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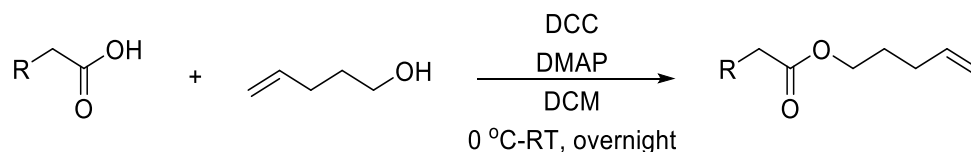
I. Supplemental Methods

1.1 General Information

All reagents were purchased from commercially available sources and used without further purification. All reactions were monitored by either ^1H NMR or thin layer chromatography (TLC) carried out on 0.25 mm pre-coated silica plates (F-254) purchased from Silicycle, Quebec, Canada, using shortwave UV light as visualizing agent and KMnO_4 or phosphomolybdic acid (PMA) as developing agents. Flash column chromatography was performed using SiliaFlash-P60 silica gel (40 – 63 μm) purchased from Silicycle, Quebec, Canada. ^1H , ^{13}C and ^{19}F NMR spectra were recorded on a Bruker DRX-600 spectrometers operating at 600 MHz for proton nuclei, 151 MHz for carbon nuclei and 565 MHz for fluorine nuclei were calibrated using residual undeuterated solvent as an internal reference (CDCl_3 : 7.26 ppm ^1H NMR and 77.00 ppm ^{13}C NMR). PR160L 390 nm LEDs (25% intensity) from Kessil Lights were used as light source. For reporting NMR peak multiplicities, the following abbreviations were used: s = singlet, d = doublet, t = triplet, q = quartet, quin = quintet, hept = heptet, m = multiplet. High-resolution mass spectra (HRMS) or mass spectra (MS) were recorded on an Agilent UHPLC TOF mass spectrometer using electrospray ionization time-of-flight (ESI-TOF), chemical ionization time-of-flight (CI-TOF), atmospheric pressure chemical ionization (APCI) or gas chromatography–mass spectrometry (GC-MS).

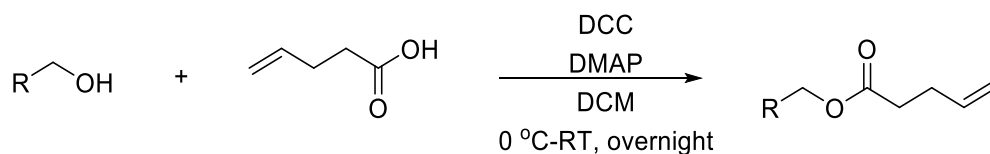
1.2 General Procedures for Substrate Synthesis

General Procedure 1 for the Synthesis of Unactivated Alkenes



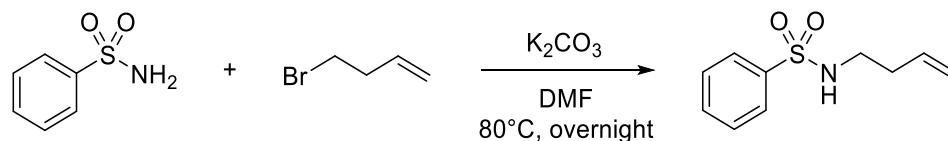
To an RB flask were added 5-pentenol (4.9 mmol, 1.96 equiv), carboxylic acid (2.5 mmol, 1.0 equiv), 4-dimethylamino pyridine (0.24 mmol, 0.097 equiv), and a stir bar. The RB flask was then evacuated and backfilled with nitrogen gas three times. Dry dichloromethane (0.225 M) was added via syringe to the RB flask, dissolving the solid components. The RB flask was then placed in an ice bath positioned on top of a stirring plate. Dicyclohexyl carbodiimide (4.85 mmol, 1.94 mmol) was added to the mixture via syringe dropwise over a period of 5 minutes. The ice bath was then removed, allowing the reaction to return to room temperature. The reaction was left to stir overnight. Following reaction, the mixture was concentrated through rotary evaporation. Subsequent flash column chromatography (hexanes/EtOAc) allowed for isolation of the ester.¹⁻²

General Procedure 2 for the Synthesis of Unactivated Alkenes



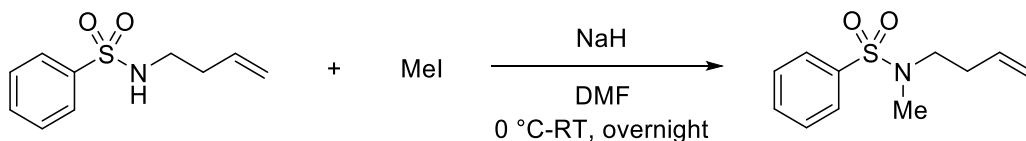
To an RB flask were added 5-pentenoic acid (2.5 mmol, 1.0 equiv), alcohol (4.9 mmol, 1.96 equiv), 4-dimethylamino pyridine (0.24 mmol, 0.097 equiv), and a stir bar. The RB flask was then evacuated and backfilled with nitrogen gas three times. Dry dichloromethane (0.225 M) was added via syringe to the RB flask, dissolving the solid components. The RB flask was then placed in an ice bath positioned on top of a stirring plate. Dicyclohexyl carbodiimide (4.85 mmol, 1.94 mmol) was added to the mixture via syringe dropwise over a period of 5 minutes. The ice bath was then removed, allowing the reaction to return to room temperature. The reaction was left to stir overnight. Following reaction, the mixture was concentrated through rotary evaporation. Subsequent flash column chromatography (hexanes/EtOAc) allowed for isolation of the ester.¹⁻²

Procedure for the Synthesis of *N*-(but-3-en-1-yl)benzenesulfonamide



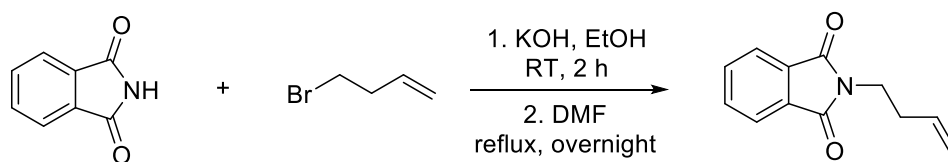
To an RB flask were added benzenesulfonamide (944 mg, 6.0 mmol), 4-bromobut-1-ene (0.8 mL, 6.0 mmol), dimethylformamide (30 mL), and a stir bar. Potassium carbonate (830 mg, 6.0 mmol) was added to the reaction mixture. After stirring overnight at 80 °C, the mixture was cooled to room temperature and quenched with water. The reaction mixture was then washed with brine and extracted with diethyl ether. The organic layer was concentrated through rotary evaporation. Subsequent flash column chromatography (hexanes/EtOAc) (3:1) allowed for isolation of *N*-(but-3-en-1-yl)benzenesulfonamide.³

Procedure for the Synthesis of *N*-(but-3-en-1-yl)-*N*-methylbenzenesulfonamide



To an RB flask were added sodium hydride (60% in mineral oil, 240 mg, 6 mmol), dimethylformamide (25 mL), a solution of *N*-(but-3-en-1-yl)benzenesulfonamide (1.20 g, 5 mmol) in DMF (5 mL), and a stir bar in an ice bath at 0 °C. The reaction mixture was brought to room temperature and stirred for 30 minutes. The reaction mixture was cooled to 0 °C in an ice bath again, and a solution of methyl iodide (1.06 g, 7.5 mmol) in DMF (5 mL) was added dropwise over a period of 5 minutes by syringe. The reaction mixture was brought to room temperature and left to run overnight. The reaction mixture was quenched with a saturated aqueous solution of sodium bicarbonate. The mixture was then washed with brine and extracted with diethyl ether. The organic phase was dried over sodium sulfate and concentrated through rotary evaporation. Subsequent flash column chromatography (hexanes/EtOAc) (10:1) produced *N*-(but-3-en-1-yl)-*N*-methylbenzenesulfonamide.⁴

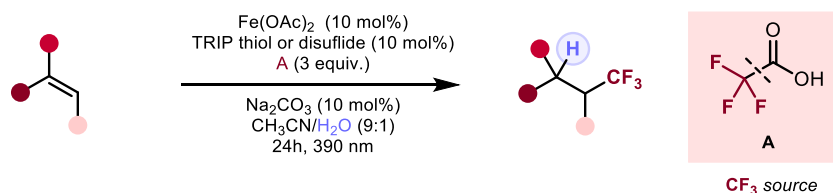
Procedure for the Synthesis of 2-(but-3-en-1-yl)isoindoline-1,3-dione



To an RB flask were added phthalimide (1.71 g, 11.6 mmol), potassium hydroxide (0.650 g, 11.6 mmol), ethyl alcohol (20 mL), and a stir bar. The reaction mixture was stirred at room temperature for 2 h and evaporated to remove EtOH. The resulting residue was then dissolved in dimethylformamide (15 mL) and 4-bromobut-1-ene (1.10 mL, 12.8 mmol) was added. The reaction mixture was stirred at reflux overnight. The reaction mixture was cooled, diluted with ethyl acetate, and quenched with saturated sodium bicarbonate. The mixture was then washed with brine. The extracted organic layer was dried over sodium sulfate and concentrated through rotary evaporation. Subsequent flash column chromatography (hexanes/EtOAc) (10:1) produced 2-(but-3-en-1-yl)isoindoline-1,3-dione.⁵

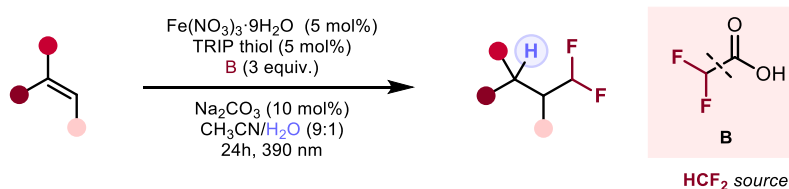
1.3 General Procedures for Photocatalytic Hydrofluoroalkylation

General Procedure A for hydrotrifluoromethylation of alkenes



$\text{Fe}(\text{OAc})_2$ (10 mol%, 0.1 equiv.), Na_2CO_3 (10 mol%, 0.1 equiv.) and TRIP disulfide (10 mol%, 0.1 equiv.) (in the case using TRIP thiol, HAT reagent was added via syringe after backfilling with N_2) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by addition of alkenes (0.1 mmol, 1.0 equiv.) and CF_3COOH (0.30 mmol, 3.0 equiv) in $\text{MeCN}/\text{H}_2\text{O}$ (9:1, 0.1 M in regard to alkenes) via syringe under N_2 . The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated, and the residue was then purified by flash column chromatography to give the corresponding hydrotrifluoromethylated products.

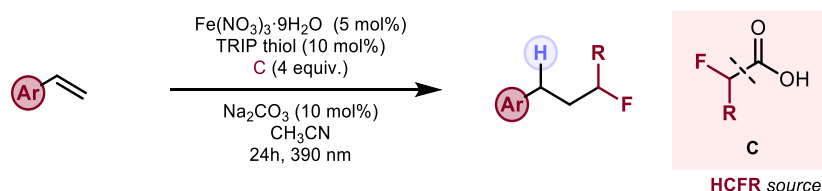
General Procedure B for hydrodifluoromethylation of alkenes



$\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ (5 mol%, 0.05 equiv.), Na_2CO_3 (10 mol%, 0.1 equiv.) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by addition of alkenes (0.1 mmol, 1.0 equiv.), HCF_2COOH (0.30 mmol, 3.0 equiv) and TRIP thiol (5 mol%, 0.05 equiv.) in $\text{MeCN}/\text{H}_2\text{O}$ (9:1, 0.1 M in regard to alkenes) via syringe under N_2 . The reaction mixture was placed under 390nm

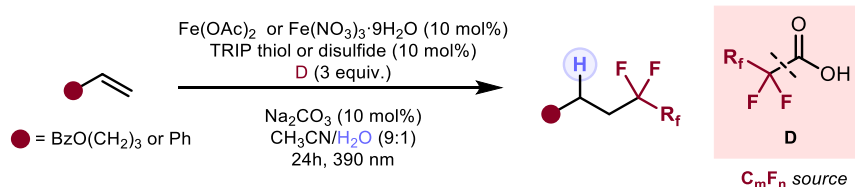
Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated, and the residue was then purified by flash column chromatography to give the corresponding hydrodifluoromethylated products.

General Procedure C for hydromonofluoromethylation of alkenes



$\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ (5 mol%, 0.05 equiv.), Na_2CO_3 (10 mol%, 0.1 equiv.) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by addition of alkenes (0.1 mmol, 1.0 equiv.), H_2CFCOOH (0.40 mmol, 4.0 equiv) and TRIP thiol (10 mol%, 0.1 equiv.) in MeCN (0.1 M in regard to alkenes) via syringe under N_2 . The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated, and the residue was then purified by flash column chromatography to give the corresponding hydromonofluoroalkylated products.

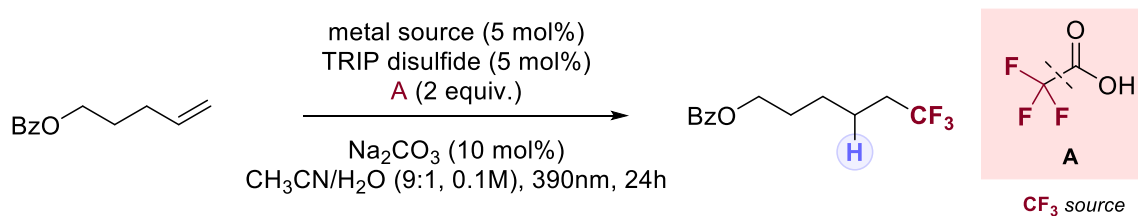
General Procedure D for hydro(perfluoro)alkylation of alkenes



Fe salt (10 mol%, 0.1 equiv.), Na₂CO₃ (10 mol%, 0.1 equiv.) and TRIP disulfide (10 mol%, 0.1 equiv.) (in the case using TRIP thiol, HAT reagent was added via syringe after backfilling with N₂) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N₂ (repeated for 4 times), followed by addition of alkenes (0.1 mmol, 1.0 equiv.) and (perfluoro)alkyl carboxylic acid (0.30 mmol, 3.0 equiv) in MeCN/H₂O (9:1, 0.1 M in regard to alkenes) via syringe under N₂. The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated, and the residue was then purified by flash column chromatography to give the corresponding hydroperfluoroalkylated products.

II. Supplemental Discussion

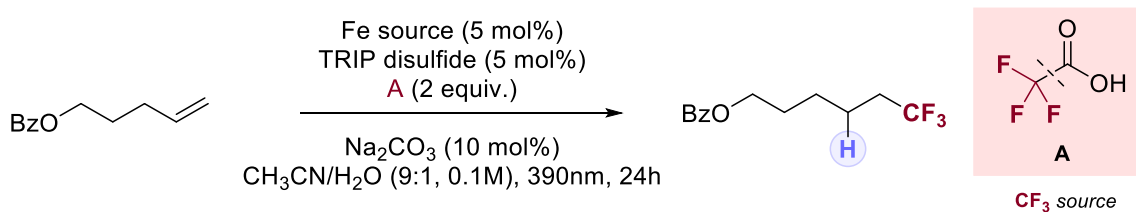
2.1 Optimization of Hydrotrifluoromethylation



entry	metal source	yield
1	Fe(acac) ₃	ND ^a
2	Fe(NO ₃) ₃ ·9H ₂ O	32
3	FeCl ₃ ·6H ₂ O	56
4	Fe(OTf) ₂	60
5	FeCl ₂	72
6	Fe(OAc) ₂	68
7	Cu(MeCN) ₄ BF ₄ ; Cu(OTf) ₂	ND ^a

^a Almost full recovery of starting material.

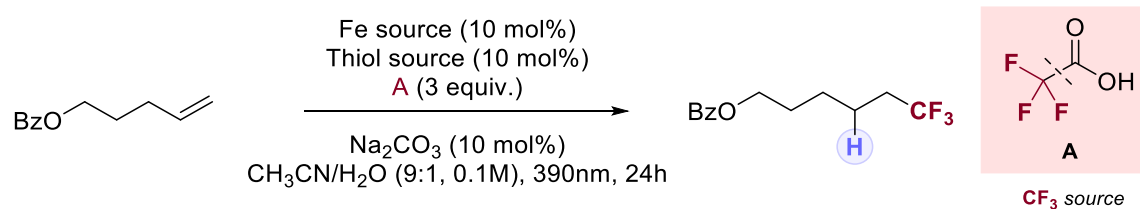
Supplemental Table 1. Catalyst screening.



entry	[Fe]	yield
a ₁	FeCl ₂	76
a,b ₂	FeCl ₂	80
a ₃	Fe(OAc) ₂	74
a,b ₄	Fe(OAc) ₂	92

^a10 mol% of [Fe] and 10 mol% of thiol/disulfide. ^bwith 3 equiv. of CF₃COOH.

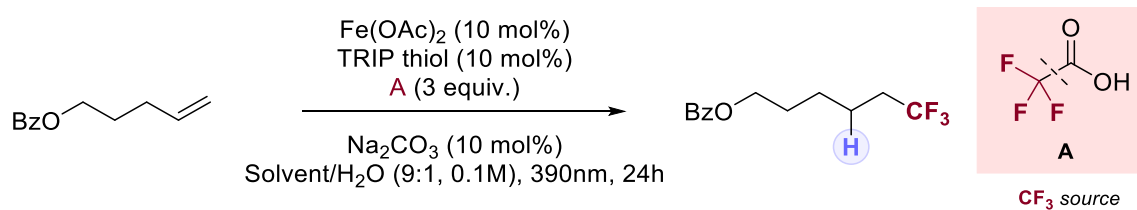
Supplemental Table 2. Catalyst, TFA loading screening.



entry	[Fe]	Thiol source	yield
1	Fe(OAc) ₂	TRIP thiol	86(84) ^a
2	Fe(OAc) ₂	TRIP disulfide	92(89) ^a

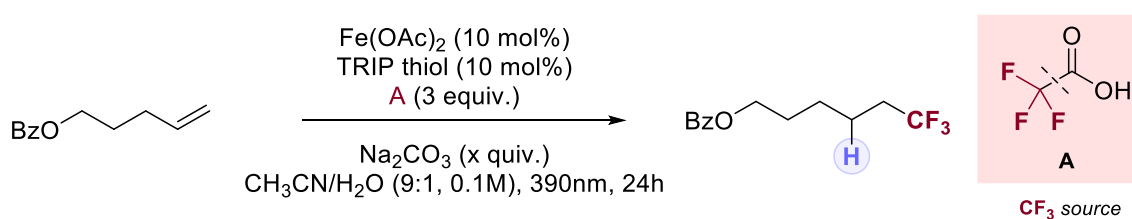
^a isolated yield in parentheses.

Supplemental Table 3. HAT reagents comparison in hydrotrifluoromethylation.



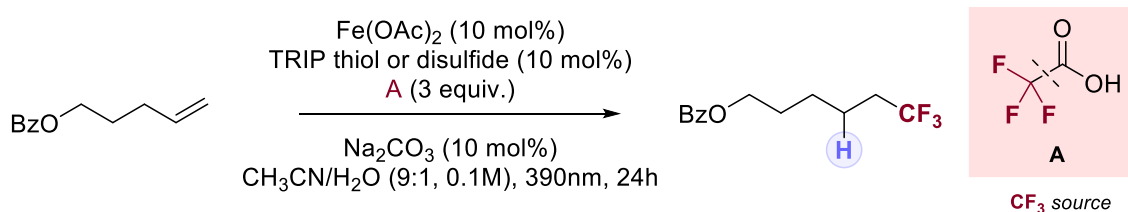
entry	Solvent	yield
1	DCM	trace
2	THF	20
3	EA	42
4	Acetone	76

Supplemental Table 4. Solvent screening.



entry	x	yield
1	no base	72
2	10 mol%	86
3	20 mol%	80
4	40 mol%	76
5	60 mol%	70
6	100 mol%	62

Supplemental Table 5. Base additive loading screening

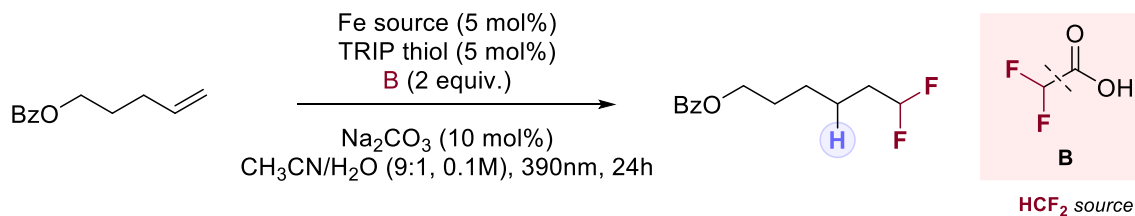


entry	Deviations from standard conditions	yield
1	no Fe	ND ^a ; ND ^b
2	no light	ND ^a ; ND ^b
3	no HAT reagent	trace ^a ; trace ^b
4	no water	18 ^a ; 24 ^b

^a with TRIP thiol. ^b with TRIP disulfide.

Supplemental Table 6. Control experiments.

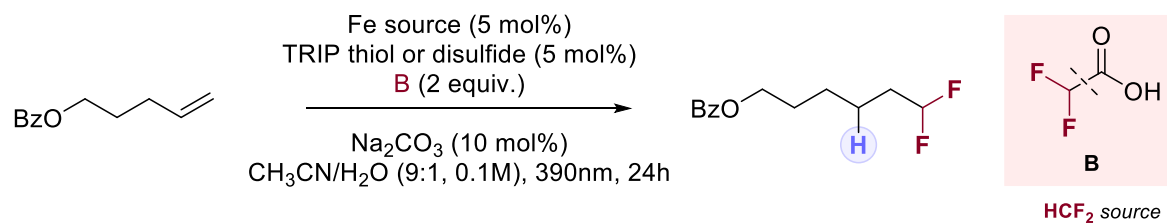
2.2 Optimization of Hydrodifluoromethylation



entry	[Fe] source	yield
1	Fe(NO ₃) ₃ 9H ₂ O	48
2	FeCl ₃ 6H ₂ O	38
3	Fe(OTf) ₃	36
4	FeCl ₂	44
5	Fe(OAc) ₂	42
^a 6	Fe(NO ₃) ₃ 9H ₂ O	35

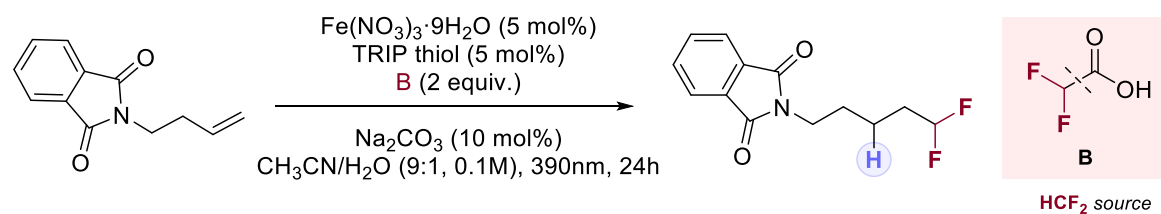
^a10 mol% of [Fe] and TRIP disulfide.

Supplemental Table 7. Iron catalyst screening



entry	[Fe]	Thiol source	yield
1	Fe(NO ₃) ₃ ·9H ₂ O	TRIP thiol	52
2	Fe(NO ₃) ₃ ·9H ₂ O	TRIP disulfide	48
3	Fe(OAc) ₂	TRIP thiol	46
4	Fe(OAc) ₂	TRIP disulfide	42

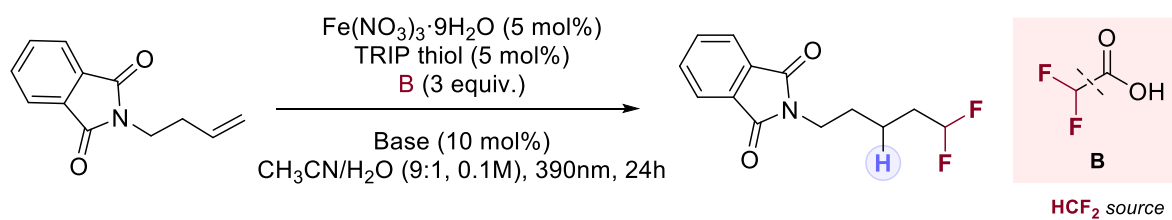
Supplemental Table 8. The comparison between TRIP thiol and TRIP disulfide as competent HAT reagents in hydrodifluoromethylation.



entry	[Fe]	yield
1	Fe(NO ₃) ₃ ·9H ₂ O	50
2	Fe(OAc) ₂	38
^a 3	Fe(NO ₃) ₃ ·9H ₂ O	76(75) ^b
^a 4	Fe(OAc) ₂	54

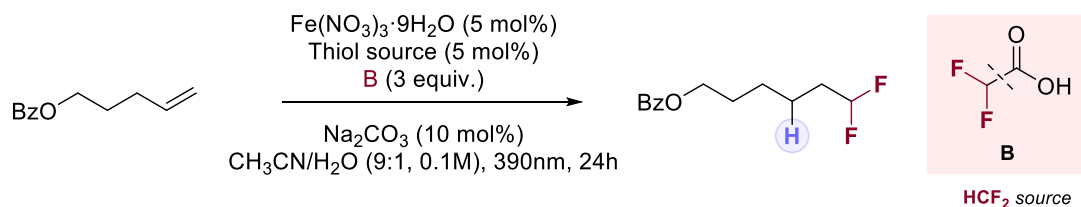
2-(but-3-en-1-yl)isoindoline-1,3-dione was used as substrate due to more precise NMR integration. ^a 3 equiv. of HCF₂COOH. ^b isolated yield in parentheses.

Supplemental Table 9. Loading of difluoroacetic acid screening



entry	Base	yield
1	Na_2CO_3	76
2	K_2CO_3	54
3	Cs_2CO_3	68

Supplemental Table 10. Base additive screening

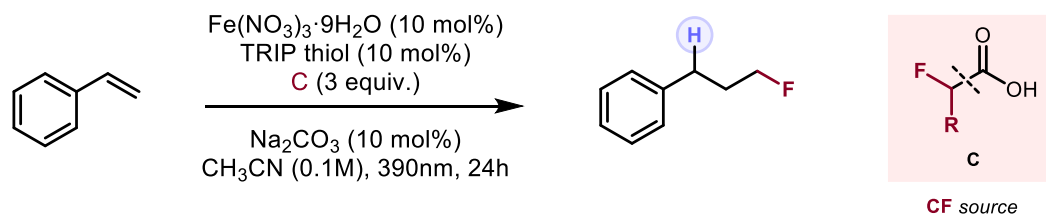


entry	thiol source	yield
1	4-CF ₃ -thiophenol	60
2	4-Cl-thiophenol	74
3	4-MeO-thiophenol	79
4	3,5-CF ₃ -thiophenol	51
5	2-MeO-thiophenol	58
6	2-CO ₂ Me-thiophenol	68
7	TRIP thiol	93(89) ^a

^aisolated yield in parentheses.

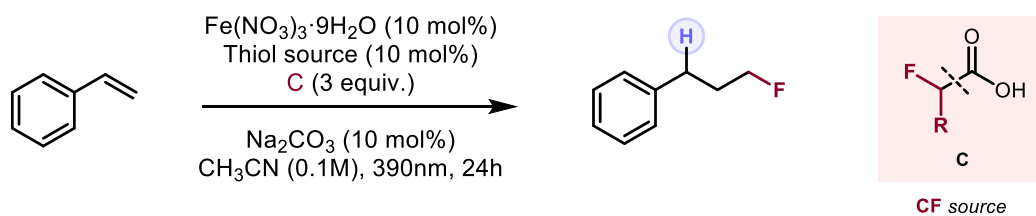
Supplemental Table 11. Aryl thiol screening (with pent-4-en-1-yl benzoate as substrate).

2.3 Optimization of Hydromonofluoromethylation



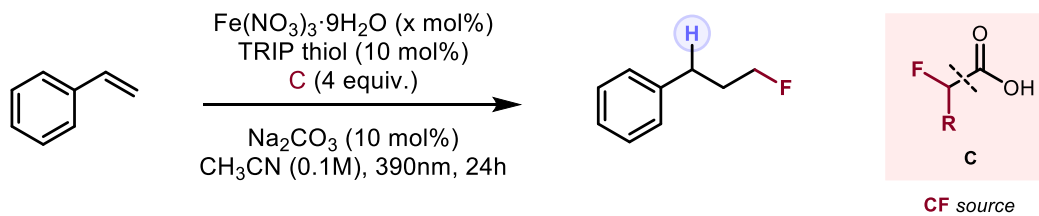
entry	[Fe]	yield
1	Fe(NO ₃) ₃ ·9H ₂ O	25
2	FeCl ₃ ·6H ₂ O	trace
3	Fe(OAc) ₂	13
4	Fe ₂ (C ₂ O ₄) ₃ ·6H ₂ O (5 mol%)	trace
5	Fe ₂ (SO ₄) ₃ ·5H ₂ O (5 mol%)	trace

Supplemental Table 12. Iron catalyst screening.



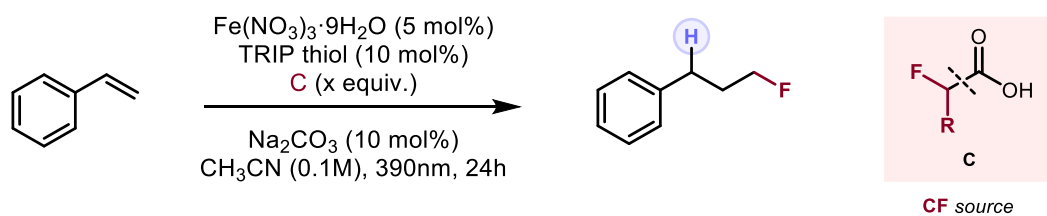
entry	[Fe]	yield
1	TRIP thiol	25
2	2-MeO-thiolphenol	messy
3	4-F-thiolphenol	messy
4	TRIP silanethiol (5 mol%)	messy
5	TRIP disulfide (5 mol%)	messy

Supplemental Table 13. Thiol/disulfide screening.



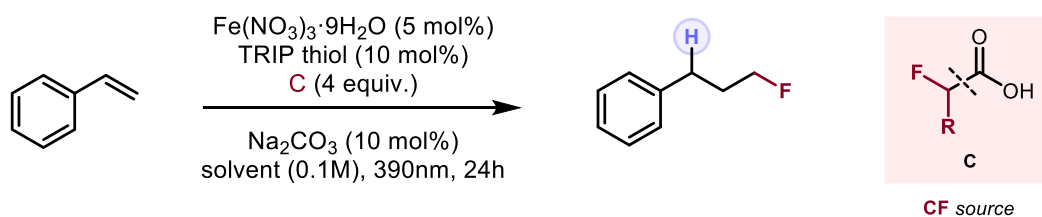
entry	x	yield
1	2.5	21
2	5	43
3	10	25

Supplemental Table 14. Catalyst loading screening.



entry	x	yield
1	3	28
2	4	43
3	6	42

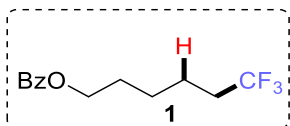
Supplemental Table 15. Monofluoroacetic acid loading screening.



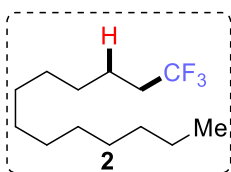
entry	Solvent	yield
1	CH_3CN	43
2	$\text{CH}_3\text{CN}/\text{H}_2\text{O}$ (9:1)	42
3	DCE; DCE/ H_2O (1:1)	ND

Supplemental Table 16. Solvent screening.

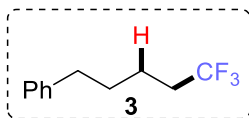
2.4 Characterization of Corresponding Products



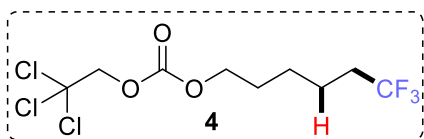
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.12 – 7.97 (m, 2H), 7.61 – 7.51 (m, 1H), 7.44 (t, J = 7.8 Hz, 2H), 4.34 (t, J = 6.5 Hz, 2H), 2.17 – 2.05 (m, 2H), 1.85 – 1.77 (m, 2H), 1.70 – 1.61 (m, 2H), 1.57 – 1.50 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.62, 132.94, 130.33, 129.55, 128.39, 127.13 (q, J = 276.3 Hz), 64.56, 33.67 (q, J = 28.6 Hz), 28.43, 25.32, 21.69 (q, J = 3.0 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.38 (t, J = 11.0 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.⁶



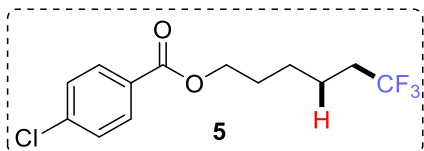
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 2.10 – 2.00 (m, 2H), 1.59-1.50 (m, 2H), 1.41 – 1.26 (m, 18H), 0.88 (t, J = 6.9 Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 127.35 (q, J = 276.2 Hz), 33.78 (q, J = 28.3 Hz), 31.96, 29.67, 29.63, 29.60, 29.41, 29.39, 29.23, 28.75, 22.73, 21.88 (q, J = 2.9 Hz), 14.13. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.48 (t, J = 11.0 Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{13}\text{H}_{25}\text{F}_3$: 238.1908; Found 238



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.29 (t, J = 7.6 Hz, 2H), 7.23 – 7.14 (m, 3H), 2.64 (t, J = 7.6 Hz, 2H), 2.16 – 2.03 (m, 2H), 1.74-1.66 (m, 2H), 1.63 – 1.57 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 141.69, 128.42, 128.36, 127.18 (q, J = 276.3 Hz), 125.95, 35.50, 33.63 (q, J = 28.4 Hz), 30.47, 21.54 (q, J = 2.9 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.32 (t, J = 11.0 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.⁷

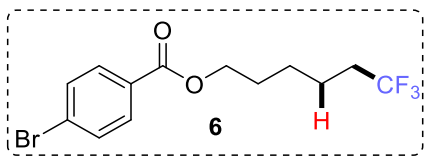


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 4.77 (s, 2H), 4.25 (t, J = 6.5 Hz, 2H), 2.15-2.03 (m, 2H), 1.81-1.70 (m, 2H), 1.66-1.58 (m, 2H), 1.55-1.44 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 154.03, 127.06 (q, J = 276.3 Hz), 94.46, 76.76, 68.74, 33.60 (q, J = 28.6 Hz), 28.22, 24.86, 21.57 (q, J = 2.9 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.39 (t, J = 10.7 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_9\text{H}_{13}\text{Cl}_3\text{F}_3\text{O}_3$: 330.9877; Found 330.9870

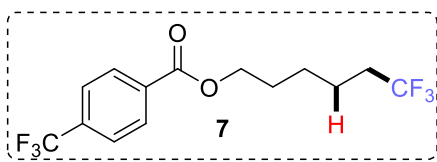


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.03 – 7.92 (m, 2H), 7.49 – 7.38 (m, 2H), 4.33 (t, J = 6.5 Hz, 2H), 2.18 – 2.03 (m, 2H), 1.87 – 1.74 (m, 2H), 1.69-1.60 (m, 2H), 1.56 – 1.48 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 165.75, 139.41, 130.94, 128.74, 127.11 (q, J = 276.3 Hz), 64.82, 33.64 (q, J = 28.4 Hz), 28.37, 25.28, 21.67 (q, J = 2.8 Hz). ^{19}F NMR

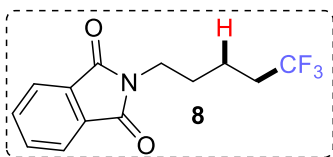
(565 MHz, Chloroform-*d*) δ -66.36 (t, J = 11.0 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.⁸



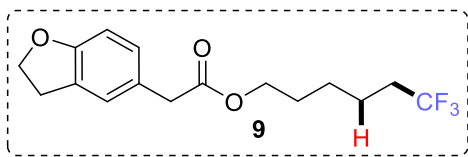
Prepared according to General Procedure A and obtained as colorless oil. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.94 – 7.85 (m, 2H), 7.63 – 7.55 (m, 2H), 4.33 (t, J = 6.5 Hz, 2H), 2.17 – 2.04 (m, 2H), 1.84 – 1.76 (m, 2H), 1.70 – 1.62 (m, 2H), 1.56 – 1.48 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 165.88, 131.74, 131.08, 129.20, 128.07, 127.10 (q, J = 276.2 Hz), 64.84, 33.63 (q, J = 28.5 Hz), 28.36, 25.27, 21.67 (q, J = 3.0 Hz). ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -66.36 (t, J = 10.8 Hz). HRMS APCI: [M+H]⁺ calcd. for C₁₃H₁₅BrF₃O₂: 339.0202; Found 339.0199



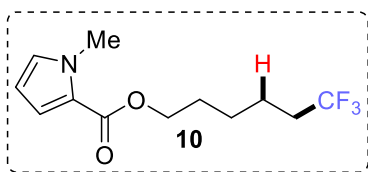
Prepared according to General Procedure A and obtained as colorless oil. ¹H NMR (600 MHz, Chloroform-*d*) δ 8.15 (d, J = 8.1 Hz, 2H), 7.71 (d, J = 8.2 Hz, 2H), 4.37 (t, J = 6.5 Hz, 2H), 2.20 – 2.04 (m, 2H), 1.87-1.79 (m, 2H), 1.71 – 1.60 (m, 2H), 1.58-1.50 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 165.39, 134.49 (q, J = 32.7 Hz), 133.52, 129.96, 127.10 (q, J = 276.3 Hz), 125.45 (q, J = 3.8 Hz), 123.65 (q, J = 272.8 Hz), 65.14, 33.64 (q, J = 28.6 Hz), 28.35, 25.26, 21.68 (q, J = 2.8 Hz). ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -63.13, -66.37 (t, J = 11.0 Hz). HRMS APCI: [M+H]⁺ calcd. for C₁₃H₁₅F₆O₂: 329.0971; Found 329.0967



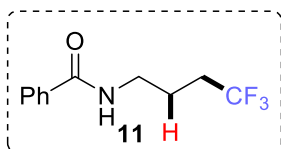
Prepared according to General Procedure A and obtained as white solid. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.85 (dd, J = 5.4, 3.0 Hz, 2H), 7.73 (dd, J = 5.5, 3.0 Hz, 2H), 3.72 (t, J = 7.1 Hz, 2H), 2.21 – 2.07 (m, 2H), 1.82-1.73 (m, 2H), 1.66-1.57 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 168.37, 134.04, 132.05, 126.97 (q, J = 276.4 Hz), 123.31, 37.27, 33.25 (q, J = 28.7 Hz), 27.66, 19.31 (q, J = 3.0 Hz). ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -66.35 (t, J = 11.0 Hz). m.p.: 75-76 °C. The compound characterization was reported in literature and the NMR data matched with previous characterization.⁹



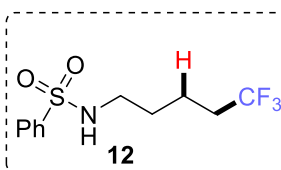
Prepared according to General Procedure A and obtained as colorless oil. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.14 – 7.09 (m, 1H), 6.99 (dd, J = 8.2, 1.9 Hz, 1H), 6.73 (d, J = 8.1 Hz, 1H), 4.56 (t, J = 8.7 Hz, 2H), 4.09 (t, J = 6.5 Hz, 2H), 3.53 (s, 2H), 3.19 (t, J = 8.7 Hz, 2H), 2.13 – 1.98 (m, 2H), 1.69-1.60 (m, 3H), 1.60 – 1.53 (m, 2H), 1.43-1.36 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 172.15, 159.27, 128.85, 127.39, 127.11 (q, J = 276.5 Hz), 125.91, 125.78, 109.19, 71.29, 64.35, 40.77, 33.60 (q, J = 28.4 Hz), 29.70, 28.20, 25.11, 21.56 (q, J = 3.0 Hz). ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -66.36 (t, J = 11.0 Hz). HRMS APCI: [M+H]⁺ calcd. for C₁₆H₂₀F₃O₃: 317.1359; Found 317.1355



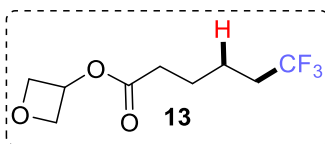
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 6.93 (dd, $J = 4.0, 1.8$ Hz, 1H), 6.78 (t, $J = 2.2$ Hz, 1H), 6.11 (dd, $J = 3.9, 2.5$ Hz, 1H), 4.23 (t, $J = 6.5$ Hz, 2H), 3.92 (s, 3H), 2.18 – 2.01 (m, 2H), 1.79 – 1.71 (m, 2H), 1.67-1.59 (m, 2H), 1.55-1.46 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 161.32, 129.52, 127.14 (q, $J = 276.3$ Hz), 122.51, 117.72, 107.83, 63.34, 36.81, 33.67 (q, $J = 28.5$ Hz), 28.52, 25.33, 21.66 (q, $J = 3.0$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.38 (t, $J = 11.0$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{12}\text{H}_{17}\text{F}_3\text{NO}_2$: 264.1206; Found 264.1201



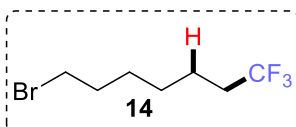
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.76 (d, $J = 7.7$ Hz, 2H), 7.51 (t, $J = 7.4$ Hz, 1H), 7.44 (t, $J = 7.6$ Hz, 2H), 6.28 (s, 1H), 3.60-3.48 (m, 2H), 2.26 – 2.11 (m, 2H), 1.95-1.86 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 167.78, 134.28, 131.68, 127.90, 126.98 (q, $J = 276.2$ Hz), 126.85, 38.77, 31.36 (q, $J = 29.1$ Hz), 22.59 (q, $J = 2.8$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.13 (t, $J = 10.8$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.⁸



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.87 (d, $J = 7.7$ Hz, 2H), 7.60 (t, $J = 7.4$ Hz, 1H), 7.53 (t, $J = 7.6$ Hz, 2H), 4.61 (t, $J = 6.4$ Hz, 1H), 2.99 (q, $J = 5.9$ Hz, 2H), 2.07-1.97 (m, 2H), 1.59 – 1.53 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 139.81, 132.79, 129.21, 127.01, 126.88 (q, $J = 276.3$ Hz), 42.70, 33.16 (q, $J = 28.7$ Hz), 28.73, 19.05 (q, $J = 3.2$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.34 (t, $J = 11.0$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{15}\text{F}_3\text{NO}_2\text{S}$: 282.0770; Found 282.0765

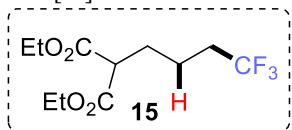


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.50 – 5.40 (m, 1H), 4.89 (ddd, $J = 7.5, 6.3, 1.0$ Hz, 2H), 4.63 (ddd, $J = 7.5, 5.3, 1.0$ Hz, 2H), 2.40 (t, $J = 7.3$ Hz, 2H), 2.16 – 2.04 (m, 2H), 1.76-1.68 (m, 2H), 1.65 – 1.57 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.27, 126.95 (q, $J = 276.3$ Hz), 77.54, 67.92, 33.45 (q, $J = 28.8$ Hz), 33.41, 23.77, 21.43 (q, $J = 2.9$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.37 (t, $J = 10.6$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_9\text{H}_{14}\text{F}_3\text{O}_3$: 227.0890; Found 227.0885

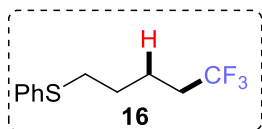


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 3.41 (t, $J = 6.7$ Hz, 2H), 2.12 – 2.04 (m, 2H), 1.91-1.81 (m, 2H), 1.64-1.54 (m, 2H), 1.53 – 1.44 (m, 2H), 1.44-1.37 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 127.19 (q, $J = 276.3$ Hz), 33.63 (q, $J = 28.3$ Hz),

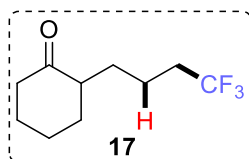
33.60, 32.44, 27.86, 27.74, 21.76 (q, $J = 3.0$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -66.41 (t, $J = 11.0$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_7\text{H}_{12}\text{BrF}_3$: 232.0074; Found 232.0



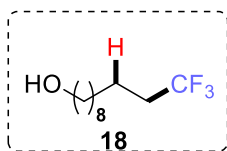
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 4.28-4.15 (m, 4H), 3.33 (t, $J = 7.4$ Hz, 1H), 2.19 – 2.06 (m, 2H), 1.97 (q, $J = 7.7$ Hz, 2H), 1.67-1.58 (m, 2H), 1.27 (t, $J = 7.1$ Hz, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 168.99, 126.84 (q, $J = 276.3$ Hz), 61.57, 51.59, 33.47 (q, $J = 28.8$ Hz), 27.69, 19.89 (q, $J = 3.1$ Hz), 14.06. ^{19}F NMR (565 MHz, Chloroform- d) δ -66.40 (t, $J = 10.9$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{18}\text{F}_3\text{O}_4$: 271.1152; Found 271.1146



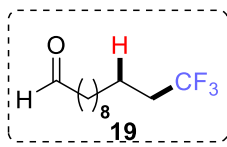
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.36 – 7.32 (m, 2H), 7.31-7.27 (m, 2H), 7.22 – 7.16 (m, 1H), 2.98 – 2.89 (m, 2H), 2.15 – 2.01 (m, 2H), 1.76-1.66 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 136.13, 129.44, 128.96, 127.01 (q, $J = 276.4$ Hz), 126.15, 33.34 (q, $J = 28.8$ Hz), 33.32, 28.16, 21.06 (q, $J = 3.0$ Hz). ^{19}F NMR (565 MHz,) δ -66.35 (t, $J = 10.8$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{11}\text{H}_{13}\text{F}_3\text{S}$: 234.0690; Found 234.1



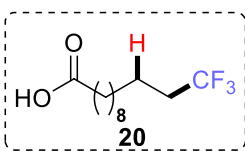
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 2.44-2.37 (m, 1H), 2.34 – 2.24 (m, 2H), 2.16 – 1.99 (m, 4H), 1.91 – 1.80 (m, 2H), 1.71-1.64 (m, 2H), 1.57 – 1.51 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 212.61, 127.11 (q, $J = 276.3$ Hz), 50.41, 42.14, 33.98, 33.97 (d, $J = 28.5$ Hz), 28.58, 28.00, 25.08, 19.78 (q, $J = 3.1$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -66.38 (t, $J = 10.7$ Hz). GC-MS: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{10}\text{H}_{16}\text{F}_3\text{O}$: 209.1148; Found 209.0



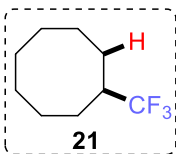
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 3.57 (t, $J = 6.7$ Hz, 2H), 2.04 – 1.92 (m, 2H), 1.53-1.44 (m, 4H), 1.34-1.19 (m, 13H). ^{13}C NMR (151 MHz, CDCl_3) δ 127.30 (q, $J = 276.3$ Hz), 63.04, 33.72 (q, $J = 28.3$ Hz), 32.77, 29.48, 29.37, 29.28, 29.16, 28.68, 25.72, 21.83 (q, $J = 2.8$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -66.44 (t, $J = 11.0$ Hz). HRMS APCI: $[\text{M}-\text{H}_2\text{O}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{20}\text{F}_3$: 209.1512; Found 209.1507



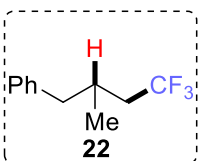
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 9.77 (s, 1H), 2.42 (t, $J = 7.4$ Hz, 2H), 2.10-2.00 (m, 2H), 1.66-1.60 (m, 2H), 1.55-1.49 (m, 2H), 1.39 – 1.26 (m, 16H). ^{13}C NMR (151 MHz, CDCl_3) δ 202.91, 127.30 (d, $J = 276.2$ Hz), 43.91, 33.72 (q, $J = 28.3$ Hz), 29.30, 29.26, 29.14, 29.13, 28.67, 22.06, 21.83 (q, $J = 2.9$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -66.41 (t, $J = 11.0$ Hz). GC-MS: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{13}\text{F}_3\text{S}$: 239.1618; Found 239.0



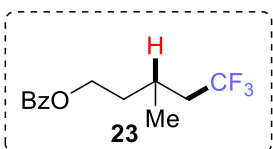
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 2.35 (t, $J = 7.5$ Hz, 2H), 2.10-2.02 (m, 2H), 1.68-1.59 (m, 2H), 1.59-1.50 (m, 2H), 1.38 – 1.25 (m, 14H). ^{13}C NMR (151 MHz, CDCl_3) δ 179.46, 127.31 (q, $J = 276.3$ Hz), 33.92, 33.73 (d, $J = 28.3$ Hz), 29.30, 29.27, 29.17, 29.14, 29.02, 28.68, 24.66, 21.83 (t, $J = 3.0$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.42 (t, $J = 11.0$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.⁸



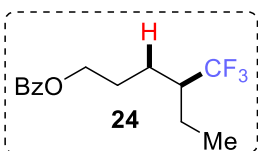
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 2.27-2.18 (m, 1H), 1.90-1.81 (m, 2H), 1.80 – 1.71 (m, 2H), 1.62 – 1.49 (m, 10H). ^{13}C NMR (151 MHz, CDCl_3) δ 128.97 (q, $J = 279.5$ Hz), 41.92 (q, $J = 24.4$ Hz), 26.38, 26.29, 25.57 (q, $J = 2.6$ Hz), 25.11. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -73.34 (d, $J = 9.8$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁰



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.30 (t, $J = 7.5$ Hz, 2H), 7.22 (t, $J = 7.4$ Hz, 1H), 7.15 (d, $J = 7.5$ Hz, 2H), 2.67 (dd, $J = 13.6, 6.6$ Hz, 1H), 2.53 (dd, $J = 13.6, 7.5$ Hz, 1H), 2.21 – 2.09 (m, 2H), 1.97 – 1.86 (m, 1H), 1.02 (d, $J = 6.4$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 139.56, 129.17, 128.42, 127.19 (q, $J = 277.5$ Hz), 126.29, 43.22, 39.54 (q, $J = 27.1$ Hz), 29.65 (q, $J = 2.3$ Hz), 19.61. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -63.11 (t, $J = 11.2$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{11}\text{H}_{13}\text{F}_3$: 202.0969; Found 202.1

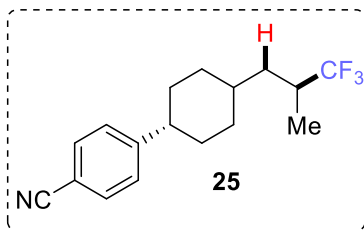


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.06 – 8.00 (m, 2H), 7.59 – 7.54 (m, 1H), 7.47-7.41 (m, 2H), 4.42 – 4.34 (m, 2H), 2.26-2.15 (m, 1H), 2.15 – 2.08 (m, 1H), 2.08-1.97 (m, 1H), 1.97 – 1.88 (m, 1H), 1.76-1.66 (m, 1H), 1.12 (d, $J = 6.7$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 164.67, 131.13, 128.30, 127.66, 126.54, 125.11 (q, $J = 277.4$ Hz), 60.59, 38.33 (q, $J = 27.2$ Hz), 33.41, 23.22 (q, $J = 2.5$ Hz), 17.75. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -63.30 (t, $J = 11.1$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.⁶

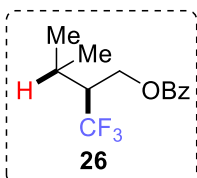


Prepared according to General Procedure A and obtained as colorless oil. 1.3:1 mixture of regioisomers. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.00-7.91 (m, 2H), 7.52-7.46 (m, 1H), 7.41-7.31 (m, 2H), 4.38-4.29 (m, 0.84H), 4.28-4.23 (m, 1.16H), 2.24-2.18 (m, 0.38H), 2.09 – 1.92 (m, 1.14H), 1.89 – 1.75 (m, 1.74H), 1.73 – 1.51 (m, 2.63H), 1.51 – 1.32 (m, 2.09H), 0.96-0.90 (m, 1.74H), 0.89-0.83 (m, 1.26H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.57, 166.41, 133.09, 132.98, 130.23, 130.03, 129.55, 129.53, 128.51 (q, $J = 280.6$ Hz), 128.43, 128.36 (q, $J = 280.2$ Hz), 128.40, 64.57, 62.35, 43.63 (q, $J = 24.8$ Hz), 39.69 (q, $J = 25.7$ Hz), 30.09 (q, $J = 2.5$ Hz), 27.24 (q, $J = 2.6$ Hz),

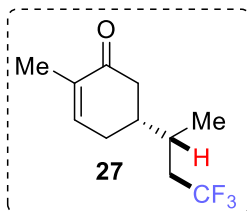
26.05, 23.83 (q, $J = 2.6$ Hz), 20.66 (q, $J = 2.7$ Hz), 19.95, 14.00, 11.16. ^{19}F NMR (565 MHz, Chloroform- d) δ -69.85 (d, $J = 9.6$ Hz), -70.50 (d, $J = 9.7$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{18}\text{F}_3\text{O}_2$: 275.1253; Found 275.1247



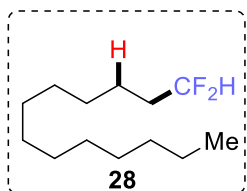
Prepared according to General Procedure A and obtained as colorless oil. 7:1 mixture of regioisomers. ^1H NMR (600 MHz, Chloroform- d) δ 7.58 (d, $J = 8.0$ Hz, 1H), 7.30 (d, $J = 8.3$ Hz, 1H), 2.60-2.50 (m, 1H), 2.32 – 2.21 (m, 0.78H), 1.98 – 1.84 (m, 4.25H), 1.69-1.62 (m, 0.5H), 1.58-1.50 (m, 1.21H), 1.50 – 1.40 (m, 3.12H), 1.32-1.26 (m, 1H), 1.20-1.13 (m, 0.86H), 1.13-1.09 (m, 2.23H), 1.06 – 1.00 (m, 1.37H). ^{13}C NMR (151 MHz, CDCl_3) δ 152.80, 152.55, 132.28, 132.25, 128.69 (q, $J = 282.1$ Hz), 128.63 (q, $J = 279.2$ Hz), 127.66, 127.63, 119.13, 119.11, 109.86, 109.80, 49.41 (q, $J = 23.3$ Hz), 44.56, 44.31, 36.71 (q, $J = 2.4$ Hz), 36.54 (q, $J = 1.6$ Hz), 35.15 (q, $J = 26.2$ Hz), 34.00, 33.95, 33.89, 33.80, 33.64, 33.47, 32.08, 30.11, 29.31, 18.45 (q, $J = 2.7$ Hz), 12.95 (q, $J = 3.2$ Hz), 12.83. ^{19}F NMR (565 MHz, Chloroform- d) δ -65.61 (d, $J = 10.3$ Hz), -73.47 (d, $J = 8.8$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{17}\text{H}_{21}\text{F}_3\text{N}$: 296.1621; Found 296.1615



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.04 (d, $J = 7.7$ Hz, 2H), 7.58 (t, $J = 7.4$ Hz, 1H), 7.45 (t, $J = 7.6$ Hz, 2H), 4.52 (qd, $J = 11.9, 5.5$ Hz, 2H), 2.53-2.41 (m, 1H), 2.39-2.19 (m, 1H), 1.15 (d, $J = 7.0$ Hz, 3H), 1.08 (d, $J = 7.0$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.22, 133.24, 129.71, 129.67, 128.47, 127.33 (d, $J = 281.7$ Hz), 59.96, 59.94, 47.97 (q, $J = 24.1$ Hz), 25.65 (d, $J = 2.2$ Hz), 20.92, 19.02. ^{19}F NMR (565 MHz, Chloroform- d) δ -65.99 (d, $J = 9.8$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹¹

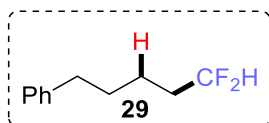


Prepared according to General Procedure A and obtained as colorless oil. 1:1 mixture of diastereomers. ^1H NMR (600 MHz, Chloroform- d) δ 6.74 (d, $J = 6.1$, 1H), 2.52-2.43 (m, 1H), 2.38-2.27 (m, 1H), 2.23 – 2.02 (m, 4H), 1.99 – 1.87 (m, 2H), 1.82 – 1.76 (m, 3H), 1.05 (dd, $J = 6.6, 4.2$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 199.43, 199.39, 144.40, 144.31, 135.68, 135.65, 127.10 (q, $J = 277.3$ Hz), 41.91, 40.33, 40.18, 40.13, 37.77 (q, $J = 27.5$ Hz), 37.58 (q, $J = 27.7$ Hz), 31.47 (q, $J = 2.2$ Hz), 31.40 (q, $J = 2.2$ Hz), 29.90, 28.29, 16.32, 16.21, 15.66, 15.64. ^{19}F NMR (565 MHz, Chloroform- d) δ -63.52 (t, $J = 11.0$ Hz), -63.54 (t, $J = 11.0$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹²

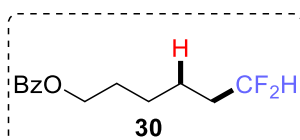


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 5.79 (tt, $J = 57.0, 4.6$ Hz, 1H), 1.88-1.73 (m, 2H), 1.48-1.40 (m, 2H), 1.37 – 1.25 (m, 18H), 0.88 (t,

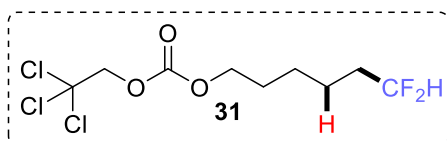
$J = 6.9$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 117.52 (t, $J = 238.6$ Hz), 34.10 (t, $J = 20.6$ Hz), 31.92, 29.65, 29.63, 29.59, 29.44, 29.37, 29.35, 29.07, 22.69, 22.13 (t, $J = 5.4$ Hz), 14.11. ^{19}F NMR (565 MHz, Chloroform- d) δ -115.73 (dt, $J = 57.1$, 17.6 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹³



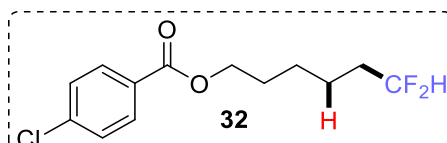
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.31 – 7.26 (m, 2H), 7.21 – 7.14 (m, 3H), 5.78 (tt, $J = 56.9$, 4.5 Hz, 1H), 2.68 – 2.58 (m, 2H), 1.92 – 1.78 (m, 2H), 1.75 – 1.61 (m, 2H), 1.52 – 1.44 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 141.98, 128.38, 125.87, 117.34 (t, $J = 238.7$ Hz), 35.67, 33.98 (t, $J = 20.7$ Hz), 30.87, 21.78 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.72 (dt, $J = 56.4$, 17.3 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁴



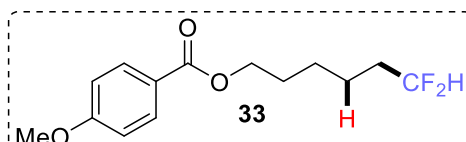
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.07 – 8.01 (m, 2H), 7.58 – 7.54 (m, 1H), 7.47 – 7.41 (m, 2H), 5.81 (tt, $J = 56.8$, 4.4 Hz, 1H), 4.33 (t, $J = 6.5$ Hz, 2H), 1.92 – 1.75 (m, 4H), 1.58 – 1.46 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.64, 132.91, 130.38, 129.54, 128.37, 117.23 (t, $J = 238.9$ Hz), 64.68, 33.98 (t, $J = 20.7$ Hz), 28.56, 25.64, 21.83 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.88 (dt, $J = 57.0$, 17.6 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{17}\text{F}_2\text{O}_2$: 243.1191; Found 243.1190



Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 5.81 (tt, $J = 56.8$, 4.4 Hz, 1H), 4.77 (s, 2H), 4.24 (t, $J = 6.6$ Hz, 2H), 1.90–1.79 (m, 2H), 1.78 – 1.72 (m, 2H), 1.54 – 1.44 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 154.04, 117.13 (t, $J = 238.8$ Hz), 94.47, 76.75, 68.90, 33.89 (t, $J = 20.8$ Hz), 28.35, 25.19, 21.69 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.96 (dt, $J = 56.8$, 17.4 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_9\text{H}_{14}\text{Cl}_3\text{F}_2\text{O}_3$: 312.9971; Found 312.9967

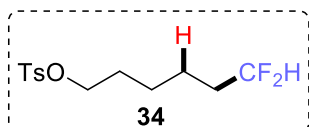


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.02 – 7.92 (m, 2H), 7.47 – 7.37 (m, 2H), 5.81 (tt, $J = 56.9$, 4.4 Hz, 1H), 4.32 (t, $J = 6.6$ Hz, 2H), 1.91 – 1.76 (m, 4H), 1.57 – 1.47 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 165.77, 139.37, 130.94, 128.80, 128.73, 117.20 (t, $J = 238.9$ Hz), 64.94, 33.95 (t, $J = 20.9$ Hz), 28.51, 25.61, 21.79 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.90 (dt, $J = 57.2$, 18.0 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{16}\text{ClF}_2\text{O}_2$: 277.0801; Found 277.0796

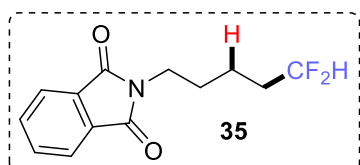


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.04 – 7.94 (m, 2H), 6.99 – 6.86 (m, 2H), 5.81 (tt, $J = 56.9$, 4.5 Hz, 1H),

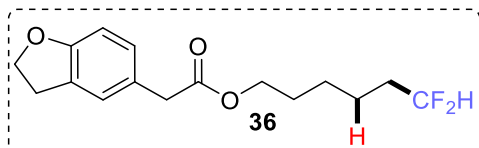
4.30 (t, $J = 6.5$ Hz, 2H), 3.86 (s, 3H), 1.91-1.82 (m, 2H), 1.81 – 1.76 (m, 2H), 1.57-1.47 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.40, 163.34, 131.55, 122.81, 117.25 (t, $J = 238.7$ Hz), 113.61, 64.38, 55.44, 33.98 (t, $J = 20.8$ Hz), 28.60, 25.66, 21.84 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.86 (dt, $J = 56.7, 17.4$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{19}\text{F}_2\text{O}_3$: 273.1297; Found 273.1293



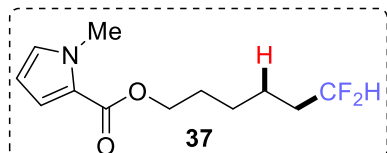
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.83 – 7.75 (m, 2H), 7.39 – 7.32 (m, 2H), 5.76 (tt, $J = 56.8, 4.4$ Hz, 1H), 4.03 (t, $J = 6.3$ Hz, 2H), 2.45 (s, 3H), 1.82 – 1.72 (m, 2H), 1.70-1.63 (m, 2H), 1.44 – 1.34 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 144.81, 133.11, 129.89, 129.87, 127.88, 117.06 (t, $J = 238.9$ Hz), 70.14, 33.81 (t, $J = 20.8$ Hz), 28.62, 24.93, 21.63, 21.44 (t, $J = 5.6$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -116.00 (dt, $J = 57.0, 17.7$ Hz). HRMS APCI: $[\text{M}+\text{NH}_4]^+$ calcd. for $\text{C}_{13}\text{H}_{22}\text{F}_2\text{NO}_3\text{S}$: 310.1283; Found 310.1279



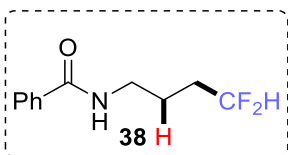
Prepared according to General Procedure B and obtained as white solid. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.85 (dd, $J = 5.4, 3.0$ Hz, 2H), 7.72 (dd, $J = 5.5, 3.0$ Hz, 2H), 5.80 (tt, $J = 56.7, 4.4$ Hz, 1H), 3.71 (t, $J = 7.2$ Hz, 2H), 1.95 – 1.83 (m, 2H), 1.79-1.72 (m, 2H), 1.55 – 1.49 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 168.40, 133.99, 132.08, 123.26, 117.03 (t, $J = 239.0$ Hz), 37.49, 33.55 (t, $J = 21.0$ Hz), 28.04, 19.44 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.97 (dt, $J = 56.8, 17.5$ Hz). m.p.: 76-77 °C. HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{14}\text{F}_2\text{NO}_2$: 254.0987; Found 254.0984



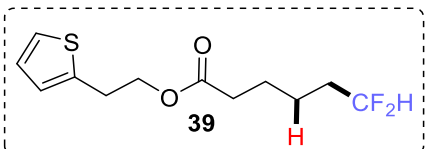
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.11 (d, $J = 2.0$ Hz, 1H), 6.99 (dd, $J = 8.1, 1.9$ Hz, 1H), 6.72 (d, $J = 8.1$ Hz, 1H), 5.78 (tt, $J = 56.8, 4.4$ Hz, 1H), 4.55 (t, $J = 8.7$ Hz, 2H), 4.09 (t, $J = 6.6$ Hz, 2H), 3.53 (s, 2H), 3.19 (t, $J = 8.7$ Hz, 2H), 1.86-1.74 (m, 2H), 1.69 – 1.60 (m, 2H), 1.50 – 1.40 (m, 2H), 1.40-1.34 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.16, 159.26, 128.85, 127.38, 125.95, 125.78, 117.21 (t, $J = 238.8$ Hz), 109.17, 71.29, 64.47, 40.78, 33.92 (t, $J = 20.8$ Hz), 29.70, 28.35, 25.43, 21.72 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.86 (dt, $J = 56.5, 17.8$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{21}\text{F}_2\text{O}_3$: 299.1453; Found 299.1448



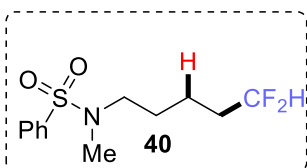
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 6.93 (dd, $J = 4.0, 1.8$ Hz, 1H), 6.78 (t, $J = 2.2$ Hz, 1H), 6.11 (dd, $J = 3.9, 2.5$ Hz, 1H), 5.81 (tt, $J = 56.9, 4.5$ Hz, 1H), 4.22 (t, $J = 6.5$ Hz, 2H), 3.92 (s, 3H), 1.91 – 1.79 (m, 2H), 1.78-1.71 (m, 2H), 1.55 – 1.44 (m, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 161.35, 129.48, 122.56, 117.70, 117.26 (t, $J = 238.7$ Hz), 107.82, 63.46, 36.81, 33.99 (t, $J = 20.7$ Hz), 28.65, 25.65, 21.83 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.85 (dt, $J = 56.8, 17.7$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{12}\text{H}_{18}\text{F}_2\text{NO}_2$: 246.1300; Found 242.1296



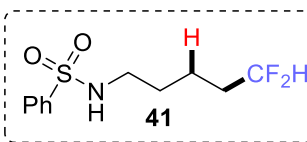
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.79 – 7.73 (m, 2H), 7.53 – 7.48 (m, 1H), 7.48-7.40 (m, 2H), 6.23 (s, 1H), 5.88 (tt, J = 56.6, 4.2 Hz, 1H), 3.53 (q, J = 6.7 Hz, 2H), 2.00-1.86 (m, 2H), 1.86-1.76 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 167.68, 134.42, 131.58, 128.64, 126.82, 116.87 (t, J = 239.1 Hz), 39.21, 31.47 (t, J = 21.3 Hz), 22.46 (t, J = 5.2 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.99 (dt, J = 56.9, 17.6 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{12}\text{H}_{14}\text{F}_2\text{NO}$: 214.1038; Found 214.1034



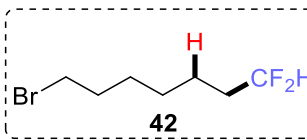
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.16 (dd, J = 5.1, 1.2 Hz, 1H), 6.94 (dd, J = 5.1, 3.4 Hz, 1H), 6.86 (dt, J = 3.3, 1.1 Hz, 1H), 5.79 (tt, J = 56.8, 4.4 Hz, 1H), 4.31 (t, J = 6.7 Hz, 2H), 3.16 (td, J = 6.7, 0.9 Hz, 2H), 2.35 (t, J = 7.4 Hz, 2H), 1.90 – 1.76 (m, 2H), 1.74-1.64 (m, 2H), 1.52 – 1.43 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 173.07, 139.94, 126.87, 125.52, 124.03, 117.09 (t, J = 238.9 Hz), 64.59, 33.94, 33.76 (t, J = 20.8 Hz), 29.32, 21.62 (t, J = 5.5 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.91 (dt, J = 57.1, 17.6 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{12}\text{H}_{17}\text{F}_2\text{O}_2\text{S}$: 263.0912; Found 263.0909



Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.83 – 7.75 (m, 2H), 7.62 – 7.57 (m, 1H), 7.56 – 7.50 (m, 2H), 5.81 (tt, J = 56.8, 4.4 Hz, 1H), 3.02 (t, J = 6.9 Hz, 2H), 2.73 (s, 3H), 1.93 – 1.78 (m, 2H), 1.64 – 1.57 (m, 2H), 1.57 – 1.48 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 137.47, 132.60, 129.09, 127.33, 117.13 (t, J = 239.0 Hz), 49.65, 34.68, 33.45 (t, J = 20.9 Hz), 27.00, 19.06 (t, J = 5.6 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.85 (dt, J = 57.0, 17.9 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{12}\text{H}_{18}\text{F}_2\text{NO}_2\text{S}$: 278.1021; Found 278.1016

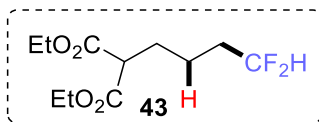


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.92 – 7.82 (m, 2H), 7.62 – 7.57 (m, 1H), 7.56-7.51 (m, 2H), 5.75 (tt, J = 56.7, 4.4 Hz, 1H), 4.70 (t, J = 6.3 Hz, 1H), 2.97 (q, J = 6.7 Hz, 2H), 1.83 – 1.72 (m, 2H), 1.57-1.49 (m, 2H), 1.50 – 1.41 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 139.86, 132.74, 129.19, 127.02, 116.93 (t, J = 239.0 Hz), 42.85, 33.41 (t, J = 21.0 Hz), 29.09, 19.08 (t, J = 5.6 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -116.05 (dt, J = 57.0, 17.9 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{16}\text{F}_2\text{NO}_2\text{S}$: 264.0864; Found 264.0860

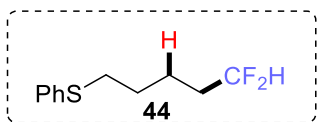


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.80 (tt, J = 56.9, 4.5 Hz, 1H), 3.41 (t, J = 6.8 Hz, 2H), 1.90 – 1.78 (m, 4H), 1.51 – 1.44 (m, 4H), 1.42-1.35 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 117.31 (t, J = 238.7 Hz), 33.95 (t, J = 20.7 Hz), 33.71,

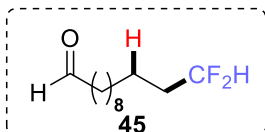
32.51, 28.21, 27.89, 21.92 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.85 (dt, $J = 56.7, 17.8$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁵



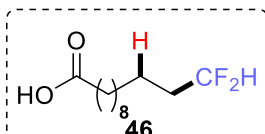
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 5.81 (tt, $J = 56.7, 4.4$ Hz, 1H), 4.25-4.16 (m, 4H), 3.33 (t, $J = 7.4$ Hz, 1H), 1.95 (q, $J = 7.7$ Hz, 2H), 1.92-1.81 (m, 2H), 1.54 – 1.47 (m, 2H), 1.27 (t, $J = 7.1$ Hz, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 169.16, 116.85 (t, $J = 239.1$ Hz), 61.50, 51.77, 33.72 (t, $J = 21.1$ Hz), 28.10, 19.98 (t, $J = 5.6$ Hz), 14.07. ^{19}F NMR (565 MHz, Chloroform- d) δ -116.12 (dt, $J = 56.8, 17.9$ Hz). GC-MS: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{19}\text{F}_2\text{O}_4$: 253.1246; Found 253



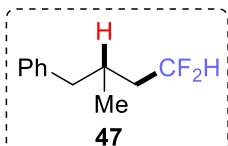
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.36 – 7.31 (m, 2H), 7.31-7.27 (m, 2H), 7.20 – 7.16 (m, 1H), 5.79 (tt, $J = 56.8, 4.4$ Hz, 1H), 2.93 (t, $J = 7.2$ Hz, 2H), 1.89 – 1.78 (m, 2H), 1.73-1.66 (m, 2H), 1.64 – 1.57 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 136.35, 129.29, 128.93, 126.03, 117.09 (t, $J = 239.0$ Hz), 33.61 (t, $J = 20.9$ Hz), 33.41, 28.55, 21.25 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.89 (dt, $J = 57.2, 17.8$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{15}\text{F}_2\text{S}$: 217.0857; Found 217.0854



Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 9.77 (s, 1H), 5.79 (tt, $J = 57.0, 4.6$ Hz, 1H), 2.42 (t, $J = 7.4$ Hz, 2H), 1.87 – 1.76 (m, 2H), 1.66-1.60 (m, 2H), 1.47-1.39 (m, 2H), 1.36 – 1.27 (m, 12H). ^{13}C NMR (151 MHz, CDCl_3) δ 202.93, 117.50 (t, $J = 238.7$ Hz), 43.92, 34.08 (t, $J = 20.6$ Hz), 29.33, 29.32, 29.14, 29.03, 22.10 (t, $J = 5.4$ Hz), 22.07. ^{19}F NMR (565 MHz, Chloroform- d) δ -115.75 (dt, $J = 57.0, 17.6$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁶

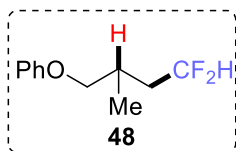


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 5.79 (tt, $J = 57.0, 4.6$ Hz, 1H), 2.35 (t, $J = 7.5$ Hz, 2H), 1.89 – 1.74 (m, 2H), 1.67-1.59 (m, 2H), 1.47-1.40 (m, 2H), 1.35 – 1.27 (m, 12H). ^{13}C NMR (151 MHz, CDCl_3) δ 180.04, 117.50 (t, $J = 238.7$ Hz), 34.08 (t, $J = 20.7$ Hz), 34.03, 29.32, 29.18, 29.03, 24.65, 22.10 (t, $J = 5.5$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -115.73 (dt, $J = 57.0, 17.5$ Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁶

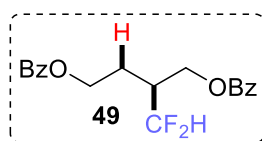


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.32-7.27 (m, 2H), 7.23 – 7.19 (m, 1H), 7.17 – 7.12 (m, 2H), 5.95 – 5.72 (m, 1H), 2.64 (dd, $J = 13.5, 6.8$ Hz, 1H), 2.51 (dd, $J = 13.5, 7.7$ Hz, 1H), 2.10-1.98 (m, 1H), 1.95-1.82 (m, 1H), 1.74-1.60 (m, 1H), 0.98 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 139.95, 129.15, 128.35, 126.16, 116.99 (t, $J = 238.7$ Hz), 43.52, 40.37 (t,

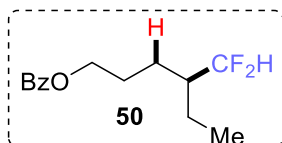
$J = 20.1$ Hz), 29.91 (t, $J = 5.2$ Hz), 19.61. ^{19}F NMR (565 MHz, Chloroform- d) δ -114.61 (dt, $J = 56.9, 17.5$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{11}\text{H}_{14}\text{F}_2$: 184.1064; Found 184.1



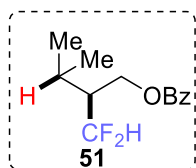
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.32 – 7.26 (m, 2H), 6.98–6.92 (m, 1H), 6.91 – 6.86 (m, 2H), 5.99 (tt, $J = 56.8, 4.8$ Hz, 1H), 3.86 (dd, $J = 9.1, 5.4$ Hz, 1H), 3.79 (dd, $J = 9.1, 6.6$ Hz, 1H), 2.30 – 2.21 (m, $J = 6.7$ Hz, 1H), 2.17–2.06 (m, 1H), 1.89–1.77 (m, 1H), 1.12 (d, $J = 6.9$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 158.79, 129.50, 120.88, 116.89 (t, $J = 238.7$ Hz), 114.47, 72.27, 38.14 (t, $J = 20.7$ Hz), 28.42 (t, $J = 5.3$ Hz), 17.16. ^{19}F NMR (565 MHz, Chloroform- d) δ -113.17 – -116.09 (m). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{11}\text{H}_{14}\text{F}_2\text{O}$: 200.1013; Found 200.0



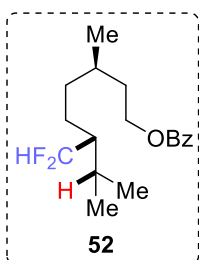
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.15–7.92 (m, 4H), 7.61–7.49 (m, 2H), 7.46–7.36 (m, 4H), 6.16 – 5.92 (m, 1H), 4.56–4.43 (m, 4H), 2.61 – 2.48 (m, 1H), 2.25–2.16 (m, 1H), 2.06–1.96 (m, 1H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.37, 166.12, 133.33, 133.13, 129.89, 129.62, 129.58, 129.55, 128.51, 128.45, 116.68 (t, $J = 242.9$ Hz), 62.29, 62.12 (t, $J = 5.7$ Hz), 39.88 (t, $J = 20.0$ Hz), 24.36 (t, $J = 4.0$ Hz). ^{19}F NMR (565 MHz, Chloroform- d) δ -121.50 – -125.84 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{19}\text{H}_{19}\text{F}_2\text{O}_4$: 349.1246; Found 349.1239



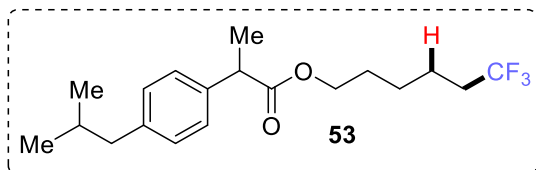
Prepared according to General Procedure B and obtained as colorless oil. 1.2:1 mixture of regioisomers. ^1H NMR (600 MHz, Chloroform- d) δ 8.07–7.98 (m, 2H), 7.61–7.55 (m, 1H), 7.50–7.41 (m, 2H), 5.94–5.63 (m, 1H), 5.78 (m, 0.88H), 4.43–4.38 (m, 1.12H), 4.35–4.31 (t, $J = 6.5$ Hz, 1H), 2.06–1.97 (m, 1H), 1.91 – 1.73 (m, 2.26H), 1.72 – 1.63 (m, 0.74H), 1.62 – 1.41 (m, 4H), 0.96 (dt, $J = 24.7, 7.4$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.62, 166.50, 133.02, 132.94, 130.31, 130.16, 129.54, 128.42, 128.39, 118.80 (t, $J = 242.5$ Hz), 118.53 (t, $J = 242.6$ Hz), 64.86, 62.78, 43.12 (t, $J = 18.5$ Hz), 39.25 (t, $J = 18.9$ Hz), 30.19 (dd, $J = 5.7, 3.5$ Hz), 26.74 (t, $J = 4.6$ Hz), 26.17, 23.62 (t, $J = 4.5$ Hz), 20.76 (dd, $J = 6.0, 4.4$ Hz), 19.96, 14.17, 11.15. ^{19}F NMR (565 MHz, Chloroform- d) δ -112.85 – -131.83 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{19}\text{F}_2\text{O}_2$: 257.1348; Found 257.1344



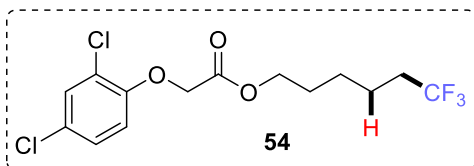
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.06 – 7.99 (m, 2H), 7.59 – 7.55 (m, 1H), 7.47–7.42 (m, 2H), 6.01 (td, $J = 55.9, 3.5$ Hz, 1H), 4.55 – 4.44 (m, 2H), 2.18–2.07 (m, 2H), 1.11 (d, $J = 6.7$ Hz, 3H), 1.07 (d, $J = 6.5$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.34, 133.14, 129.93, 129.61, 128.46, 117.24 (t, $J = 243.0$ Hz), 60.72 (t, $J = 5.7$ Hz), 47.46 (t, $J = 18.1$ Hz), 25.53 (t, $J = 3.7$ Hz), 20.62, 19.77. ^{19}F NMR (565 MHz, Chloroform- d) δ -117.23 – -125.25 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{17}\text{F}_2\text{O}_2$: 243.1191; Found 243.1188



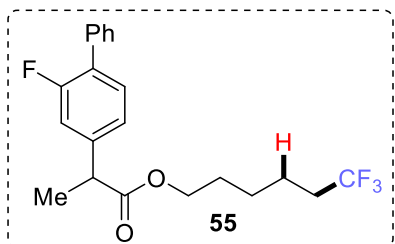
Prepared according to General Procedure B and obtained as colorless oil. 1:1 mixture of diastereomers. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.04 (d, J = 7.7 Hz, 2H), 7.55 (t, J = 7.4 Hz, 1H), 7.44 (t, J = 7.7 Hz, 2H), 5.78 (td, J = 56.5, 3.6 Hz, 1H), 4.43 – 4.30 (m, 2H), 1.97-1.87 (m, 1H), 1.87-1.79 (m, 1H), 1.65 – 1.53 (m, 4H), 1.53 – 1.40 (m, 2H), 1.38 – 1.30 (m, 1H), 1.01-0.97 (m, 3H), 0.97-0.91 (m, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.68, 132.85, 130.46, 129.54, 128.34, 119.00 (t, J = 242.4 Hz), 118.93 (t, J = 242.3 Hz), 63.40, 63.38, 47.83 (t, J = 17.4 Hz), 35.51, 35.45, 35.38, 30.40, 30.36, 27.11 (dd, J = 5.7, 3.6 Hz), 27.01 (dd, J = 5.8, 3.5 Hz), 21.94 (t, J = 4.4 Hz), 21.89 (t, J = 4.5 Hz), 19.75, 19.71, 19.58, 19.41, 19.39. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -116.39 – -124.91 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{18}\text{H}_{27}\text{F}_2\text{O}_2$: 313.1974; Found 313.1969



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.19 (d, J = 8.1 Hz, 2H), 7.09 (d, J = 8.1 Hz, 2H), 4.14 – 3.98 (m, 2H), 3.68 (q, J = 7.2 Hz, 1H), 2.44 (d, J = 7.2 Hz, 2H), 2.04 – 1.92 (m, 2H), 1.89-1.79 (m, 1H), 1.60-1.56 (m, 2H), 1.53 – 1.47 (m, 5H), 1.32-1.25 (m, 2H), 0.89 (d, J = 6.6 Hz, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 174.76, 140.55, 137.83, 129.30, 127.14, 127.09 (q, J = 276.3 Hz), 64.13, 45.19, 45.01, 33.58 (q, J = 28.4 Hz), 30.19, 28.16, 24.99, 22.36, 21.48 (q, J = 3.0 Hz), 18.35. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.42 (t, J = 10.9 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{19}\text{H}_{28}\text{F}_3\text{O}_2$: 345.2036; Found 345.2031

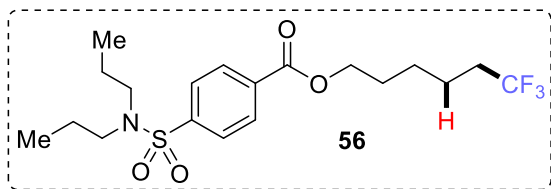


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.32 (d, J = 2.5 Hz, 1H), 7.09 (dd, J = 8.8, 2.6 Hz, 1H), 6.71 (d, J = 8.8 Hz, 1H), 4.62 (s, 2H), 4.13 (t, J = 6.5 Hz, 2H), 2.04 – 1.94 (m, 2H), 1.64 – 1.56 (m, 2H), 1.52-1.46 (m, 2H), 1.36-1.26 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 168.14, 152.41, 130.38, 127.56, 127.14, 127.06 (q, J = 276.4 Hz), 124.29, 114.67, 66.38, 65.11, 33.56 (q, J = 28.5 Hz), 28.14, 24.99, 21.54 (q, J = 2.9 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.36 (t, J = 10.9 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{16}\text{Cl}_2\text{F}_3\text{O}_3$: 359.0423; Found 359.0427

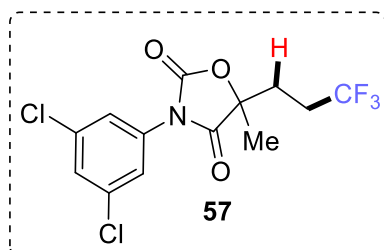


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.57-7.50 (m, 2H), 7.47 – 7.41 (m, 2H), 7.41 – 7.33 (m, 2H), 7.17 – 7.09 (m, 2H), 4.15-4.07 (m, 2H), 3.75 (q, J = 7.1 Hz, 1H), 2.08 – 1.96 (m, 2H), 1.67-1.59 (m, 2H), 1.57 – 1.52 (m, 5H), 1.40-1.32 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 173.98, 159.68 (d, J = 248.4 Hz), 141.86 (d, J = 7.7 Hz), 135.46, 130.78 (d, J = 3.9 Hz), 128.93 (d, J = 3.1 Hz), 128.47, 127.83 (q, J = 13.6 Hz), 127.08 (q, J = 276.2 Hz), 123.52 (d, J = 3.3 Hz),

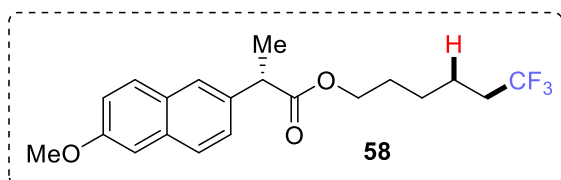
115.23 (d, $J = 23.5$ Hz), 64.54, 45.08, 33.59 (q, $J = 28.4$ Hz), 28.17, 25.06, 21.53 (q, $J = 3.0$ Hz), 18.28. ^{19}F NMR (565 MHz, Chloroform- d) δ -66.36 (t, $J = 11.0$ Hz), -117.67 (dd, $J = 13.0$, 8.8 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{21}\text{H}_{23}\text{F}_4\text{O}_2$: 383.1629; Found 383.1623



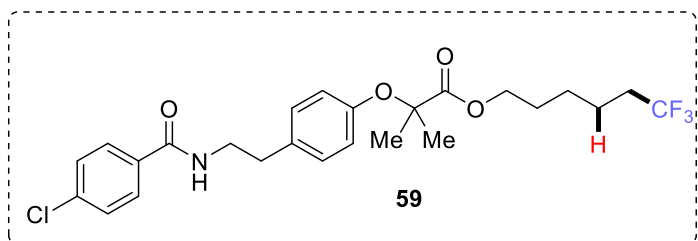
Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 8.15 (d, $J = 8.4$ Hz, 2H), 7.88 (d, $J = 8.4$ Hz, 2H), 4.37 (t, $J = 6.5$ Hz, 2H), 3.16 – 3.06 (m, 4H), 2.17 – 2.06 (m, 2H), 1.86 – 1.78 (m, 2H), 1.68–1.62 (m, 2H), 1.60–1.50 (m, 6H), 0.87 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 165.28, 144.32, 133.59, 130.19, 127.11 (q, $J = 276.3$ Hz), 127.04, 65.24, 49.95, 33.62 (q, $J = 28.5$ Hz), 28.34, 25.26, 21.95, 21.67 (q, $J = 3.0$ Hz), 11.16. ^{19}F NMR (565 MHz, Chloroform- d) δ -66.35 (t, $J = 11.0$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{19}\text{H}_{29}\text{F}_3\text{NO}_4\text{S}$: 424.1764; Found 424.1754



Prepared according to General Procedure A and obtained as white solid. ^1H NMR (600 MHz, Chloroform- d) δ 7.42–7.32 (m, 3H), 2.30 – 2.24 (m, 1H), 2.19 – 2.08 (m, 3H), 1.64 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.71, 151.80, 135.70, 132.29, 129.26, 126.06 (q, $J = 276.2$ Hz), 123.65, 84.08, 29.37 (q, $J = 3.2$ Hz), 28.23 (q, $J = 30.4$ Hz), 22.25. ^{19}F NMR (565 MHz, Chloroform- d) δ -66.40 (t, $J = 9.8$ Hz). m.p.: 114–115 $^\circ\text{C}$. HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{11}\text{Cl}_2\text{F}_3\text{NO}_2$: 356.0063; Found 356.0067

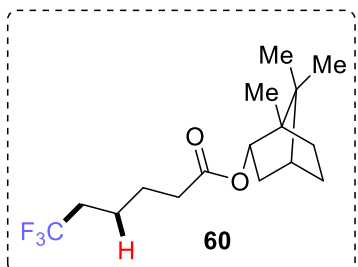


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.70 (d, $J = 8.7$ Hz, 2H), 7.67 – 7.64 (m, 1H), 7.39 (dd, $J = 8.5$, 1.8 Hz, 1H), 7.14 (dd, $J = 8.9$, 2.6 Hz, 1H), 7.11 (d, $J = 2.5$ Hz, 1H), 4.12 – 4.02 (m, 2H), 3.91 (s, 3H), 3.84 (q, $J = 7.1$ Hz, 1H), 1.89 – 1.82 (m, 2H), 1.58–1.56 (m, 4H), 1.48–1.41 (m, 2H), 1.26 – 1.20 (m, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 174.68, 157.68, 135.76, 133.69, 129.22, 128.92, 127.11, 127.05 (q, $J = 276.5$ Hz) 126.20, 125.94, 119.04, 105.57, 64.25, 55.31, 45.51, 33.48 (q, $J = 28.5$ Hz), 28.19, 24.99, 21.48 (q, $J = 3.0$ Hz), 18.30. ^{19}F NMR (565 MHz, Chloroform- d) δ -66.44 (t, $J = 10.9$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{20}\text{H}_{24}\text{F}_3\text{O}_3$: 369.1672; Found 369.1665

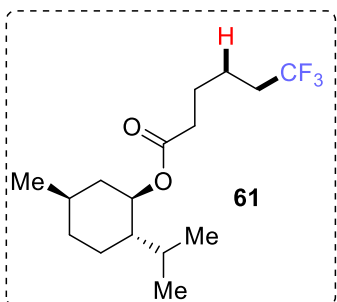


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform- d) δ 7.65 – 7.59 (m, 2H), 7.43 – 7.35 (m, 2H), 7.14 – 7.03

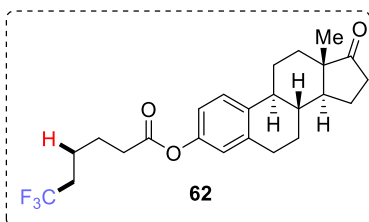
(m, 2H), 6.89 – 6.75 (m, 2H), 6.13-6.02 (m, 1H), 5.30 (s, 6H), 4.16 (t, $J = 6.5$ Hz, 2H), 3.66 (q, $J = 6.6$ Hz, 2H), 2.85 (t, $J = 6.9$ Hz, 2H), 2.04 – 1.94 (m, 2H), 1.66 – 1.63 (m, 2H), 1.56 – 1.48 (m, 2H), 1.33 (p, $J = 7.9$ Hz, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 174.30, 166.37, 154.20, 137.66, 132.99, 132.29, 129.48, 128.83, 128.24, 127.05 (q, $J = 276.3$ Hz), 119.26, 79.12, 64.97, 53.43, 41.23, 34.72, 33.55 (q, $J = 28.5$ Hz), 28.09, 25.38, 25.01, 21.50 (q, $J = 3.0$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.33 (t, $J = 10.9$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{25}\text{H}_{30}\text{NF}_3\text{ClO}_4$: 500.1810; Found 500.1816



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 4.93-4.87 (m, 1H), 2.44 – 2.30 (m, 3H), 2.19 – 2.01 (m, 2H), 1.94-1.87 (m, 1H), 1.78 – 1.66 (m, 4H), 1.66 – 1.59 (m, 2H), 1.35-1.28 (m, 1H), 1.27 – 1.19 (m, 2H), 0.91 (s, 3H), 0.88 (s, 3H), 0.83 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 173.36, 173.33, 127.05 (q, $J = 276.8$ Hz), 80.01, 79.98, 48.77, 48.74, 47.83, 47.80, 44.91, 44.88, 36.86, 36.84, 34.17, 34.14, 33.52 (q, $J = 28.7$ Hz), 28.05, 28.03, 27.13, 27.11, 24.16, 24.14, 21.47, 19.72, 19.69, 18.85, 18.83, 13.50, 13.48. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.39 (t, $J = 10.9$ Hz). HRMS APCI: $[\text{M}+\text{NH}_4]^+$ calcd. for $\text{C}_{16}\text{H}_{29}\text{F}_3\text{NO}_2$: 324.2145; Found 234.2143

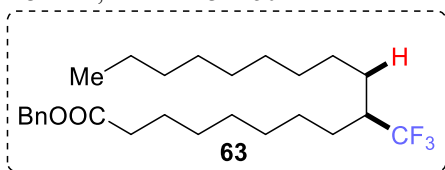


Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 4.69 (td, $J = 10.9, 4.4$ Hz, 1H), 2.36 – 2.29 (m, 2H), 2.14 – 2.05 (m, 2H), 2.01-1.95 (m, 1H), 1.88-1.81 (m, 1H), 1.73-1.65 (m, 4H), 1.63 – 1.58 (m, 2H), 1.52-1.44 (m, 1H), 1.40-1.34 (m, 1H), 1.26 – 1.18 (m, 1H), 1.10-1.01 (m, 1H), 0.93-0.87 (m, 7H), 0.76 (d, $J = 7.0$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.62, 127.03 (q, $J = 276.2$ Hz), 74.28, 47.02, 40.94, 34.25, 34.18, 33.50 (q, $J = 28.4$ Hz), 31.39, 26.32, 24.13, 23.41, 22.01, 21.45 (q, $J = 3.2$ Hz), 20.74, 16.25. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.43 (t, $J = 10.8$ Hz). HRMS APCI: $[\text{M}+\text{NH}_4]^+$ calcd. for $\text{C}_{16}\text{H}_{31}\text{F}_3\text{NO}_2$: 326.2301; Found 326.2292



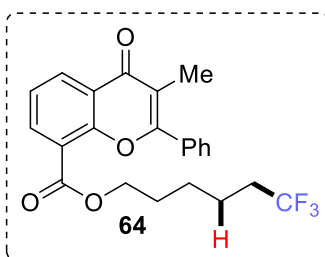
Prepared according to General Procedure A and obtained as white solid. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.24-7.20 (m, 1H), 6.79-6.75 (m, 1H), 6.75-6.71 (m, 1H), 2.88 – 2.80 (m, 2H), 2.51 (t, $J = 7.3$ Hz, 2H), 2.47-2.40 (m, 1H), 2.37-2.31 (m, 1H), 2.26-2.18 (m, 1H), 2.13-2.03 (m, 3H), 2.02 – 1.91 (m, 2H), 1.91-1.87 (m, 1H), 1.76 (p, $J = 7.5$ Hz, 2H), 1.66 – 1.60 (m, 2H), 1.59 – 1.36 (m, 6H), 0.84 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.83, 148.47, 138.08, 137.47, 127.0 (q, $J = 276.3$ Hz), 126.45, 121.51, 118.68, 50.44, 47.96, 44.16, 38.00, 35.87, 33.85, 33.50 (q, $J = 28.6$ Hz), 31.56, 29.41, 26.34, 25.76, 23.97, 21.60, 21.47 (q, $J = 3.1$ Hz), 13.84. ^{19}F NMR

(565 MHz, Chloroform-*d*) δ -66.32 (t, J = 10.8 Hz). m.p.: 94-95 °C. HRMS APCI: $[M+H]^+$ calcd. for C₂₄H₃₀F₃O₃: 423.2142; Found 423.2150



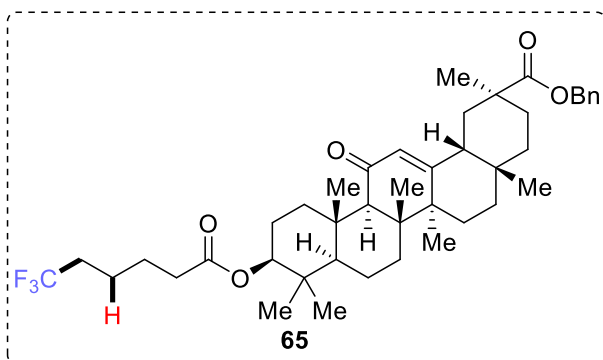
Prepared according to General Procedure A and obtained as colorless oil.

