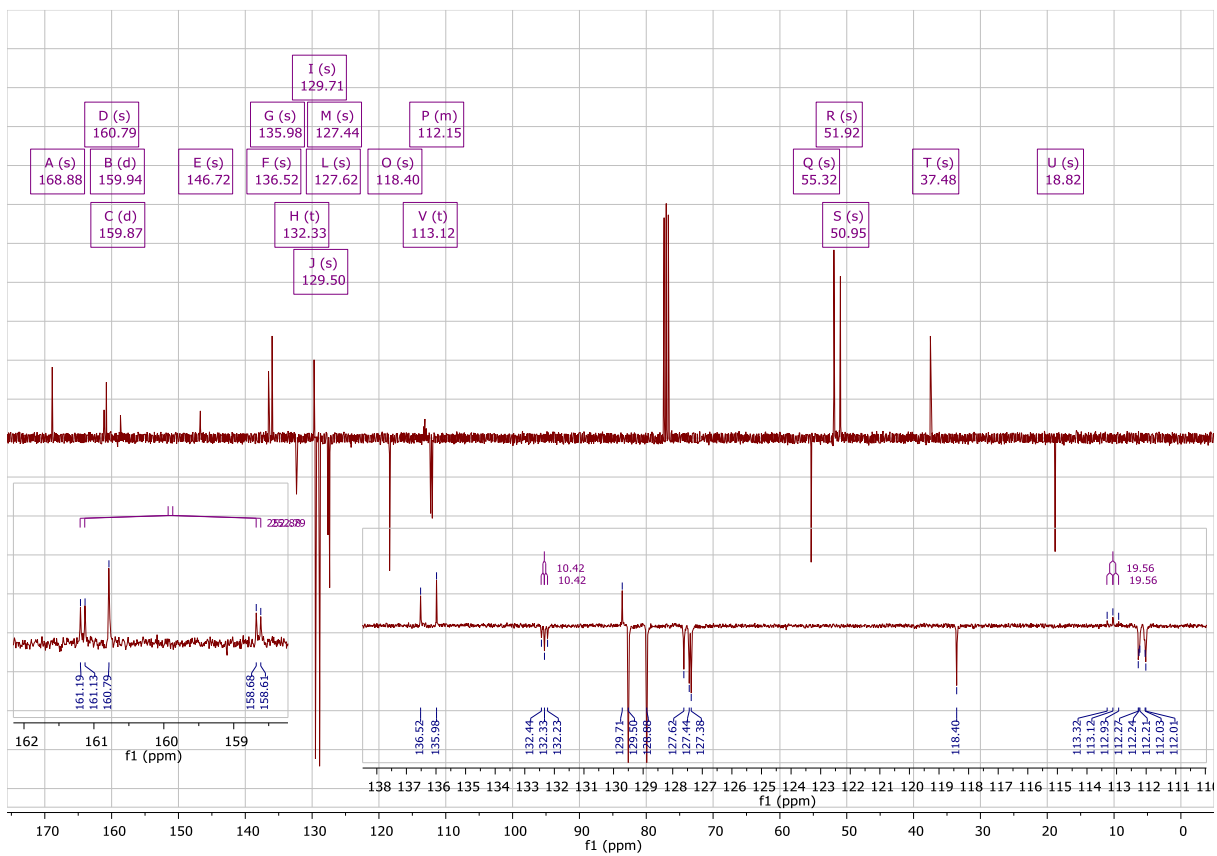
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HPLC-Trace 12 HPLC-purity 19

