

## Supporting Information

### **Carboxylated Nanoparticle Surfaces Enhance Association with Mucoic *P. aeruginosa* Biofilms**

*Elad Deiss-Yehiely*<sup>1, 2</sup>, *Abigail E. Dzordzorme*<sup>3</sup>, *Maggie Elizabeth Loisel*<sup>4, 5</sup>, *Lael M. Yonker*<sup>4-6</sup>, and *Paula T. Hammond*<sup>2, 7, 8, \*</sup>

<sup>1</sup>Department of Materials Science and Engineering, Massachusetts Institute of Technology, Cambridge, MA, 02139, United States

<sup>2</sup>Koch Institute for Integrative Cancer Research, Massachusetts Institute of Technology, Cambridge, MA, 02139, United States

<sup>3</sup>Department of Biological Engineering, Massachusetts Institute of Technology, Cambridge, MA, 02139, United States

<sup>4</sup>Mucosal Immunology and Biology Research Center, Division of Infectious Disease, Massachusetts General Hospital, Boston, MA, 02114, United States

<sup>5</sup>Department of Pediatrics, Division of Infectious Disease, Massachusetts General Hospital, Boston, MA, 02114, United States

<sup>6</sup>Harvard Medical School, Boston, MA, 02115, United States

<sup>7</sup>Institute for Soldier Nanotechnologies, Massachusetts Institute of Technology, Cambridge, MA, 02139, United States

<sup>8</sup>Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge, MA, 02139, United States

\* Corresponding author: [hammond@mit.edu](mailto:hammond@mit.edu)

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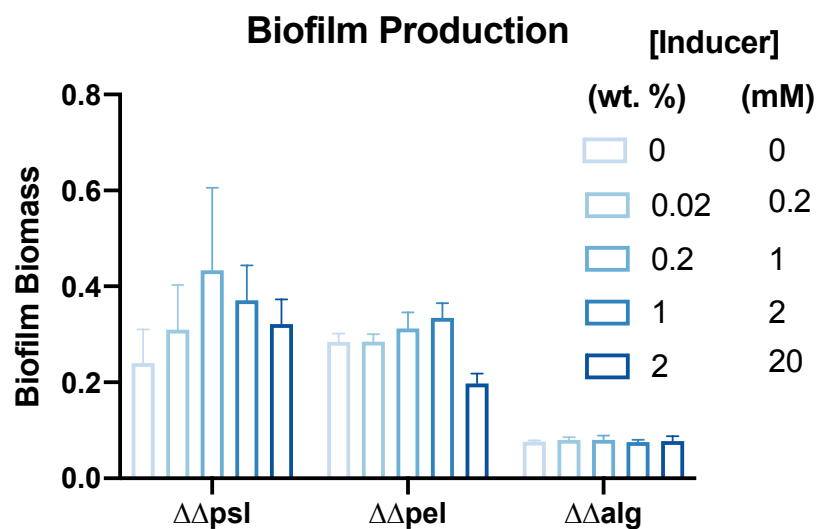


Figure S1. Variable biofilm biomass production based on inducer concentration for all three strains. An inducer concentration of 0.2 weight % arabinose was chosen for  $\Delta\Delta psl$  and  $\Delta\Delta pel$ , whereas an inducer concentration of 1 mM IPTG was chosen from  $\Delta\Delta alg$ . The chosen concentrations, as shown in green, are due to maximized biomass production and previous literature results. N = 12 technical replicates.

### Layer-by-Layer Nanoparticle Panel

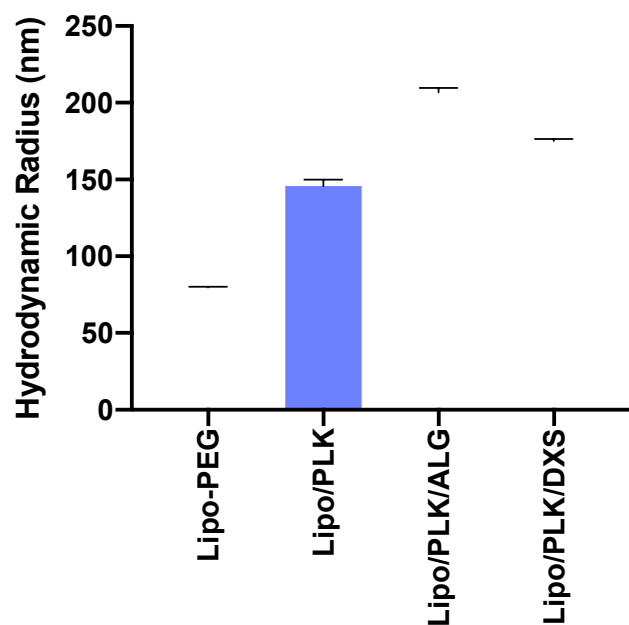


Figure S2. The low polydispersity index ( $< 0.3$ ) of LbL NPs indicates monodispersity.  $N = 3$  technical replicates of the same NP synthesis.