1:1 mixture of regioisomers. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.43 – 7.30 (m, 5H), 5.11 (s, 2H), 2.35 (t, J = 7.5 Hz, 2H), 2.01 – 1.93 (m, 1H), 1.67-1.61 (m, 2H), 1.57-1.52 (m, 2H), 1.42 – 1.26 (m, 24H), 0.88 (t, J = 6.9 Hz, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 173.68, 173.64, 136.14, 136.13, 128.79 (q, J = 280.6 Hz), 128.55, 128.19, 66.10, 66.08, 42.56 (q, J = 24.6 Hz), 34.32, 34.29, 31.89, 31.85, 29.72, 29.65, 29.55, 29.53, 29.42, 29.38, 29.30, 29.25, 29.21, 29.18, 29.08, 29.05, 29.03, 27.85, 27.84, 27.82, 26.88, 26.86, 26.82, 24.93, 24.90, 22.68, 22.66, 14.11. ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -70.05– -70.18 (m, 3F). HRMS APCI: $[M+H]^+$ calcd. for C₂₆H₄₂F₃O₂: 443.3131; Found 443.3131



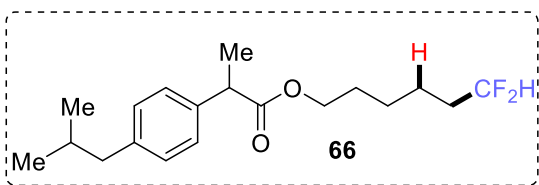
Prepared according to General Procedure A and obtained as colorless oil. ¹H NMR

(600 MHz, Chloroform-*d*) δ 8.47 (dd, J = 7.9, 1.8 Hz, 1H), 8.26 (dd, J = 7.5, 1.8 Hz, 1H), 7.77 (dt, J = 6.8, 2.2 Hz, 2H), 7.54 (dd, J = 5.1, 1.9 Hz, 3H), 7.45 (t, J = 7.7 Hz, 1H), 4.36 (t, J = 6.5 Hz, 2H), 2.23 (s, 3H), 2.02 – 1.92 (m, 2H), 1.76 – 1.70 (m, 2H), 1.55 – 1.48 (m, 2H), 1.44-1.35 (m, 2H). ¹³C NMR (151 MHz, CDCl₃) δ 178.27, 164.63, 161.08, 154.45, 136.16, 133.11, 130.85, 130.51, 129.31, 128.44, 127.05 (q, J = 276.3 Hz), 124.05, 123.34, 120.72, 117.76, 65.22, 33.56 (q, J = 28.4 Hz), 28.31, 25.22, 21.57 (q, J = 3.0 Hz), 11.75. ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -66.38 (t, J = 11.0 Hz). HRMS APCI: $[M+H]^+$ calcd. for C₂₃H₂₂F₃O₄: 419.1465; Found 419.1457

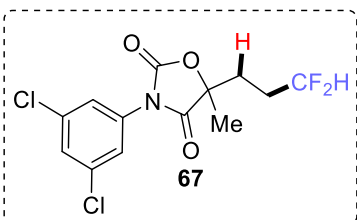


Prepared according to General Procedure A and obtained

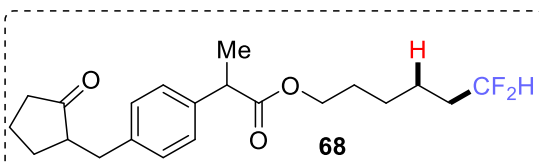
as white solid. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.50 – 7.29 (m, 5H), 5.54 (s, 1H), 5.20 (d, J = 12.2 Hz, 1H), 5.09 (d, J = 12.2 Hz, 1H), 4.53 (dd, J = 11.8, 4.7 Hz, 1H), 2.83-2.76 (m, 1H), 2.38 – 2.30 (m, 3H), 2.15-2.07 (m, 2H), 2.06 – 1.97 (m, 3H), 1.97-1.91 (m, 1H), 1.84-1.76 (m, 1H), 1.74 – 1.59 (m, 9H), 1.49 – 1.26 (m, 9H), 1.16 (s, 6H), 1.11 (s, 3H), 1.09 – 0.97 (m, 2H), 0.91-0.85 (m, 6H), 0.82-0.76 (m, 1H), 0.73 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ 199.99, 176.22, 172.81, 169.11, 136.14, 128.63, 128.47, 128.31, 128.26, 127.03 (q, J = 276.3 Hz), 80.72, 66.23, 61.68, 55.01, 48.24, 45.37, 44.00, 43.18, 41.06, 38.78, 38.08, 37.65, 36.93, 34.25, 33.49 (q, J = 28.4 Hz), 32.70, 31.79, 31.18, 28.42, 28.29, 28.08, 26.46, 26.40, 24.14, 23.59, 23.31, 21.50 (q, J = 3.1 Hz), 18.68, 17.38, 16.75, 16.41. ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -66.36 (t, J = 11.0 Hz). m.p.: 184-185 °C HRMS APCI: $[M+H]^+$ calcd. for C₄₃H₆₃F₃O₅: 713.4387; Found 713.4375



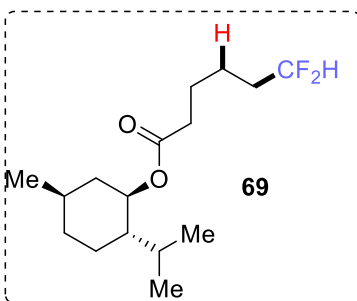
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.22 – 7.17 (m, 2H), 7.11 – 7.06 (m, 2H), 5.74 (tt, J = 56.9, 4.5 Hz, 1H), 4.13 – 4.00 (m, 2H), 3.68 (q, J = 7.1 Hz, 1H), 2.44 (d, J = 7.1 Hz, 2H), 1.90-1.80 (m, 1H), 1.80 – 1.69 (m, 2H), 1.60 – 1.57 (m, 2H), 1.48 (d, J = 7.2 Hz, 3H), 1.44 – 1.35 (m, 2H), 1.30-1.25 (m, 2H), 0.89 (d, J = 6.6 Hz, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 174.79, 140.54, 137.85, 129.30, 127.14, 117.21 (t, J = 238.7 Hz), 64.27, 45.19, 45.01, 33.91 (t, J = 20.7 Hz), 30.19, 28.31, 25.32, 22.37, 21.68 (t, J = 5.5 Hz), 18.39. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.86 (dt, J = 57.0, 17.9 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{19}\text{H}_{29}\text{F}_2\text{O}_2$: 327.2130; Found 327.2125



Prepared according to General Procedure B and obtained as white solid. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.40-7.33 (m, 3H), 5.83 (tt, J = 56.1, 3.9 Hz, 1H), 2.13 – 2.05 (m, 2H), 2.06 – 1.95 (m, 1H), 1.90 – 1.79 (m, 1H), 1.63 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 173.07, 152.03, 135.65, 132.41, 129.18, 123.71, 115.58 (t, J = 240.2 Hz), 84.80, 29.08 (t, J = 5.7 Hz), 28.02 (t, J = 22.2 Hz), 22.40. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -116.87 (dt, J = 56.1, 17.0 Hz). m.p.: 121-122 °C. HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{12}\text{Cl}_2\text{F}_2\text{NO}_3$: 338.0157; Found 338.0161

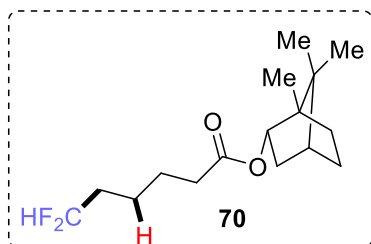


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.21 (d, J = 7.9 Hz, 2H), 7.12 (d, J = 7.9 Hz, 2H), 5.75 (tt, J = 56.9, 4.4 Hz, 1H), 4.06 (tt, J = 8.1, 4.1 Hz, 2H), 3.68 (q, J = 7.2 Hz, 1H), 3.12 (dd, J = 14.0, 4.2 Hz, 1H), 2.51 (dd, J = 13.9, 9.5 Hz, 1H), 2.38 – 2.29 (m, 2H), 2.14 – 2.05 (m, 2H), 1.99-1.92 (m, 1H), 1.81 – 1.69 (m, 3H), 1.61 – 1.53 (m, 3H), 1.48 (d, J = 7.2 Hz, 3H), 1.44-1.37 (m, 2H), 1.34 – 1.27 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 220.18, 174.65, 138.87, 138.45, 129.11, 129.09, 127.52, 117.18 (t, J = 238.8 Hz), 64.35, 50.97, 45.17, 38.18, 35.19, 33.90 (t, J = 20.8 Hz), 29.24, 28.29, 25.33, 21.66 (t, J = 5.5 Hz), 20.53, 18.41. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.86 (dt, J = 56.7, 17.7 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{21}\text{H}_{29}\text{F}_2\text{O}_3$: 367.2079; Found 367.2075

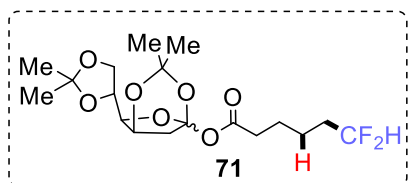


Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.80 (tt, J = 56.8, 4.5 Hz, 1H), 4.69 (td, J = 10.9, 4.4 Hz, 1H), 2.34 – 2.29 (m, 2H), 2.00-1.94 (m, 1H), 1.90 – 1.79 (m, 3H), 1.72 – 1.64 (m, 4H), 1.53-1.45 (m, 3H), 1.39 – 1.34 (m, 1H), 1.10-1.01 (m, 1H), 1.00-0.93 (m, 1H), 0.92 – 0.85 (m, 7H), 0.76 (d, J = 6.9 Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.83, 117.10 (t, J

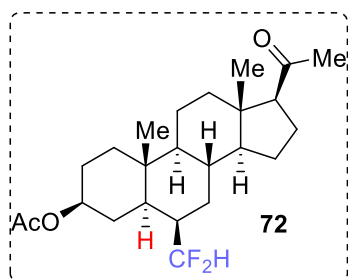
= 238.9 Hz), 74.18, 47.02, 40.95, 34.36, 34.26, 33.78 (t, $J = 20.8$ Hz), 31.39, 26.31, 24.50, 23.42, 22.02, 21.63 (t, $J = 5.6$ Hz), 20.75, 16.27. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.96 (dt, $J = 56.6, 17.5$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{29}\text{F}_2\text{O}_2$: 291.2130; Found 291.2126



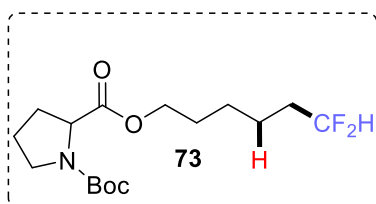
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.81 (tt, $J = 56.7, 4.5$ Hz, 1H), 4.89 (dq, $J = 10.0, 1.9$ Hz, 1H), 2.39-2.31 (m, 3H), 1.94 – 1.81 (m, 3H), 1.77 – 1.67 (m, 4H), 1.55-1.47 (m, 2H), 1.34 – 1.20 (m, 3H), 0.91 (s, 3H), 0.87 (s, 3H), 0.83 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 173.56, 117.10 (t, $J = 238.9$ Hz), 79.89, 48.74, 47.80, 44.88, 36.84, 34.33, 33.79 (t, $J = 21.0$ Hz), 28.04, 27.12, 24.51, 21.64 (t, $J = 5.5$ Hz), 19.70, 18.84, 13.50. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.93 (dt, $J = 57.1, 18.0$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{27}\text{F}_2\text{O}_2$: 289.1974; Found 289.1972



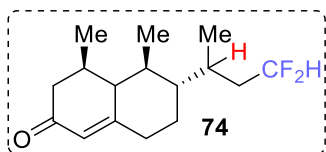
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 6.14 (s, 1H), 5.82 (tt, $J = 56.7, 4.4$ Hz, 1H), 4.86 (dd, $J = 5.9, 3.6$ Hz, 1H), 4.69 (d, $J = 5.8$ Hz, 1H), 4.44-4.37 (m, 1H), 4.12-4.07 (m, 1H), 4.07-3.97 (m, 2H), 2.40-2.27 (m, 2H), 1.91-1.79 (m, 2H), 1.69 (p, $J = 7.5$ Hz, 1H), 1.53 – 1.47 (m, 5H), 1.46 (s, 3H), 1.38 (s, 3H), 1.35 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.61, 117.00 (t, $J = 239.0$ Hz), 113.32, 109.38, 100.74, 85.07, 82.28, 79.32, 72.87, 66.82, 33.90, 33.71 (t, $J = 21.0$ Hz), 26.99, 25.95, 25.13, 24.67, 24.08, 21.50 (t, $J = 5.6$ Hz). ^{19}F NMR (564 MHz, Chloroform-*d*) δ -116.01 (dt, $J = 56.8, 17.5$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{18}\text{H}_{29}\text{F}_2\text{O}_7$: 395.1876; Found 395.1873



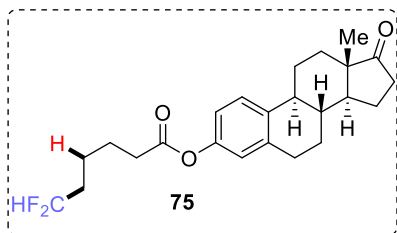
Prepared according to General Procedure B and obtained as colorless oil. 8.1:1 mixture of diastereomers. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.91 (td, $J = 56.6, 7.6$ Hz, 0.89H), 5.69 (td, $J = 56.4, 2.5$ Hz, 0.11H), 4.66 – 4.51 (m, 1H), 2.46 (t, $J = 9.0$ Hz, 1H), 2.15 – 2.03 (m, 4H), 2.00 – 1.93 (m, 4H), 1.93-1.86 (m, 1H), 1.86-1.80 (m, 1H), 1.79 – 1.68 (m, 4H), 1.67-1.52 (m, 4H), 1.49 – 1.25 (m, 5H), 1.17 – 0.96 (m, 4H), 0.87 (s, 2.70H), 0.79 (s, 0.33H), 0.703-0.65 (m, 1H), 0.56 (s, 2.67H), 0.55 (s, 0.33H). ^{13}C NMR (151 MHz, CDCl_3) δ 209.38, 170.69, 118.06 (dd, $J = 241.7, 239.4$ Hz), 73.86, 63.64, 56.23, 54.54, 45.06 (d, $J = 7.2$ Hz), 44.12, 43.17 (t, $J = 18.6$ Hz), 39.17, 38.77, 35.83, 32.84 (d, $J = 5.1$ Hz), 31.96, 31.78 (dd, $J = 8.1, 3.6$ Hz), 31.51, 27.56, 24.37, 22.80, 21.42, 21.03, 15.96, 13.52. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -114.20 (dddd, $J = 287.4, 57.2, 16.4$ Hz), -122.66 – -128.15 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{24}\text{H}_{37}\text{F}_2\text{O}_3$: 411.2705; Found 411.2703



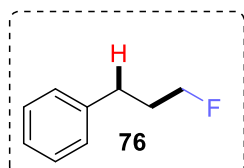
Prepared according to General Procedure B and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.73 (tt, $J = 57.0, 4.6$ Hz, 1H), 4.28 – 3.99 (m, 3H), 3.53 – 3.29 (m, 2H), 2.20-2.07 (m, 1H), 1.93 – 1.70 (m, 5H), 1.63-1.52 (m, 2H), 1.43 – 1.32 (m, 13H). ^{13}C NMR (151 MHz, CDCl_3) δ 173.30, 173.08, 154.40, 153.82, 117.25 (t, $J = 238.9$ Hz), 117.15 (t, $J = 239.0$ Hz), 79.84, 79.72, 64.61, 64.59, 59.18, 58.88, 46.56, 46.32, 33.91 (t, $J = 20.8$ Hz), 30.94, 29.98, 28.46, 28.43, 28.32, 25.49, 25.37, 24.32, 23.62, 21.76 (t, $J = 5.1$ Hz), 21.72 (d, $J = 5.2$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.93 (ddt, $J = 56.8, 26.4, 17.6$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{28}\text{F}_2\text{NO}_4$: 336.1981; Found 336.1981



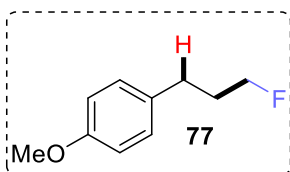
Prepared according to General Procedure B and obtained as colorless oil. 1:1 mixture of diastereomers. ^1H NMR (600 MHz, Chloroform-*d*) δ 5.99-5.77 (m, 1H), 5.77-5.73 (m, 1H), 2.51 – 2.40 (m, 1H), 2.39-2.33 (m, 1H), 2.33 – 2.20 (m, 2H), 2.07 – 1.55 (m, 8H), 1.23 – 1.13 (m, 1H), 1.11-1.06 (m, 3H), 0.99 – 0.92 (m, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 199.57, 170.51, 124.66, 124.63, 117.21 (t, $J = 239.0$ Hz), 117.17 (t, $J = 239.0$ Hz), 42.42, 42.04, 40.84, 40.56, 40.54, 39.27, 39.17, 38.56 (t, $J = 20.1$ Hz), 38.09 (t, $J = 20.1$ Hz), 37.62, 37.61, 32.99, 32.99, 32.22-32.18 (m), 32.16-32.13 (m), 29.77, 28.32, 16.93, 16.36, 16.00, 14.95, 14.94. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -111.43 – -119.70 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{25}\text{F}_2\text{O}$: 271.1868; Found 271.1870. The compound characterization was reported in literature and the NMR data matched with previous characterization.¹



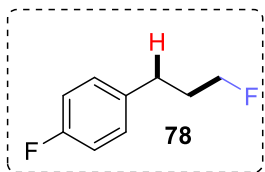
Prepared according to General Procedure B and obtained as white solid. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.31-7.27 (m, 1H), 6.88-6.83 (m, 1H), 6.81-6.78 (m, 1H), 5.83 (tt, $J = 56.8, 4.4$ Hz, 1H), 2.94-2.88 (m, 2H), 2.58 (t, $J = 7.4$ Hz, 2H), 2.54 – 2.48 (m, 1H), 2.44 – 2.36 (m, 1H), 2.34-2.25 (m, 1H), 2.20-2.10 (m, 1H), 2.10 – 1.98 (m, 2H), 1.98-1.94 (m, 1H), 1.94-1.84 (m, 2H), 1.94-1.79 (m, 2H), 1.69 – 1.41 (m, 9H), 0.91 (s, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.01, 148.50, 138.04, 137.41, 126.42, 121.52, 118.69, 118.69, 117.03 (t, $J = 239.0$ Hz), 50.42, 47.94, 44.14, 37.99, 35.85, 34.02, 33.75 (t, $J = 20.9$ Hz), 31.54, 29.40, 26.33, 25.75, 24.35, 21.63, 21.59 (t, $J = 5.5$ Hz), 21.58, 21.55, 13.82. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -115.98 (dt, $J = 56.7, 17.6$ Hz). m.p.: 95-96 °C. HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{24}\text{H}_{31}\text{F}_2\text{O}_3$: 405.2236; Found 405.2244



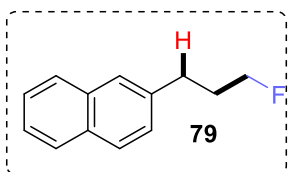
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.32 – 7.27 (m, 2H), 7.23-7.18 (m, 3H), 4.46 (dt, $J = 47.2, 6.0$ Hz, 2H), 2.78 – 2.70 (m, 2H), 2.06 – 1.97 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 141.11, 128.49, 128.47, 126.04, 83.15 (d, $J = 164.8$ Hz), 32.04 (d, $J = 19.9$ Hz), 31.33 (d, $J = 5.4$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.01 (tt, $J = 47.2, 25.3$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_9\text{H}_{11}\text{F}$: 138.0845; Found 138



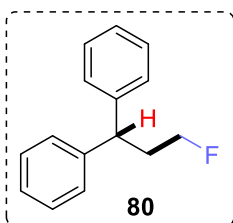
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.13 – 7.08 (m, 2H), 6.86 – 6.82 (m, 2H), 4.44 (dt, $J = 47.2, 5.9$ Hz, 2H), 3.79 (s, 3H), 2.72 – 2.66 (m, 2H), 2.04 – 1.90 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 157.93, 133.14, 129.39, 113.87, 83.14 (d, $J = 164.6$ Hz), 55.27, 32.26 (d, $J = 19.6$ Hz), 30.38 (d, $J = 5.4$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.09 (tt, $J = 47.4, 25.6$ Hz). HRMS APCI: [M-H] $^+$ calcd. for $\text{C}_{10}\text{H}_{12}\text{FO}$: 167.0867; Found 167.0865



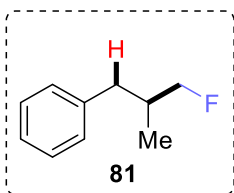
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.17 – 7.13 (m, 2H), 7.00 – 6.95 (m, 2H), 4.45 (dt, $J = 47.2, 5.9$ Hz, 2H), 2.76 – 2.70 (m, 2H), 2.05 – 1.90 (m, 2H). ^{13}C NMR (151 MHz, Chloroform-*d*) δ 161.39 (d, $J = 243.8$ Hz), 136.68 (d, $J = 3.3$ Hz), 129.83 (d, $J = 7.7$ Hz), 115.21 (d, $J = 21.0$ Hz), 82.93 (d, $J = 164.7$ Hz), 32.14 (d, $J = 19.7$ Hz), 30.53 (d, $J = 5.4$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -117.45 (tt, $J = 9.7, 5.7$ Hz), -220.32 (tt, $J = 46.5, 25.5$ Hz). GC-MS: [M] $^+$ calcd. for $\text{C}_9\text{H}_{10}\text{F}_2$: 156.0751; Found 156



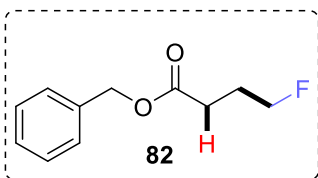
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.84 – 7.75 (m, 3H), 7.64 (d, $J = 1.6$ Hz, 1H), 7.44 (dddd, $J = 18.4, 8.1, 6.8, 1.4$ Hz, 2H), 7.34 (dd, $J = 8.4, 1.8$ Hz, 1H), 4.49 (dt, $J = 47.2, 5.9$ Hz, 2H), 2.96 – 2.88 (m, 2H), 2.17 – 2.03 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 138.57, 133.60, 132.07, 128.06, 127.62, 127.43, 127.20, 126.61, 126.01, 125.29, 83.10 (d, $J = 165.0$ Hz), 31.93 (d, $J = 19.7$ Hz), 31.46 (d, $J = 5.4$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -215.61 – -231.03 (m). GC-MS: [M] $^+$ calcd. for $\text{C}_{13}\text{H}_{13}\text{F}$: 188.1001; Found 188



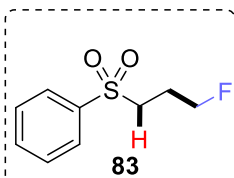
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.29 (dd, $J = 8.1, 7.1$ Hz, 4H), 7.25 (dd, $J = 4.5, 2.9$ Hz, 4H), 7.21 – 7.17 (m, 2H), 4.39 (dt, $J = 47.0, 6.0$ Hz, 2H), 4.17 (t, $J = 8.0$ Hz, 1H), 2.49-2.39 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 143.86, 128.60, 127.87, 126.44, 82.06 (d, $J = 164.6$ Hz), 46.43 (d, $J = 5.4$ Hz), 36.05 (d, $J = 19.8$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.88 (tt, $J = 47.5, 24.2$ Hz). GC-MS: [M] $^+$ calcd. for $\text{C}_{15}\text{H}_{15}\text{F}$: 214.1158; Found 214



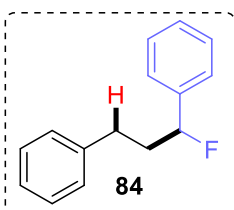
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.32 – 7.27 (m, 2H), 7.22 – 7.19 (m, 1H), 7.18 – 7.15 (m, 2H), 4.27 (dd, $J = 47.5, 5.5$ Hz, 2H), 2.77 (dd, $J = 13.6, 6.5$ Hz, 1H), 2.47 (dd, $J = 13.5, 8.0$ Hz, 1H), 2.18 – 1.96 (m, 1H), 0.95 (dd, $J = 6.8, 1.1$ Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 139.88, 129.19, 128.31, 126.05, 87.42 (d, $J = 168.8$ Hz), 38.68 (d, $J = 5.6$ Hz), 36.09 (d, $J = 18.2$ Hz), 15.65 (d, $J = 6.0$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -224.20 (td, $J = 47.3, 20.4$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{10}\text{H}_{13}\text{F}$: 152.1001; Found 152



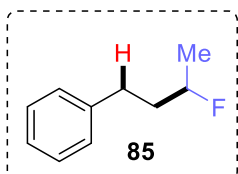
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.40 – 7.29 (m, 5H), 5.13 (s, 2H), 4.48 (dt, $J = 47.1, 5.8$ Hz, 2H), 2.52 (t, $J = 7.4$ Hz, 2H), 2.09-1.99 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.72, 135.87, 128.60, 128.30, 128.23, 82.89 (d, $J = 165.3$ Hz), 66.40, 29.96 (d, $J = 5.1$ Hz), 25.76 (d, $J = 20.4$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.97 (dp, $J = 73.7, 25.6$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{11}\text{H}_{14}\text{FO}_2$: 197.0973; Found 197.0972



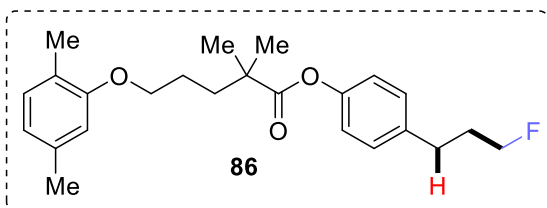
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.95 – 7.89 (m, 2H), 7.71 – 7.65 (m, 1H), 7.61 – 7.57 (m, 2H), 4.52 (dt, $J = 46.8, 5.7$ Hz, 2H), 3.27 – 3.21 (m, 2H), 2.20 – 2.09 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 138.91, 133.93, 129.43, 128.05, 81.55 (d, $J = 167.9$ Hz), 52.53 (d, $J = 4.3$ Hz), 24.09 (d, $J = 20.8$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.52 (tt, $J = 46.9, 25.9$ Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_9\text{H}_{12}\text{FO}_2\text{S}$: 203.0537; Found 203.0533



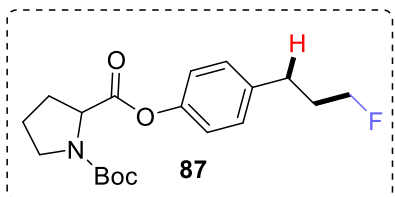
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.39 – 7.35 (m, 2H), 7.34 – 7.31 (m, 3H), 7.31 – 7.27 (m, 2H), 7.22 – 7.18 (m, 3H), 5.43 (ddd, $J = 47.8, 8.5, 4.4$ Hz, 1H), 2.87 – 2.68 (m, 2H), 2.38 – 2.24 (m, 1H), 2.18 – 2.05 (m, 1H). ^{13}C NMR (151 MHz, CDCl_3) δ 141.13, 140.16 (d, $J = 19.7$ Hz), 128.48, 128.46, 128.30 (d, $J = 2.0$ Hz), 126.05, 125.57 (d, $J = 6.8$ Hz), 93.65 (d, $J = 170.9$ Hz), 38.82 (d, $J = 23.9$ Hz), 31.32 (d, $J = 4.3$ Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -176.01 (ddd, $J = 46.5, 30.0, 15.9$ Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{15}\text{H}_{15}\text{F}$: 214.1158; Found 214



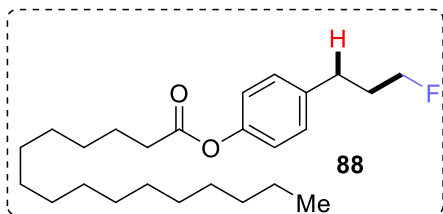
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.31 – 7.27 (m, 2H), 7.20 (dd, J = 7.5, 1.4 Hz, 3H), 4.74 – 4.57 (m, 1H), 2.84-2.76 (m, 1H), 2.72-2.64 (m, 1H), 2.04-1.92 (m, 1H), 1.90 – 1.74 (m, 1H), 1.34 (dd, J = 23.9, 6.2 Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 141.50, 128.44, 125.95, 90.05 (d, J = 165.0 Hz), 38.67 (d, J = 20.8 Hz), 31.37 (d, J = 4.9 Hz), 21.00 (d, J = 22.6 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -174.22 (ddqd, J = 47.9, 31.8, 23.7, 16.0 Hz). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{10}\text{H}_{13}\text{F}$: 152.1001; Found 152



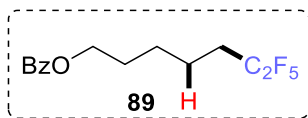
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.21 – 7.15 (m, 2H), 7.02-6.98 (m, 1H), 6.98 – 6.92 (m, 2H), 6.69 – 6.64 (m, 1H), 6.64-6.60 (m, 1H), 4.45 (dt, J = 47.2, 5.9 Hz, 2H), 4.01-3.94 (m, 2H), 2.79 – 2.68 (m, 2H), 2.30 (s, 3H), 2.17 (s, 3H), 2.05 – 1.94 (m, 2H), 1.36 (s, 6H). ^{13}C NMR (151 MHz, CDCl_3) δ 176.47, 156.88, 149.25, 138.45, 136.48, 130.34, 129.35, 123.62, 121.44, 120.74, 111.95, 82.97 (d, J = 165.0 Hz), 67.80, 42.40, 37.16, 32.04 (d, J = 19.8 Hz), 30.72 (d, J = 5.4 Hz), 25.28, 25.16, 21.40, 15.79. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.24 (tt, J = 47.4, 25.6 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{24}\text{H}_{32}\text{FO}_3$: 387.2330; Found 387.2325



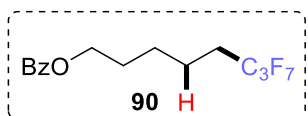
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.23-7.16 (m, 2H), 7.06-7.00 (m, 2H), 4.55 – 4.39 (m, 3H), 3.65 – 3.41 (m, 2H), 2.78-2.69 (m, 2H), 2.44 – 2.28 (m, 1H), 2.22 – 2.12 (m, 1H), 2.08 – 1.91 (m, 4H), 1.47 (d, J = 7.4 Hz, 9H). ^{13}C NMR (151 MHz, CDCl_3) δ 171.74, 154.45, 153.76, 149.04, 148.83, 138.78, 138.58, 129.48, 129.33, 121.39, 121.06, 82.98 (d, J = 164.8 Hz), 82.95 (d, J = 165.4 Hz), 80.20, 79.95, 59.18, 59.06, 46.62, 46.44, 32.02 (d, J = 19.9 Hz), 31.99 (d, J = 19.6 Hz), 31.06, 30.74, 30.71, 28.43, 24.48, 23.70. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -219.84 – -220.64 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{19}\text{H}_{27}\text{FNO}_4$: 352.1919; Found 352.1916



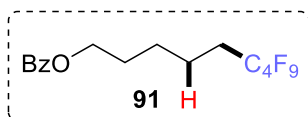
Prepared according to General Procedure C and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.21-7.16 (m, 2H), 7.02 – 6.97 (m, 2H), 4.45 (dt, J = 47.2, 5.9 Hz, 2H), 2.79 – 2.67 (m, 2H), 2.54 (t, J = 7.5 Hz, 2H), 2.06 – 1.93 (m, 2H), 1.74 (p, J = 7.5 Hz, 2H), 1.42 – 1.25 (m, 24H), 0.88 (t, J = 7.0 Hz, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 172.49, 149.02, 138.50, 129.37, 121.51, 82.99 (d, J = 164.9 Hz), 34.42, 32.02 (d, J = 19.9 Hz), 31.94, 30.73 (d, J = 5.1 Hz), 29.71, 29.69, 29.67, 29.66, 29.61, 29.47, 29.37, 29.27, 29.12, 24.98, 22.70, 14.13. ^{19}F NMR (565 MHz, Chloroform-*d*) δ -220.23 (tt, J = 47.5, 25.5 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{25}\text{H}_{42}\text{FO}_2$: 393.3164; Found 393.3148



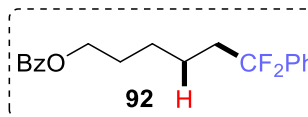
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.07 – 8.00 (m, 2H), 7.59 – 7.54 (m, 1H), 7.44 (t, J = 7.8 Hz, 2H), 4.34 (t, J = 6.5 Hz, 2H), 2.10-1.99 (m, 2H), 1.86-1.78 (m, 2H), 1.73-1.64 (m, 2H), 1.57 – 1.52 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.64, 132.97, 130.33, 129.56, 128.40, 119.24 (qt, J = 285.2, 36.4 Hz), 115.76 (tq, J = 251.5, 37.5 Hz), 64.58, 30.62 (t, J = 22.2 Hz), 28.47, 25.75, 20.14 (t, J = 3.7 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -85.45, -118.22 (t, J = 18.5 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{16}\text{F}_5\text{O}_2$: 311.1065; Found 311.1060



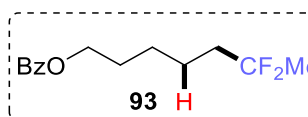
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.08 – 8.00 (m, 2H), 7.59 – 7.54 (m, 1H), 7.44 (t, J = 7.8 Hz, 2H), 4.34 (t, J = 6.5 Hz, 2H), 2.15 – 2.03 (m, 2H), 1.87 – 1.77 (m, 2H), 1.73-1.66 (m, 2H), 1.59 – 1.51 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.65, 132.98, 130.34, 129.57, 128.41, 117.93 (qt, J = 287.5, 34.1 Hz), 117.71 (tt, J = 253.2, 30.9 Hz), 108.93 (tq, J = 263.4, 37.0 Hz), 64.59, 30.58 (t, J = 22.4 Hz), 28.49, 25.77, 19.97 (t, J = 3.8 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -80.61 (t, J = 10.1 Hz), -115.36 (tdt, J = 20.0, 14.7, 10.0 Hz), -127.83 (d, J = 6.0 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{15}\text{H}_{16}\text{F}_7\text{O}_2$: 361.1003; Found 361.1029



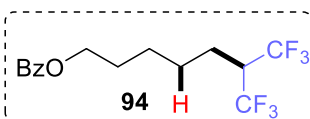
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.08 – 7.99 (m, 2H), 7.61 – 7.52 (m, 1H), 7.44 (t, J = 7.6 Hz, 2H), 4.34 (t, J = 6.5 Hz, 2H), 2.16-2.03 (m, 2H), 1.87 – 1.77 (m, 2H), 1.77-1.65 (m, 2H), 1.60 – 1.49 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.61, 132.93, 130.31, 129.53, 128.37, 120.54-106.87 (m, C_4F_9), 64.55, 30.72 (t, J = 22.4 Hz), 28.46, 25.73, 19.95 (t, J = 3.8 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -81.09 (t, J = 11.2 Hz), -114.59 (dd, J = 37.7, 18.2 Hz), -124.50 (d, J = 11.6 Hz), -125.98 – -126.13 (m). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{16}\text{H}_{16}\text{F}_9\text{O}_2$: 411.1001; Found 411.0992



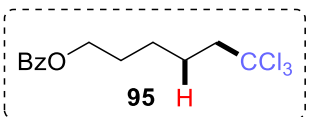
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.08 – 7.99 (m, 2H), 7.59 – 7.52 (m, 1H), 7.49 – 7.36 (m, 7H), 4.29 (t, J = 6.6 Hz, 2H), 2.23 – 2.09 (m, 2H), 1.81 – 1.71 (m, 2H), 1.56-1.43 (m, 2.3 Hz, 4H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.61, 137.36 (t, J = 26.6 Hz), 132.86, 130.38, 129.61, 129.52, 128.38, 128.34, 124.89 (t, J = 6.2 Hz), 122.97 (t, J = 242.1 Hz), 64.76, 39.00 (t, J = 27.6 Hz), 28.52, 25.80, 22.28 (t, J = 4.0 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -95.51 (t, J = 16.3 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{19}\text{H}_{24}\text{F}_2\text{NO}_2$: 336.1770; Found 336.1762



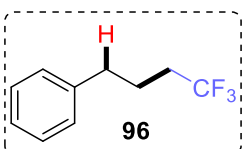
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.04 (d, J = 7.7 Hz, 2H), 7.56 (t, J = 7.4 Hz, 1H), 7.44 (t, J = 7.6 Hz, 2H), 4.33 (t, J = 6.6 Hz, 2H), 1.92-1.75 (m, 4H), 1.60 – 1.46 (m, 7H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.66, 132.89, 130.41, 129.54, 128.36, 124.23 (t, J = 237.7 Hz), 64.78, 37.85 (t, J = 25.4 Hz), 28.57, 25.88, 23.28 (t, J = 28.1 Hz), 22.46 (t, J = 4.7 Hz). ^{19}F NMR (564 MHz, Chloroform-*d*) δ -90.59 (q, J = 17.5 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{14}\text{H}_{19}\text{F}_2\text{O}_2$: 257.1348; Found 257.1344



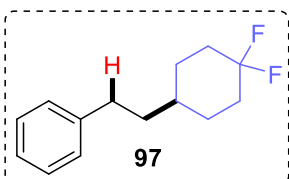
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.07 – 7.98 (m, 2H), 7.59 – 7.53 (m, 1H), 7.45 (t, J = 7.8 Hz, 2H), 4.33 (t, J = 6.5 Hz, 2H), 2.90-2.79 (m, 1H), 1.90 – 1.70 (m, 4H), 1.64-1.57 (m, 2H), 1.54 – 1.47 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.64, 132.95, 130.32, 129.54, 128.38, 123.97 (q, J = 281.3 Hz), 123.95 (q, J = 281.6 Hz), 64.63, 47.99 (hept, J = 27.8 Hz), 28.35, 26.93, 25.95, 23.65 (hept, J = 1.9 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -67.05 (d, J = 8.1 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{15}\text{H}_{17}\text{F}_6\text{O}_2$: 343.1127; Found 343.1121



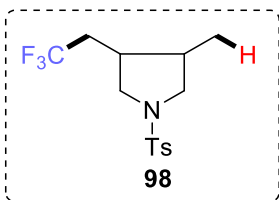
Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 8.08 – 8.02 (m, 2H), 7.59-7.54 (m, 1H), 7.49 – 7.41 (m, 2H), 4.36 (t, J = 6.5 Hz, 2H), 2.76 – 2.67 (m, 2H), 1.91 – 1.79 (m, 4H), 1.60-1.55 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 166.63, 132.94, 130.29, 129.55, 128.37, 99.88, 64.64, 55.00, 28.50, 26.17, 24.99. GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{13}\text{H}_{15}\text{Cl}_3\text{O}_2$: 308.0138; Found 308



Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.32 – 7.28 (m, 2H), 7.23 – 7.19 (m, 1H), 7.19 – 7.14 (m, 2H), 2.69 (t, J = 7.6 Hz, 2H), 2.13 – 2.03 (m, 2H), 1.93 – 1.86 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ 140.68, 128.56, 128.38, 127.19 (q, J = 276.5 Hz), 126.27, 34.64, 33.10 (q, J = 28.5 Hz), 23.50 (q, J = 2.9 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -66.18 (t, J = 10.8 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁷

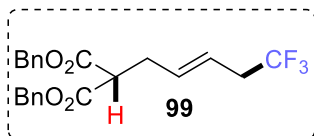


Prepared according to General Procedure D and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.30 – 7.26 (m, 2H), 7.21 – 7.15 (m, 3H), 2.68 – 2.59 (m, 2H), 2.13 – 2.02 (m, 2H), 1.88 – 1.78 (m, 2H), 1.77 – 1.62 (m, 2H), 1.61 – 1.57 (m, 2H), 1.39 – 1.29 (m, 3H). ^{13}C NMR (151 MHz, CDCl_3) δ 142.38, 128.39, 128.29, 125.79, 123.80 (dd, J = 241.9, 239.5 Hz), 37.52 (d, J = 2.6 Hz), 35.23, 33.61, 33.45 (d, J = 2.9 Hz), 33.42, 33.30, 28.89 (d, J = 9.4 Hz). ^{19}F NMR (565 MHz, Chloroform-*d*) δ -81.46 – -111.74 (m). GC-MS: $[\text{M}]^+$ calcd. for $\text{C}_{14}\text{H}_{18}\text{F}_2$: 224.1377; Found 224



Prepared according to General Procedure A and obtained as colorless oil. ^1H NMR (600 MHz, Chloroform-*d*) δ 7.76 – 7.67 (m, 2H), 7.34 (dd, J = 8.2, 2.3 Hz, 2H), 3.64-3.50 (m, 0.21H), 3.56-3.47 (m, 1.06H), 3.38-3.33 (m, 0.79H), 3.11 – 3.04 (m, 1.59H), 3.00-2.95 (m, 0.23H), 2.81-2.76 (m, 0.22H), 2.46-2.41 (m, 3H), 2.34 – 2.23 (m, 1.85H), 2.16 – 2.07 (m, 0.80H), 1.95 – 1.74 (m, 1.55H), 0.97 (d, J = 6.2 Hz, 0.65H), 0.77 (d, J = 6.7 Hz, 2.35H). ^{13}C NMR (151 MHz, CDCl_3) δ 143.62, 143.59, 133.91, 133.67, 129.76, 129.74, 127.50, 127.40, 126.59 (q, J = 277.0 Hz), 126.35 (q, J = 276.9 Hz), 54.46, 53.96, 53.01, 50.41, 39.68 (q, J = 2.6 Hz), 38.69, 35.85 (q, J = 28.9 Hz), 35.73 (q, J = 2.6 Hz), 35.23, 32.41 (q, J = 28.8 Hz), 21.55, 21.54, 15.68, 13.25. ^{19}F NMR (565 MHz, Chloroform-*d*)

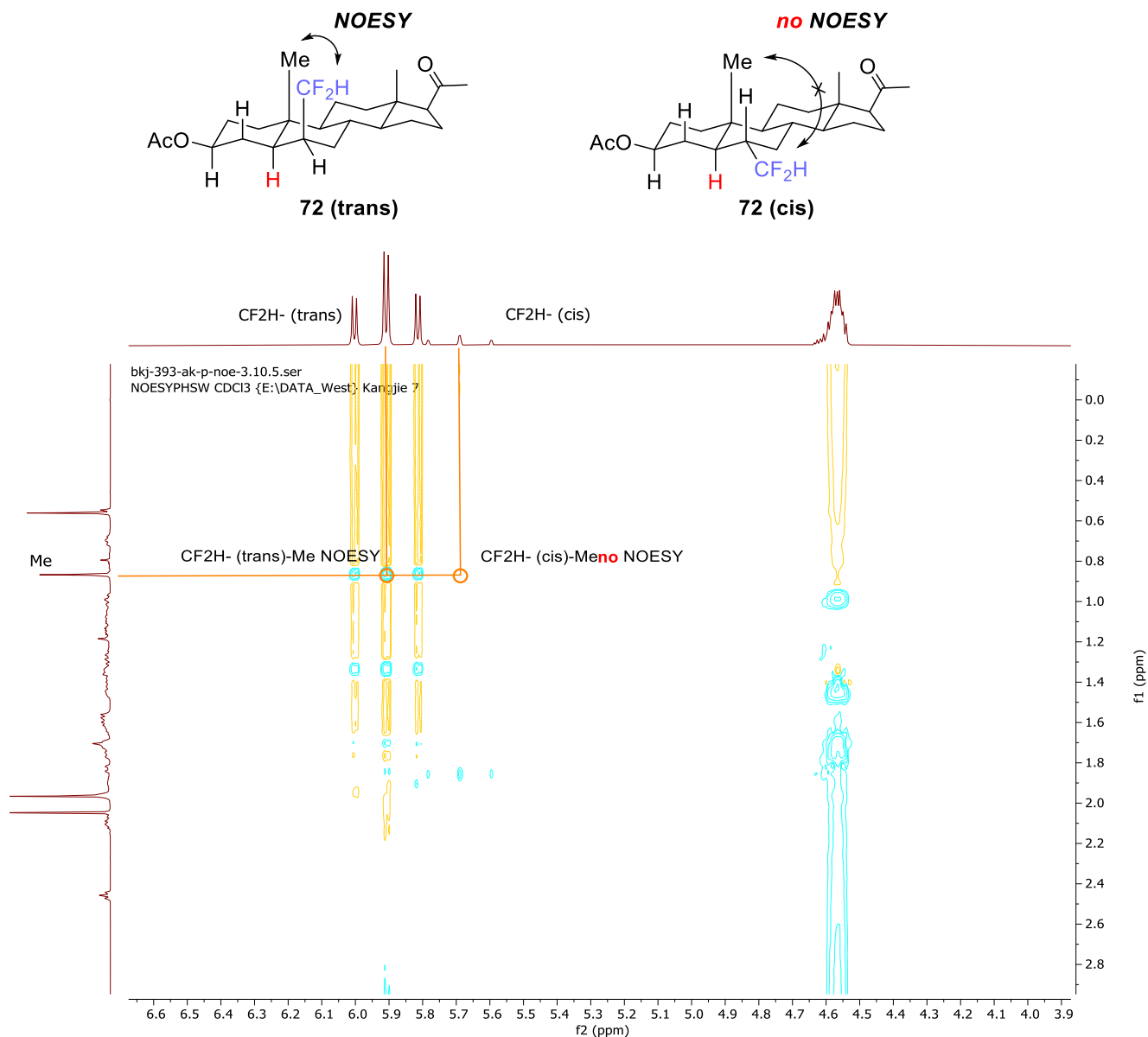
δ -64.99 (t, J = 10.9 Hz), -65.03 (t, J = 10.9 Hz). HRMS APCI: $[M+H]^+$ calcd. for $C_{14}H_{19}F_3NO_2S$: 322.1083; Found 322.1077



Prepared according to General Procedure A and obtained as colorless oil. ¹H NMR (600 MHz, Chloroform-*d*) δ 7.41 – 7.25 (m, 10H), 5.64 (dt, J = 14.6, 7.0 Hz, 1H), 5.44 (dt, J = 14.9, 7.0 Hz, 1H), 5.20 – 5.09 (m, 4H), 3.61-3.44 (m, 1H), 2.78 – 2.59 (m, 4H). ¹³C NMR (151 MHz, CDCl₃) δ 168.37, 135.27, 133.06, 128.58, 128.43, 128.28, 125.75 (q, J = 276.5 Hz), 121.37 (q, J = 3.6 Hz), 67.26, 51.55, 37.19 (q, J = 29.7 Hz), 31.61. ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -66.51 (t, J = 10.9 Hz). The compound characterization was reported in literature and the NMR data matched with previous characterization.¹⁸

2.5 Relative configuration of compound **72**

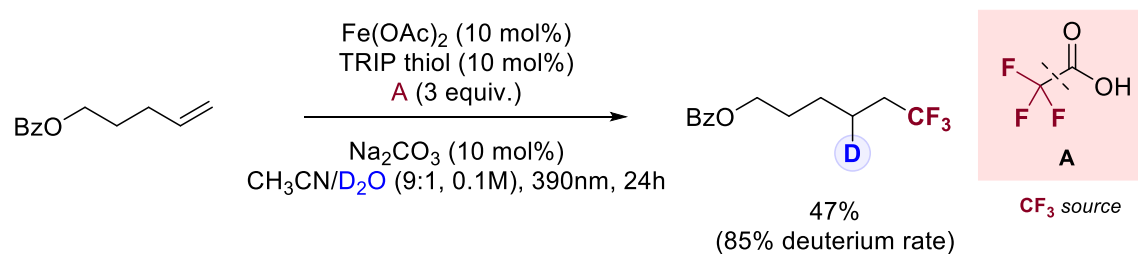
On the basis of NMR experiment, we have assigned the major diastereomers of products **72** as shown below:

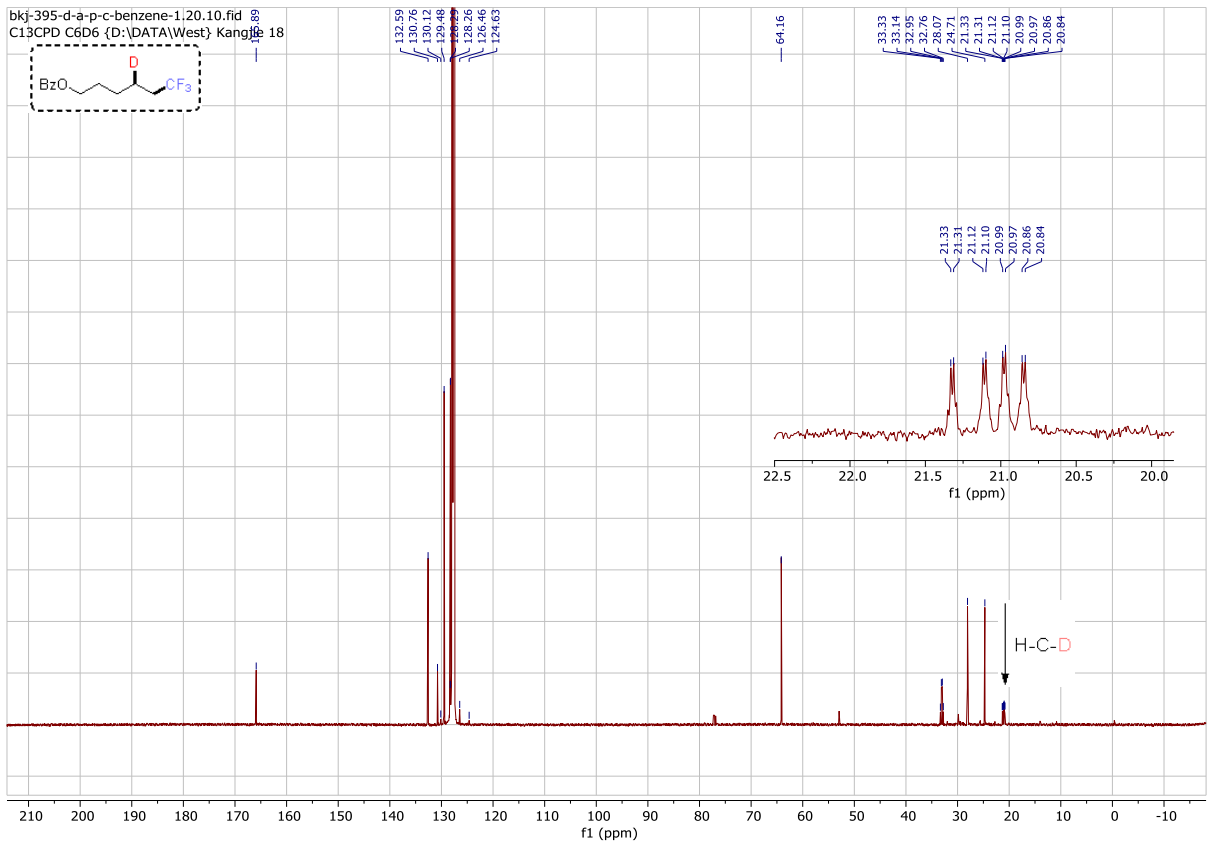


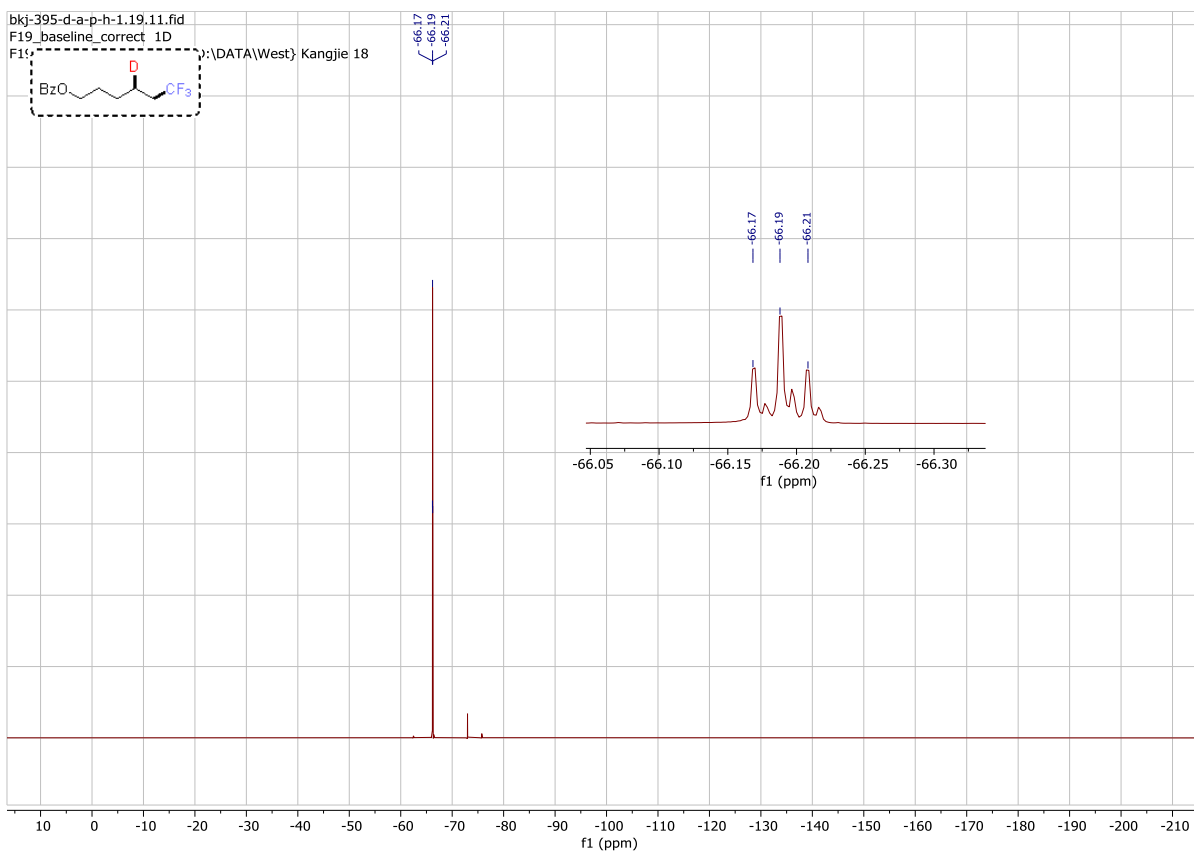
For **72-trans**, a strong NOE was observed for CF₂H (trans)/Me. However, there is no significant NOE observed for CF₂H (cis)/Me of **72-cis**. Therefore, major isomer of **72** should be the trans-product, which also correlates the finding by Wu and co-workers.¹⁹

2.6 Deuterium labelling and kinetic isotope effect (KIE) studies

Deuterium labelling experiments



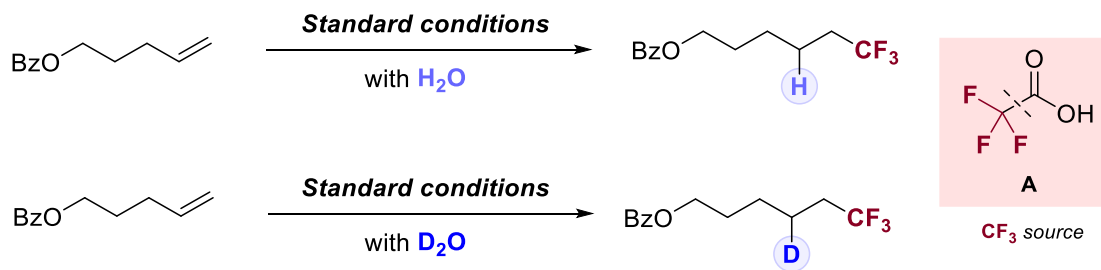




Chemical structure: BzOCCCC[C@H](D)CC(F)(F)F

^1H NMR (600 MHz, Benzene- d_6) δ 8.22 – 8.12 (m, 2H), 7.15 – 7.03 (m, 3H), 4.04 (t, J = 6.6 Hz, 2H), 1.52-1.43 (m, 2H), 1.26 – 1.20 (m, 2H), 1.15-1.08 (m, 1.10H), 0.91-0.85 (m, 2H). ^{13}C NMR (151 MHz, C_6D_6) δ 165.89, 132.59, 130.76, 129.48, 128.26, 127.38 (q, J = 276.5 Hz), 64.16, 33.05 (q, J = 28.3 Hz), 28.07, 24.71, 21.33, 21.31, 21.12, 21.10, 20.99, 20.97, 20.86, 20.84. ^{19}F NMR (565 MHz, Benzene- d_6) δ -66.19 (t, J = 11.3 Hz). HRMS APCI: $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{13}\text{H}_{15}\text{DF}_3\text{O}_2$: 262.1160; Found 262.1156

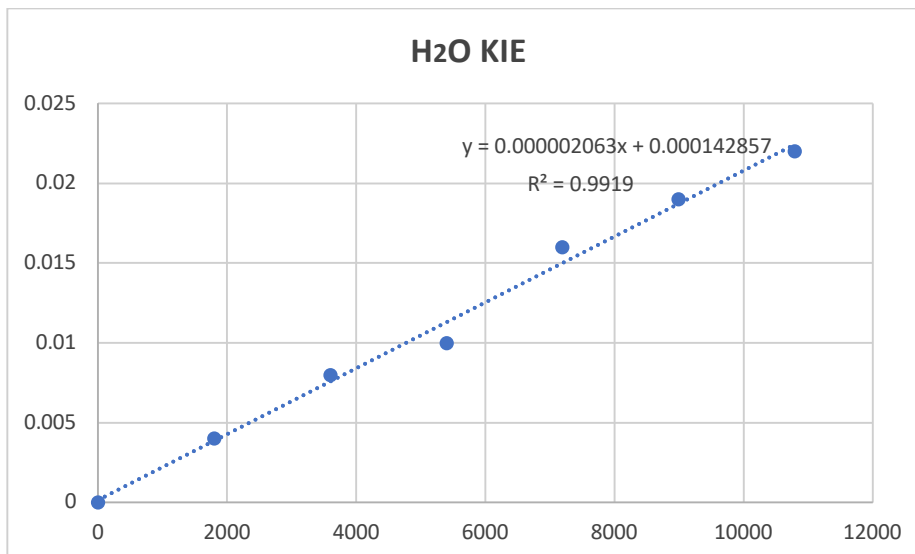
Kinetic isotope effect (KIE) experiments



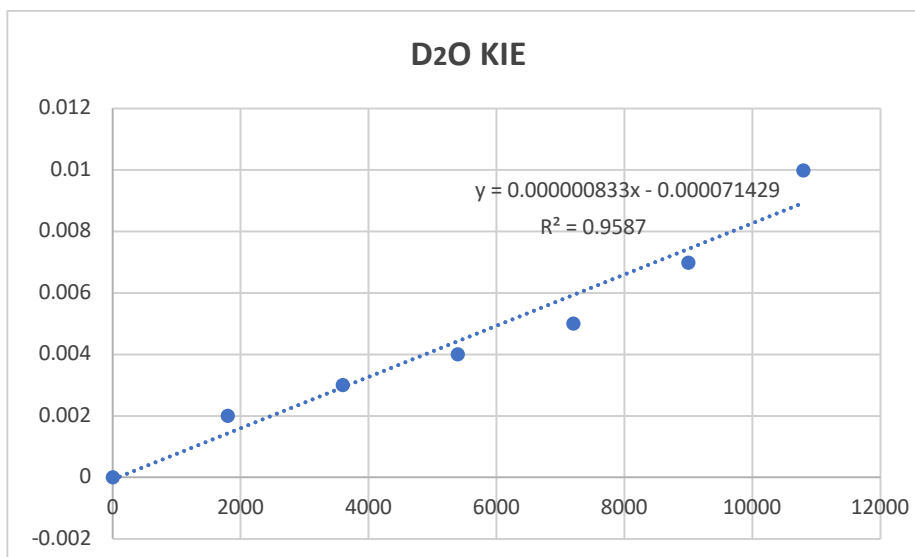
$\text{Fe}(\text{OAc})_2$ and Na_2CO_3 were added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by addition of alkenes, CF_3COOH and TRIP thiol in $\text{MeCN}/\text{H}_2\text{O}$ (9:1, 0.1 M in regard to alkenes) via syringe under N_2 . Note: Both reactions were run on 0.2 mmol of the substrate and PhCF_3 was added as internal standard. The reaction mixture was placed under 390nm Kessil® light. At different time points, the aliquot of reaction mixture is directly transferred to NMR tube and reaction progress was monitored by ^{19}F NMR.

t (s)	H_2O (M)	t (s)	D_2O (M)
0	0	0	0
1800	0.004	1800	0.002
3600	0.008	3600	0.003
5400	0.01	5400	0.004
7200	0.016	7200	0.005
9000	0.019	9000	0.007
10800	0.022	10800	0.01

entry	mmol acid	mmol alkene	mmol iron	mmol thiol	initial rate (M/s)
H ₂ O KIE	0.3	0.1	0.01	0.01	2.06×10^{-6}
D ₂ O KIE	0.3	0.1	0.01	0.01	0.83×10^{-6}



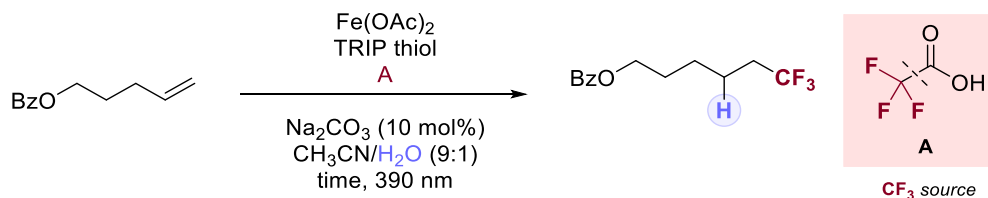
Reaction with H₂O (standard conditions): $y = (2.06 \times 10^{-6})(k_H)x + (1.4 \times 10^{-4})$, $R^2 = 0.9916$



Reaction with D₂O: $y = (0.83 \times 10^{-6})(k_D)x - (7.1 \times 10^{-4})$, $R^2 = 0.9587$

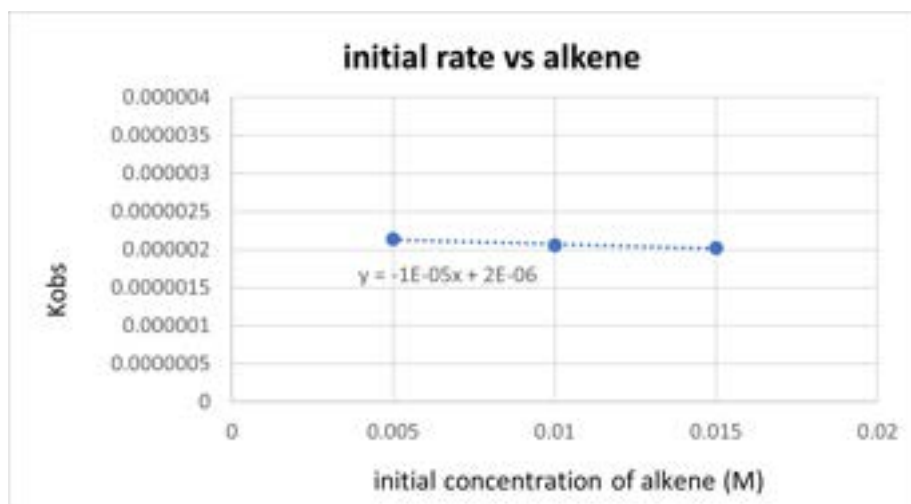
KIE: $k_H/k_D = 2.5$

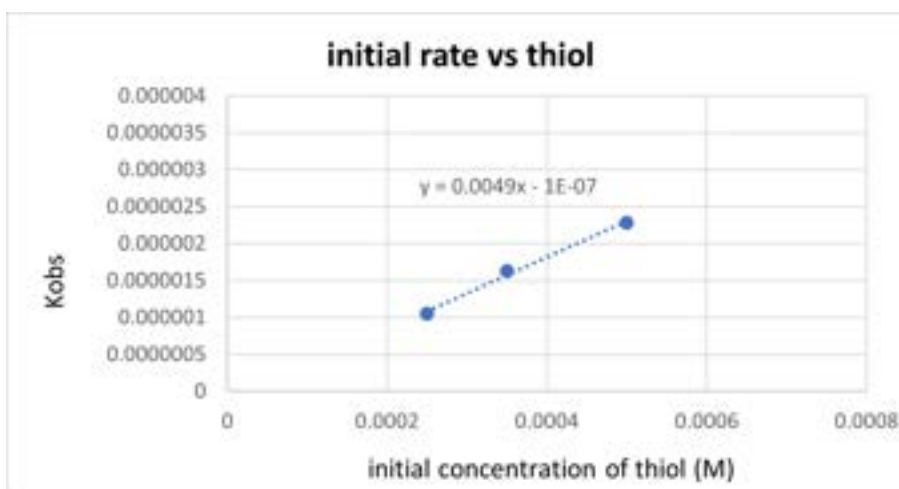
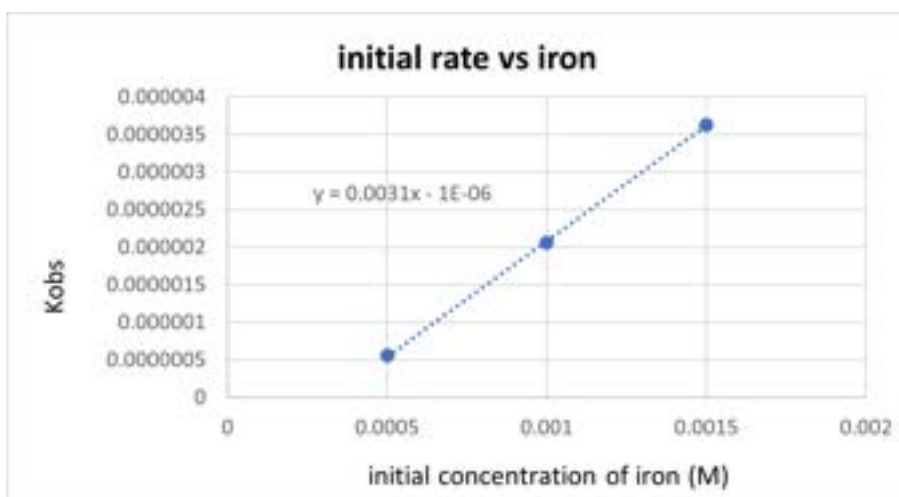
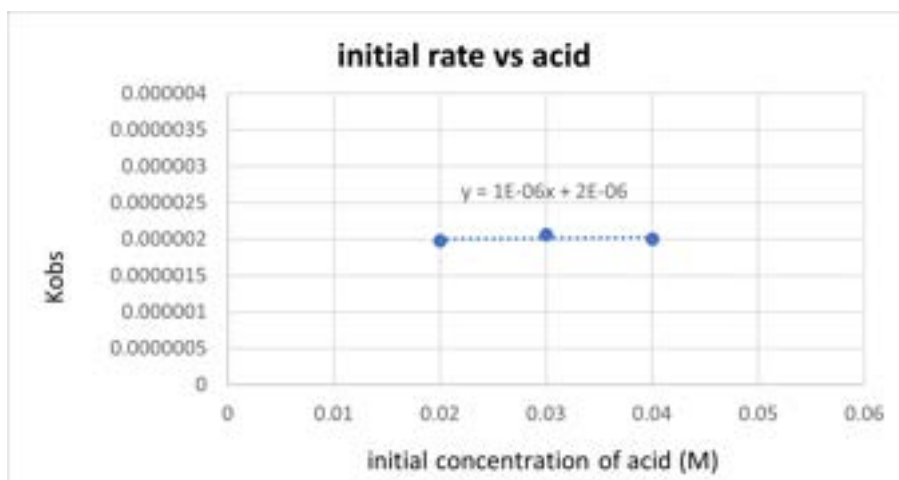
2.7 Kinetic studies to determine reaction rate law



$\text{Fe}(\text{OAc})_2$ and Na_2CO_3 were added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by addition of alkenes, CF_3COOH and TRIP thiol in $\text{MeCN}/\text{H}_2\text{O}$ (9:1, 0.1 M in regard to alkenes) via syringe under N_2 . Note: All reactions were run on 0.2 mmol of the substrate and PhCF_3 was added as internal standard. The reaction mixture was placed under 390nm Kessil® light. At different time points, the aliquot of reaction mixture is directly transferred to NMR tube and reaction progress was monitored by ^{19}F NMR.

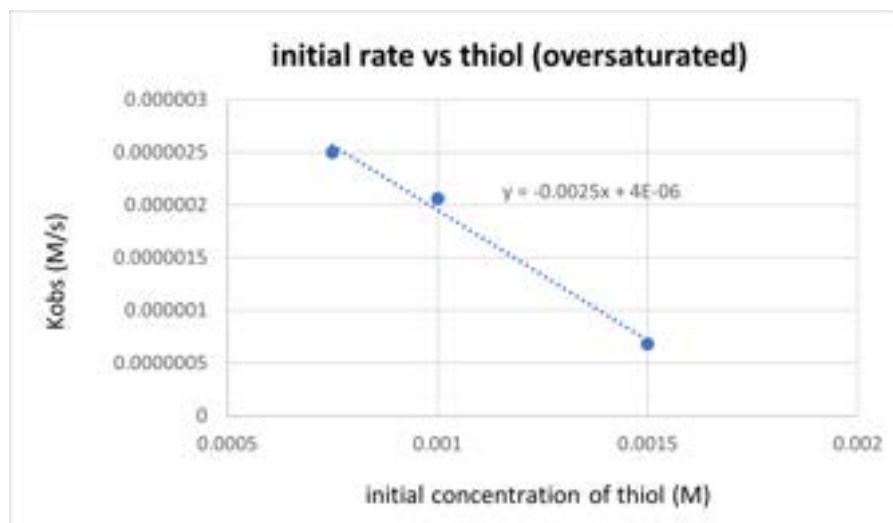
entry	mmol acid	mmol alkene	mmol iron	mmol thiol	initial rate (M/s)
1	0.3	0.1	0.01	0.01	2.06×10^{-6}
2	0.3	0.05	0.01	0.01	2.14×10^{-6}
3	0.3	0.15	0.01	0.01	2.02×10^{-6}
4	0.2	0.1	0.01	0.01	1.98×10^{-6}
5	0.4	0.1	0.01	0.01	2×10^{-6}
6	0.3	0.1	0.005	0.01	5.6×10^{-7}
7	0.3	0.1	0.015	0.01	3.63×10^{-6}
8	0.3	0.1	0.01	0.0025	1.05×10^{-6}
9	0.3	0.1	0.01	0.0035	1.63×10^{-6}
10	0.3	0.1	0.01	0.005	2.28×10^{-6}
11	0.3	0.1	0.01	0.0075	2.5×10^{-6}





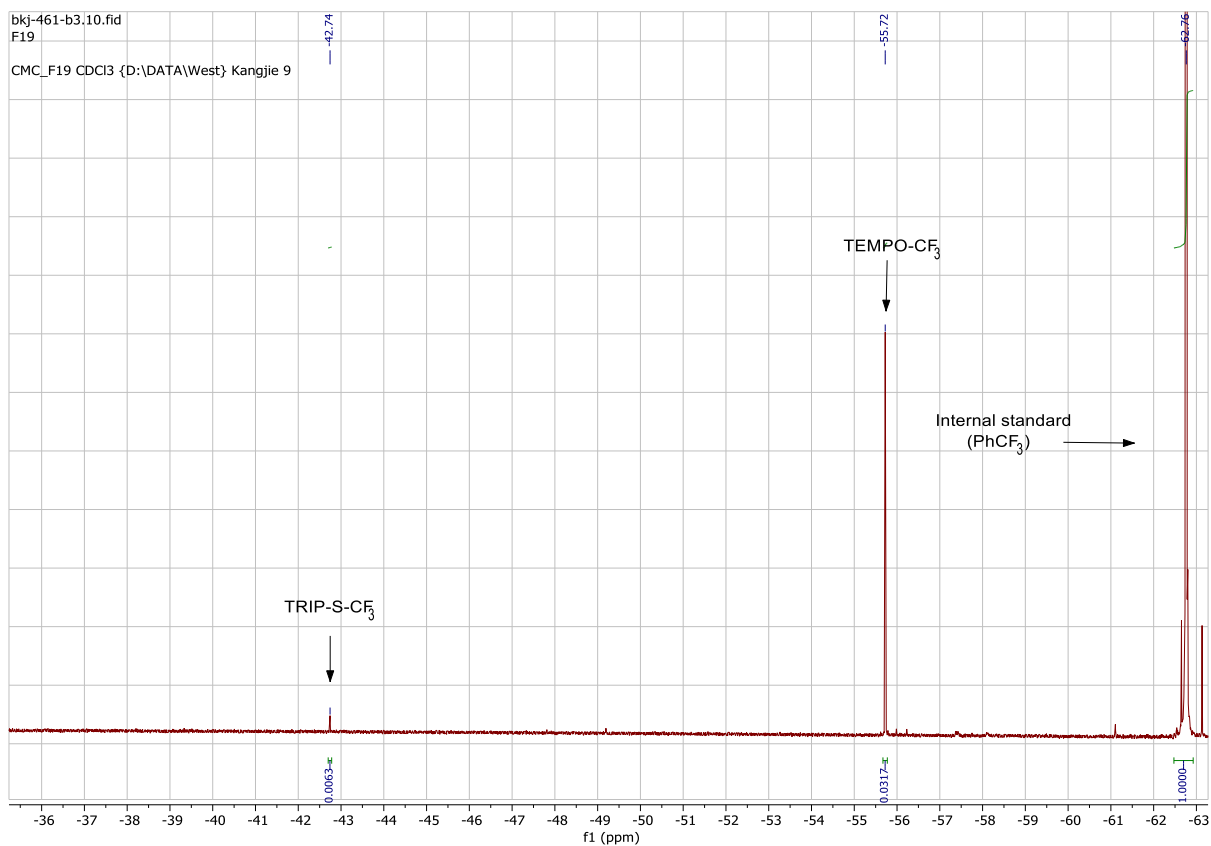
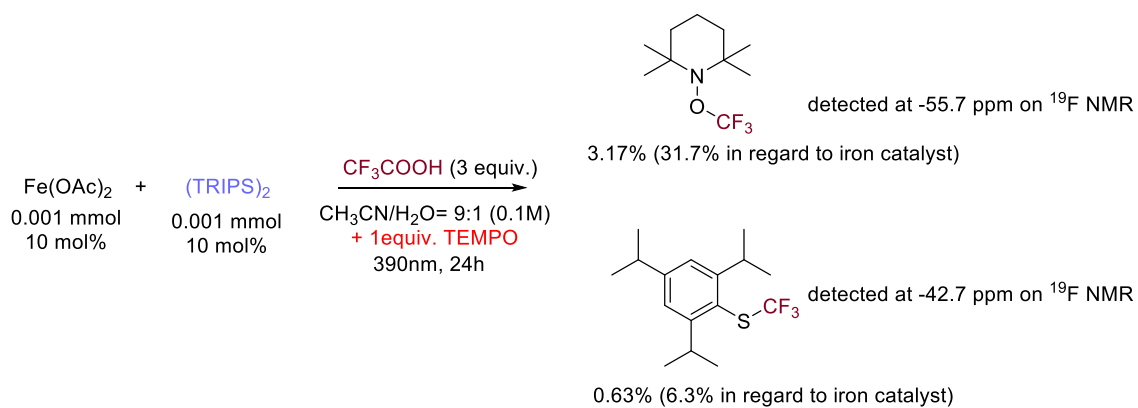
With higher loading of thiol catalyst (15 mol%), negative order of thiol is observed. The oversaturation of thiol catalyst may lead to other side reactions, consuming thiol and circumventing the following dual catalytic cycle with iron and other reactants.

entry	mmol acid	mmol alkene	mmol iron	mmol thiol	initial rate (M/s)
12	0.3	0.1	0.01	0.015	6.8×10^{-7}

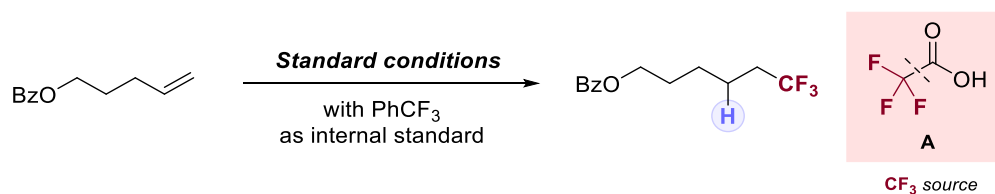


Indicated by previous reports²⁰⁻²³ and our study on disulfide formation (see 2.7 Study of disulfide formation), thiol catalyst can form disulfide under light irradiation. Furthermore, TEMPO-trapping experiment demonstrates that higher concentration of disulfide could quench CF_3 radical, indicated by peak at -42.74ppm on ^{19}F NMR, suggesting the generation of TRIP- SCF_3 .²⁴

With higher concentration of TRIP thiol in the system, it is more likely to form disulfide under light irradiation, which can sequester the CF_3 radical, leading to drastically decreased rate of thiol and showing negative order when reaching oversaturation.

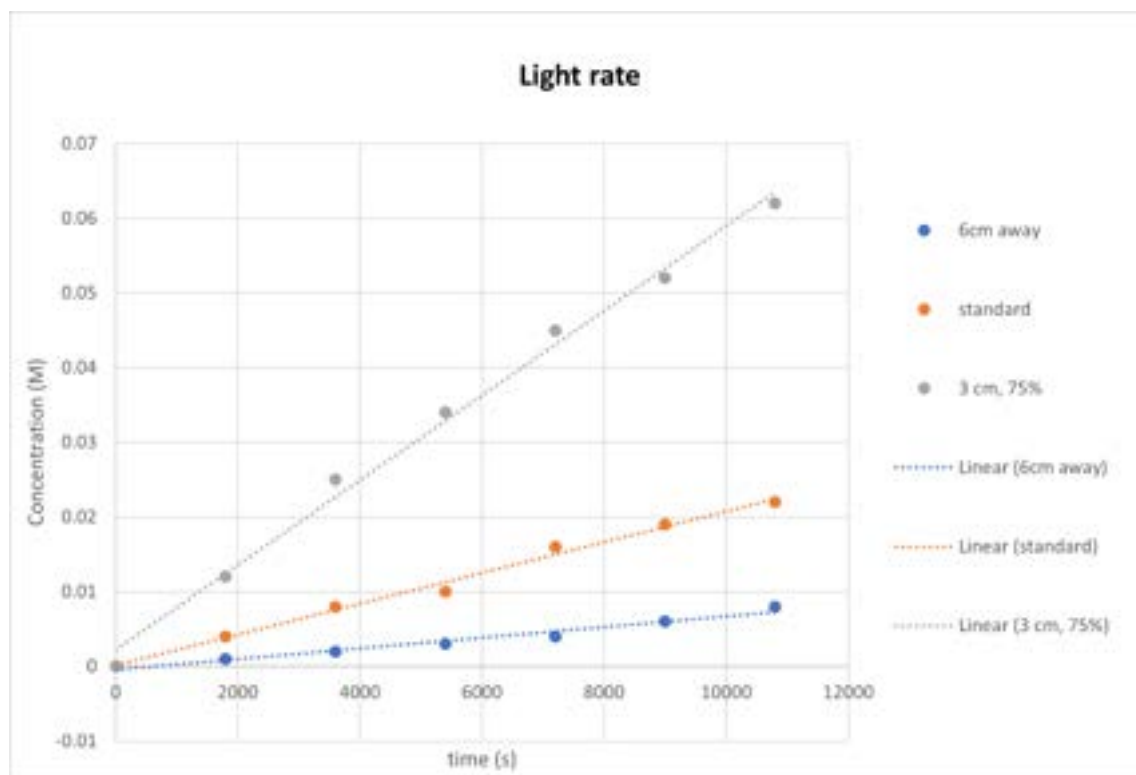


2.8 Light intensity impact on reaction rate

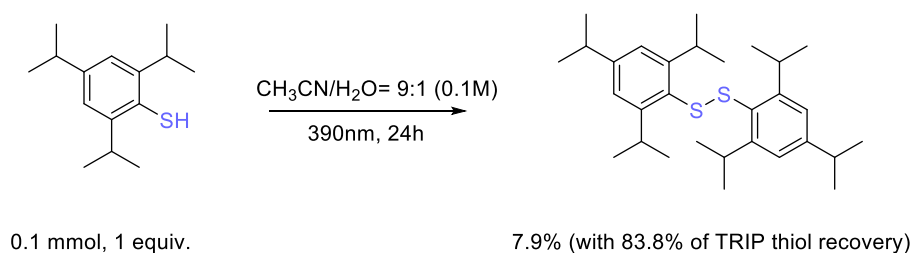


Following General Procedure A, all reactions were run on 0.2 mmol of the substrate and PhCF₃ was added as internal standard. The reaction mixture was placed under 390nm Kessil® light. At different time points, the aliquot of reaction mixture is directly transferred to NMR tube and reaction progress was monitored by ¹⁹F NMR. Reaction rate is correlated to light intensity positively, indicating a light-dependent character of the system.

entry	mmol acid	mmol alkene	mmol iron	mmol thiol	initial rate (M/s)
25%, 3cm	0.3	0.1	0.01	0.01	2.06 x 10 ⁻⁶
75%, 3cm	0.3	0.1	0.01	0.01	5.68 x 10 ⁻⁶
25%, 6cm	0.3	0.1	0.01	0.01	7.14 x 10 ⁻⁷



2.9 Study of the disulfide formation

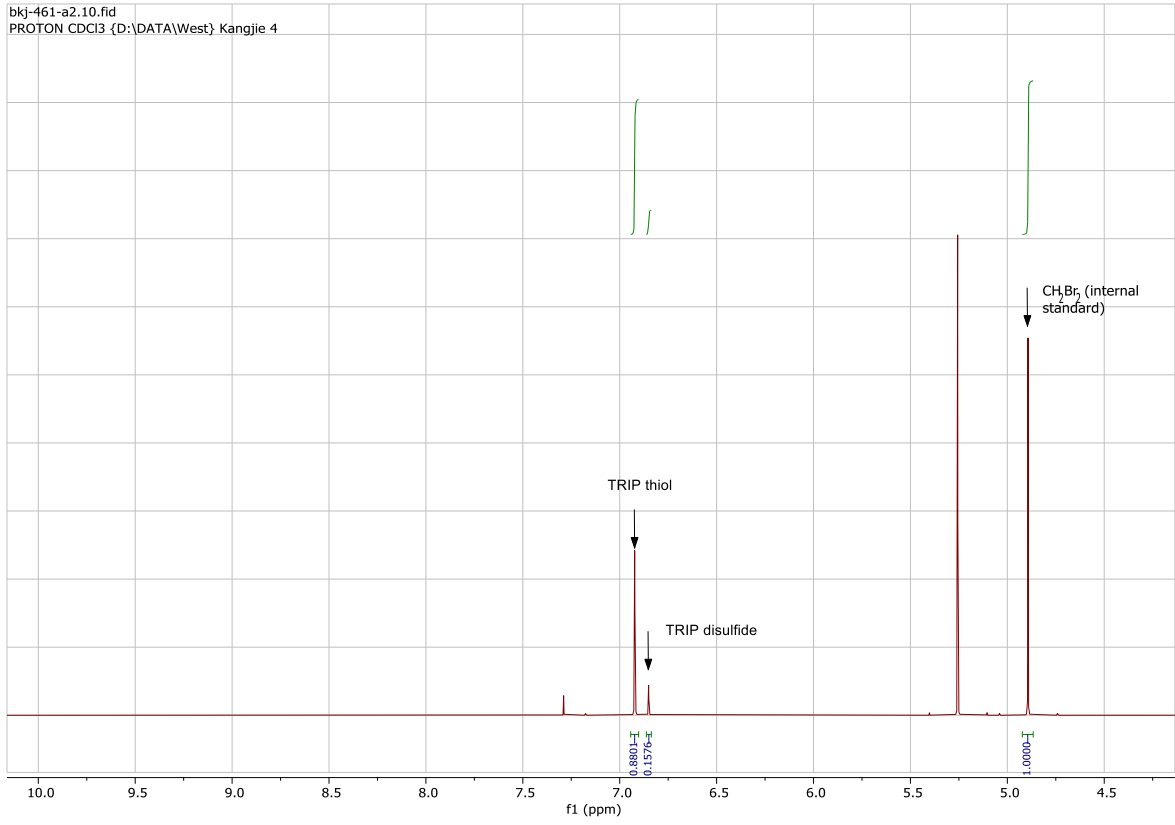


TRIP thiol (0.1 mmol, 1 equiv. was added to MeCN/H₂O (9:1, 0.1 M) via syringe after backfilling with N₂) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. With solid formation observed over time, crude NMR indicated the formation of TRIP disulfide in 7.9% yield with 83.8% TRIP thiol recovery. The formation of disulfide could further engage in homolysis under light irradiation, leading to generation of thiyl radical, which can oxidize Fe(II) catalyst to photoreactive Fe(III) species, initiating the fluoroalkyl radical generation.²⁰⁻²³

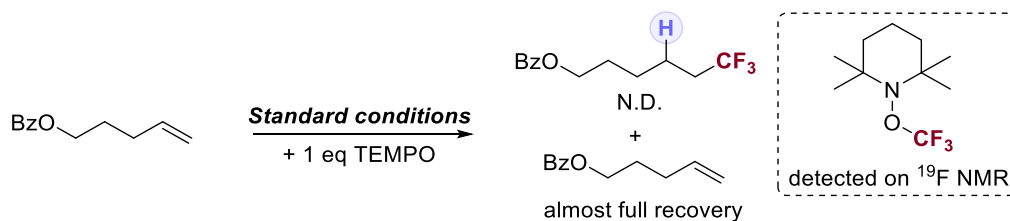


Figure S1. Left: solid formation (TRIP disulfide). Right: TLC spotting (left to right: reaction mixture, pure TRIP thiol, pure TRIP disulfide).

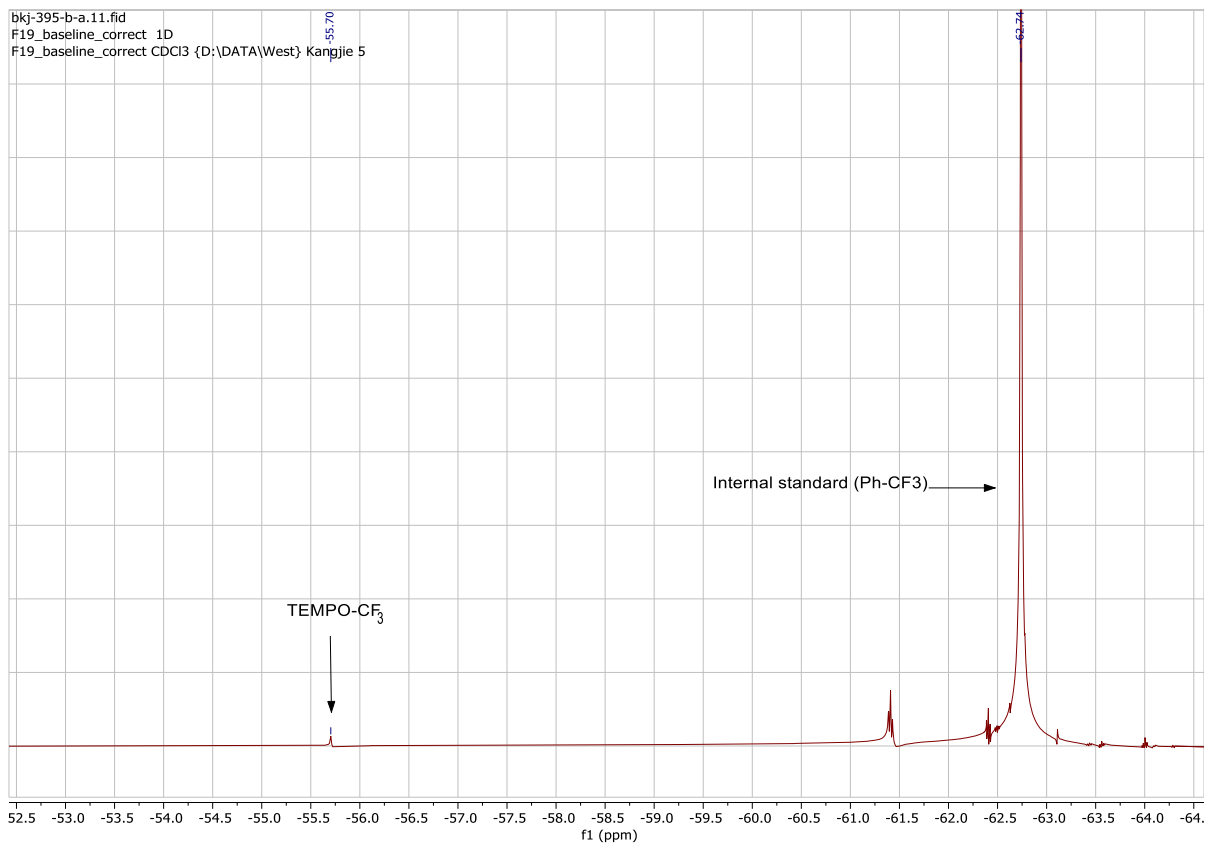
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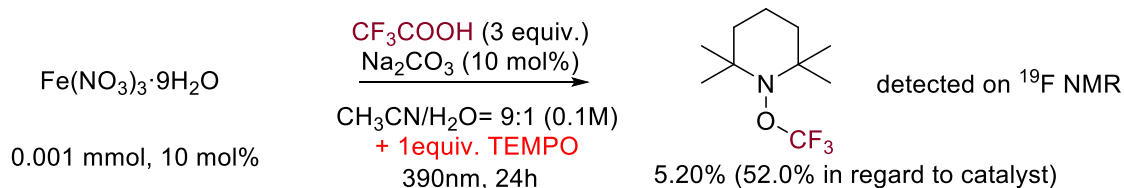
2.10 TEMPO experiment study



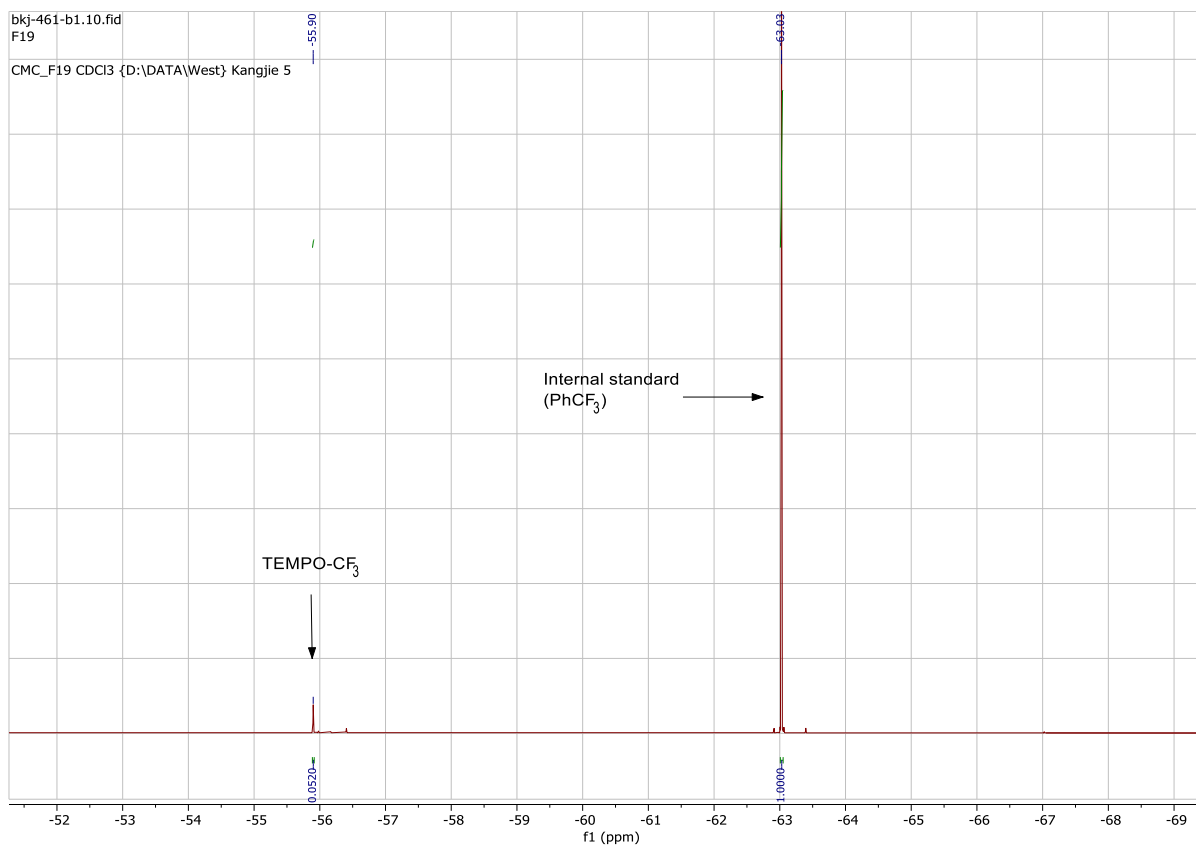
Fe(OAc)₂ (10 mol%, 0.1 equiv.), Na₂CO₃ (10 mol%, 0.1 equiv.) and TEMPO (1 equiv.) were added via syringe after backfilling with N₂) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N₂ (repeated for 4 times), followed by addition of alkenes (0.1 mmol, 1.0 equiv.) TRIP thiol (10 mol%, 0.1 equiv.) and CF₃COOH (0.30 mmol, 3.0 equiv) in MeCN/H₂O (9:1, 0.1 M in regard to alkenes) via syringe under N₂. The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated and directly applied for crude ¹⁹F NMR (with PhCF₃ as internal standard). The detection of TEMPO-CF₃ at -55.70 ppm suggested CF₃ radical generation in the system.²⁵



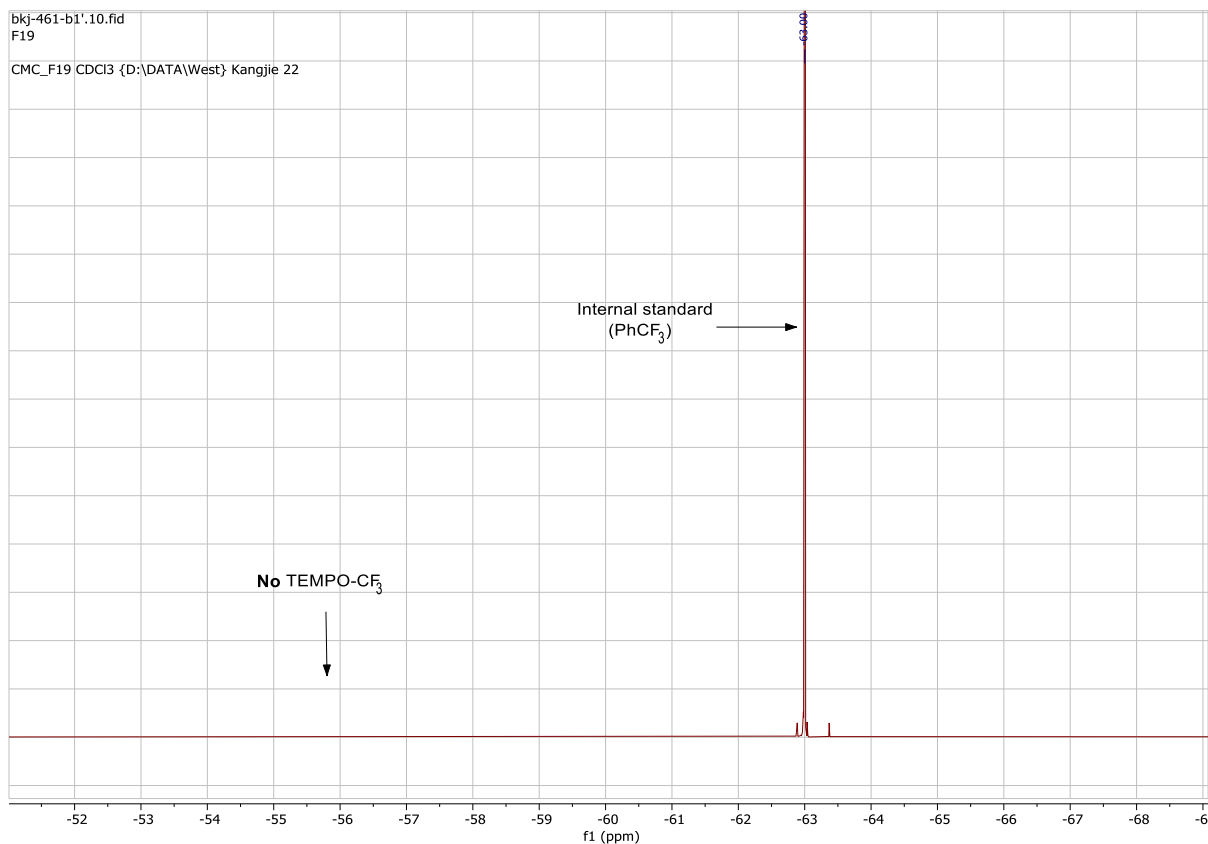
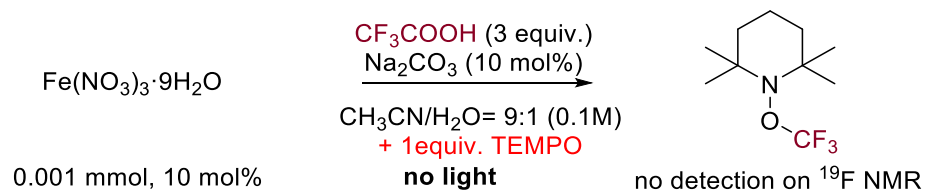
2.11 Evidence of visible-light induced homolysis of trifluoroacetic acid with iron



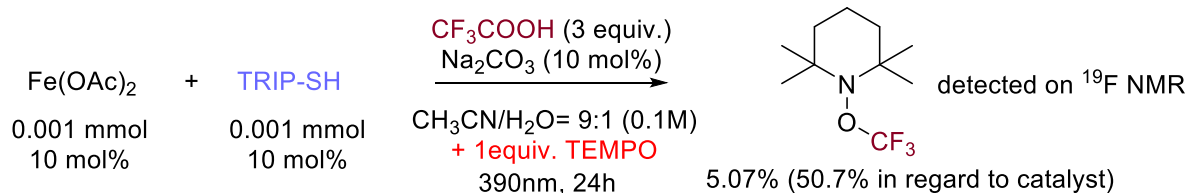
$\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ (10 mol%, 0.1 equiv.), Na_2CO_3 (10 mol%, 0.1 equiv.) and TEMPO (1 equiv.) were added via syringe after backfilling with N_2) was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by CF_3COOH (0.30 mmol, 3.0 equiv.) in MeCN/ H_2O (9:1, 0.1M) via syringe under N_2 . The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated and directly applied for crude ^{19}F NMR (with PhCF_3 as internal standard).



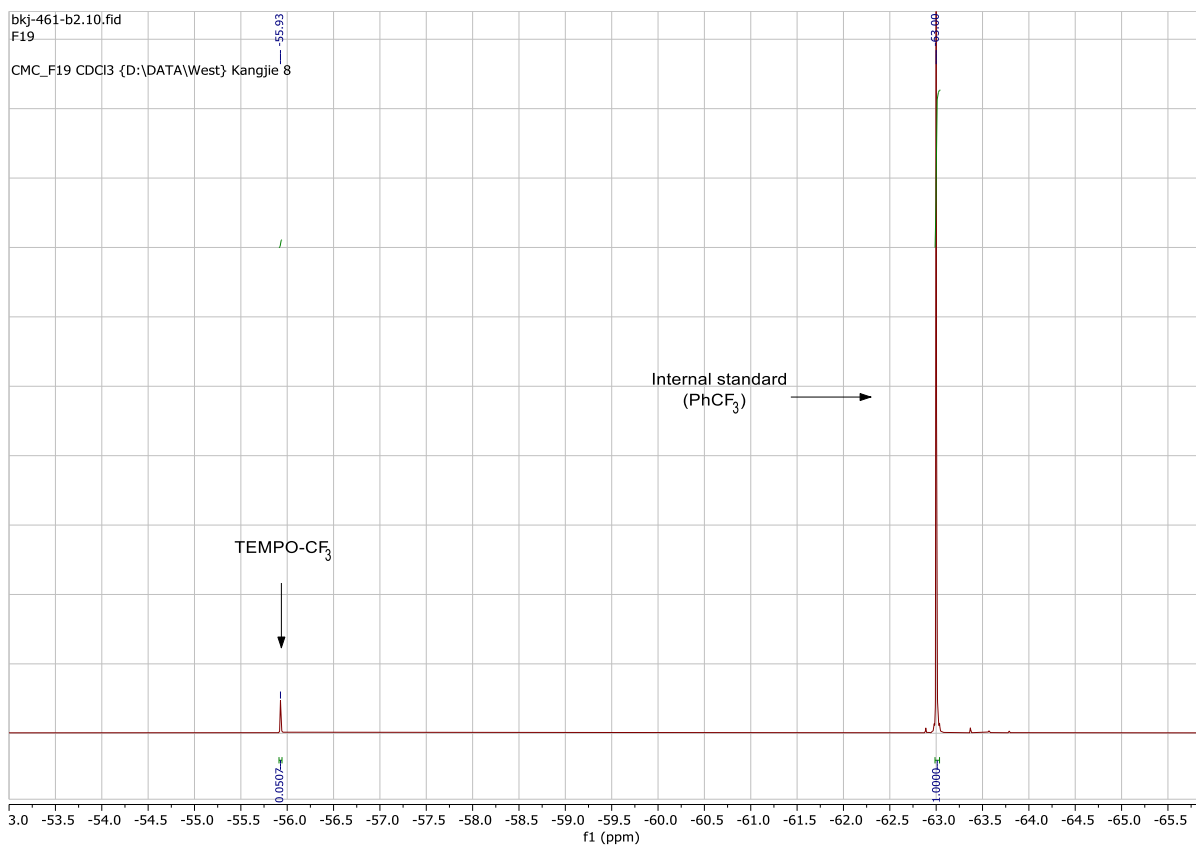
Indicated by ¹⁹F NMR, photoactive Fe(III) (Fe(NO₃)₃·9H₂O) could induce the homolysis of trifluoroacetic acid (TFA) under light irradiation, as TEMPO-CF₃ was detected at -55.9 ppm in 5.2% yield (52.0% in regard to iron).



Indicated by ^{19}F NMR, in the absence of light irradiation, the homolysis of trifluoroacetic acid (TFA) by Fe(III) ($\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$) is not observed, supporting light is necessary in promoting homolysis of TFA to afford CF_3 radical.