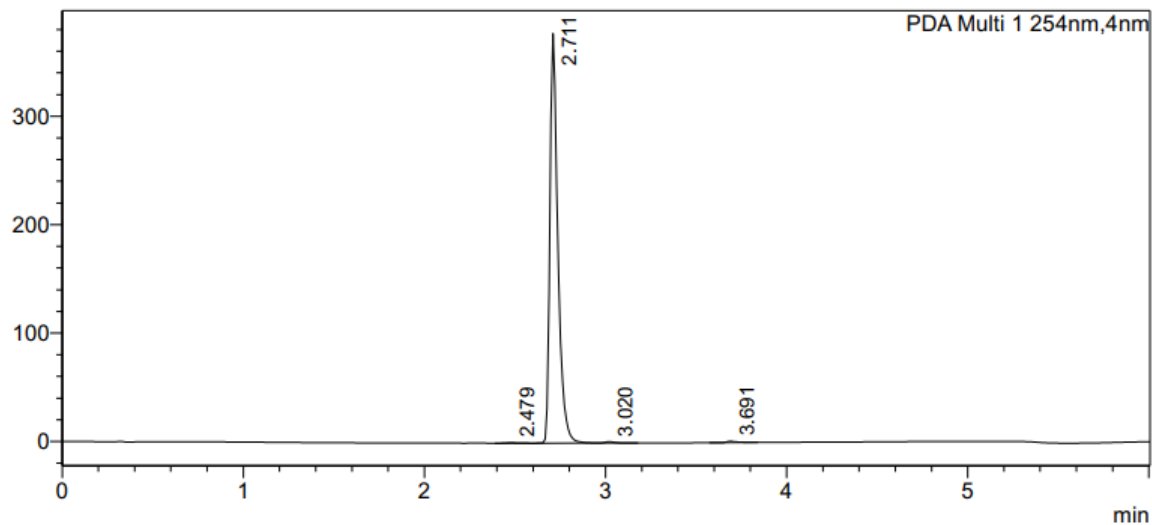


Spectrum 31 ¹H-NMR 20

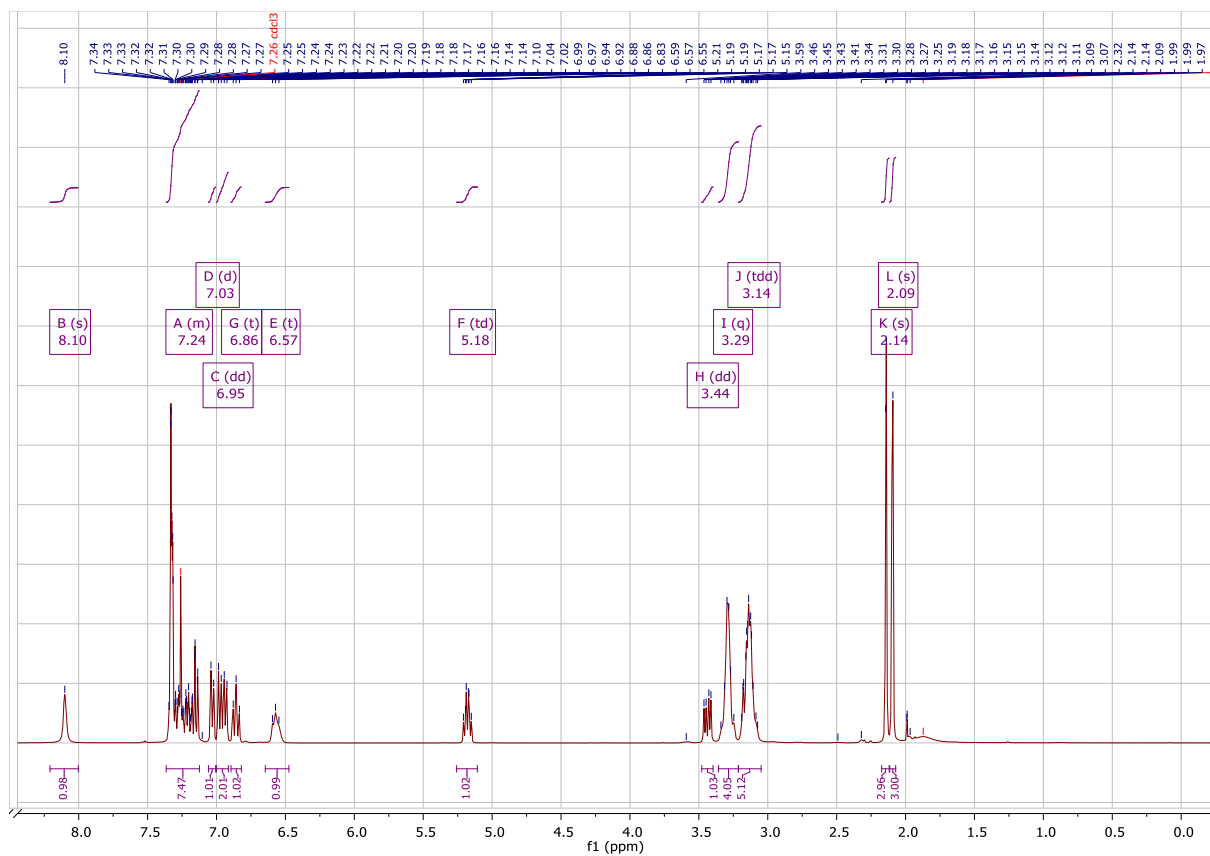


Spectrum 32 APT ¹³C-NMR 20

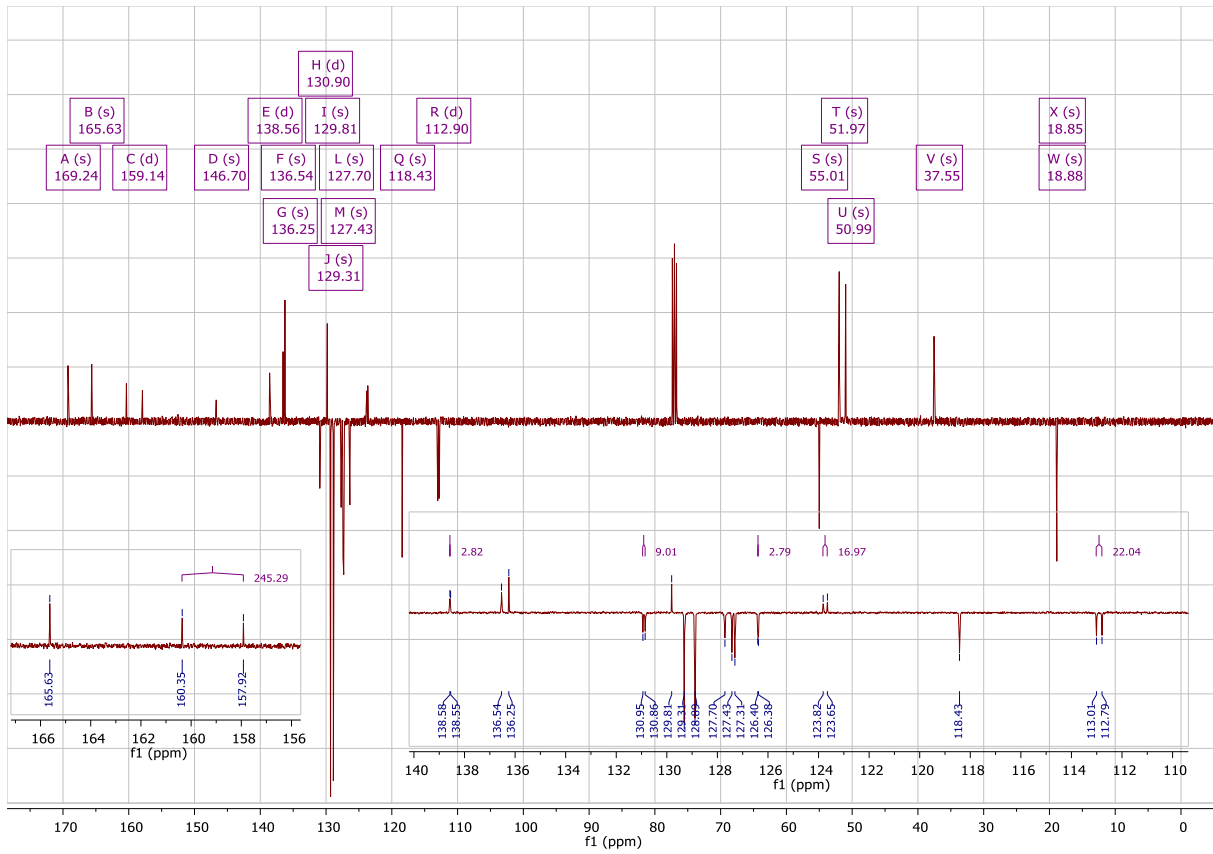
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HPLC-Trace 13 HPLC-purity 20