### Viable Microbes on MBEC Assay Device

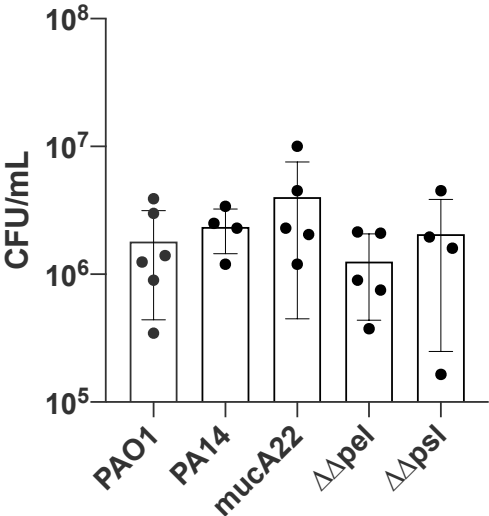


Figure S3. Insignificant biofilm CFU/mL differences between strains grown on the MBEC Assay Kit for 48 hours. N = 4-6 technical replicates.

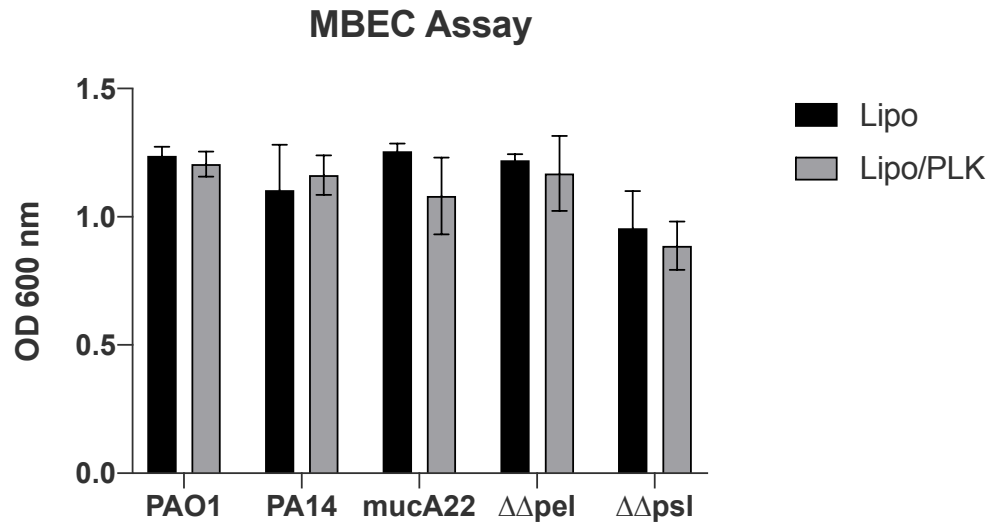


Figure S4. Liposomes and Lipo/PLK formulations are compatible with biofilms. Liposomes and Lipo/PLK formulations were tested for the MBEC at 50  $\mu\text{g}/\text{mL}$ , and showed little to no inhibitory activity, as seen by high  $\text{OD}_{600}$  measurements. N = 12 technical replicates.