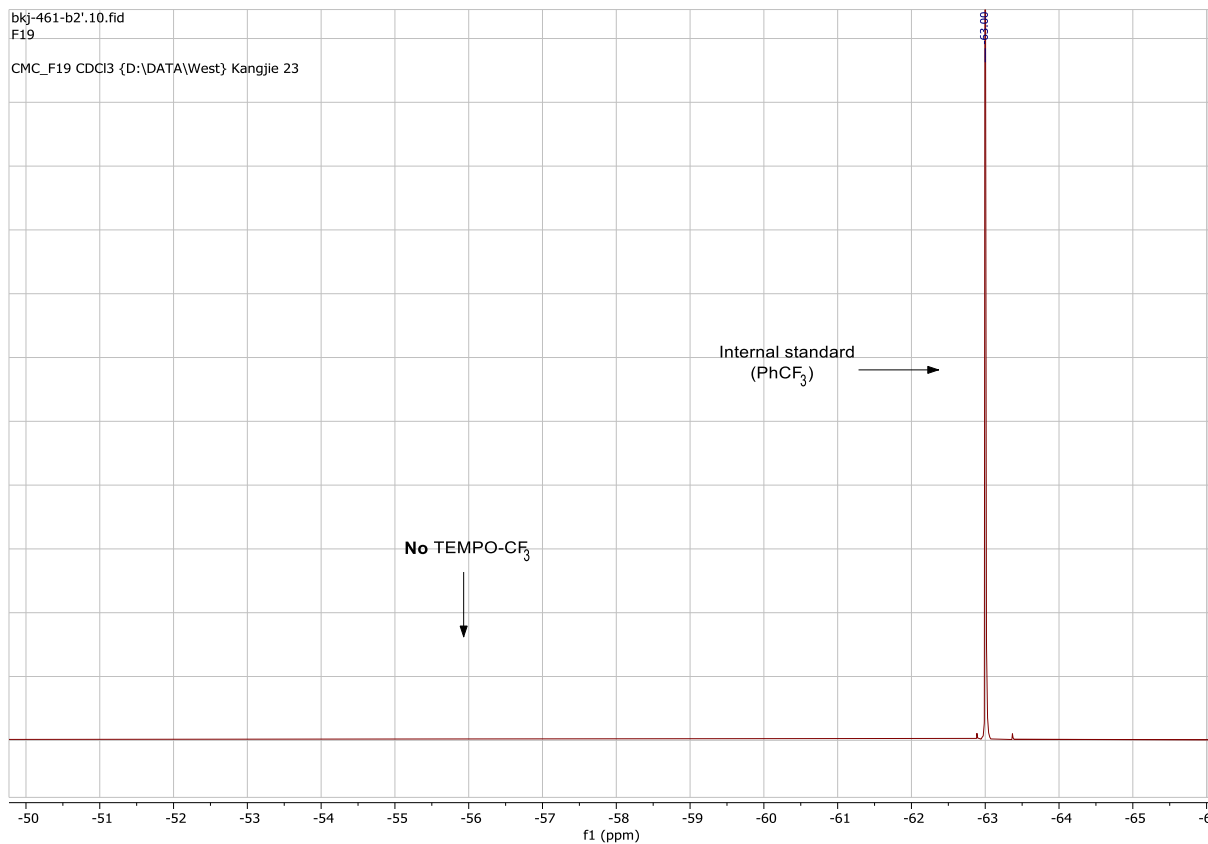
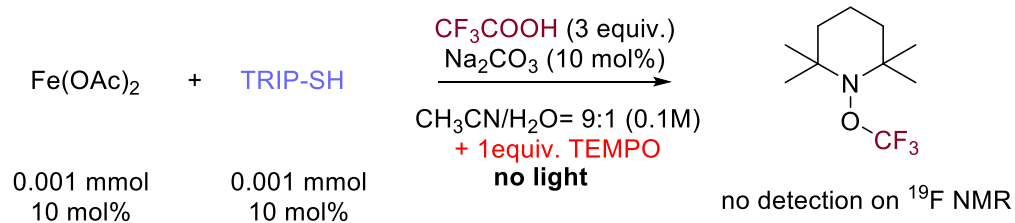


$\text{Fe}(\text{OAc})_2$ (10 mol%, 0.1 equiv.), Na_2CO_3 (10 mol%, 0.1 equiv.) and TEMPO (1 equiv.) were added via syringe after backfilling with N_2 was added in an oven-dried 8-mL test vial containing a Teflon®-coated magnetic stir bar. The vial was evacuated and backfilled with N_2 (repeated for 4 times), followed by TRIPSH (10 mol%, 0.1 equiv.), CF_3COOH (0.30 mmol, 3.0 equiv.) in $\text{MeCN}/\text{H}_2\text{O}$ (9:1, 0.1M) via syringe under N_2 . The reaction mixture was placed under 390nm Kessil® light after sealing the punctured holes of the vial cap with vacuum grease and electric tape/parafilm for better air-tight protection and allowed to react at room temperature for 24 h. Following this, the reaction mixture was filtered through a pad of celite which was subsequently rinsed with DCM. The filtrate was concentrated and directly applied for crude ^{19}F NMR (with PhCF_3 as internal standard).



As demonstrated in the section of 2.7 Study of disulfide formation and illustrated by previous reports,²⁰⁻²³ thiol or disulfide can undergo homolysis under light irradiation, which is capable of oxidizing lower-valent iron species to photoactive iron (III) species, engaging in the following photoinduced homolysis of trifluoroacetic acid.

Indicated by ¹⁹F NMR, the mixture of Fe(OAc)₂ and TRIP thiol could induce the homolysis of trifluoroacetic acid (TFA) under light irradiation, as TEMPO-CF₃ was detected at -55.9 ppm in 5.07% yield (50.7% in regard to iron).

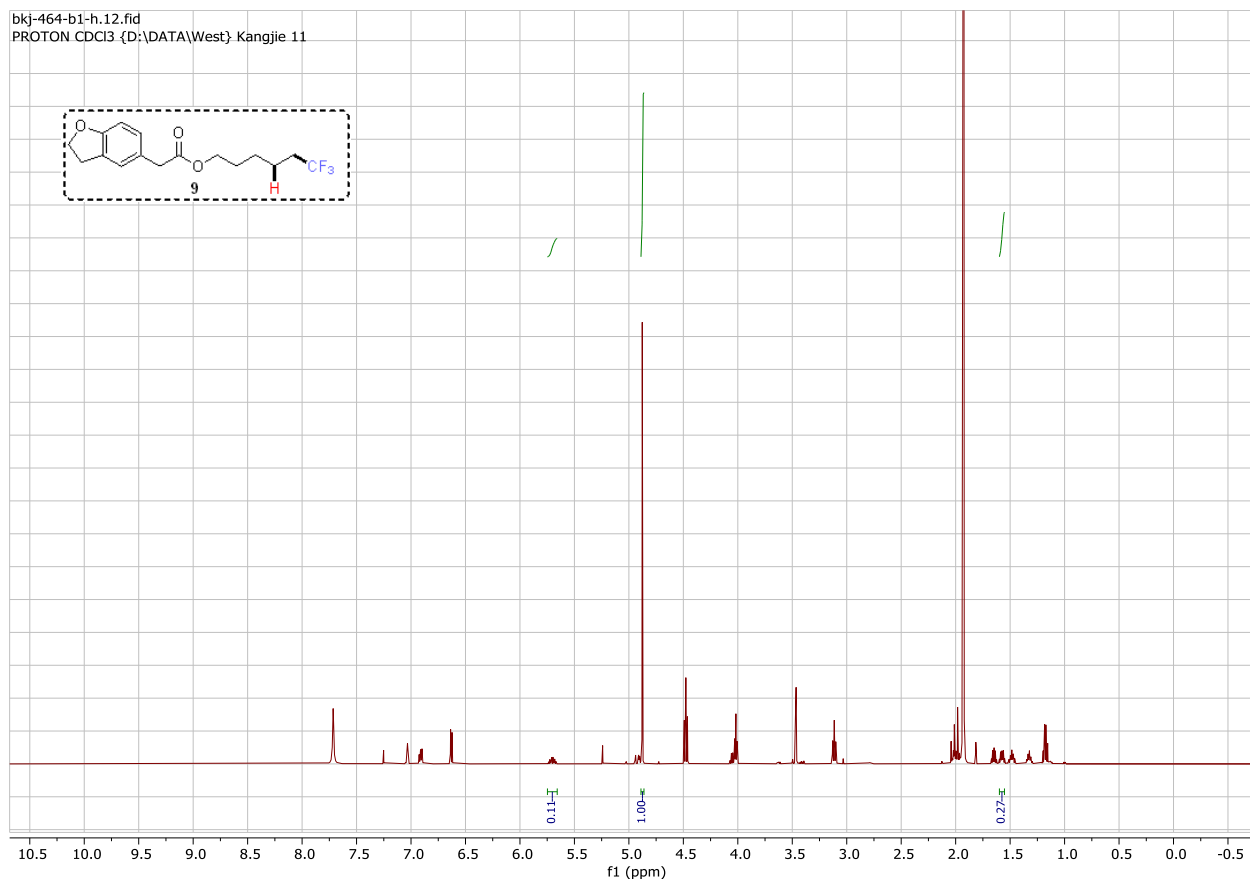


Similar to Fe(III) counterpart and indicated by ^{19}F NMR, the homolysis of trifluoroacetic acid is not observed by the combination of Fe(II) ($\text{Fe}(\text{OAc})_2$) and TRIP thiol in the absence of light irradiation, confirming light is necessary in inducing homolysis of trifluoroacetic acid to afford CF_3 radical.

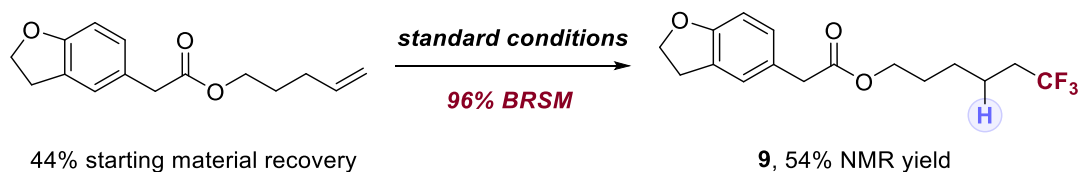
2.12 Mass balance studies of hydrofluoroalkylation of representative examples

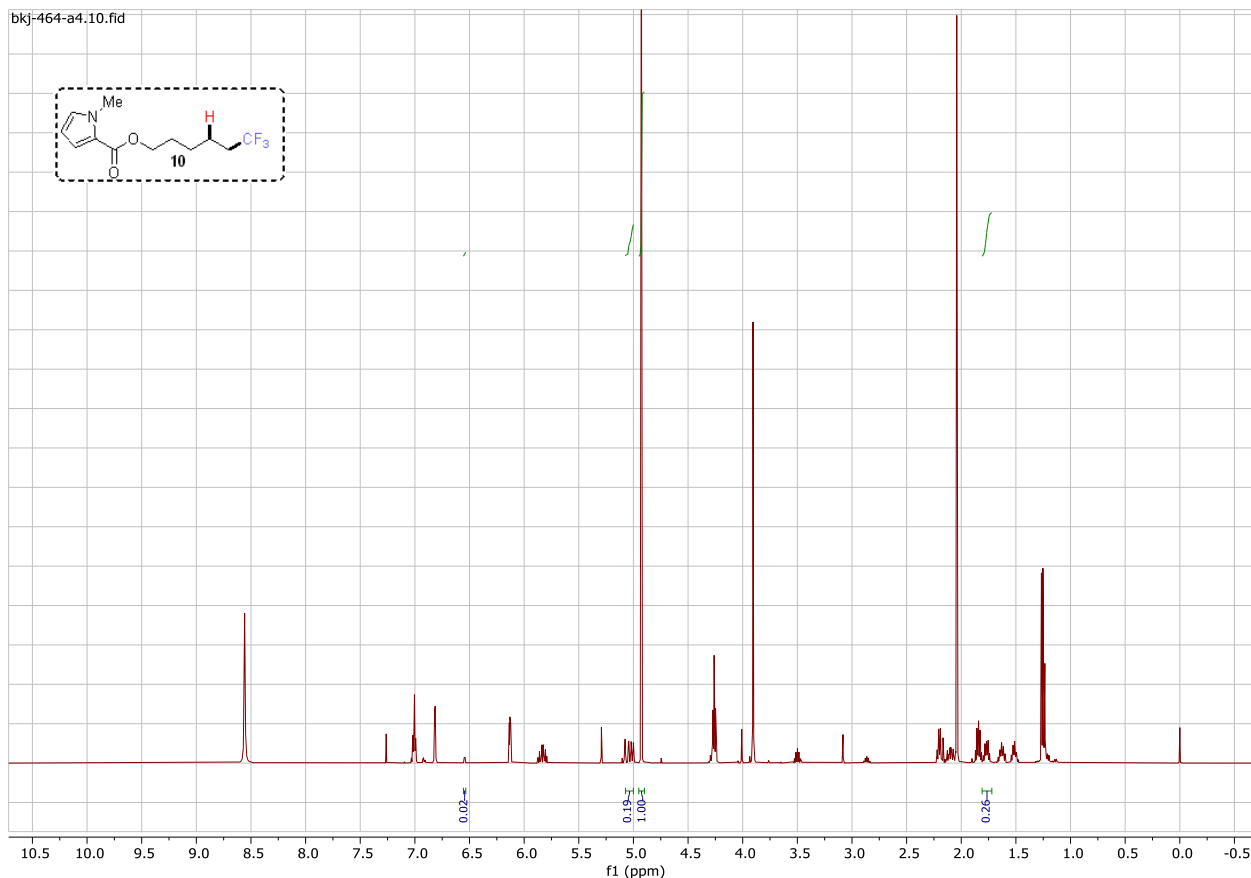
Mass balance studies of heterocycle-containing substrates

Trifluoromethylation Examples



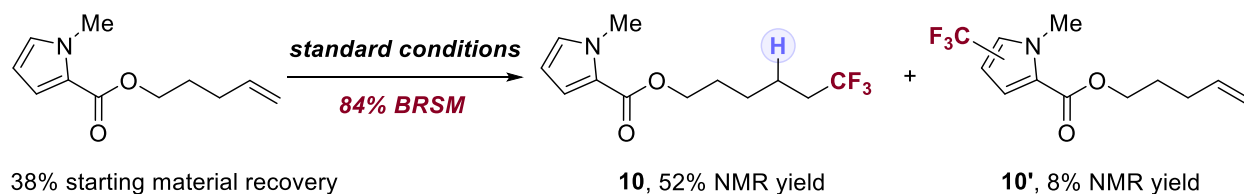
As indicated in the crude ¹H NMR, **9** is produced in 54% NMR yield at 1.60-1.55 (m, 2H) with 44% residual starting material rat (5.75-5.66, m, 1H) (in regard to 0.1 mmol CH₂Br₂ as internal standard). The mass-balance of this reaction is 98%.



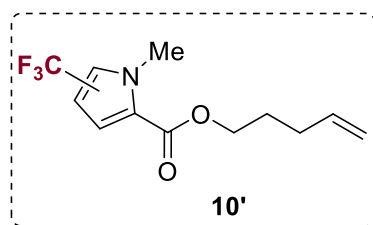


As indicated in the crude ^1H NMR, **10** is observed in 52% NMR yield at 1.81-1.71 (m, 2H) with 38% residual starting material recovery at (5.07-5.00, m, 1H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). The mass-balance is 90% from these two species.

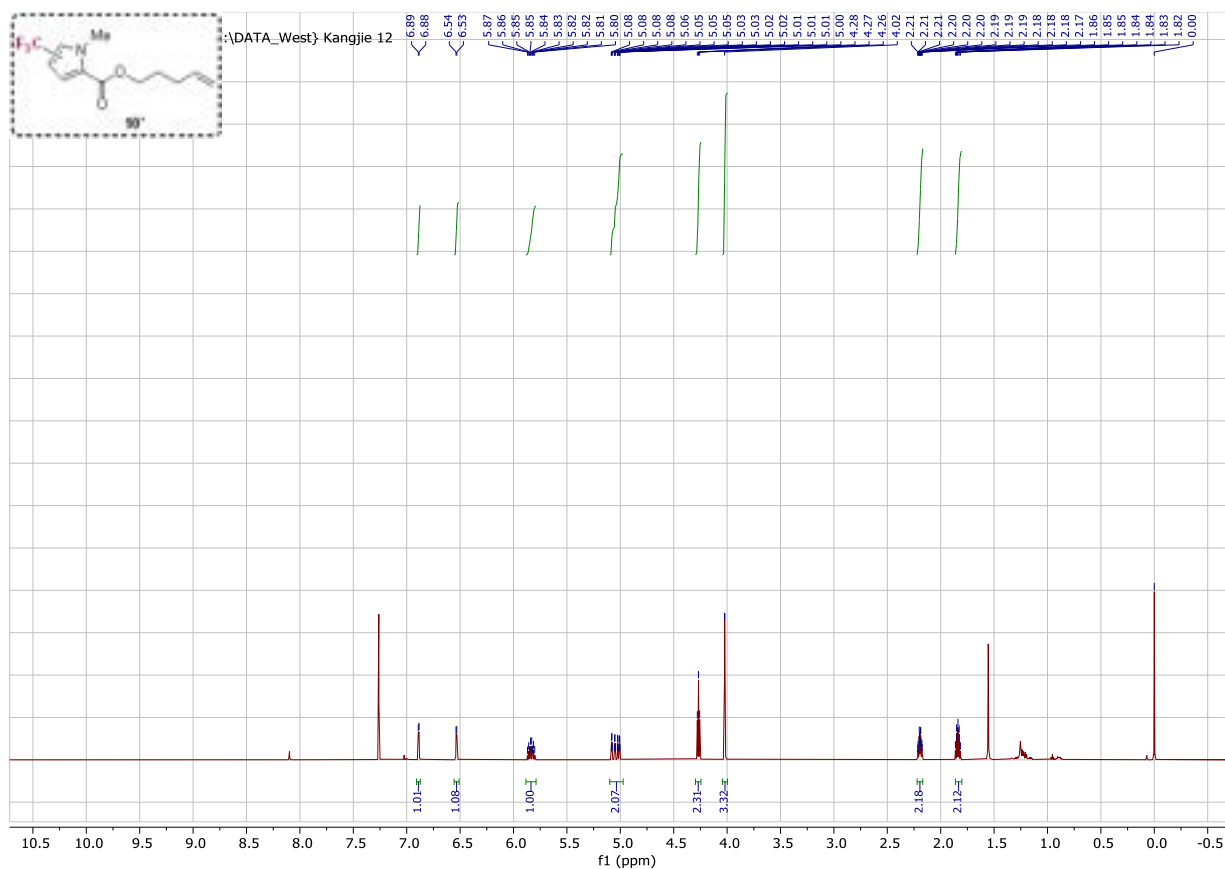
The reduced mass balance from can be explained by slightly competitive trifluoromethylation of the pyrrole ring (diagnostic peak at 6.56-6.53 (m, 1H)) to form trifluoromethylated starting material **10'** (single isomer), accounting for 8% NMR yield. This species has been isolated and characterization is provided below. With this, the overall mass balance is 98%.

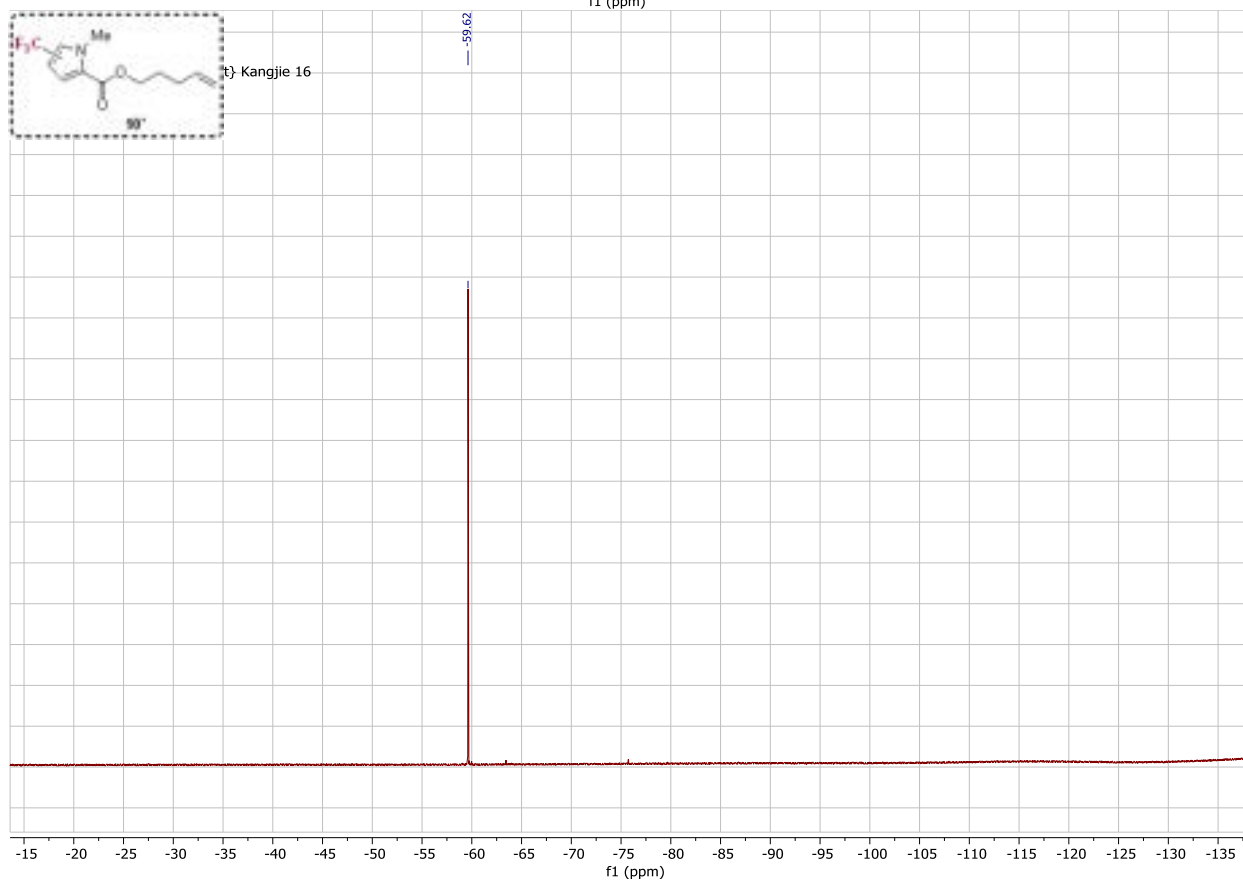
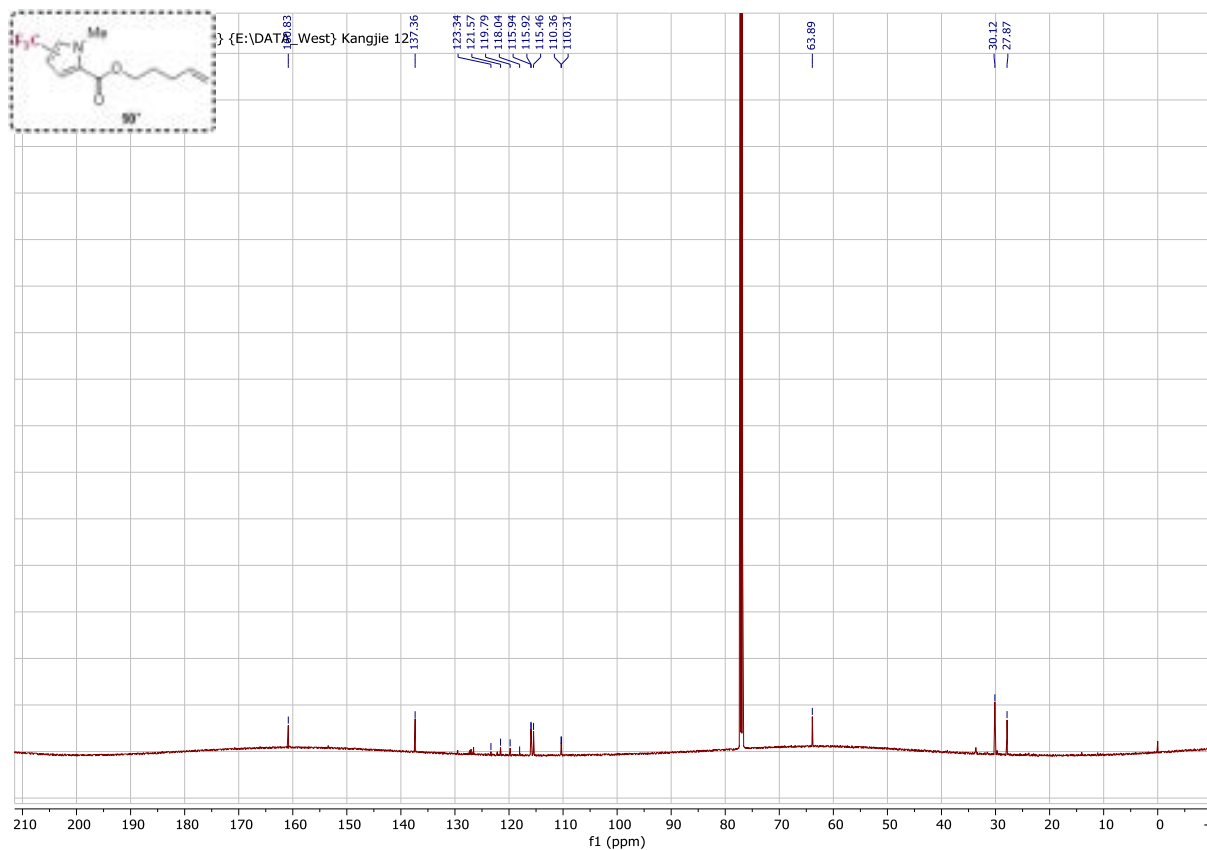


The characterization data of **10'** is shown below,

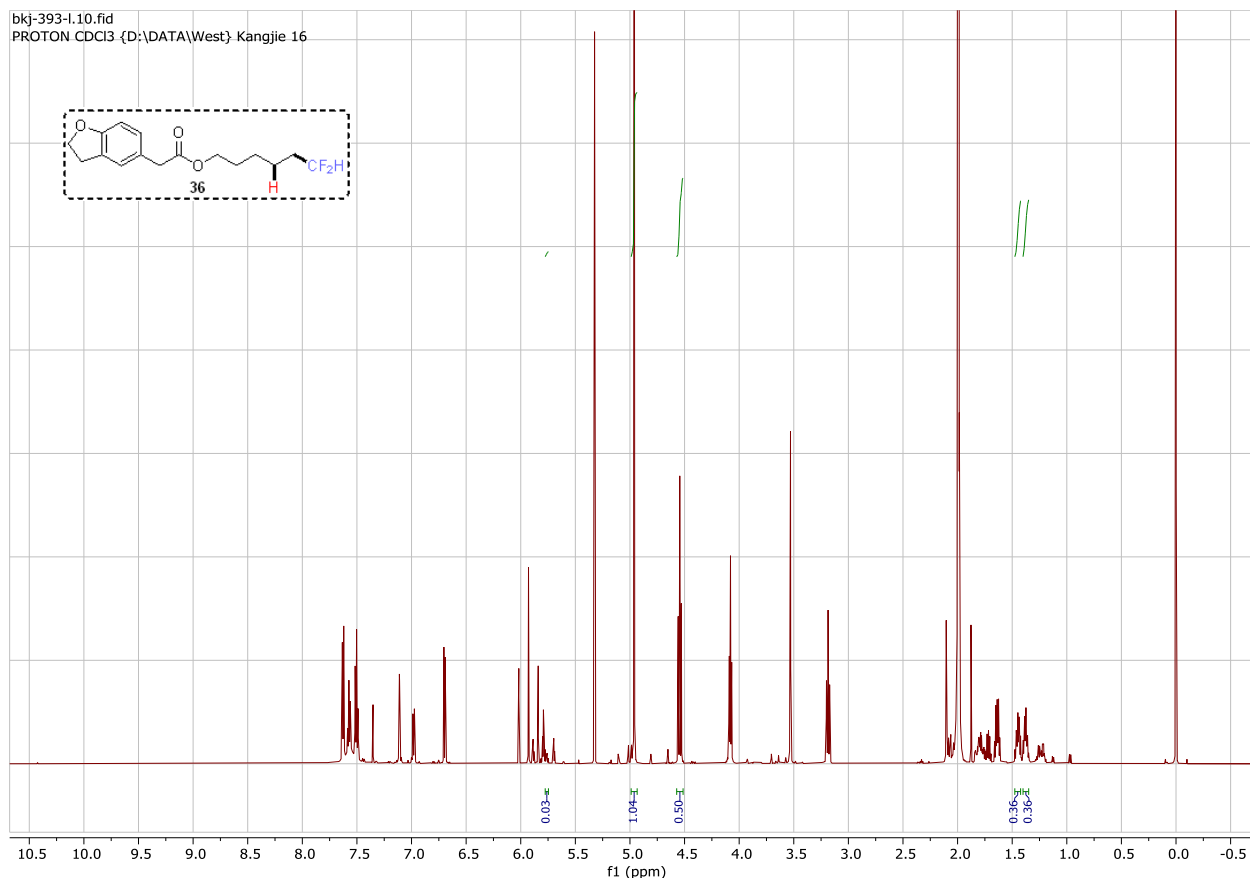


¹H NMR (600 MHz, Chloroform-*d*) δ 6.89 (d, *J* = 4.1 Hz, 1H), 6.53 (d, *J* = 4.1 Hz, 1H), 5.89-5.79 (m, 1H), 5.10 – 4.97 (m, 2H), 4.27 (t, *J* = 6.6 Hz, 2H), 4.02 (s, 3H), 2.22 – 2.17 (m, 2H), 1.86 – 1.80 (m, 2H). ¹³C NMR (151 MHz, CDCl₃, 3 mm tube) δ 160.83, 137.36, 120.68 (q, *J* = 267.8 Hz), 115.93 (q, *J* = 2.4 Hz), 115.46, 110.34 (q, *J* = 7.6 Hz), 63.89, 30.12, 27.87. ¹⁹F NMR (565 MHz, Chloroform-*d*) δ -59.62. HRMS APCI: [M+H]⁺ calcd. for C₁₂H₁₅F₃NO₂: 262.1049; Found 262.1054

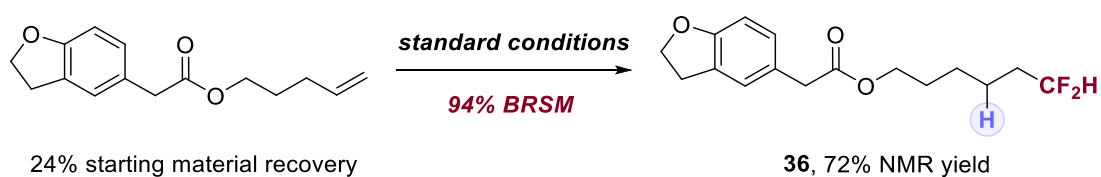


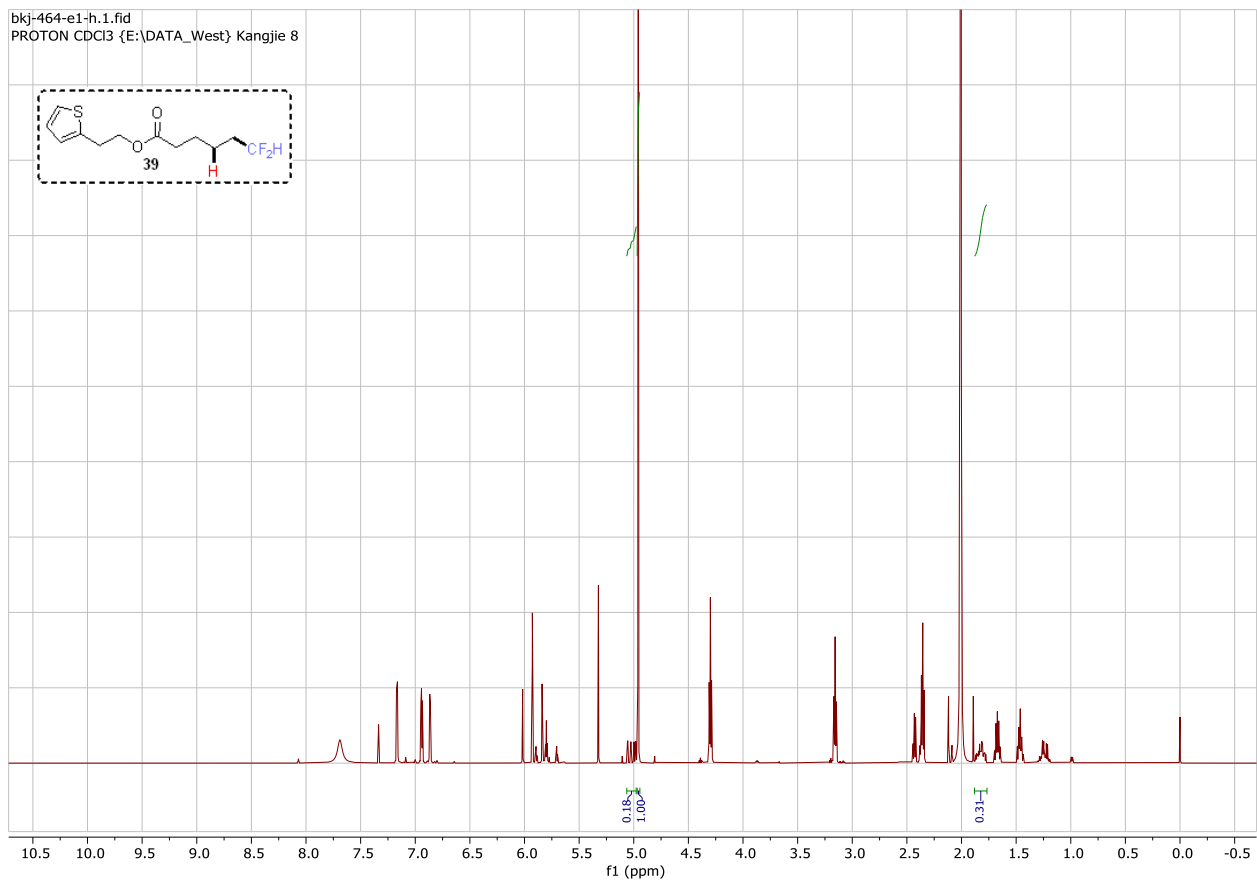


Difluoromethylation examples.

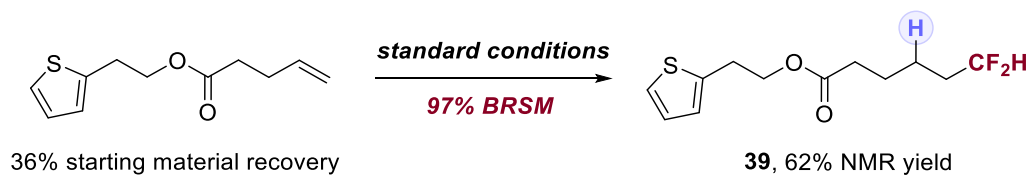


As indicated in the crude ^1H NMR, **36** is produced in 72% NMR yield (1.48-1.42, m, 2H) with 24% residual starting material at 5.81-5.74 (partially overlapped with product peak) (m, 1H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). The internal standard is slightly overlapped with substrate peak which shows integration of 1.04 at 4.9 ppm. No other significant products are observed. The mass balance is approximately 96%.



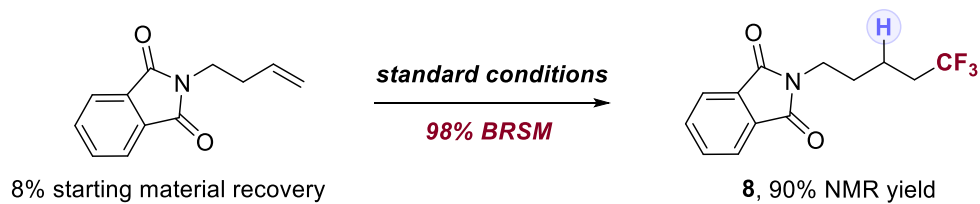
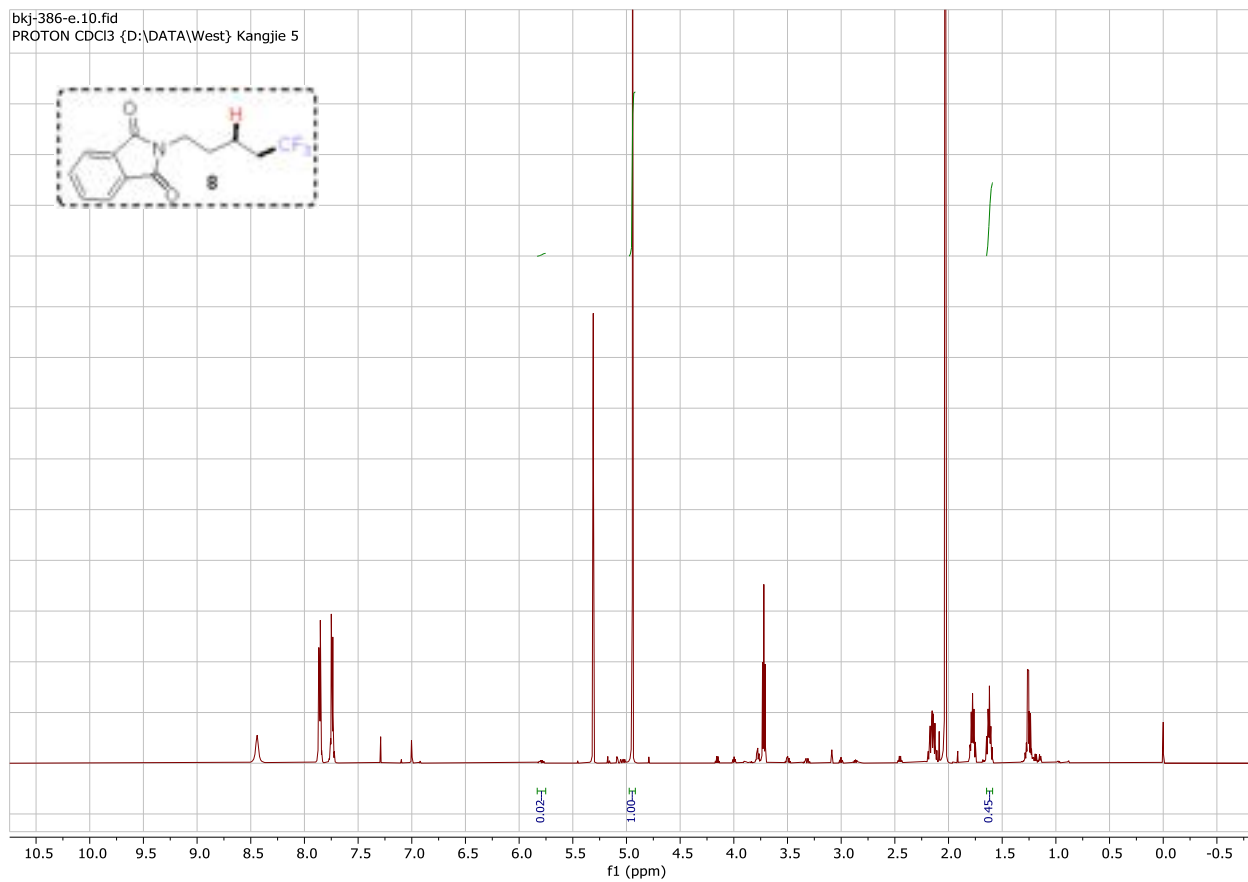


As indicated in the crude ^1H NMR, **39** is produced in 62% NMR yield (1.88-1.77, m, 2H) with 36% residual starting material at 5.07-4.98 (m, 1H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). No other significant products are observed. The mass-balance of the reaction is 98%.

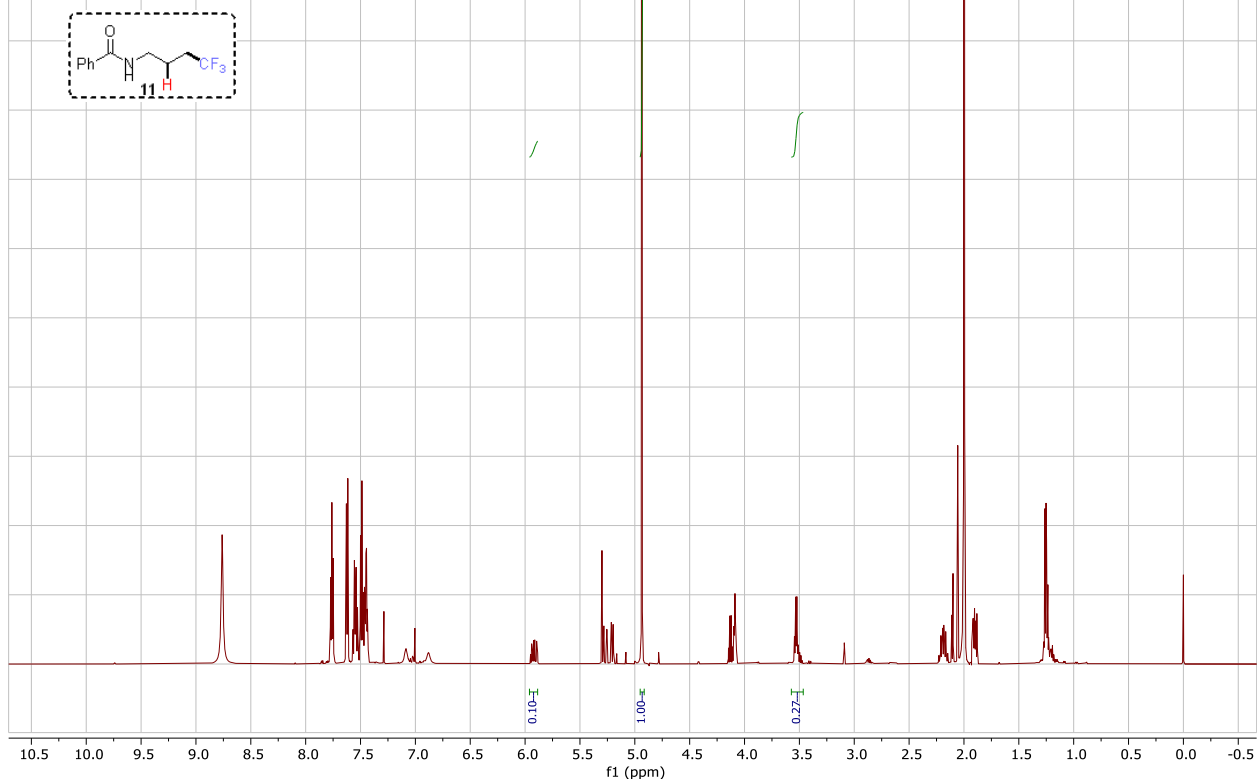


Mass balance studies of representative substrates

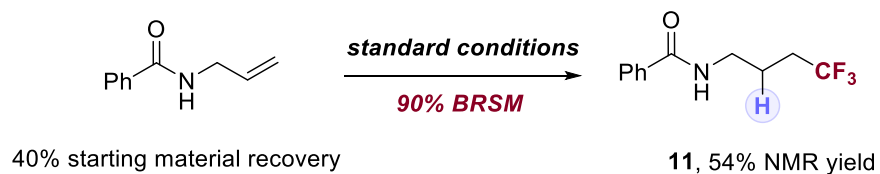
Hydrotrifluoromethylation of representative examples



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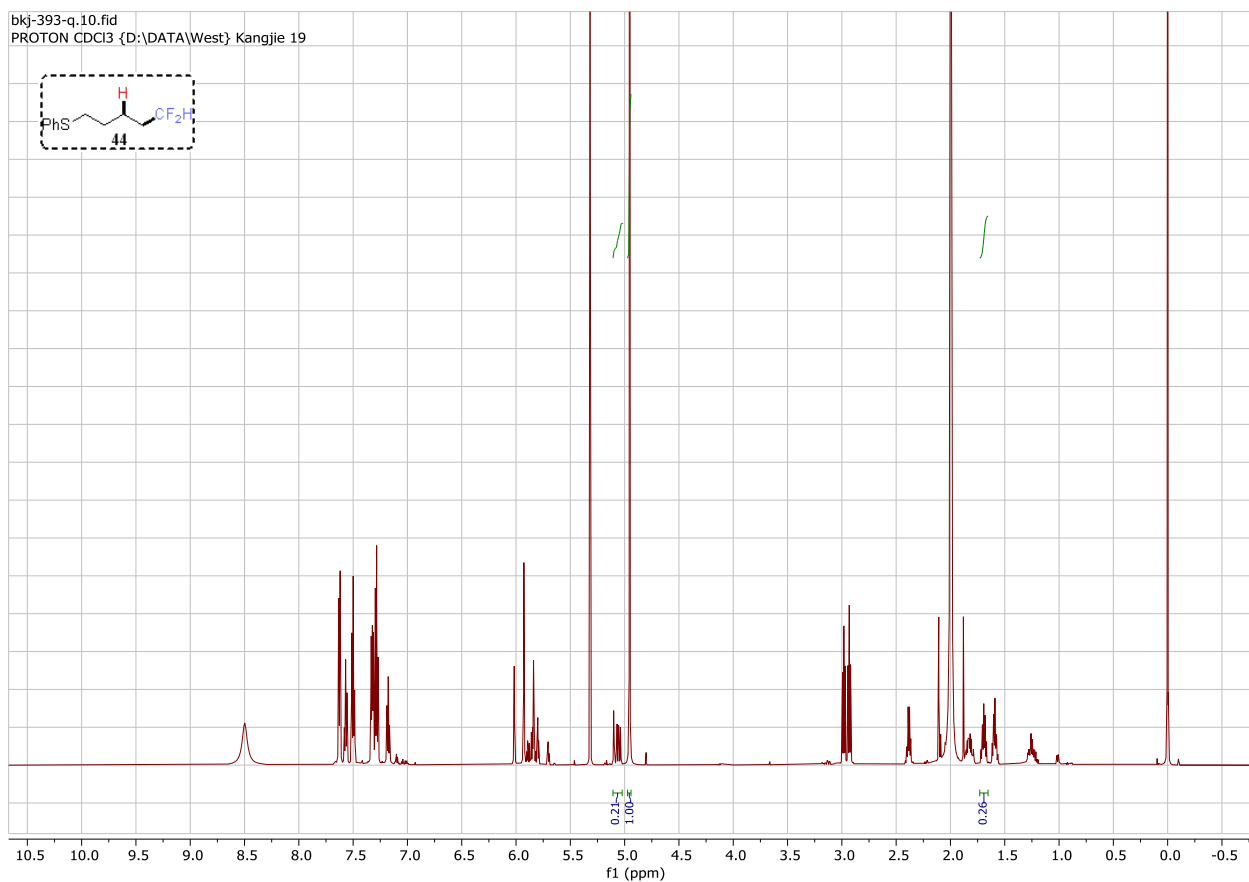


As indicated in the crude ^1H NMR, **11** is produced in 54% NMR yield at 3.59-3.45 (m, 2H) with 40% residual starting material (5.97-5.87, m, 1H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). No other significant products are observed. The mass-balance is 94%.

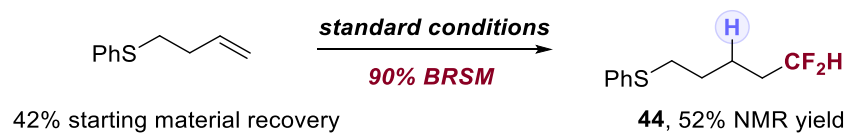


Hydrodifluoromethylation of representative examples

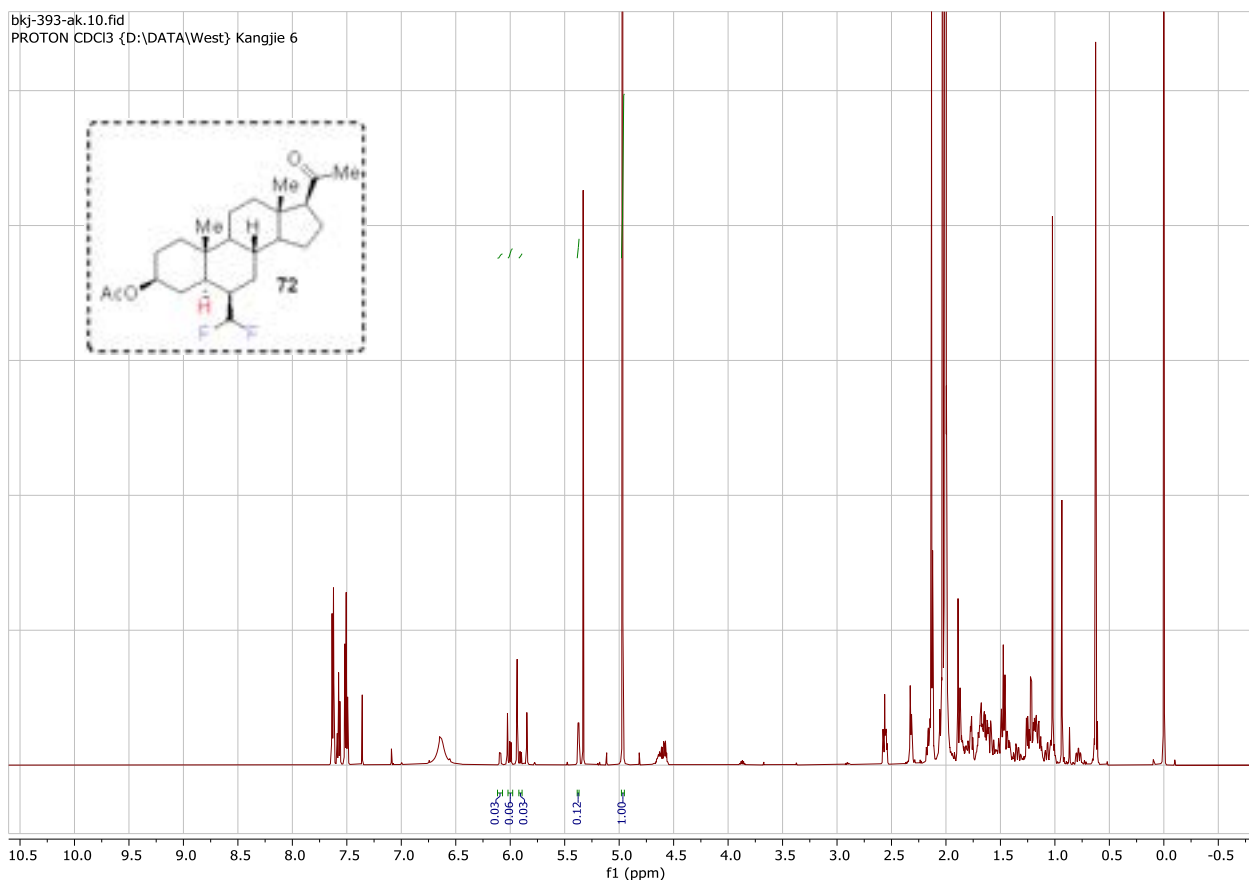
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PROTON CDCl3 {D:\DATA\West\ Kangjie 19



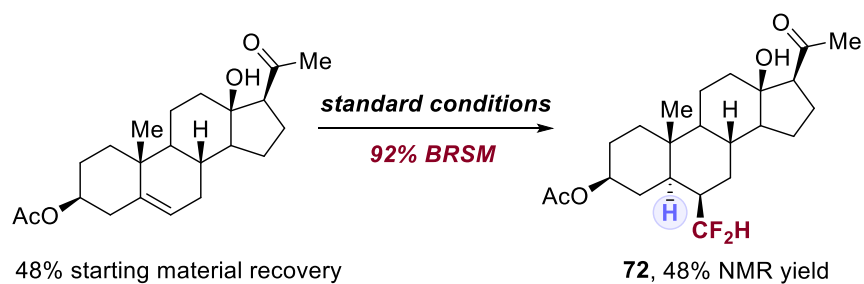
As indicated in the crude ^1H NMR, **44** is produced in 52% NMR yield at 1.73-1.65 (m, 2H) with 42% residual starting material at (5.11-5.803, m, 2H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). No other significant products are observed. The mass-balance is 94%.



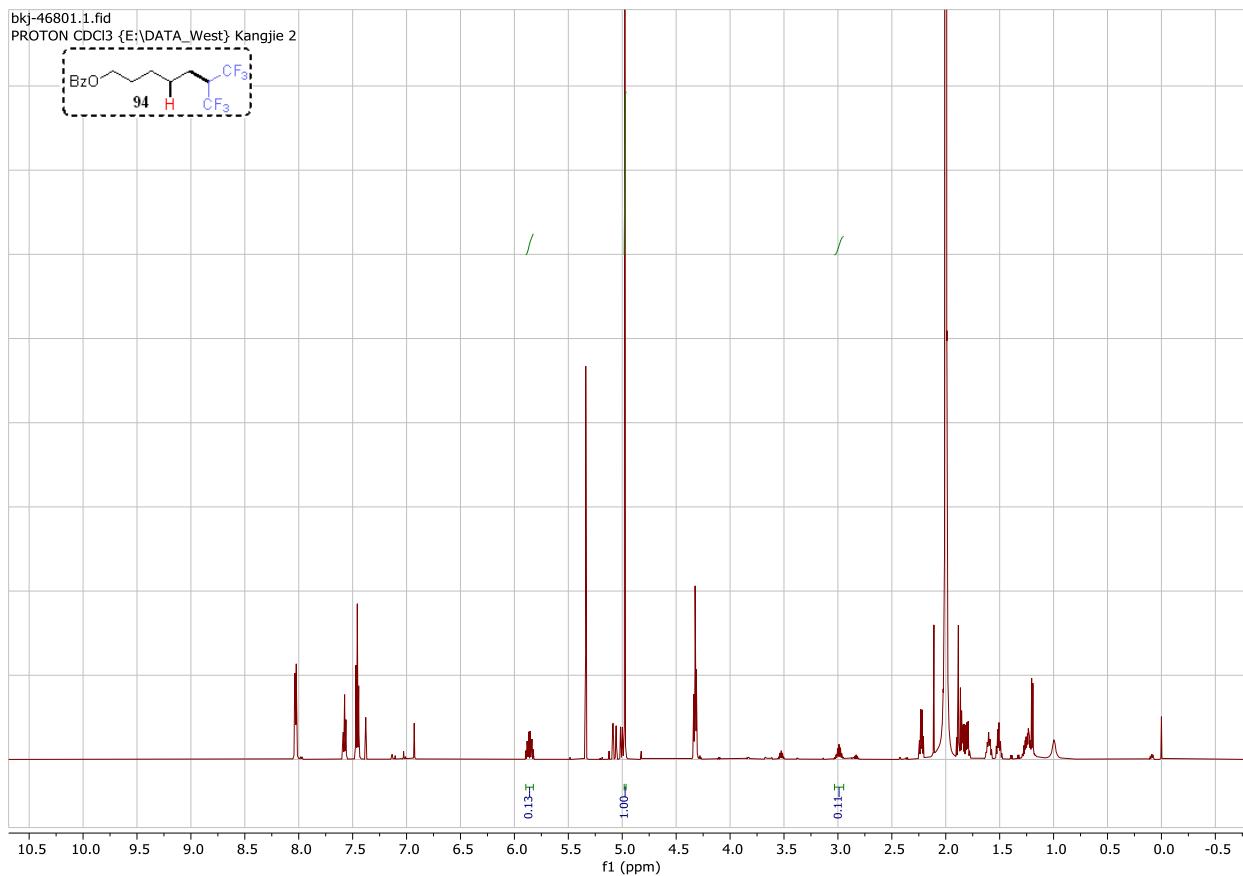
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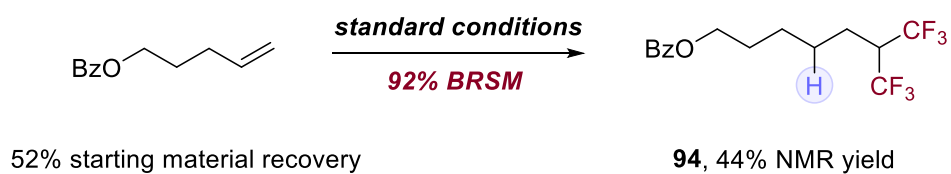
As indicated in the crude ^1H NMR, **72** is produced in 48% NMR yield at 1.73-1.65 (m, 2H) with 48% residual starting material at 5.39-5.36 (m, 1H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). No other significant products are observed. The mass-balance is 96%.



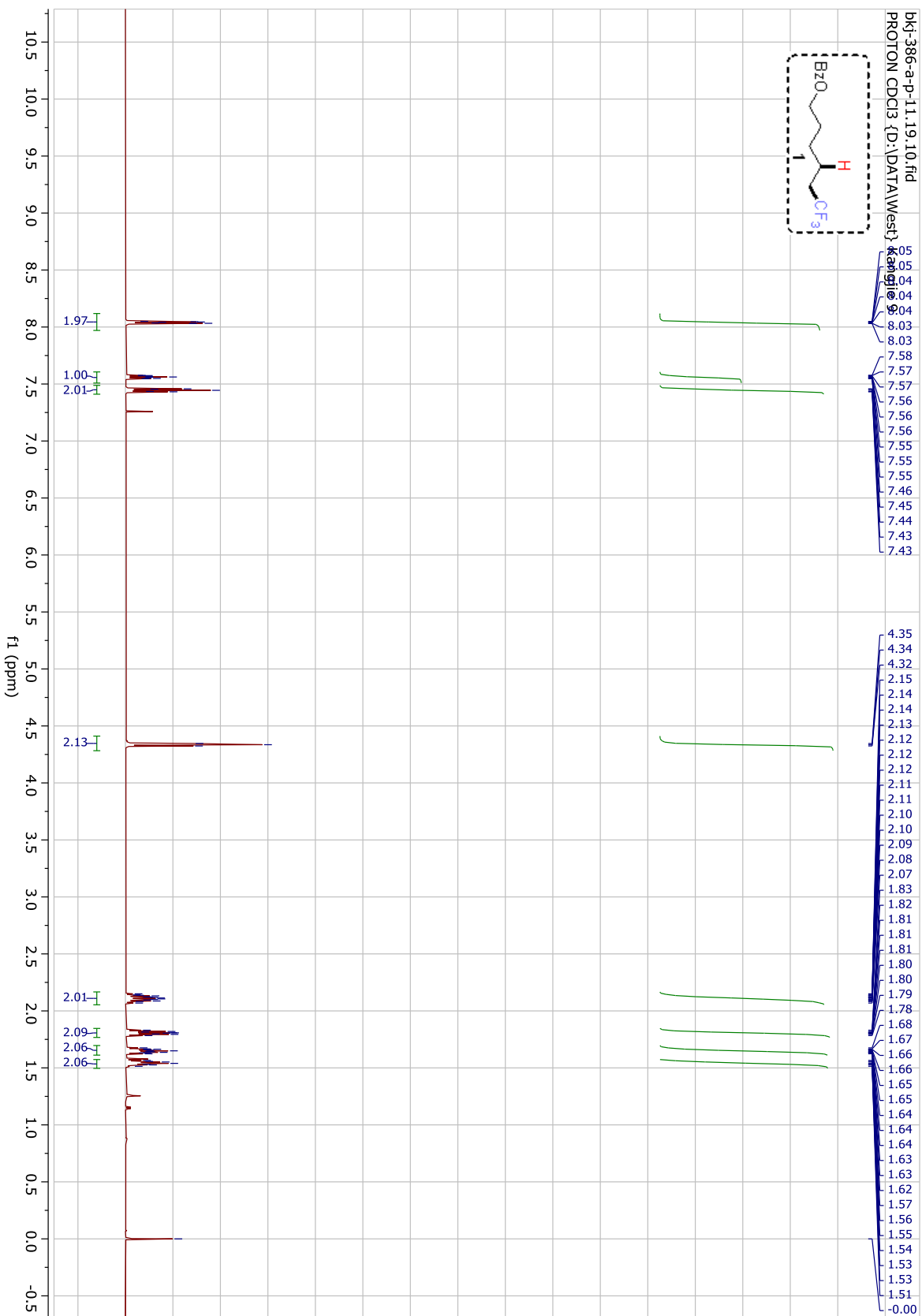
Hydrofluoroalkylation of a representative substrate



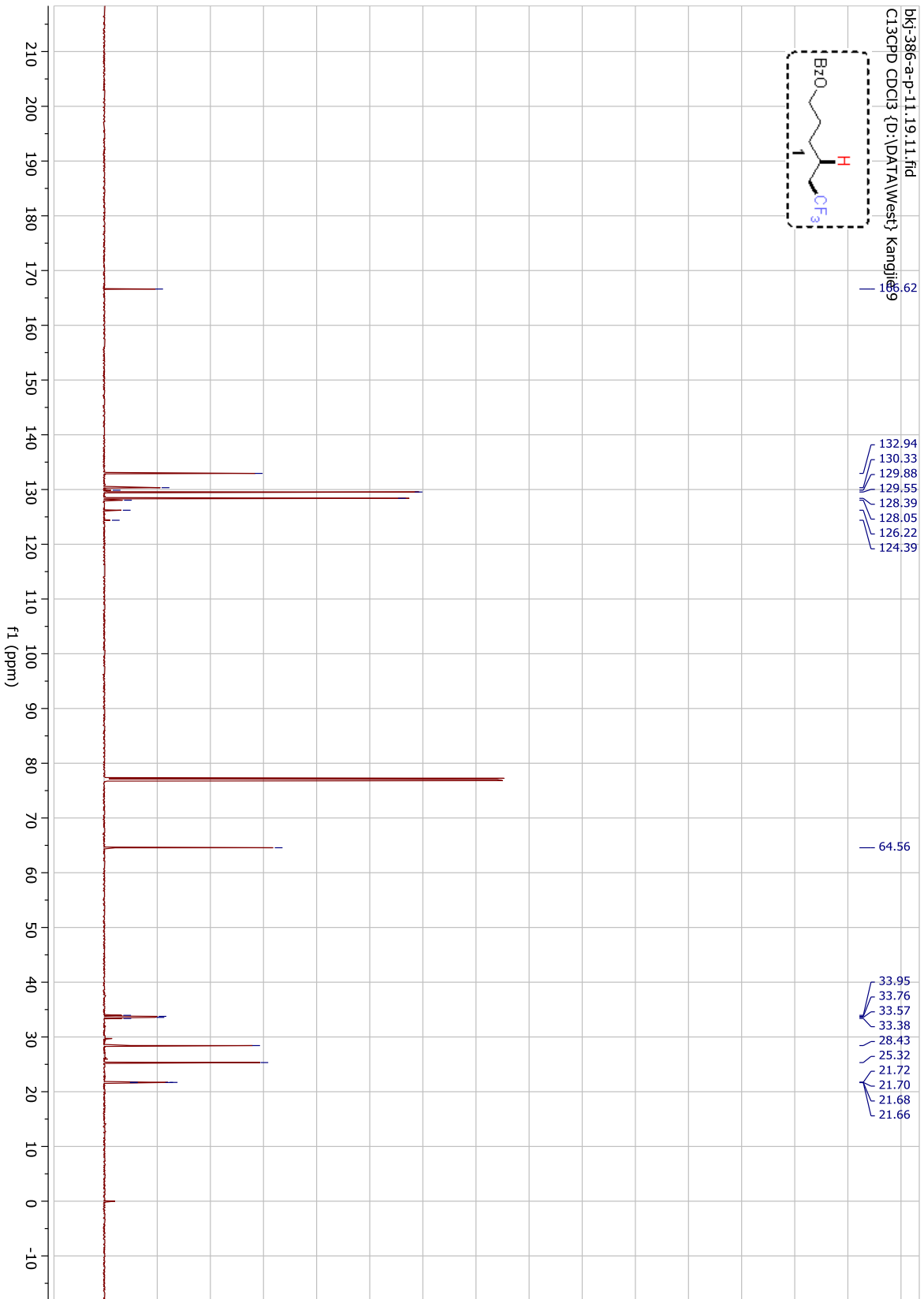
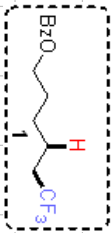
As indicated in the crude ^1H NMR, **94** is produced in 44% NMR yield at 3.03-2.95 (m, 1H) with 52% residual starting material at 5.90-5.82 (m, 1H) (in regard to 0.1 mmol CH_2Br_2 as internal standard). No other significant products are observed. The mass-balance is 96%.



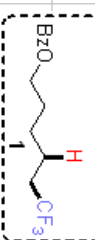
III. Supplemental Figure



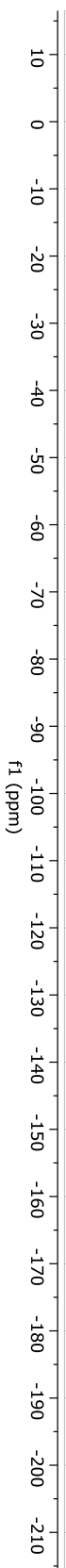
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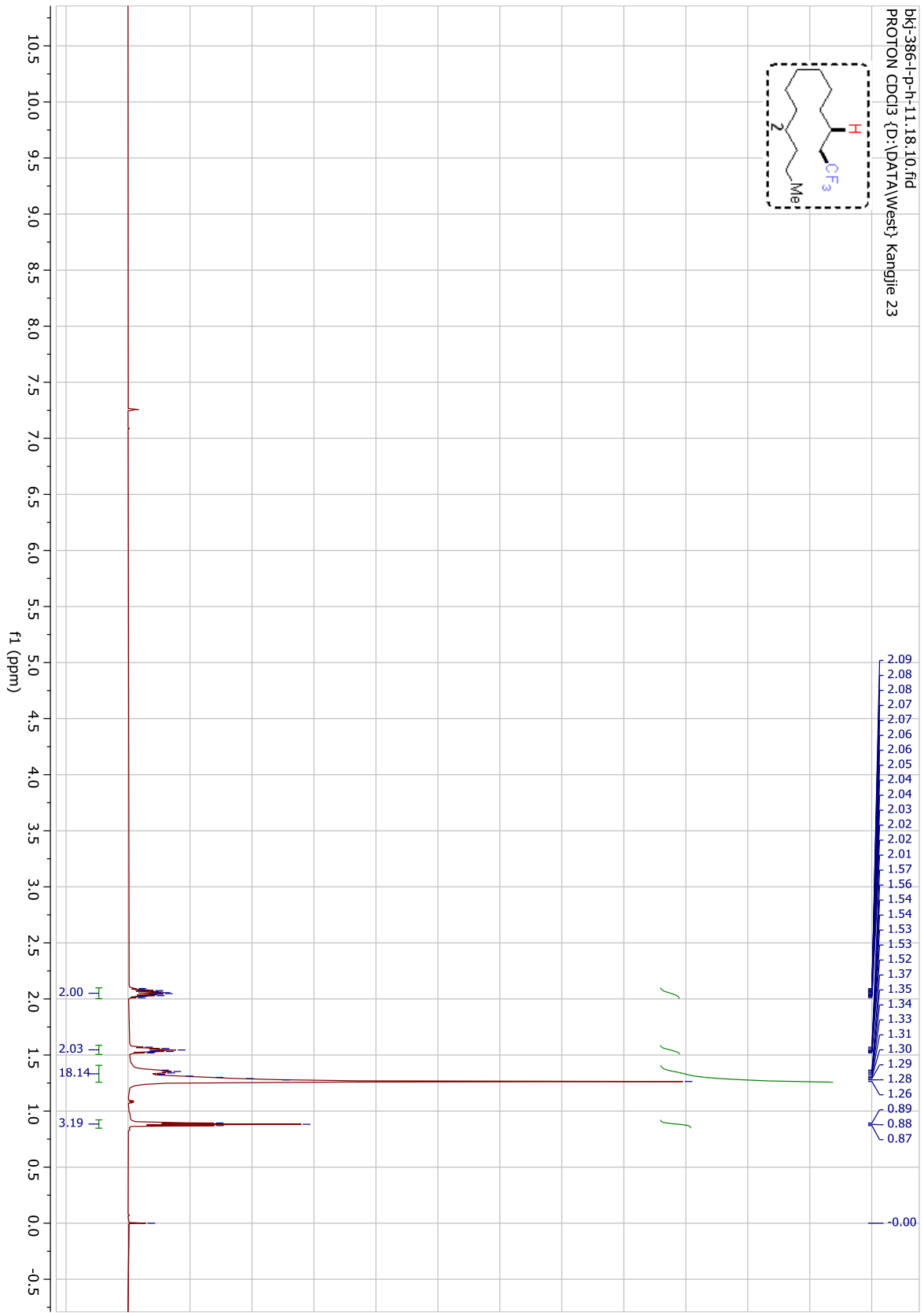
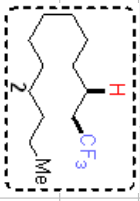
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F19_baseline_correct F1919_19\DATA\West\ Kangjie 9



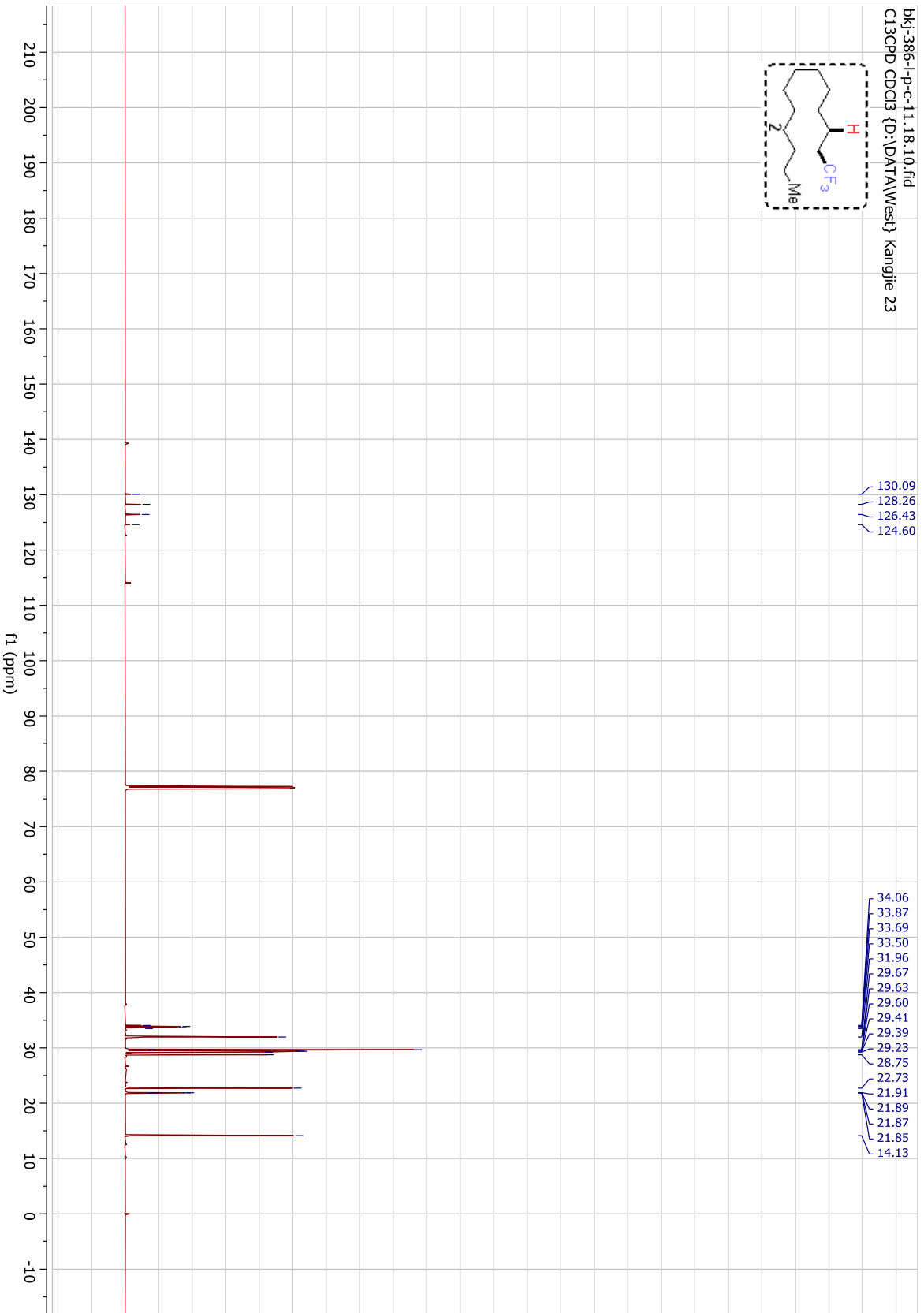
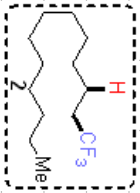
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66.38
66.40



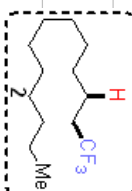
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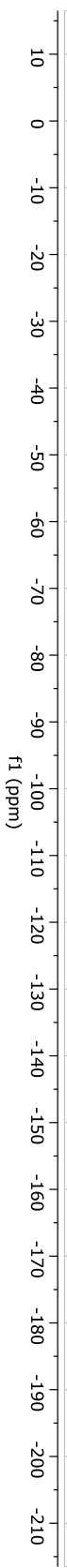
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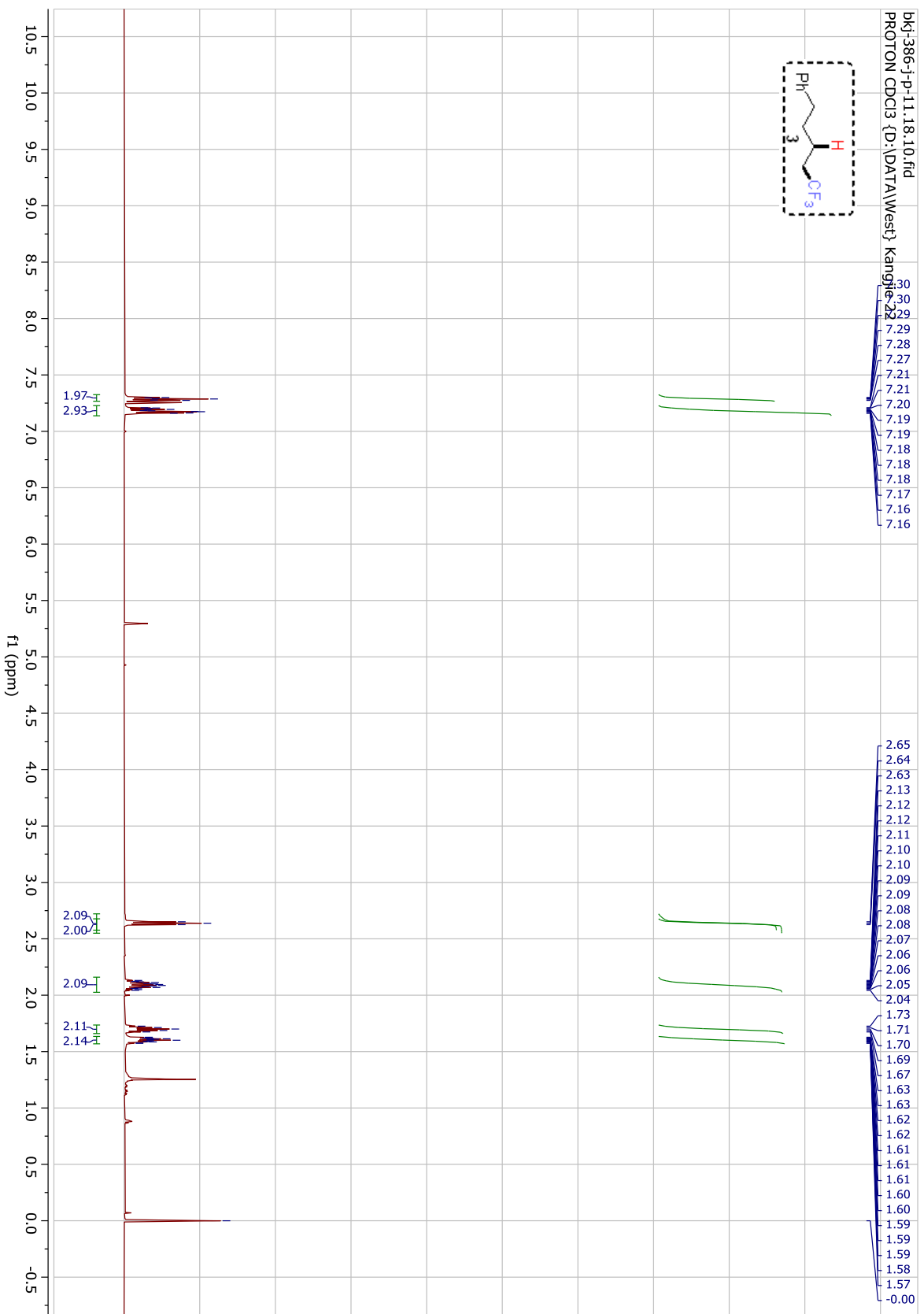


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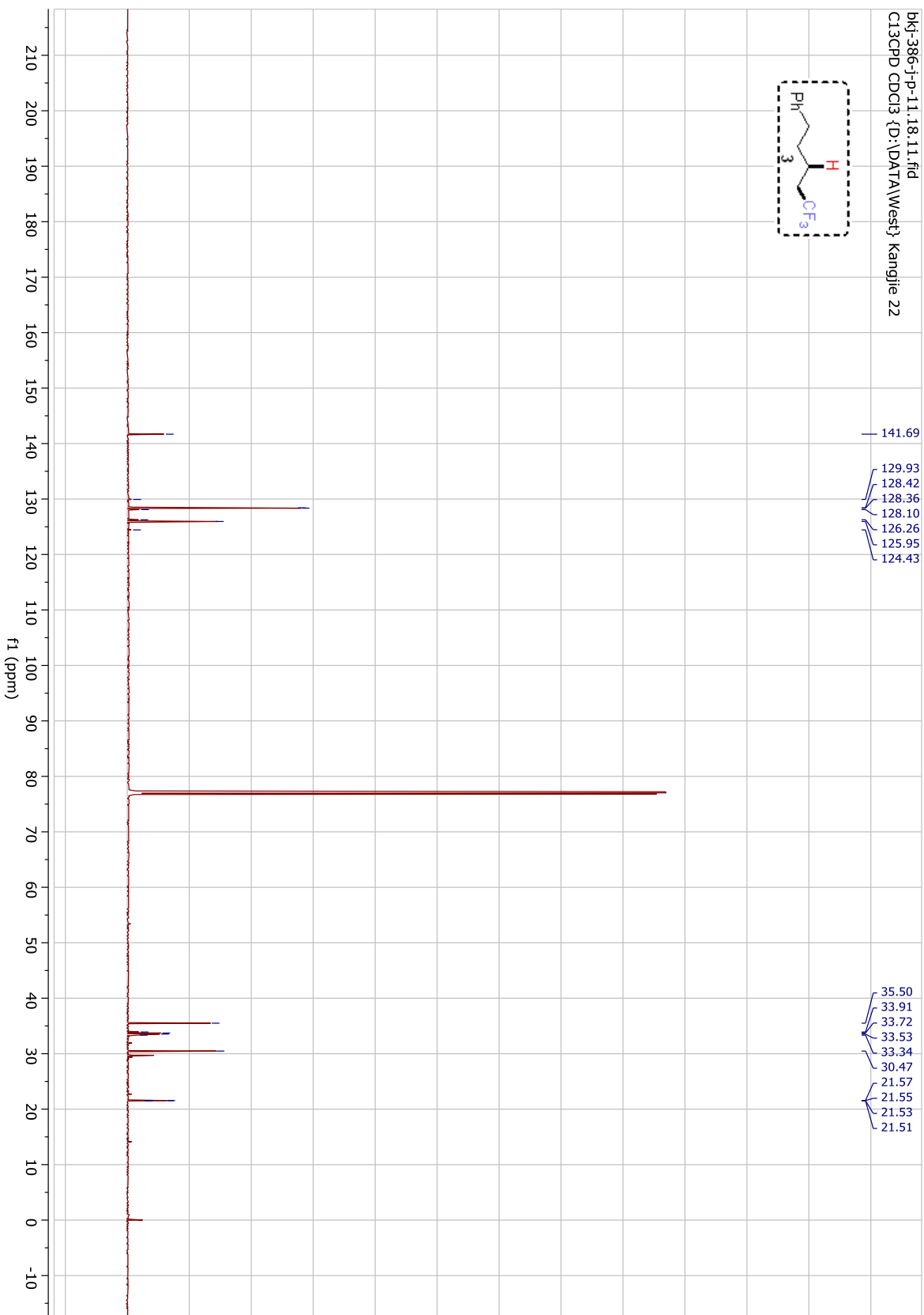
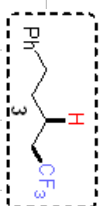


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-66.50



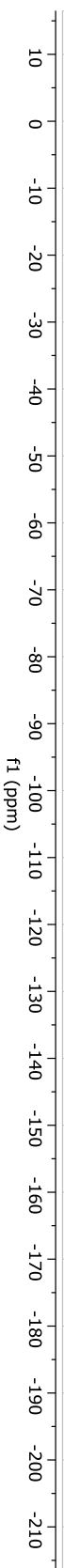
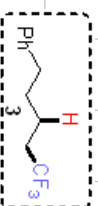


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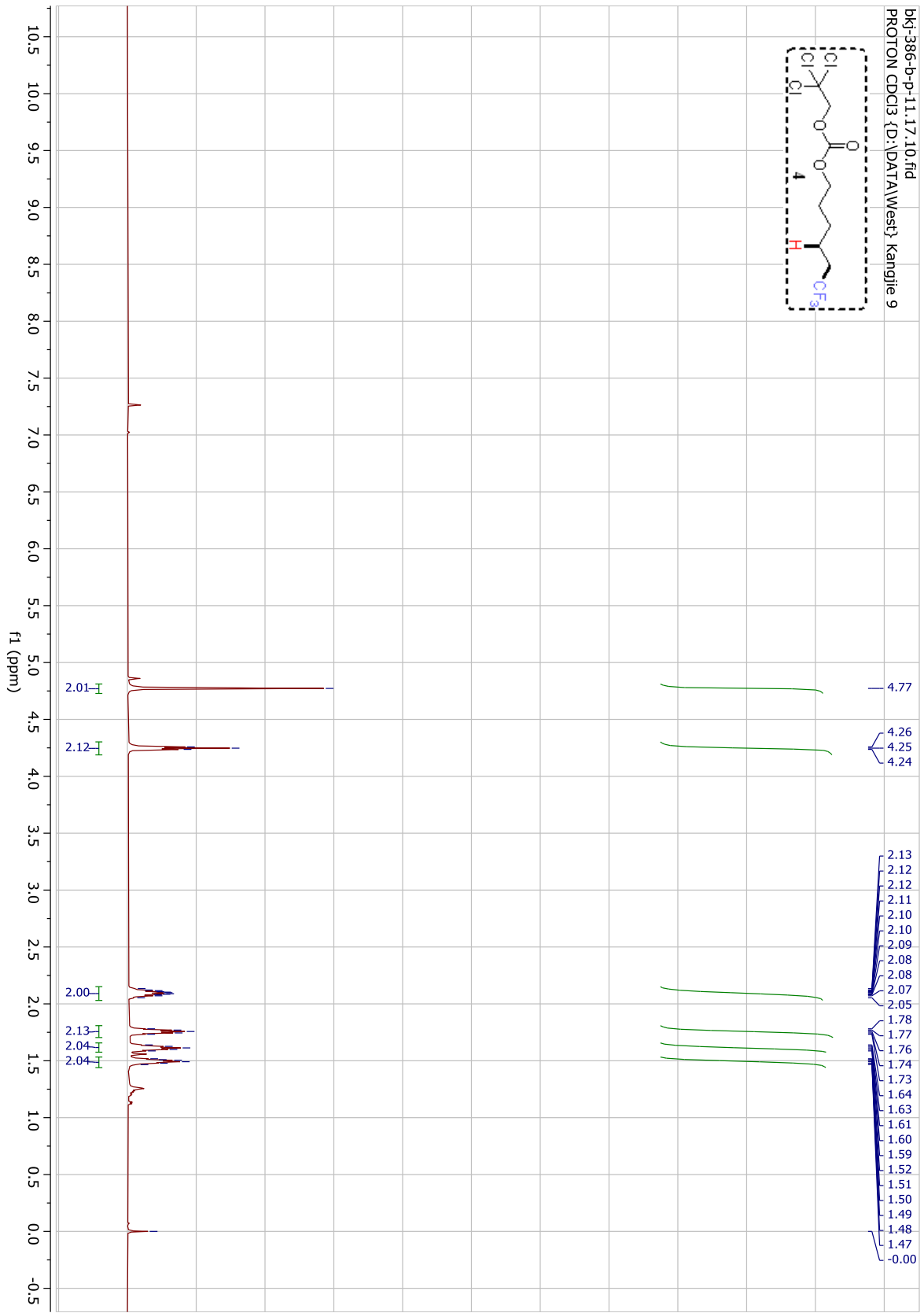
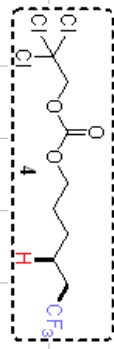


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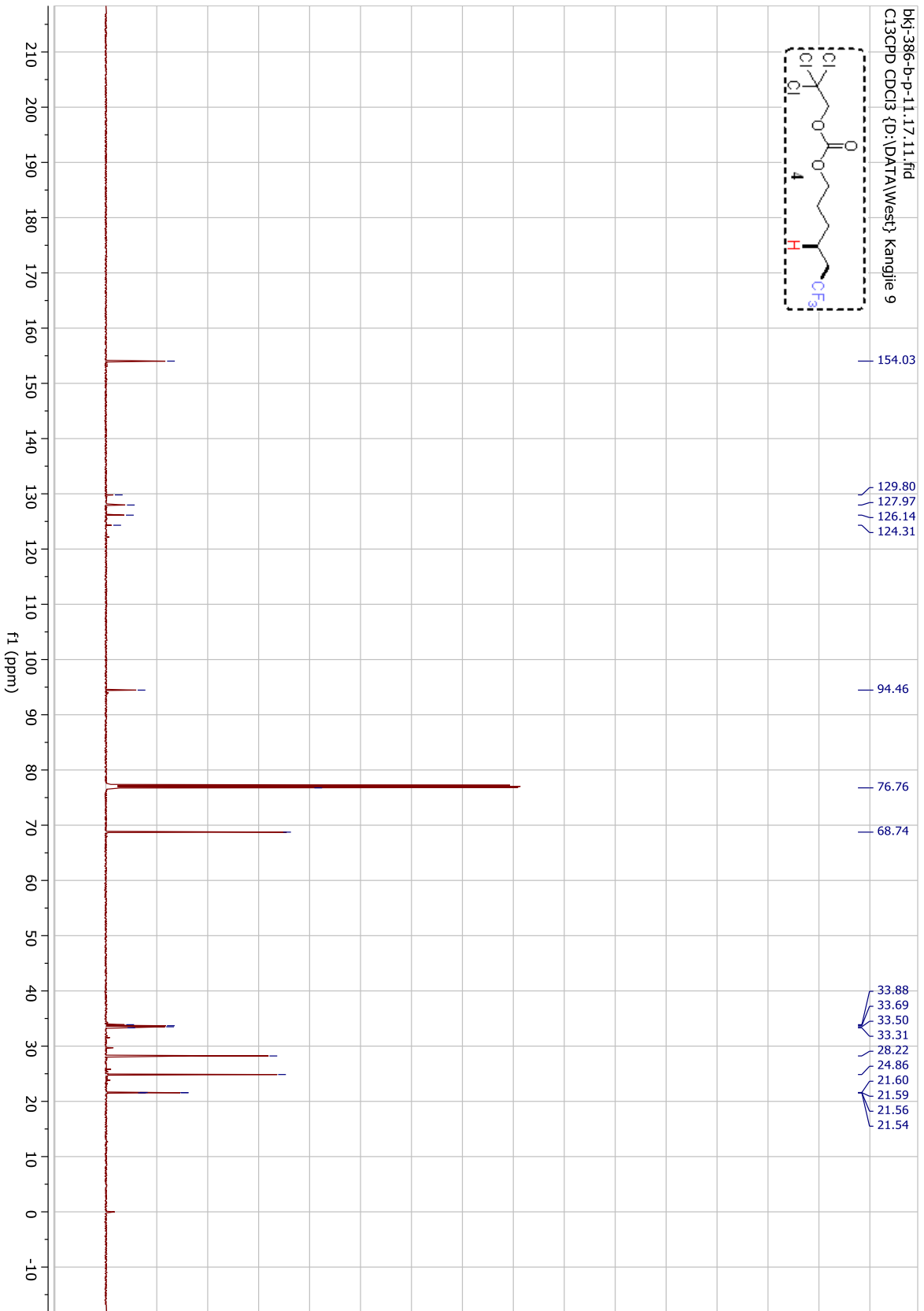
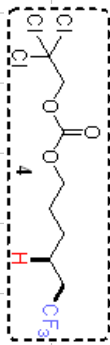
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96.34



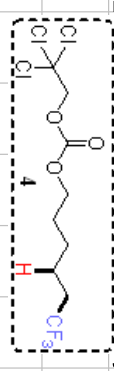
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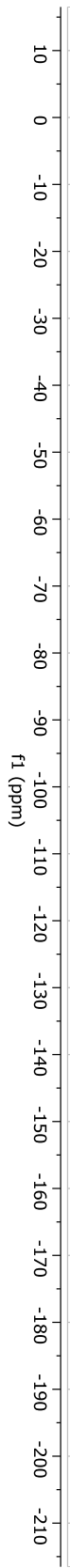
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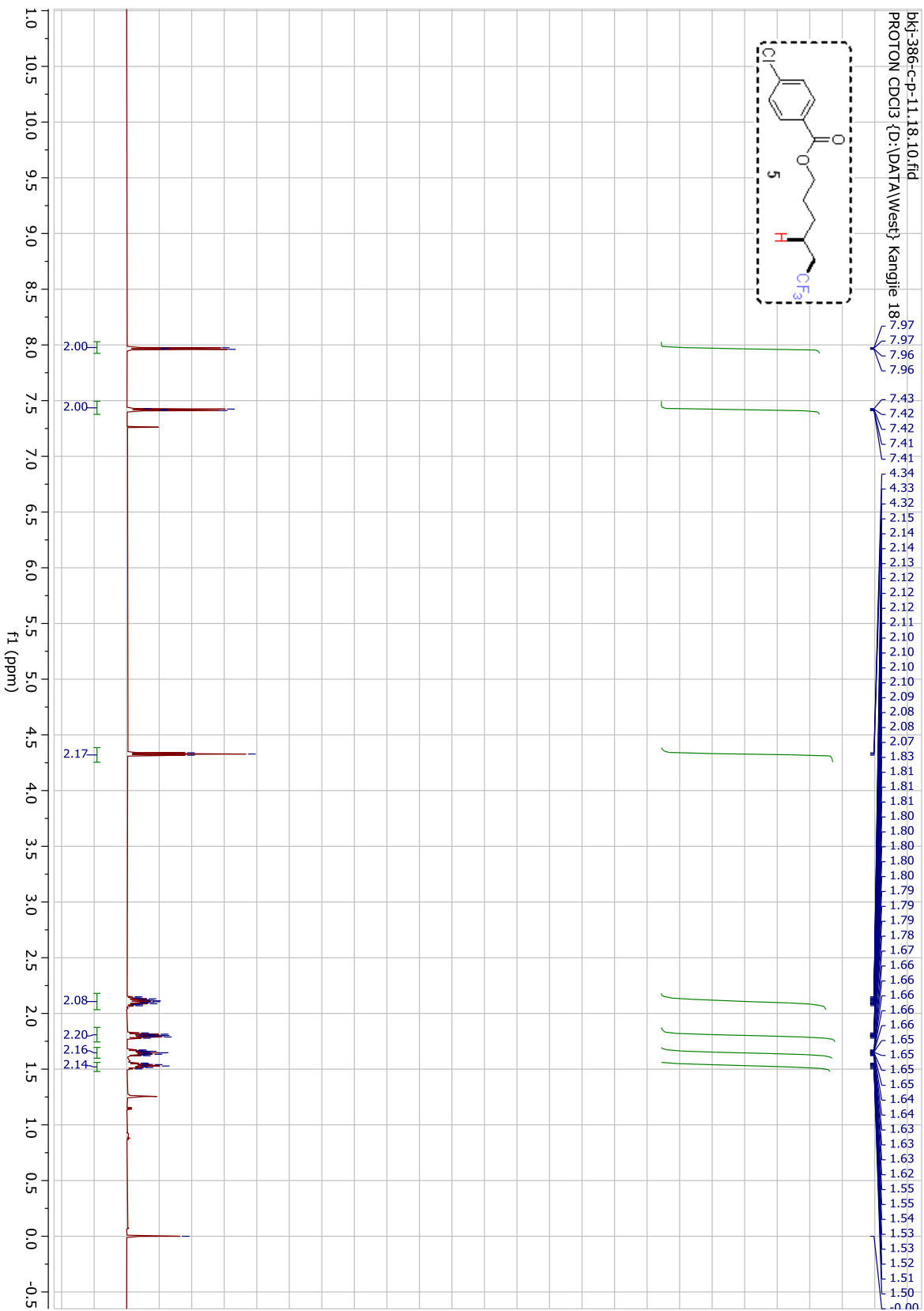


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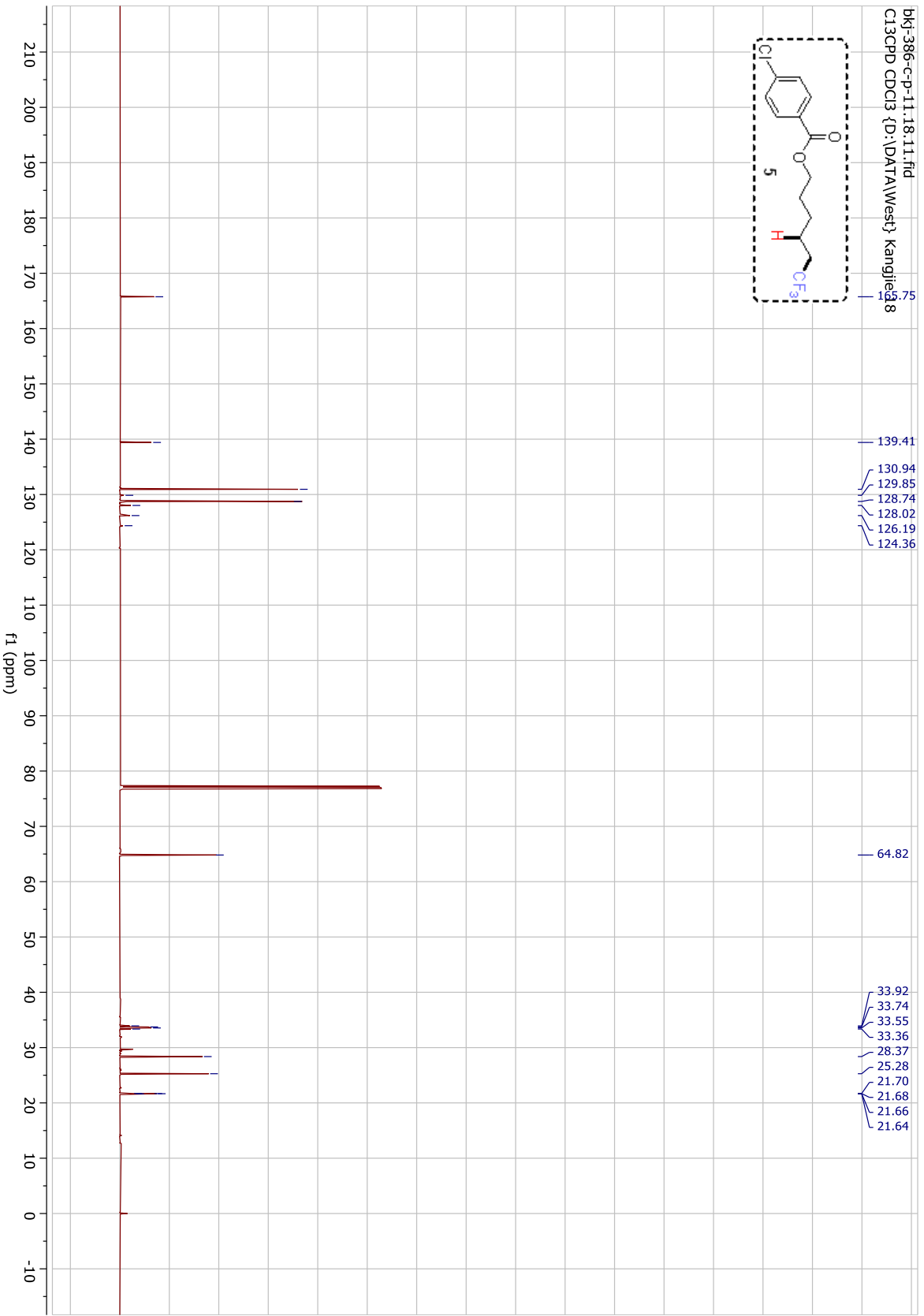
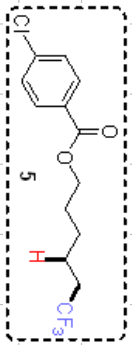


-66.37
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-66.41

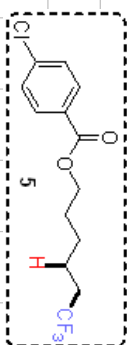




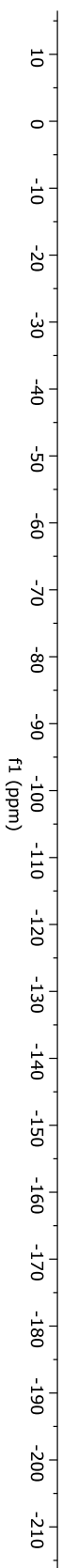
bkj-386-c-p-11:18:11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie\58

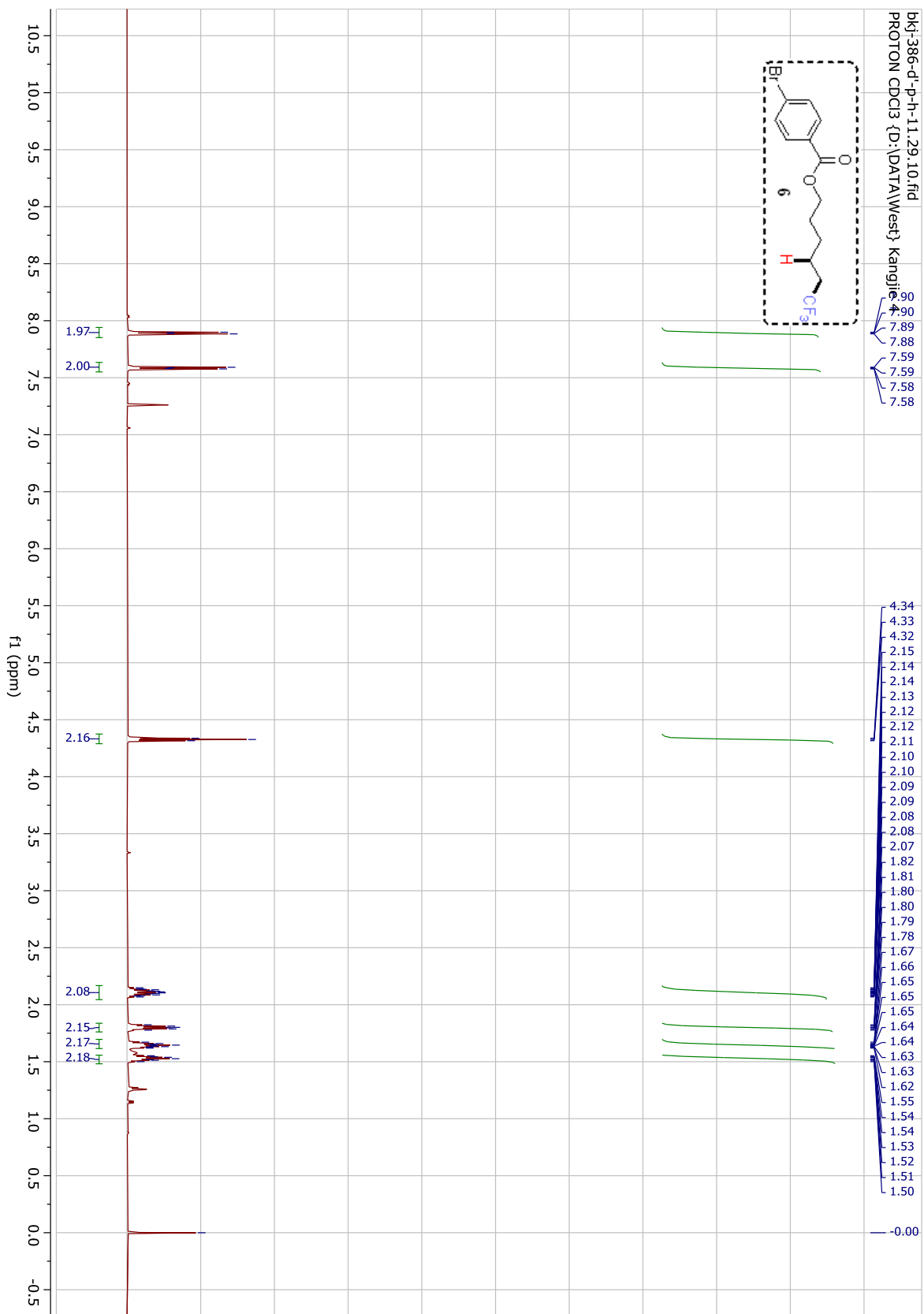


bkj-386-c-p-11.18.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 18



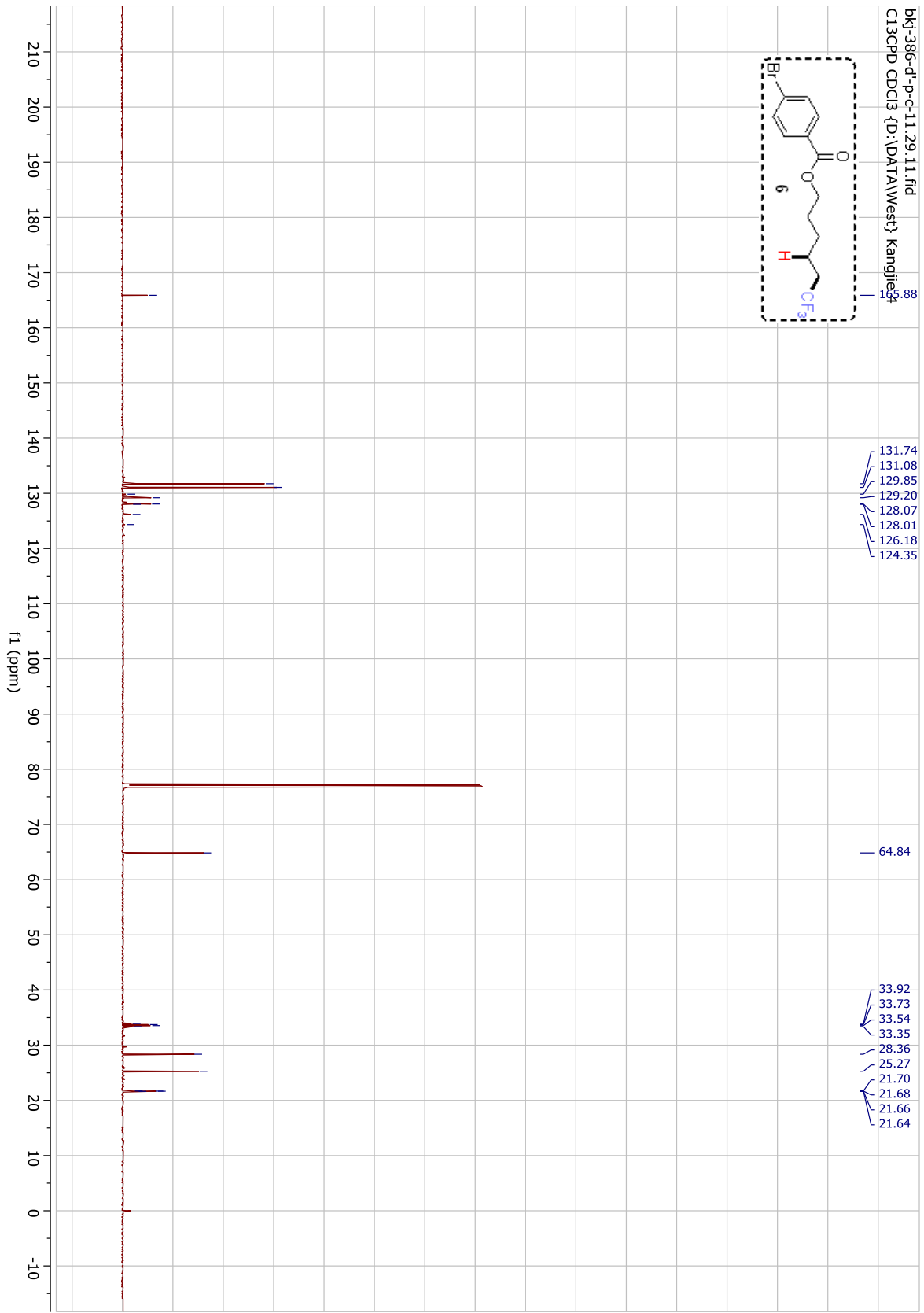
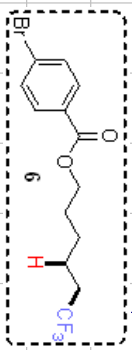
-66.34
-66.36
-66.38