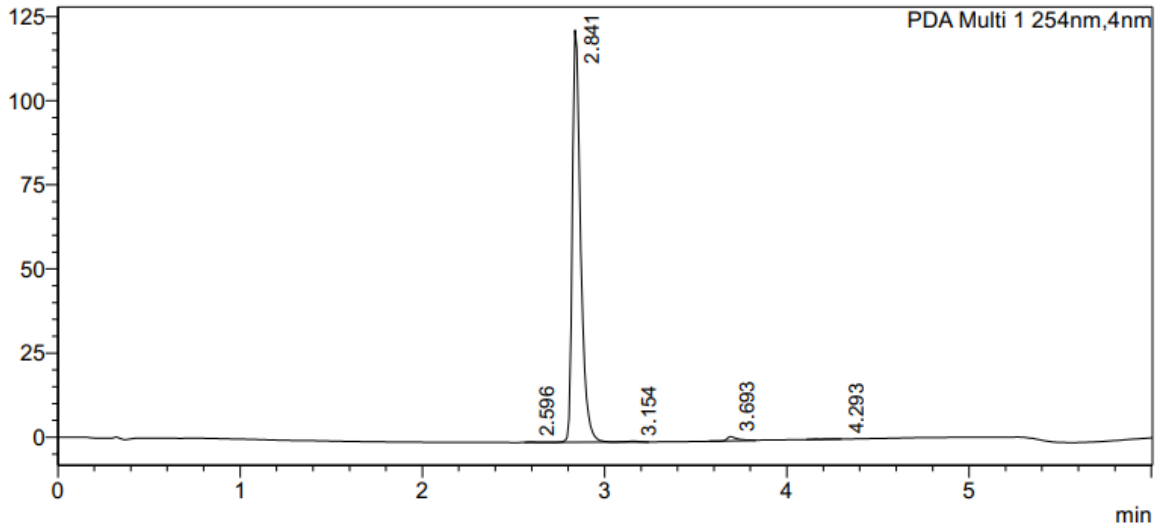


Spectrum 33 ¹H-NMR 21

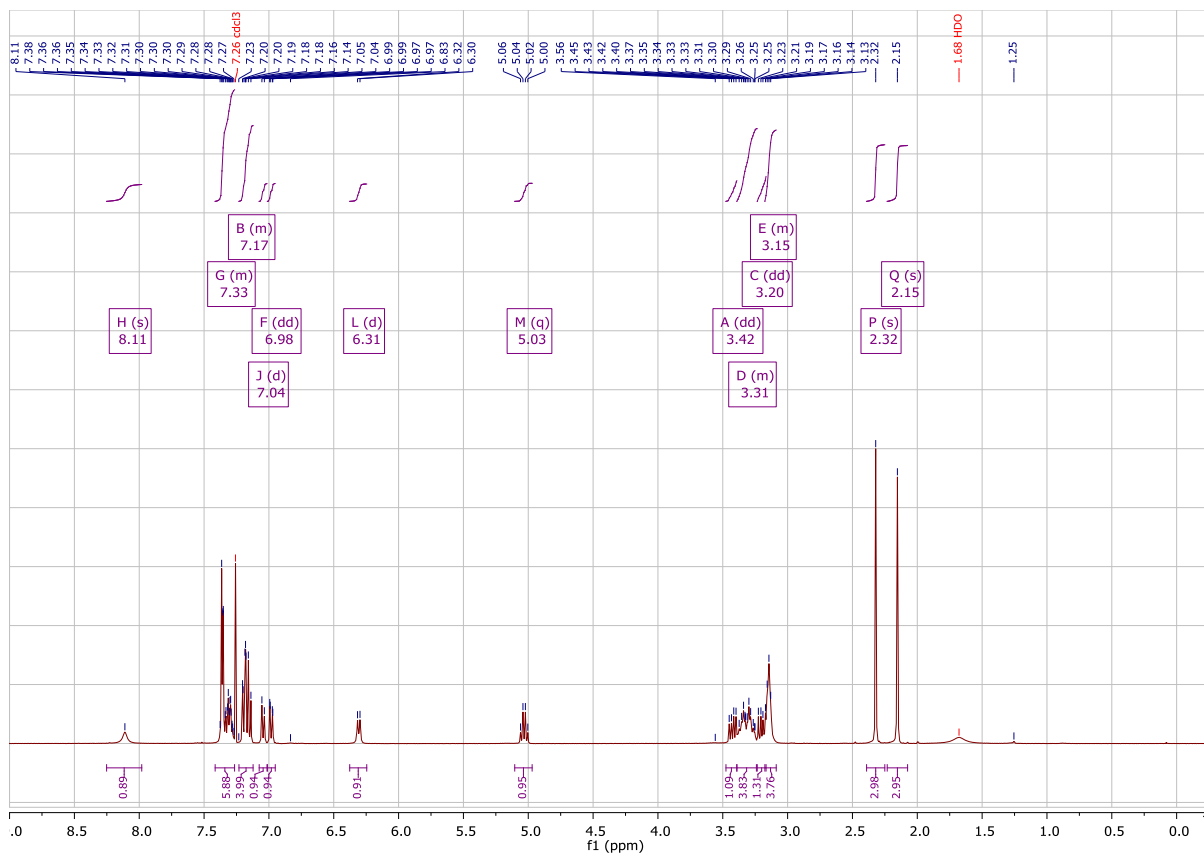


Spectrum 34 APT ¹³C-NMR 21

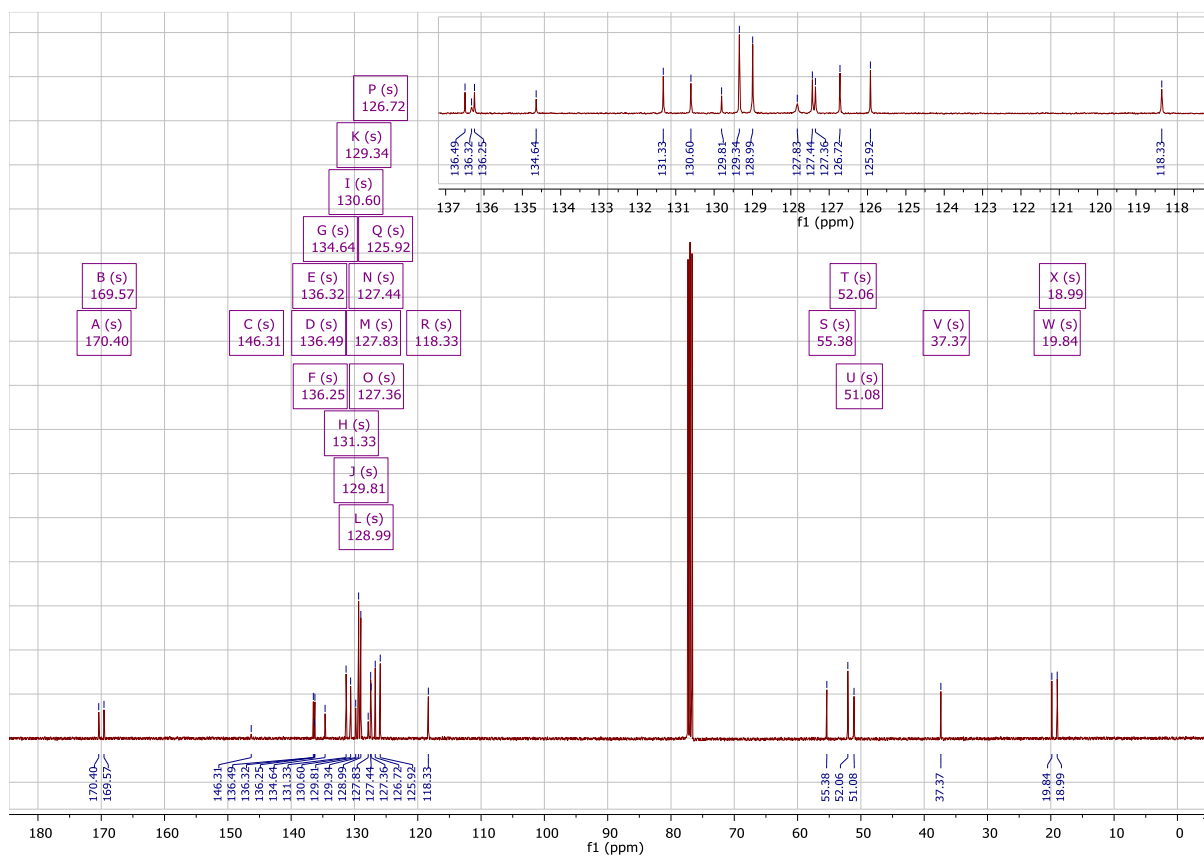
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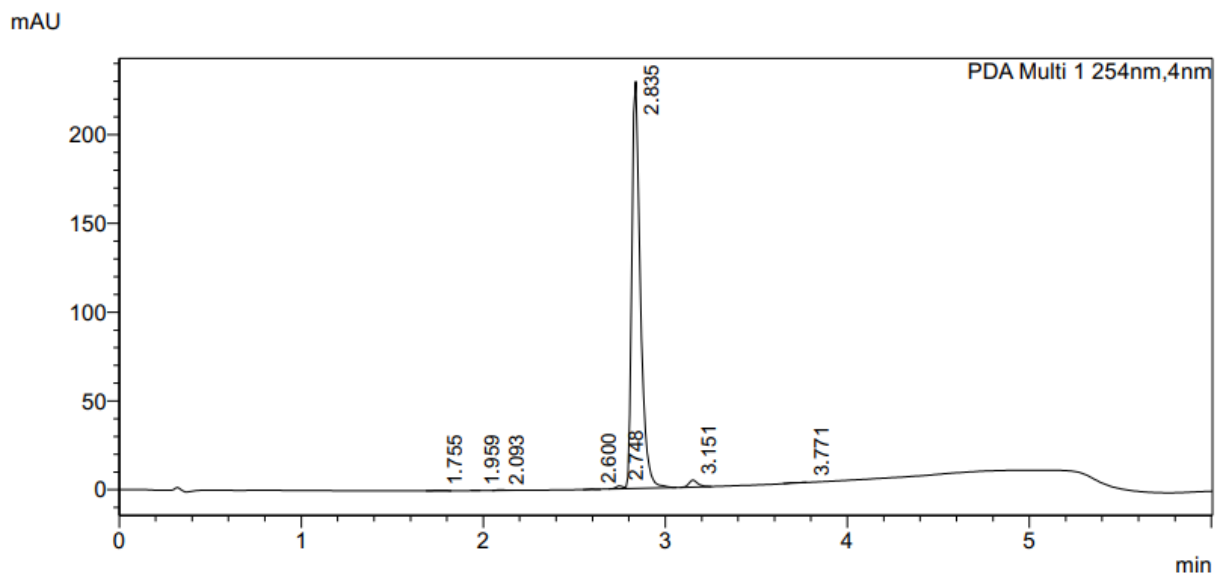
HPLC-Trace 14 HPLC-purity 21