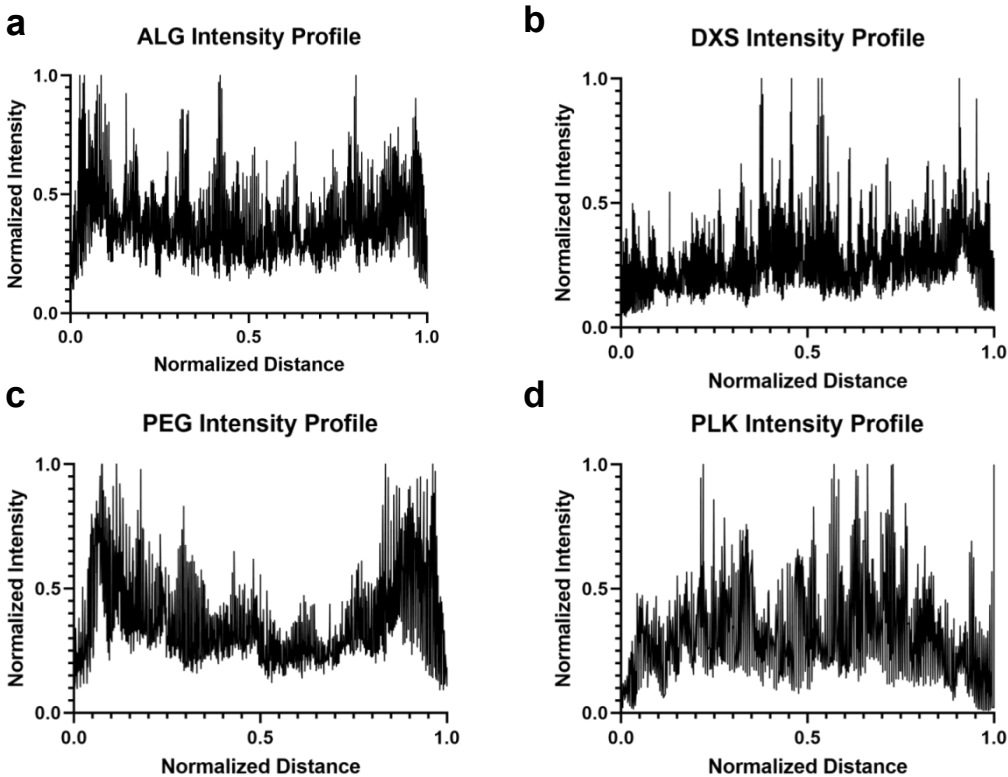


Figure S5. Pixel profile plots of all four NP systems interacting with biofilms produced by mucA22. Pixel intensity profiles for **a** ALG, **b** DXS, **c** PEG, and **d** PLK are the aggregate of five evenly spaced line width profile intensities of  $N = 3$  selected mucA22 produced biofilm technical replicate images. Pixel intensity is normalized within each LbL NP, and not against other NP systems.

## Patient Strain Biofilm Biomass

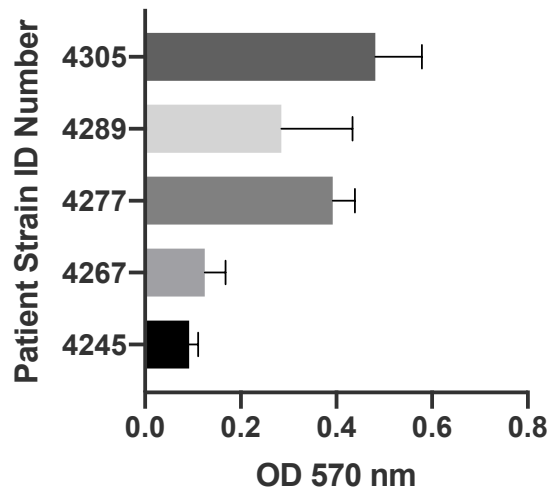


Figure S6. Crystal violet staining of patient strains. N = 3 biological replicates with at least 8 technical replicates in each, where bars and error bars are averages and standard deviations, respectively.

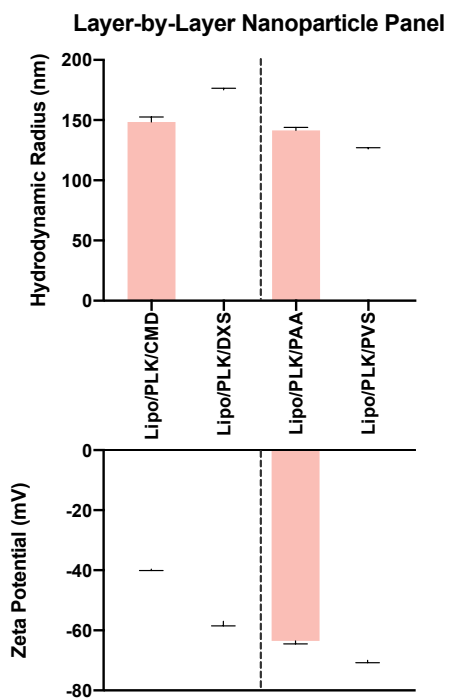


Figure S7. Size and zeta potential measurements of DXS and CMD, PVS and PAA are indistinguishable. N = 3 technical replicates for nanoparticle physiochemical characterization, where bars and error bars are averages and standard deviations, respectively.



Table S1. List of bacteria strains used, and their relative polysaccharide production during biofilm maturation. All strains besides PA14 are isogenic with a PAO1 background.

Strain name [shorthand name used]	Psl	Pel	alginate
PAO1 [PAO1]	+	+	+
PA14 [PA14]	-	+	+
$\Delta algD, \Delta pslA-O, \Delta pelA-G$ [ $\Delta\Delta\Delta$ ]	-	-	-
pDO300_mucA22 [mucA22]	+	+	+++
$\Delta wspF$ [ $\Delta wspF$ ]	+++	+++	+++
$\Delta algD, \Delta pelA-G, (pBADpslA-O)$ [ $\Delta\Delta psl$ ]	inducible	-	-
$\Delta algD, \Delta pslA-O, (pBADpelA-G)$ [ $\Delta\Delta pel$ ]	-	inducible	-
$\Delta algD, \Delta pelA-G, pslA-O (pAlgU)$ [ $\Delta\Delta alg$ ]	-	-	inducible