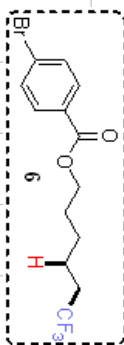


bkj-386-d'-p-c-11-29-11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjief)

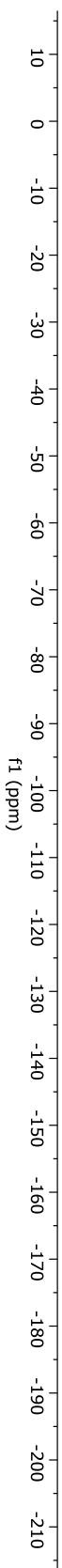
88

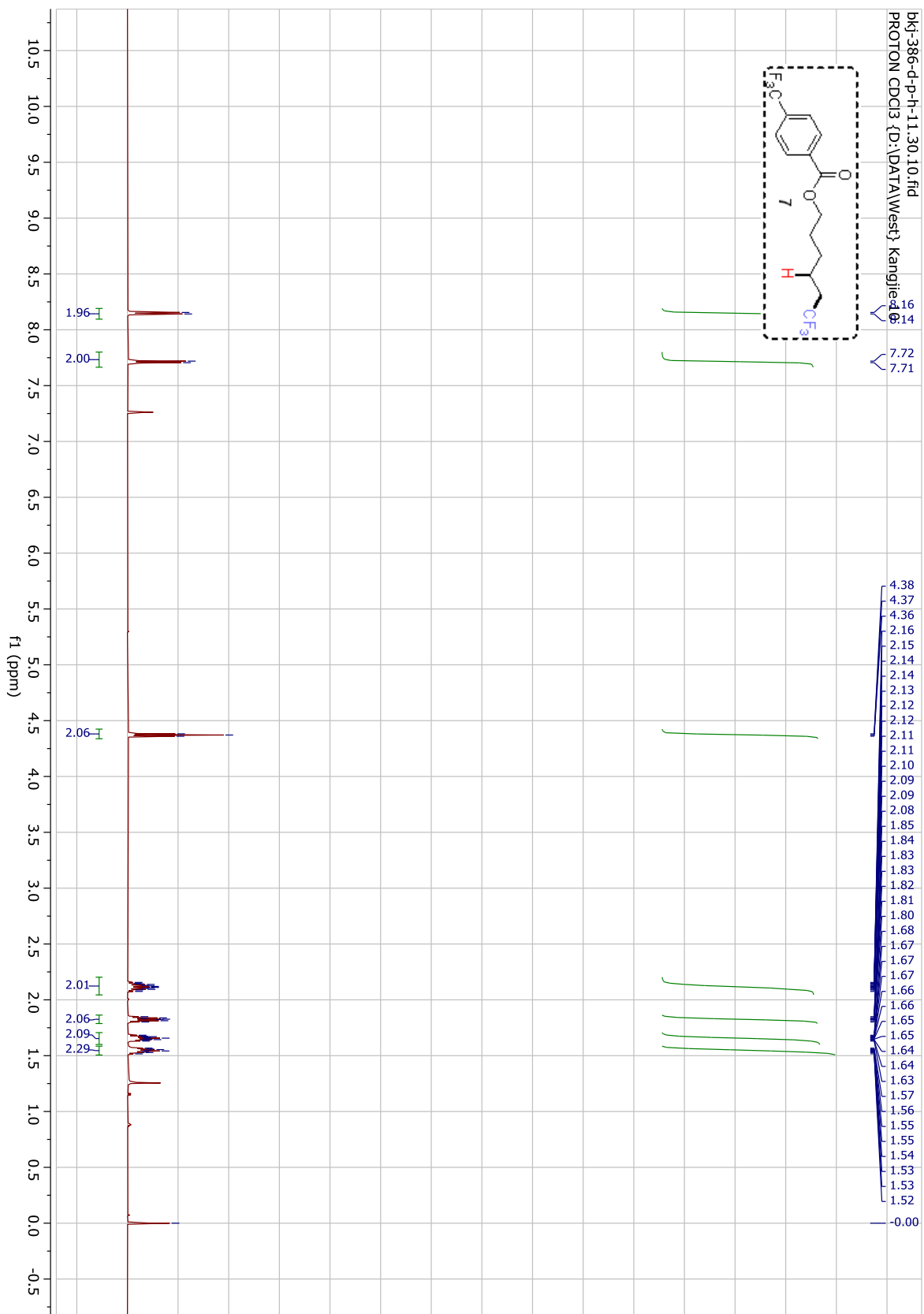


bkj-386-d-p-h-11-29-11.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 4

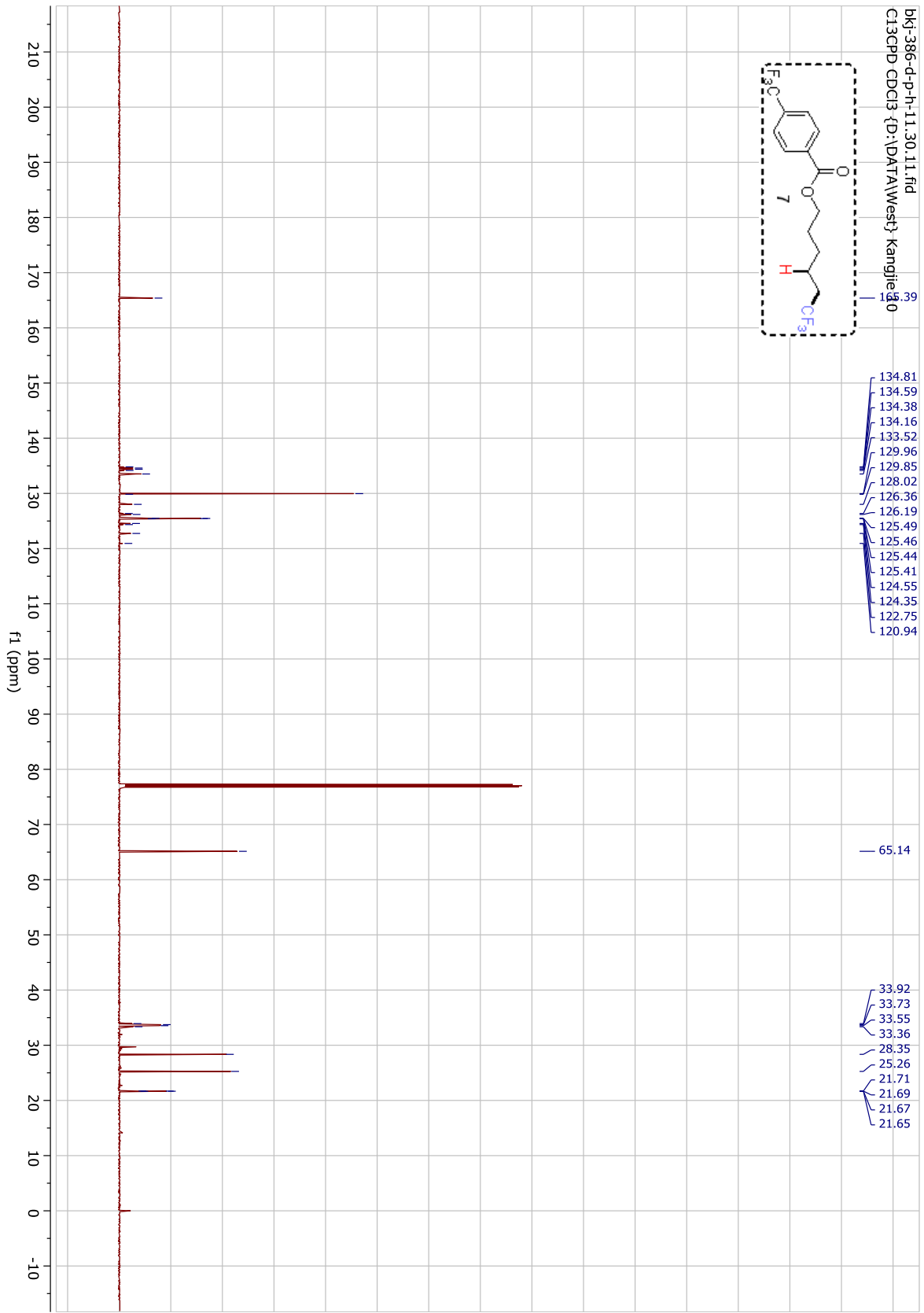
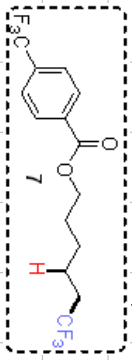


-66.34
-66.36
-66.38





bjf-386-d-p-h-11.30.11.ftd
C13CPD CDCl3 (D:\DATA\Westy\Kangjie\90

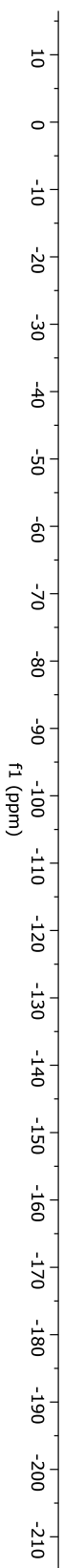
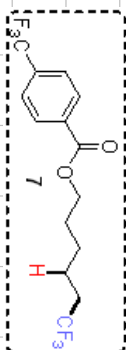


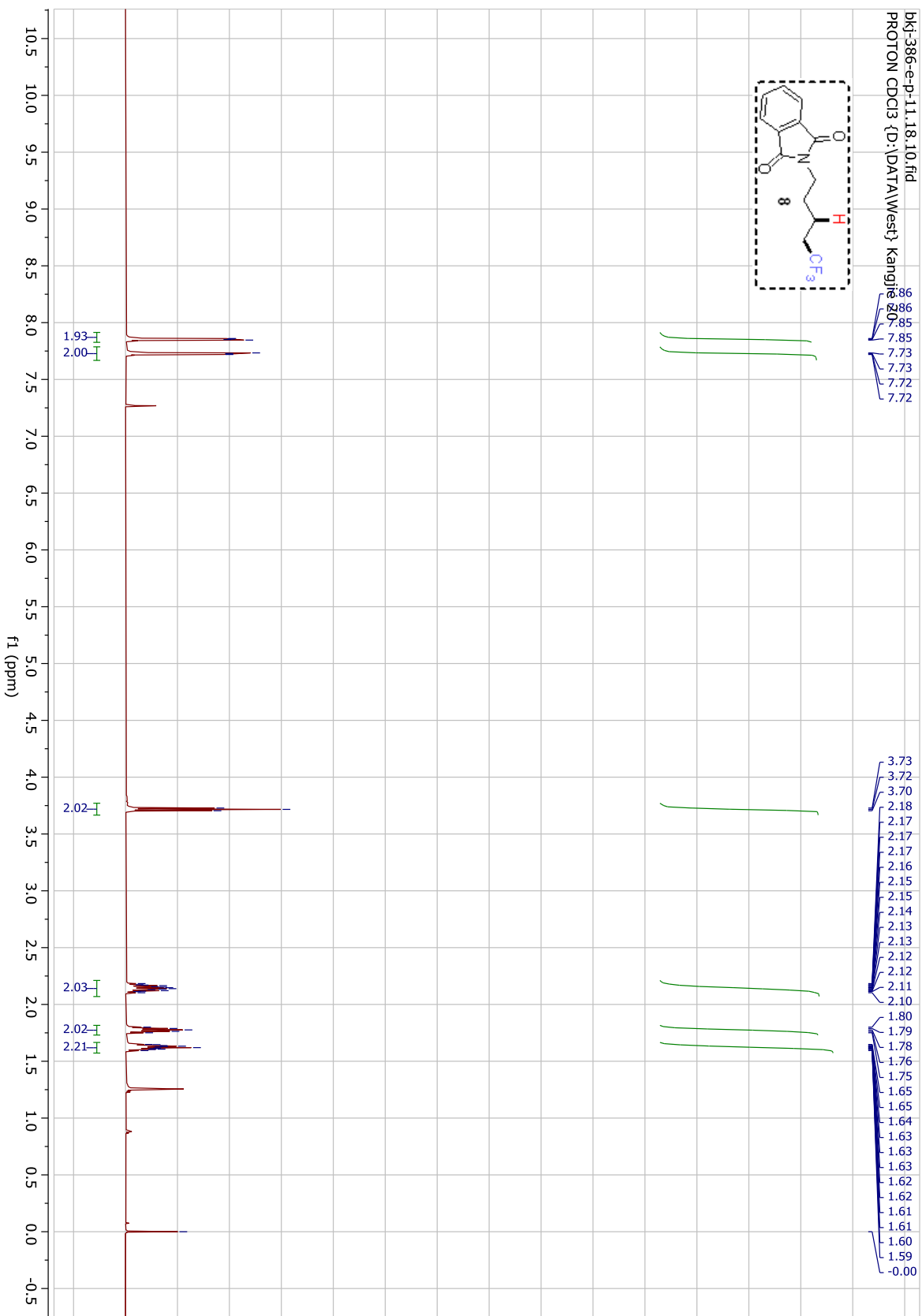
bkj-386-d-p-h-11.30.12.fid

F19_baseline_correct 1D

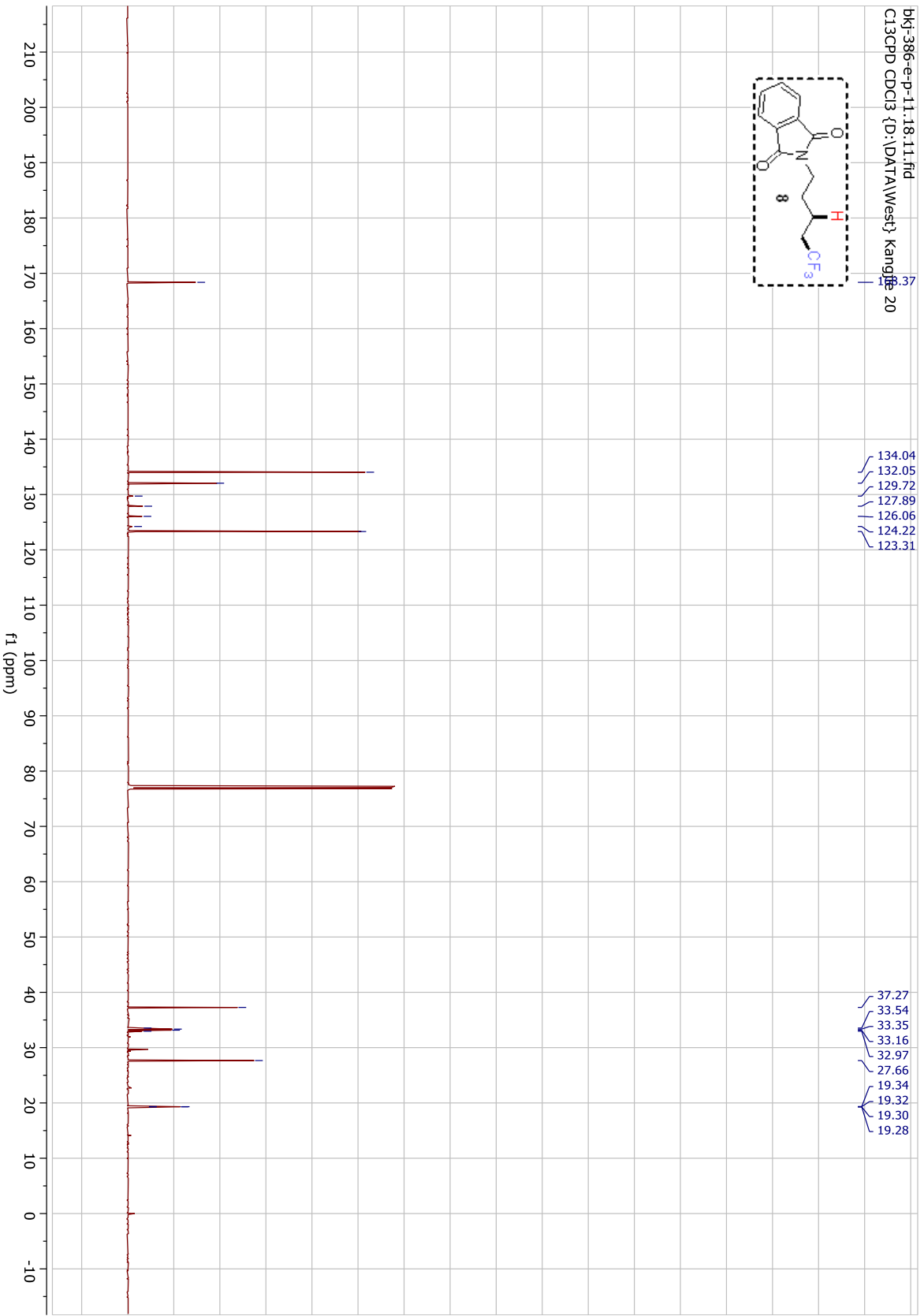
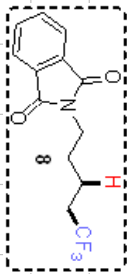
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 10

63.13
66.35
66.37
66.37
66.39
66.39

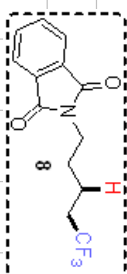




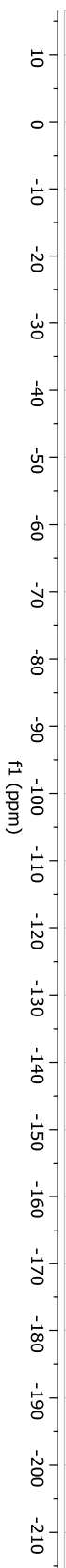
bjl-386-e-p-11-18-11.fid
C13CPD CDCl3 (D:\DATA\Westj Kang) 20



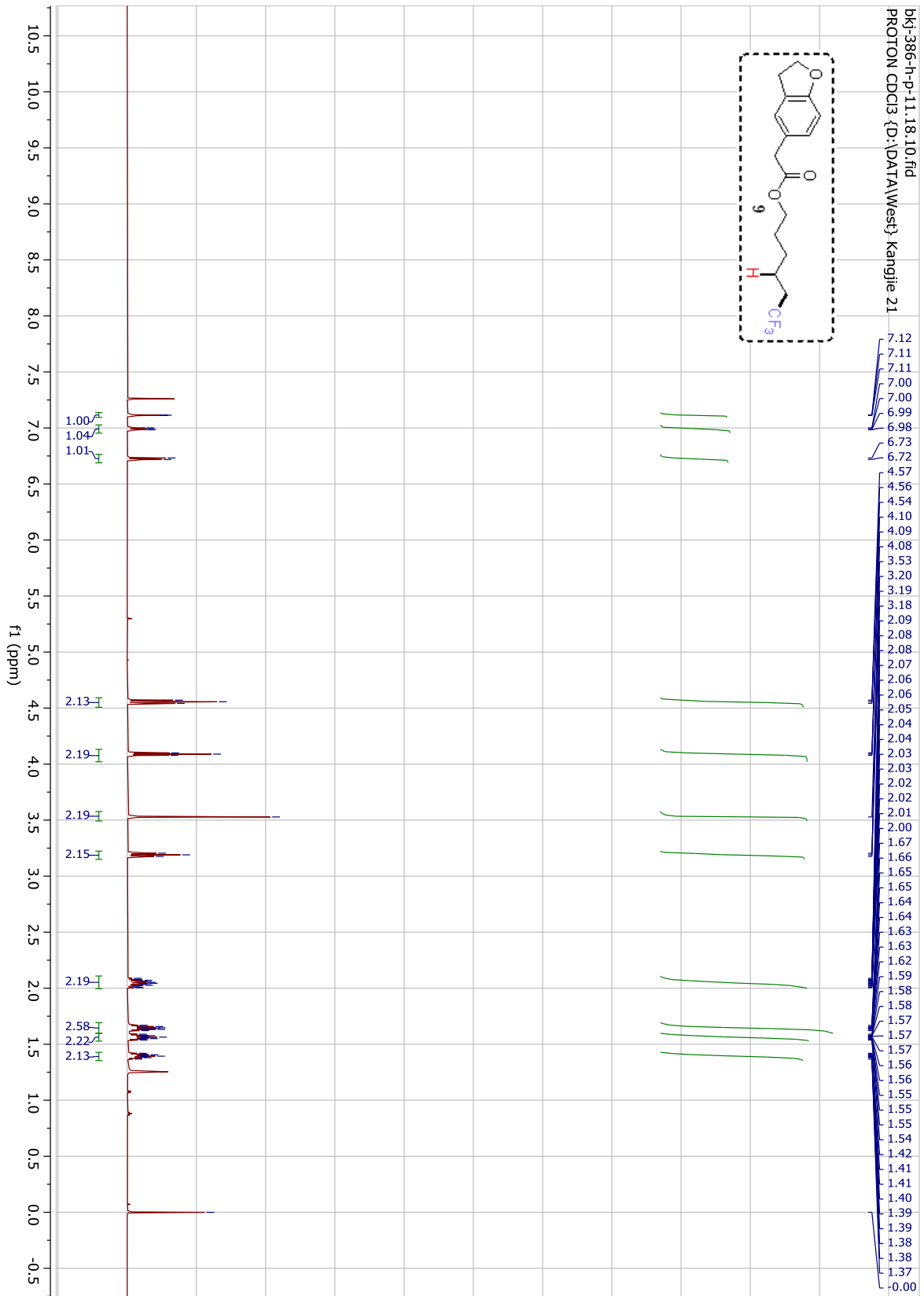
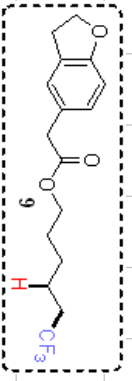
bkj-386-e-p-11.18.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 20



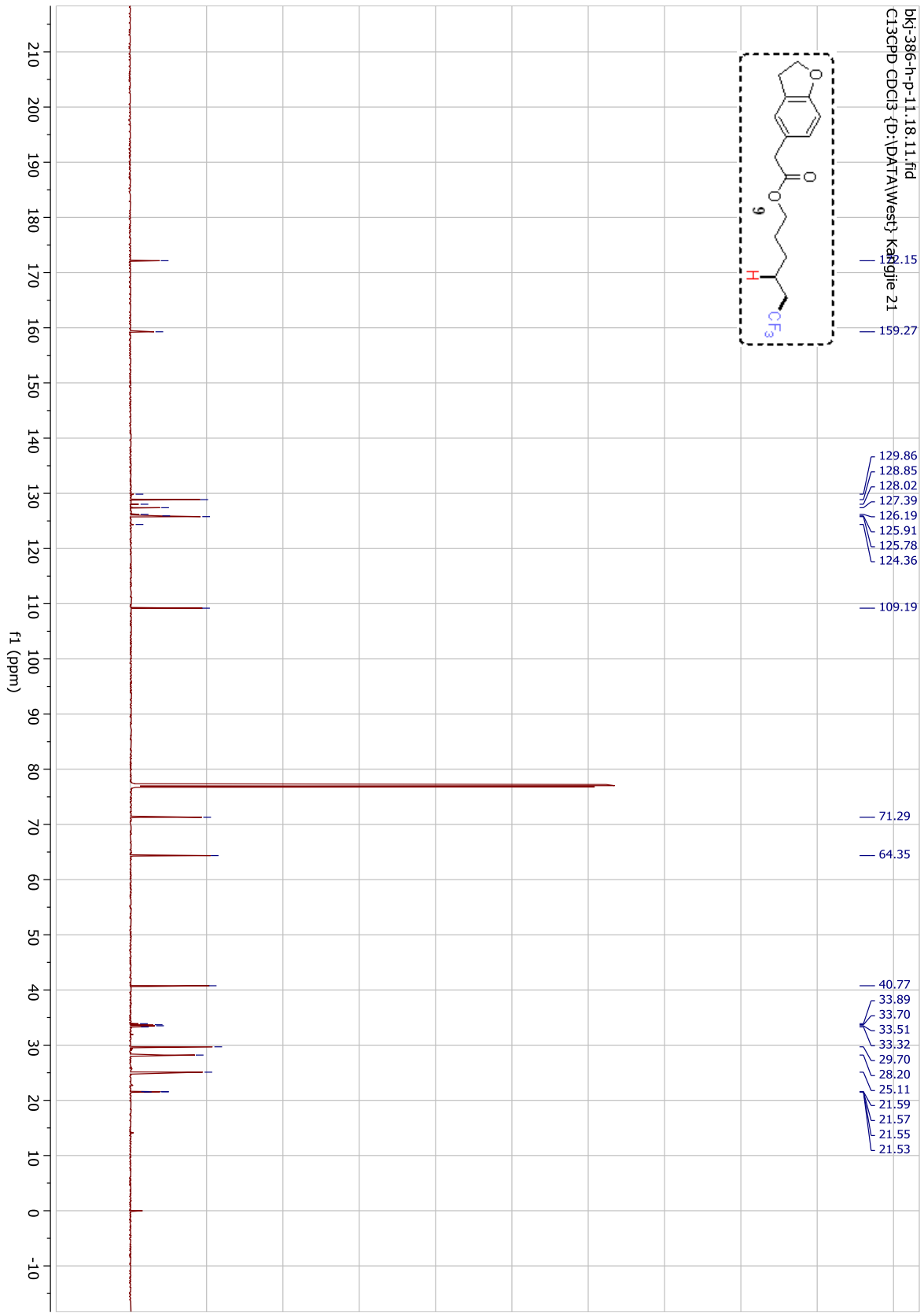
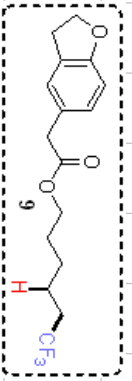
-66.33
-66.35
-66.37



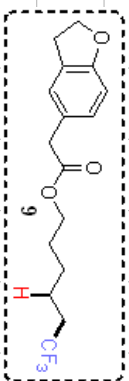
bk1-386-h-p-11.18.10.fid
PROTON CDCl3 {D:\DATAWest\Kanglie 21



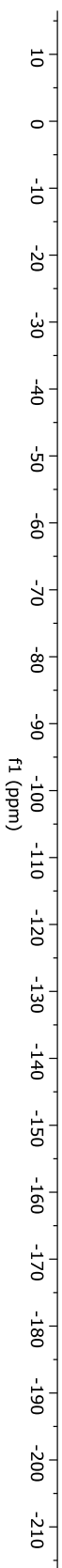
bkj-386-h-p-11.18.11.fid
C13CPD CDCl3 (D:\DATA\Westj_kapjije 21



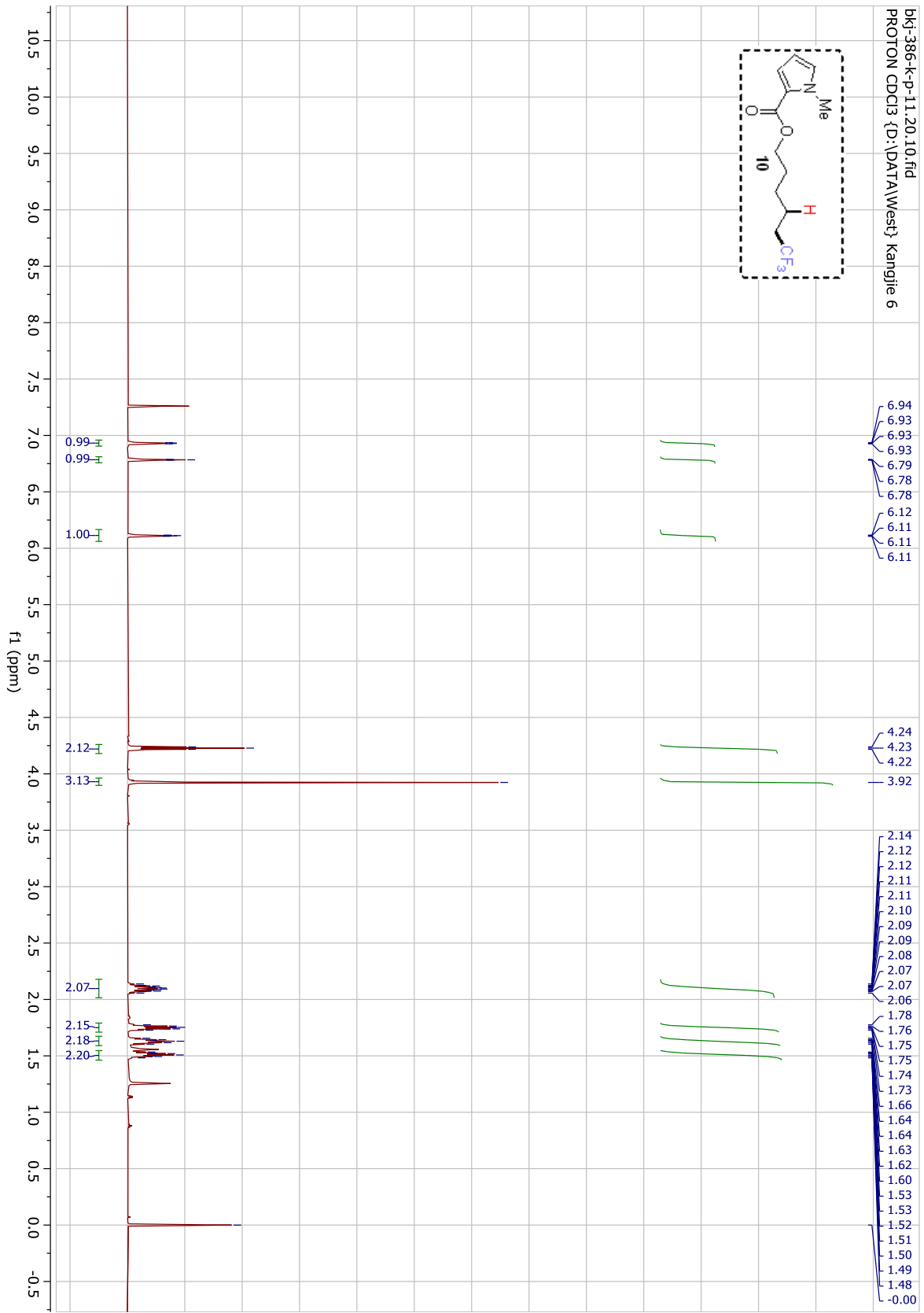
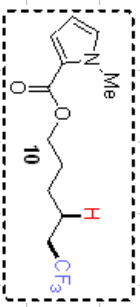
bkj-386-h-p-11.18.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West}\Kangjie 21



-66.34
-66.36
-66.38

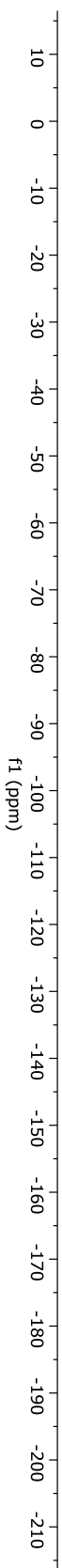
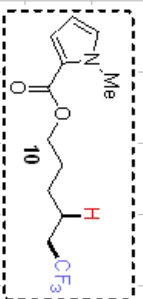


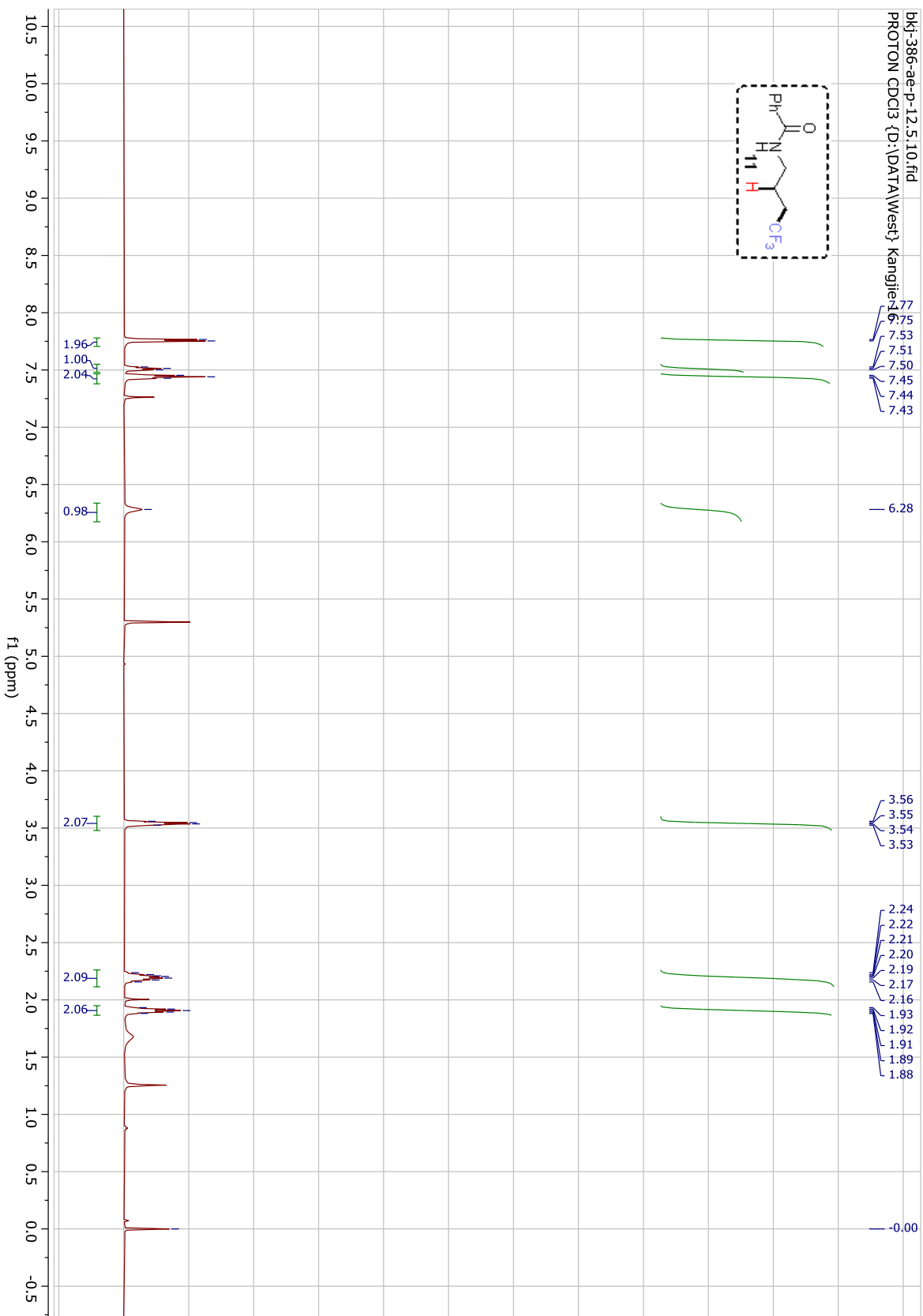
bk1-386-k-p-11.20.10.fid
PROTON CDCl3 {D:\DATA\West\} Kangjie 6



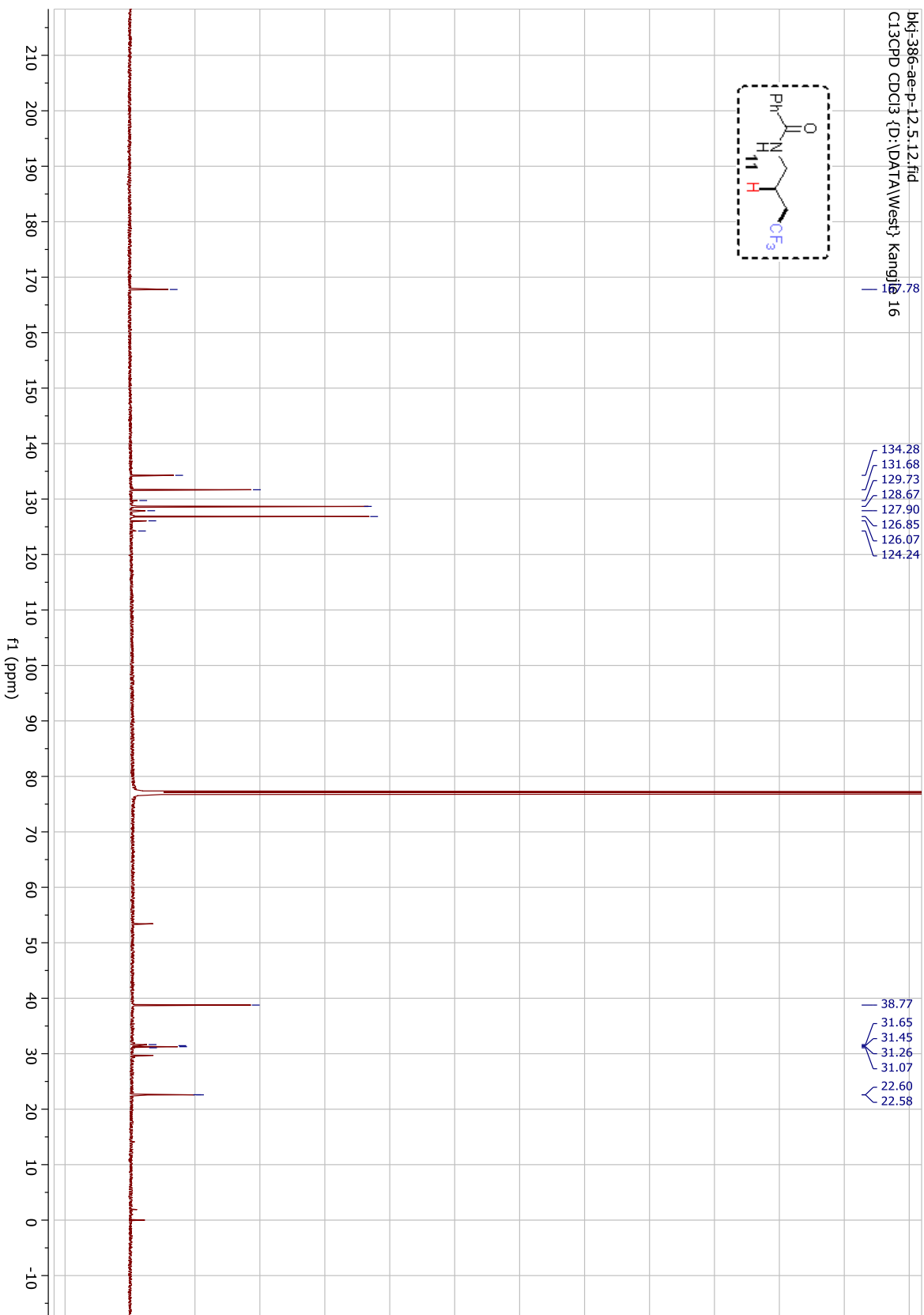
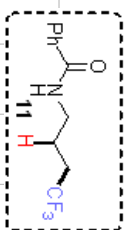
bkj-386-k-p-11.20.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 6

-66.36
-66.38
-66.39



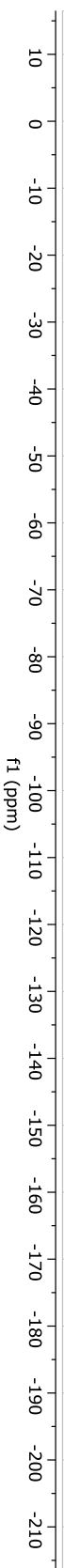
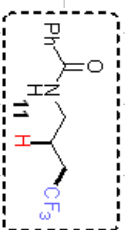


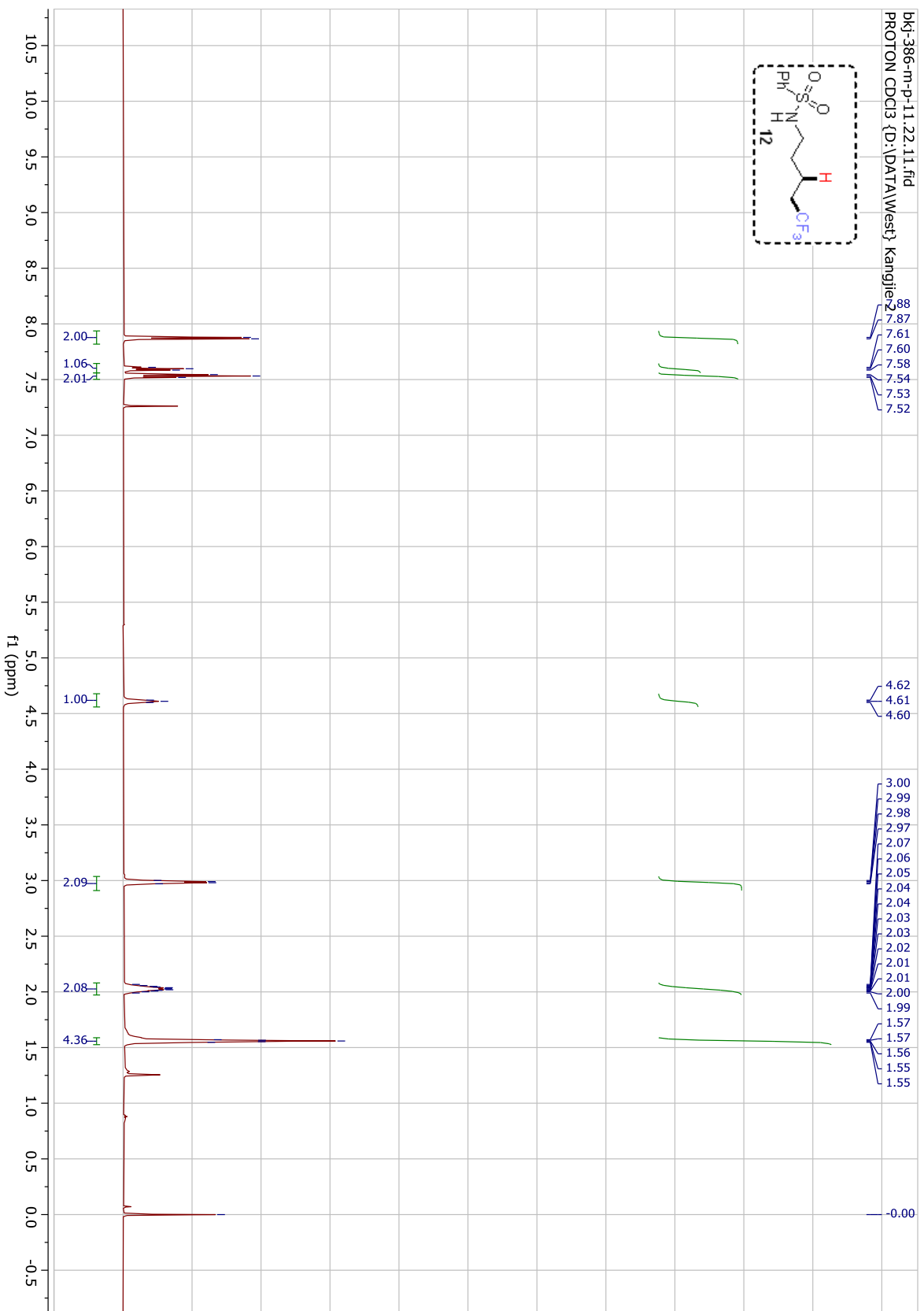
bkj-386-ae-p-12.5.12.fid
C13CPD CDCl3 (D:\DATA\Westj Kangji)
16



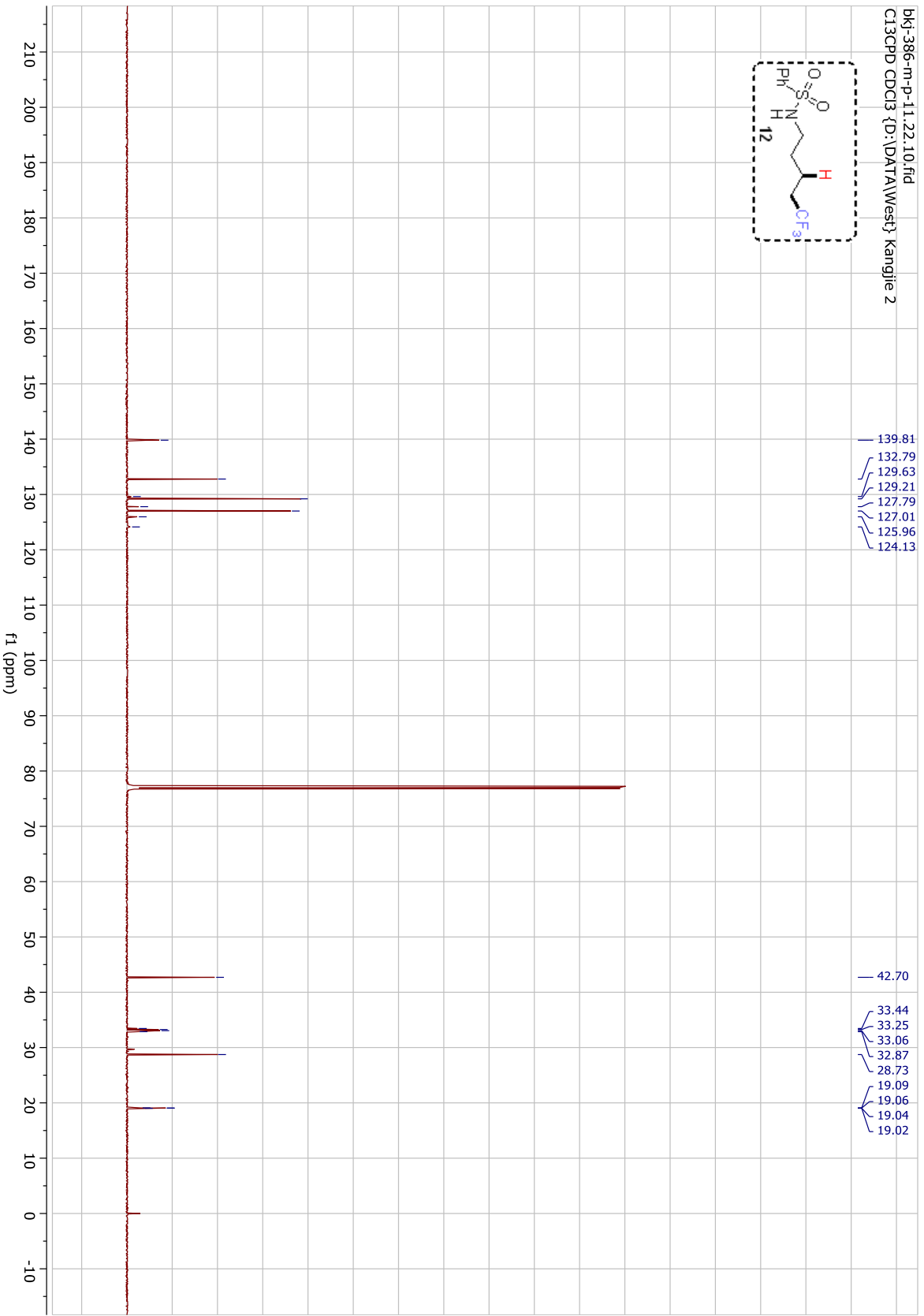
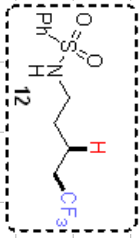
bkj-386-ae-p-12.5.11.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 16

-66.11
-66.13
-66.15



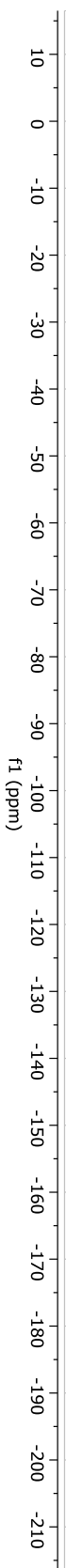
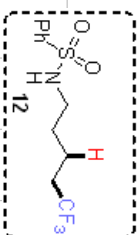


bjf-386-m-p-11-22.10.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 2

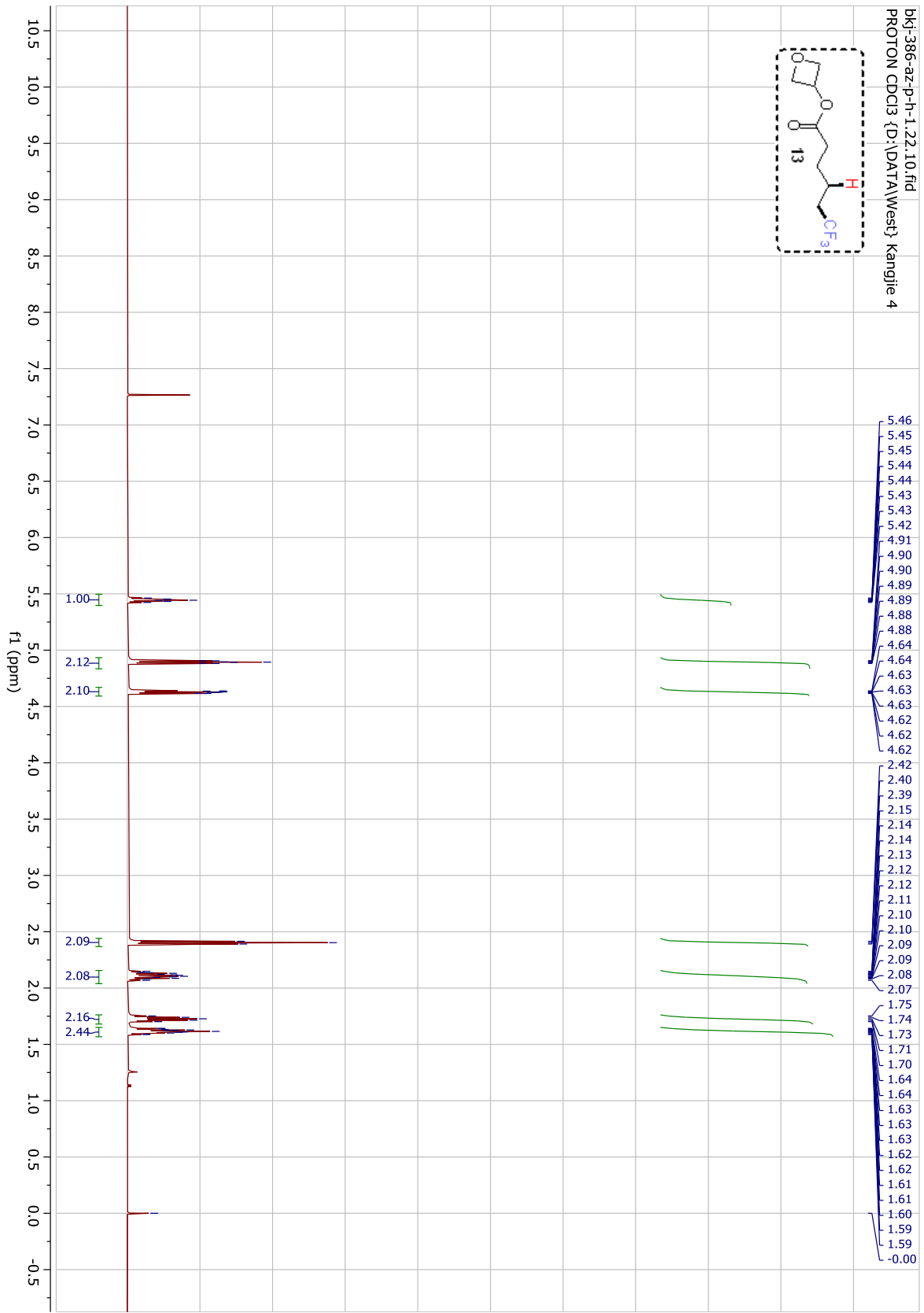
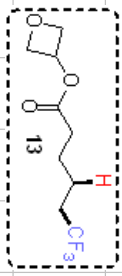


bkj-386-m-p-11.22.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 2

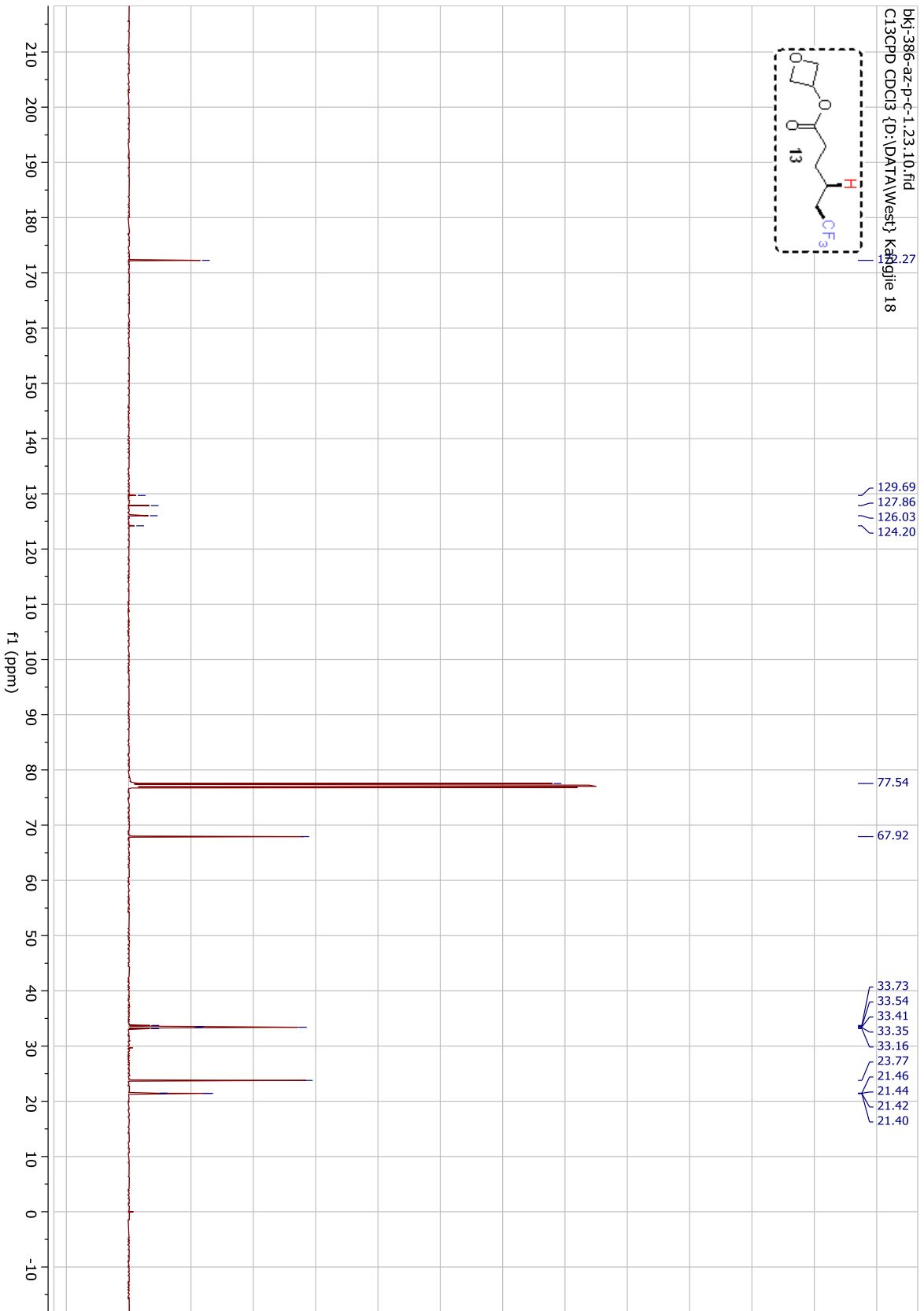
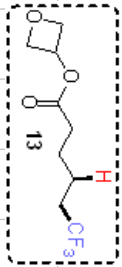
-66.32
-66.34
-66.36



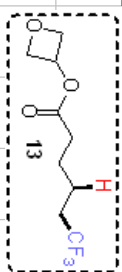
bkl-386-az-p-h-1.22.10.fid
PROTON CDCl3 {D:\DATA\West\ Kangjie 4



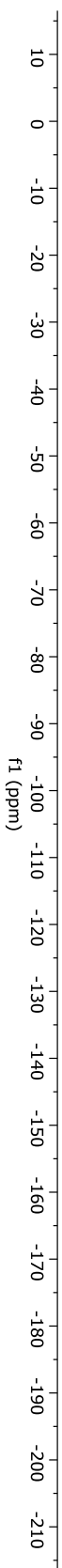
bkj-386-az-p-c-1.23.10.fid
C13CPD CDCl3 (D:\DATA\Westj) Kapjele 18



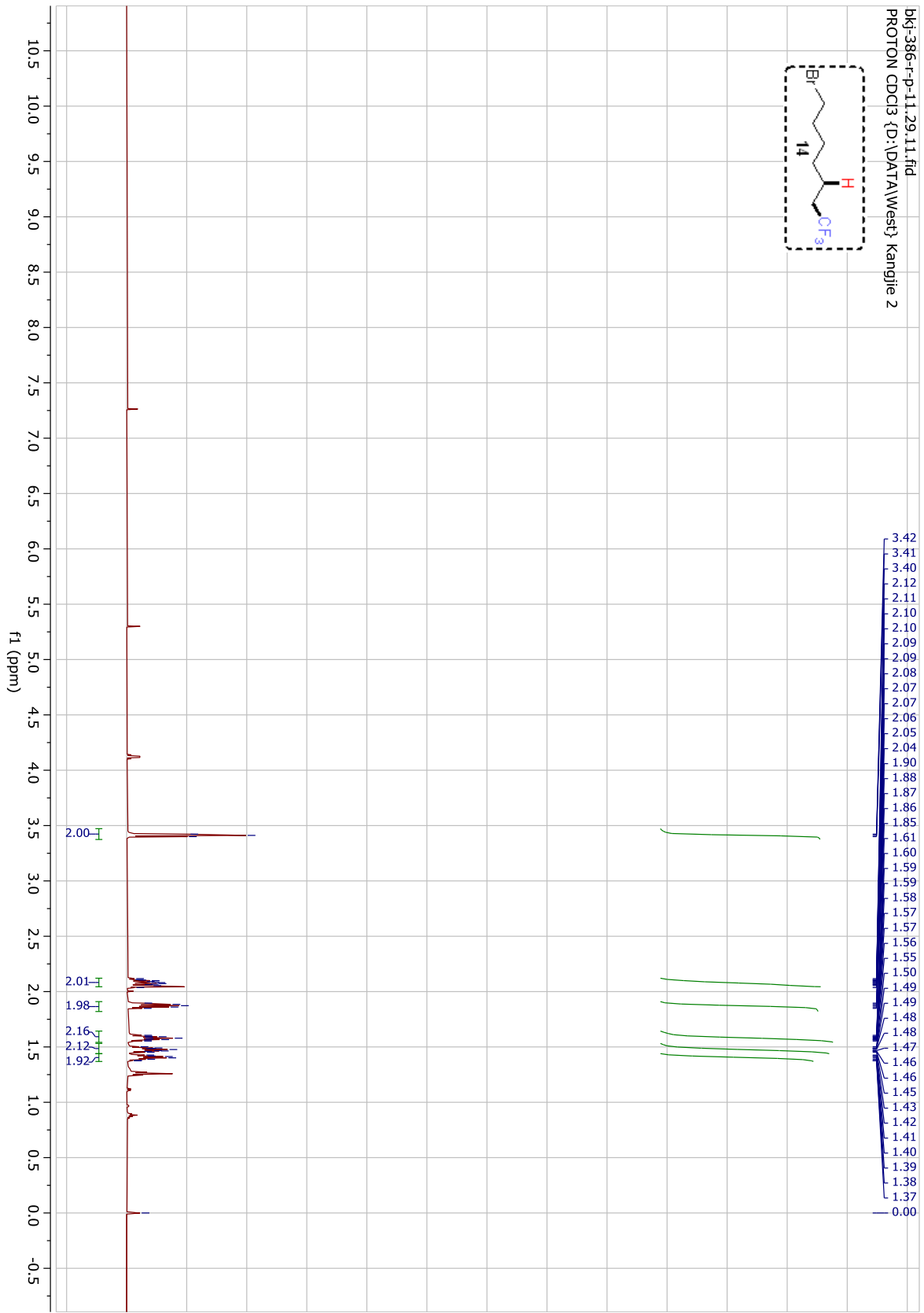
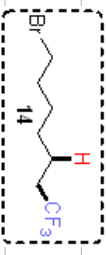
bkj-386-az-p-h-1.22.11.fid
F19_baseline_correct_ID
F19_baseline_correct_CDCl3 {D:\DATA\West\ Kangjie 4



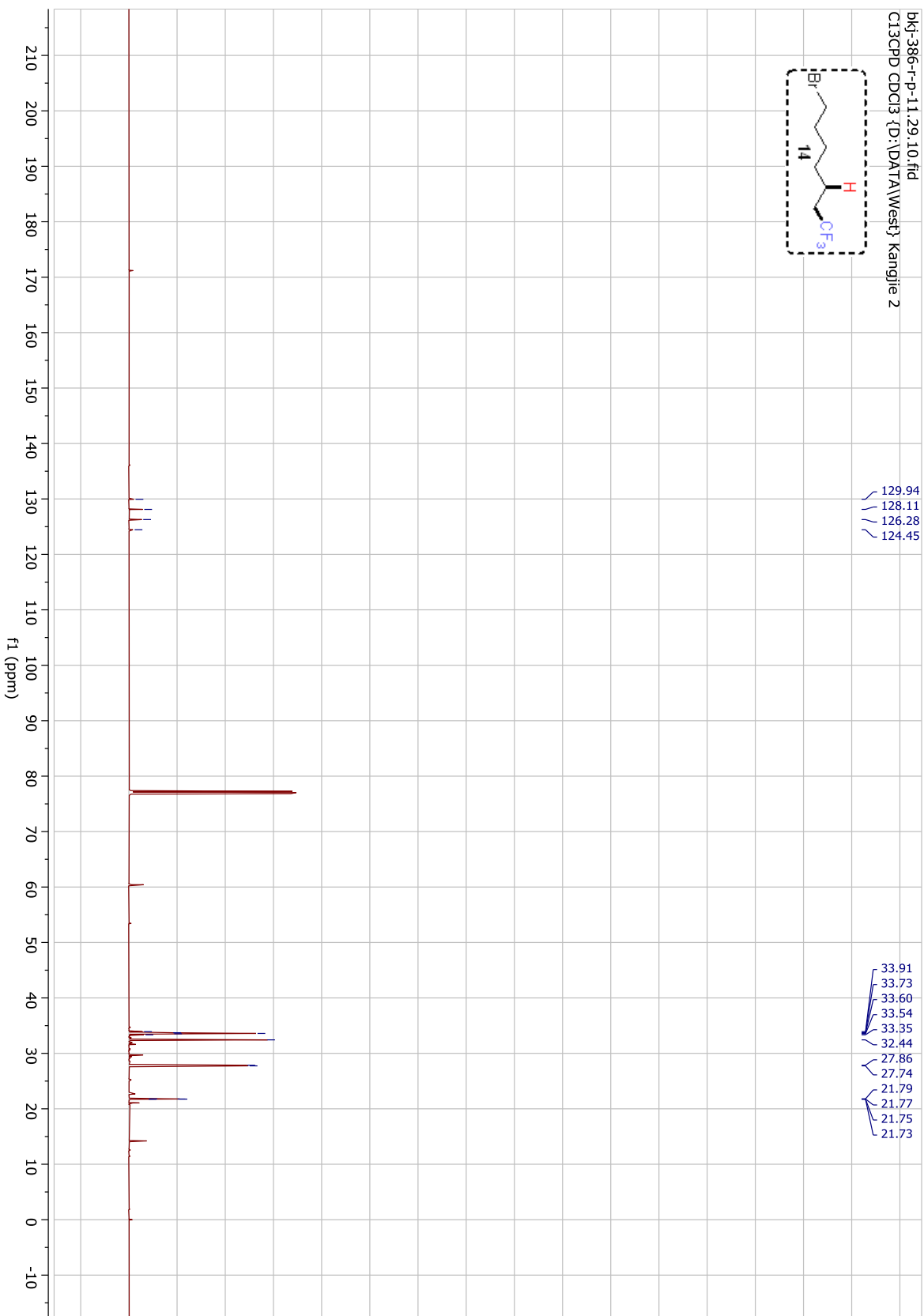
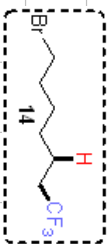
66.35
66.37
66.39



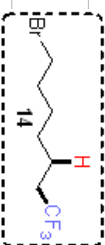
bk1-386-r-p-11.29.11.fid
PROTON CDCl3 {D:\DATA\West\} Kangjie 2



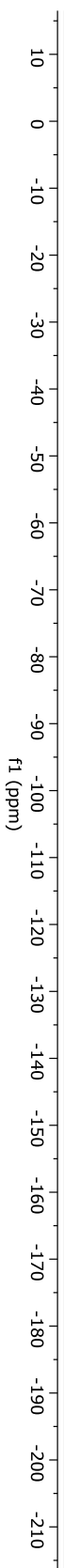
bk1-386-r-p-11.29.10.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 2



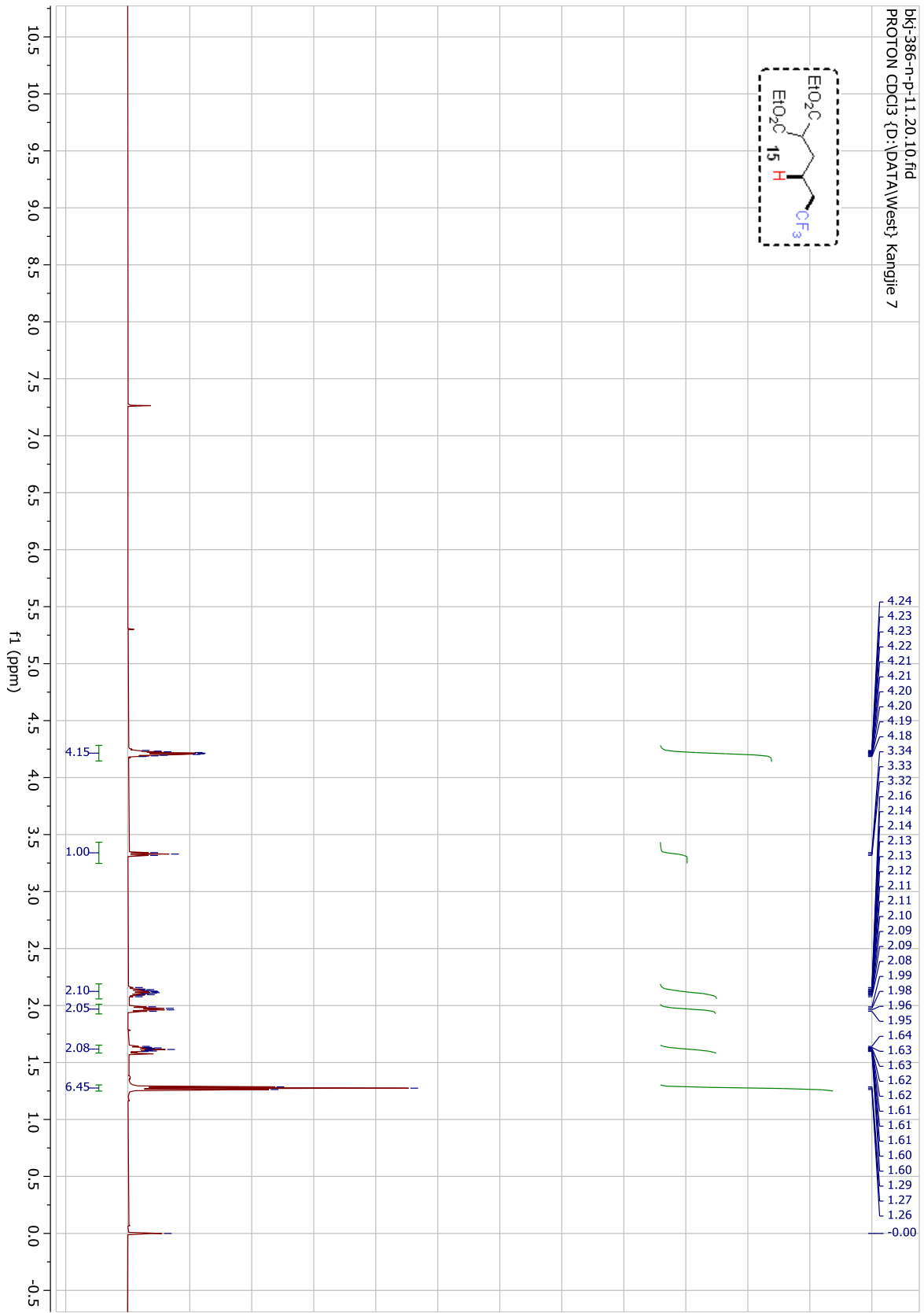
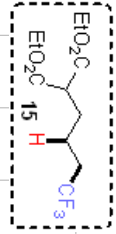
bkj-386-r-p-11.29.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 2



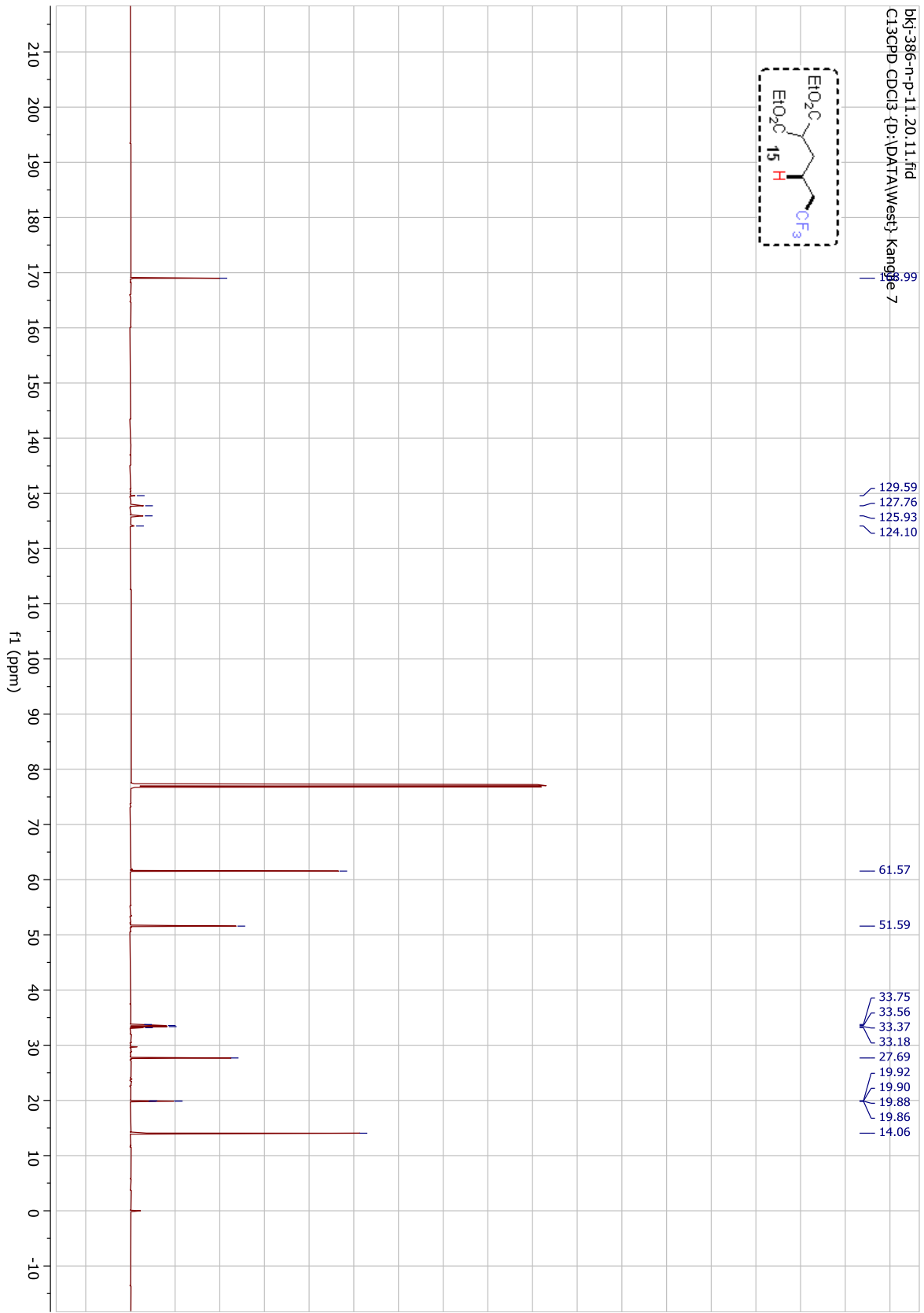
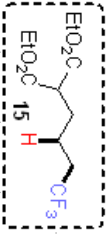
-66.39
-66.41
-66.43



bk1-386-n-p-11.20.10.fid
PROTON CDCl3 {D:\DATA\West\ Kangjie 7

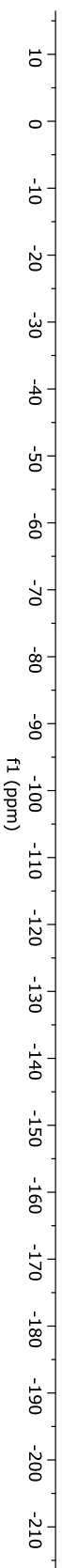
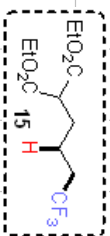


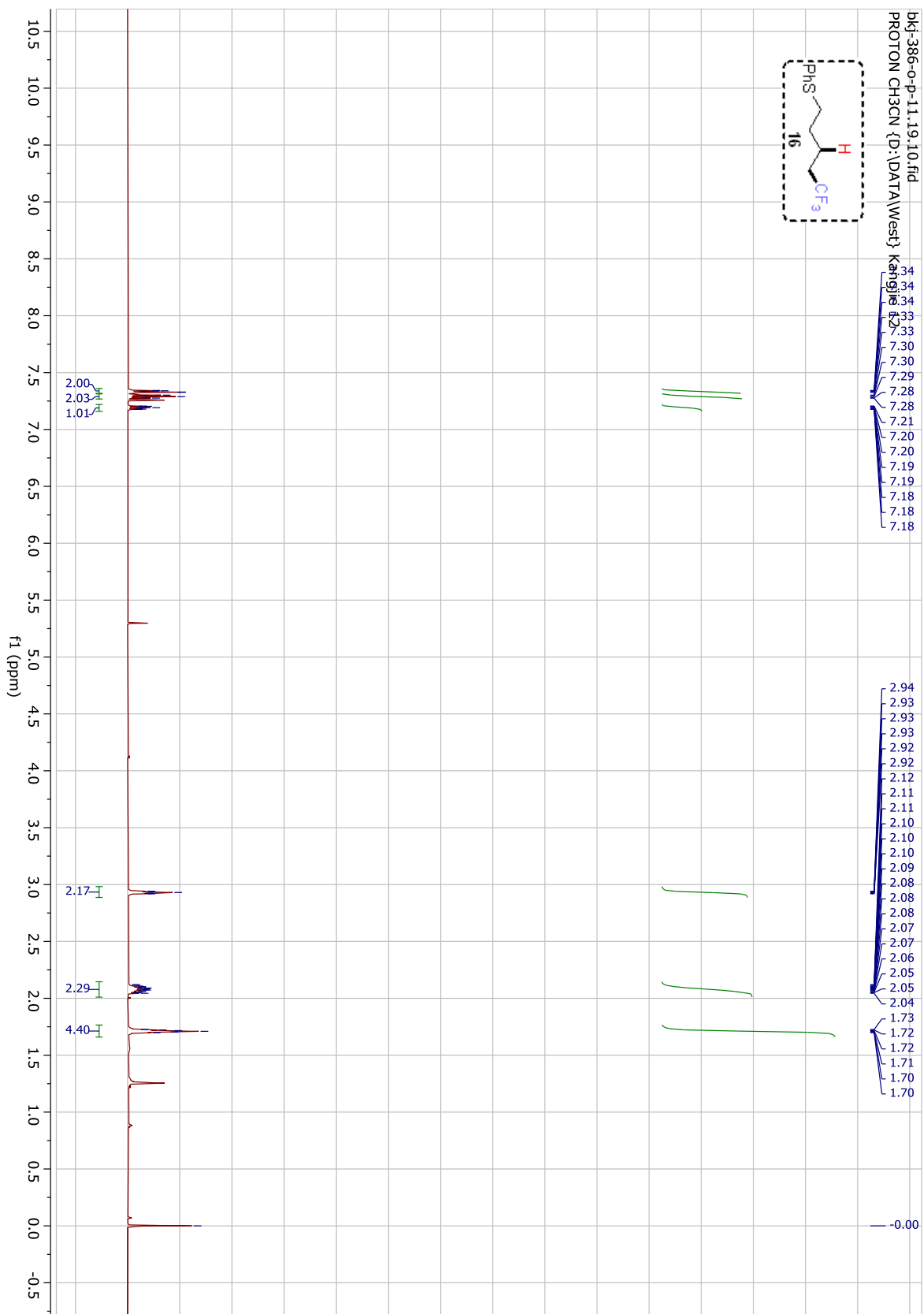
bkj-386-n-p-11.20.11.fid
C13CPD CDCl3 (D:\DATA\Westj_Kang) Page 7



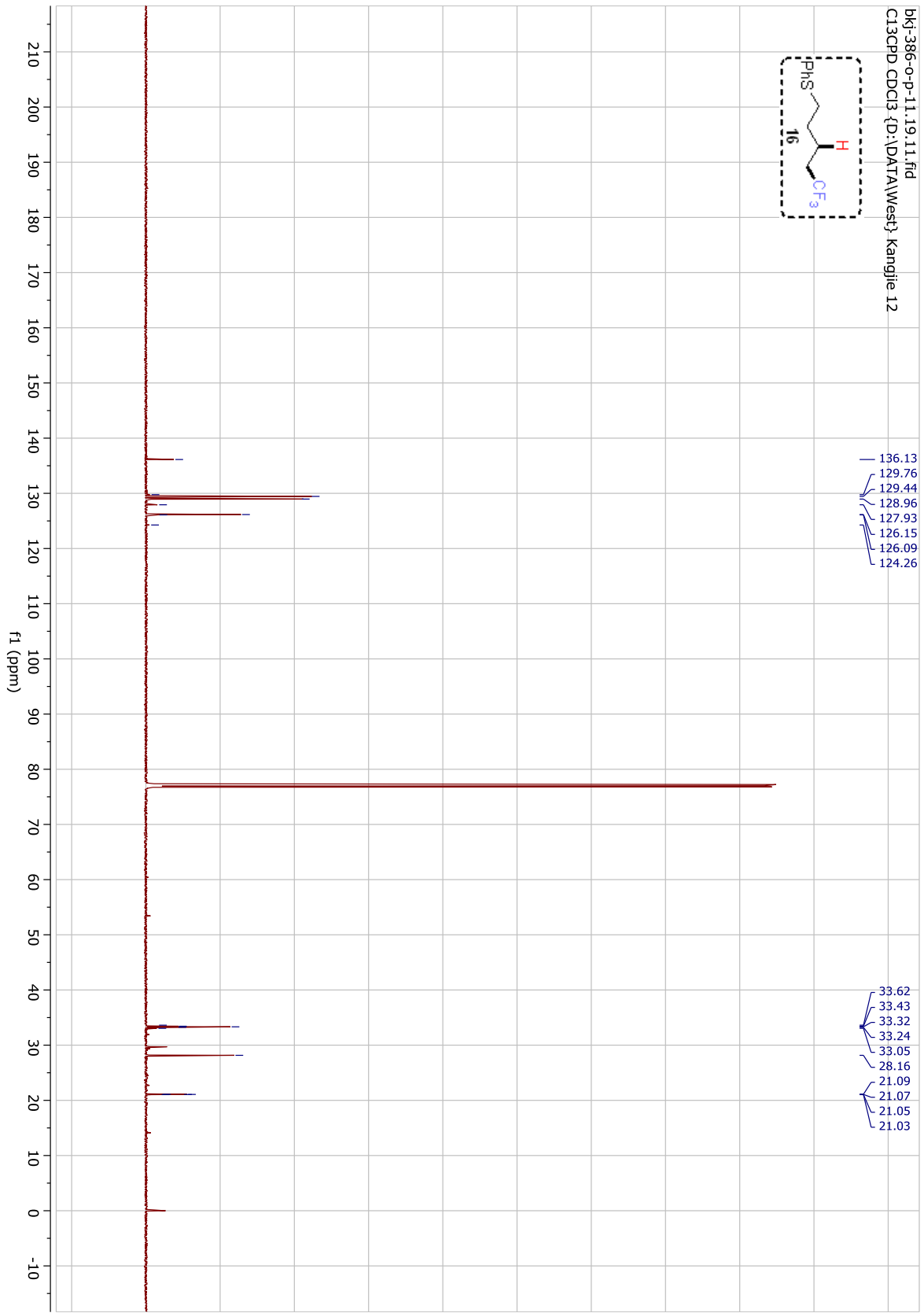
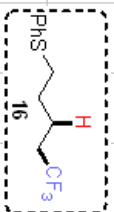
bkj-386-n-p-11.20.12.fid
F19_baseline_correct_1D
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 7

96.38
96.40
96.42

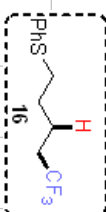




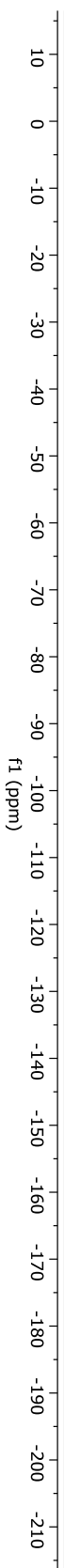
bkj-386-o-p-11.19.11.fid
C13CPD CDCl3 (D:\DATA\Westj_Kangjie 12

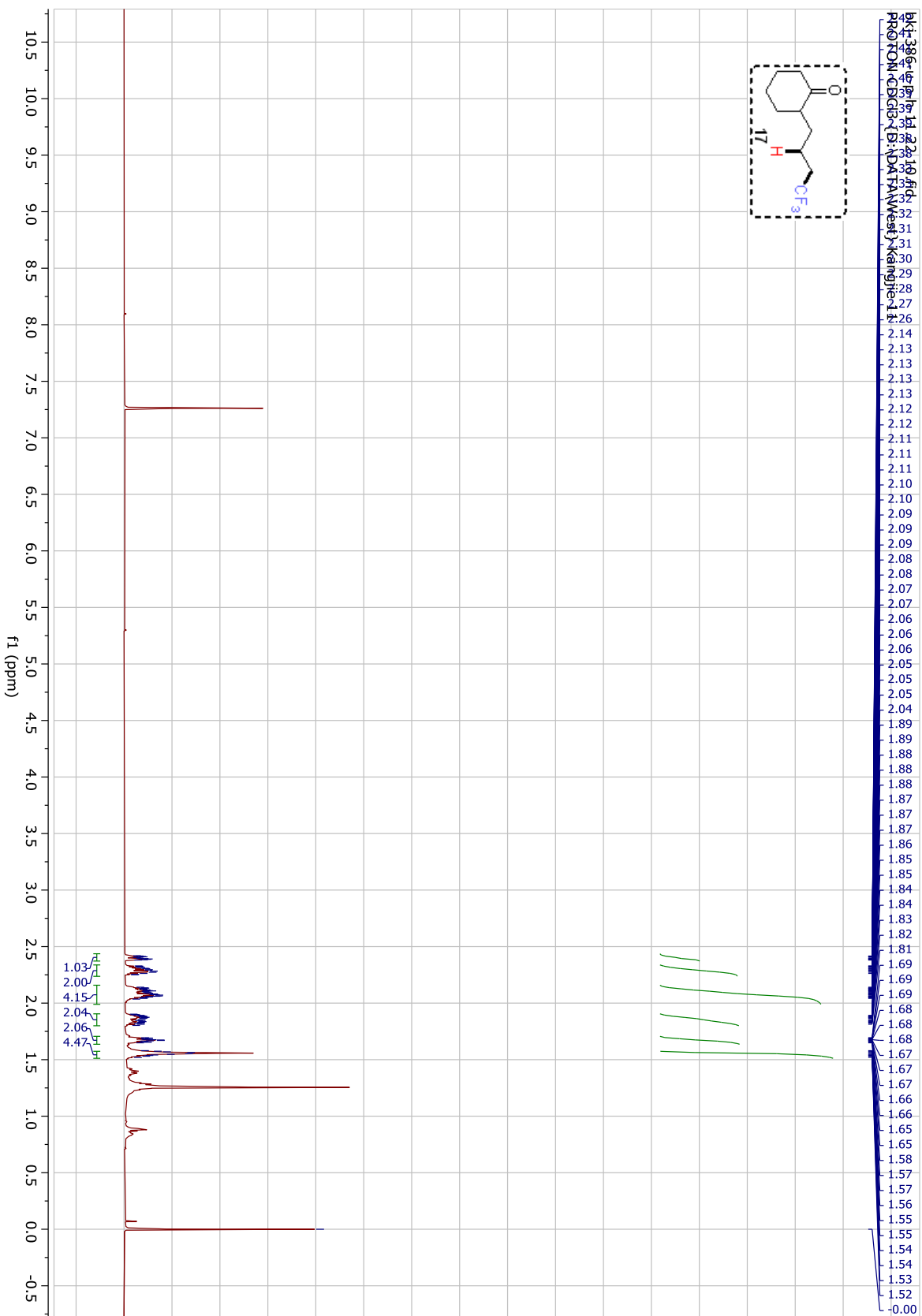


bkj-386-o-p-11.19.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 12

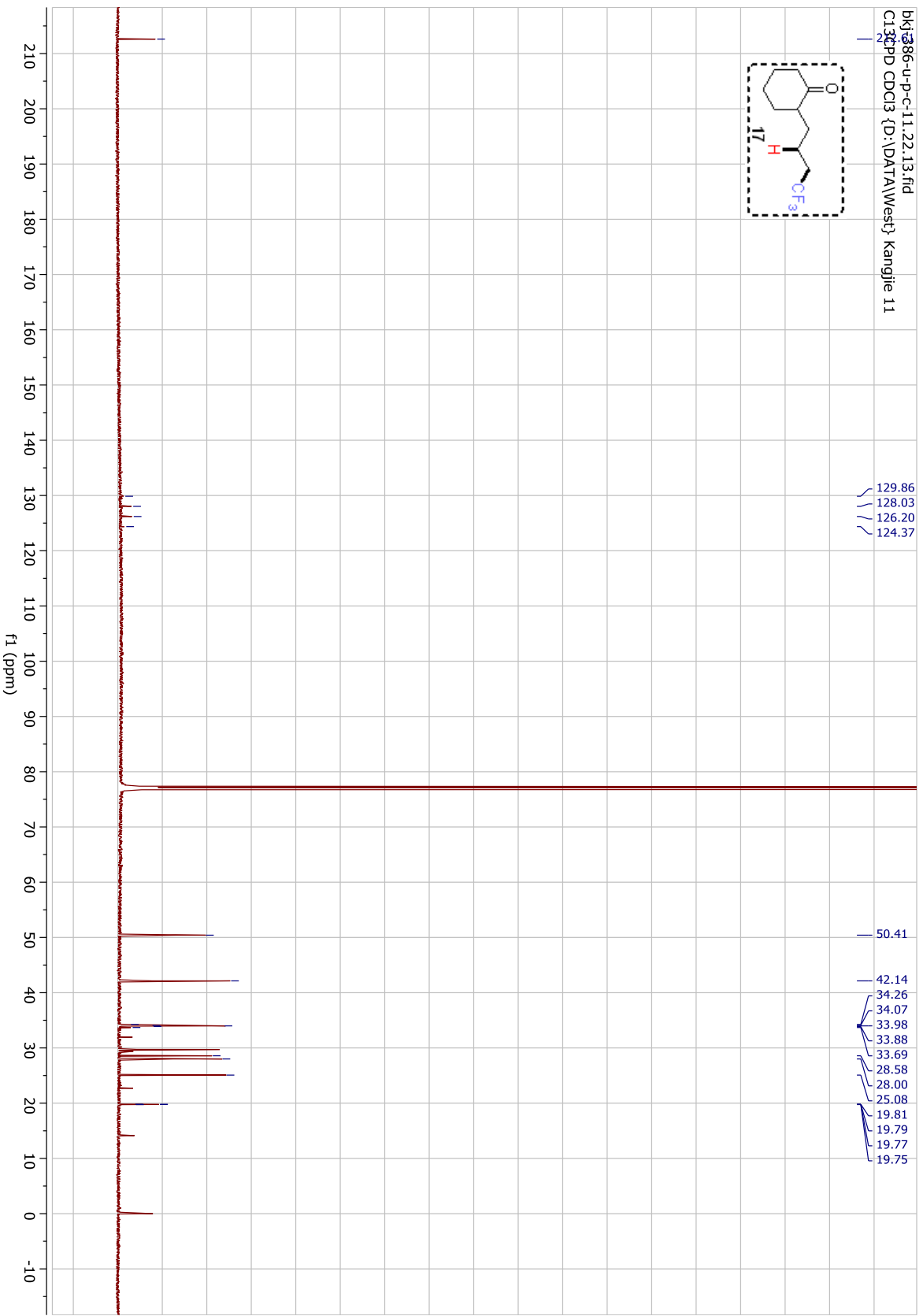
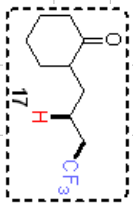


66.33
66.35
66.37



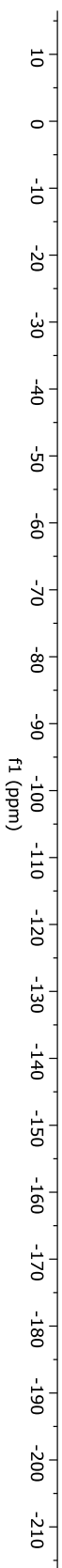
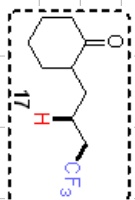


bk1386-u-p-c-11.22.13.fid
C138PD CDCl3 (D:\DATA\West\ Kangjie 11

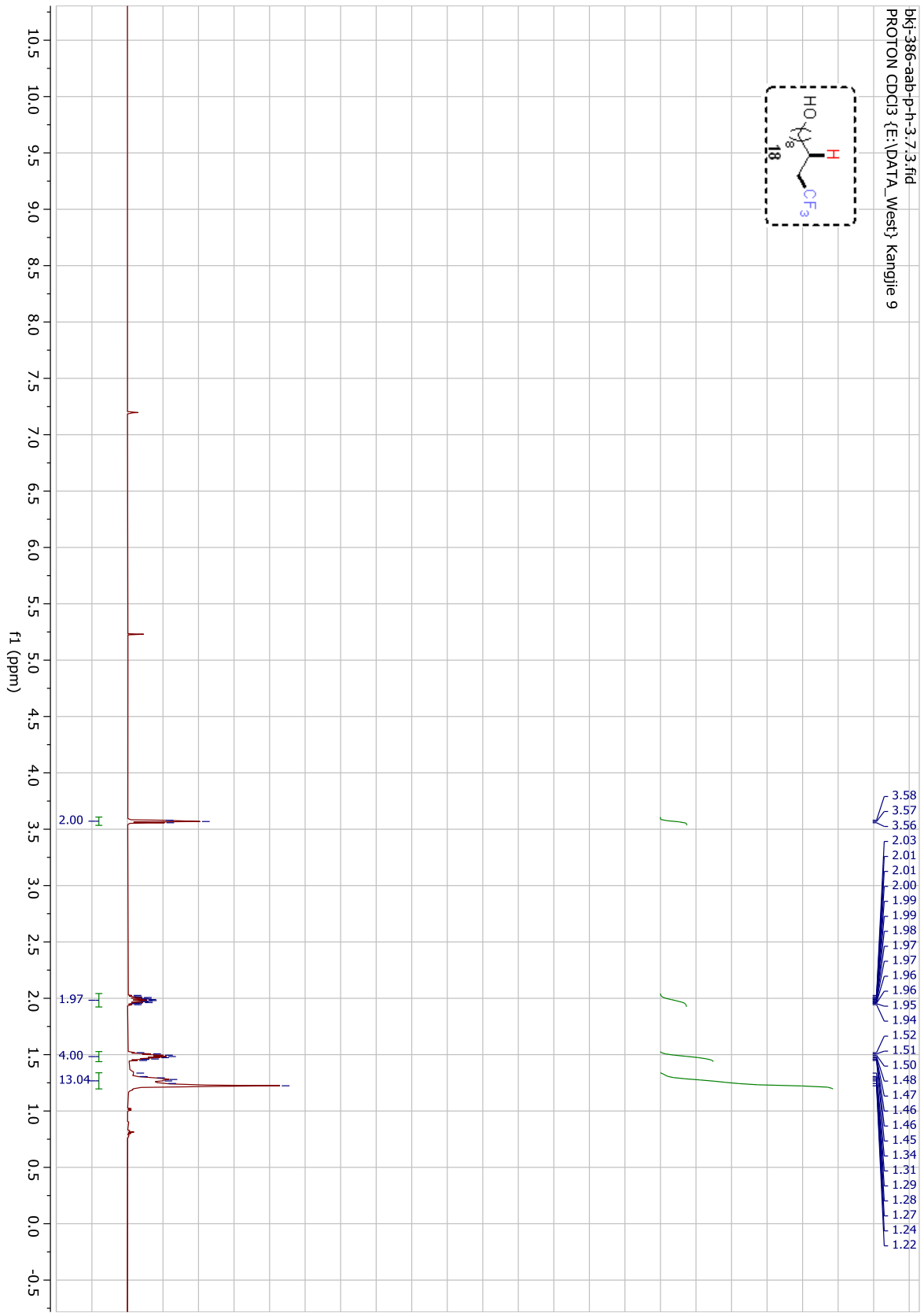
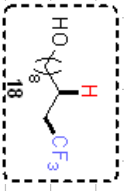


bkj-386-u-p-h-11-22-11.ftd
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West\ Kangjie 11