Spectrum 35 ¹H-NMR 22



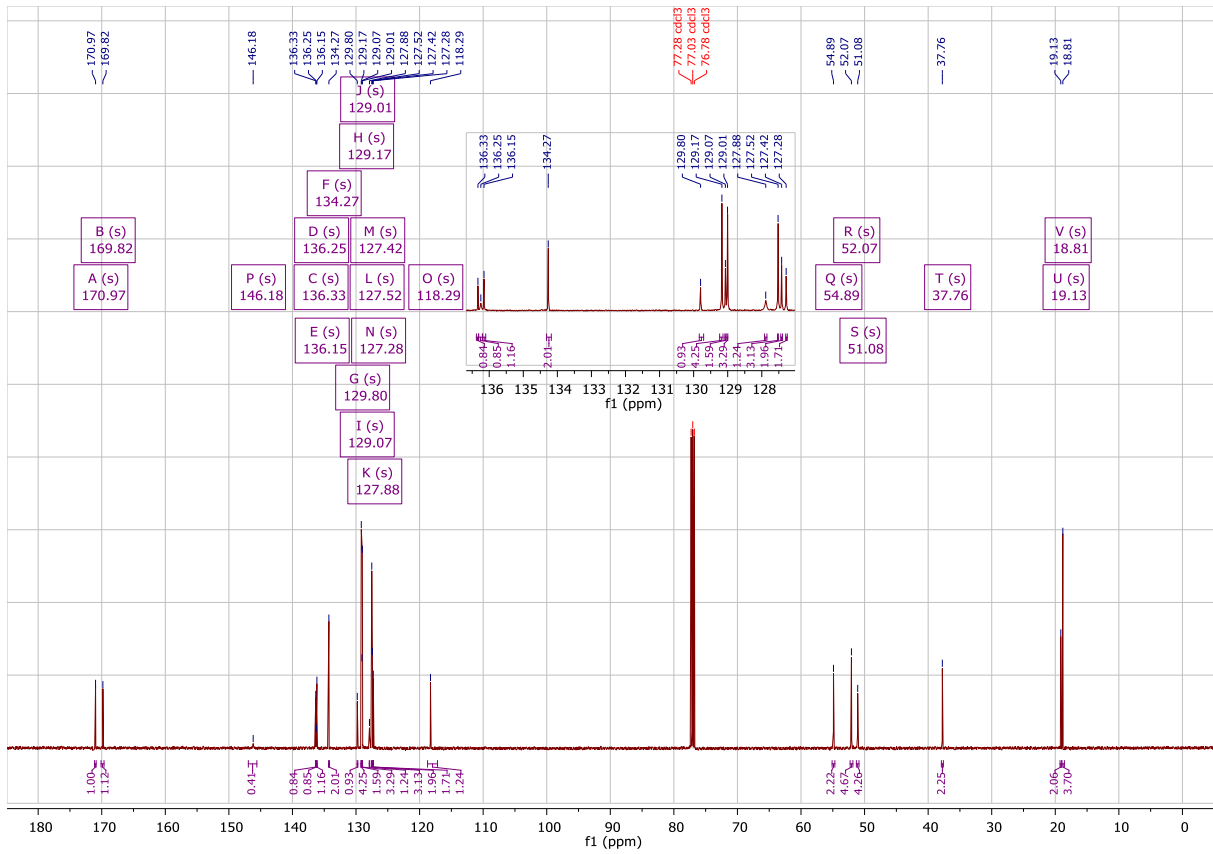
Spectrum 36 ¹³C-NMR 22



HPLC-Trace 15 HPLC-purity 22

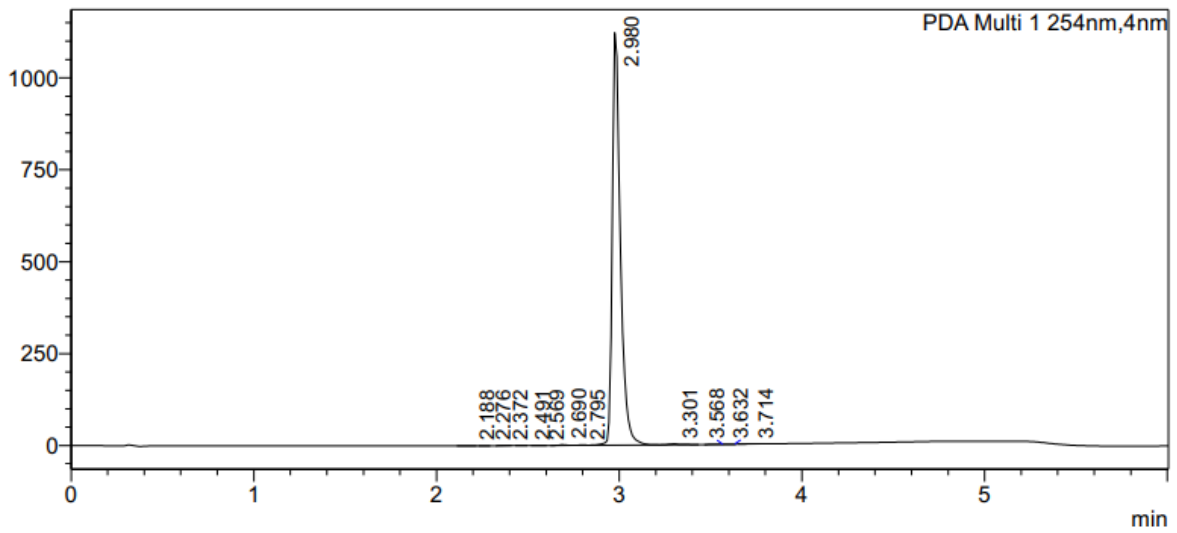


Spectrum 37 ¹H-NMR 23

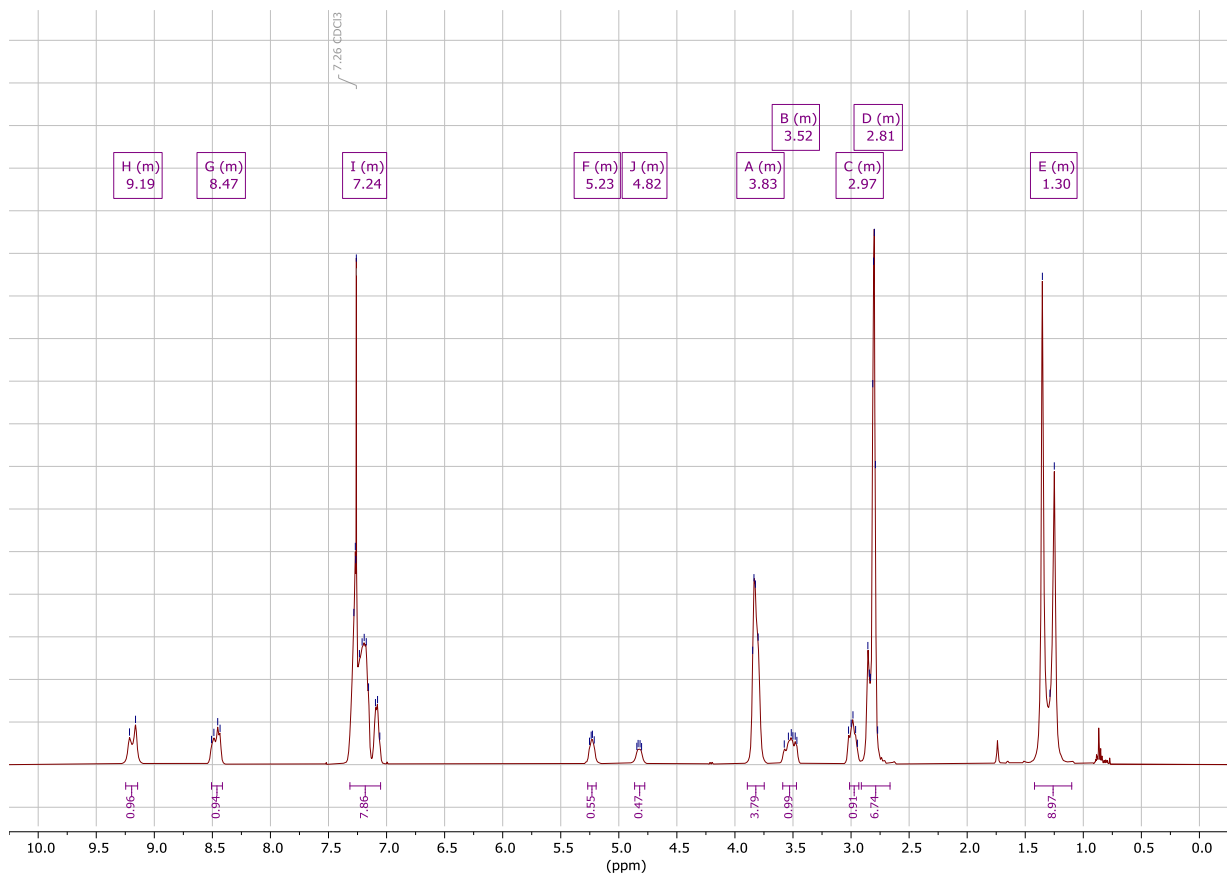