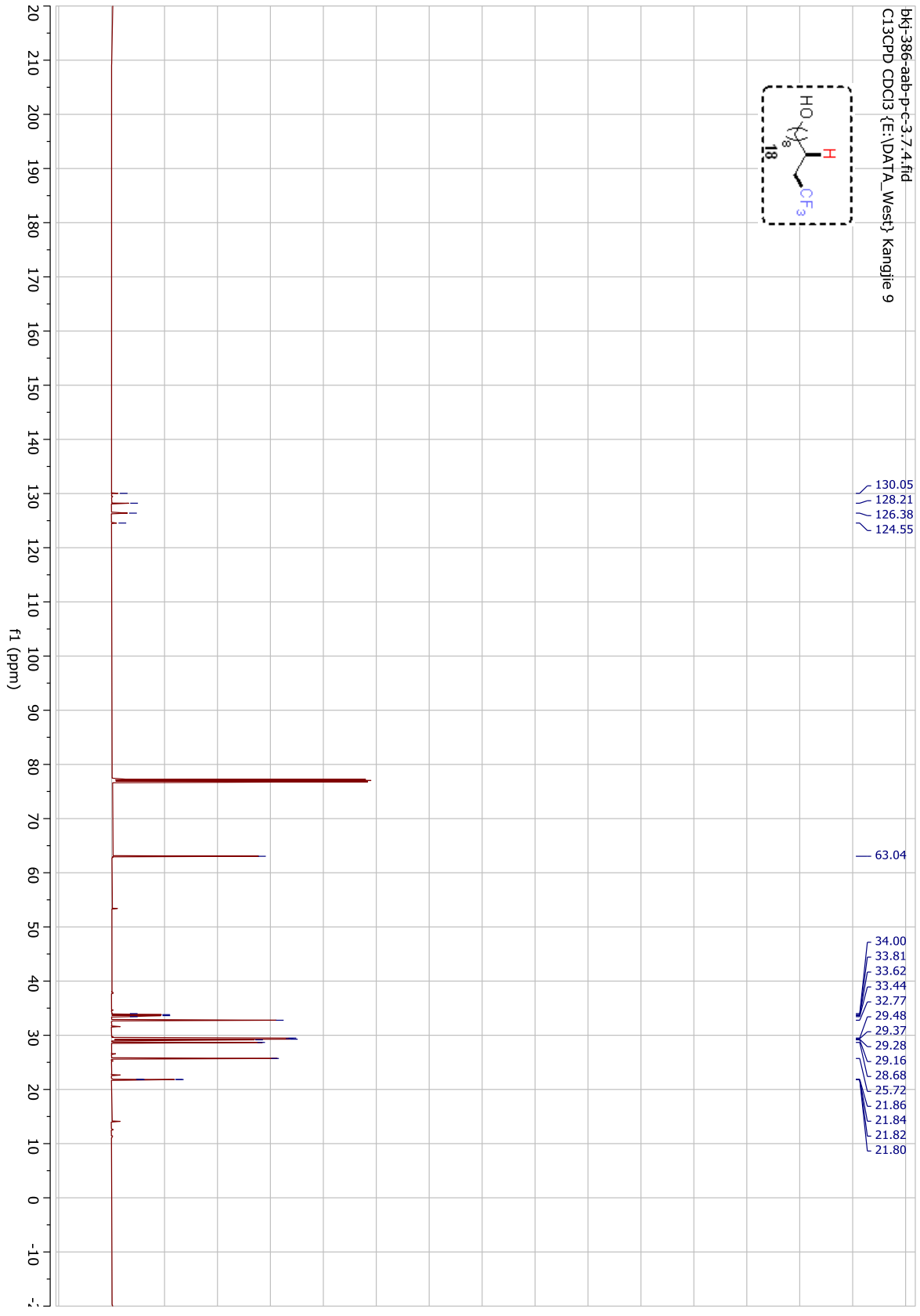
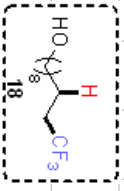
96.36
96.38
96.40



bkj-386-aab-p-h-3.7.3.fid
PROTON CDCl3 {E:\DATA_West\Kangjie 9

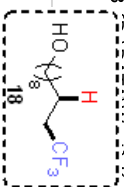


bkj-386-aab-p-c-3-7-4.ftd
C13CPD CDCl3 {E:\DATA_West\Kangjie 9

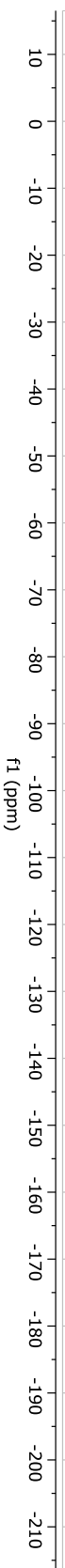


bkj-386-aab-p-f-3.7.10.fid
F19

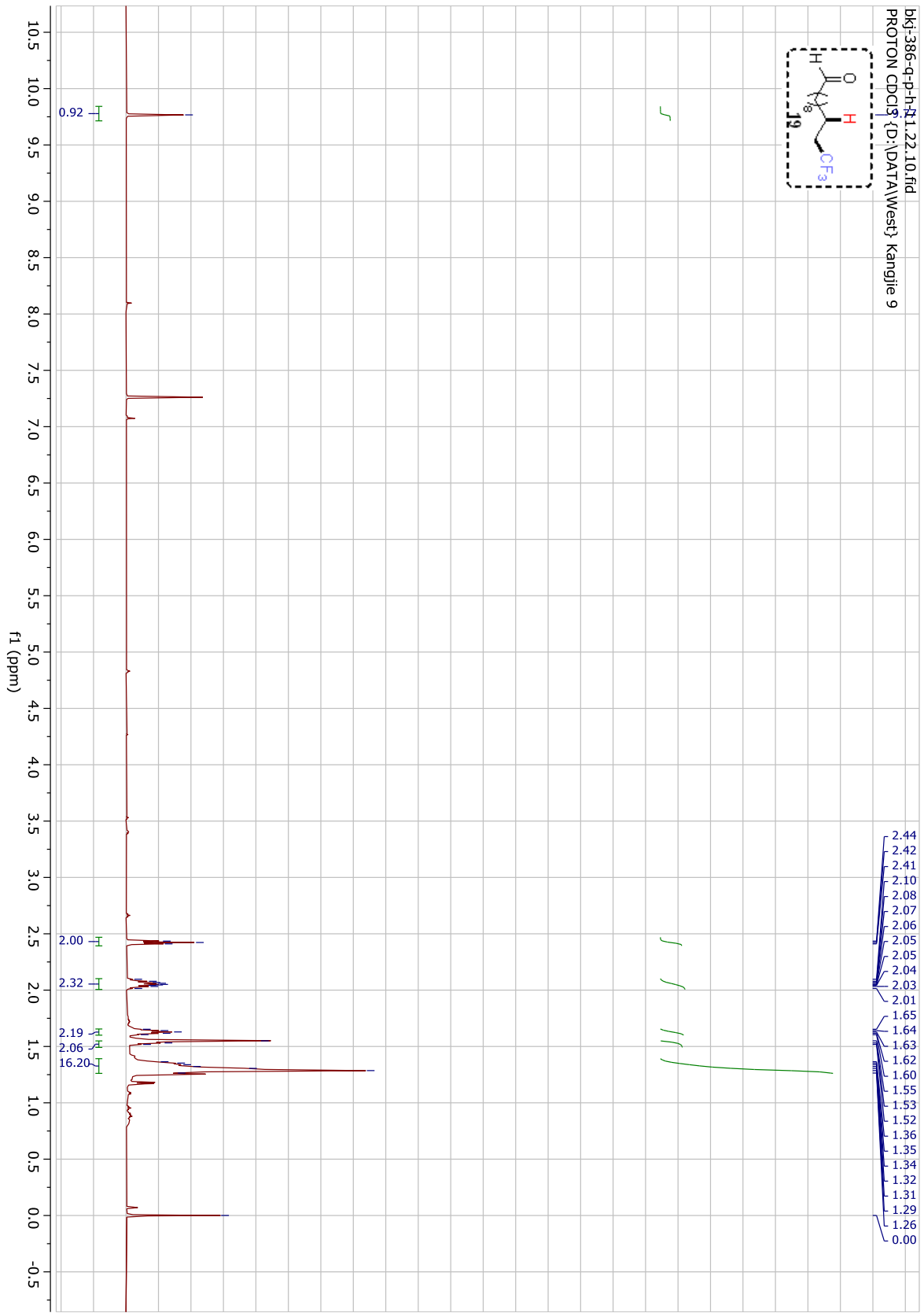
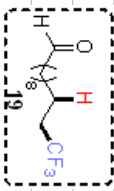
F19 CDCl3



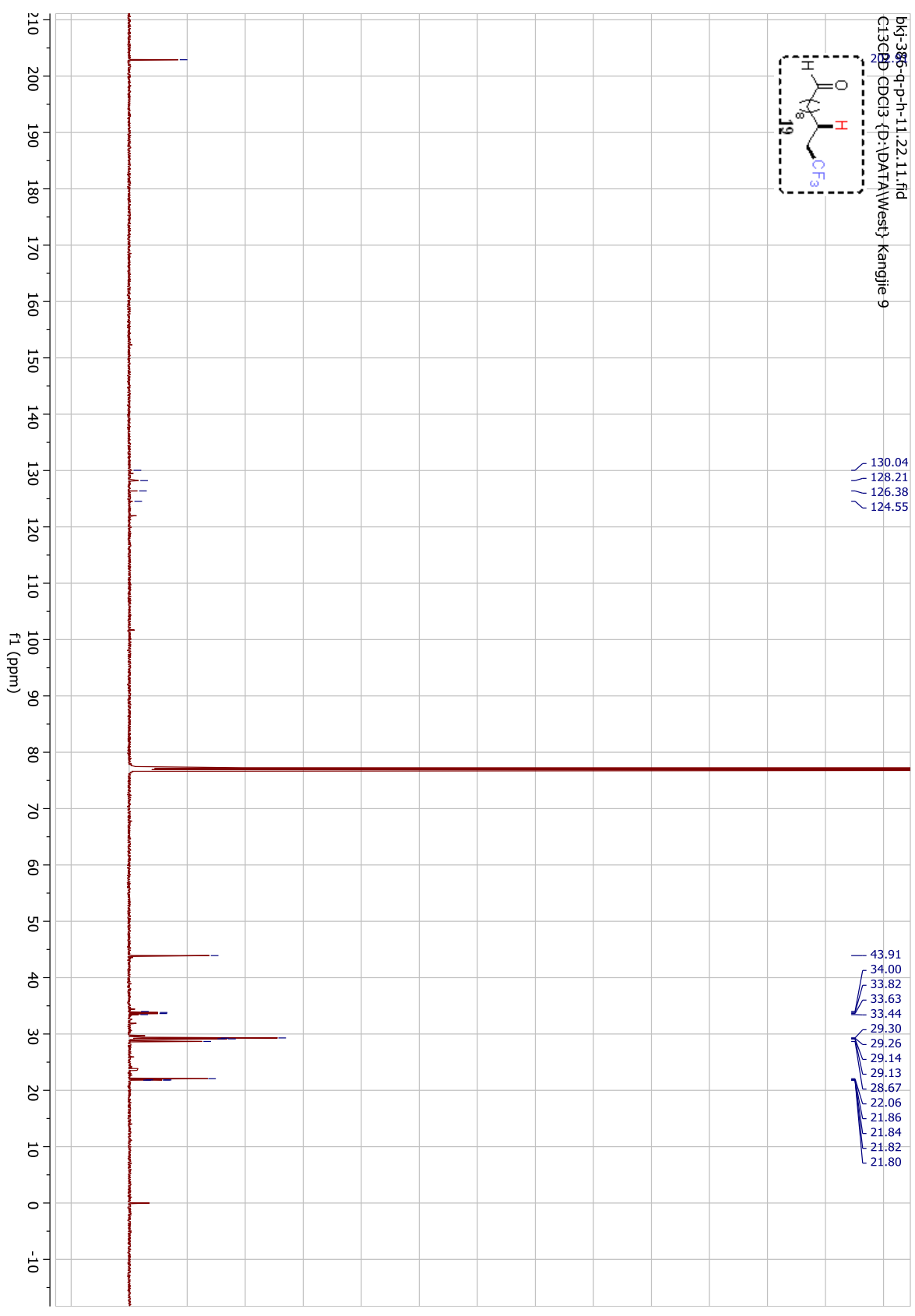
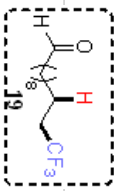
-66.42
-66.44
-66.46



bkj-386-q-p-h-1.22.10.fid
PROTON CDCl₃ {D:\DATA\West\ Kangjie 9



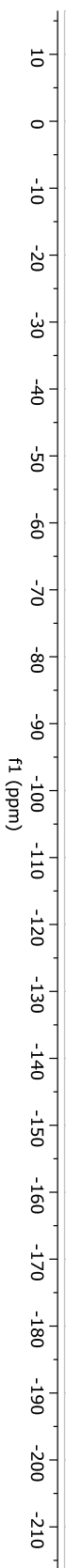
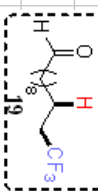
bkj-386-q-p-h-11.22.11.ftd
C130 CDCl3 (D:\DATA\Westj kangjie 9



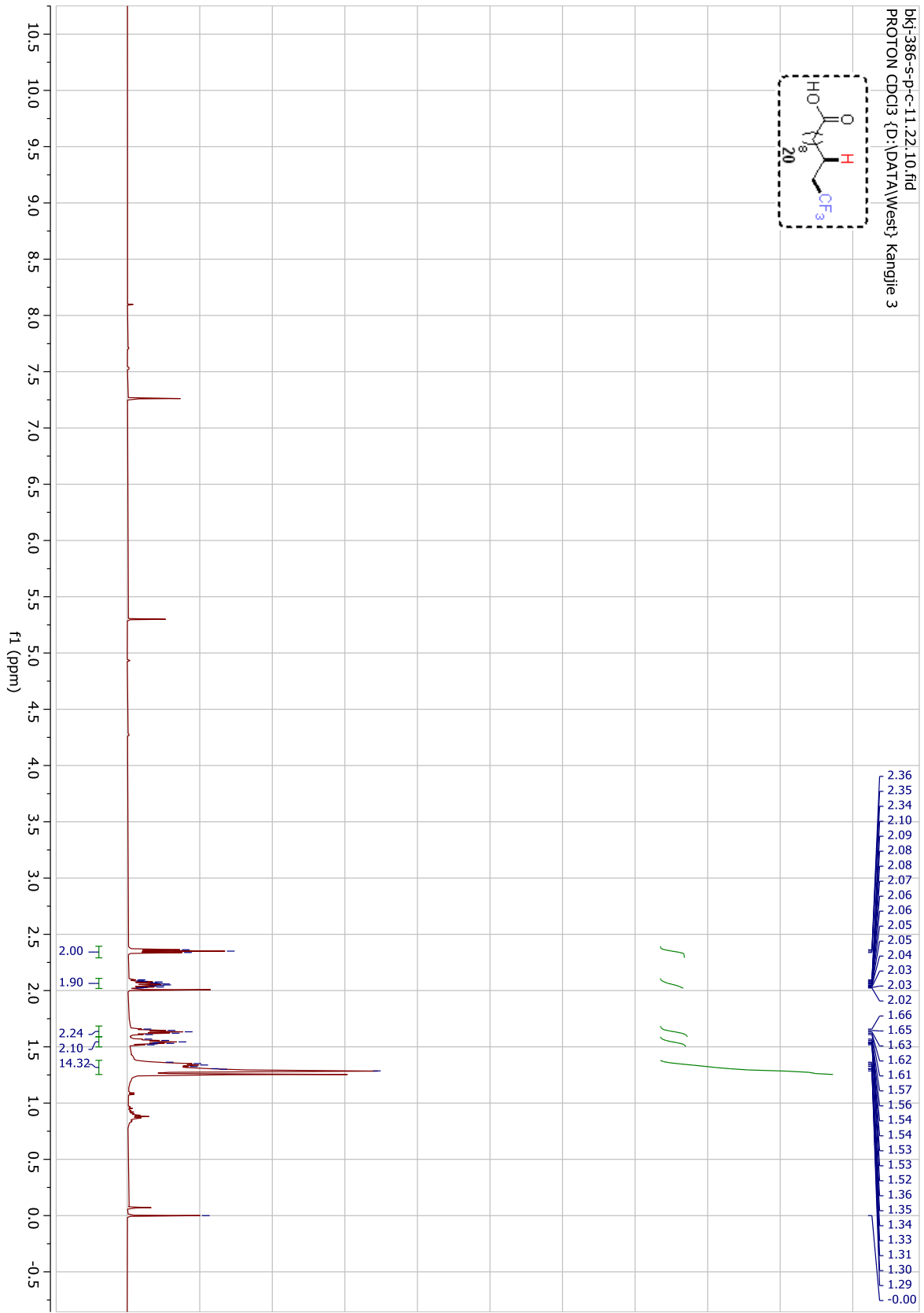
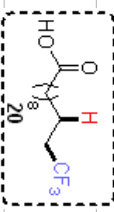
bkj-386-q-p-h-11-22-12.fid
F19_baseline_correct ID

13 {D:\DATA\West\ Kangjie 9

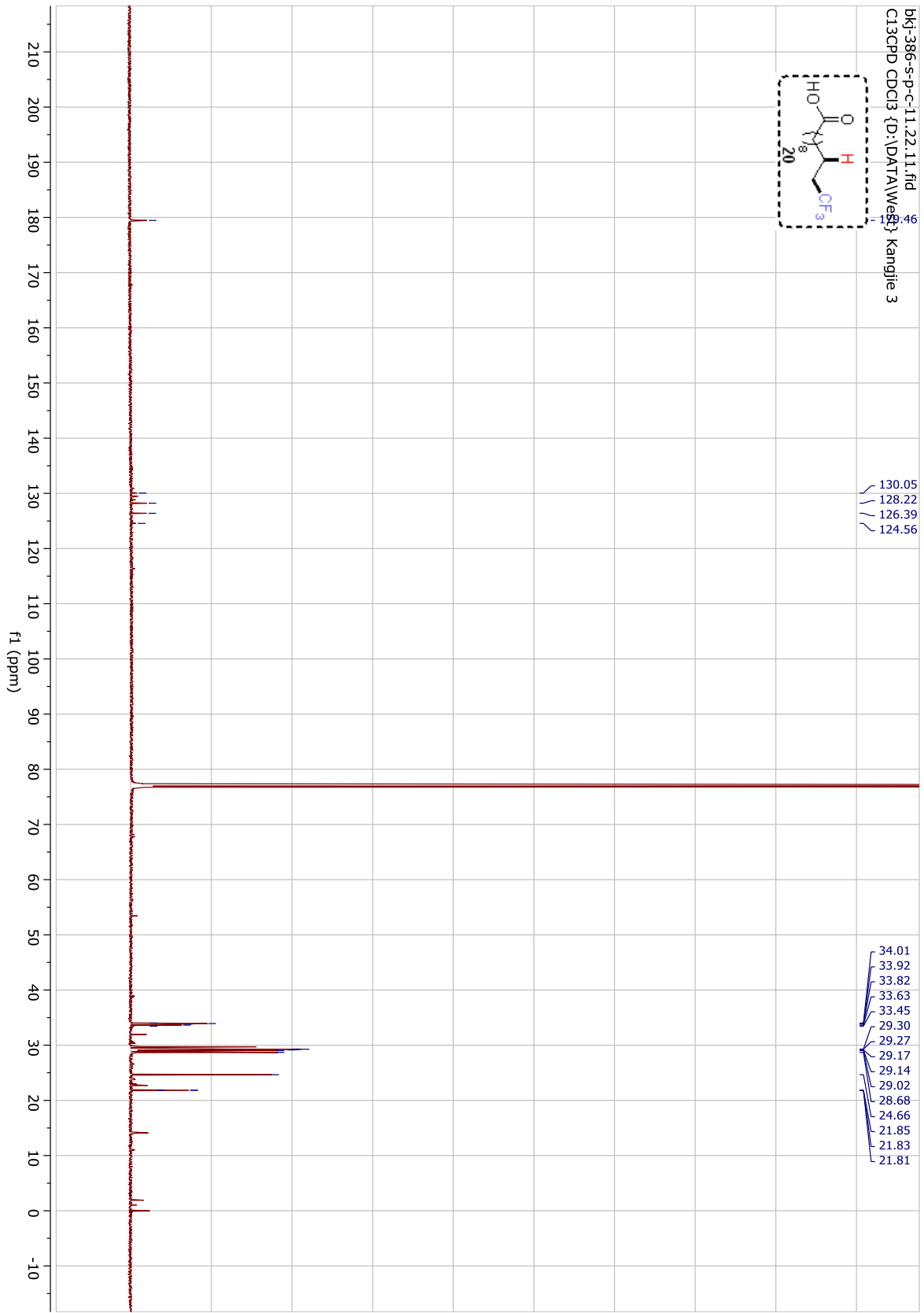
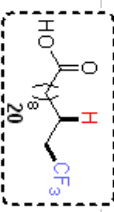
66.39
66.40
66.41
66.42
66.43



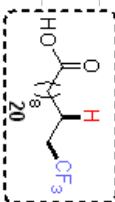
bkj-386-s-p-c-11.22.10.fid
PROTON CDQ3 {D:\DATA\West\ Kangjie 3



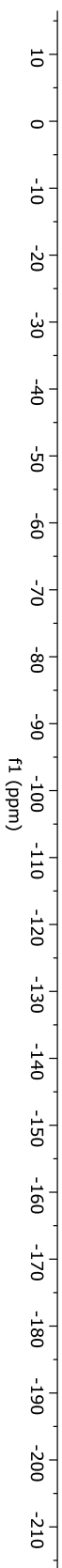
bkj-386-s-p-c-11.22.11.fid
C13CPD CDCl3 (D:\DATA\We...)
Kangjie 3



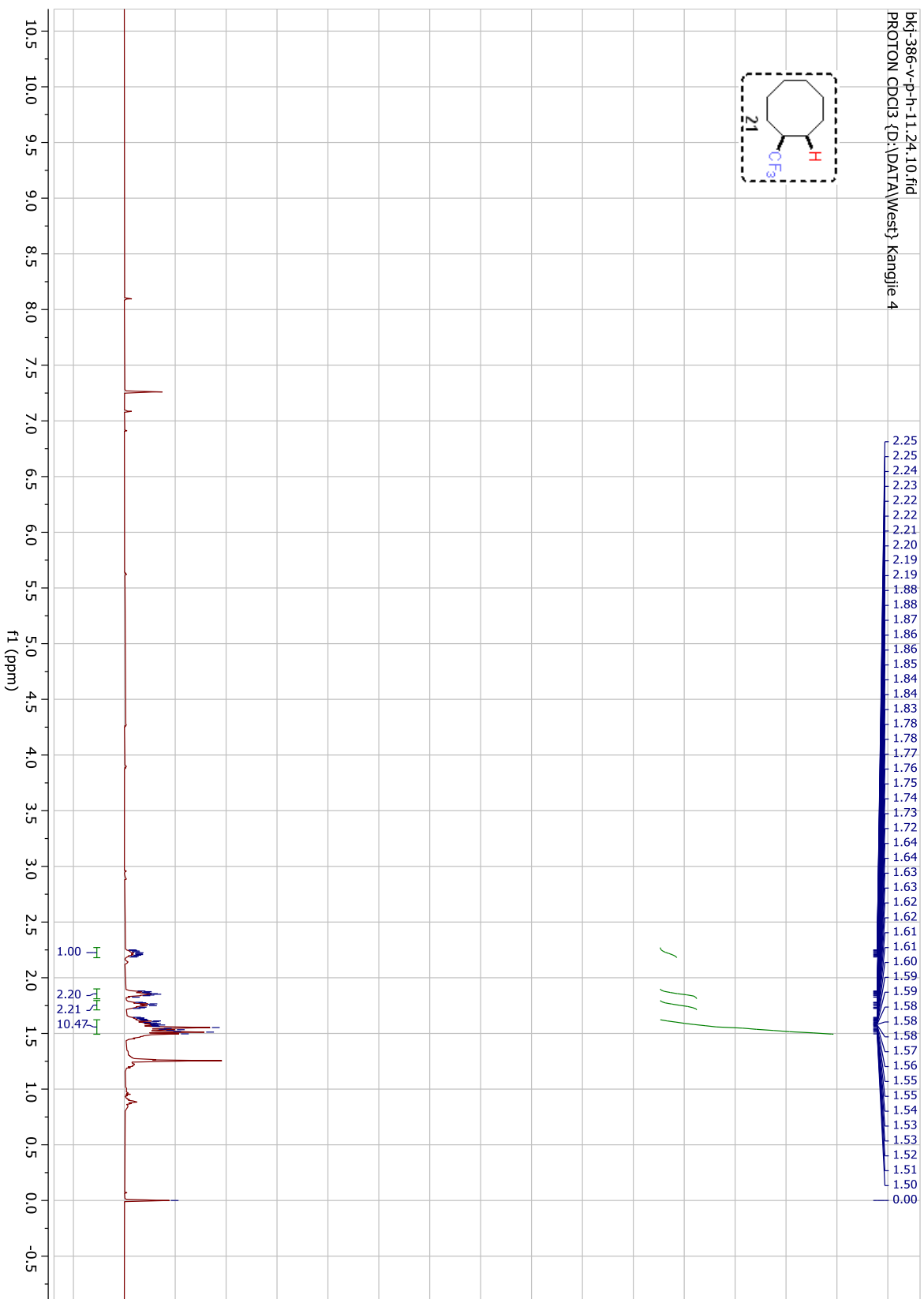
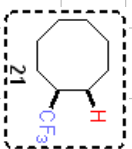
bkj-386-s-p-c-11.22.12.fid
F19_baseline_correct ID
F19_baseline_correct F1913 1D \DATA\West\ Kangjie 3



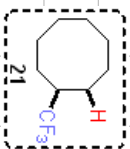
-66.40
-66.42
-66.44



bj1-386-v-p-h-11.24.10.fid
PROTON CDCl3 {D:\DATA\Westf\Kanglie 4

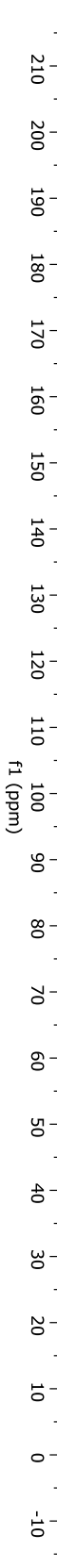


bjf-386-v-p-h-11.24.12.fid
C13CPD CDCl3 (D:\DATA\Westj_Kangjie_4



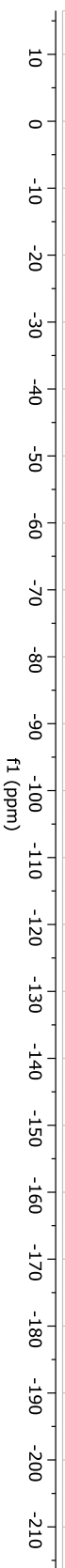
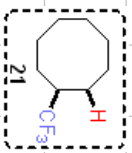
131.75
129.90
128.05
126.19

42.16
42.00
41.84
41.68
26.38
26.29
25.59
25.58
25.56
25.54
25.11

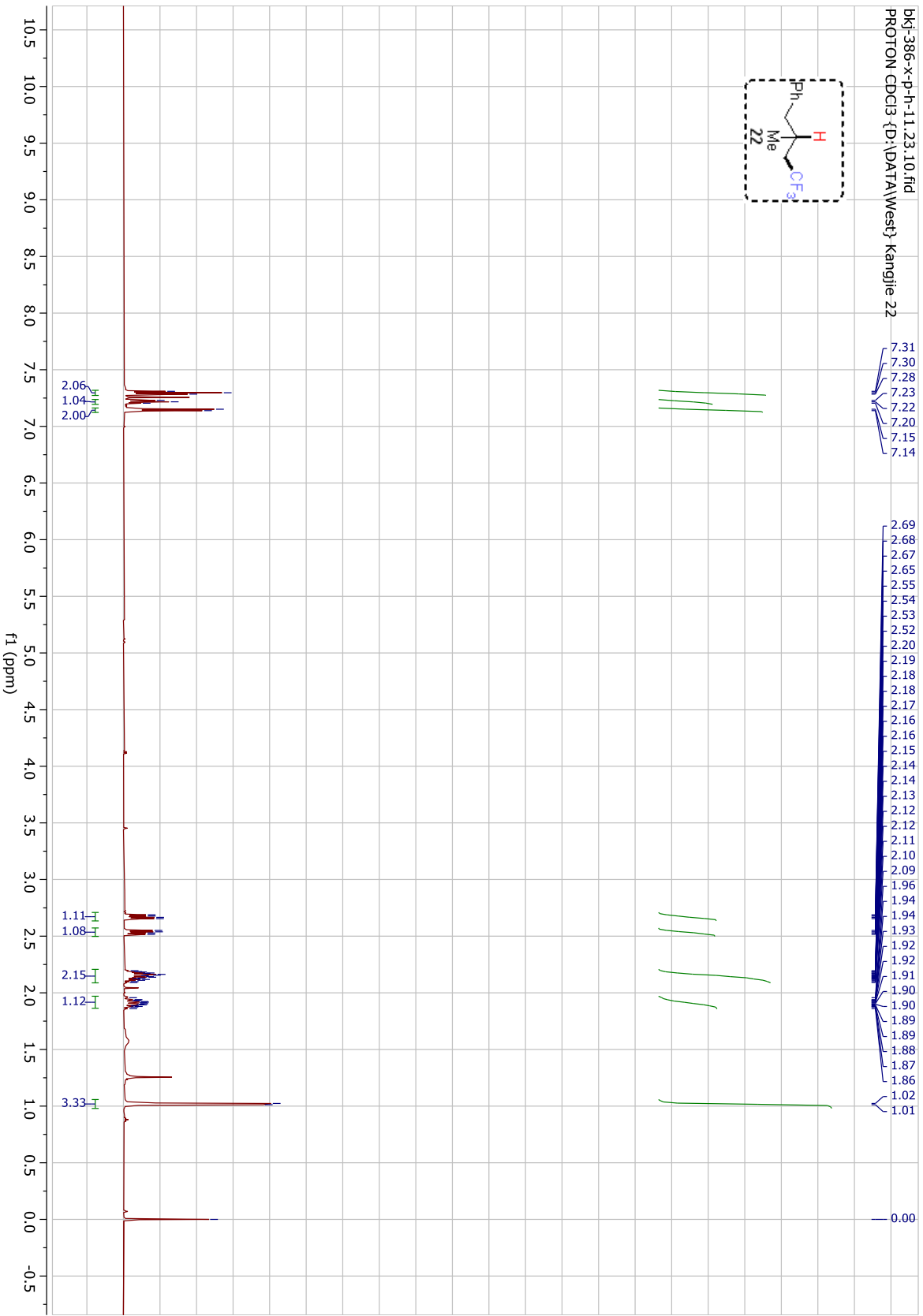
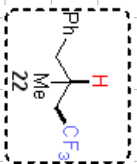


bkj-386-v-p-h-11.24.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 4

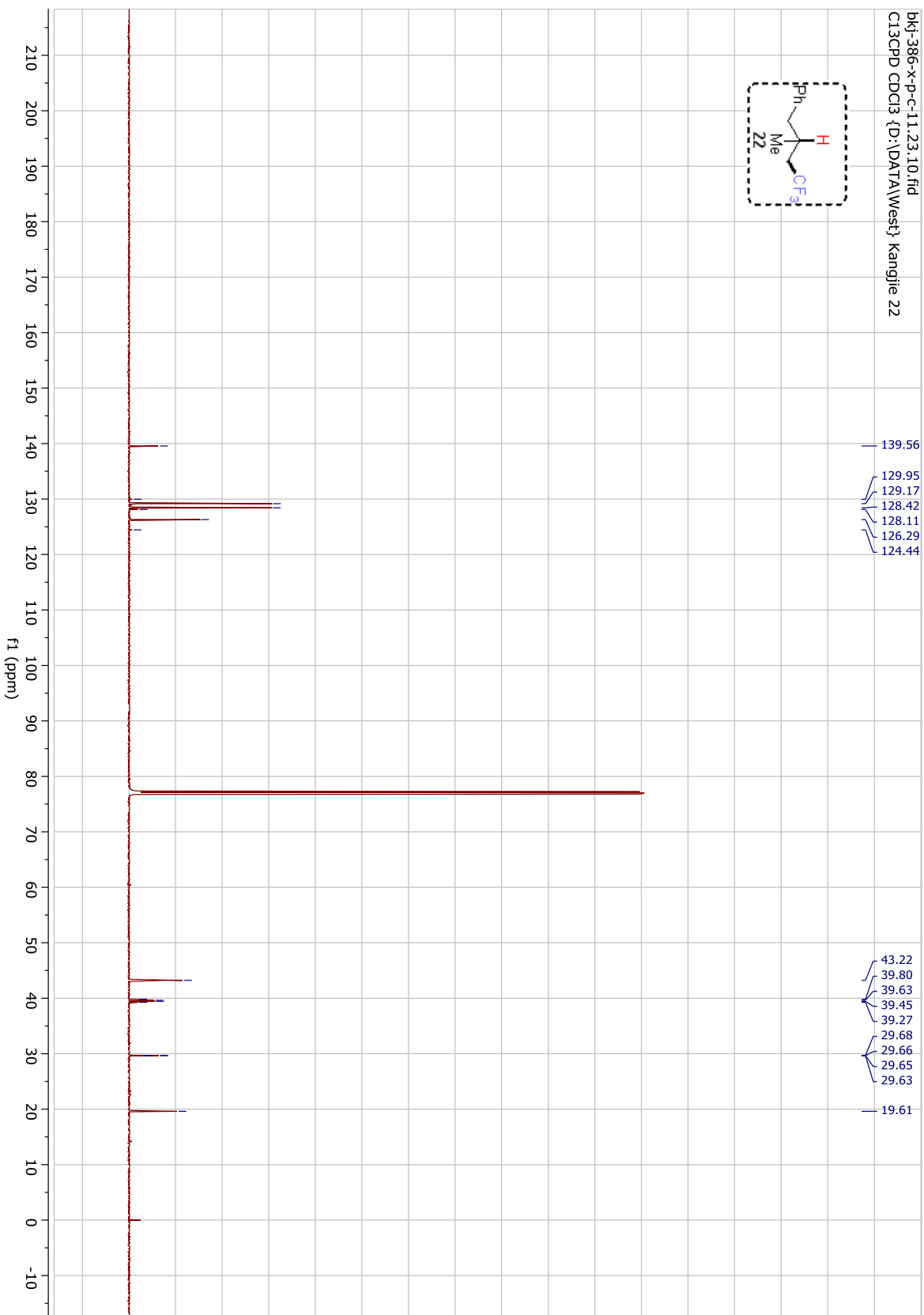
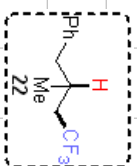
-73.33
-73.35



bk1-386-x-p-h-11.23.10.fid
PROTON CDG3 {D:\DATA\Westfj_Kangjie 22

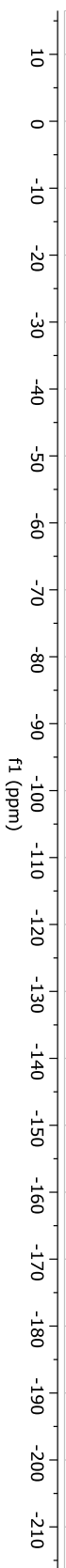
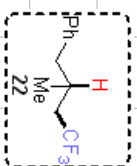


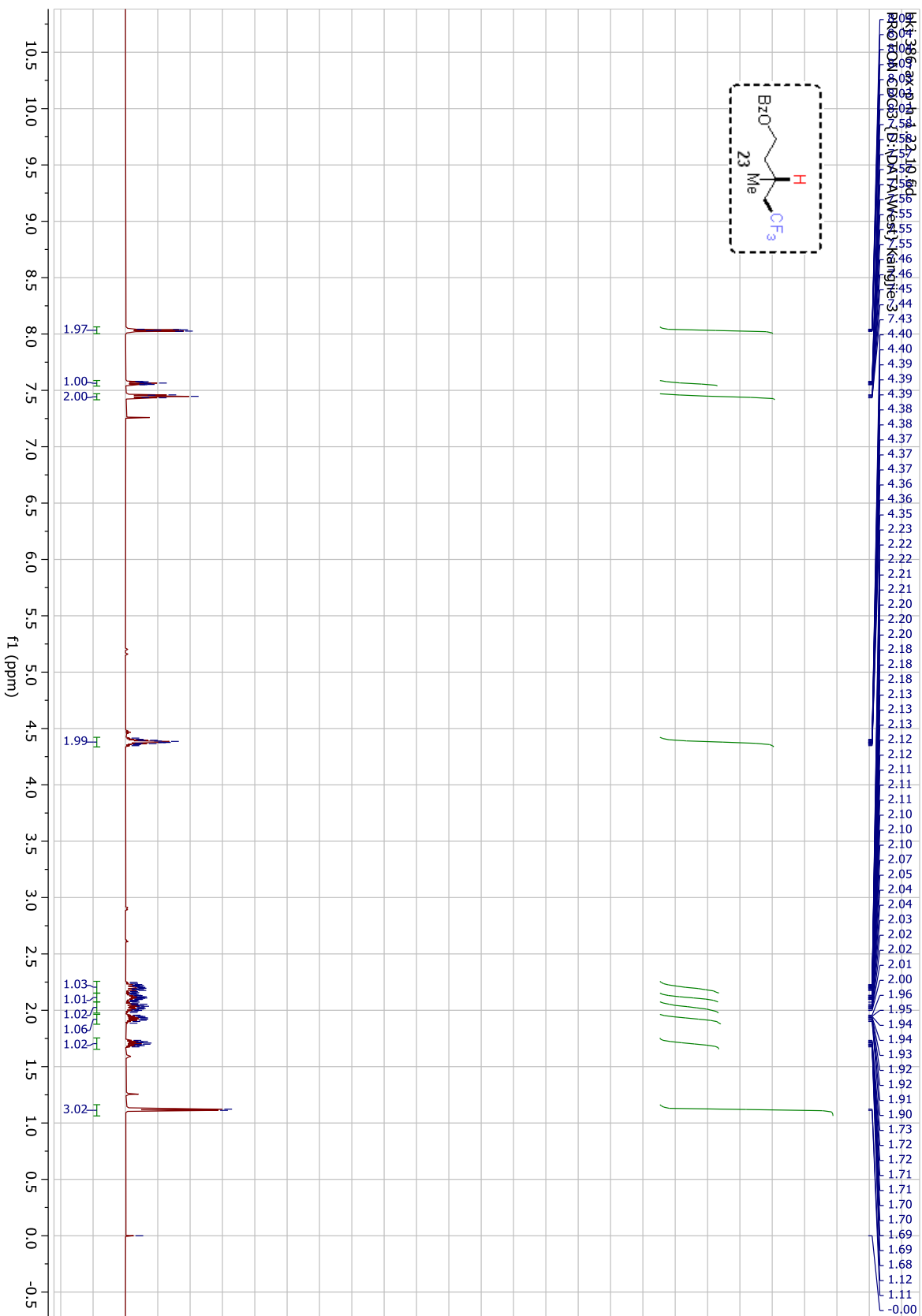
bjf-386-x-p-c-11.23.10.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 22



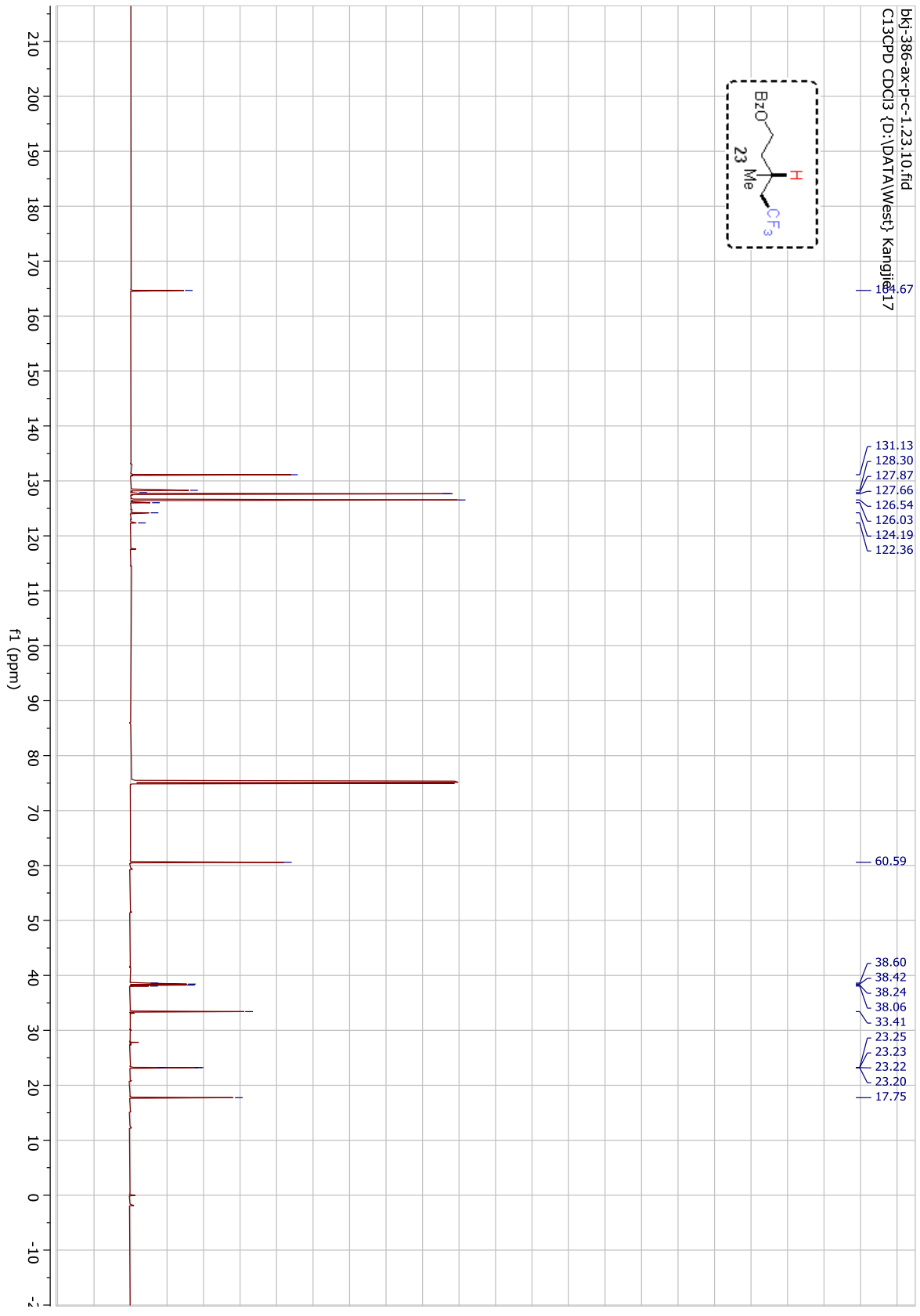
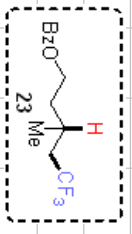
bkj-386-x-p-f-11.23.10.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 22

63.09
63.11
63.13



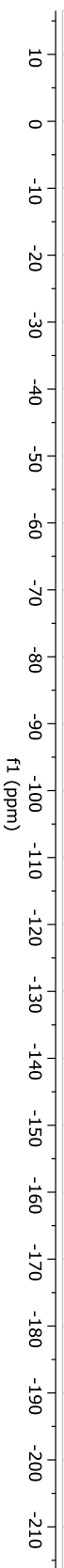
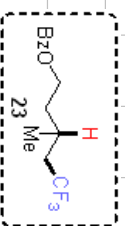


bkj-386-ax-p-c-1.23.10.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjil) 17

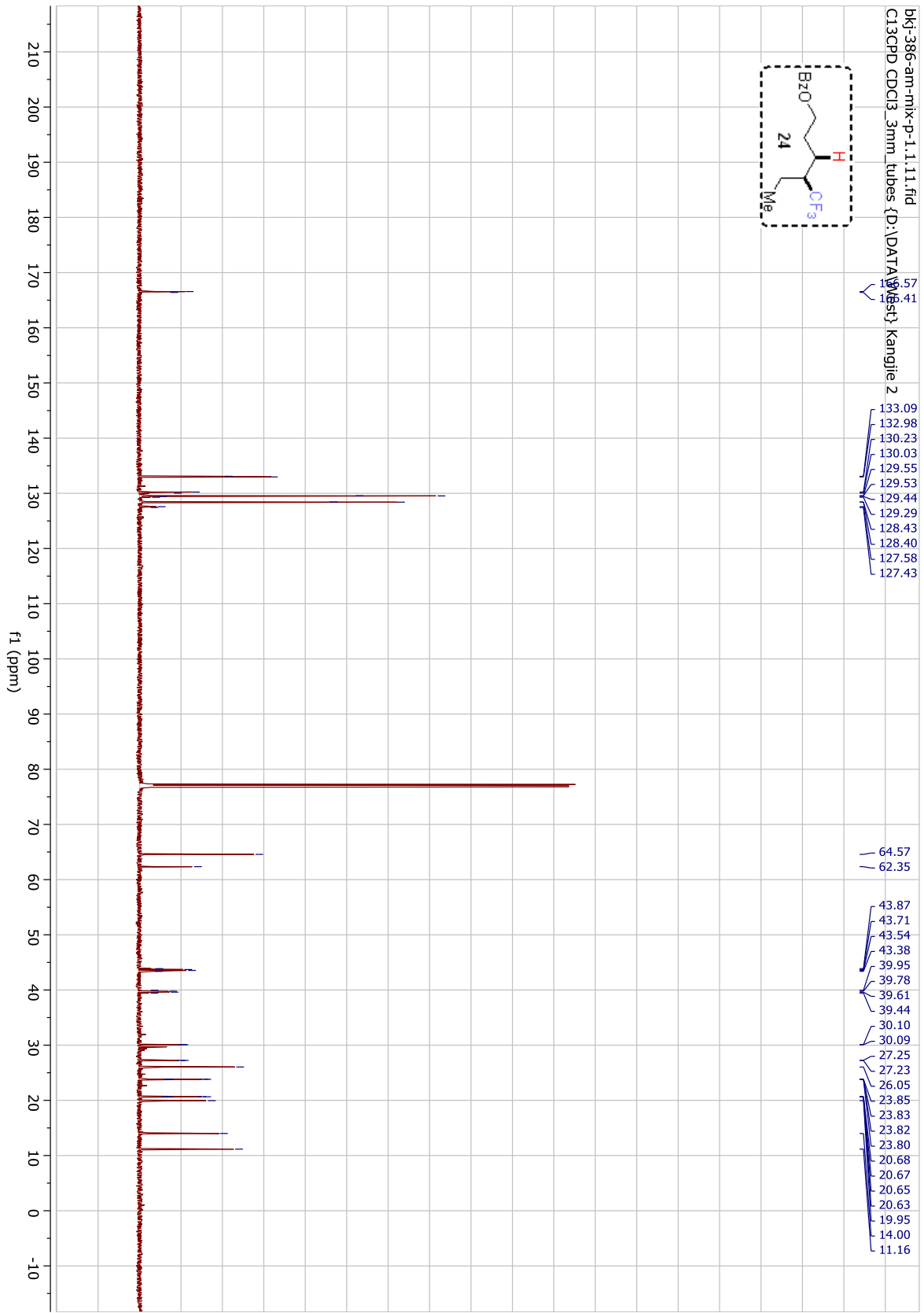
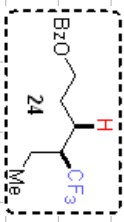


bkt-386-ax-p-h-1.22.11.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 3

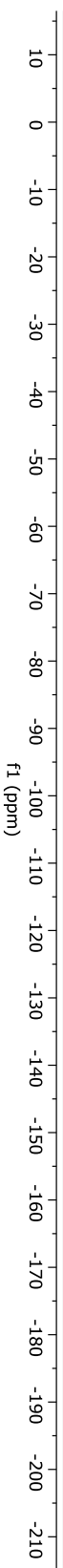
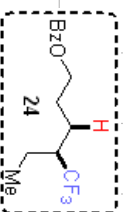
63.28
63.30
63.32

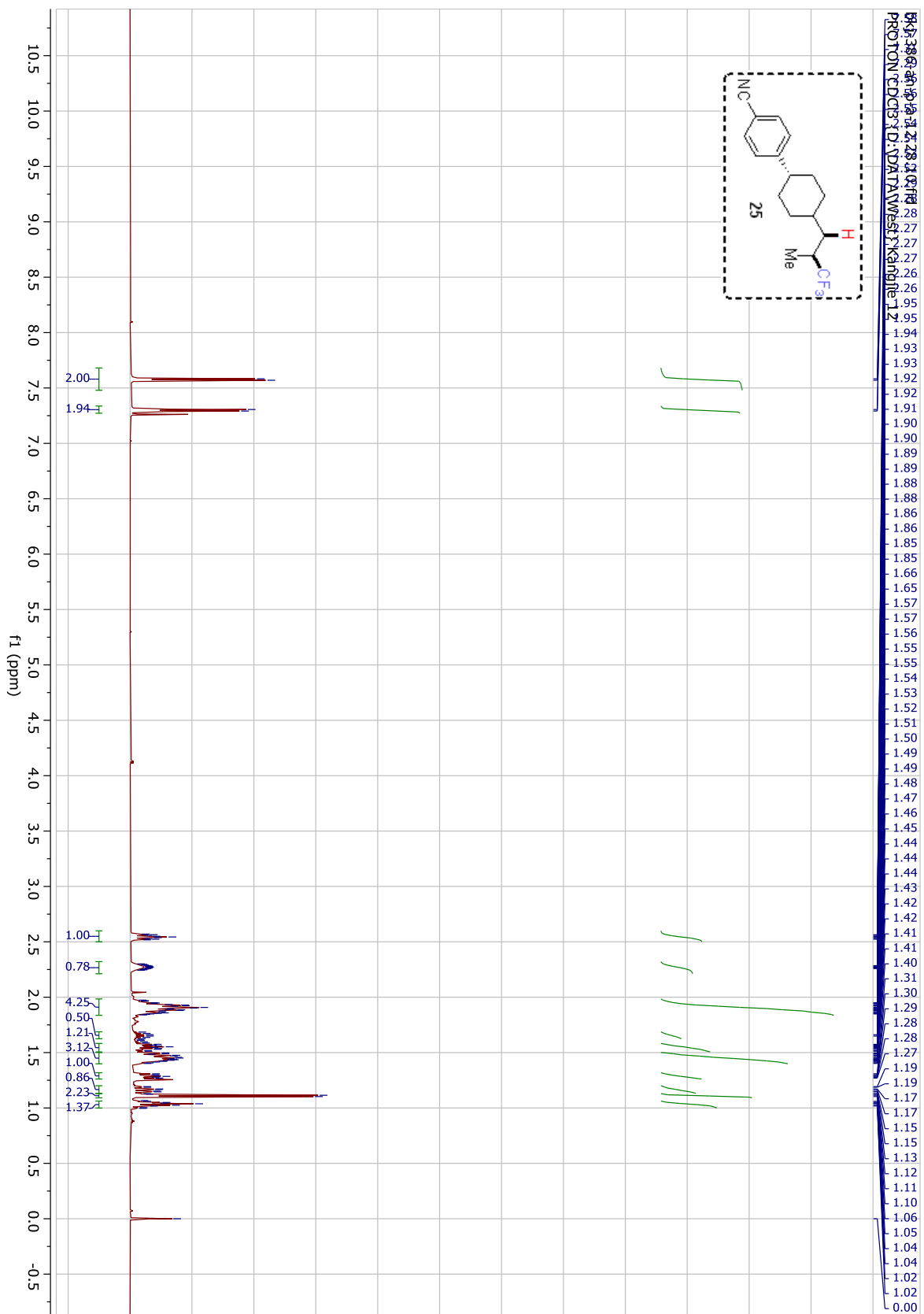


hkl-386-am-mix-p-1.1.11.fid
C13CPD CDCl3_3mm_tubes {D:\DATA\MSF\Kangjie 2

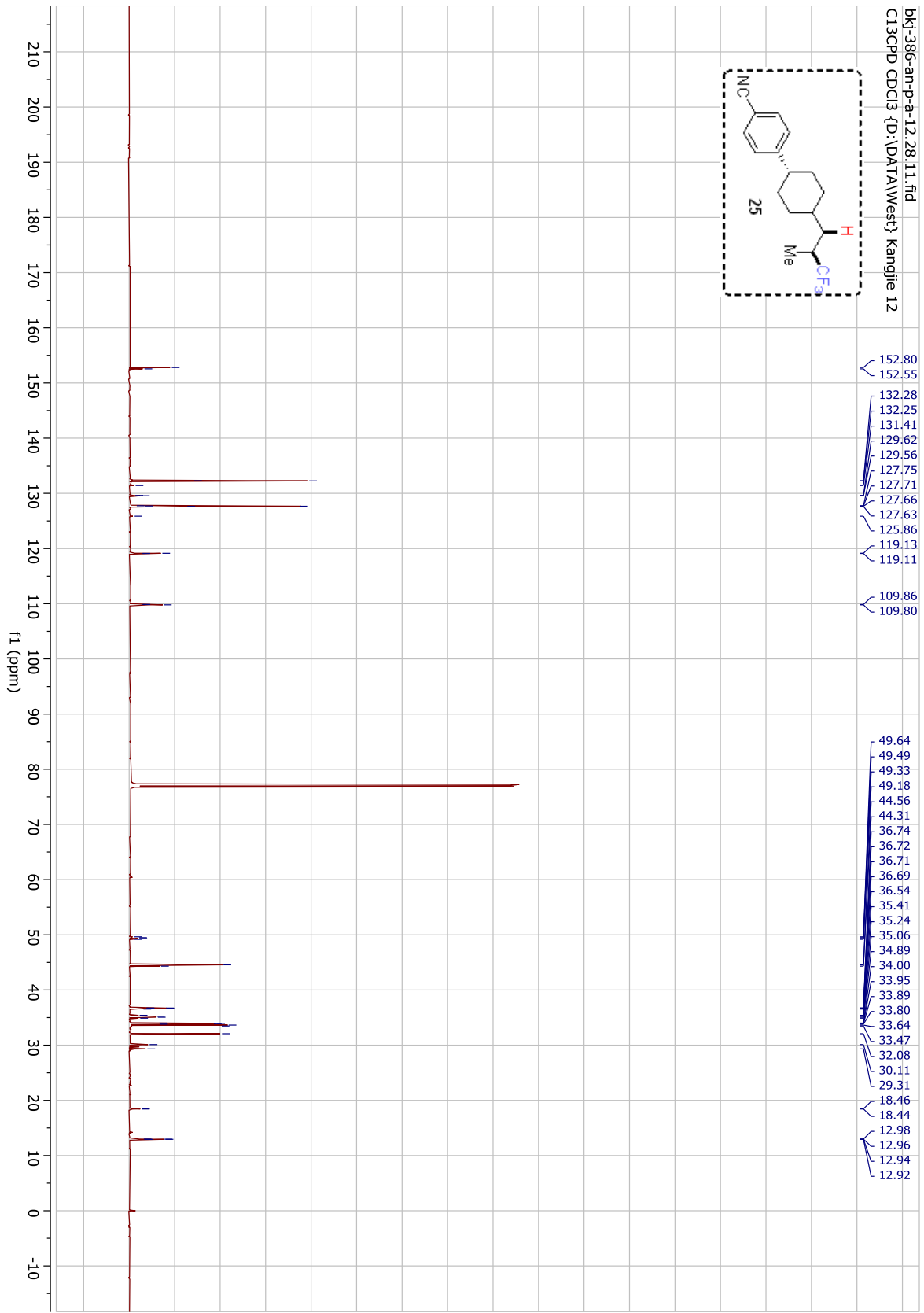
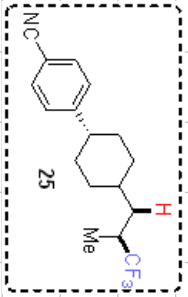


bkj-386-am-mix-p-1.1.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3_3mm_tubes {D:\DATA\West} Kangjie
-69.84
-69.85
-70.49
-70.51

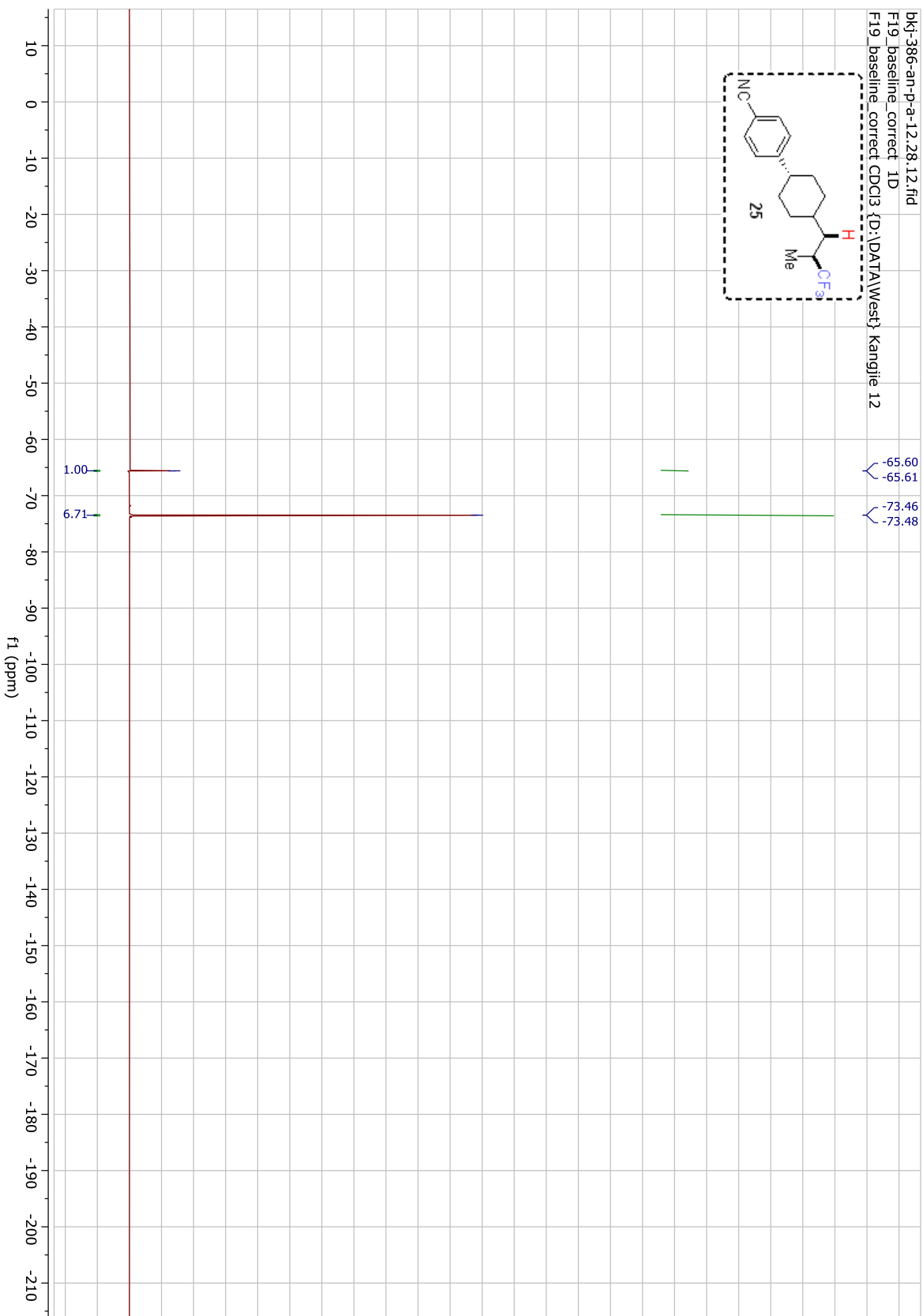
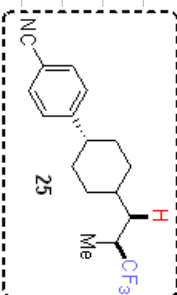


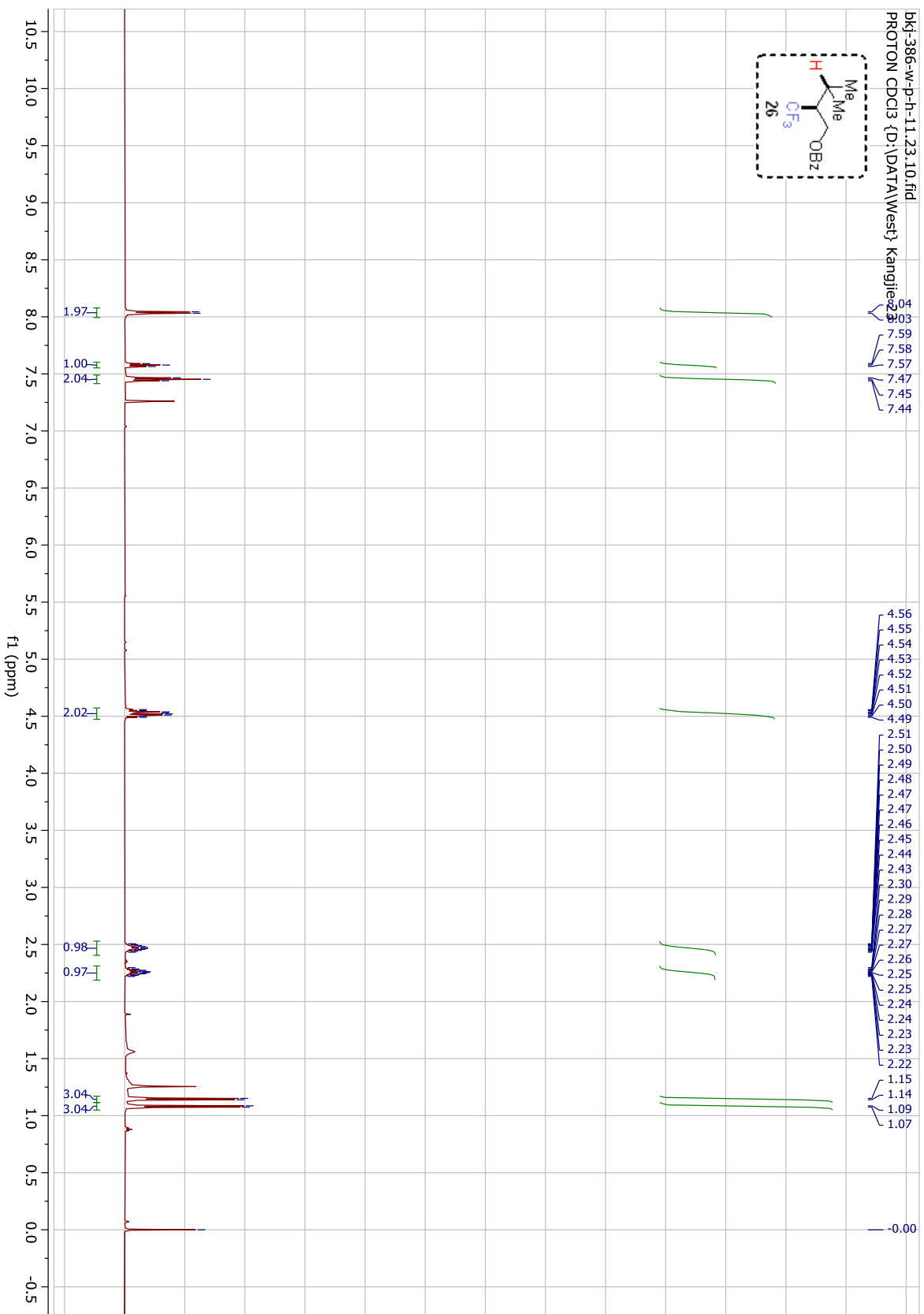


bjf-386-an-p-a-12.28.11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjle 12

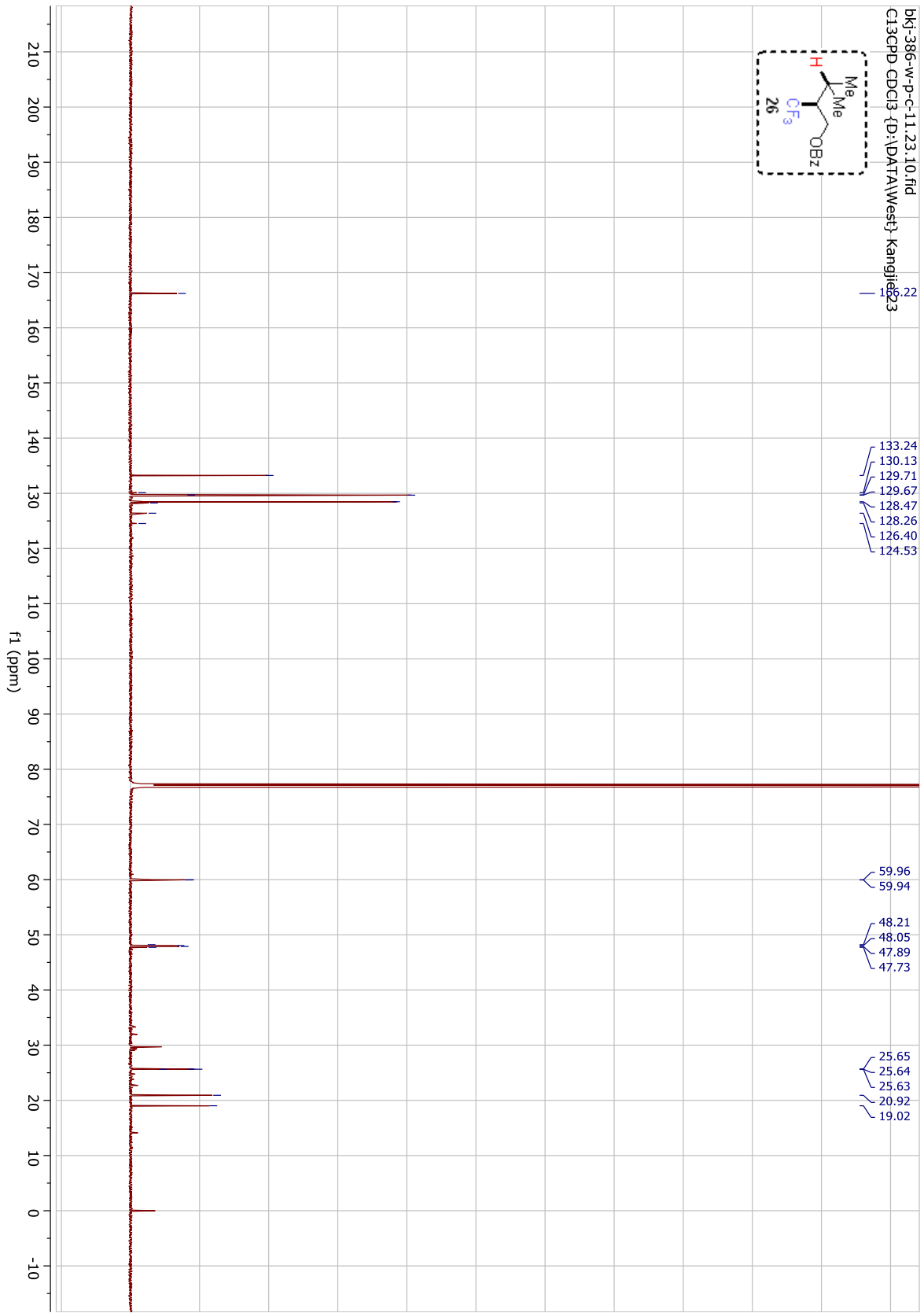
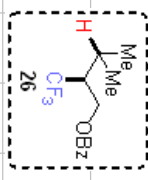


bkj-386-an-p-a-12.28.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 12



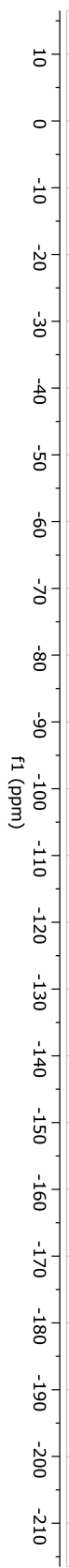
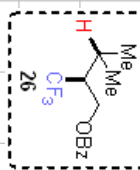


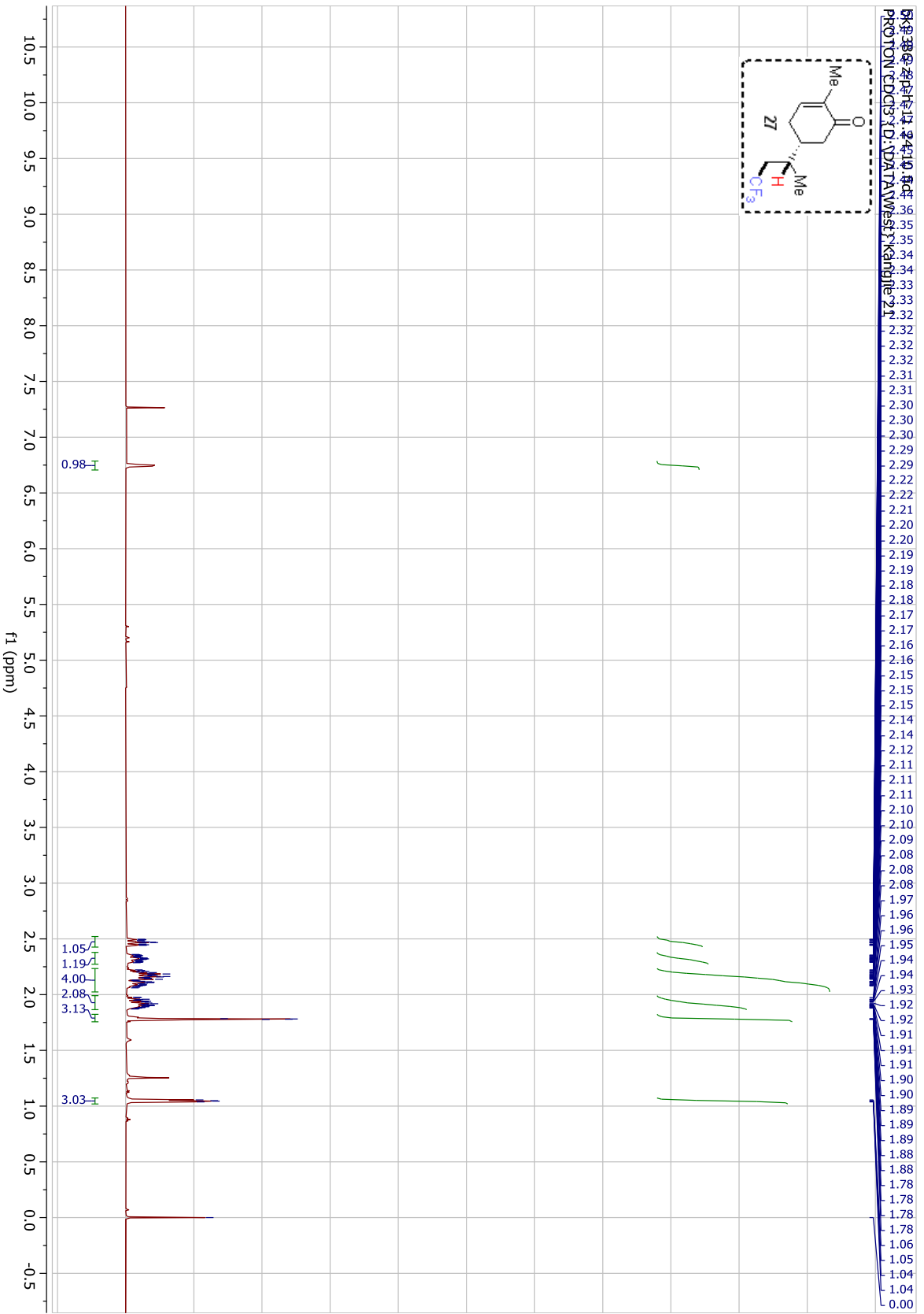
bjf-386-w-p-c-11.23.10.fid
C13CPD CDCl3 (D:\DATAWestfj Kangjief23



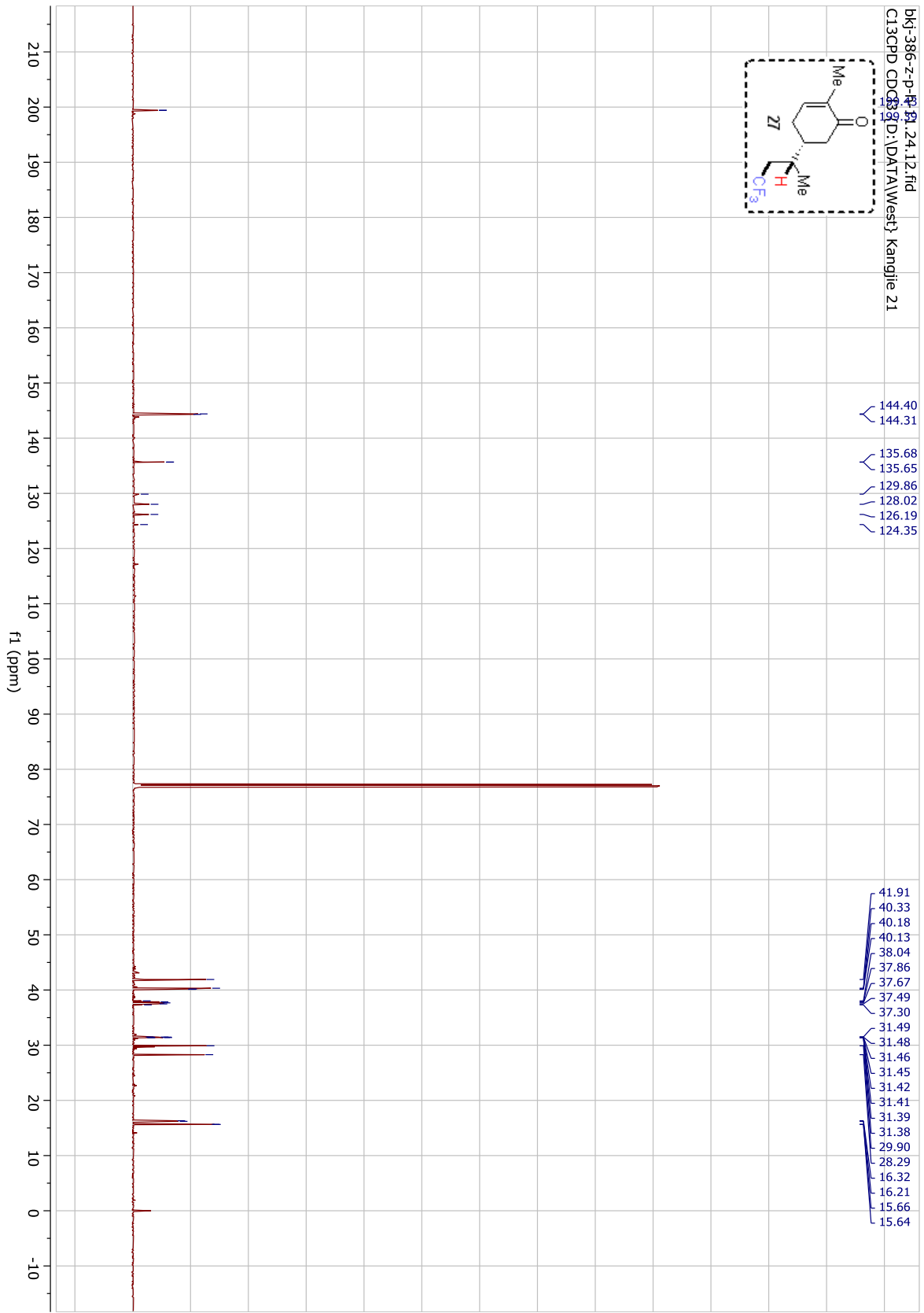
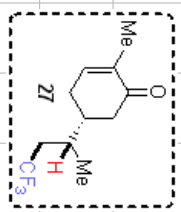
bkj-386-w-p-f-11.23.10.ftd
F19_baseline_correct ID
F19_baseline_correct FIDC13 {D:\DATA\West} Kangjie 23

-65.98
-65.99





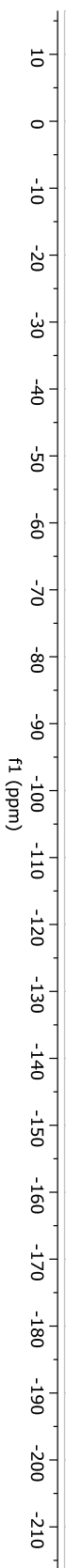
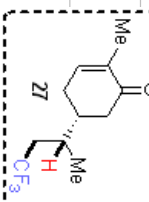
hkl-386-z-p-11-24-12.fid
C13CPD CDCl3 24.12.12



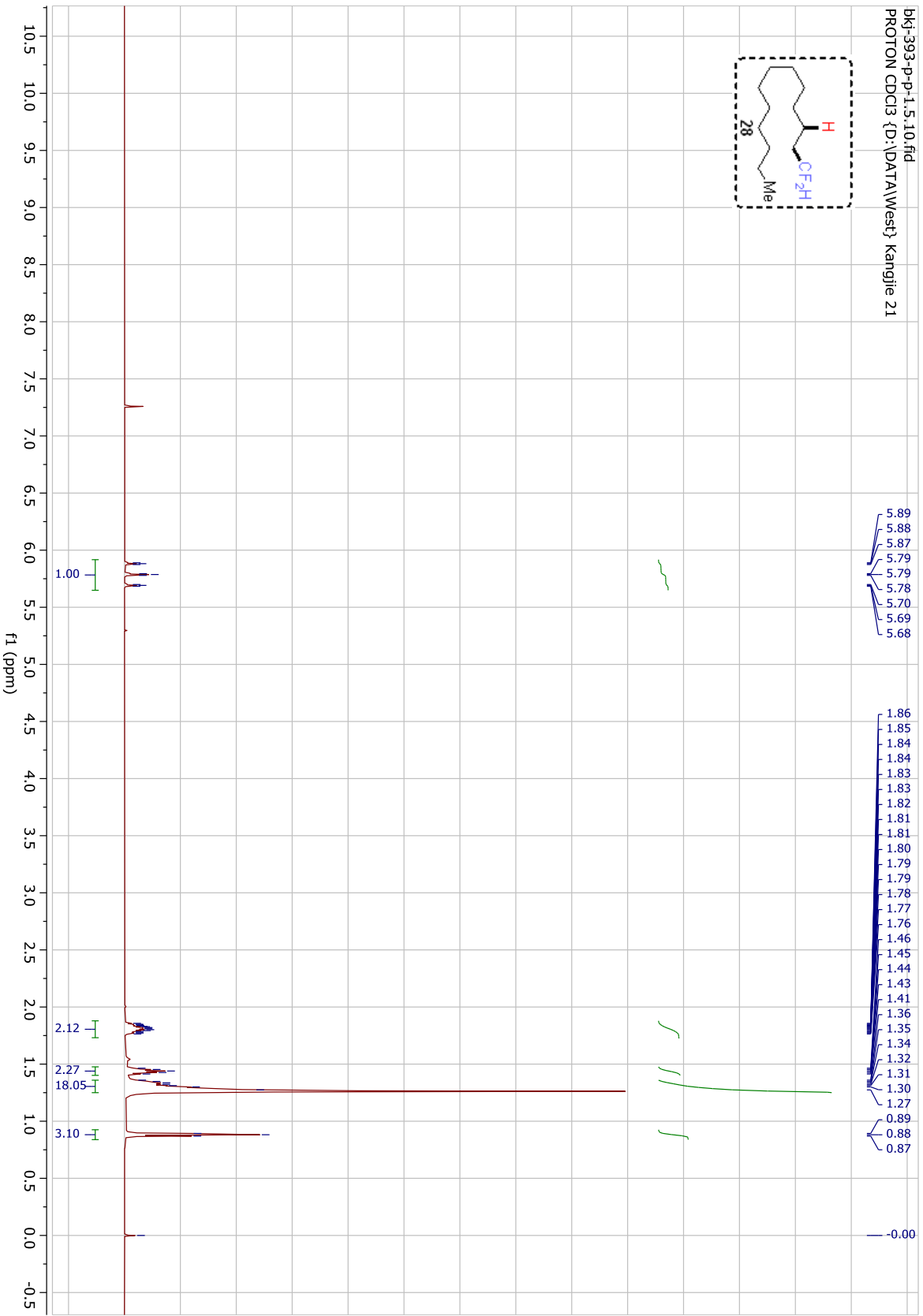
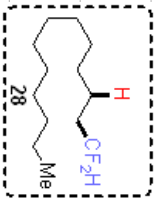
bkj-386-z-p-h-11.24.11.fid
F19_baseline_correct ID

D:\DATA\West\ Kangjie 21

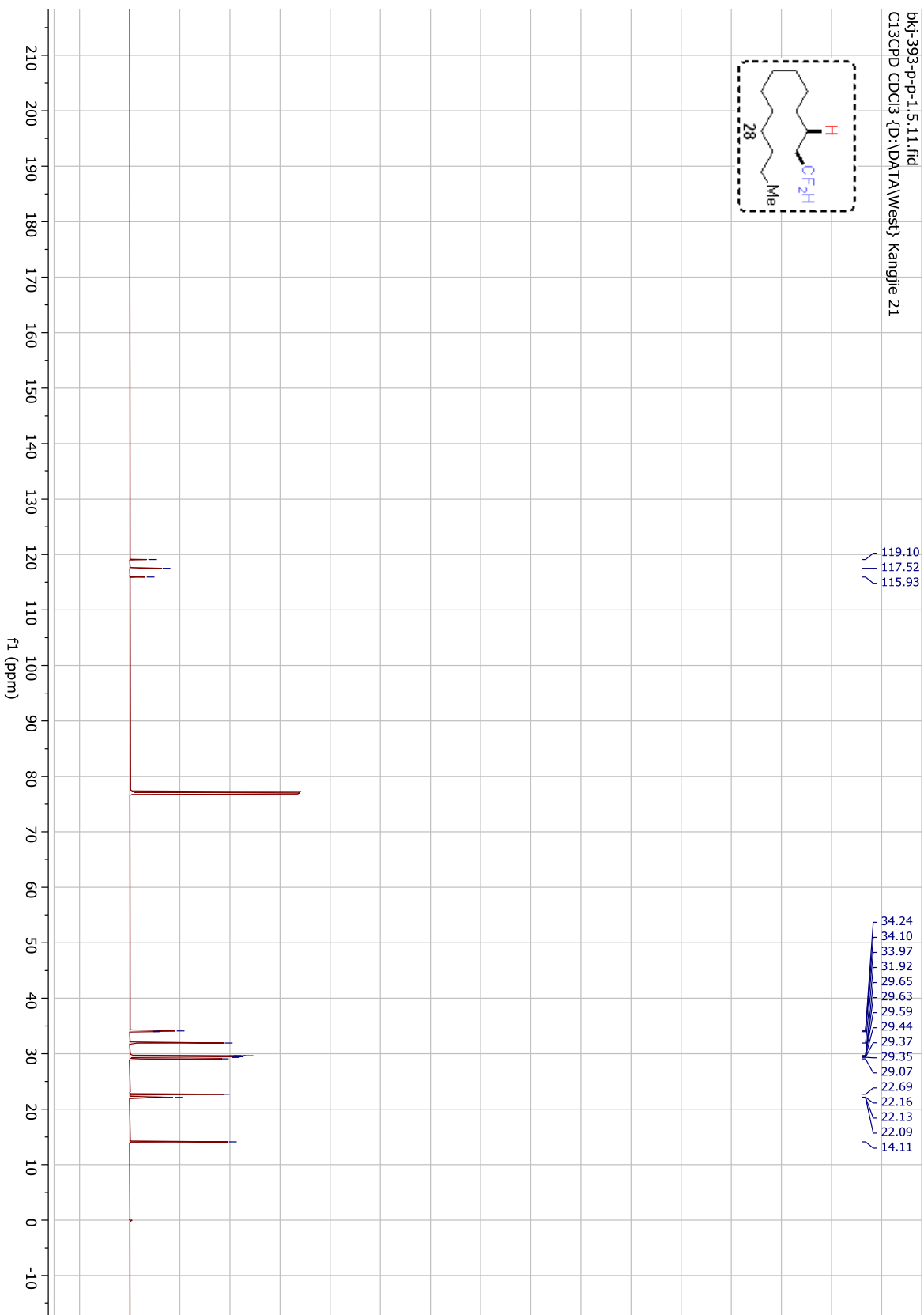
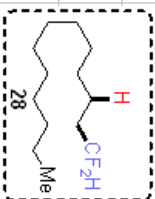
63.51
63.52
63.52
63.54
63.54
63.55



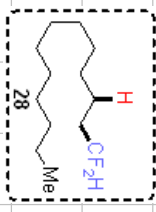
bk1-393-p-p-1.5.10.fid
PROTON CDCl3 {D:\DATA\West\Kangjie 21



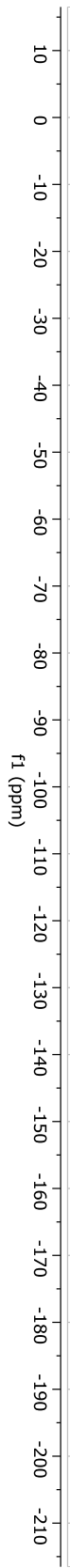
bkt-393-p-p-1,5,11.fid
C13CPD CDCl3 (D:\DATAWest\ Kangjie 21



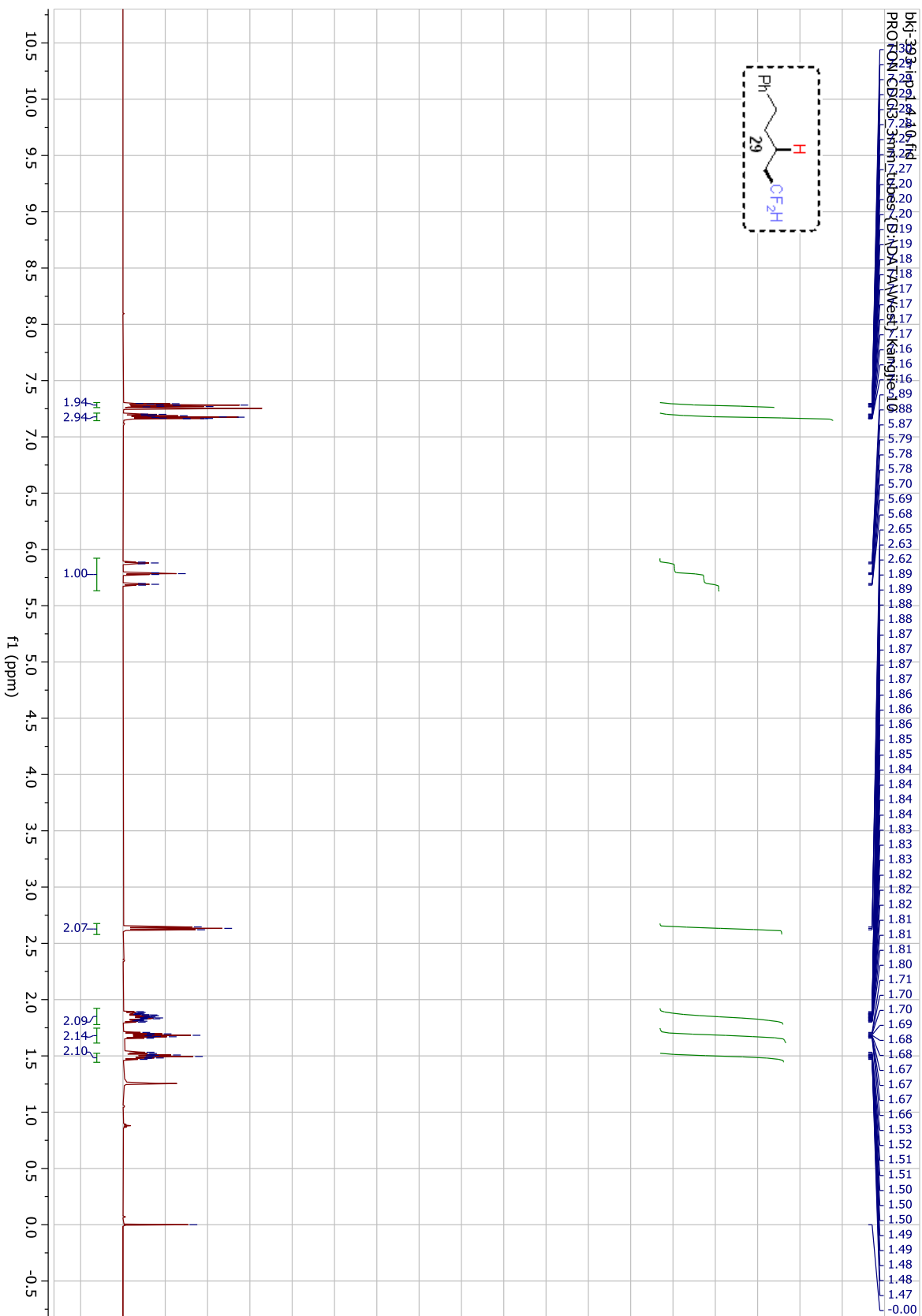
bkj-393-p-p-1.5.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 21



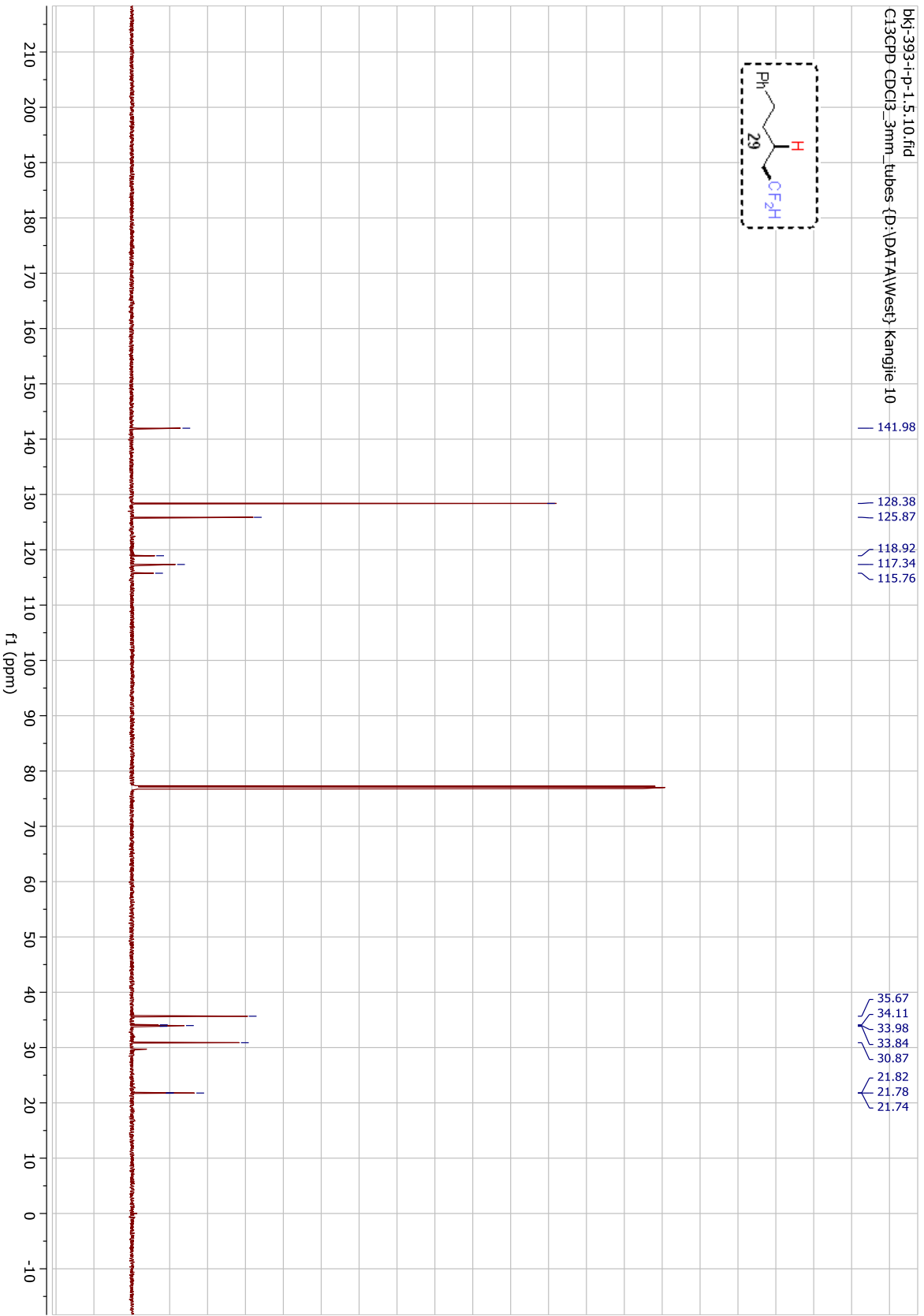
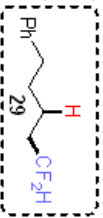
-115.65
-115.68
-115.71
-115.75
-115.78
-115.81



160

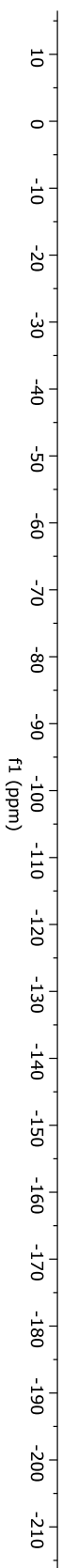
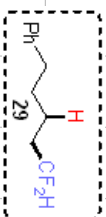


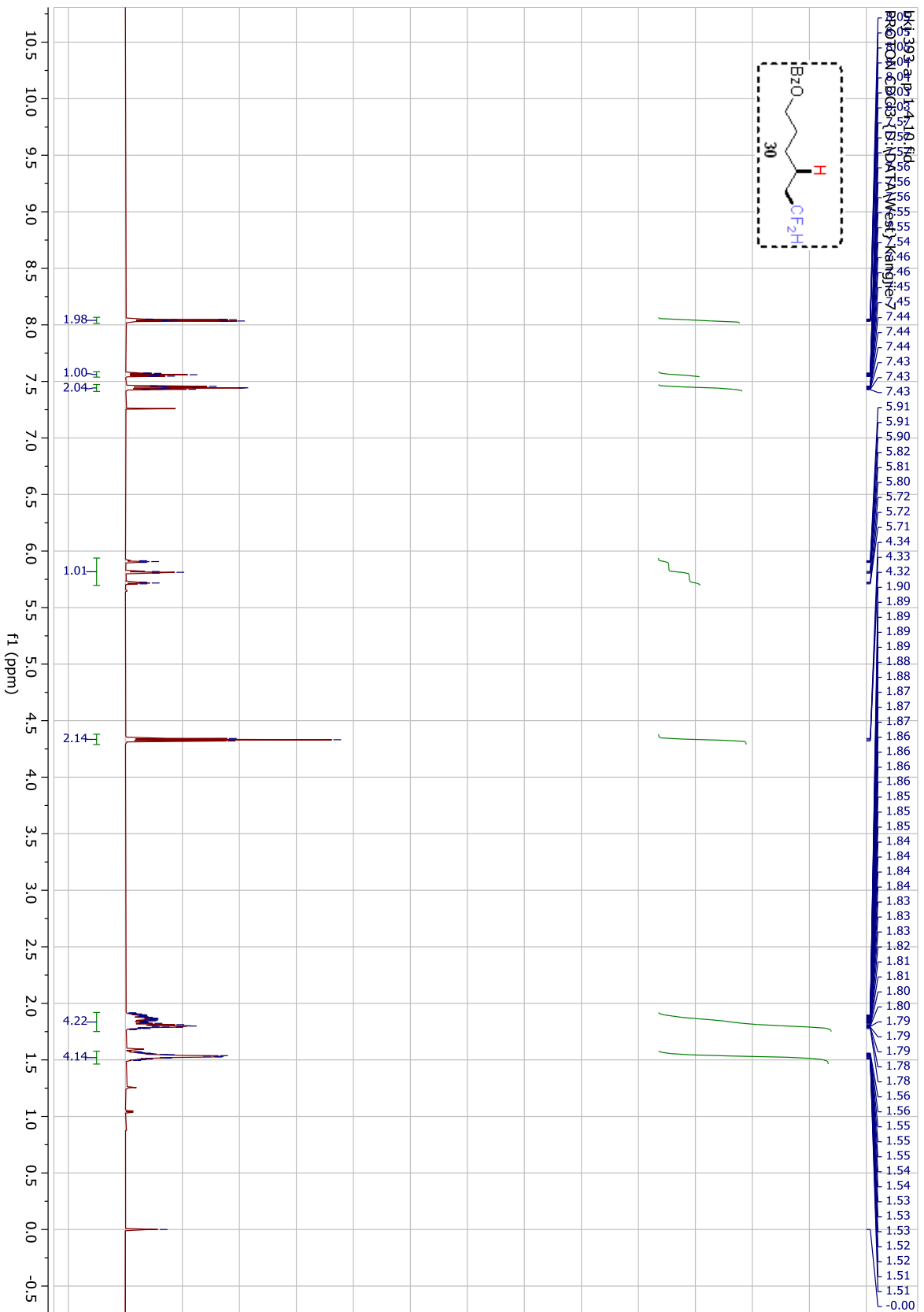
bk1-393-i-p-1.5.10.fid
C13CPD CDCl3_3mm_tubes {D:\DATA\Westf\Kangjie 10



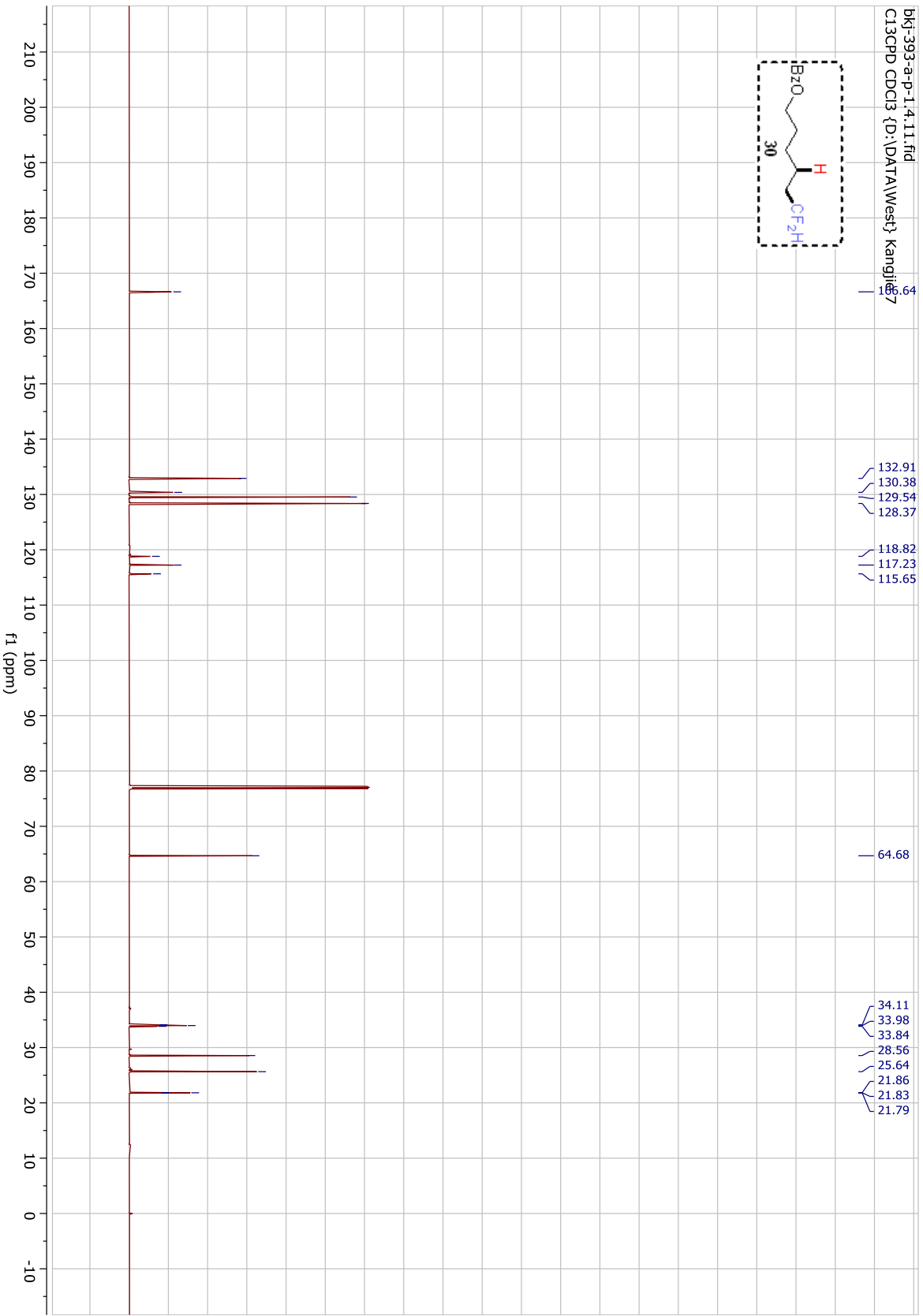
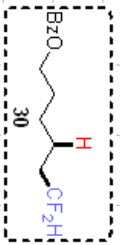
bkj-393-i-p-1.4.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3_3mm_tubes {D:\DATA\West\ Kangjie 10

-115.64
-115.67
-115.70
-115.74
-115.77
-115.80

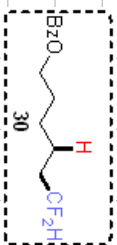




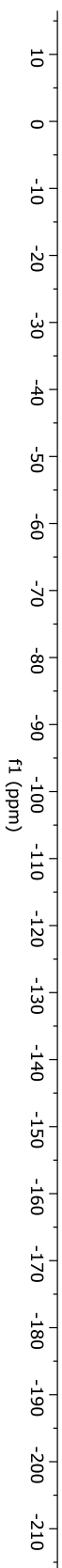
bkt-393-a-p-1,4,11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangji)



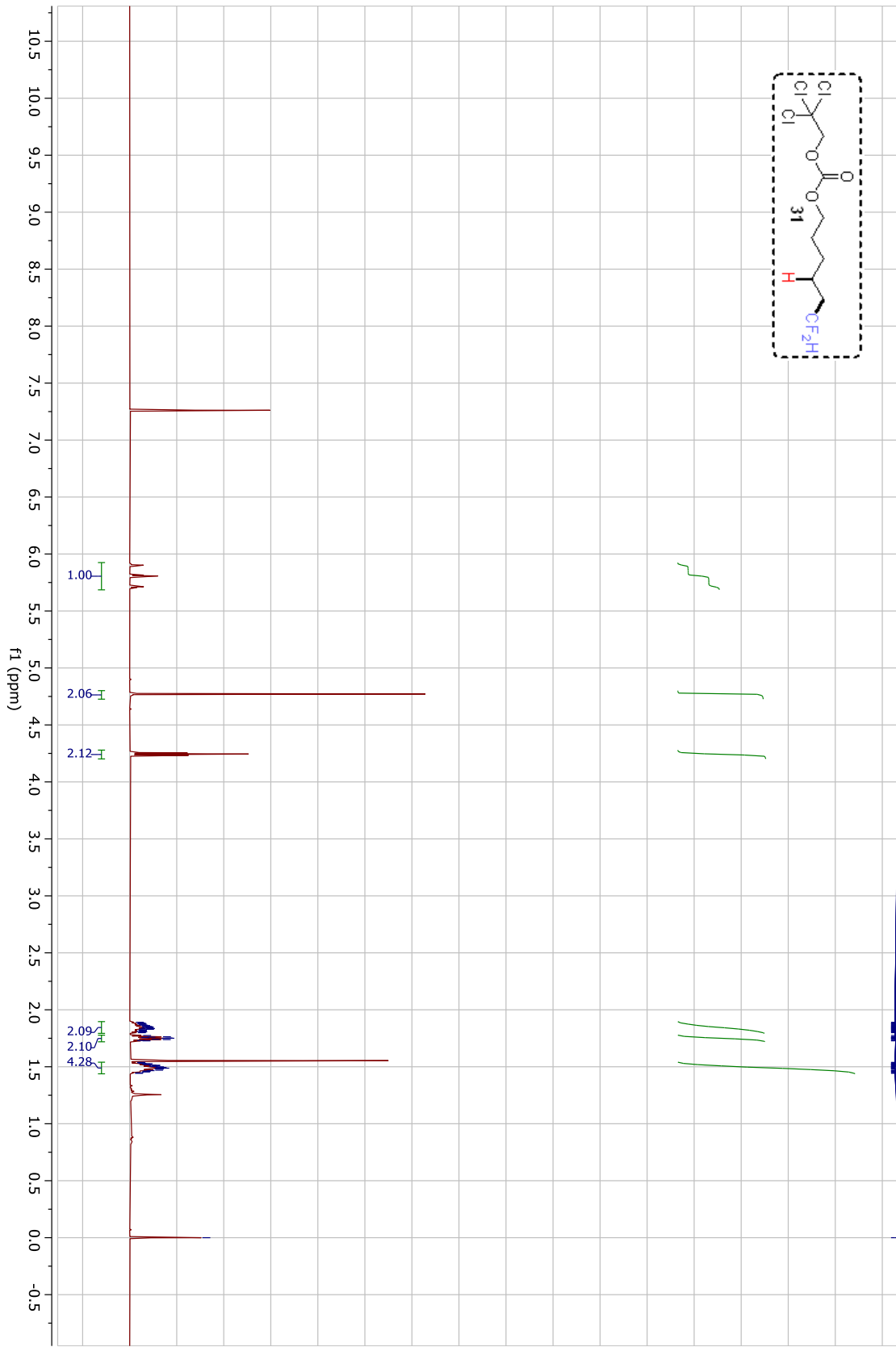
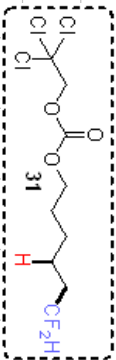
bkj-393-a-p-1-4-12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 7



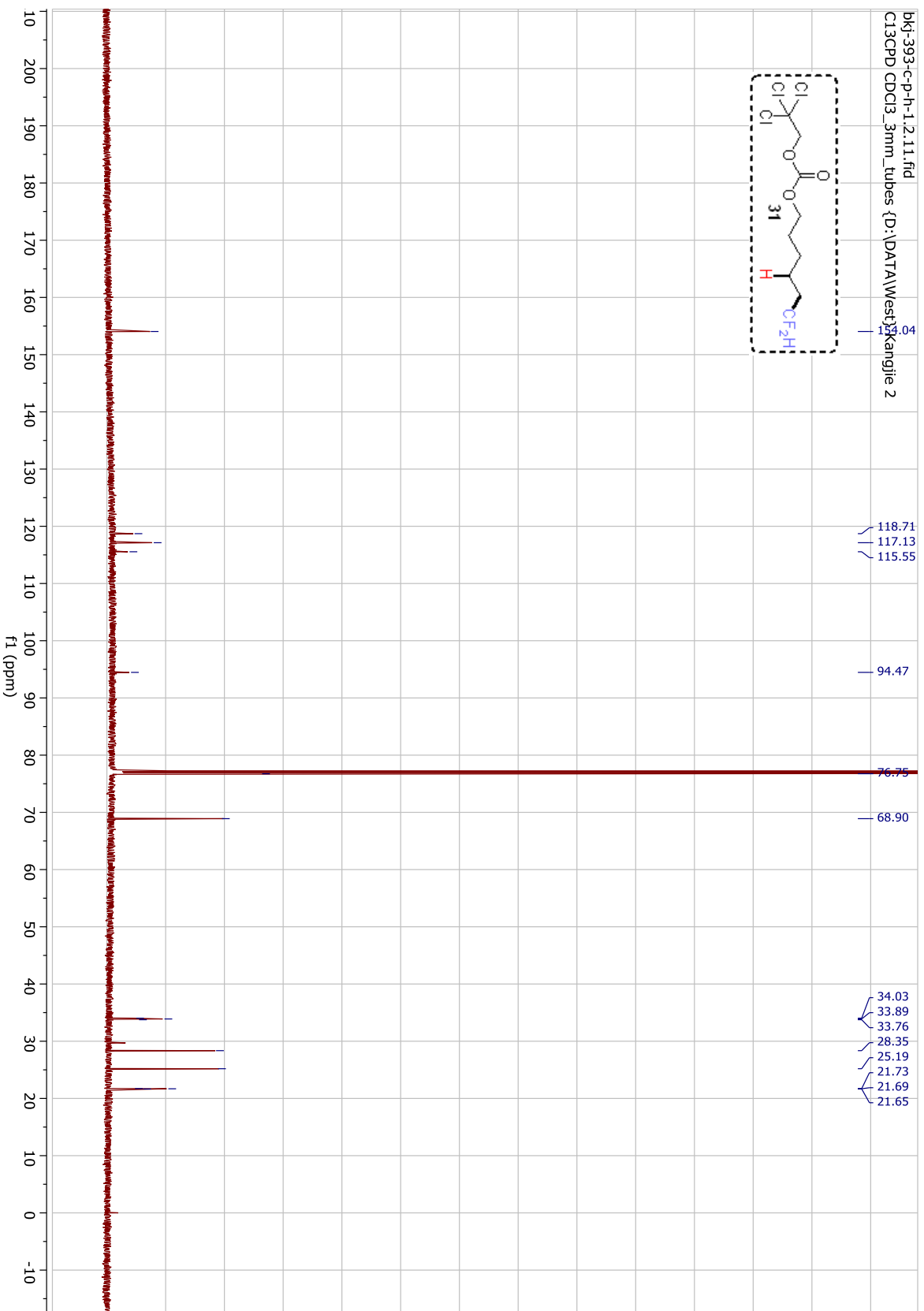
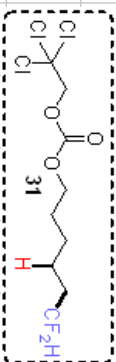
-115.80
-115.83
-115.86
-115.90
-115.93
-115.96



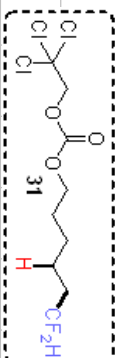
bkl-393-c-p-h-1.2.10.fid
PROTON CDCl3_3mm_tubes {D:\DATA\Wesley_Kangji\}



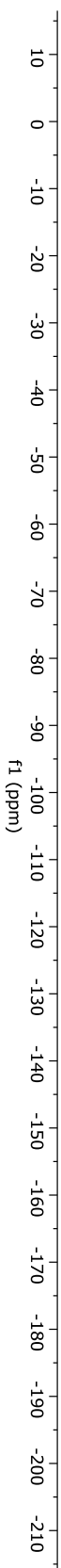
bk1-393-c-p-h-1.2.11.fid
C13CPD CDCl3_3mm_tubes {D:\DATA\Westfy\Kangjie 2

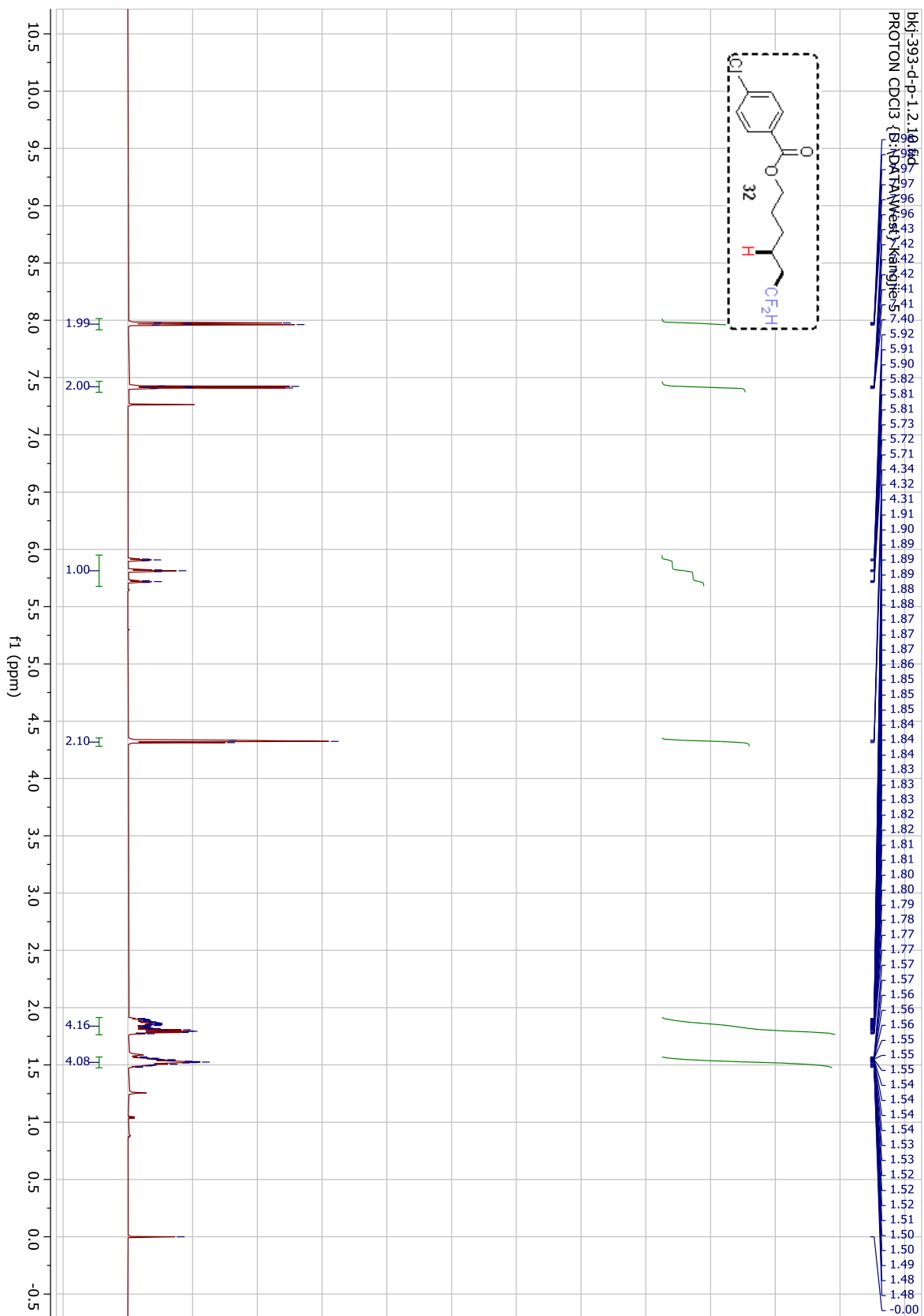


bkj-393-c-p-h-1.2.12.fid
F19_baseline_correct_1D
F19_baseline_correct CDCl3 3mm_tubes {D:\DATA\West} Kangjie 2

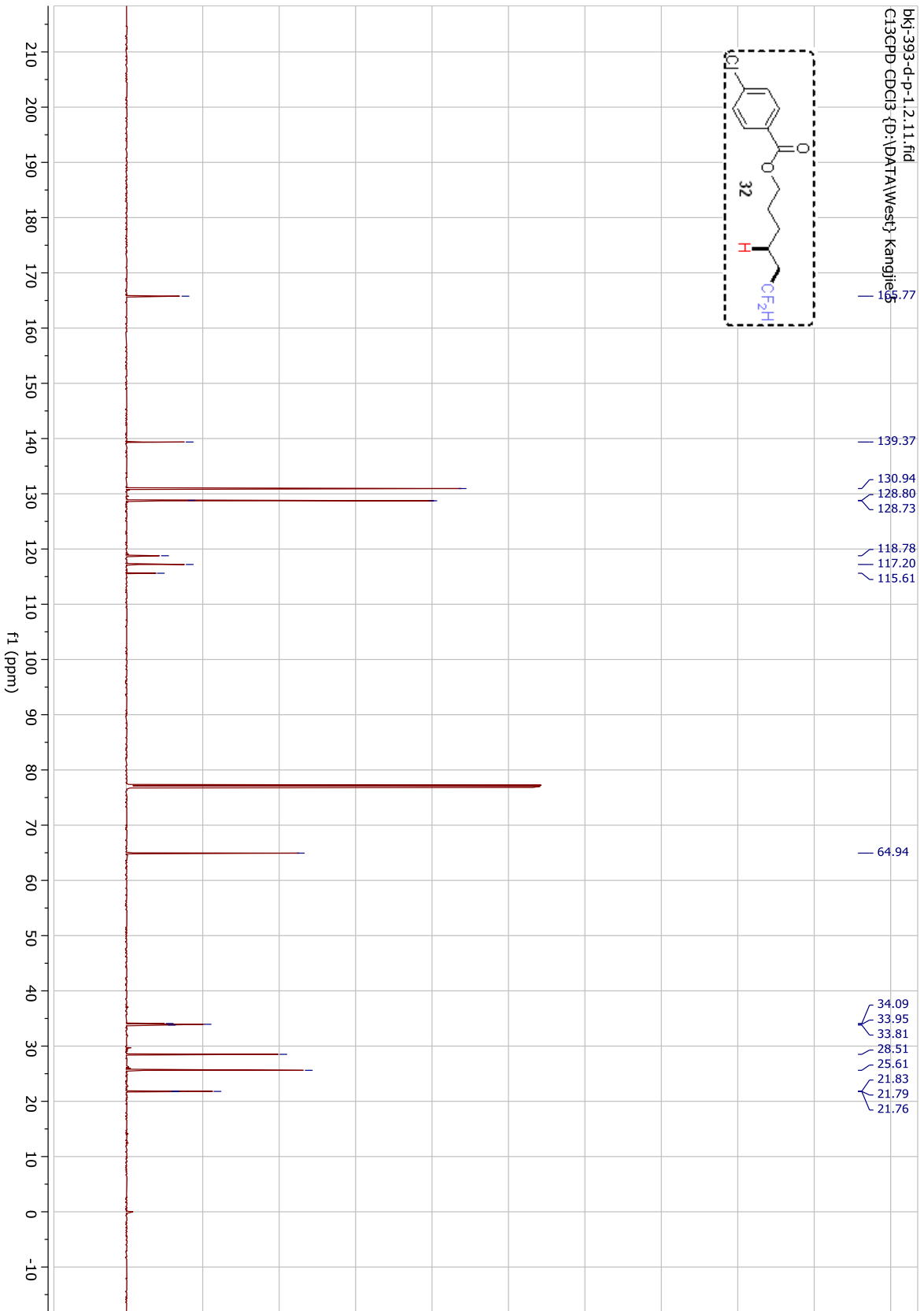
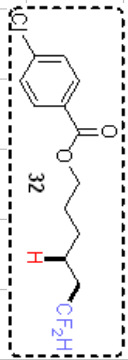


-115.88
-115.91
-115.91
-115.94
-115.94
-115.98
-116.01
-116.04

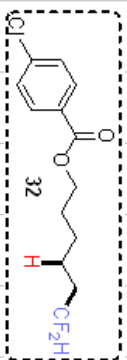




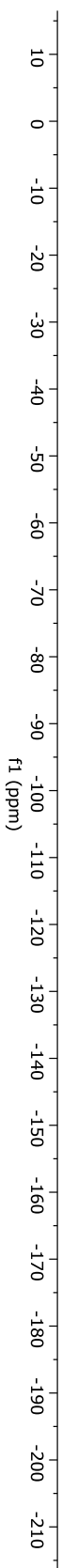
bkj-393-d-p-1,2,1,1.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjief5

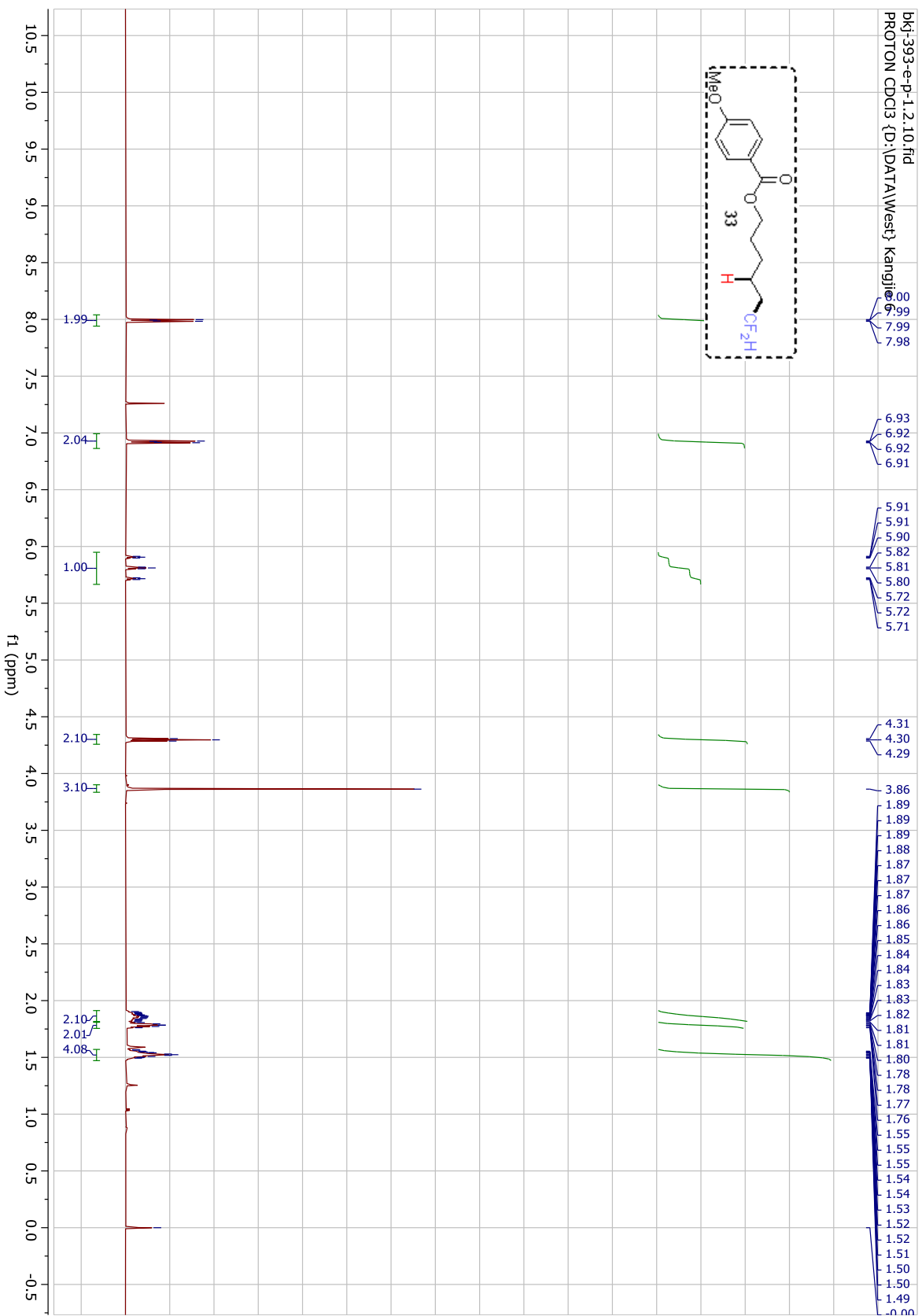


bkj-393-d-p-1.2.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 5



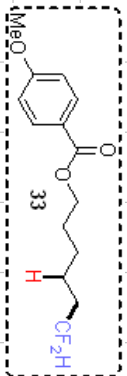
-115.82
-115.85
-115.88
-115.92
-115.92
-115.95
-115.98



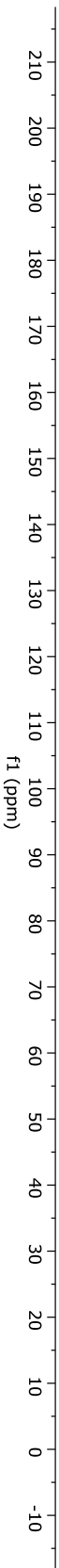


bkt-393-e-p-1,2,11.fid
C13CPD CDCl3 (D:\DATA\Westj Kanji)

40
163.34
131.55
122.81
118.83
117.25
115.67
113.61

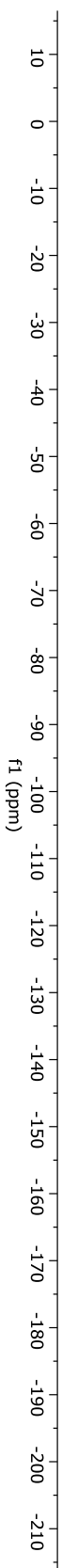
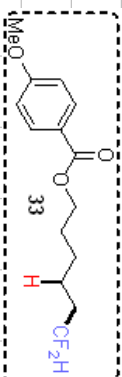


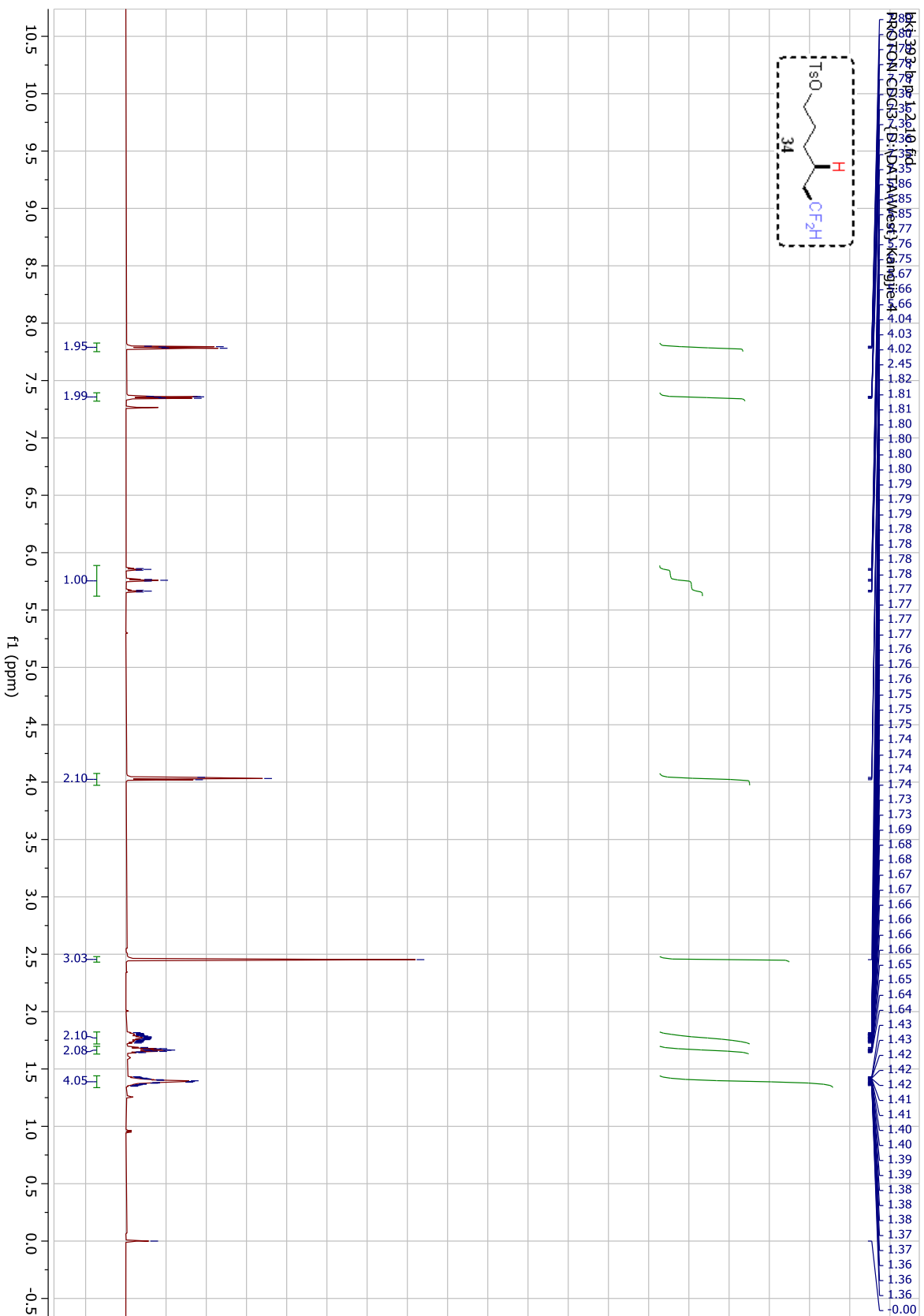
64.38
55.44
34.12
33.98
33.84
28.60
25.66
21.87
21.84
21.80



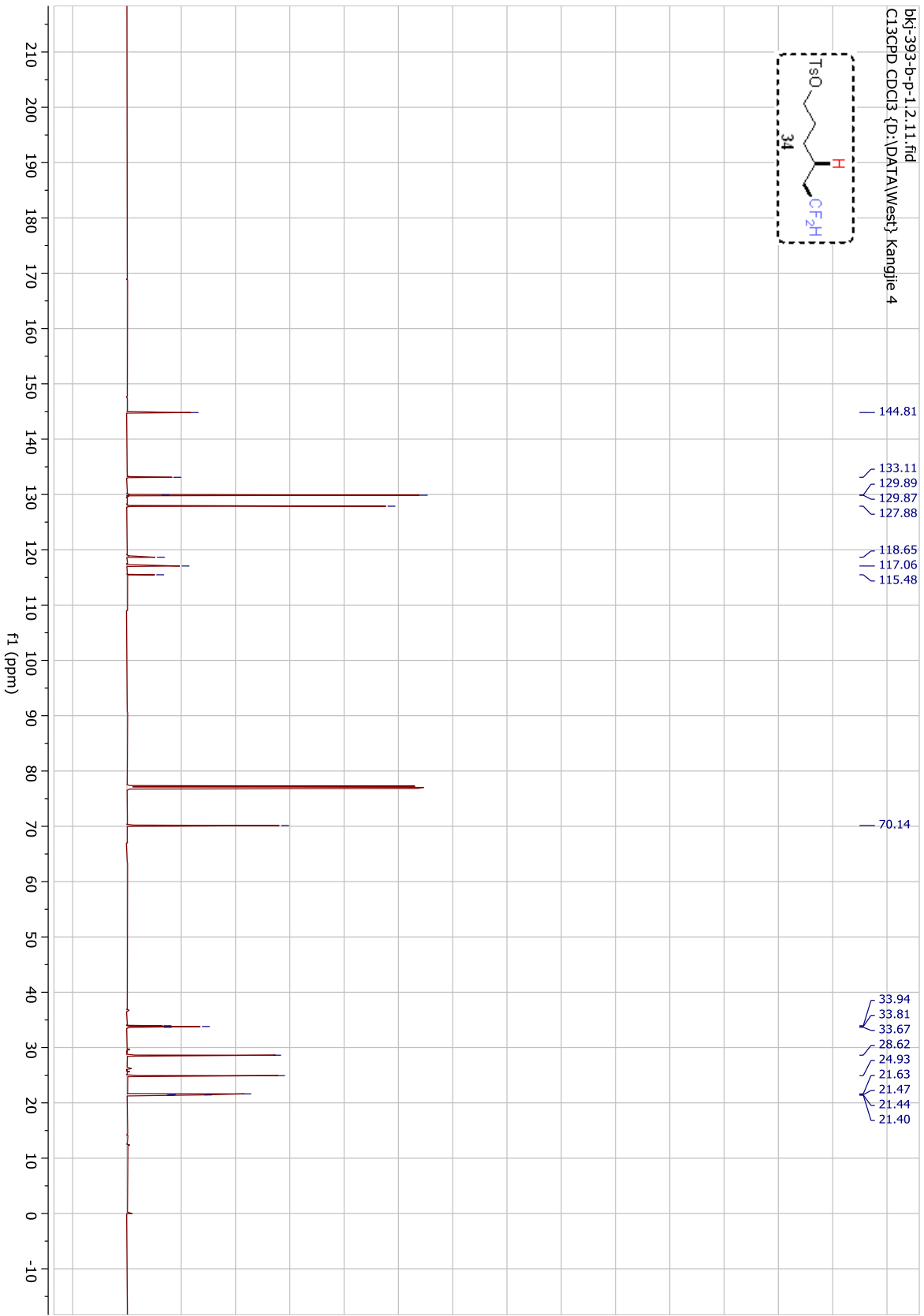
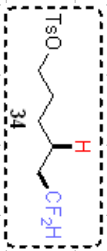
bkj-393-e-p-1.2.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 6

-115.78
-115.81
-115.84
-115.88
-115.91
-115.94

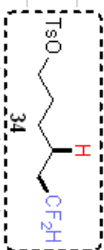




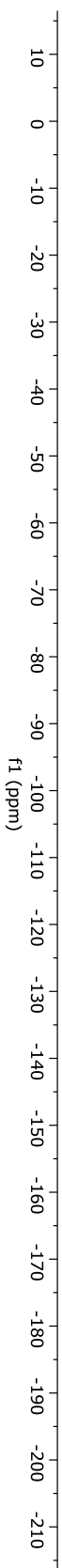
bkj-393-b-p-1,2,1,1.fid
C13CPD CDCl3 (D:\DATA\Westj_Kangjie 4

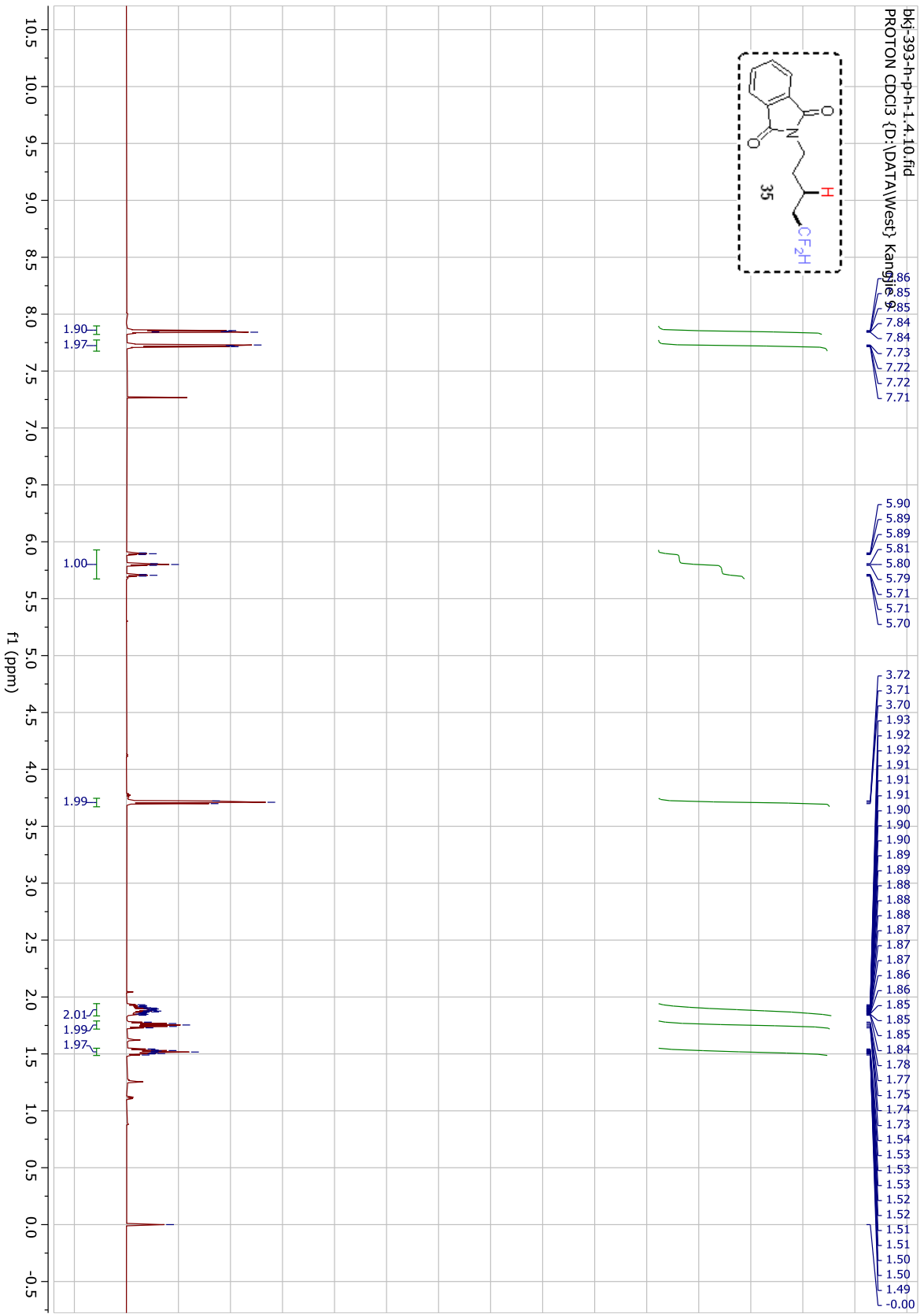


bkj-393-b-p-1.2.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 4

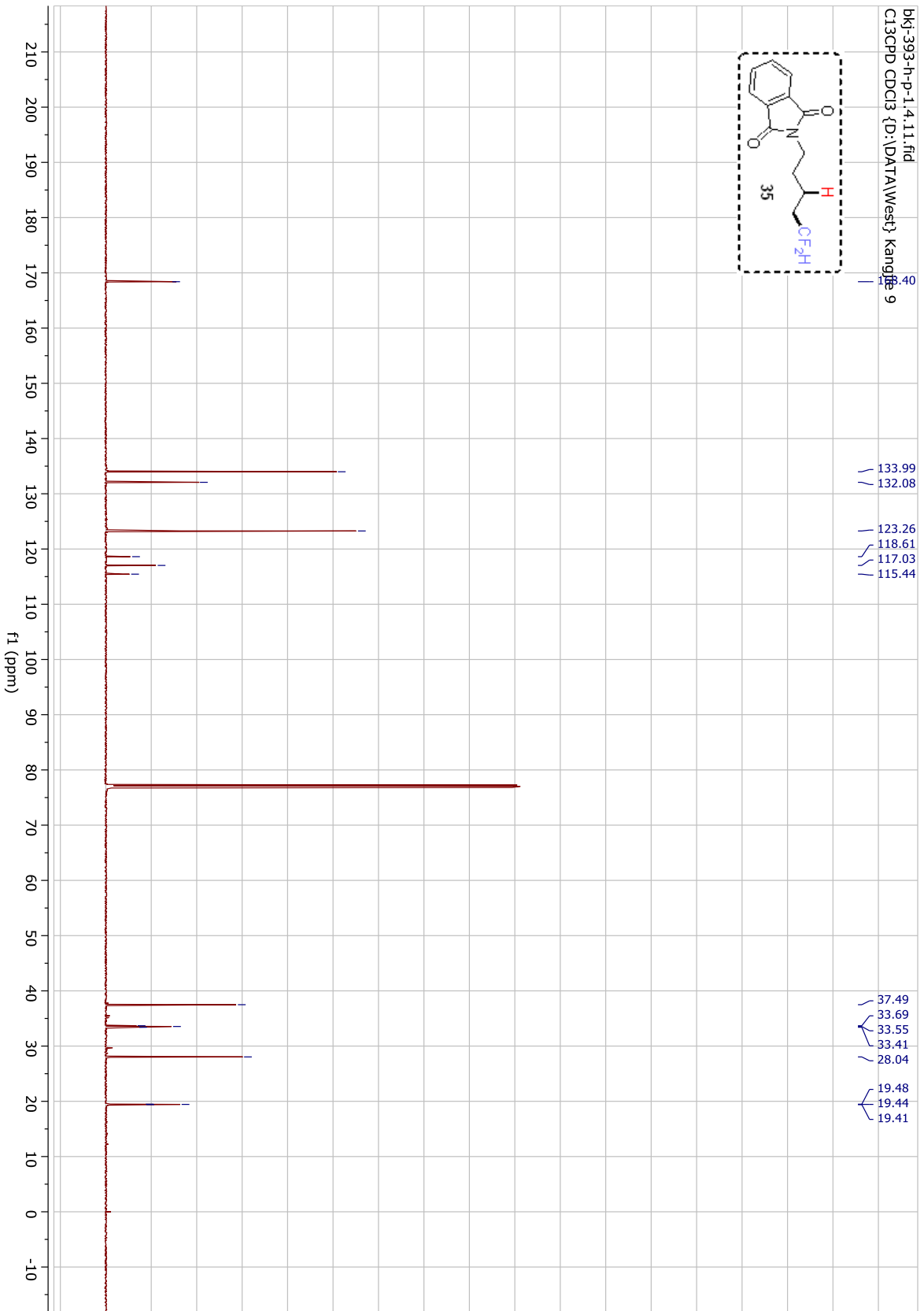
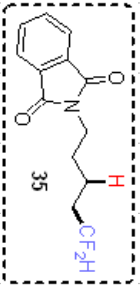


-115.92
-115.95
-115.98
-116.02
-116.05
-116.08

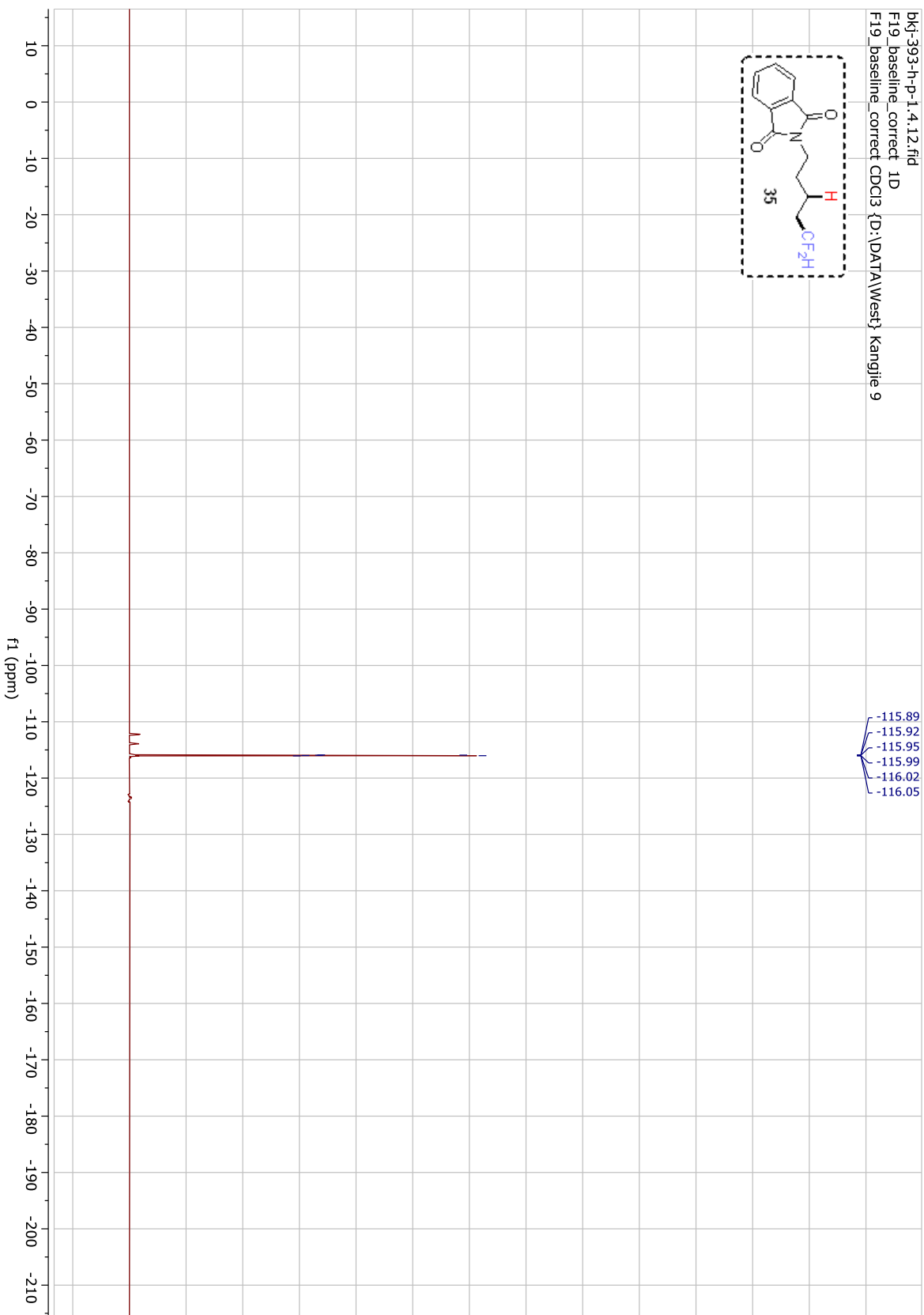
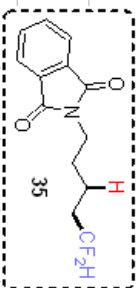


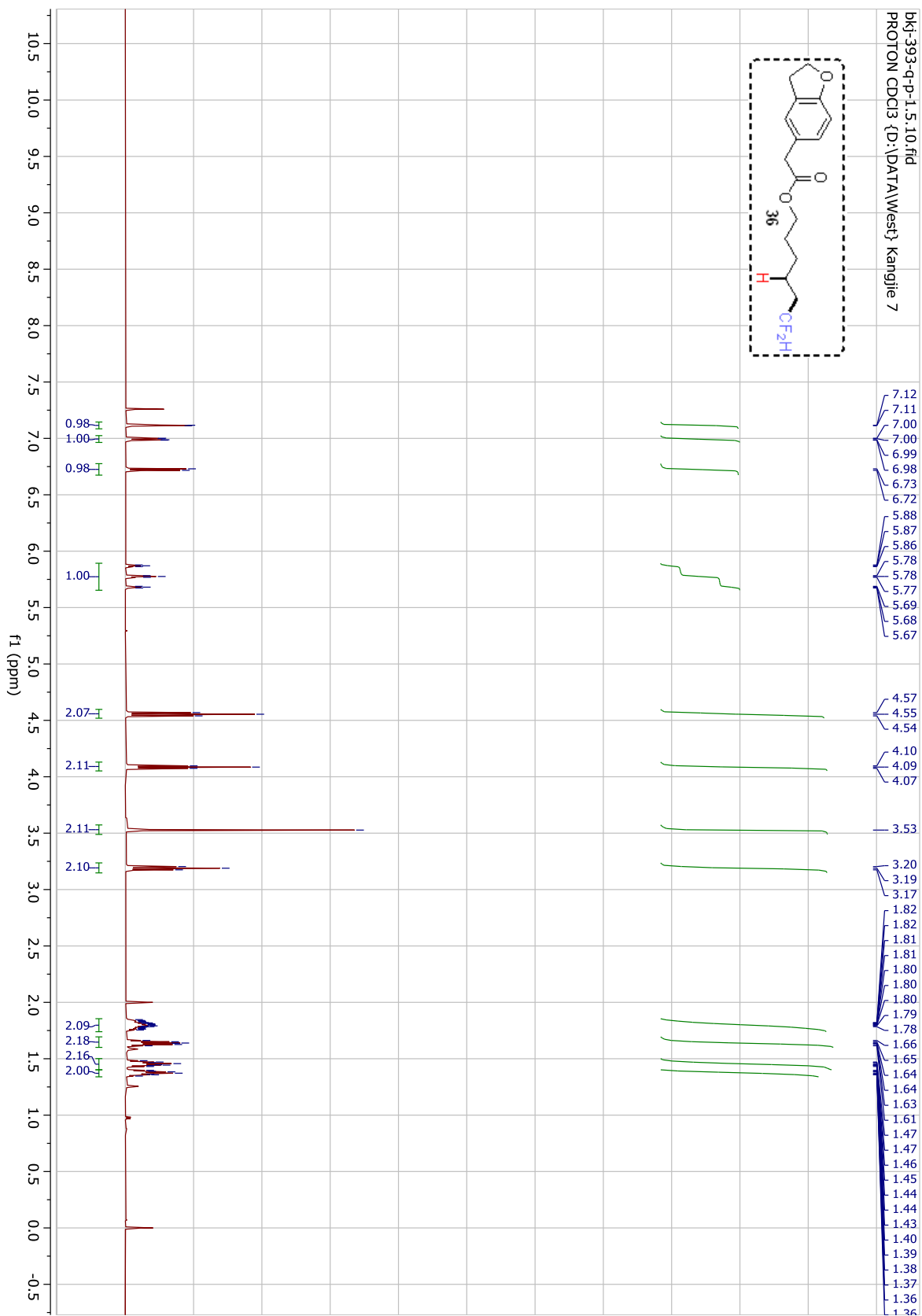


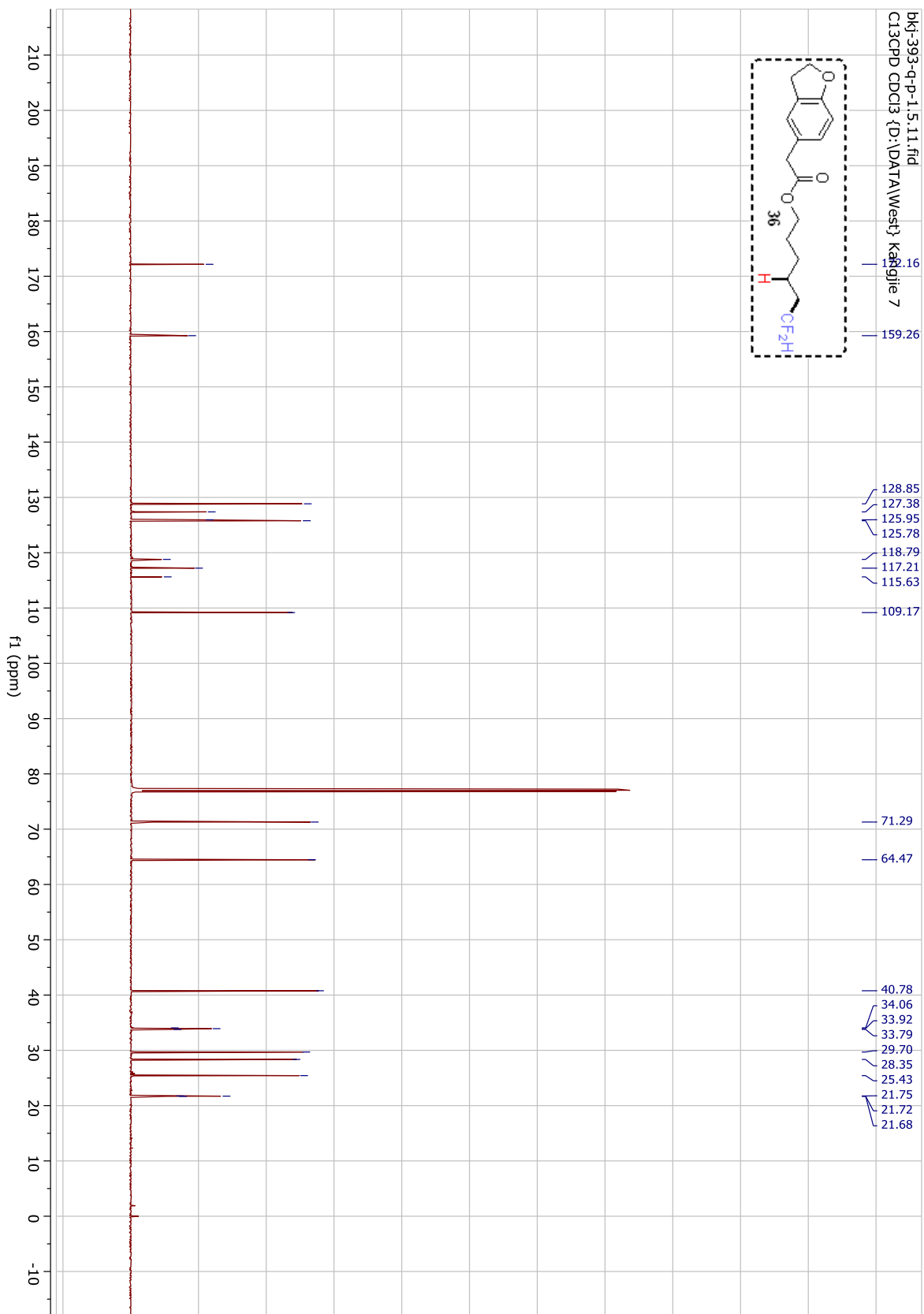
bkt-393-h-p-1,4,1,1.fid
C13CPD CDCl3 (D:\DATA\Westj_Kanfig
9



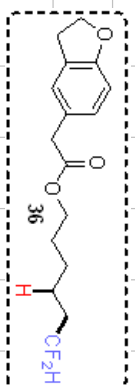
bkj-393-h-p-1-4.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 9



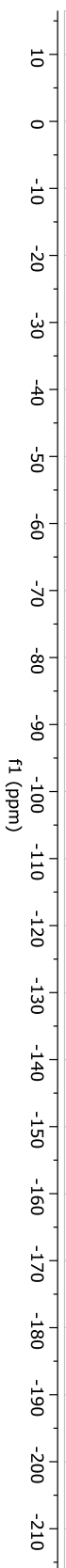




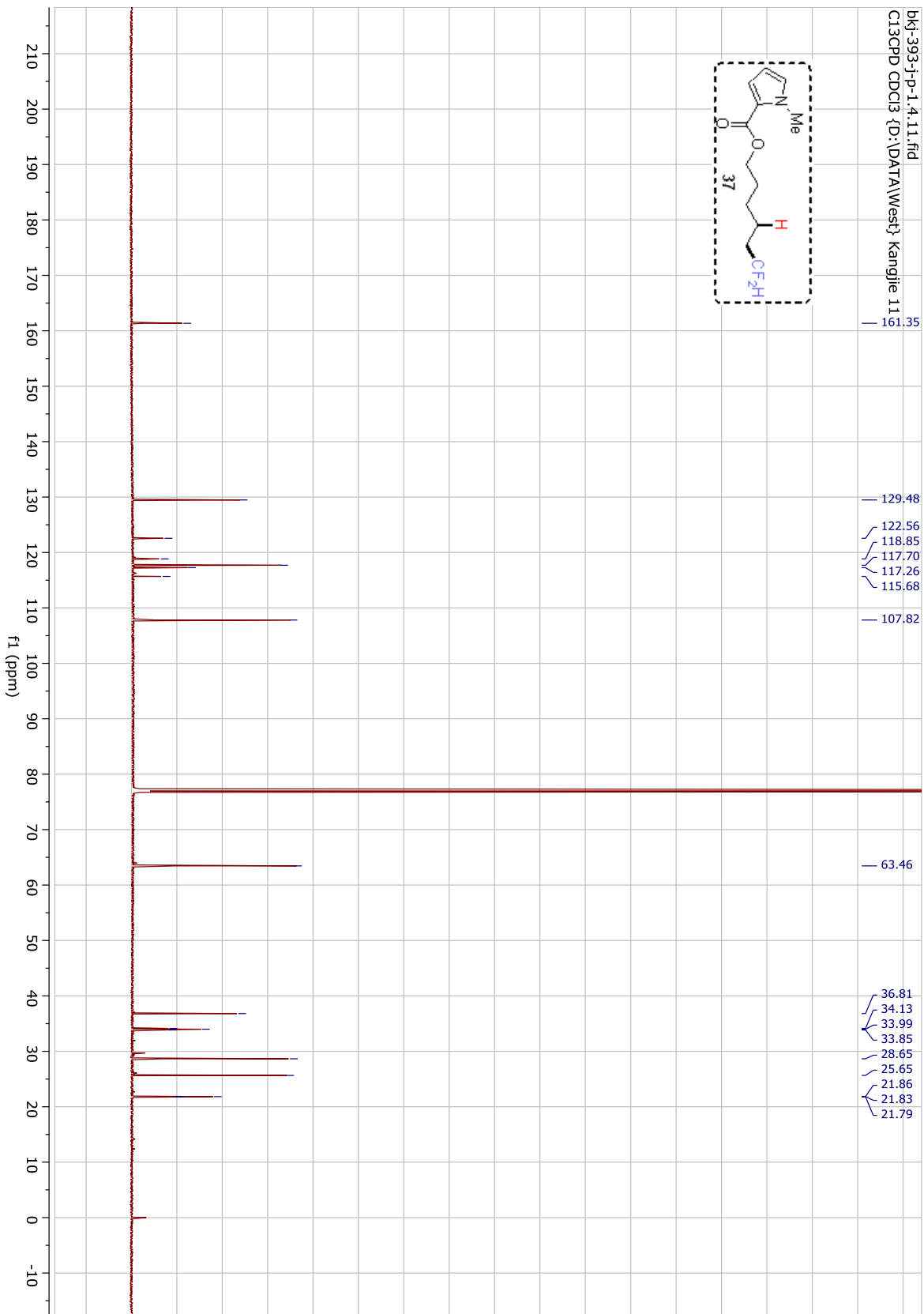
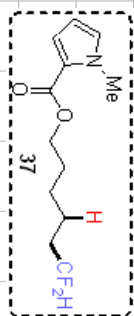
bkj-393-q-p-1.5.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 7



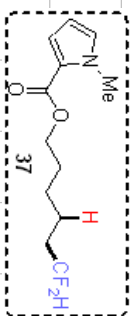
-115.78
-115.81
-115.82
-115.82
-115.85
-115.88
-115.91
-115.95



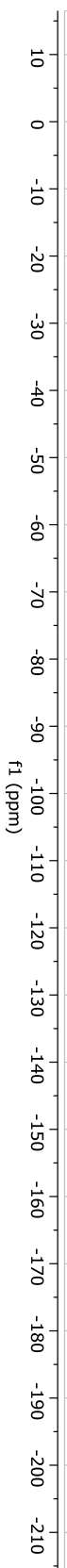
bkj-393-j-p-1,4,11.fid
C13CPD CDCl3 (D:\DATAWestj_Kangjie 11

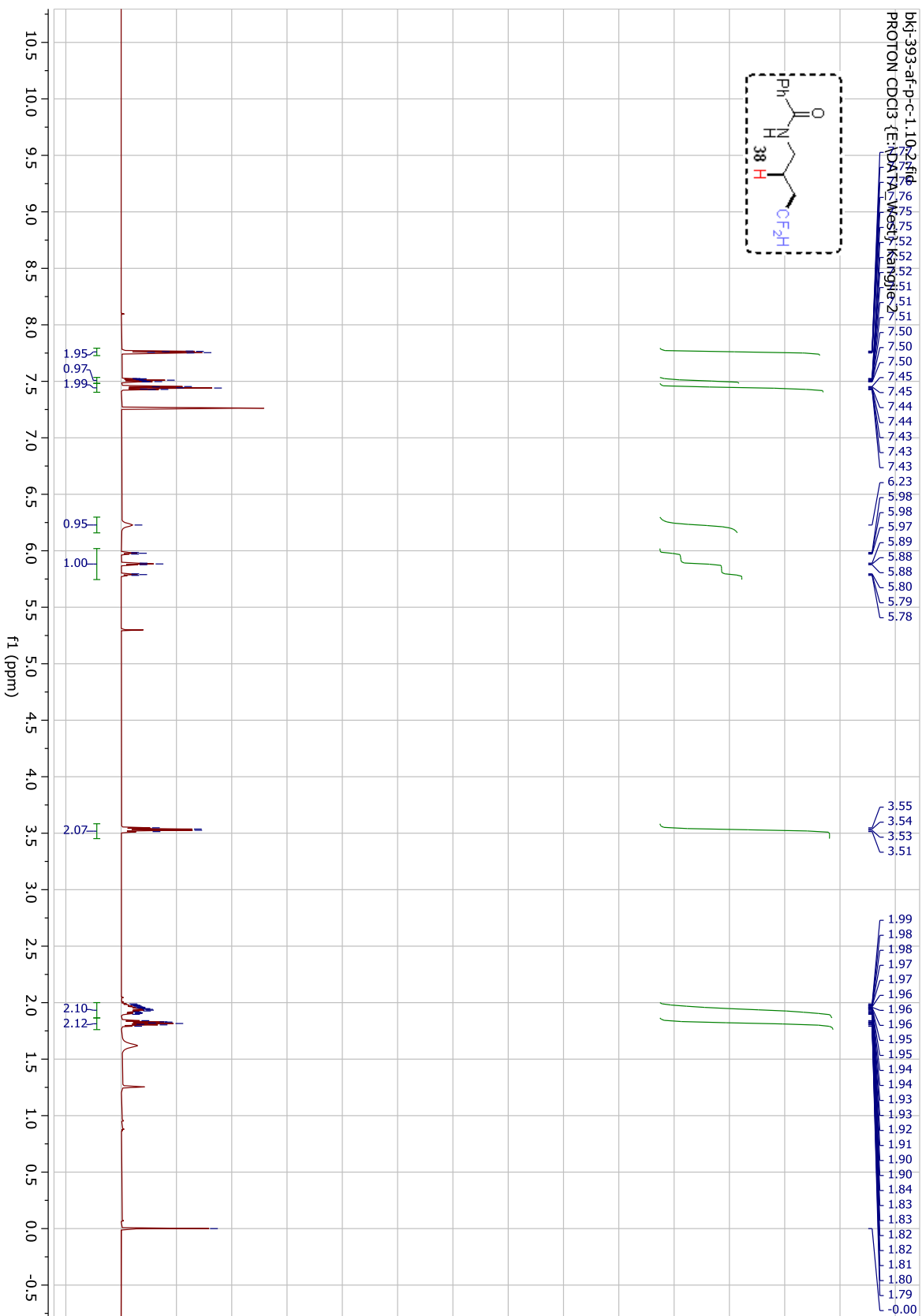


bkj-393-j-p-1.4.12.ftd
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 11

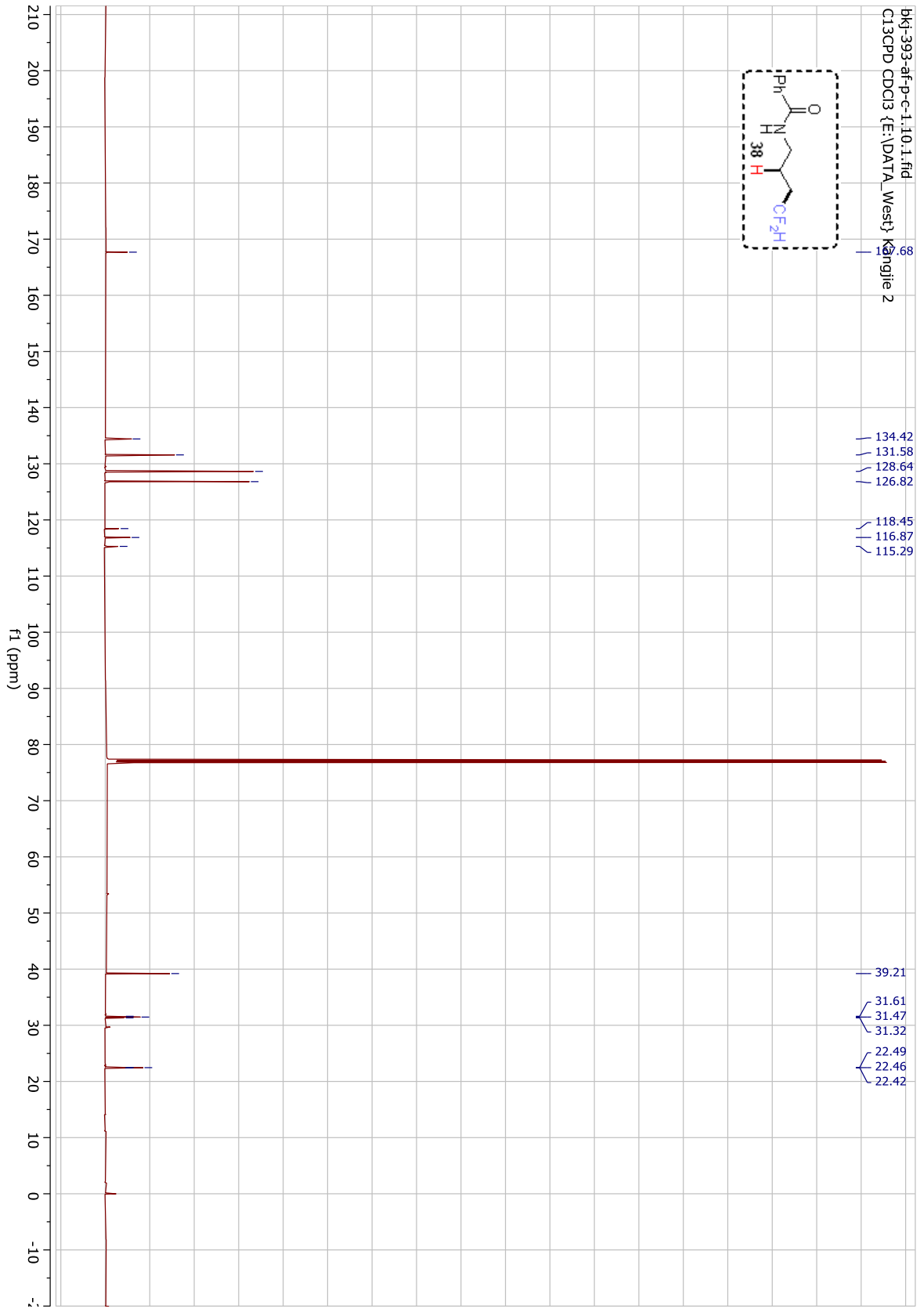
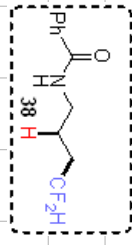


-115.77
-115.80
-115.83
-115.87
-115.90
-115.93

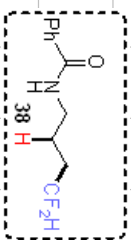




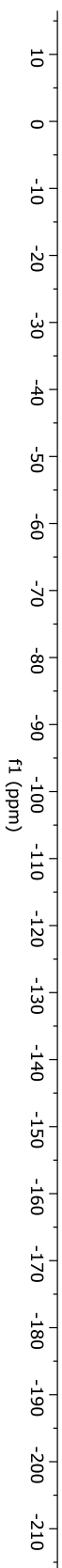
bkj-393-at-p-c-1.10.1.fid
C13CPD CDCl3 {E:\DATA_West\Kangjie 2

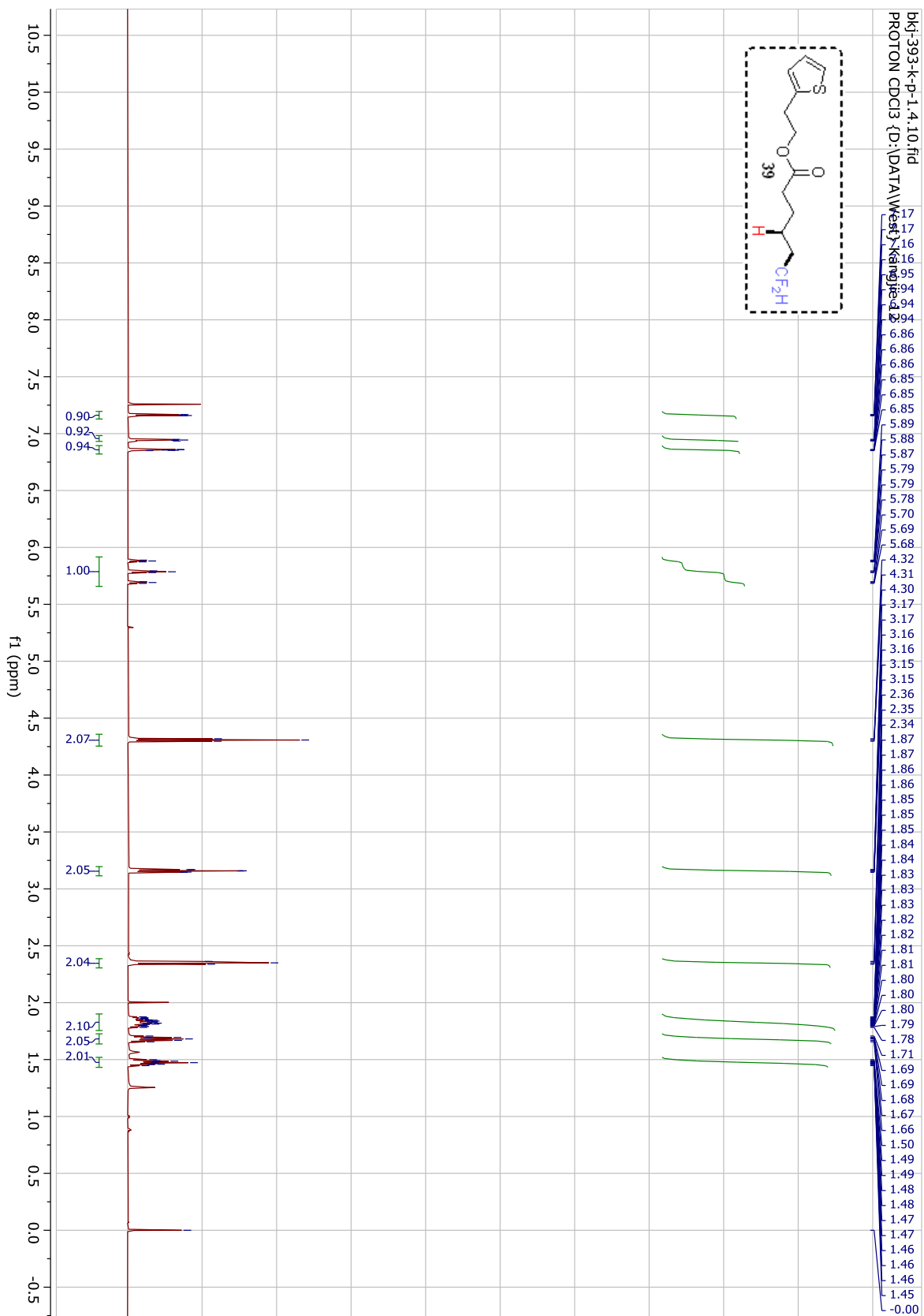


bkj-393-af-p-h-1.9.11.ftd
F19_baseline_correct_1D
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 19

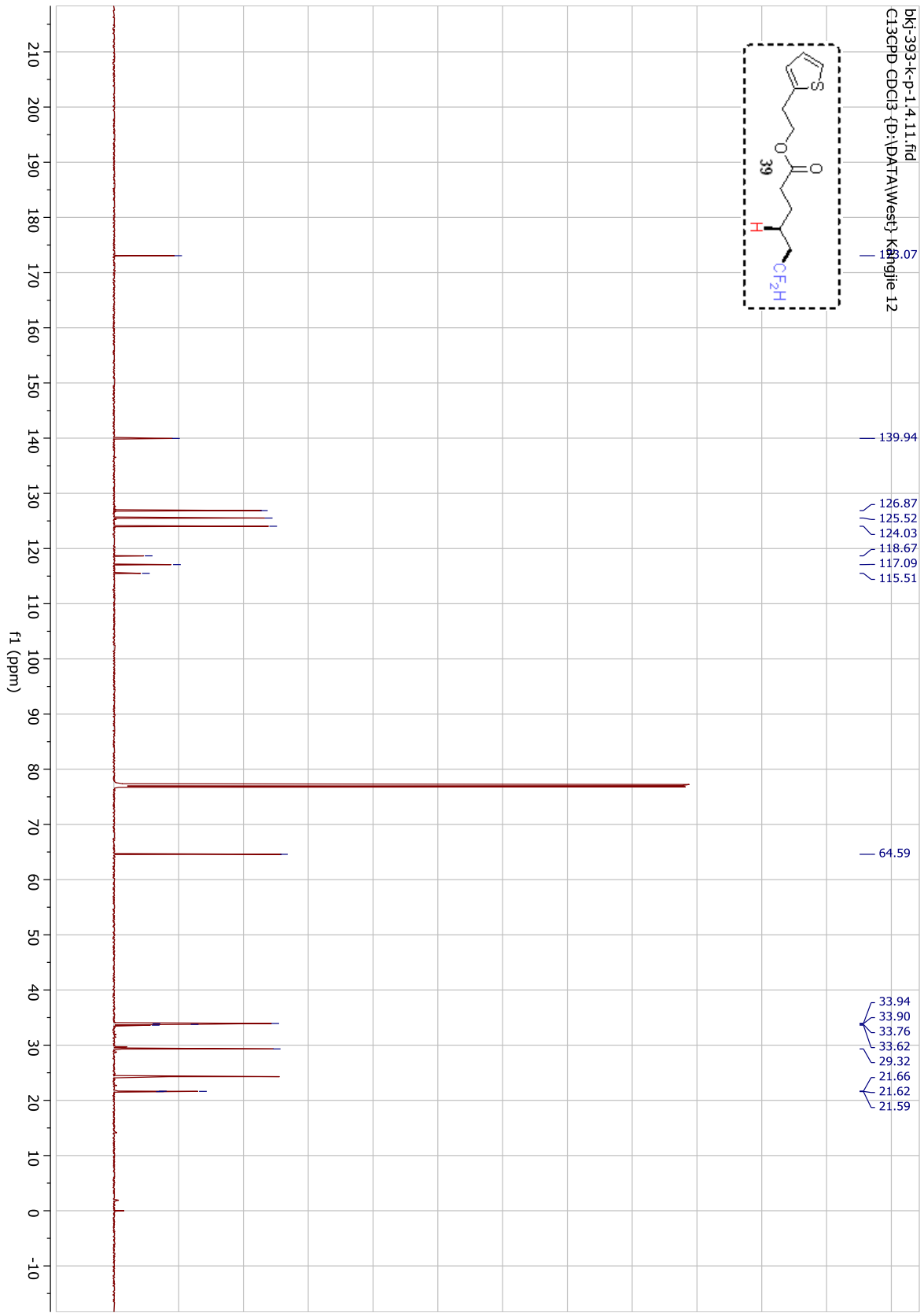
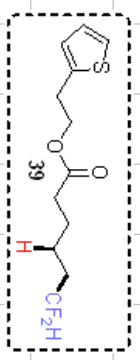


-115.91
-115.94
-115.97
-116.01
-116.04
-116.07

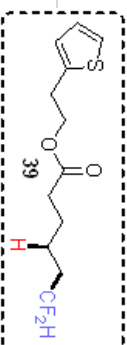




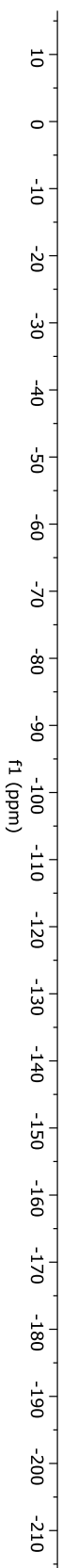
bkj-393-k-p-1,4,11.fid
C13CPD CDCl3 (D:\DATA\Westj k\ngjle-12

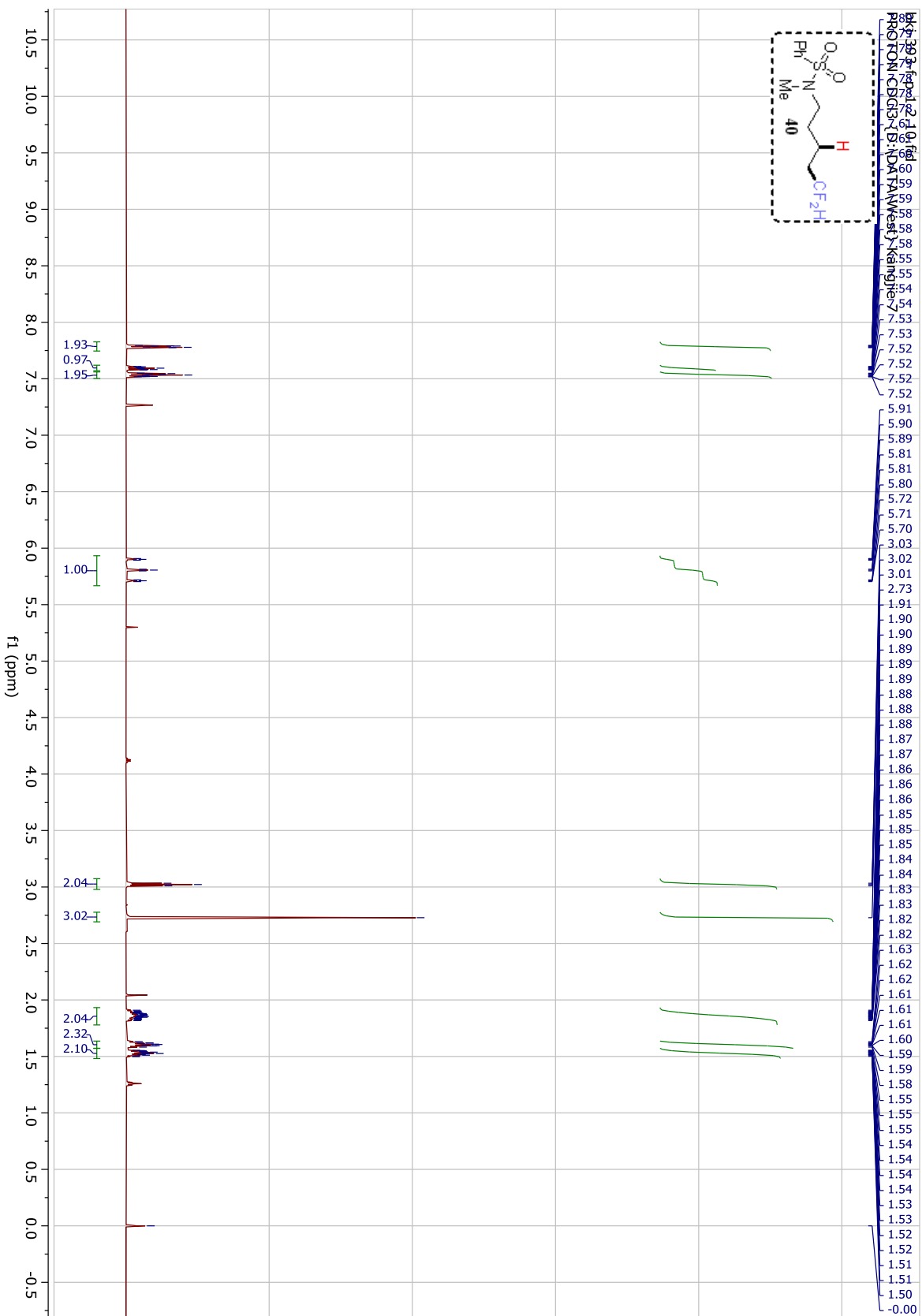


bkj-393-k-p-1-4.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 12

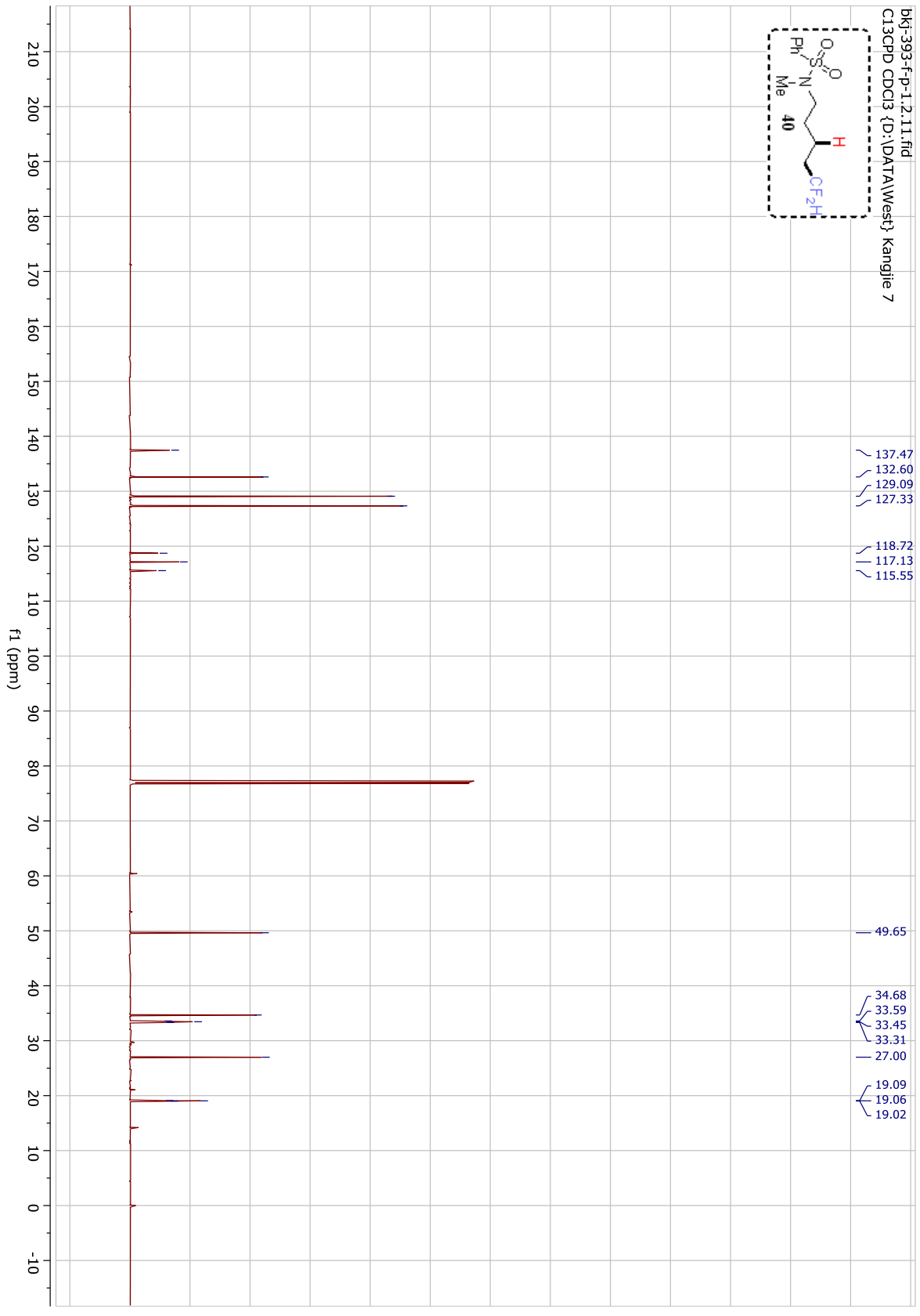
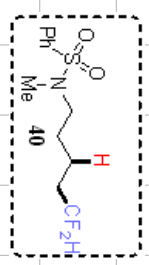


-115.82
-115.86
-115.89
-115.92
-115.96
-115.99

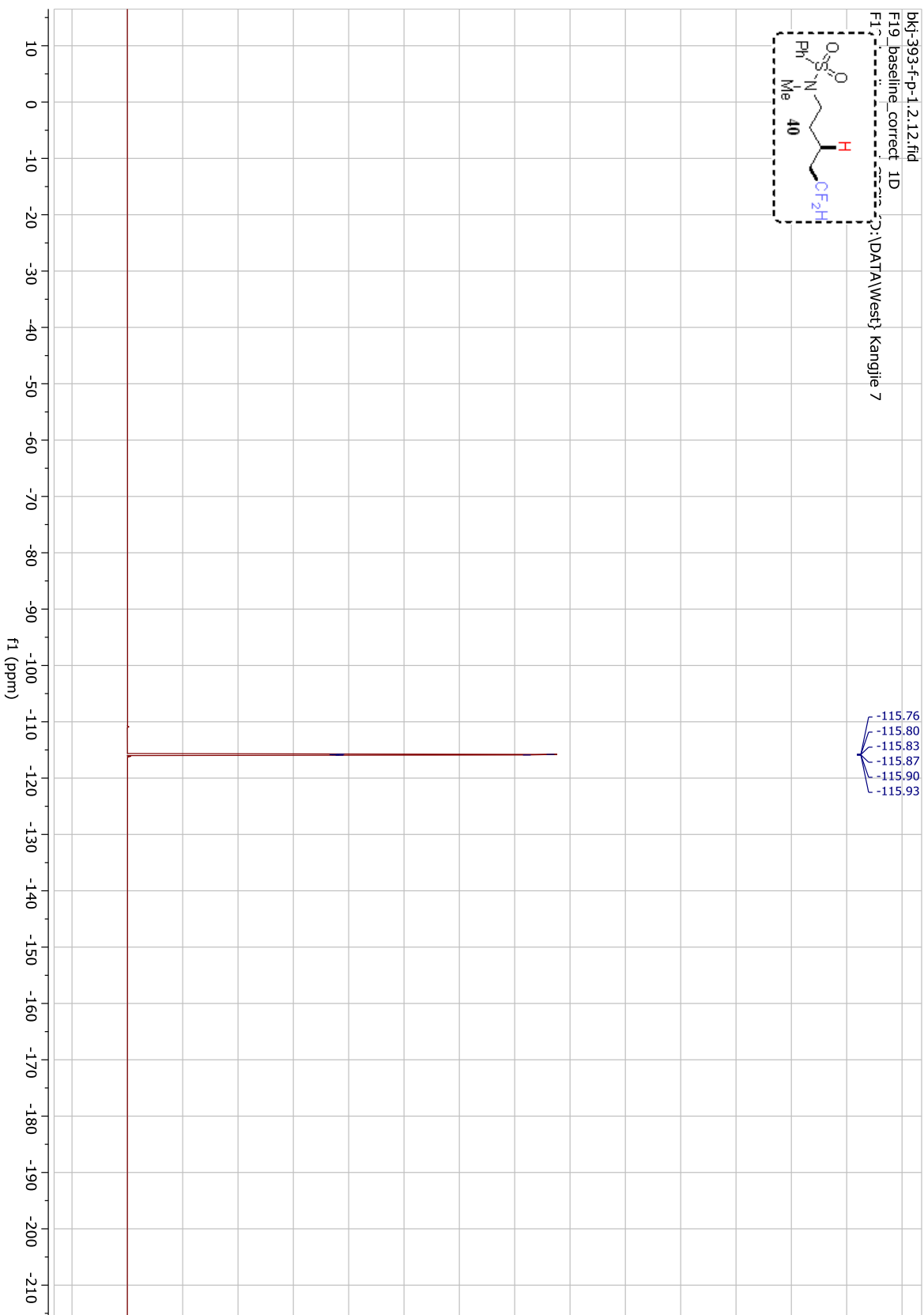
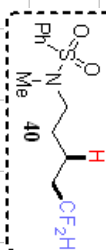




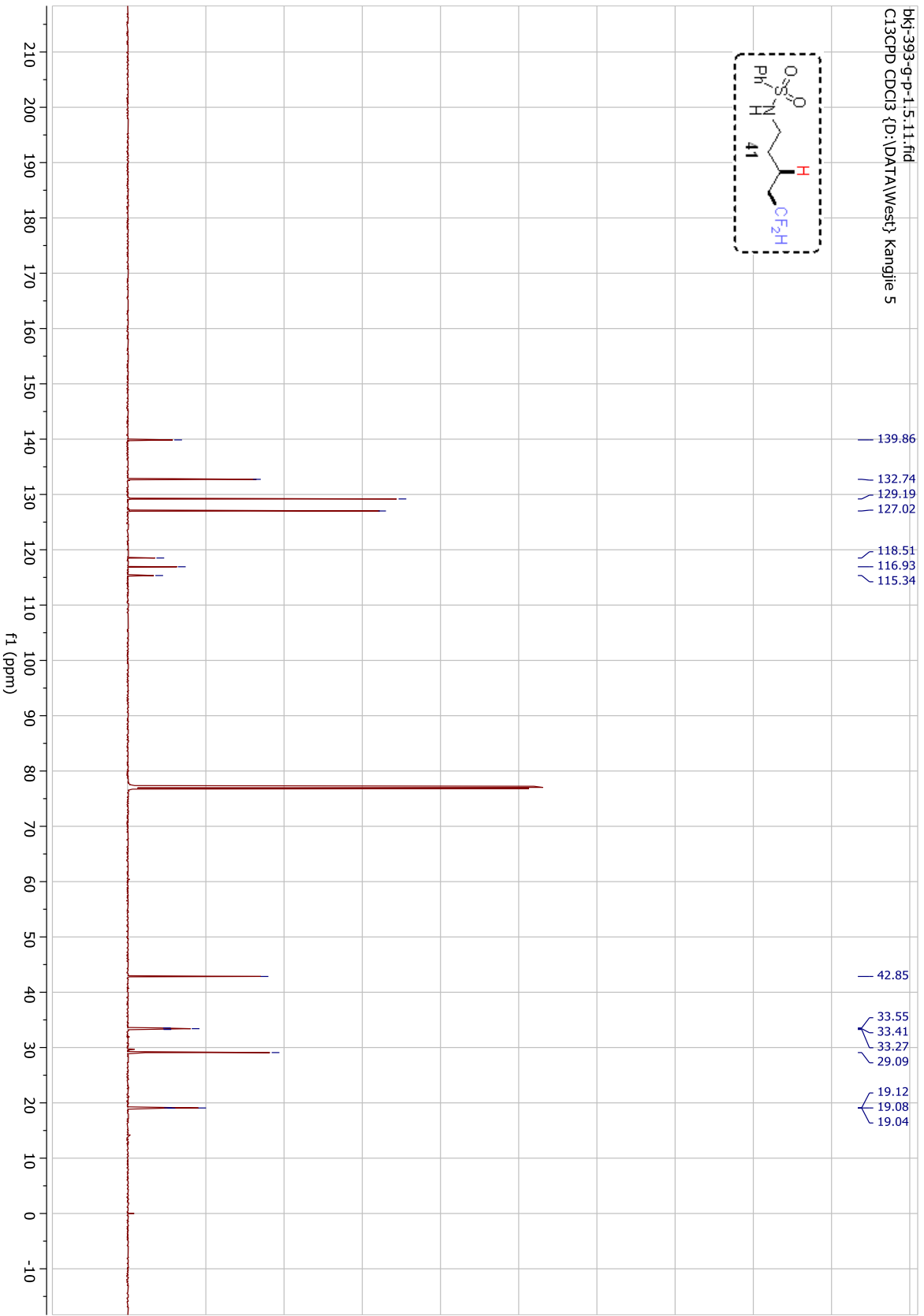
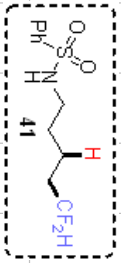
bkt-393-f-p-1.2.11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 7



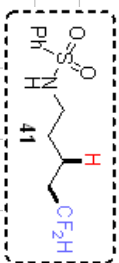
bkj-393-f-p-1.2.12.ftd
F19_baseline_correct ID
F19_20120120_001.D\\DATA\\West\\Kangjie 7



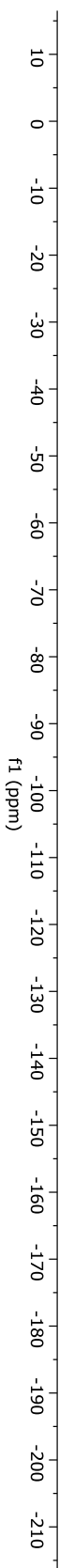
bkj-393-g-p-1-5-11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 5



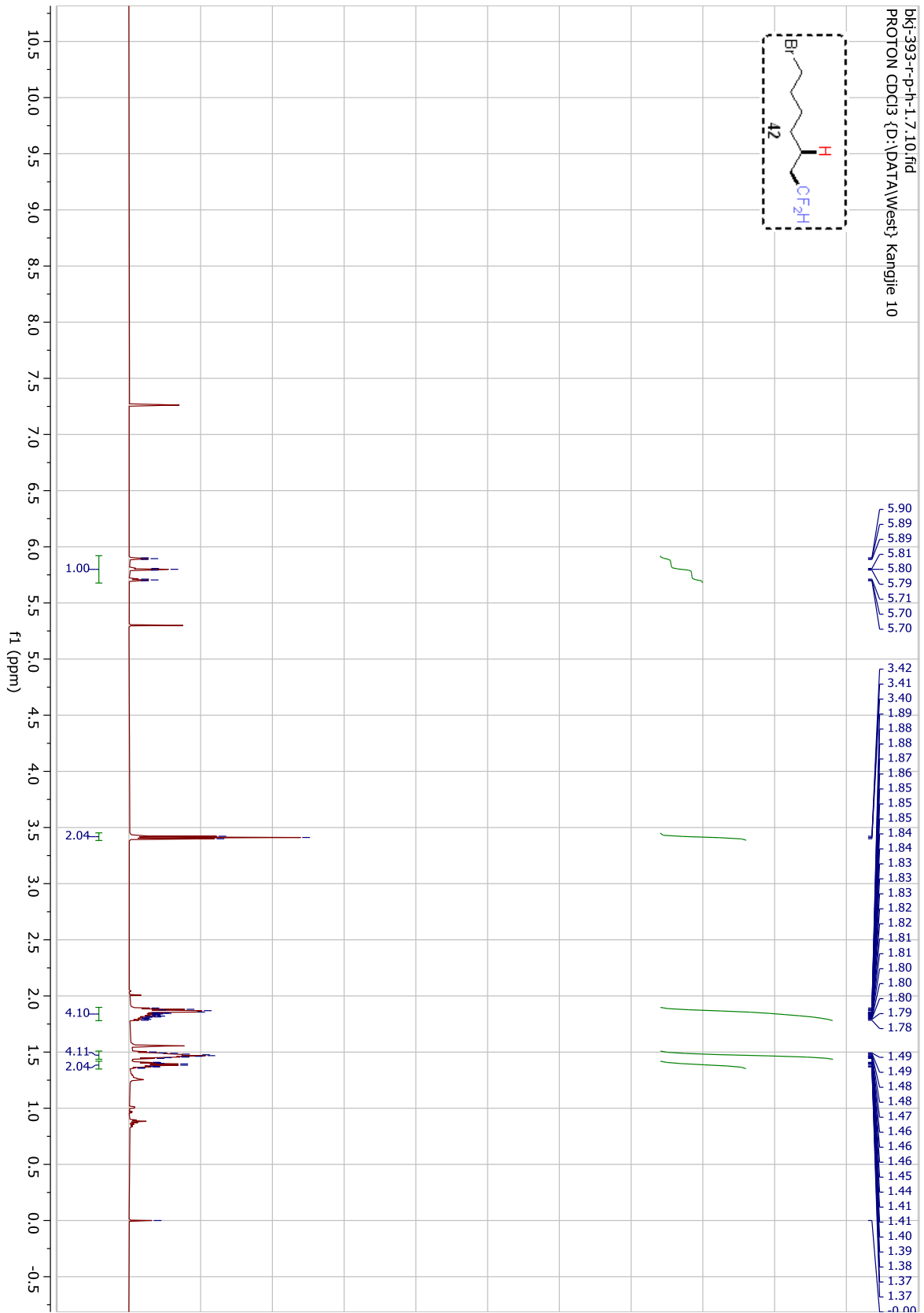
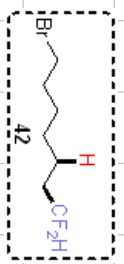
bkj-393-g-p-1.5.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 5



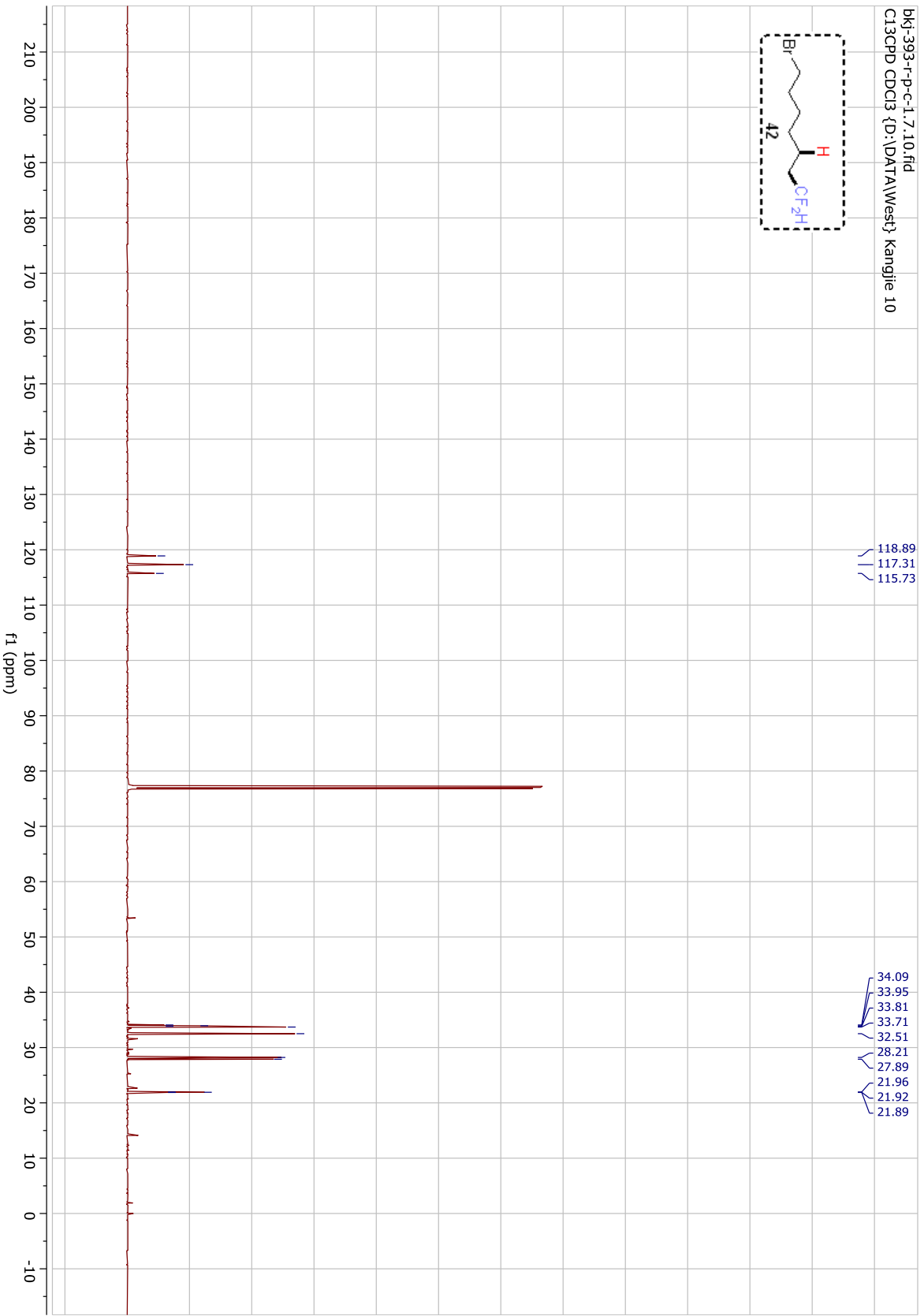
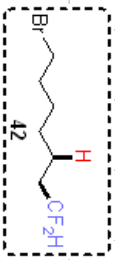
-115.97
-116.00
-116.00
-116.03
-116.07
-116.10
-116.14



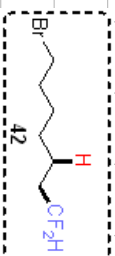
bk1-393-r-p-h-1-7.10.fid
PROTON CDCl3 {D:\DATA\West\Kangjie 10



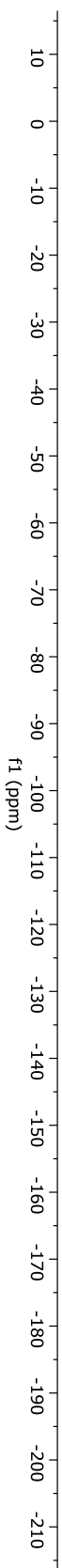
bkt-393-r-p-c-1.7.10.fid
C13CPD CDCl3 (D:\DATA\Westj) Kangle 10



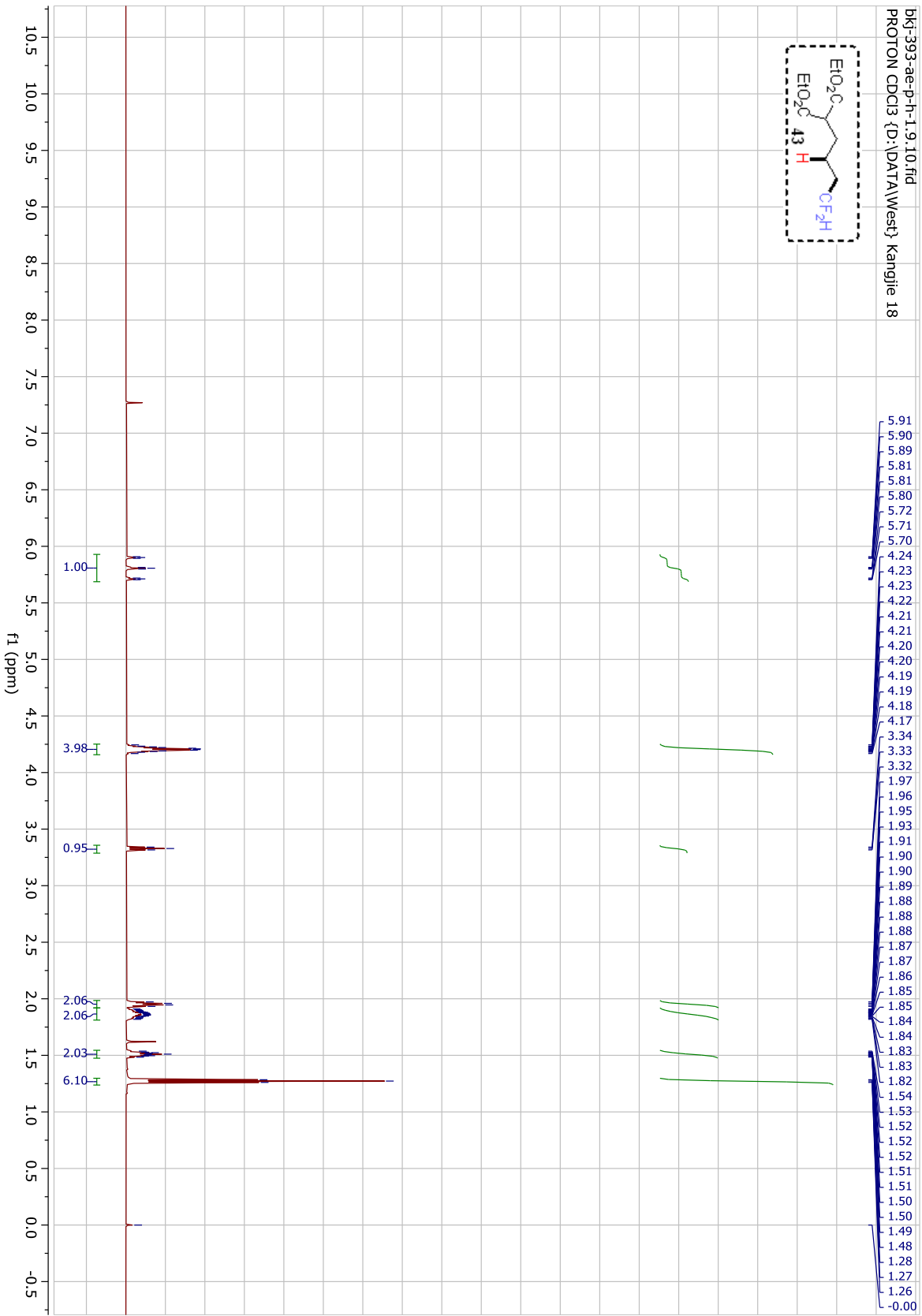
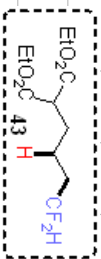
bkj-393-r-p-h-1-7.11.fid
F19_baseline_correct_1D
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 10