Spectrum 38 ¹³C-NMR 23

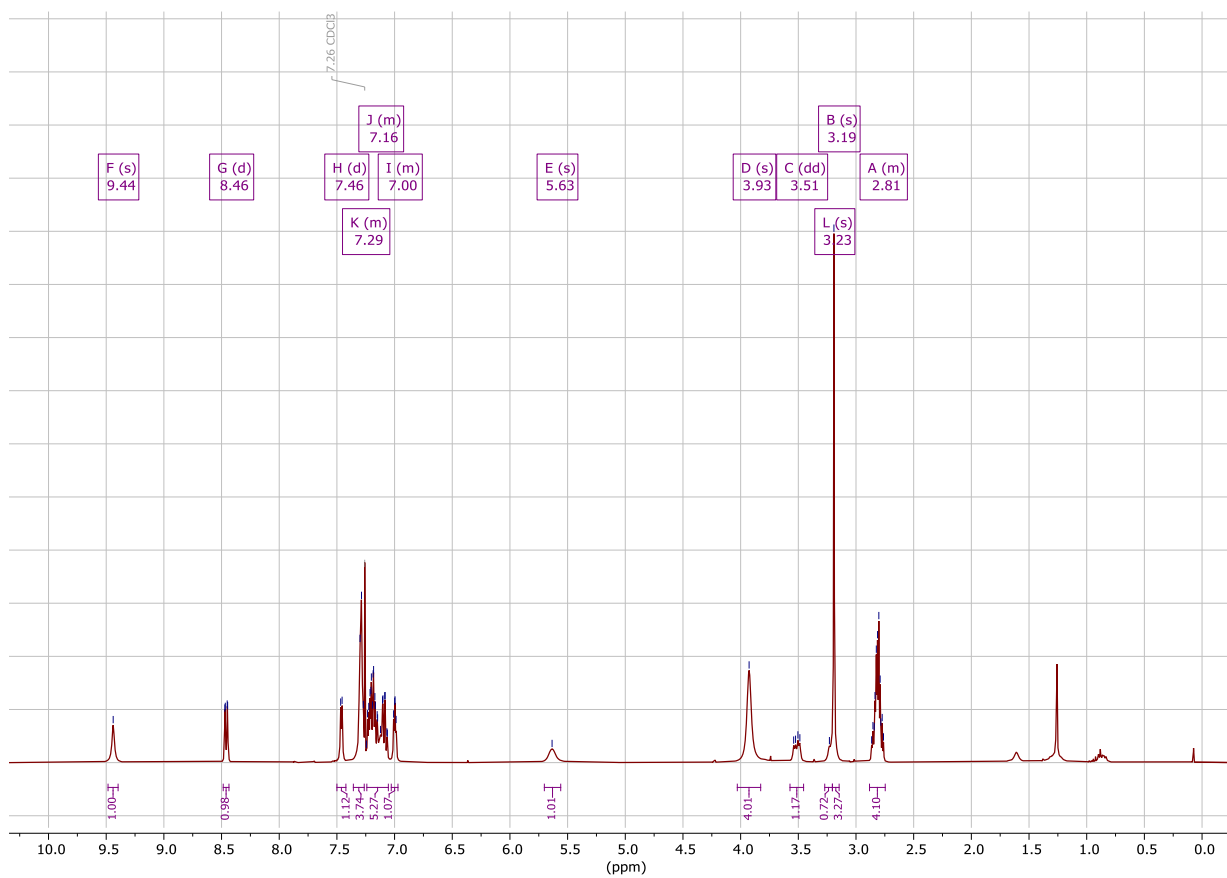
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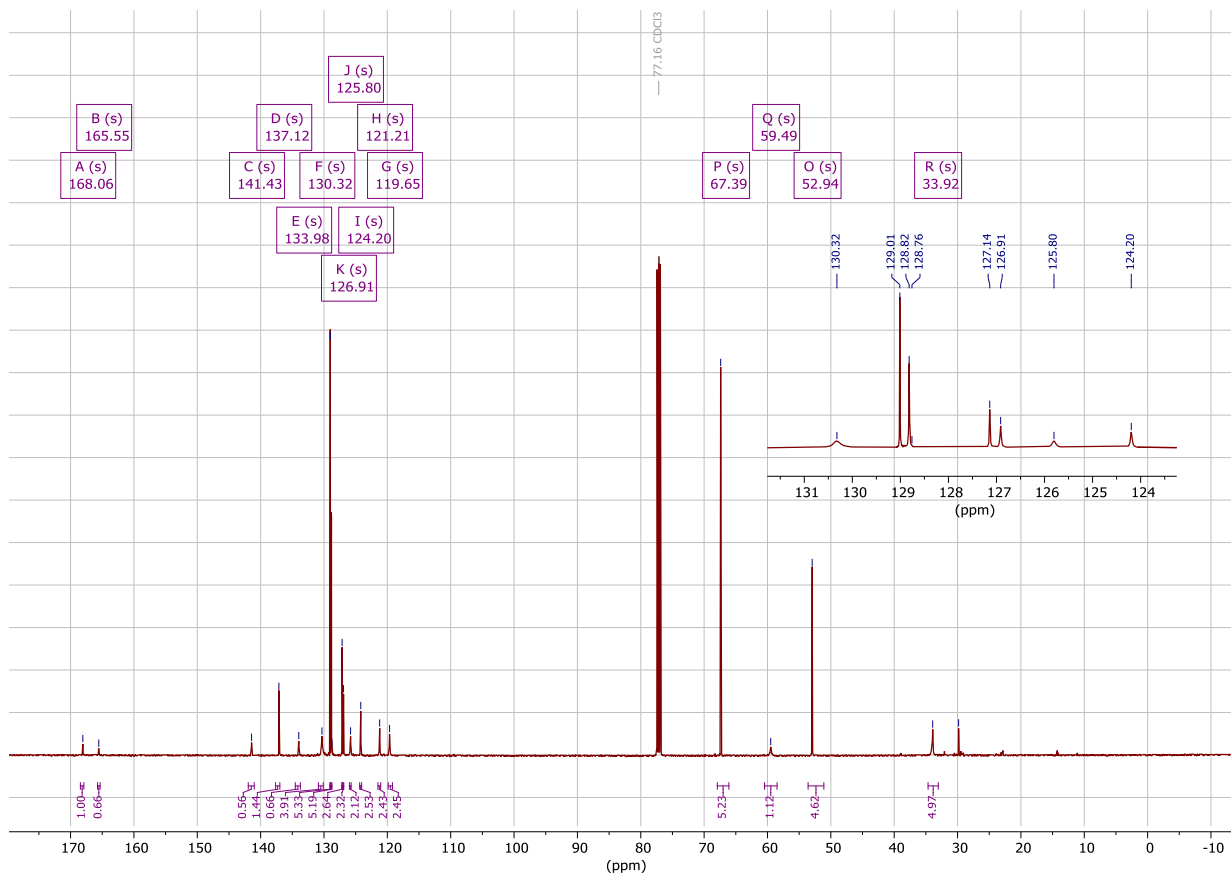
HPLC-Trace 16 HPLC-purity 23