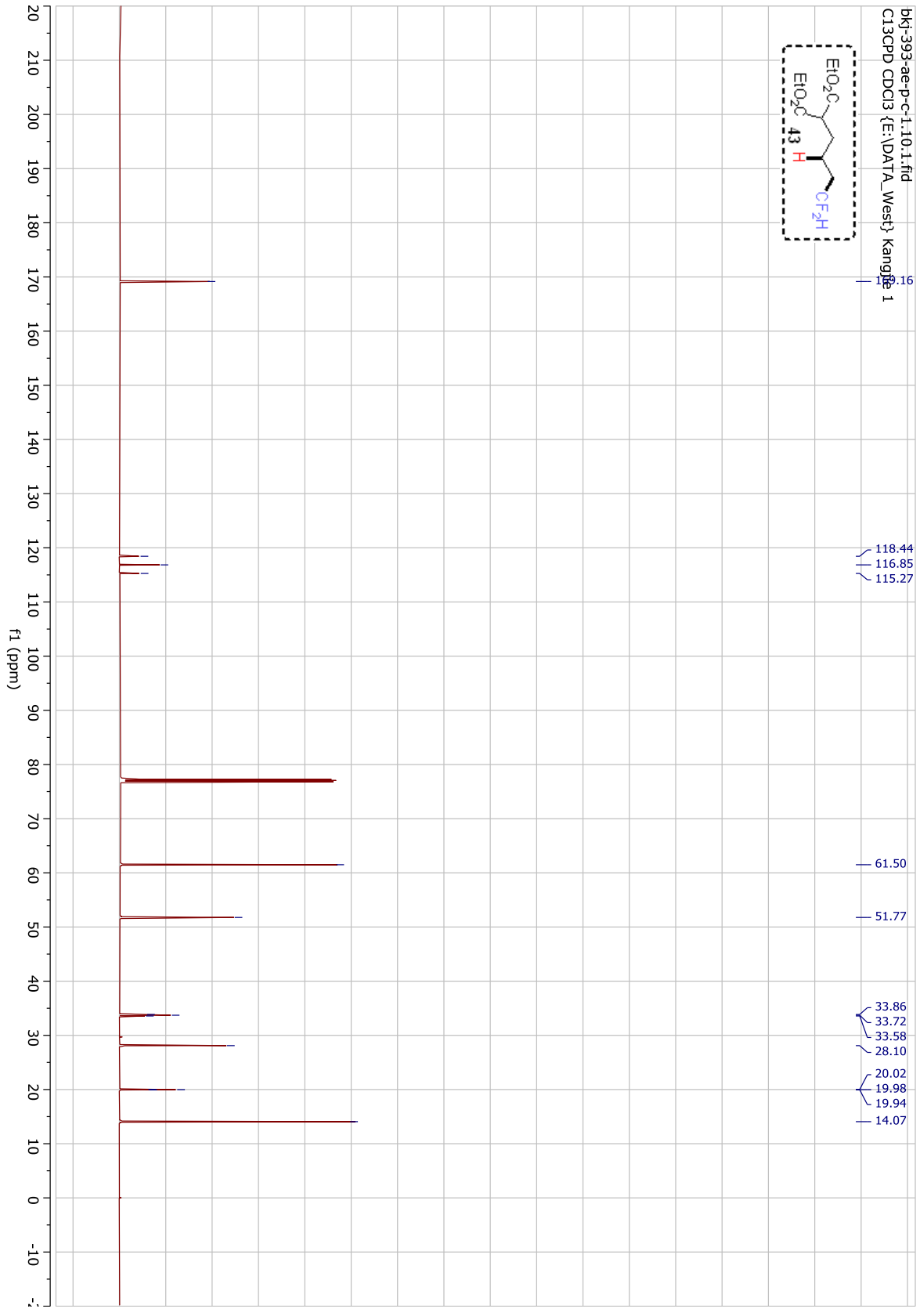
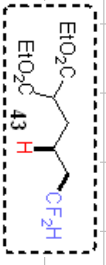
-115.76
-115.80
-115.83
-115.87
-115.90
-115.93



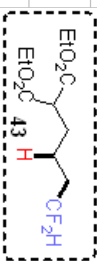
bj1-393-ae-p-h-1.9.10.fid
PROTON CDCl3 {D:\DATA\West\Kangjie 18



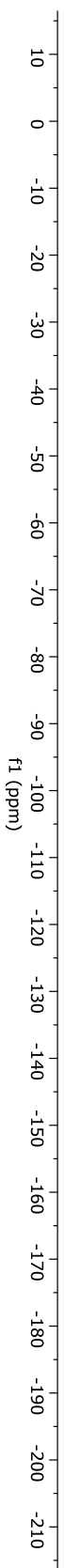
bjt-393-ae-p-c-1.10.1.fid
C13CPD CDCl3 (E:\DATA_West\ Kang) 1

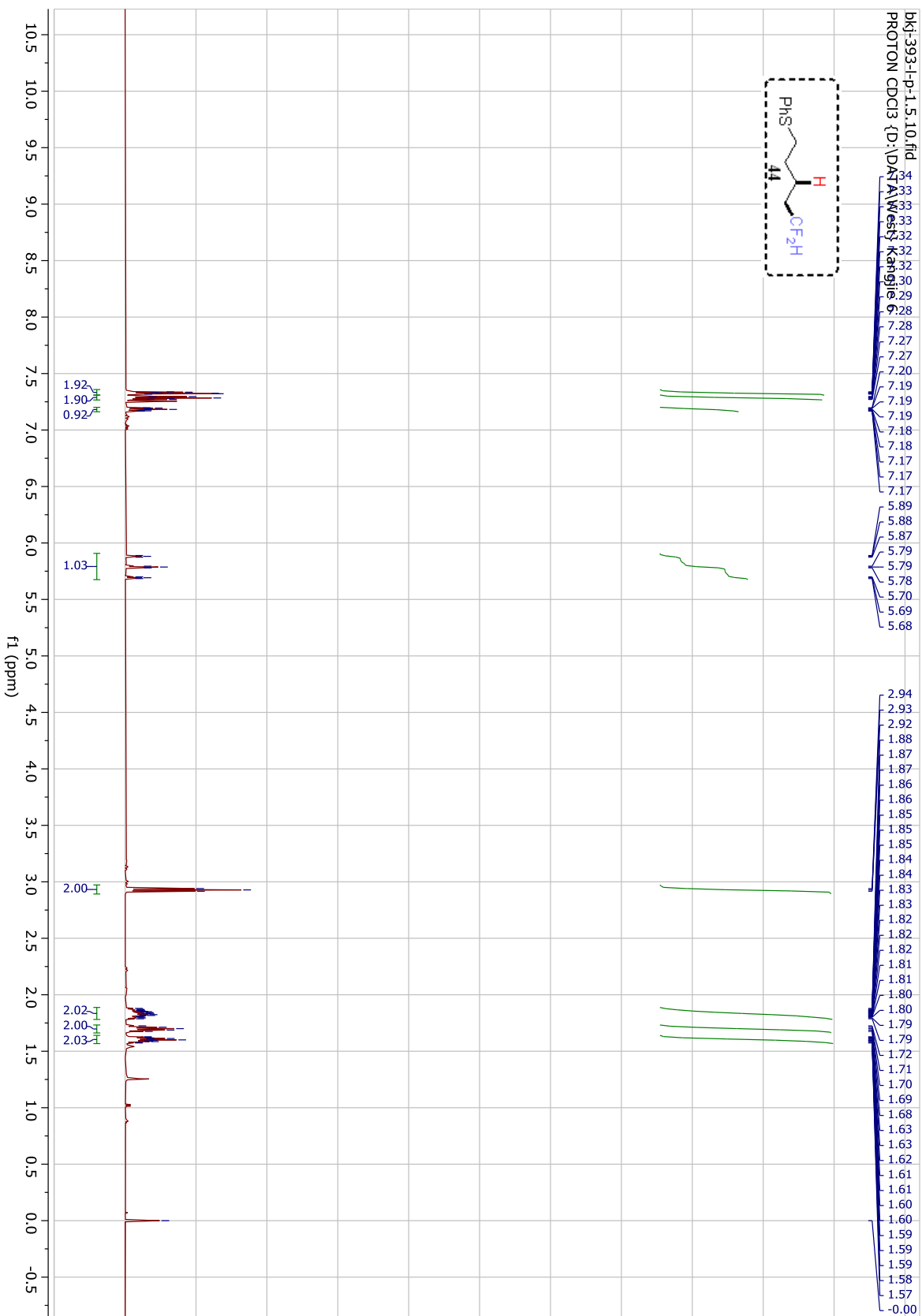


bk1-393-ae-p-h-1.9.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 18

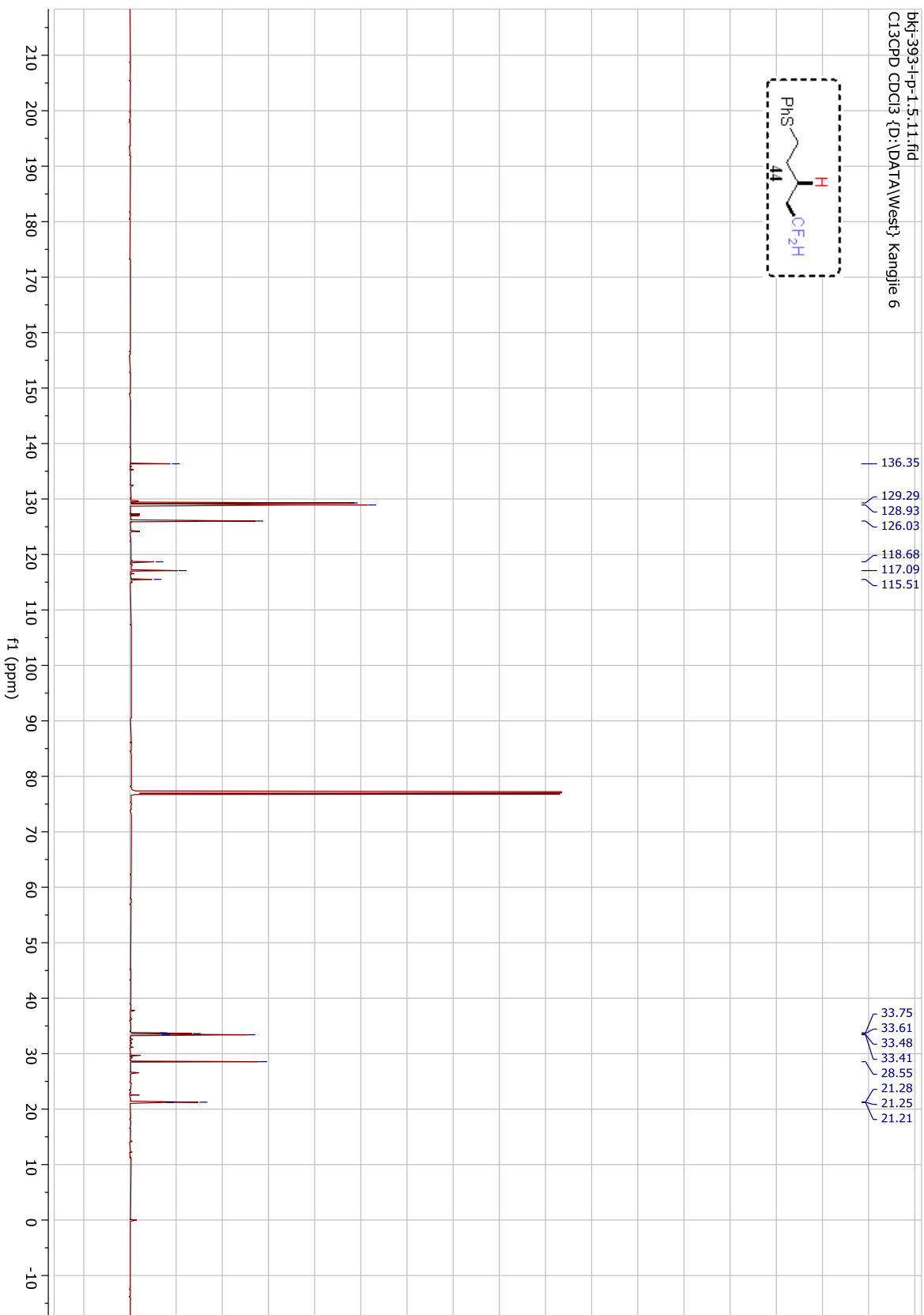
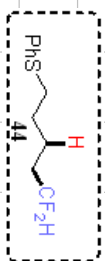


-116.04
-116.07
-116.10
-116.14
-116.17
-116.20

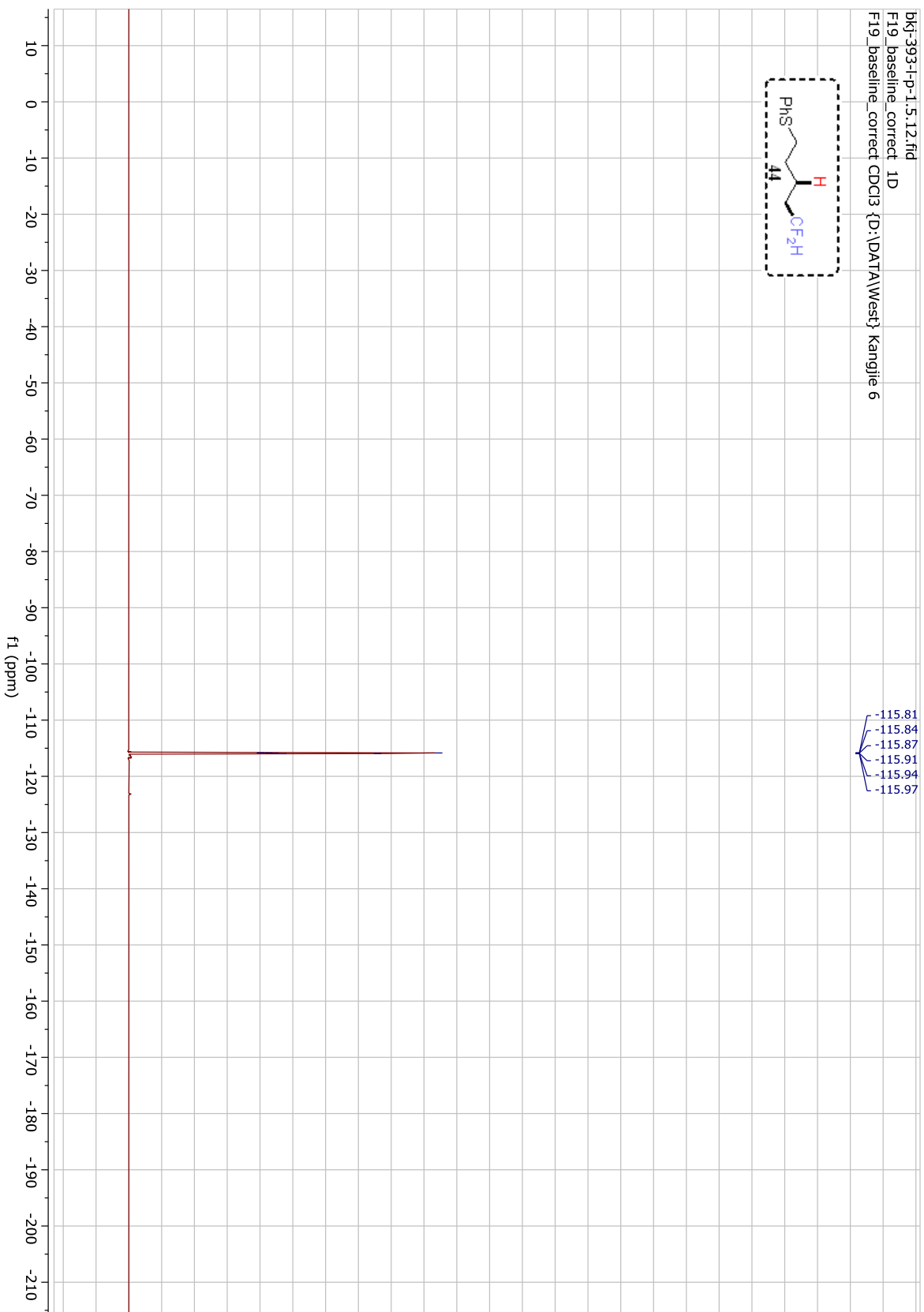
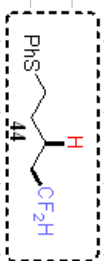




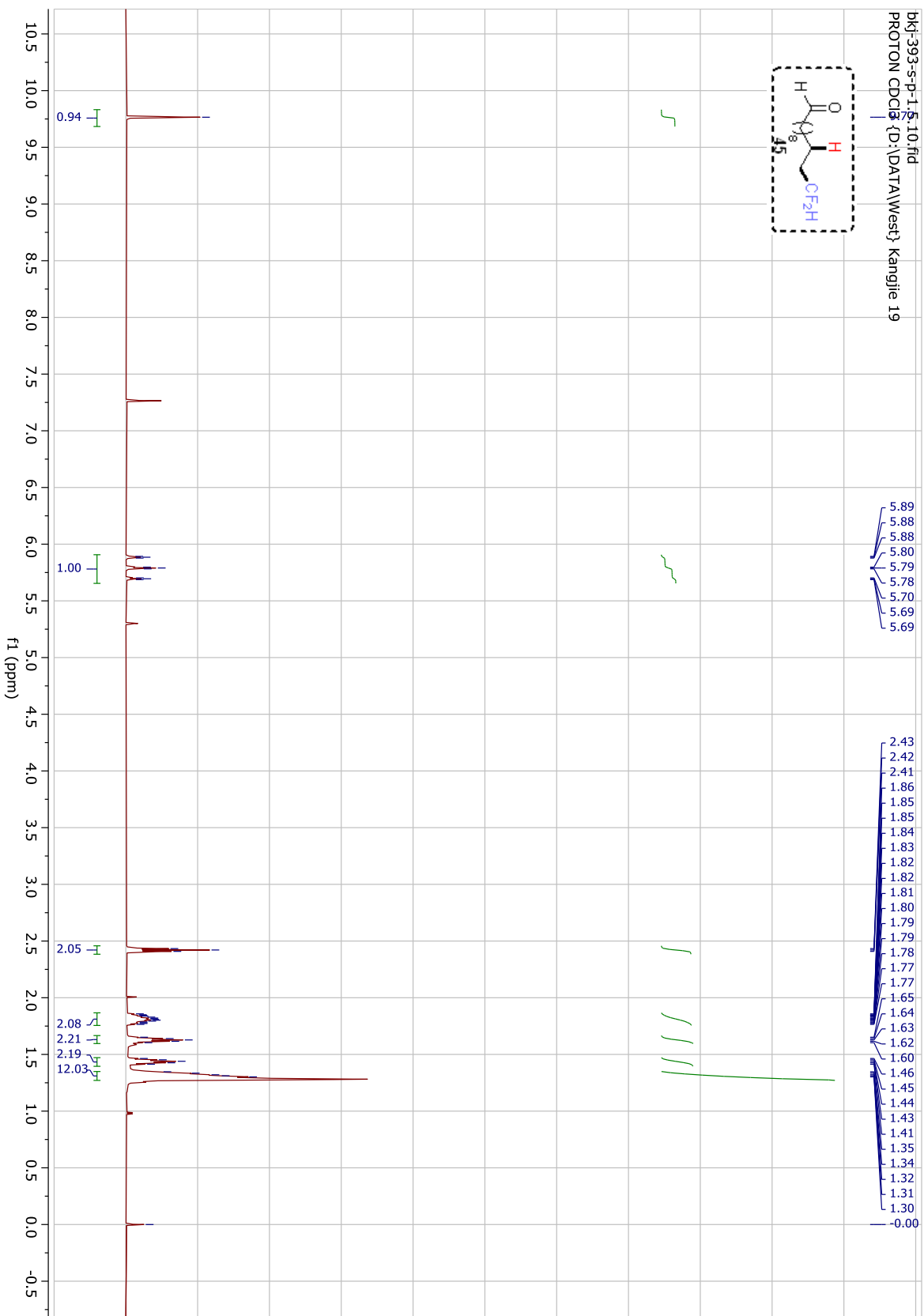
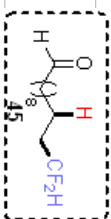
bjl-393-l-p-1-5-11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 6



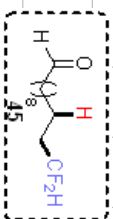
bkj-393-l-p-1.5.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 6



bkl-393-s-p-1-5-10.fid
PROTON CDCl3 {D:\DATA\West\ Kangjie 19

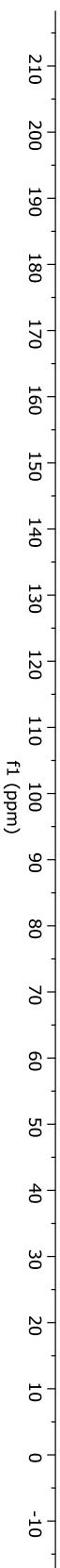


bkf-393-s-p815-11.fid
C13CPD Q823 (D:\DATAWest\ Kangjie 19

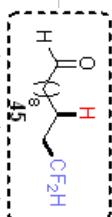


119.08
117.50
115.91

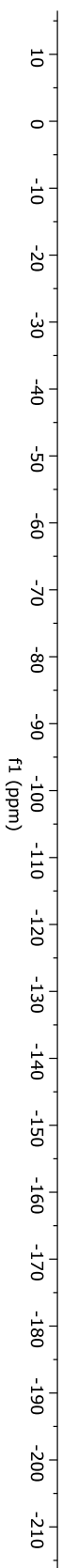
43.92
34.22
34.08
33.95
29.33
29.32
29.14
29.03
22.13
22.10
22.07



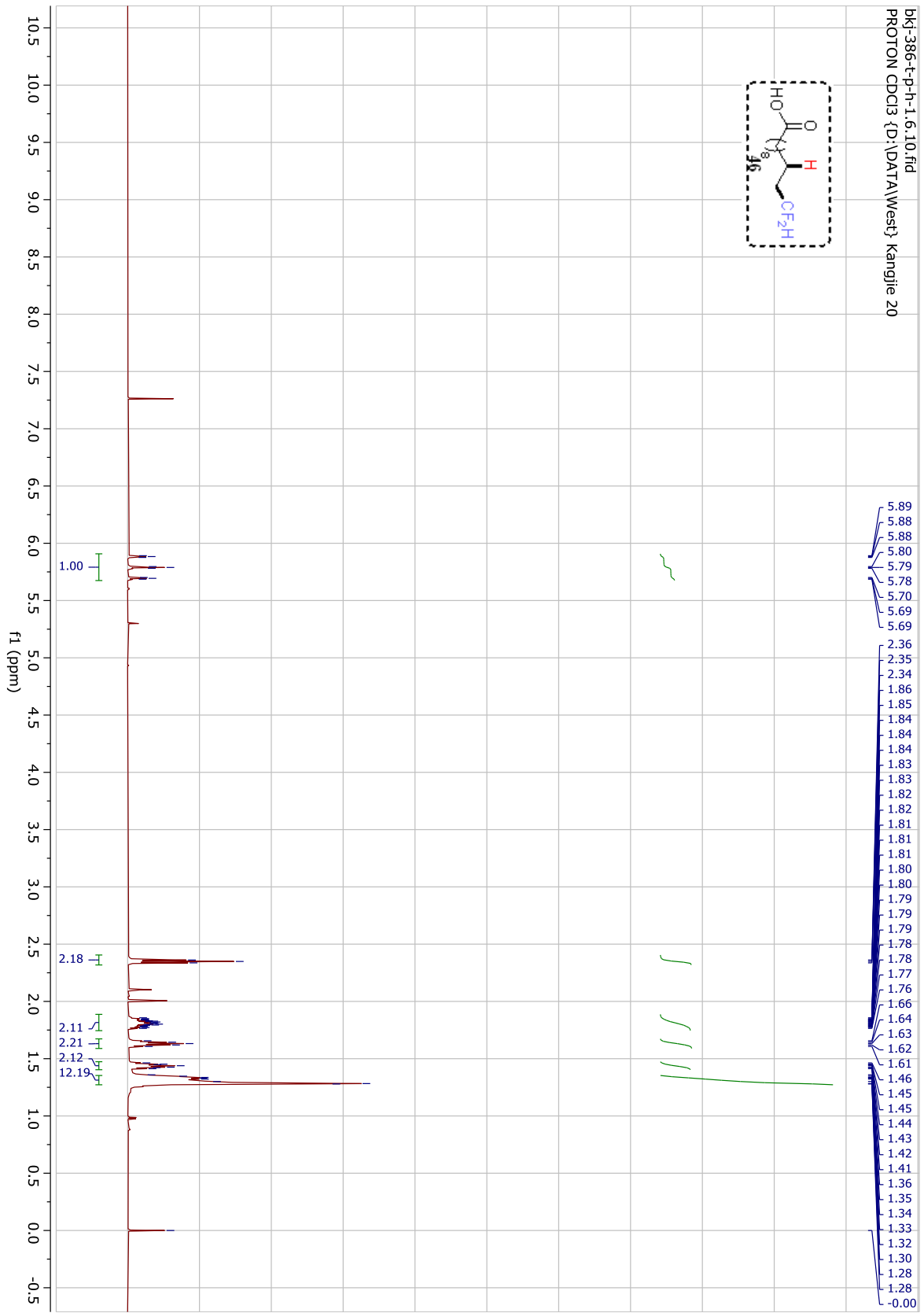
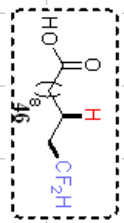
bkj-393-s-p-1.5.12.ftd
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 19



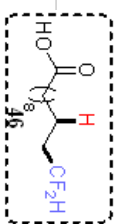
-115.67
-115.70
-115.73
-115.77
-115.80
-115.83



bk1-386-t-p-h-1.6.10.fid
PROTON CDCl3 {D:\DATA\West\ Kangjie 20

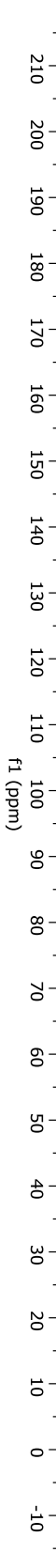


bkt-393-t-p-1-5-11.fid
C13CPD CDCl3 (D:\DATA\W...
Kangjie 20

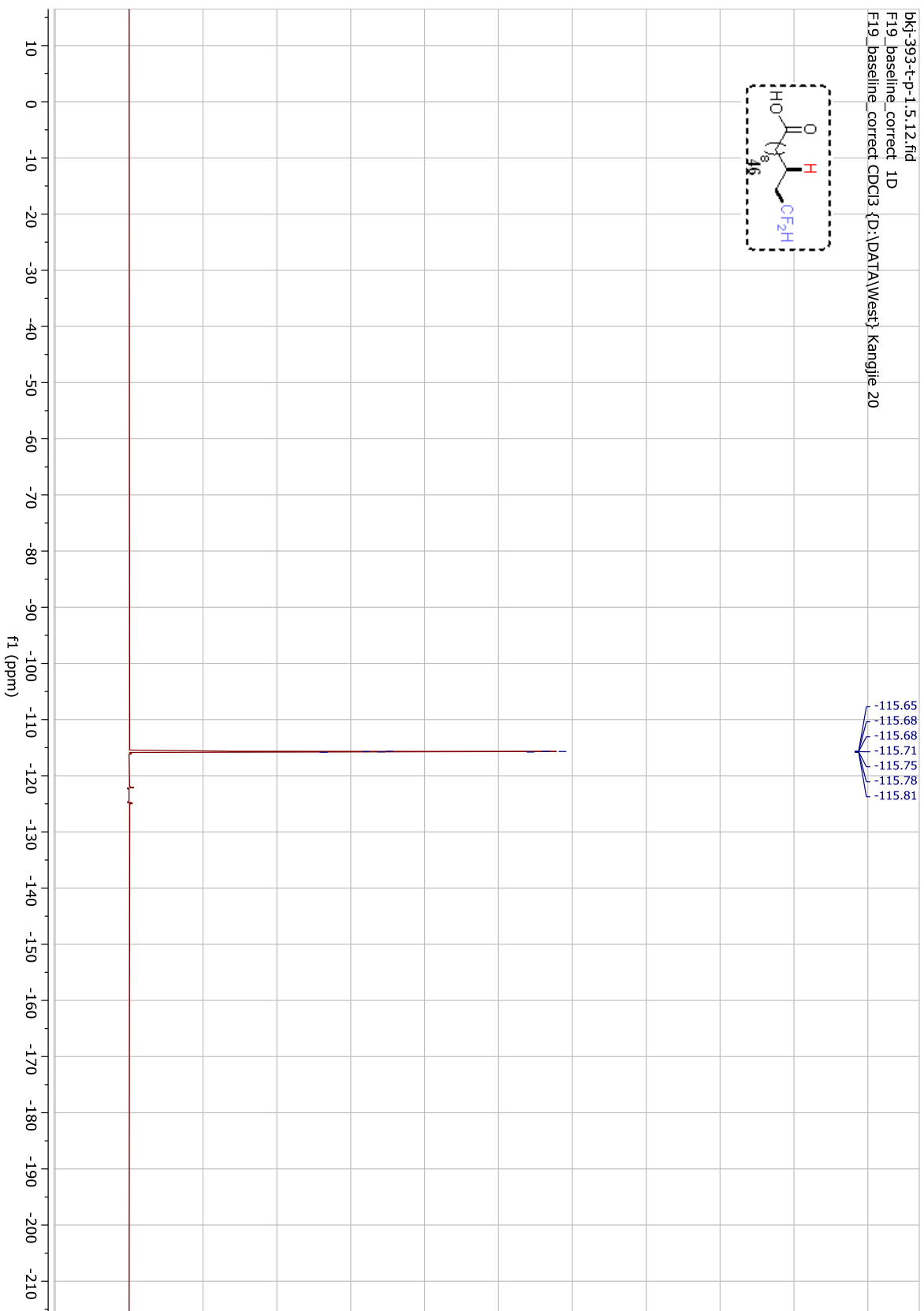
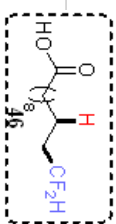


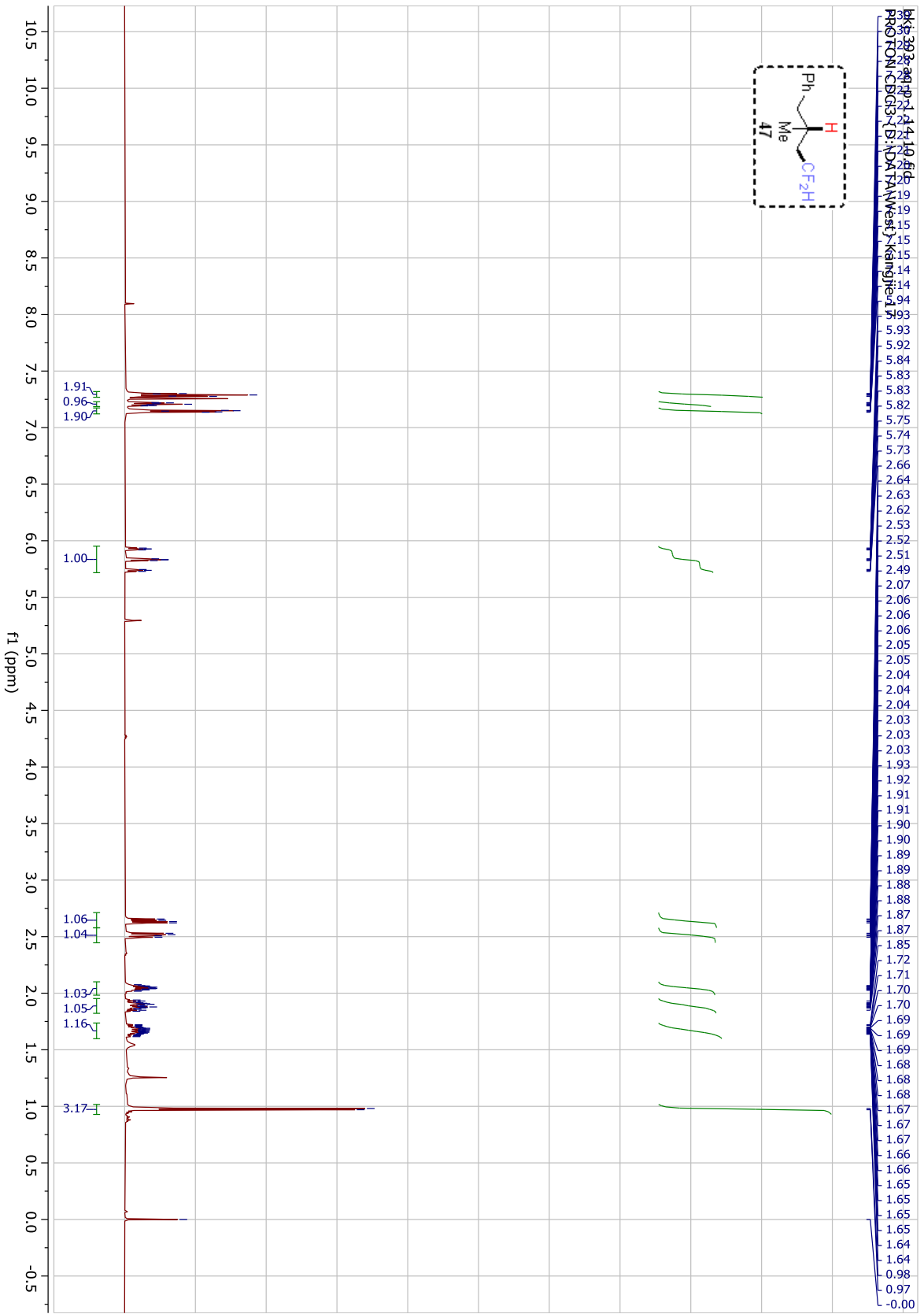
119.08
117.50
115.92

34.22
34.08
34.03
33.95
29.32
29.18
29.03
24.65
22.14
22.10
22.07

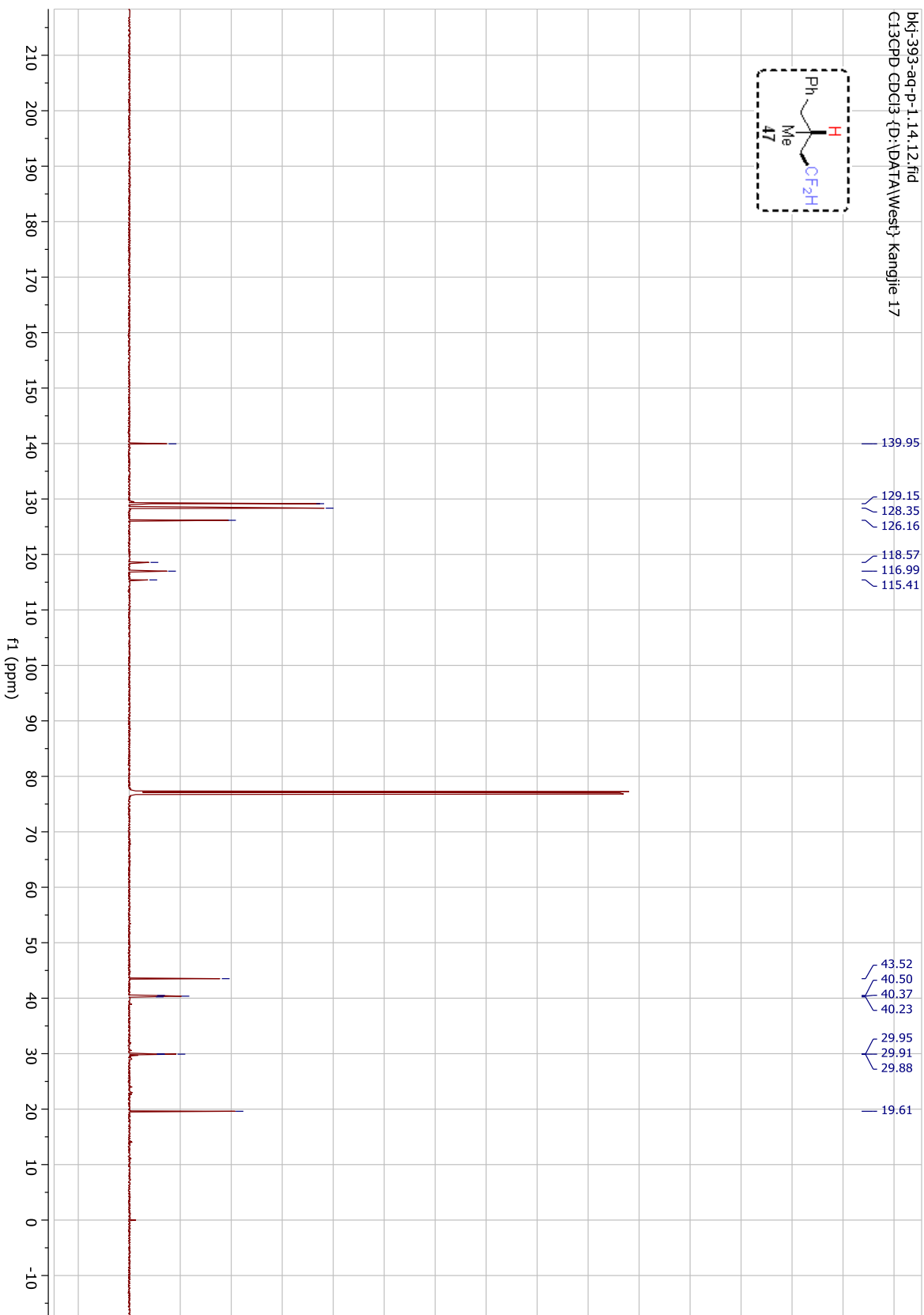
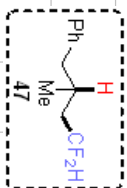


bkj-393-t-p-1.5.12.ftd
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 20

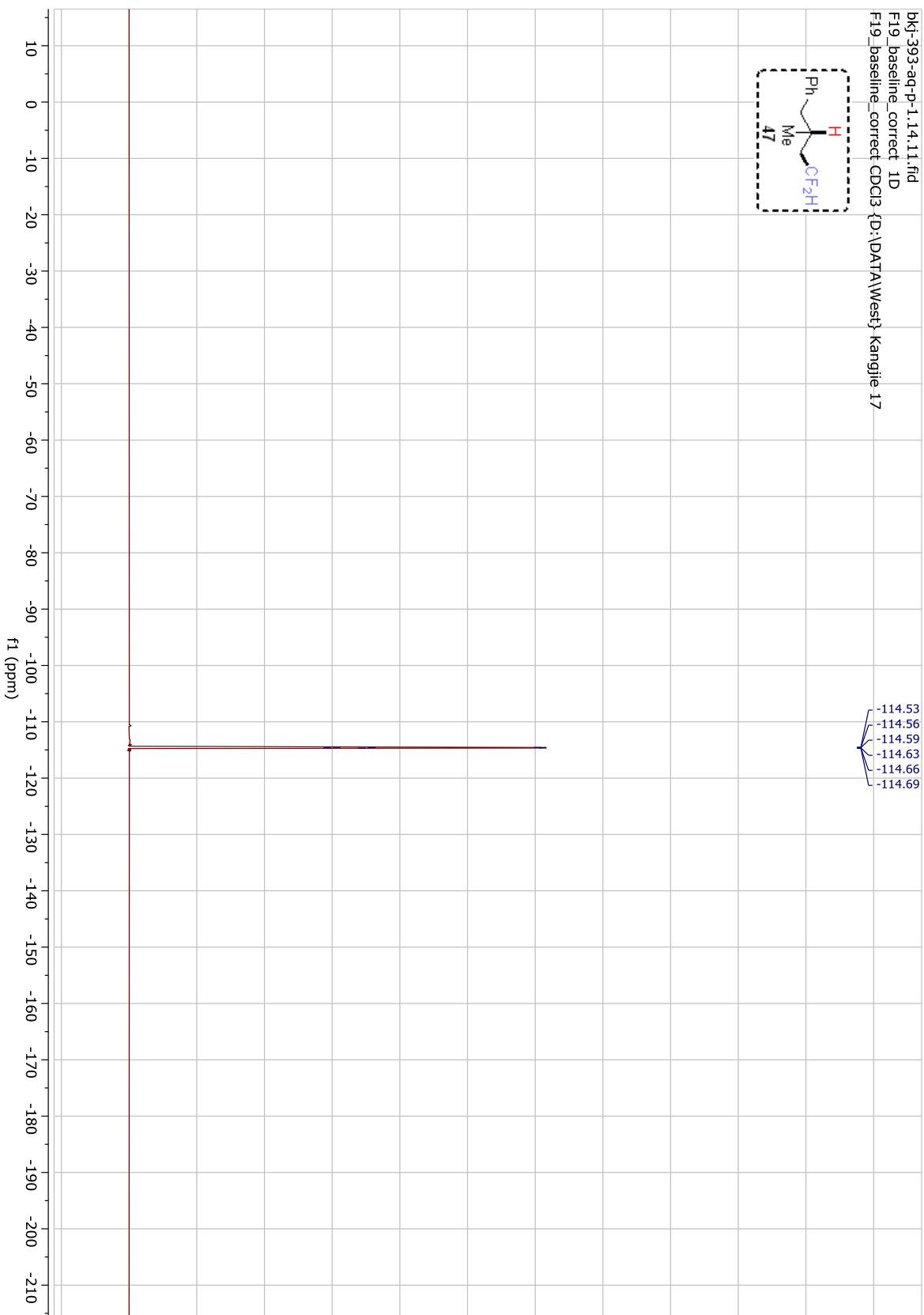
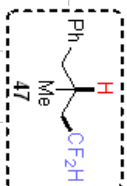


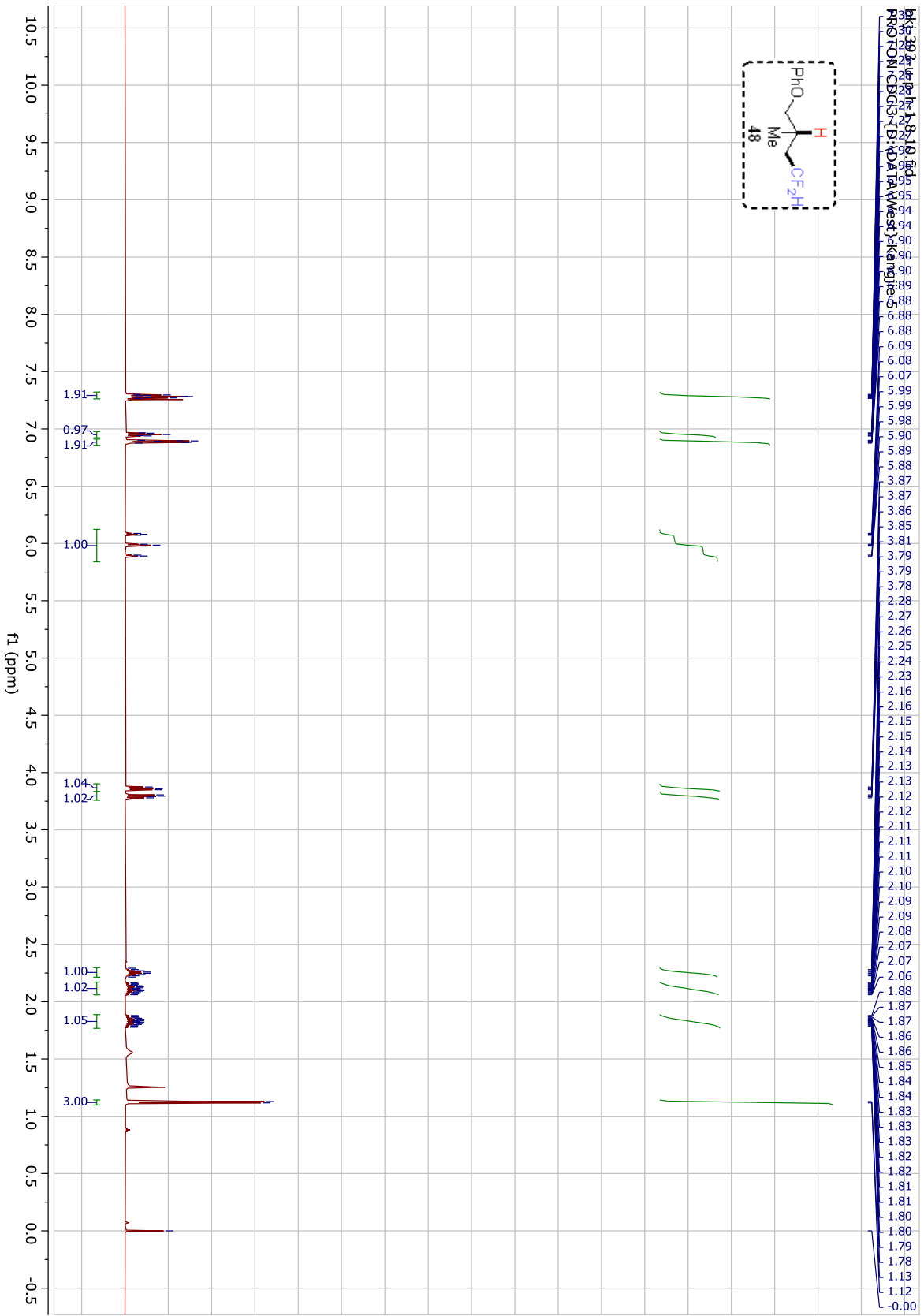


bkj-393-aq-p-1.14.12.fid
C13CPD CDCl3 (D:\DATA\Westj kangjie 17

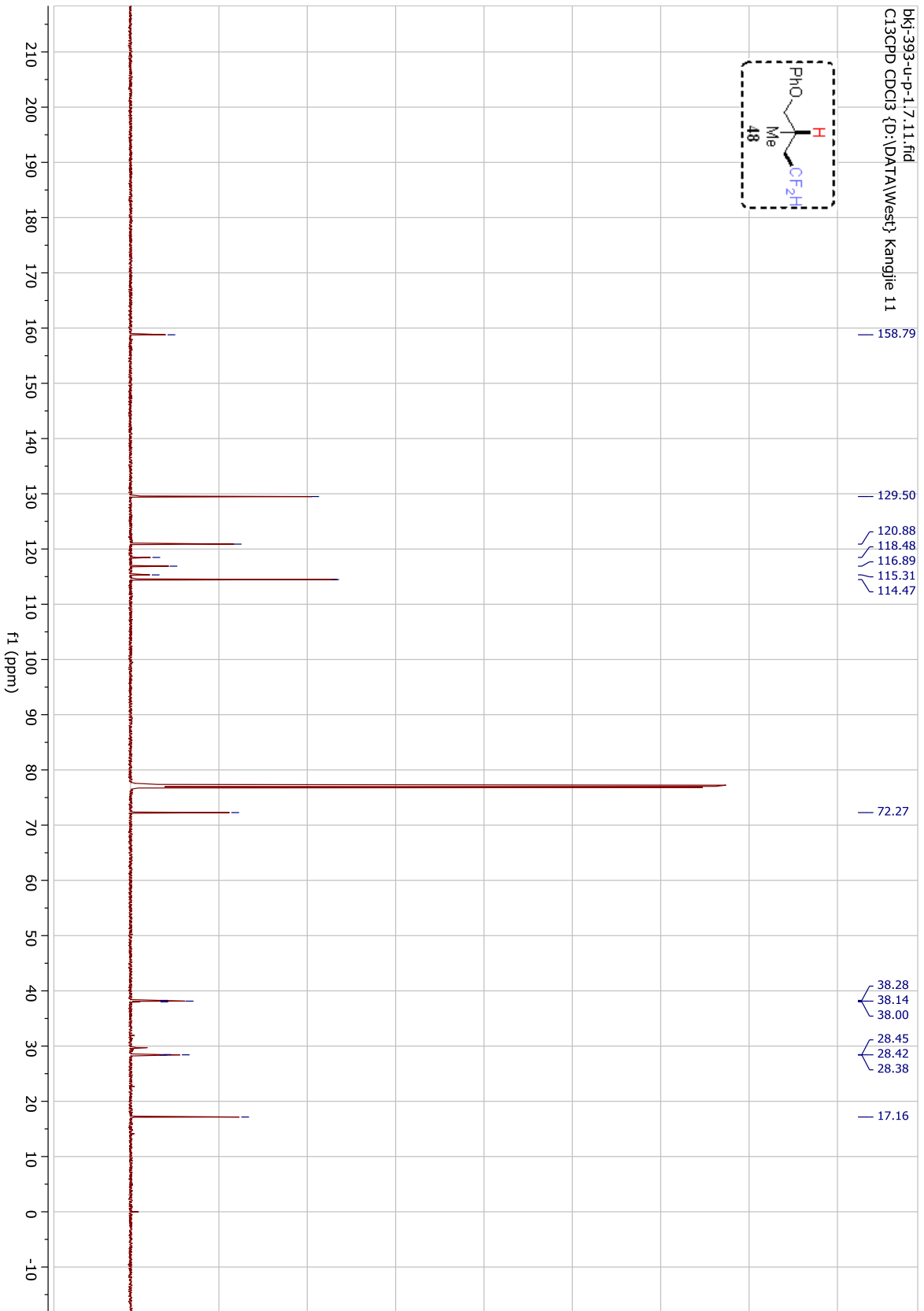
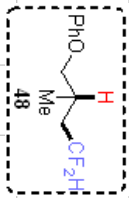


bkj-393-aq-p-1.14.11.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West}\Kangjie 17

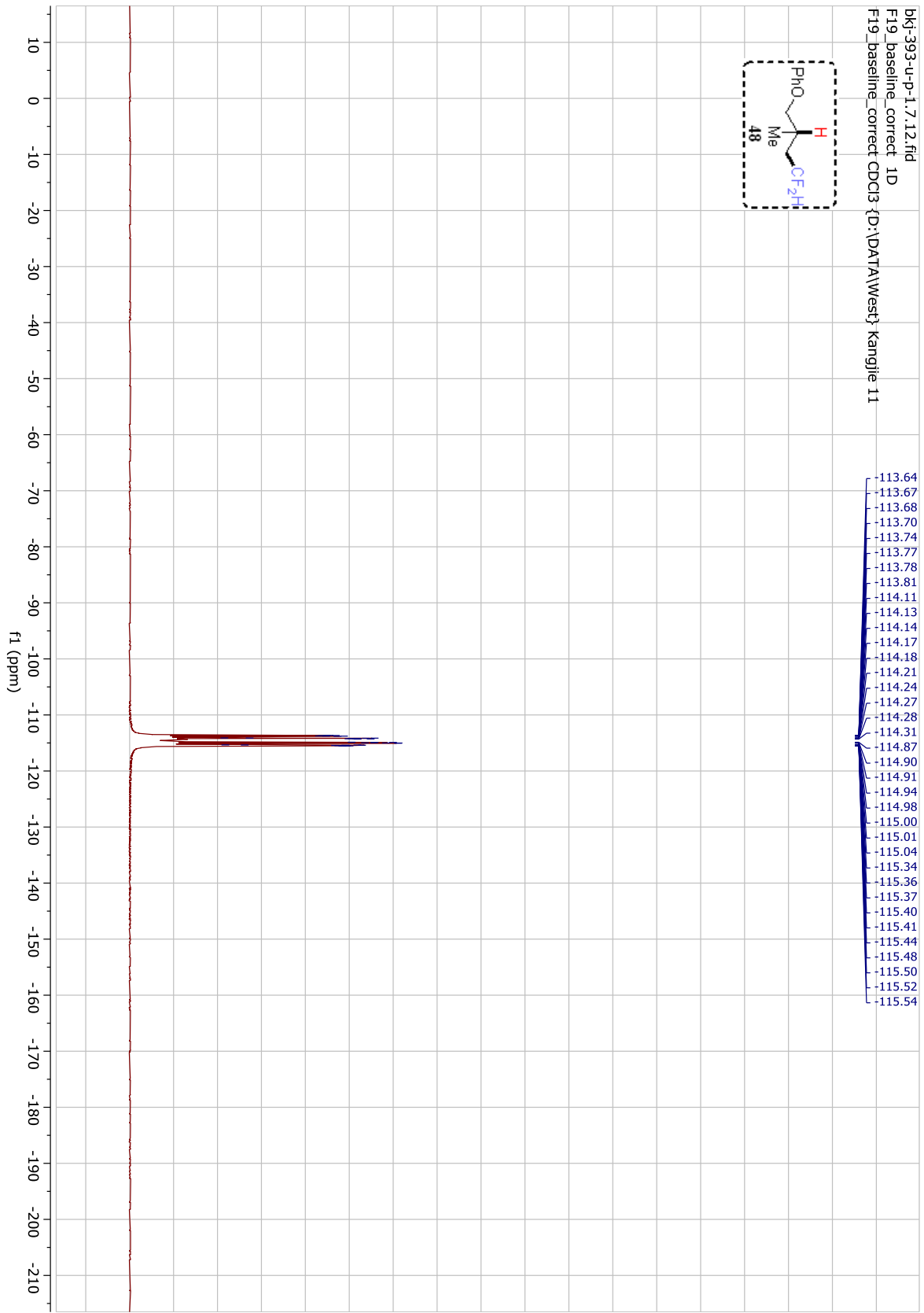
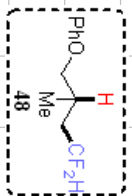


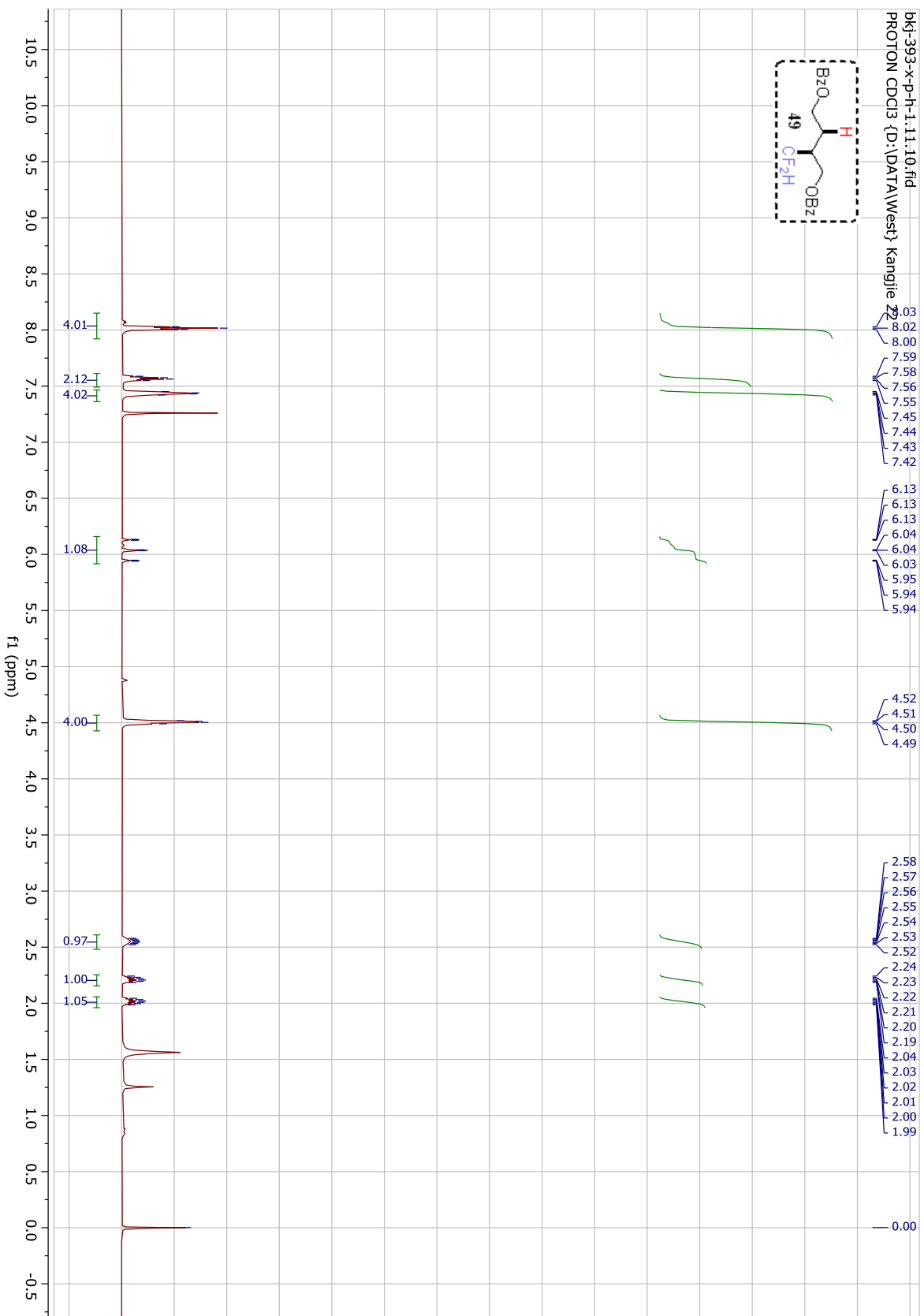


bkt-393-u-p-17.11.fid
C13CPD CDCl3 (D:\DATAWest\ Kangjie 11

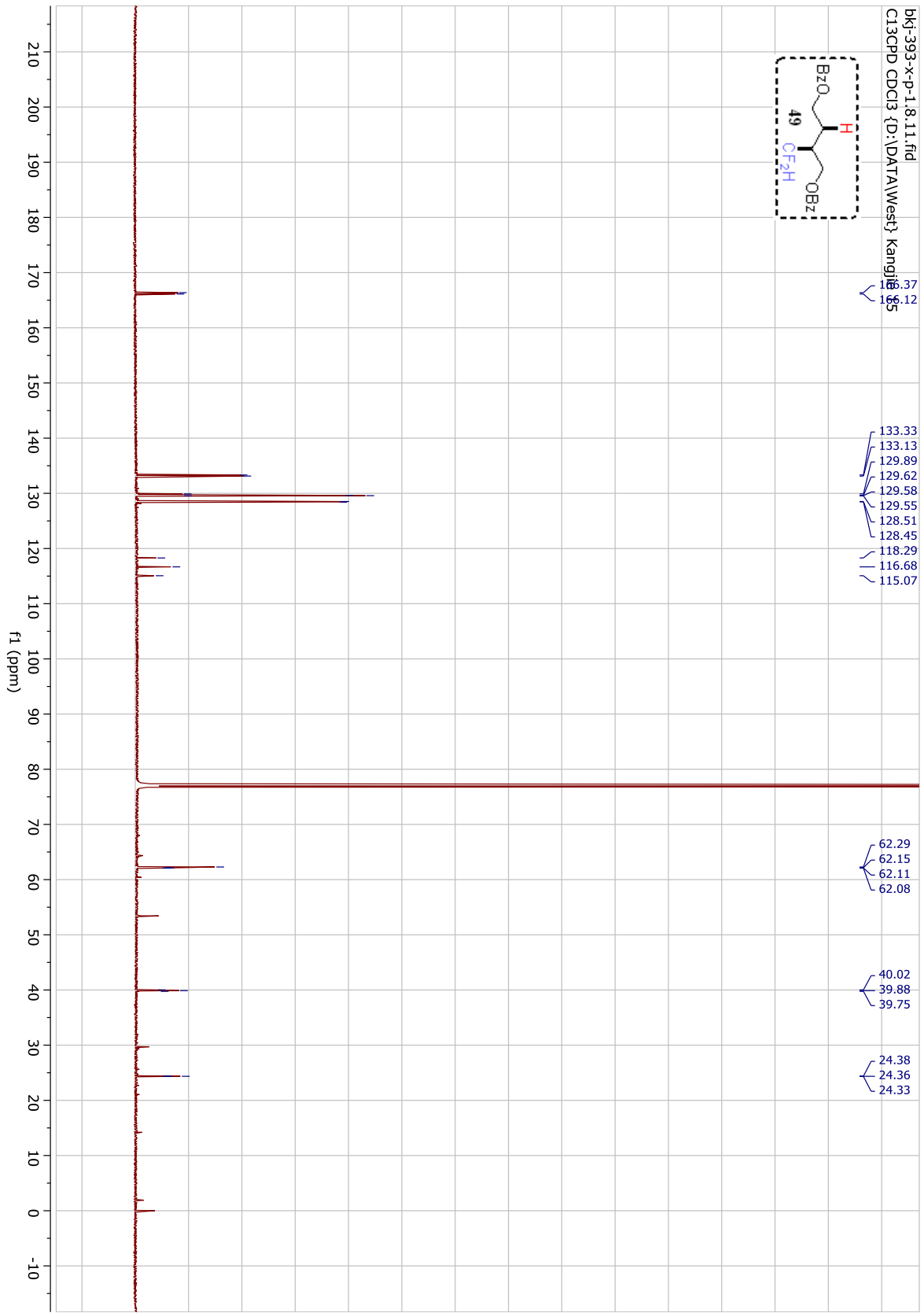
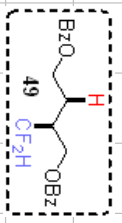


bkt-393-u-p-1.7.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 11

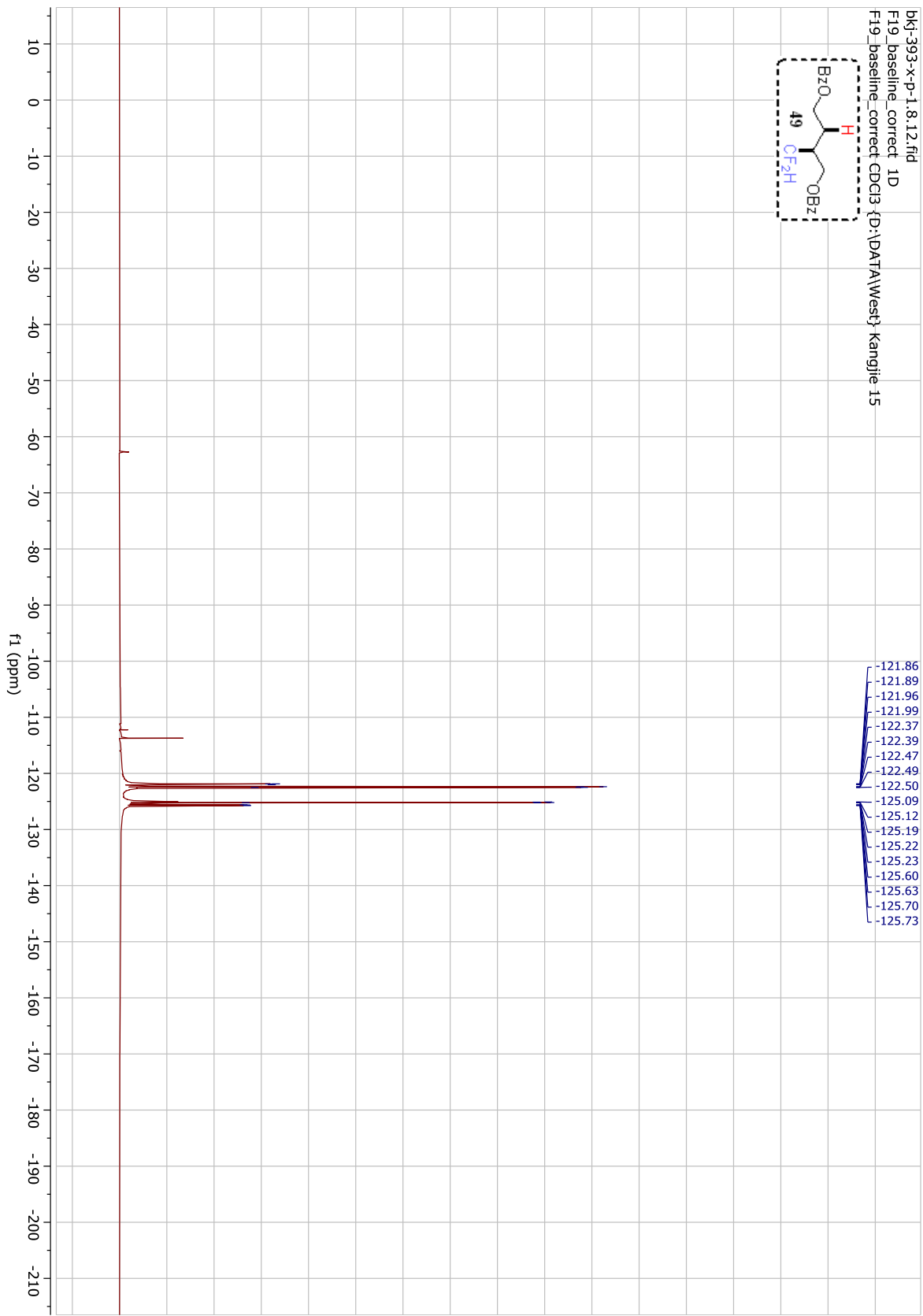
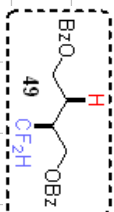


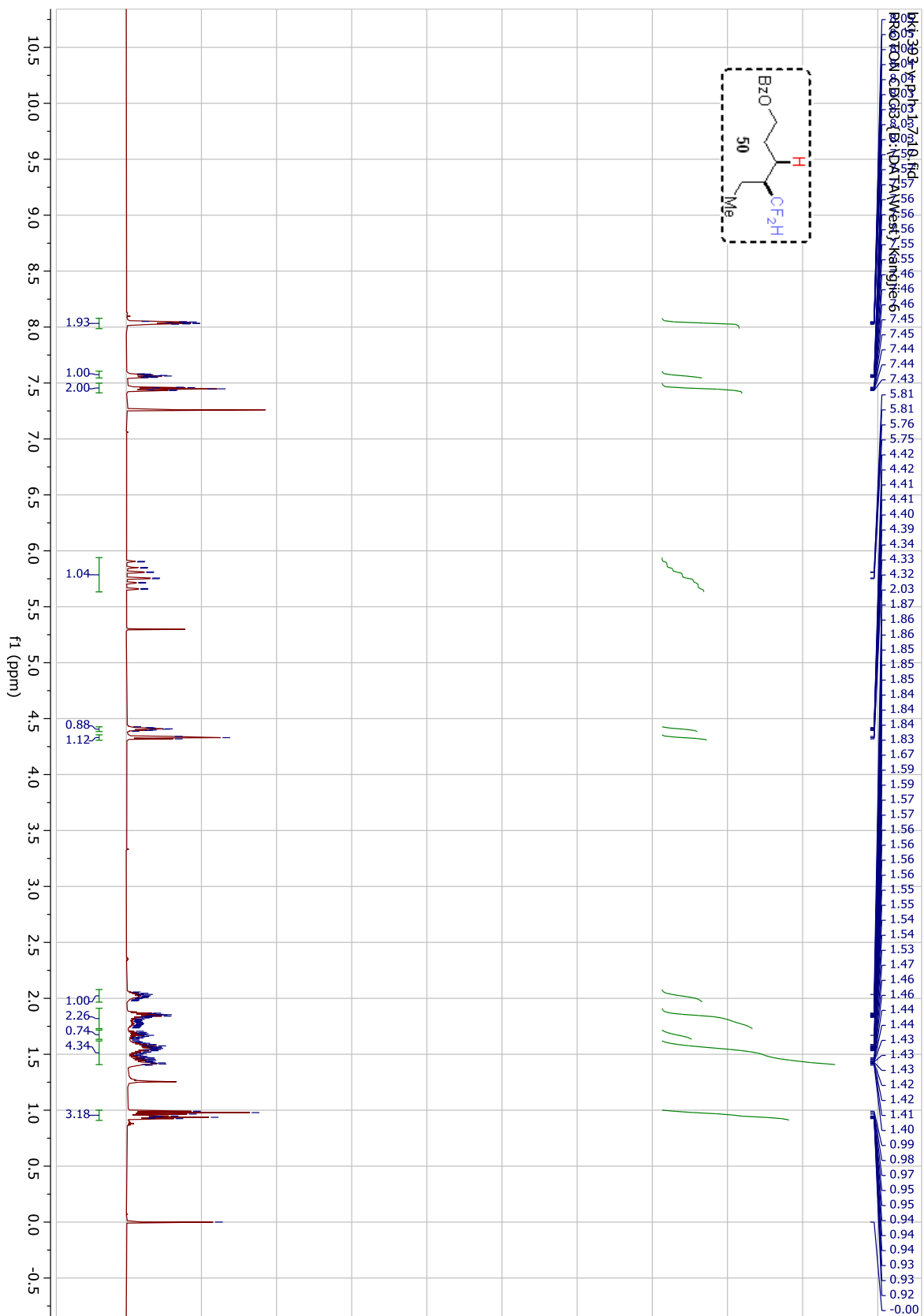


bkj-393-x-p-1-8-11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjil)

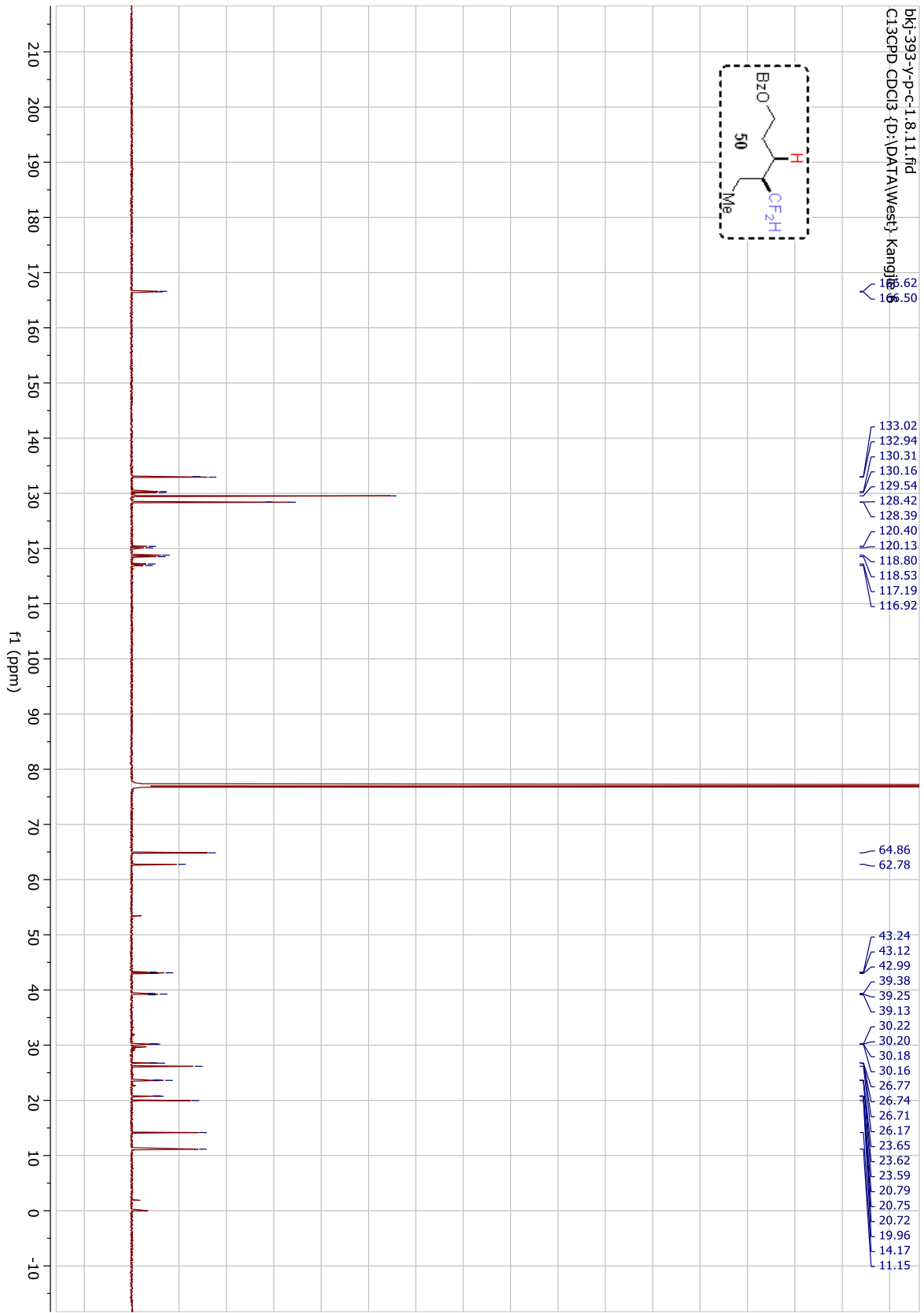
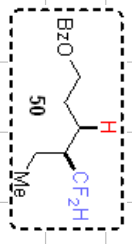


bkj-393-x-p-1.8.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 15

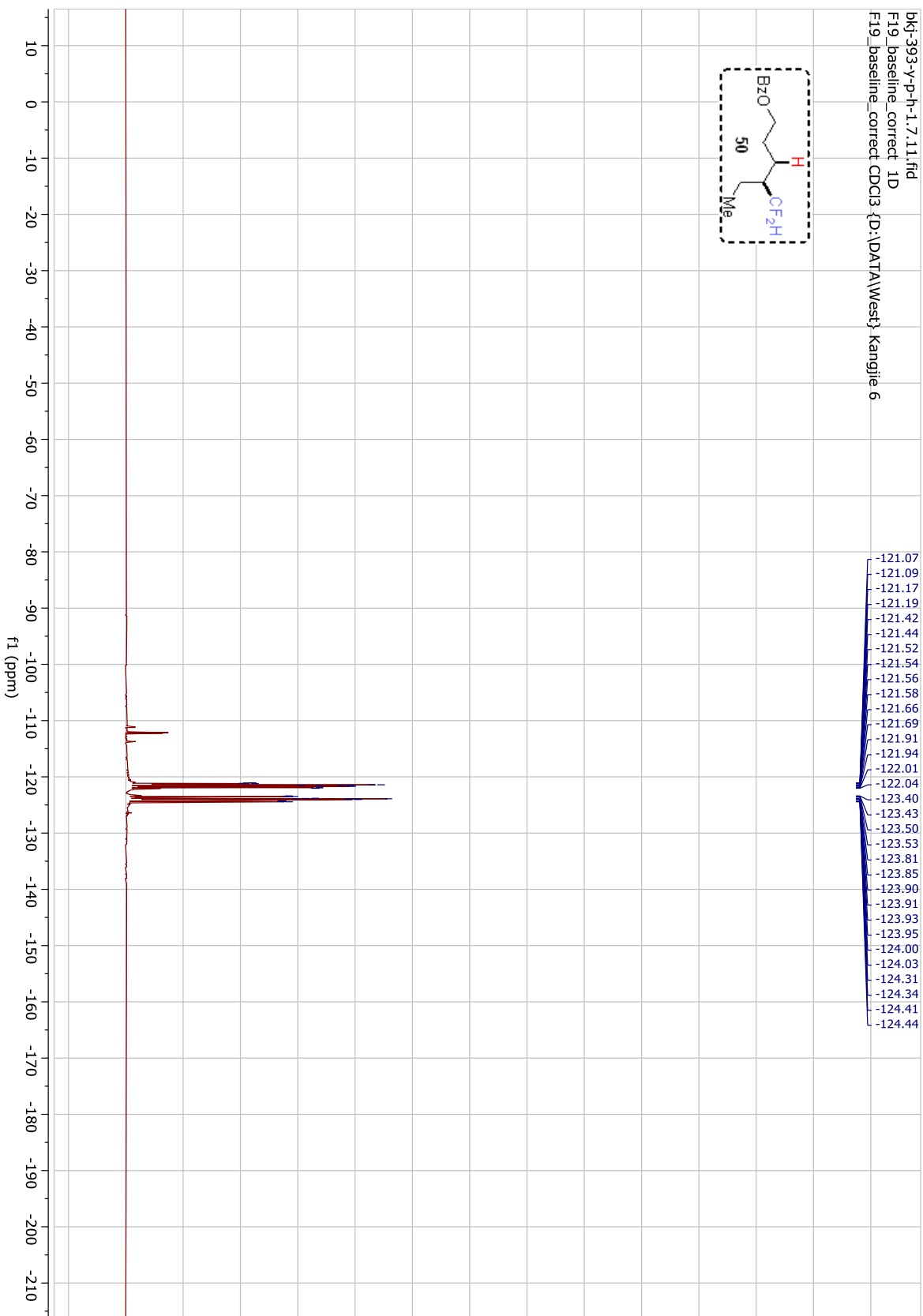
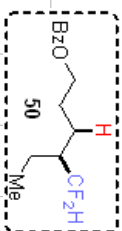


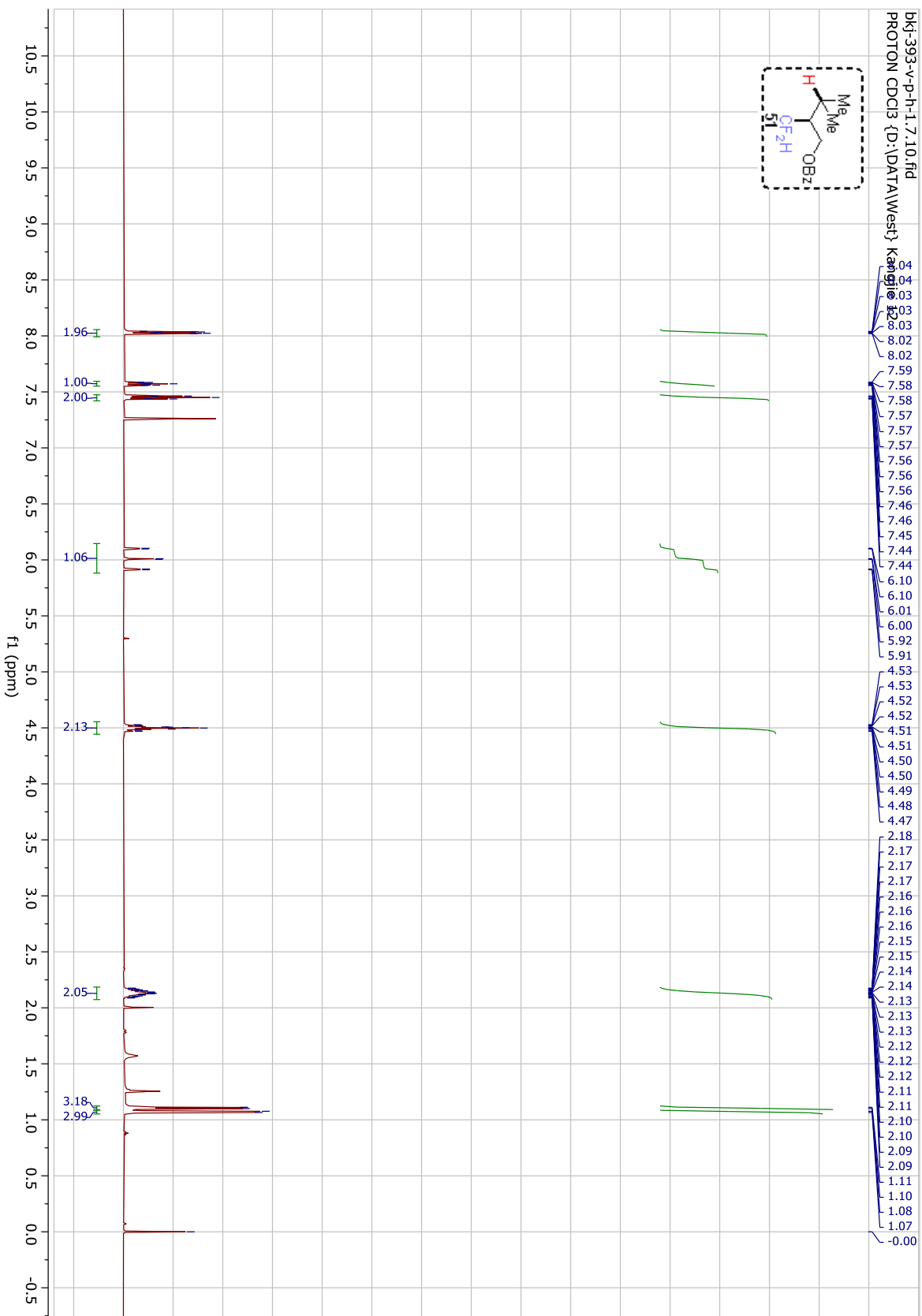


bjl-393-y-p-c-1,8,11.fid
C13CPD CDCl3 (D:\DATA\Westj_Kangjil)

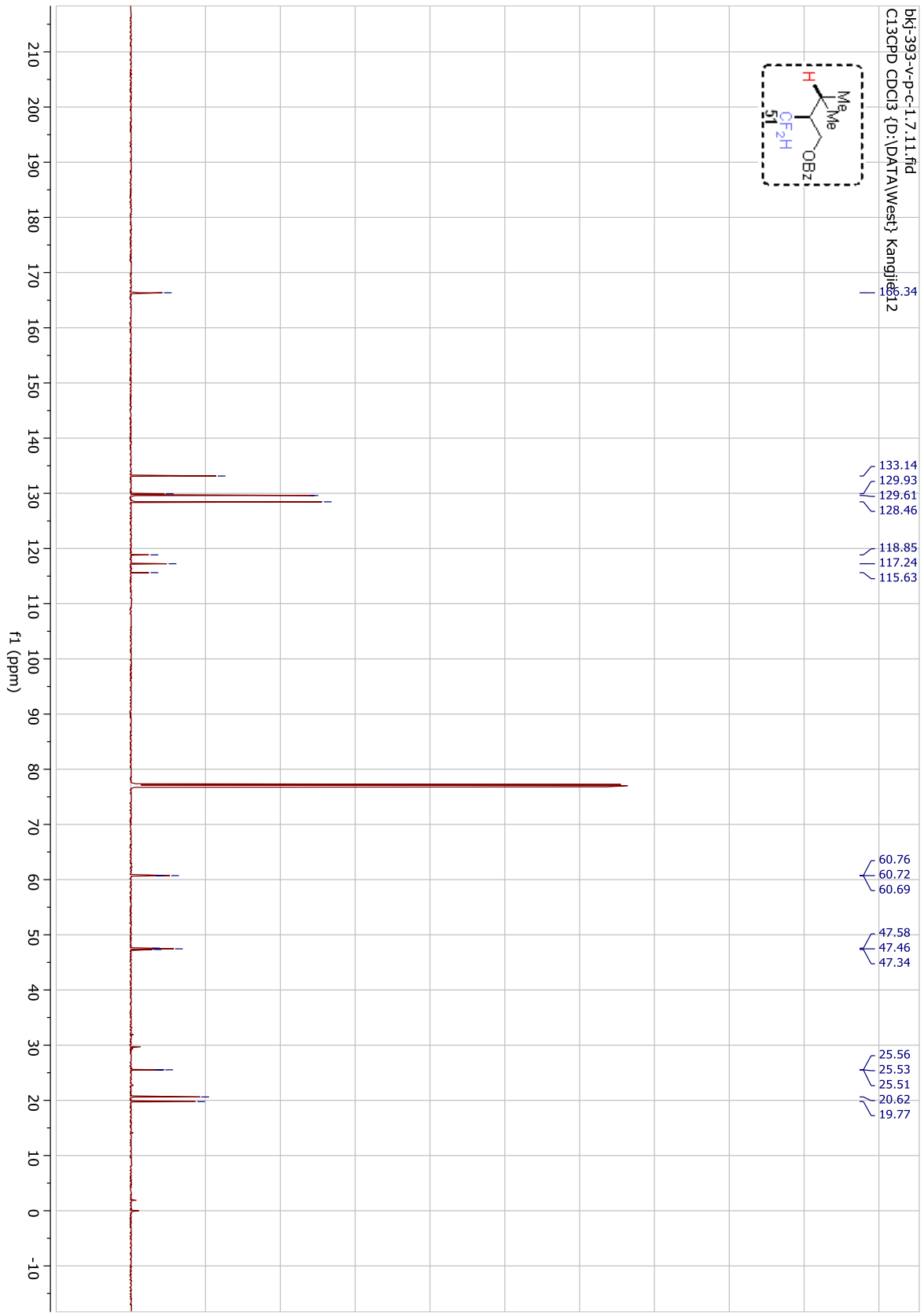
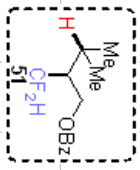


bkj-393-y-p-h-1-7.11.fid
F19_baseline_correct_1D
F19_baseline_correct_CDCl3 {D:\DATA\West} Kangjie 6

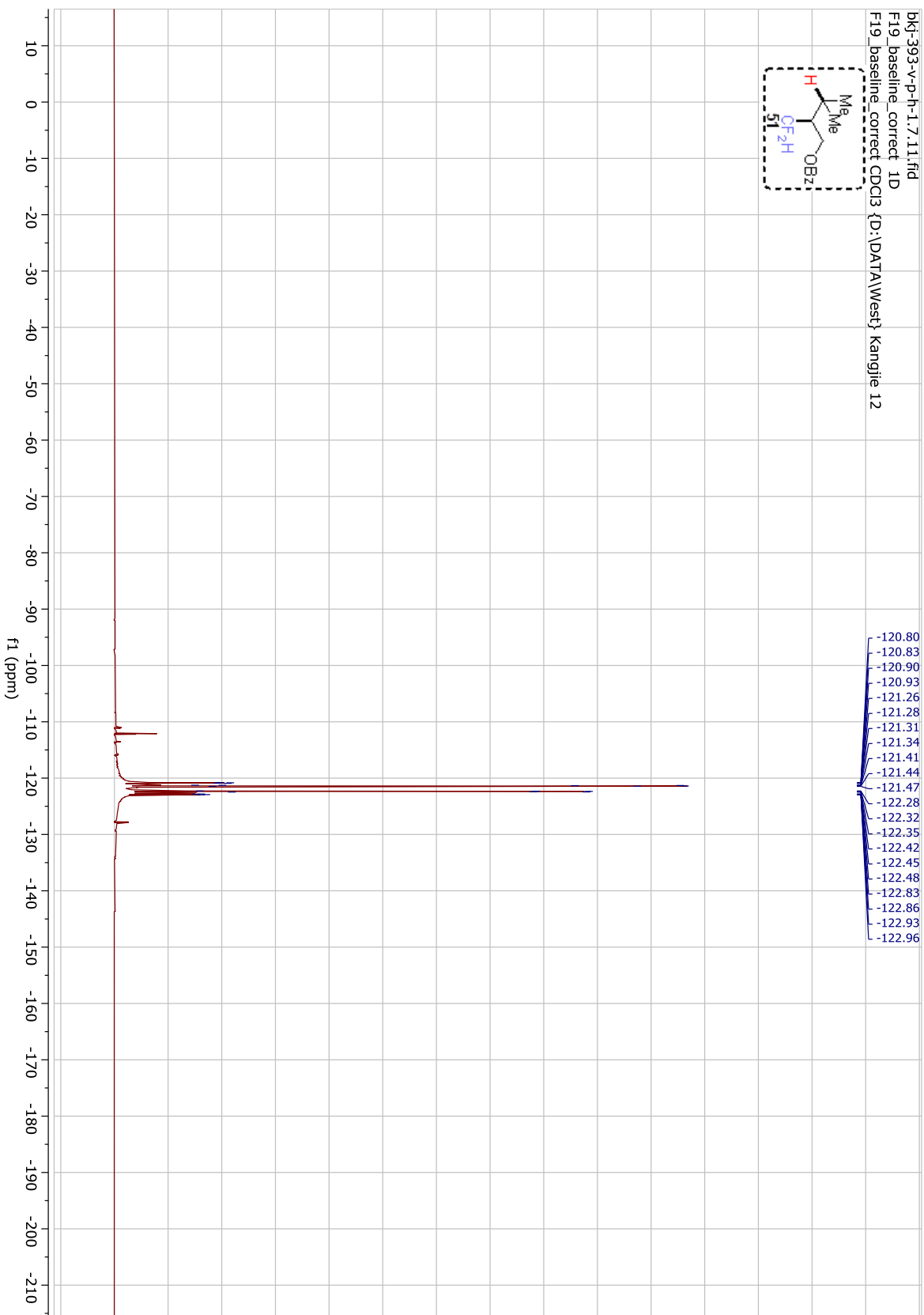
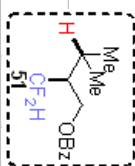




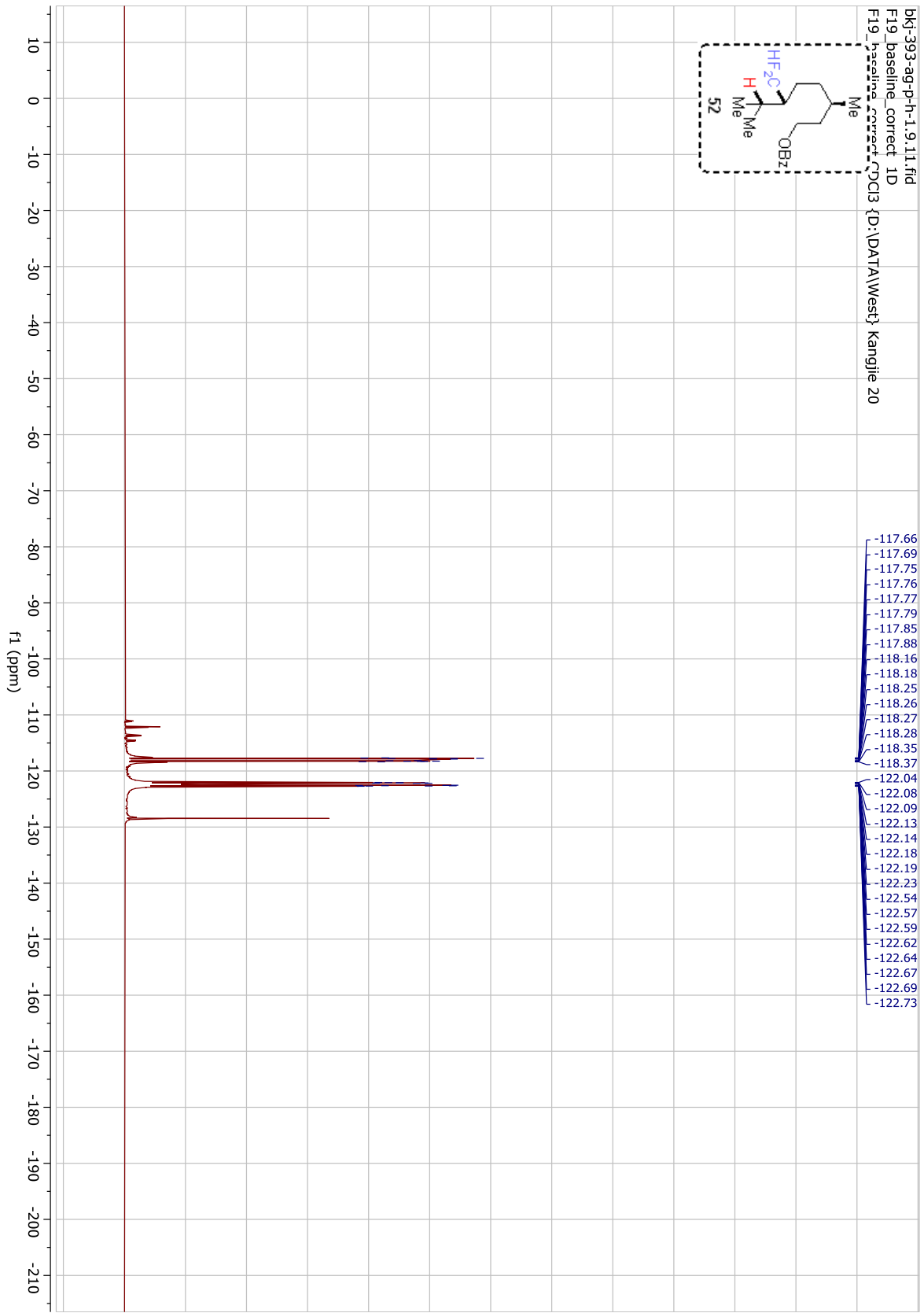
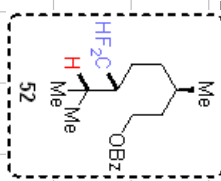
bkt-393-v-p-c-1.7.11.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjil) 812

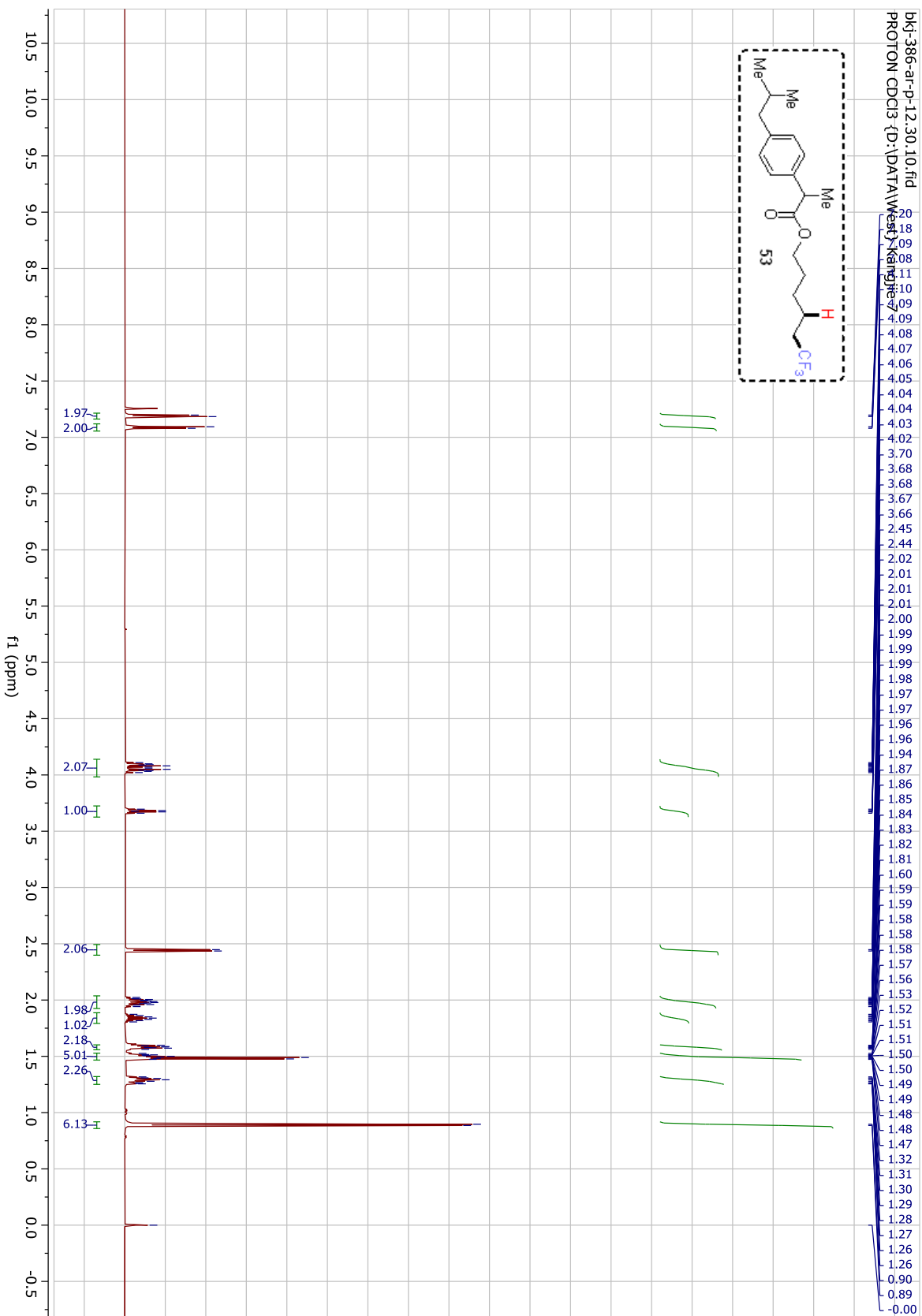


bkj-393-v-p-h-1.7.11.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 12

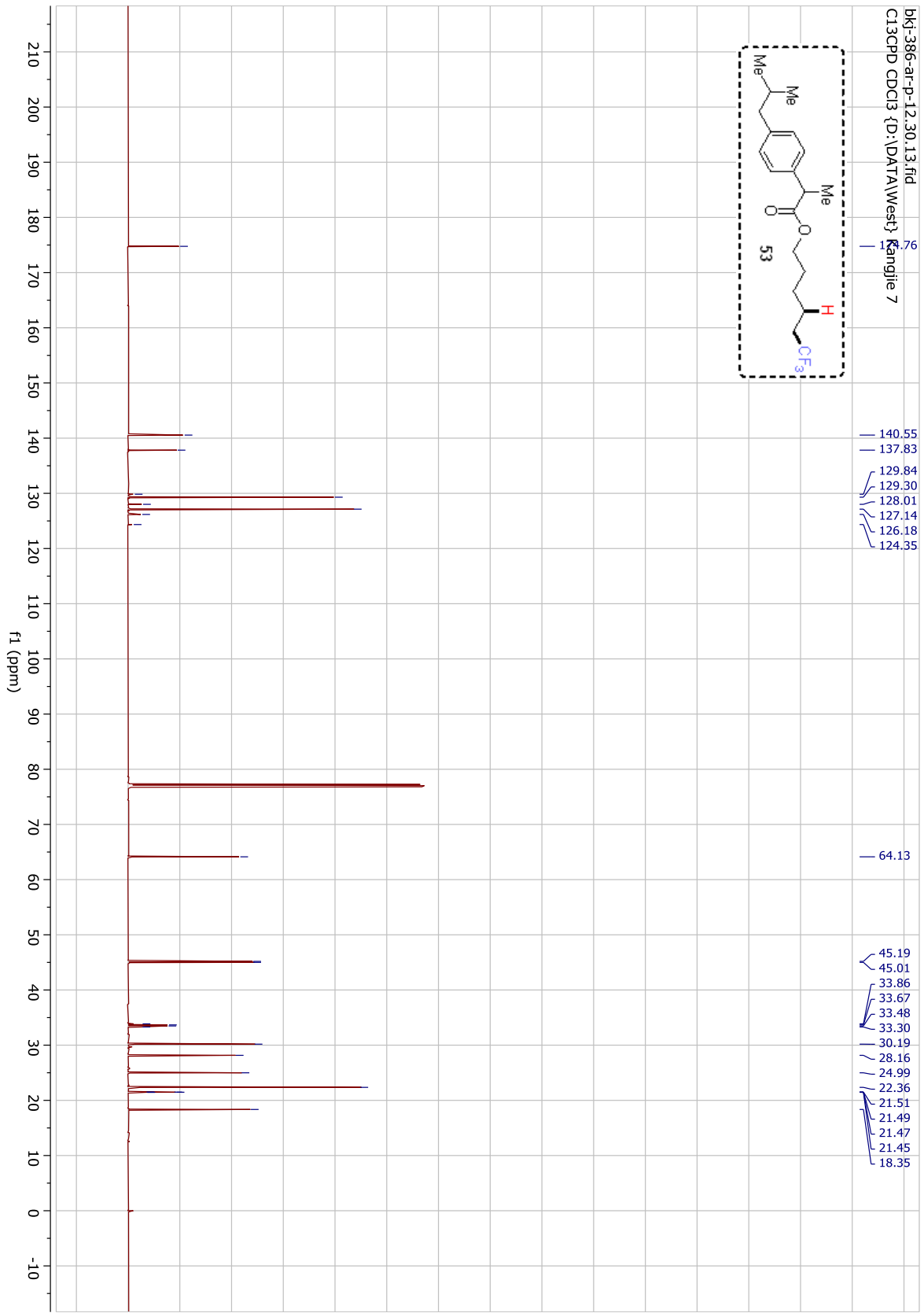
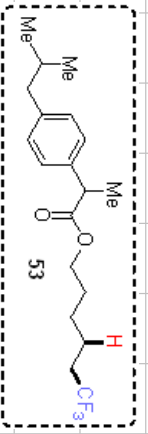


bjt-393-ag-p-h-1.9.11.fid
F19_baseline_correct_1D
F19_baseline_correct_1DCl3 {D:\DATA\West}\Kangjie 20

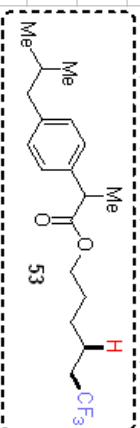




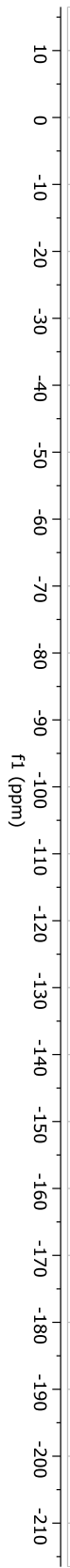
bk1-386-ar-p-12.30.13.fid
C13CPD CDCl3 (D:\DATA\West)\Kangjie 7