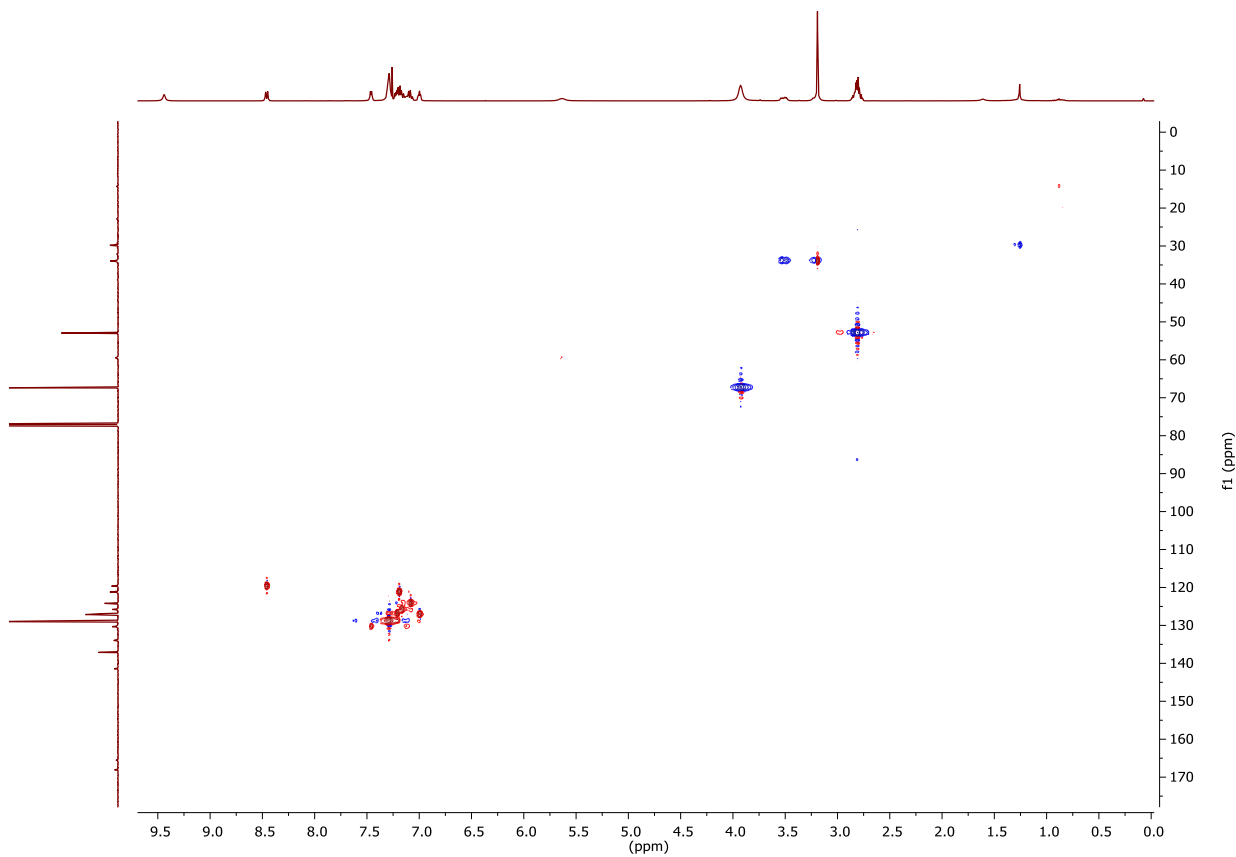
Spectrum 39 ¹H-NMR 24



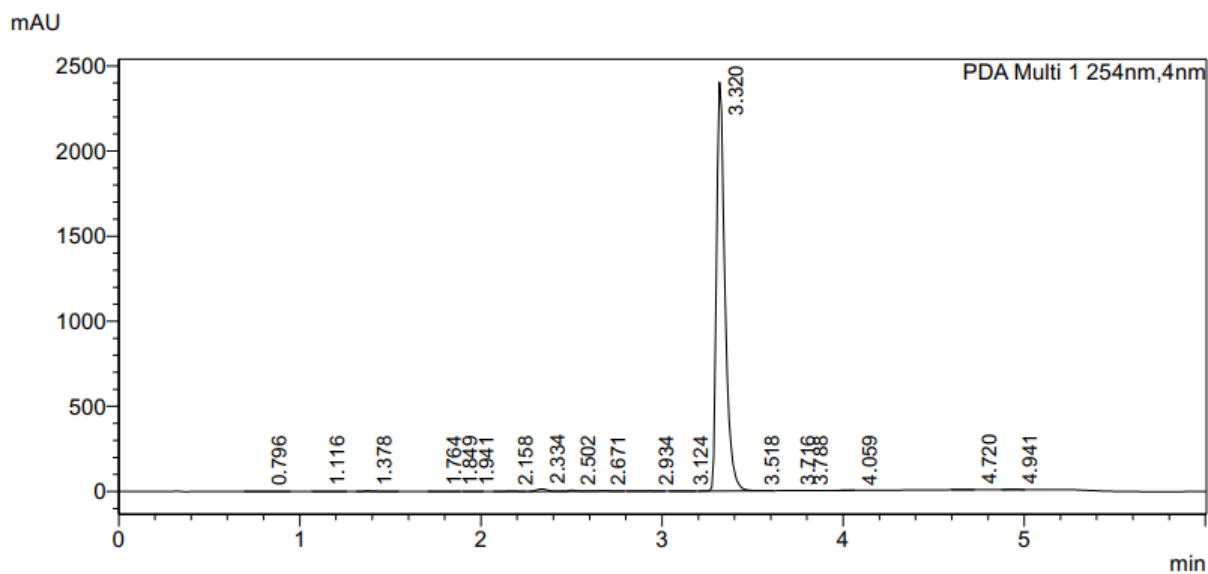
Spectrum 40 ¹H-NMR 25



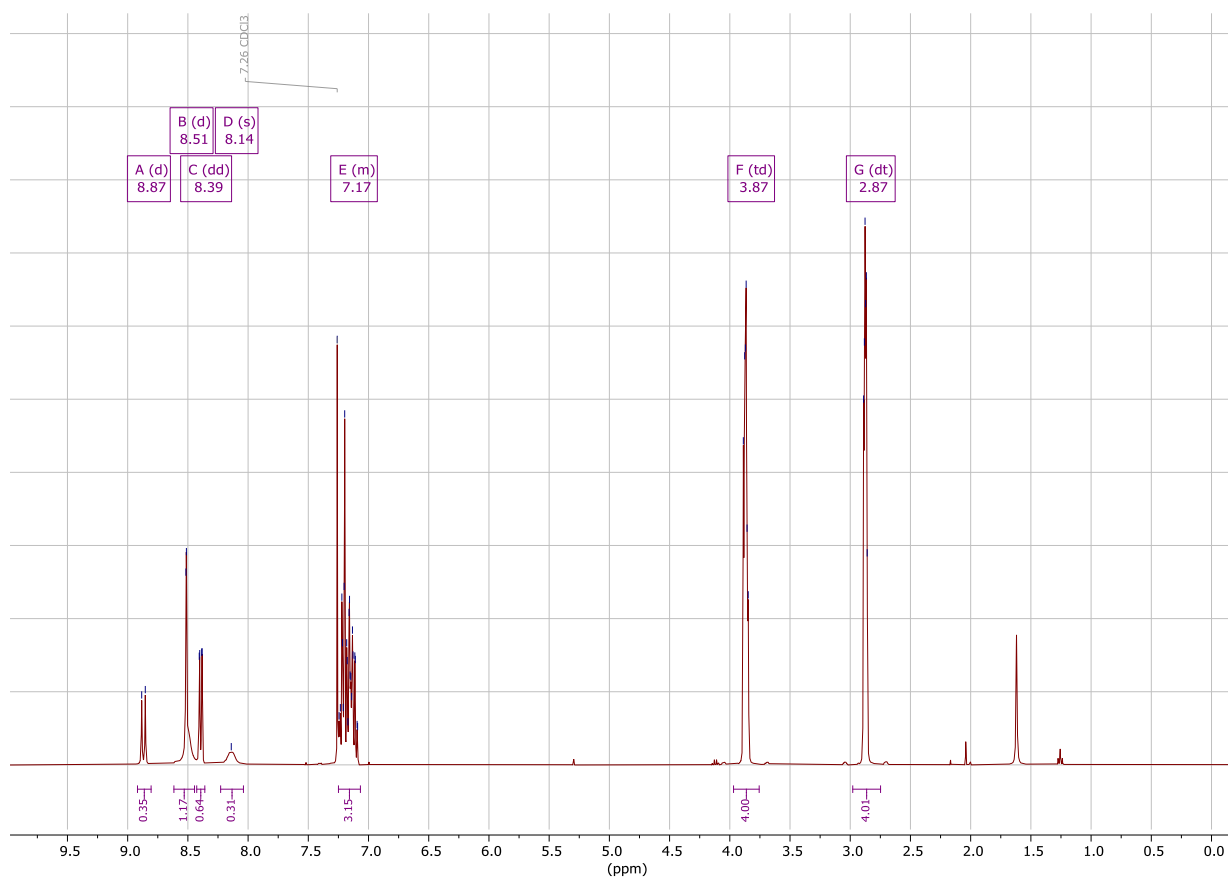
Spectrum 41 ¹³C-NMR 25



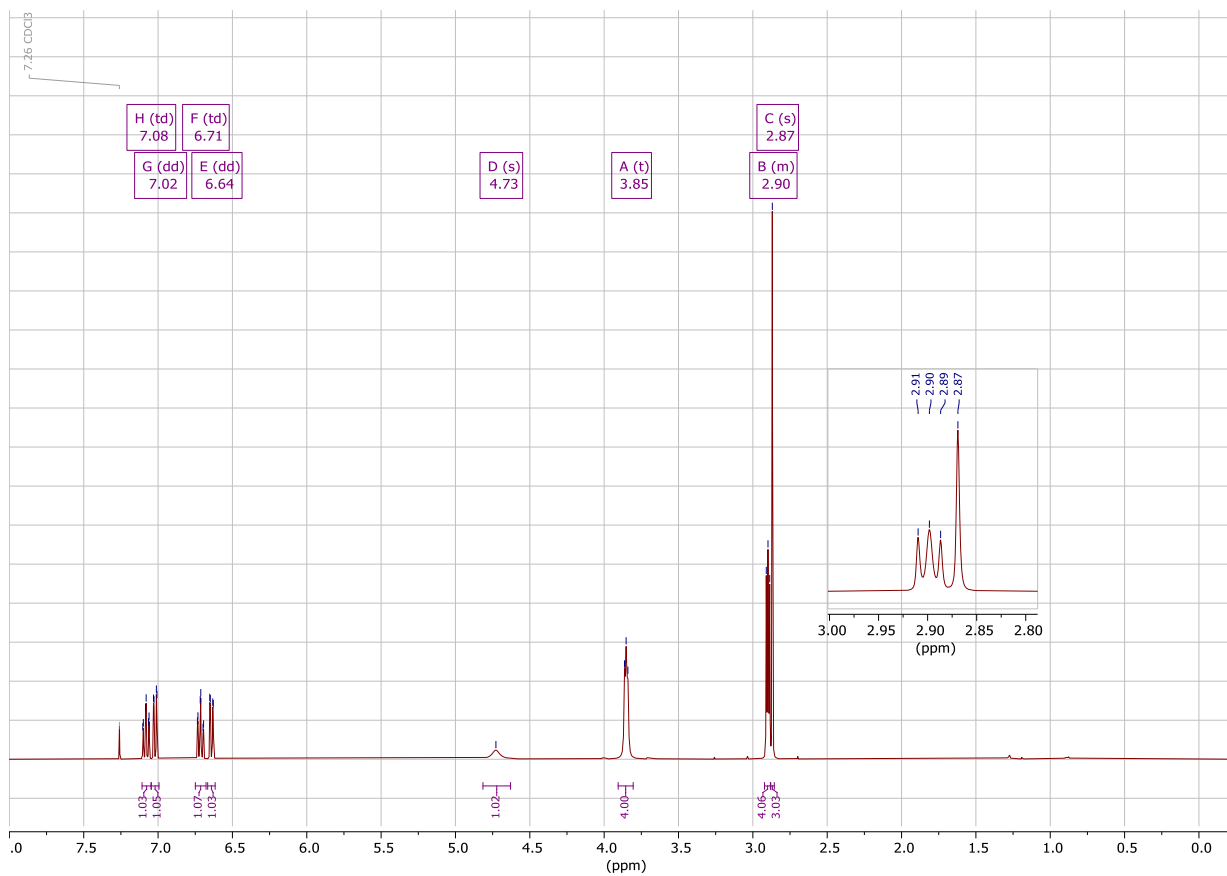
Spectrum 42 HSQC 25



HPLC-Trace 17 HPLC-purity 25



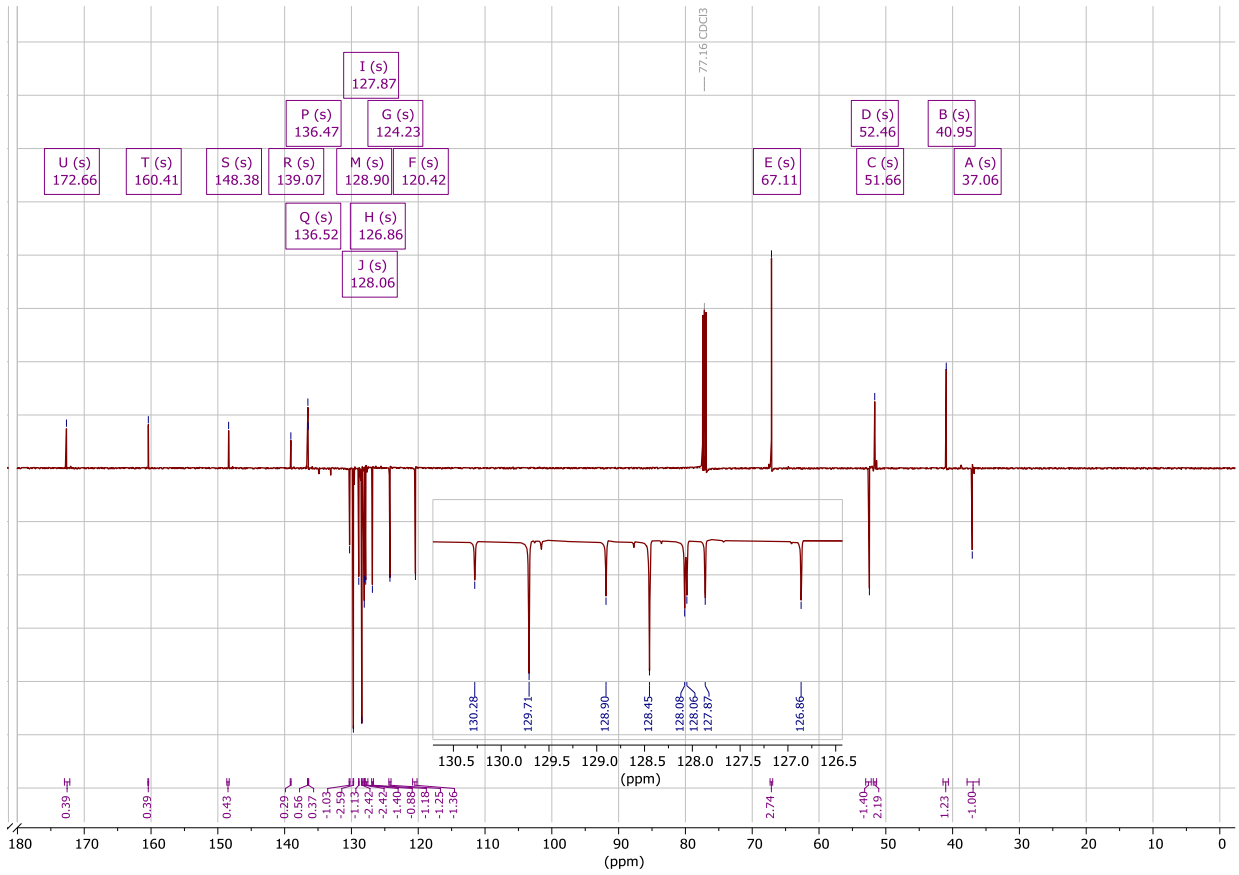
Spectrum 43 ¹H-NMR 26



Spectrum 44 ¹H-NMR 27

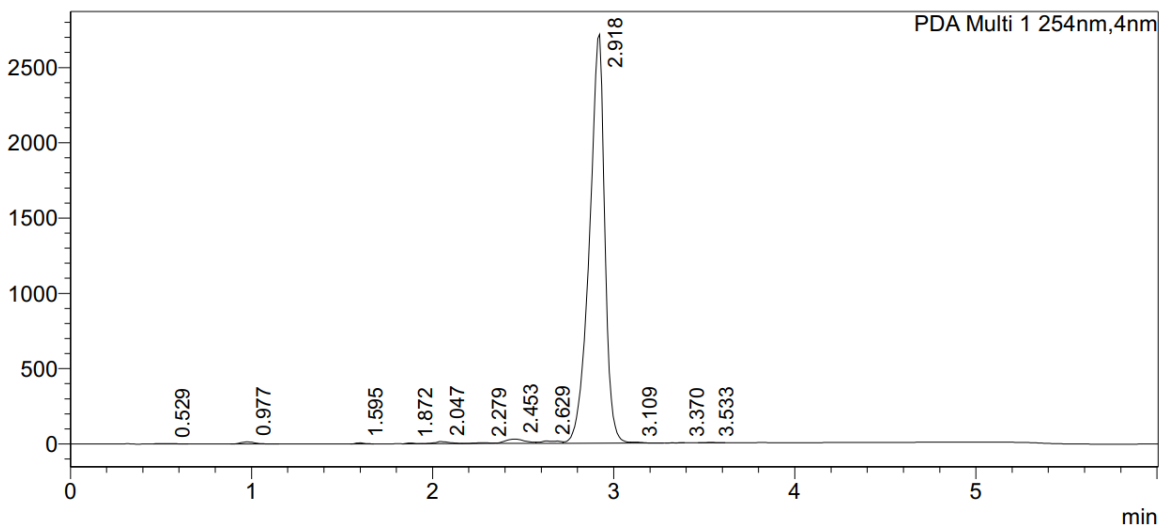


Spectrum 45 ¹H-NMR 29



Spectrum 46 APT ¹³C-NMR 29

mAU



HPLC-Trace 18 HPLC-purity 29