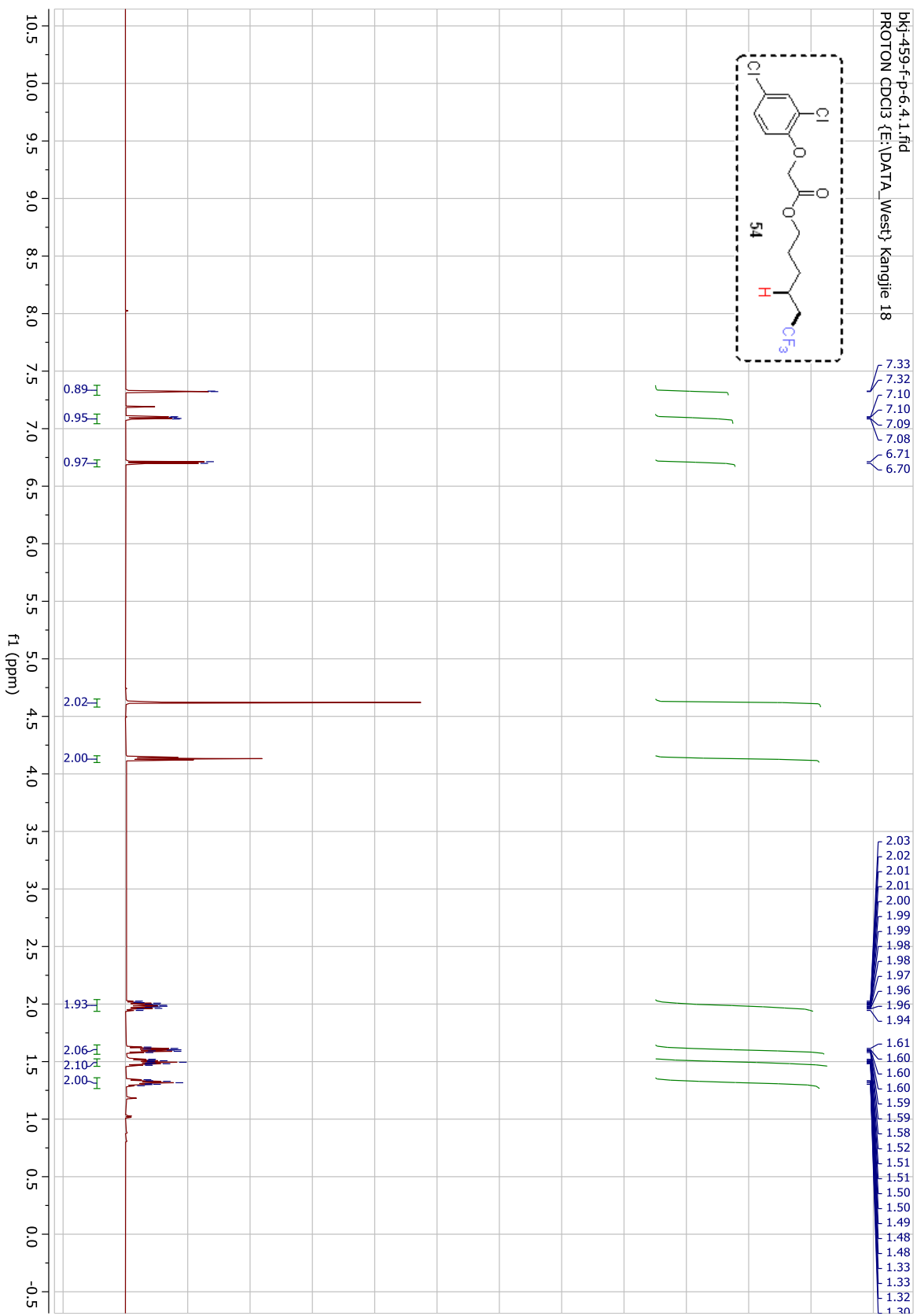


bkj-386-ar-p-12.30.12.ftd
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 7

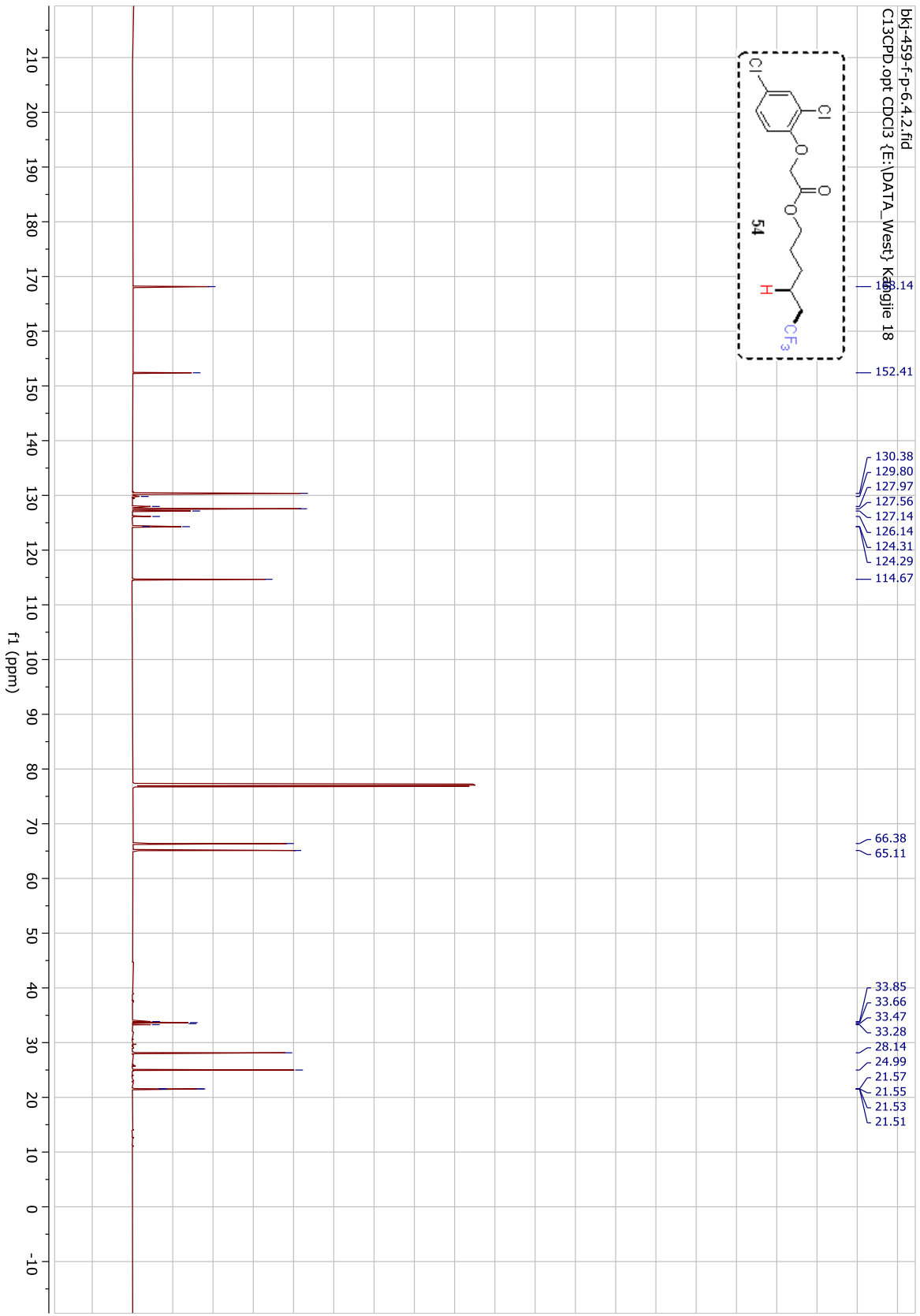
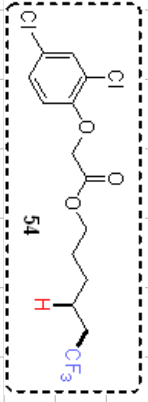


-66.40
-66.42
-66.44



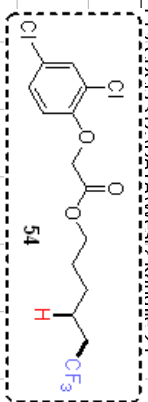


bkj-459-f-p-6-4-2.fid
C13CPD.opt CDCl3 {E:\DATA_Westf\Kaggle 18

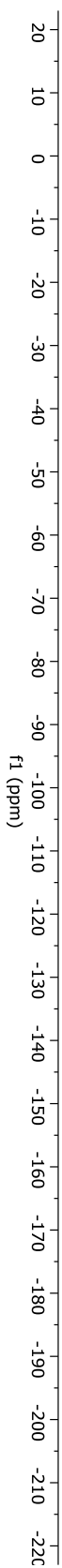


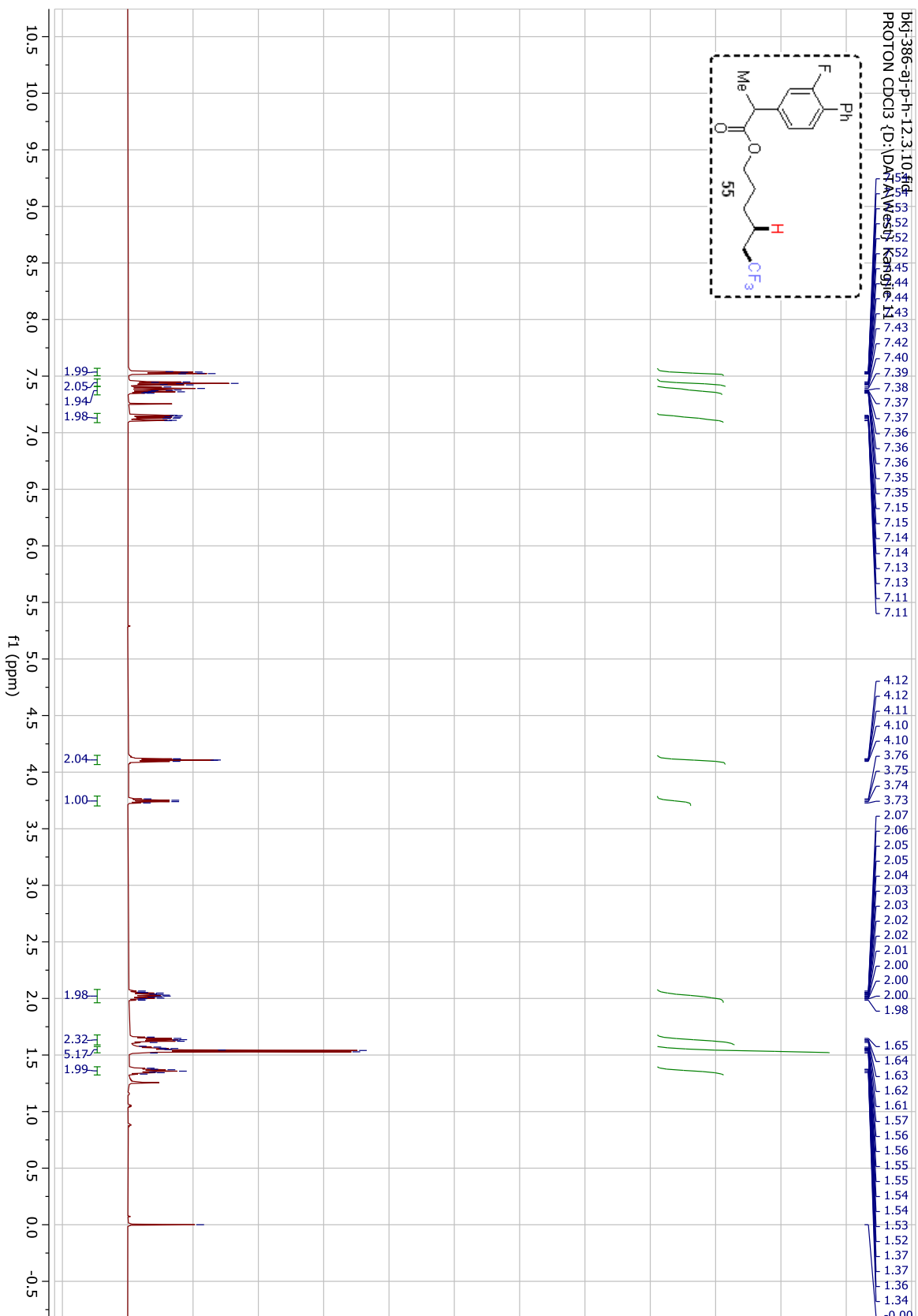
bkj-459-f-p-f-6.5.10.ftd
F19

CMC_F19.CDCI3 (D:\DATA\West\Kantile 21

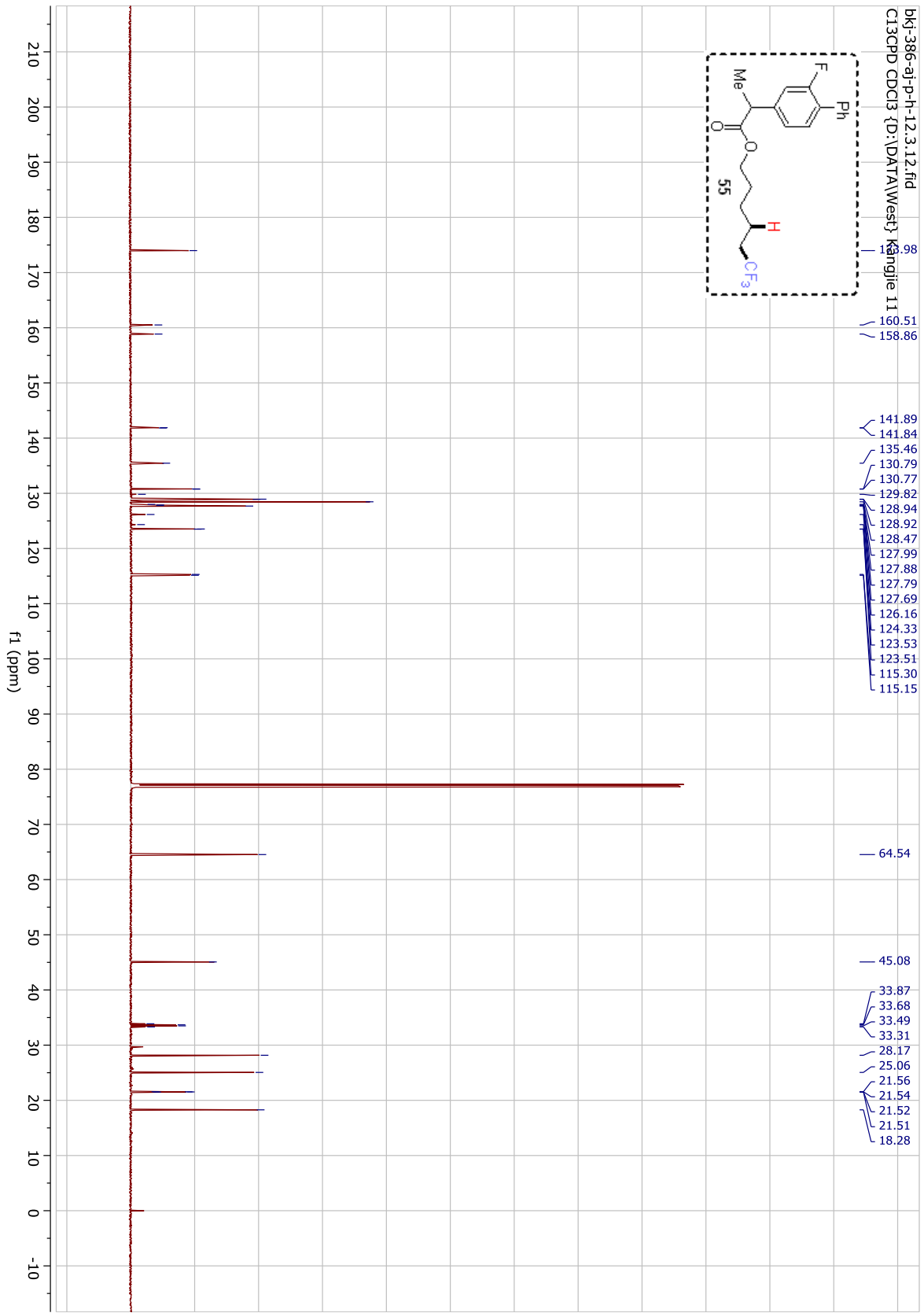
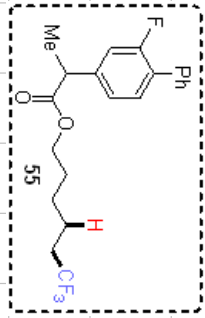


-66.34
-66.36
-66.38

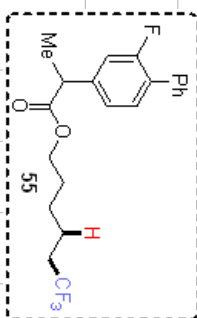




bj1-386-aj-p-h-12.3.12.fid
C13CPD CDCl3 (D:\DATA\Westy K\Engle 11

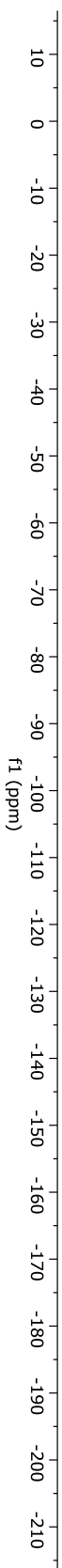


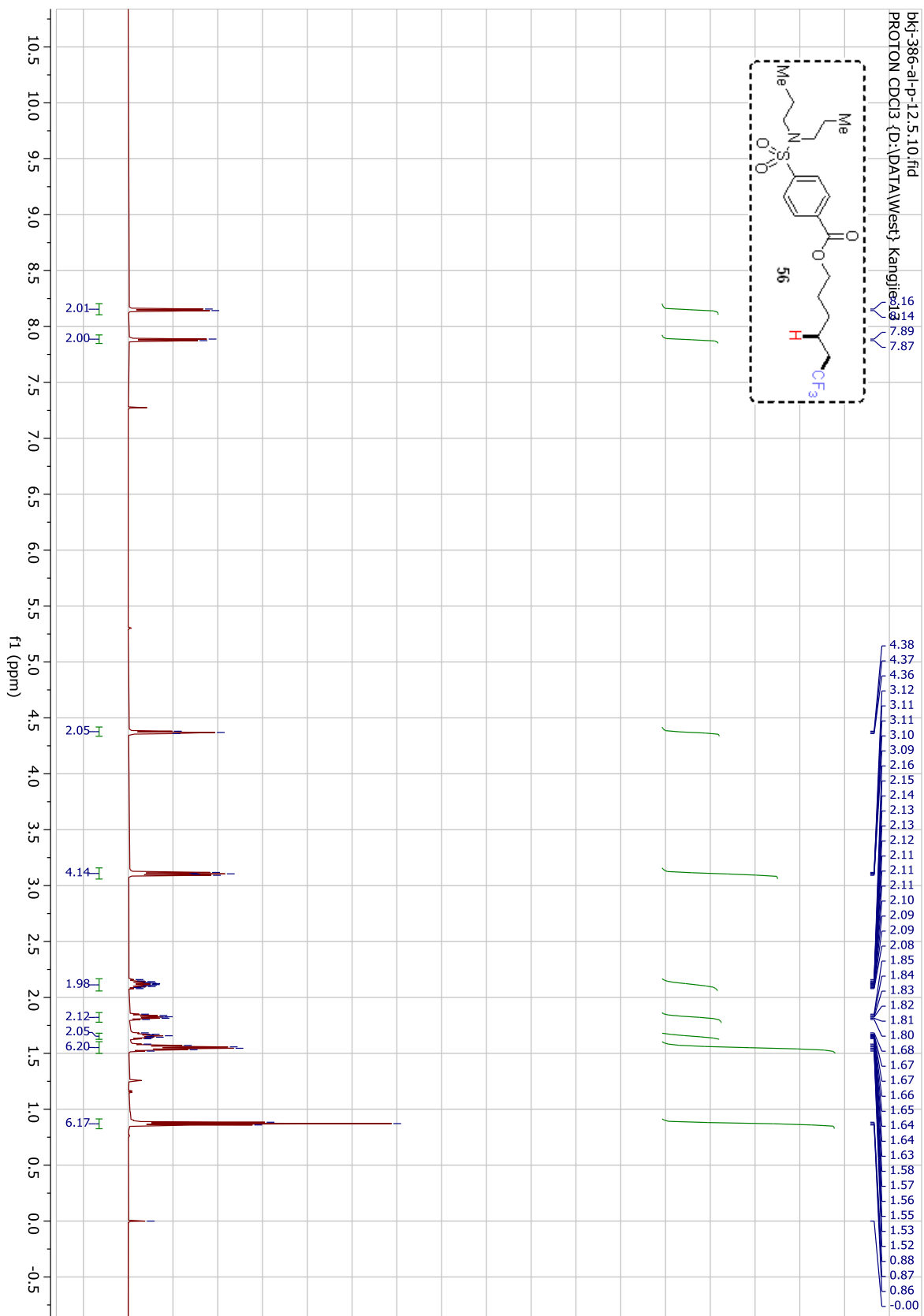
bkj-386-aj-p-h-12.3.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 11



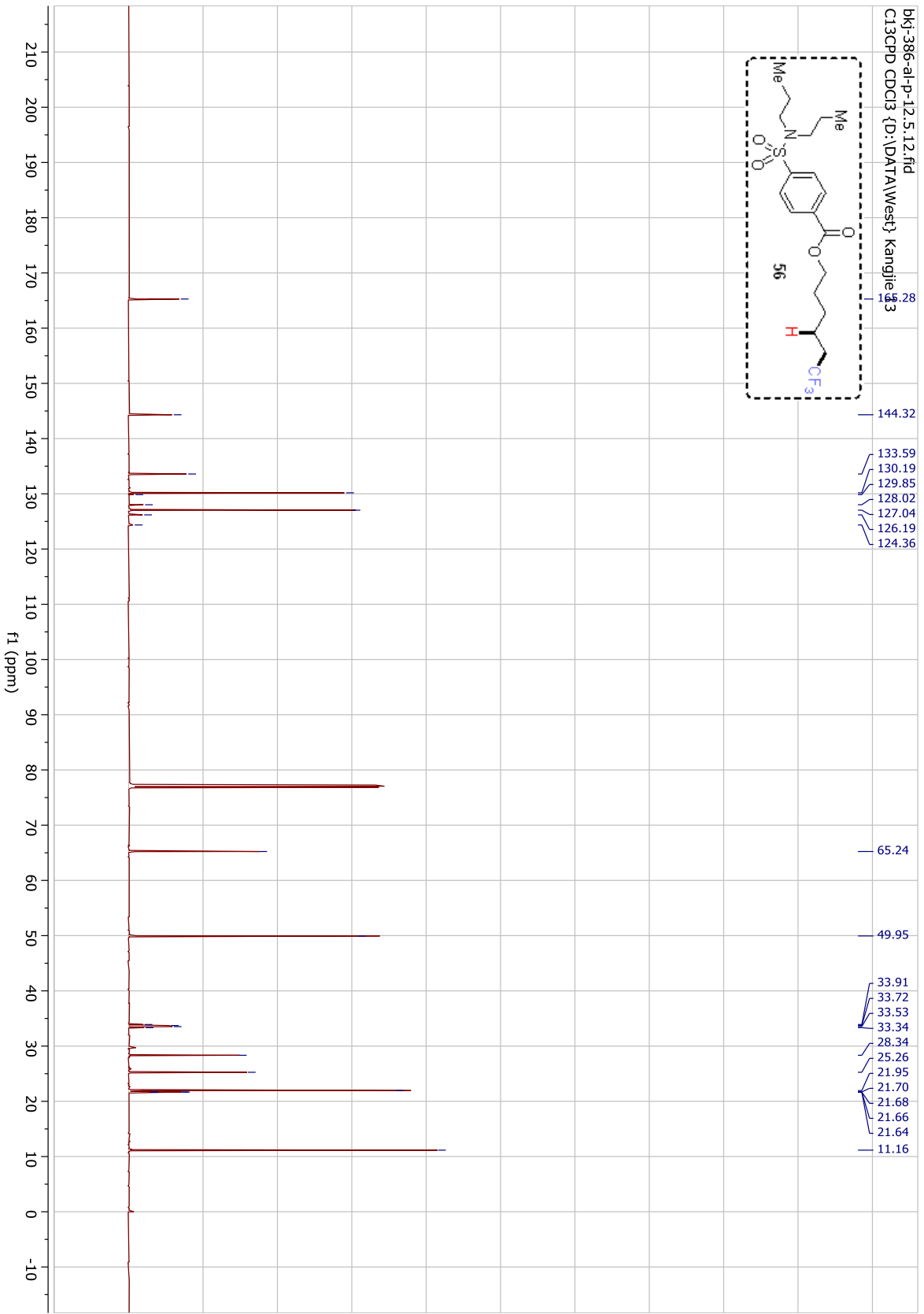
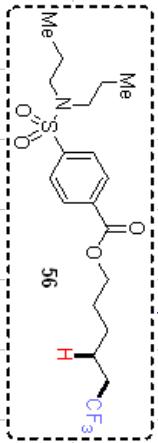
-66.34
-66.36
-66.38

-117.65
-117.67
-117.67
-117.69

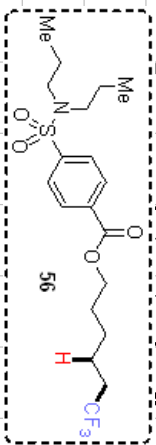




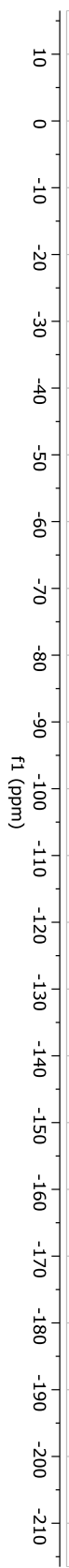
bk1-386-al-p-12.5.12.fid
C13CPD CDCl3 (D:\DATA\Westj_Kangjie_93

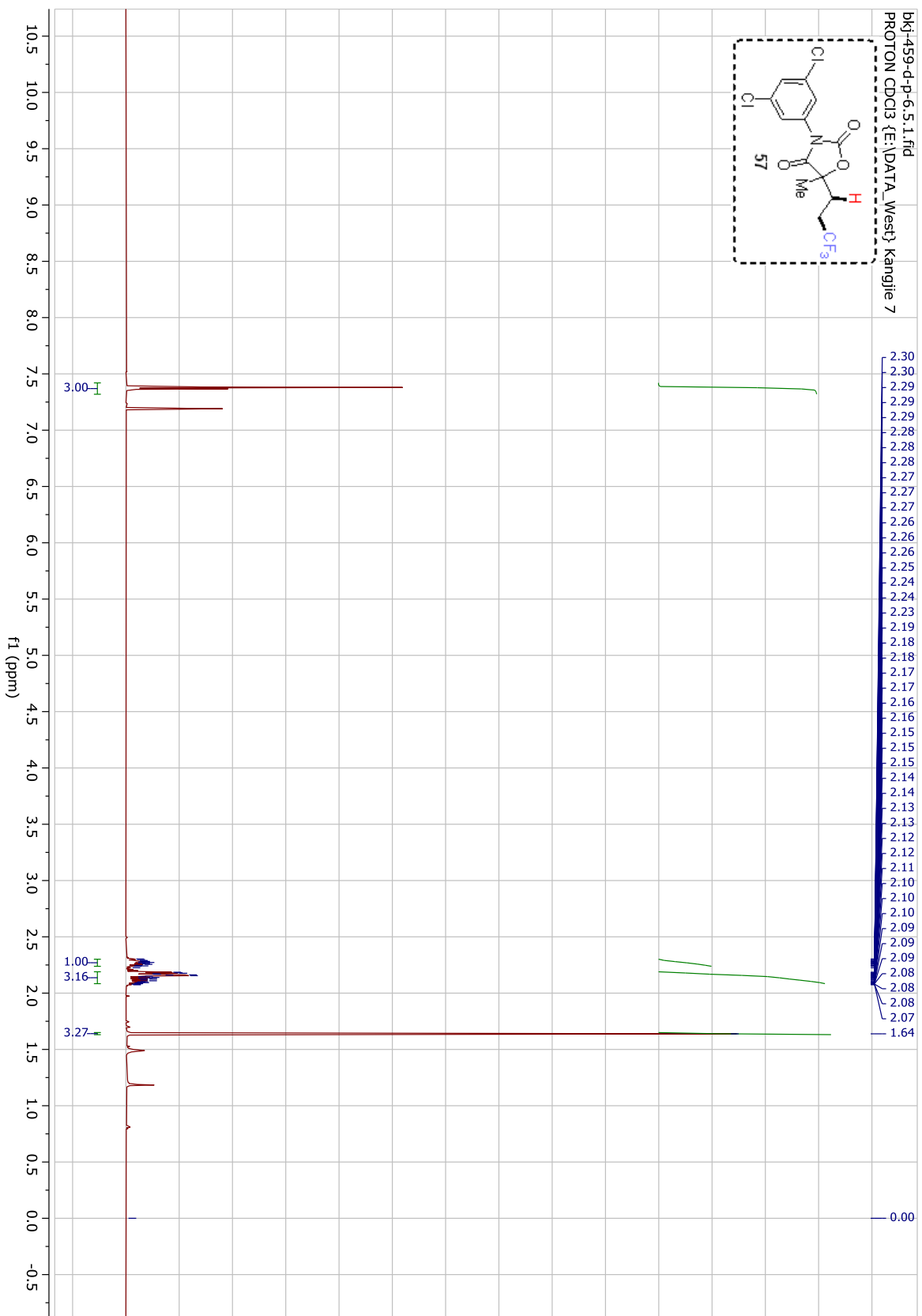


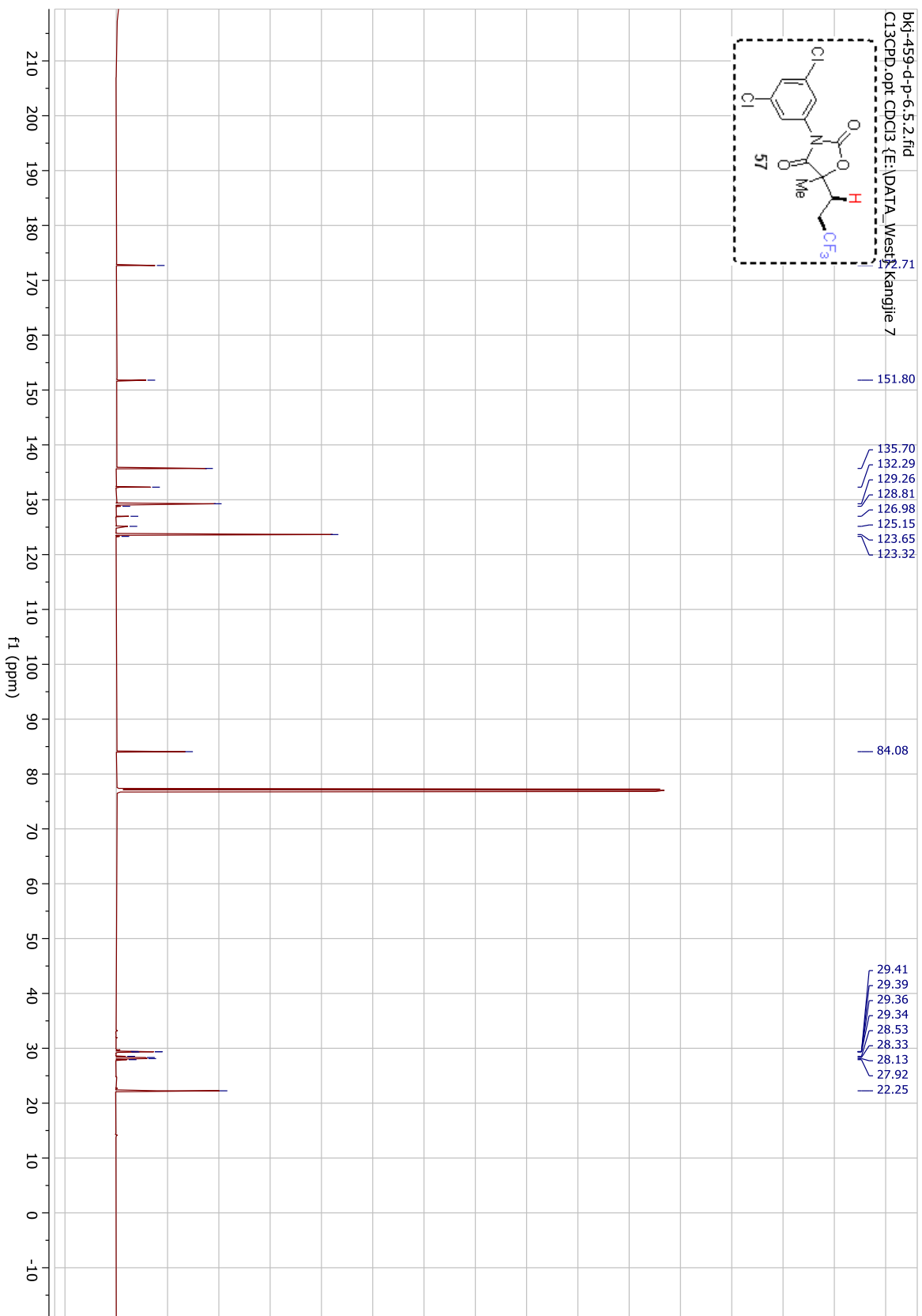
bkj-386-al-p-12.5.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West} Kangjie 13



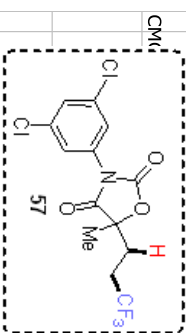
-66.33
-66.35
-66.37



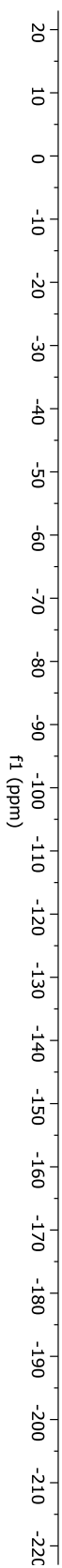


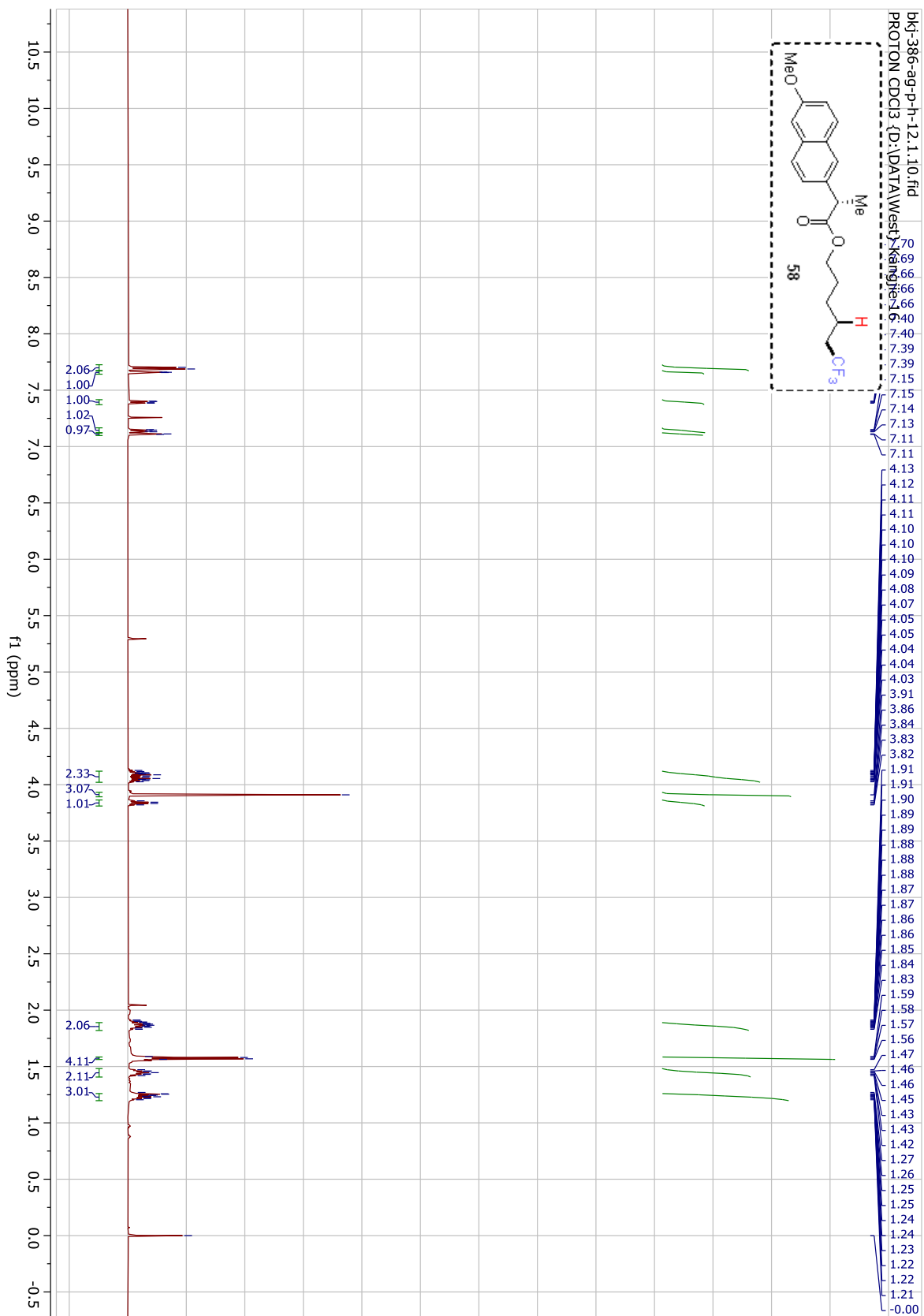


bkj-459-d-p-f-6.6.30.fid
F19

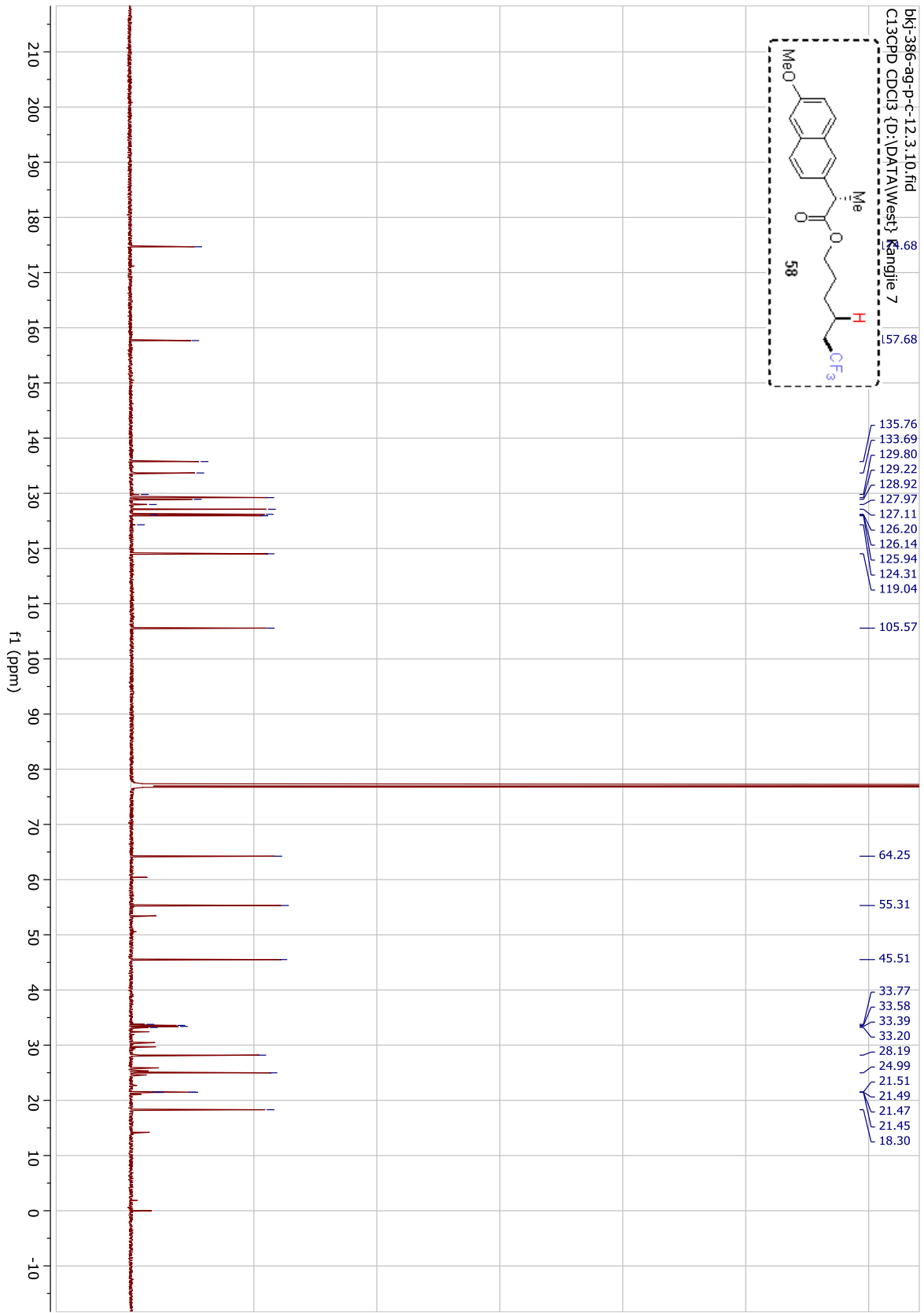
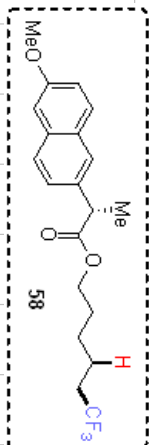


-66.38
-66.40
-66.41

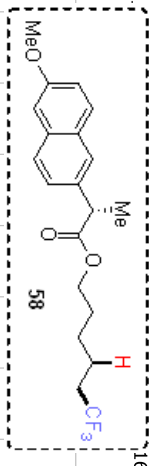




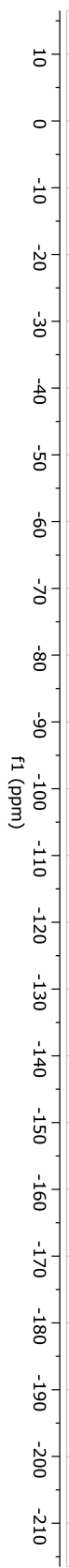
hkl-386-ag-p-c-12.3.10.fid
C13CPD CDCl3 (D:\DATA\West) Sample 7

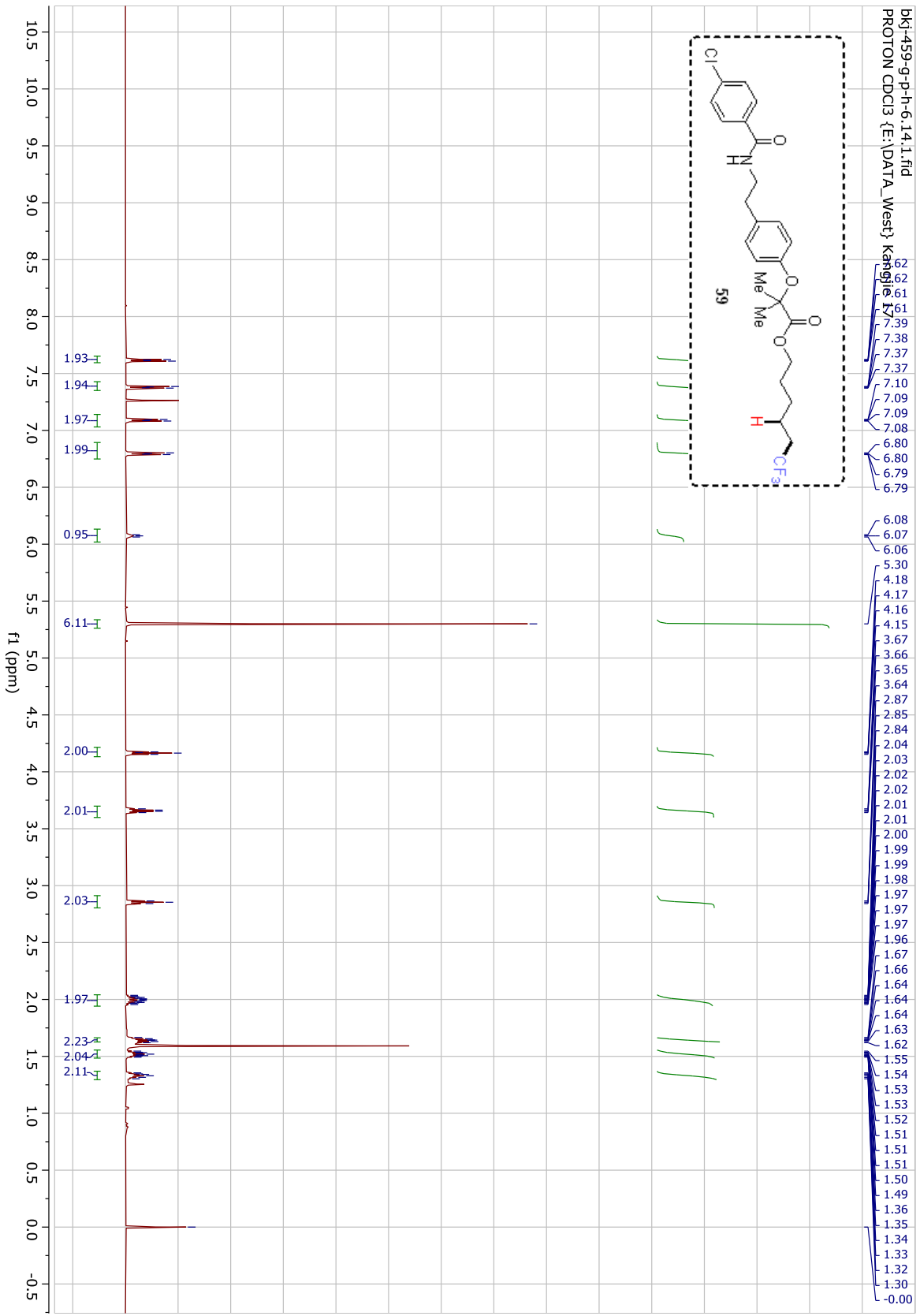


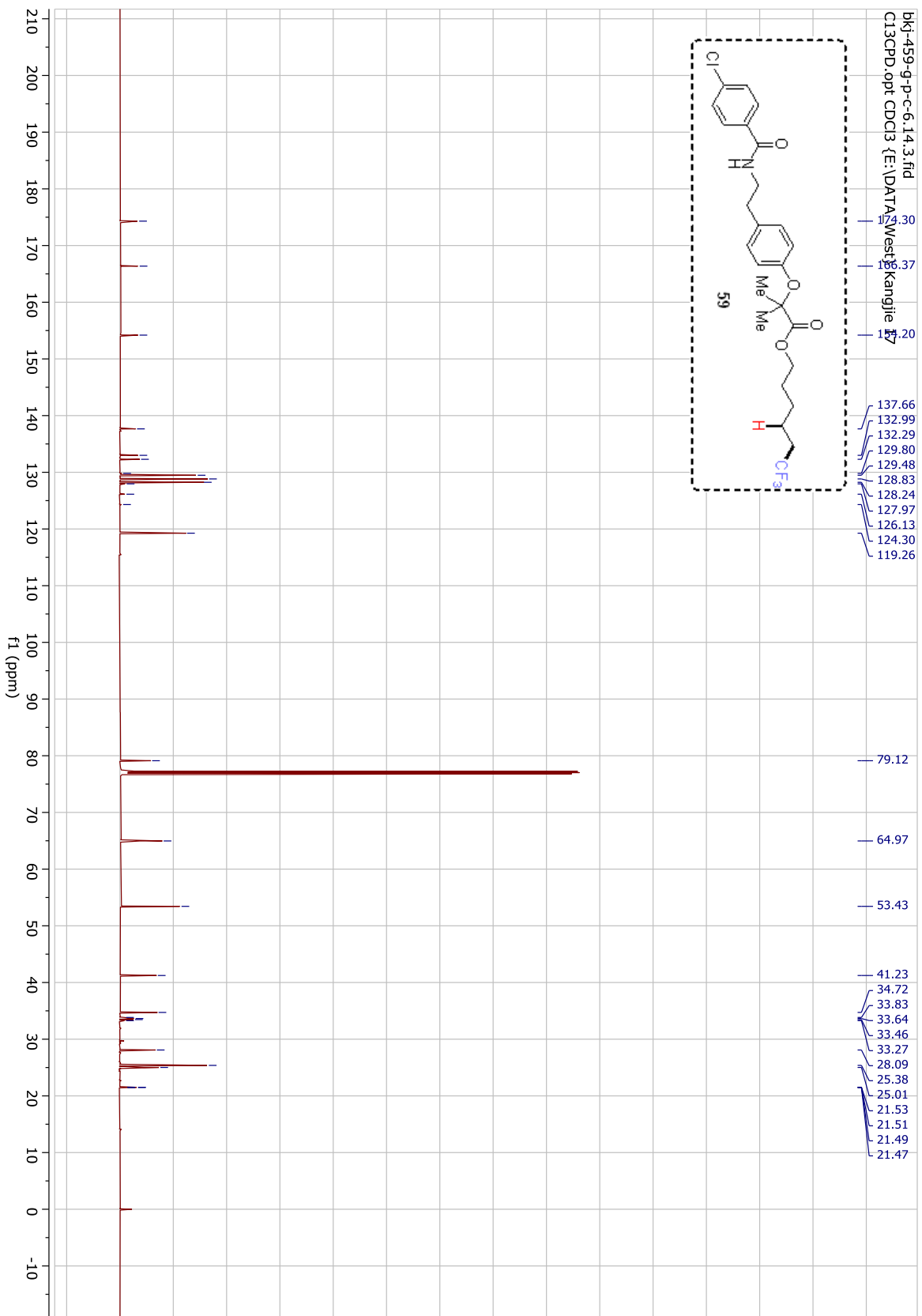
bkj-386-ag-p-h-12.1.11.ftd
F19_baseline_correct_ID



-66.42
-66.44
-66.46



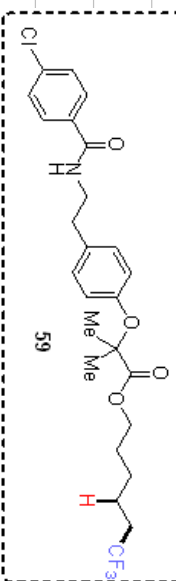


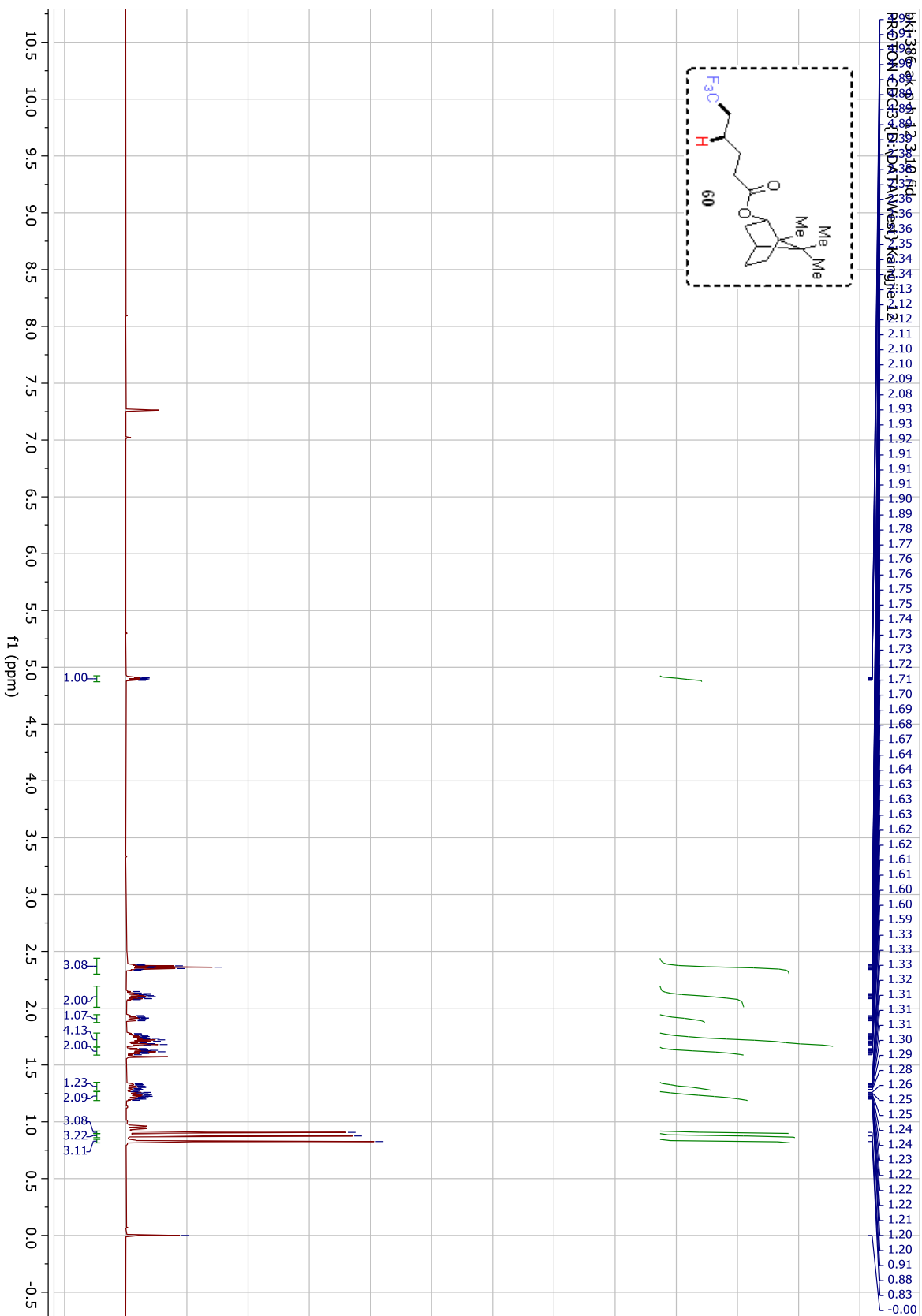


bkj-459-g-p-f-6.15.10.ftd
F19

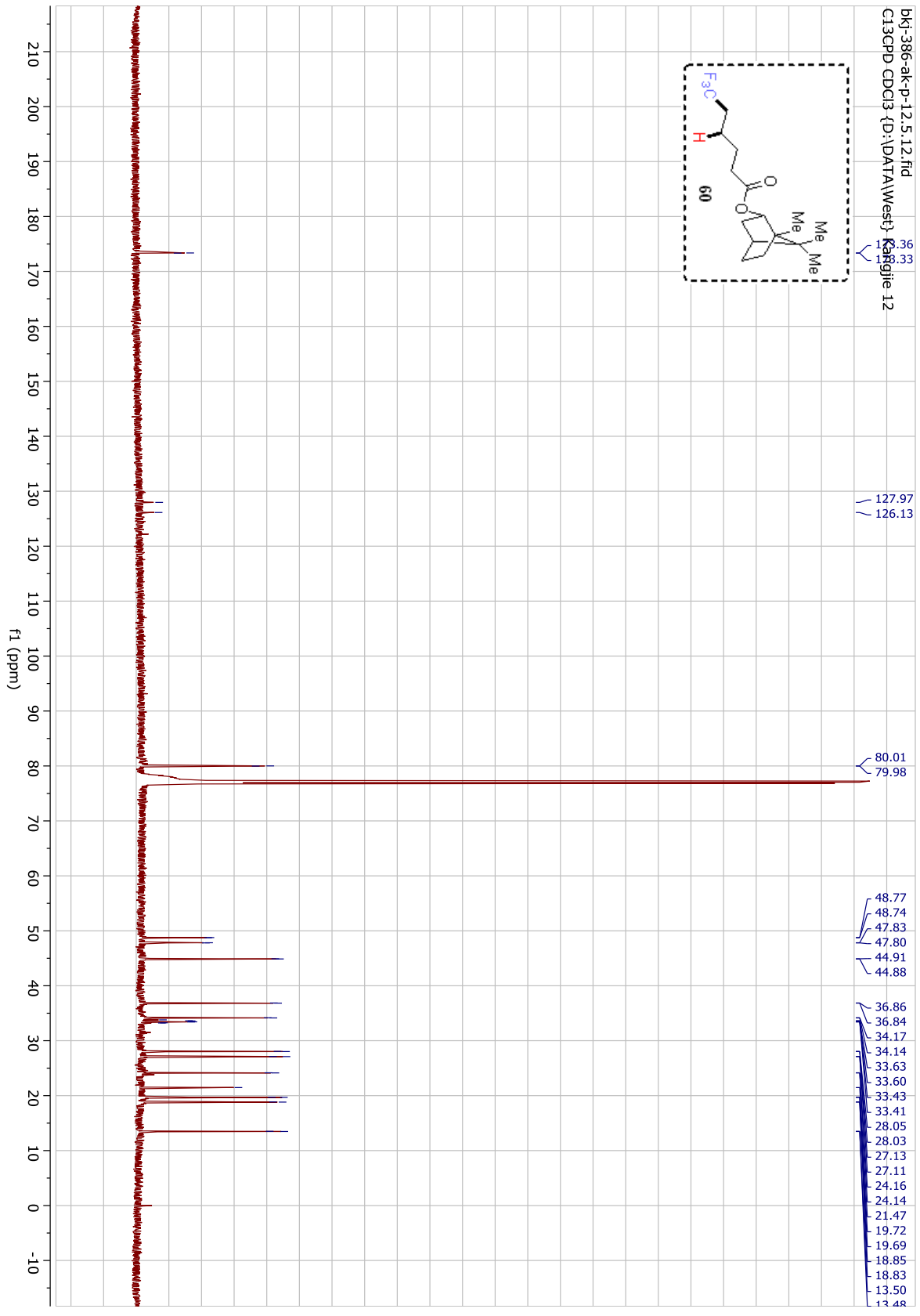
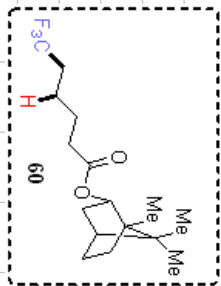
CMC-F19.CDCI3.FD\DATA\West\Kantile 17

-66.31
-66.33
-66.35



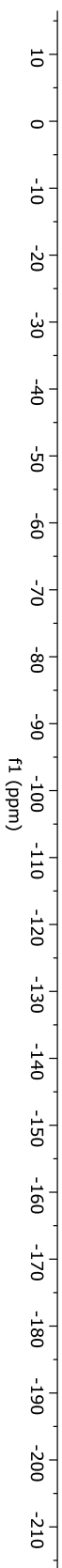
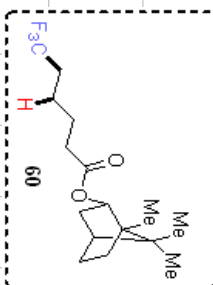


bkj-386-ak-p-12.5.12.fid
C13CPD CDCl3 (D:\DATA\West)\K28912

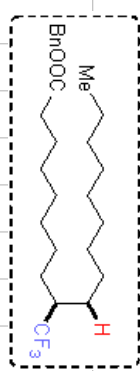


bkj-386-ak-p-h-12.3.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West\ Kangjie 12

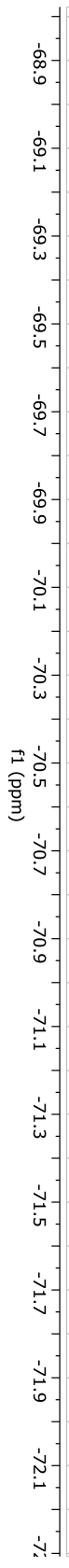
-66.37
-66.39
-66.41



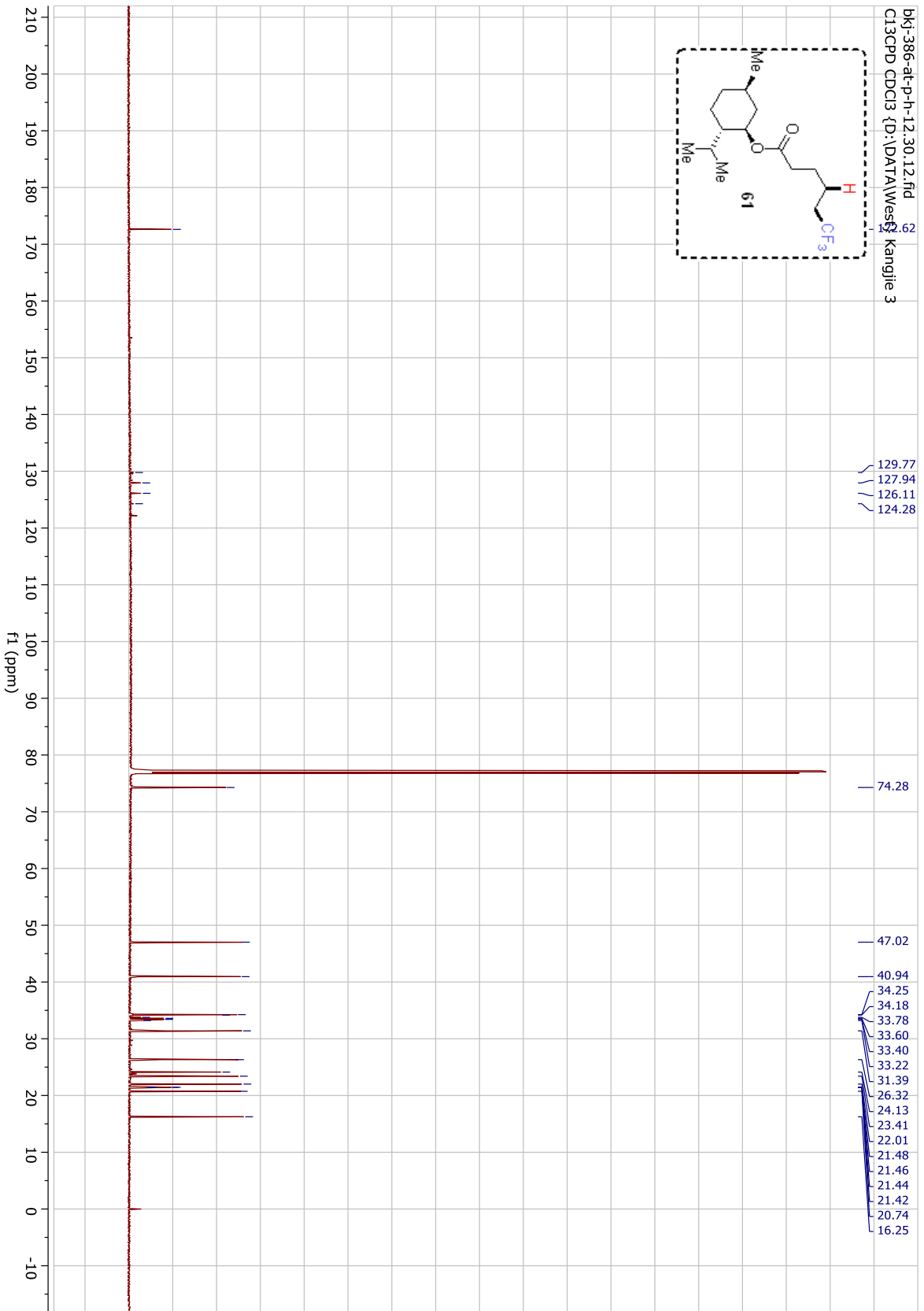
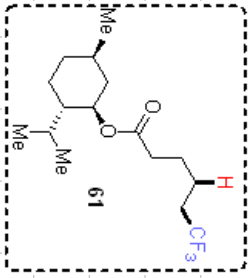
bk1-386-au"-p-h-1.2.12.fid
F191-BBMR-500MHz-1Doforom-5-70.14 (dJ = 4.3 Hz).
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 3



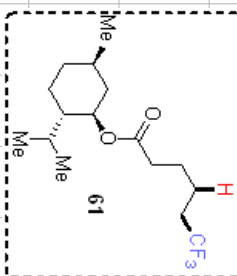
-70.12
-70.12
-70.13
-70.14



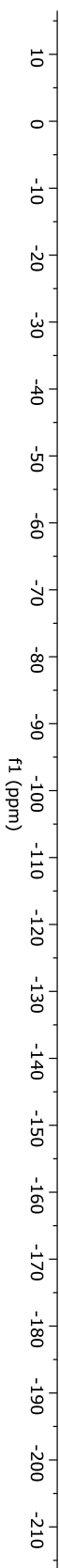
bkj-386-at-p-h-12.30.12.fid
C13CPD CDCl3 (D:\DATA\Wes) 2
Kangjie 3

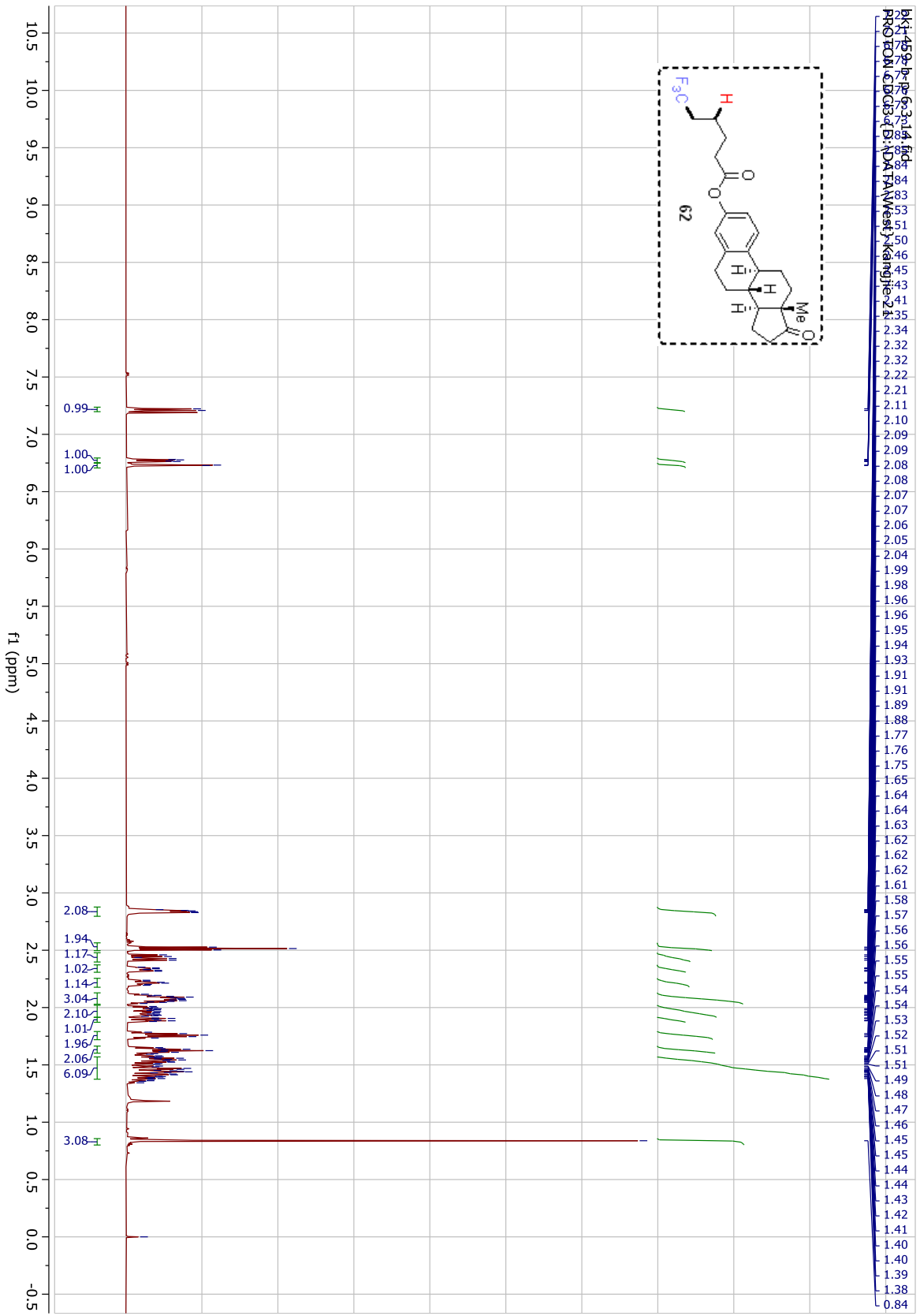


bkj-386-at-p-h-12.30.11.fid
F19_baseline_correct ID
F19_baseline_correct FID19 (DATA\West) Kangjie 3

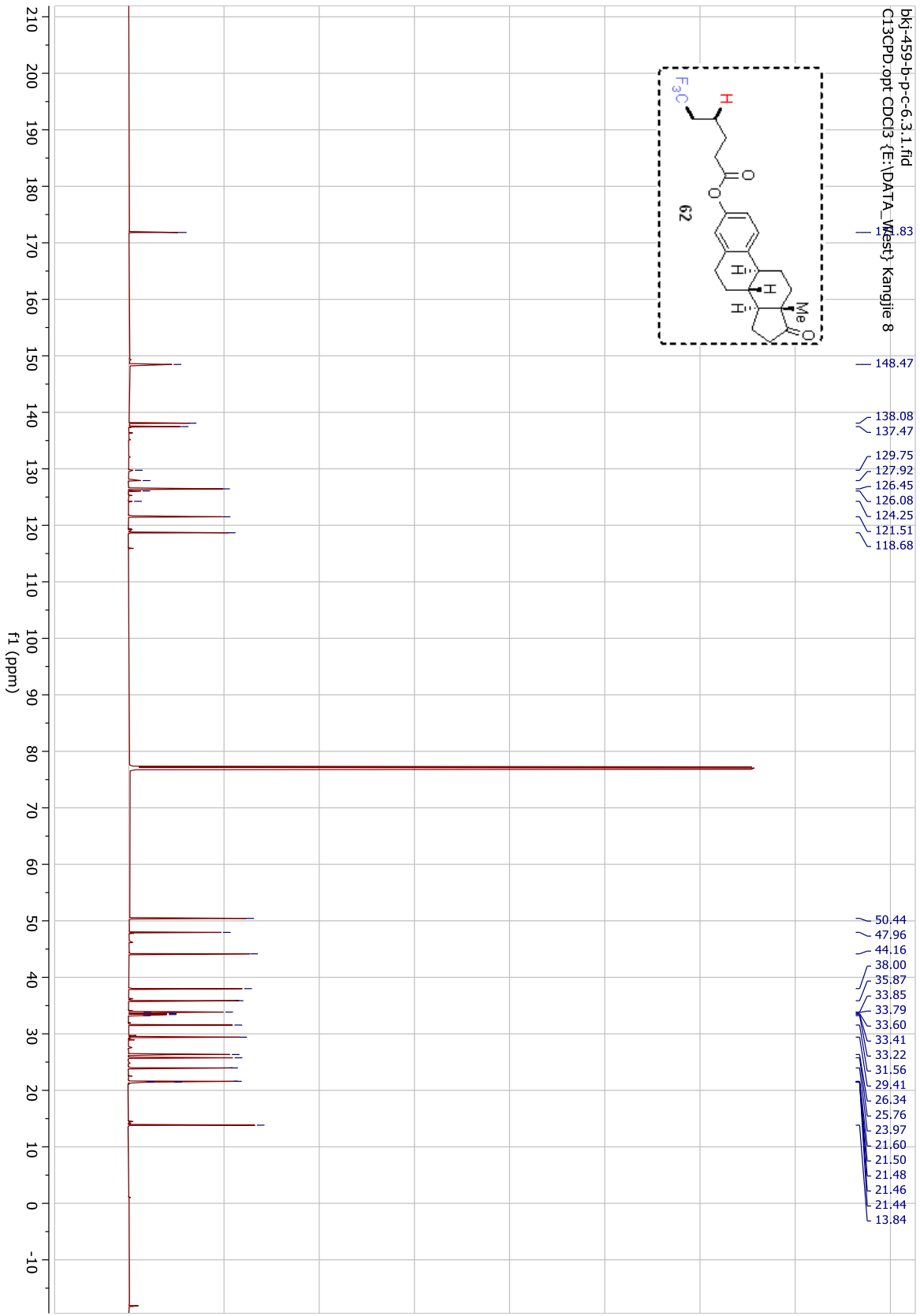
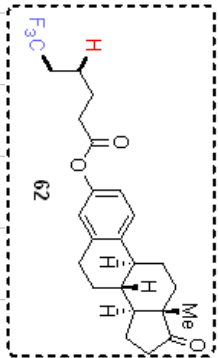


-66.41
-66.43
-66.45





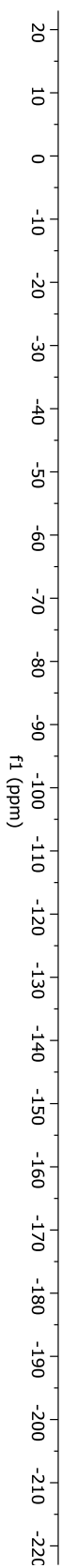
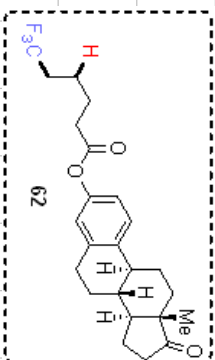
bkj-459-b-p-c-6-3-1.fid
C13CPD.opt CDCl3 {E:\DATA_Mest\ Kanjlie 8



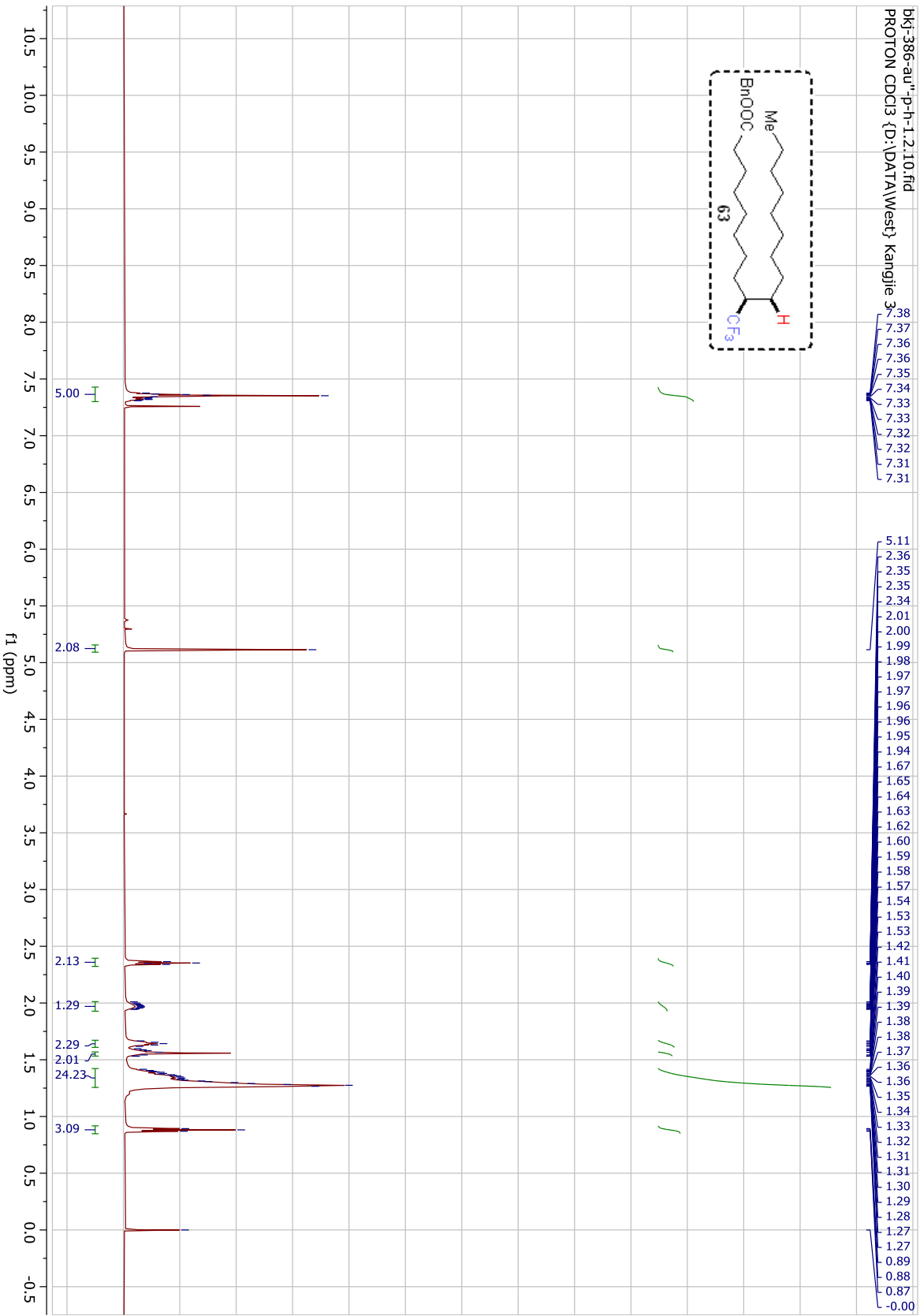
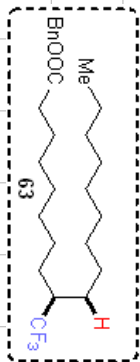
bkj-459-b-p-f-6.3.12.ftd
F19

CMC_F19 CDCl3 (D:\DATA\West) Kanjije 21

-66.30
-66.32
-66.34



bk1-386-au"-p-h-1.2.10.fid
PROTON CDCl3 {D:\DATA\West\ Kangjie 3



bjt-386-au"-p-h-1.2.11.fid
C13CPD CDCl3 (D:\DATA\West) K20170116 3

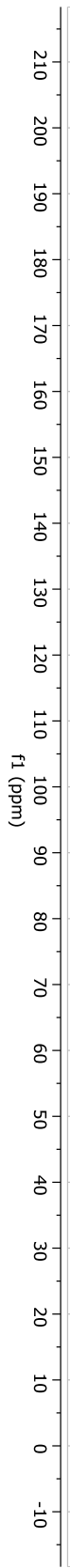
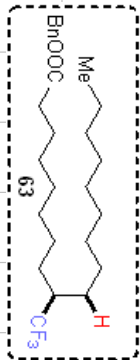
89.64

136.14
136.13
131.57
129.72
128.55
128.19
127.87
126.00

66.10
66.08

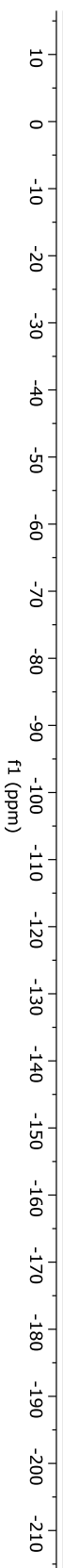
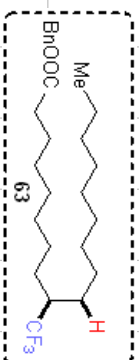
42.81
42.64
42.48
42.32

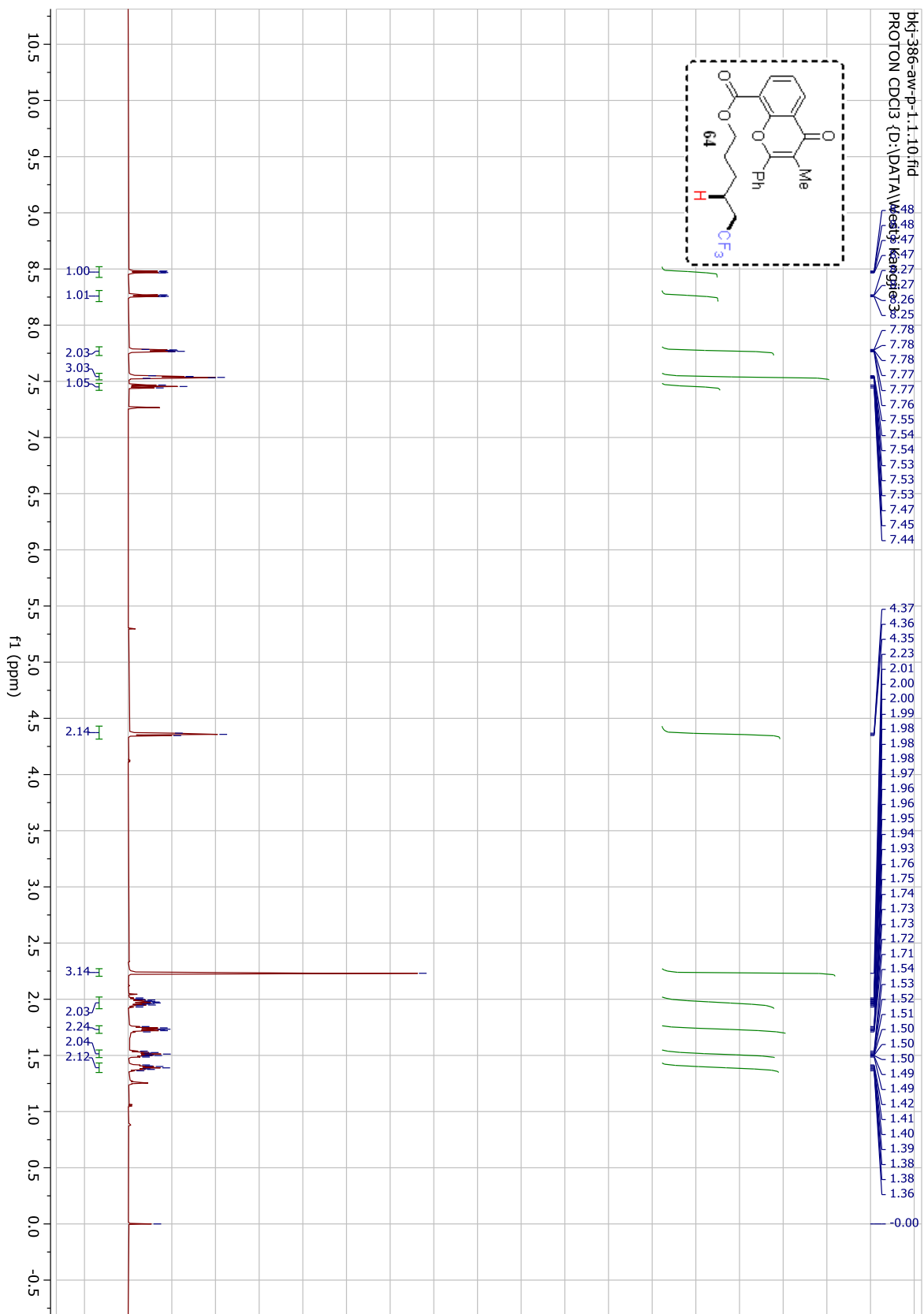
34.29
31.89
29.72
29.55
29.42
29.30
29.21
29.18
29.18
29.08
29.05
29.03
27.85
27.84
26.88
24.93
24.90
22.68
22.66
14.11

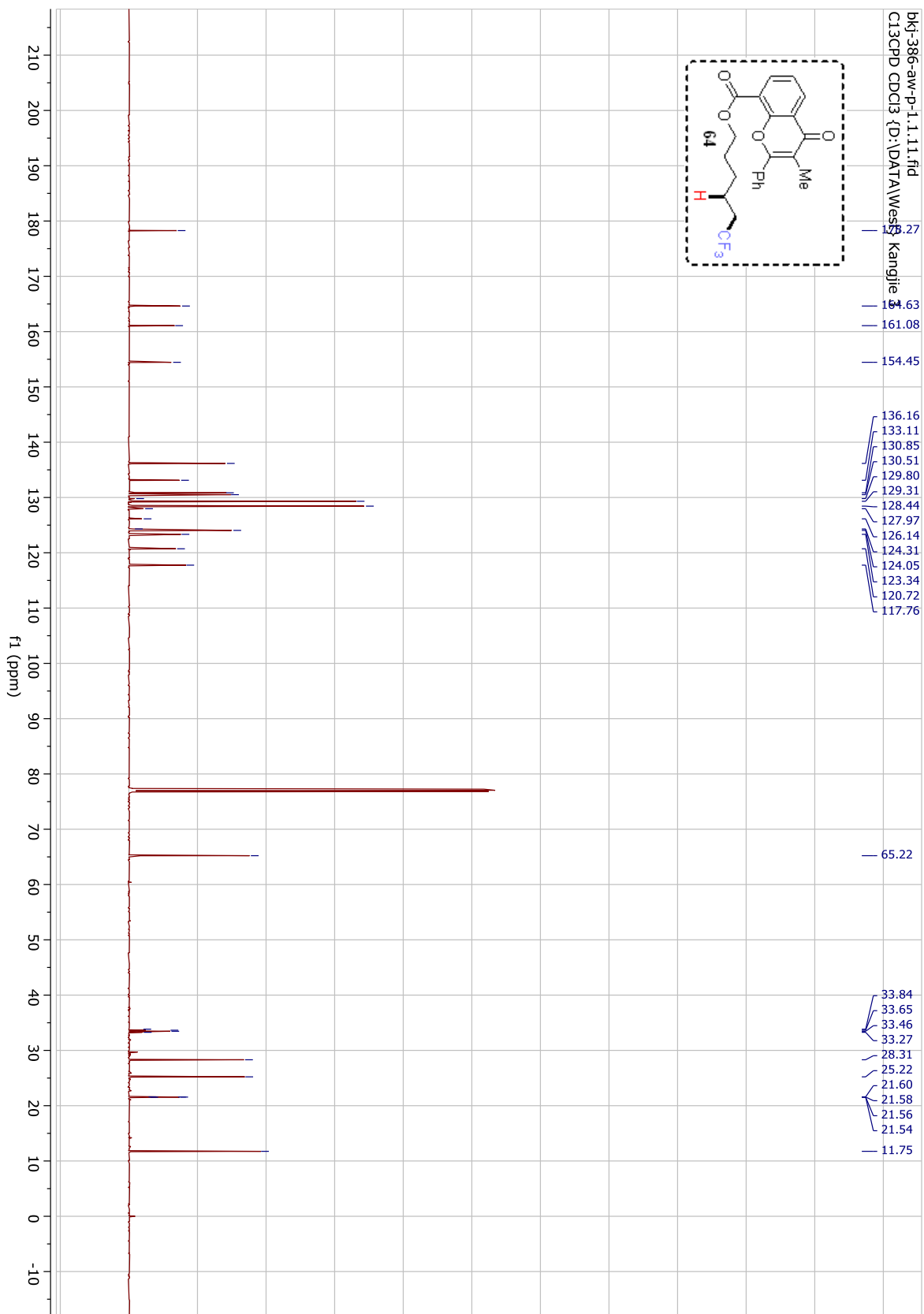


bkj-386-au"-p-h-1.2.12.fid
F19_baseline_correct_ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 3

-70.12
-70.12
-70.13
-70.14

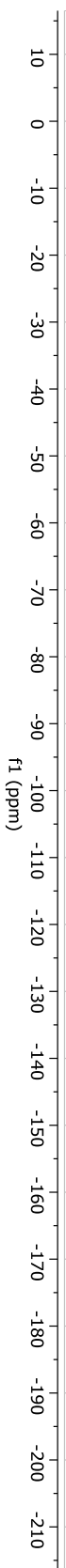
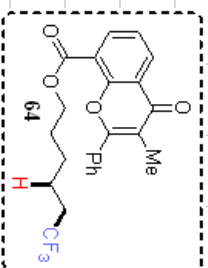




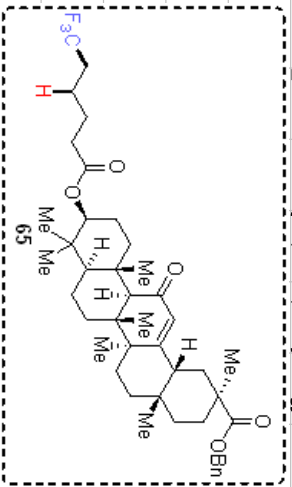


bkj-386-aw-p-1.1.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 3

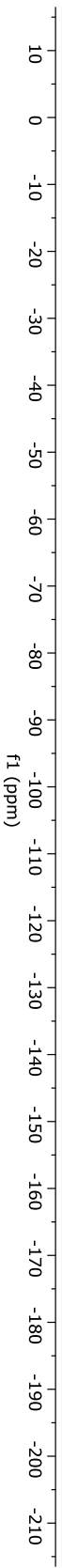
-66.36
-66.38
-66.40



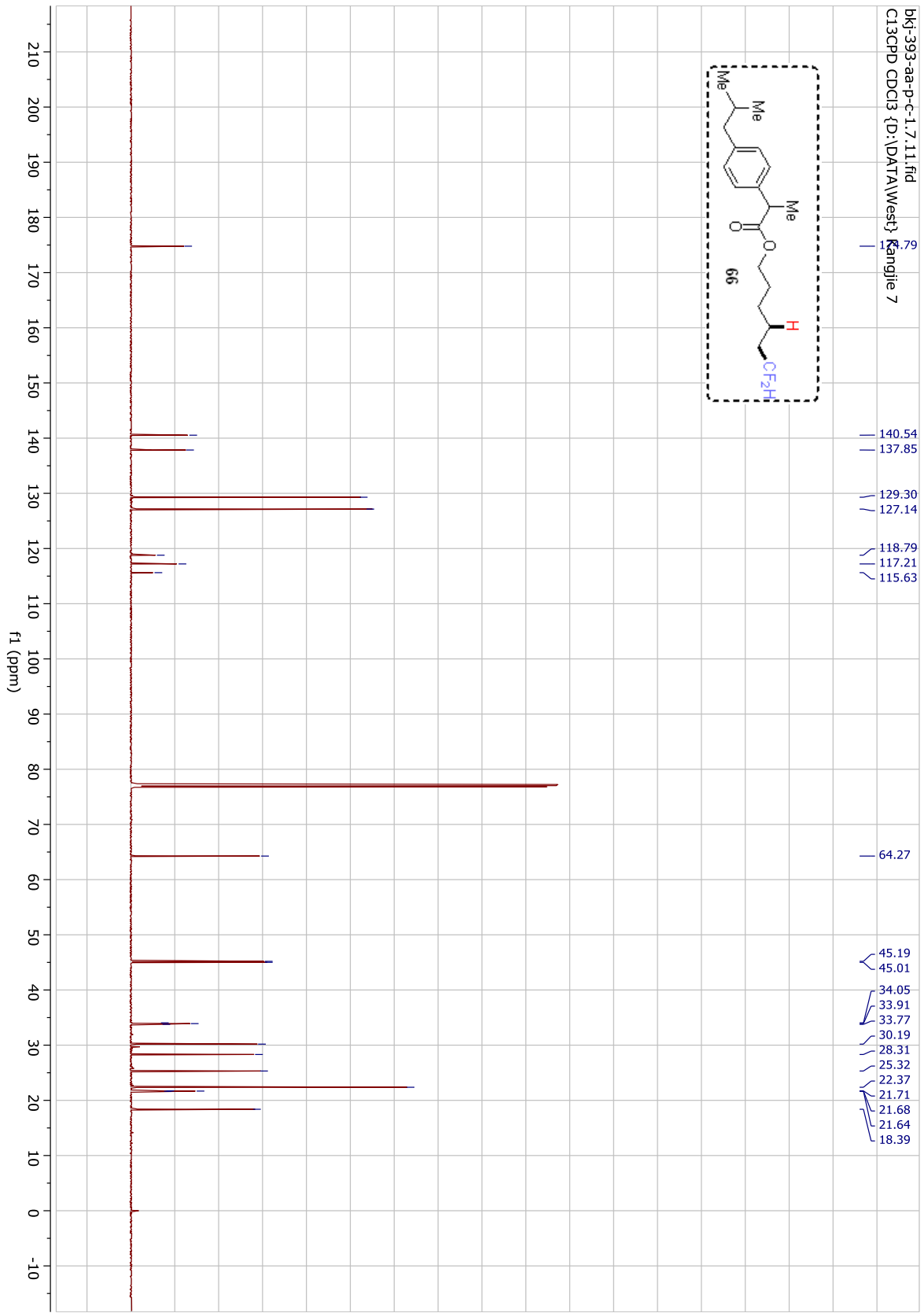
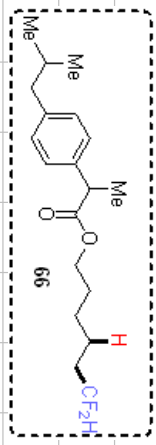
bkj-386-as-p-12.30.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 8



-66.34
-66.36
-66.38

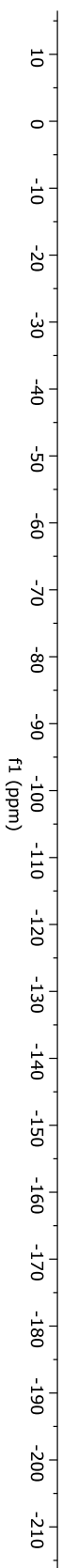
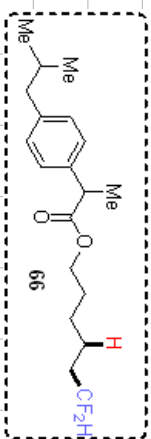


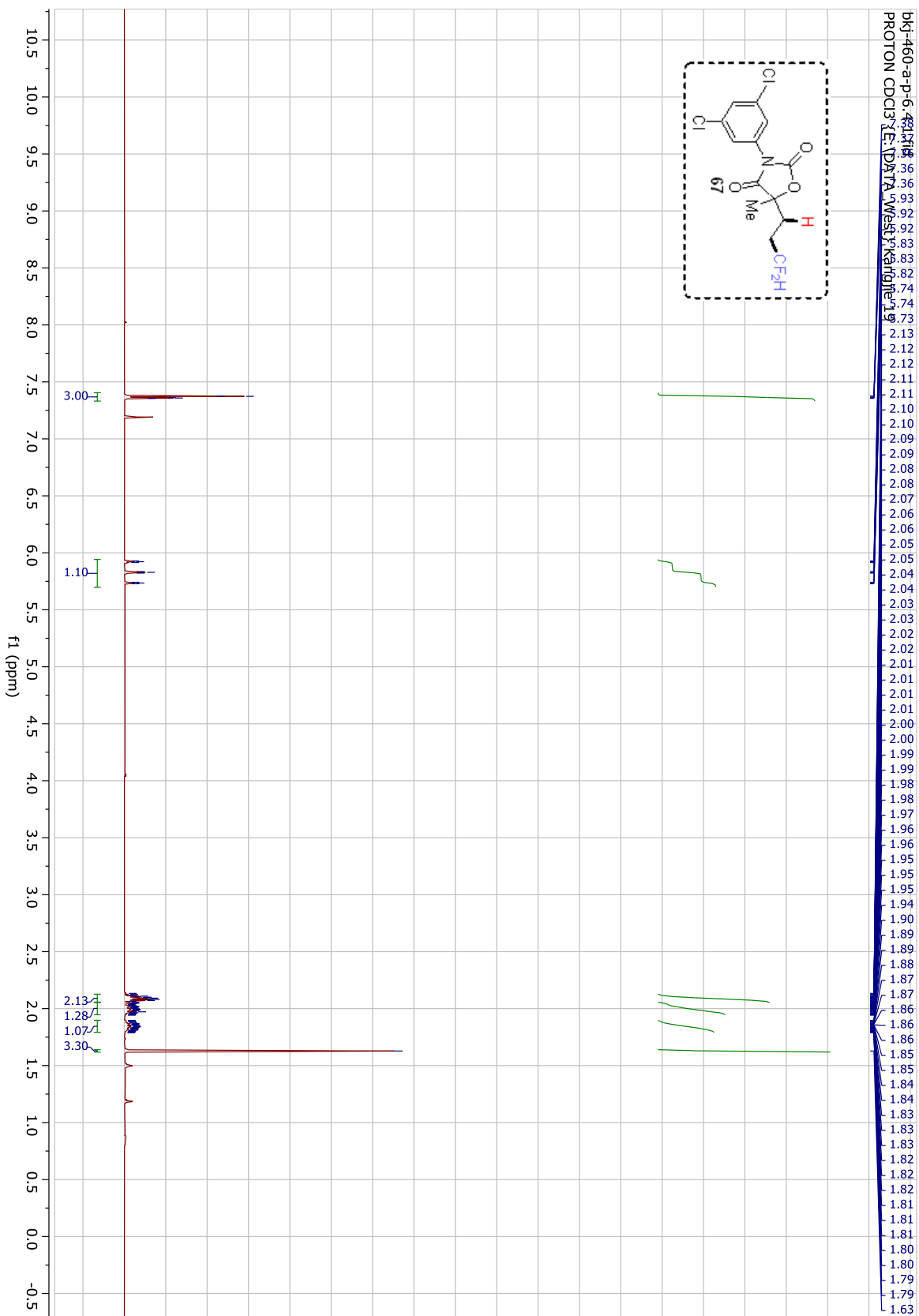
hkl-393-aa-p-c-1.7.11.fid
C13CPD CDCl3 (D:\DATA\West) Sample 7

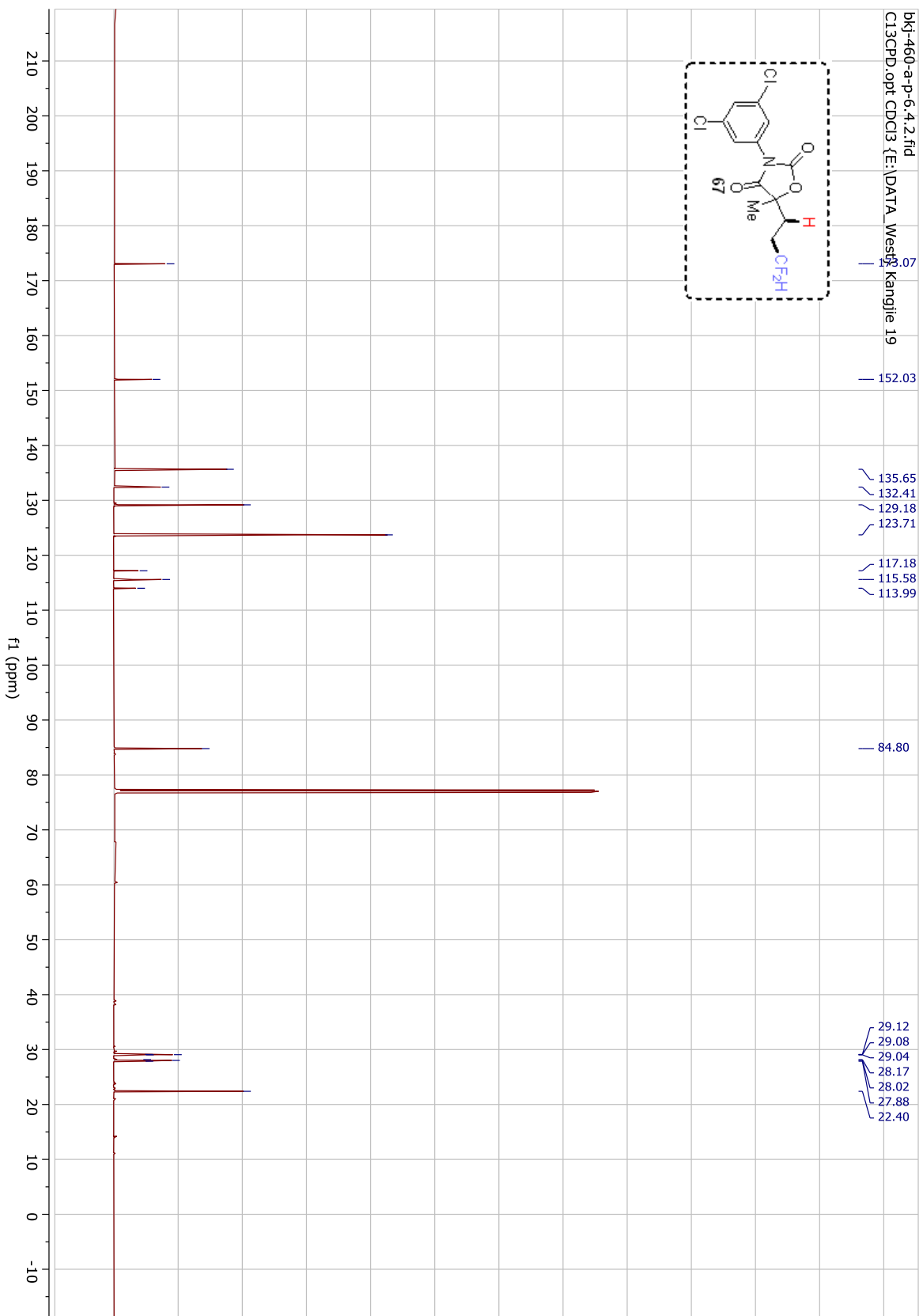


bkj-393-aa-p-h-1.7.11.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 7

-115.78
-115.81
-115.84
-115.88
-115.91
-115.94

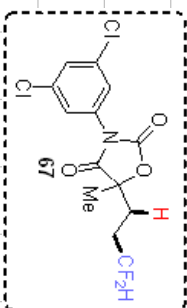




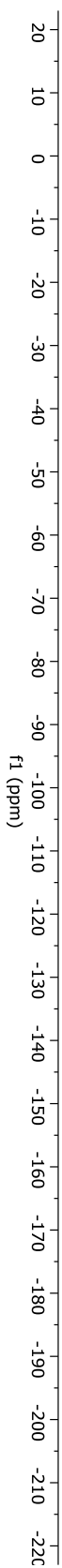


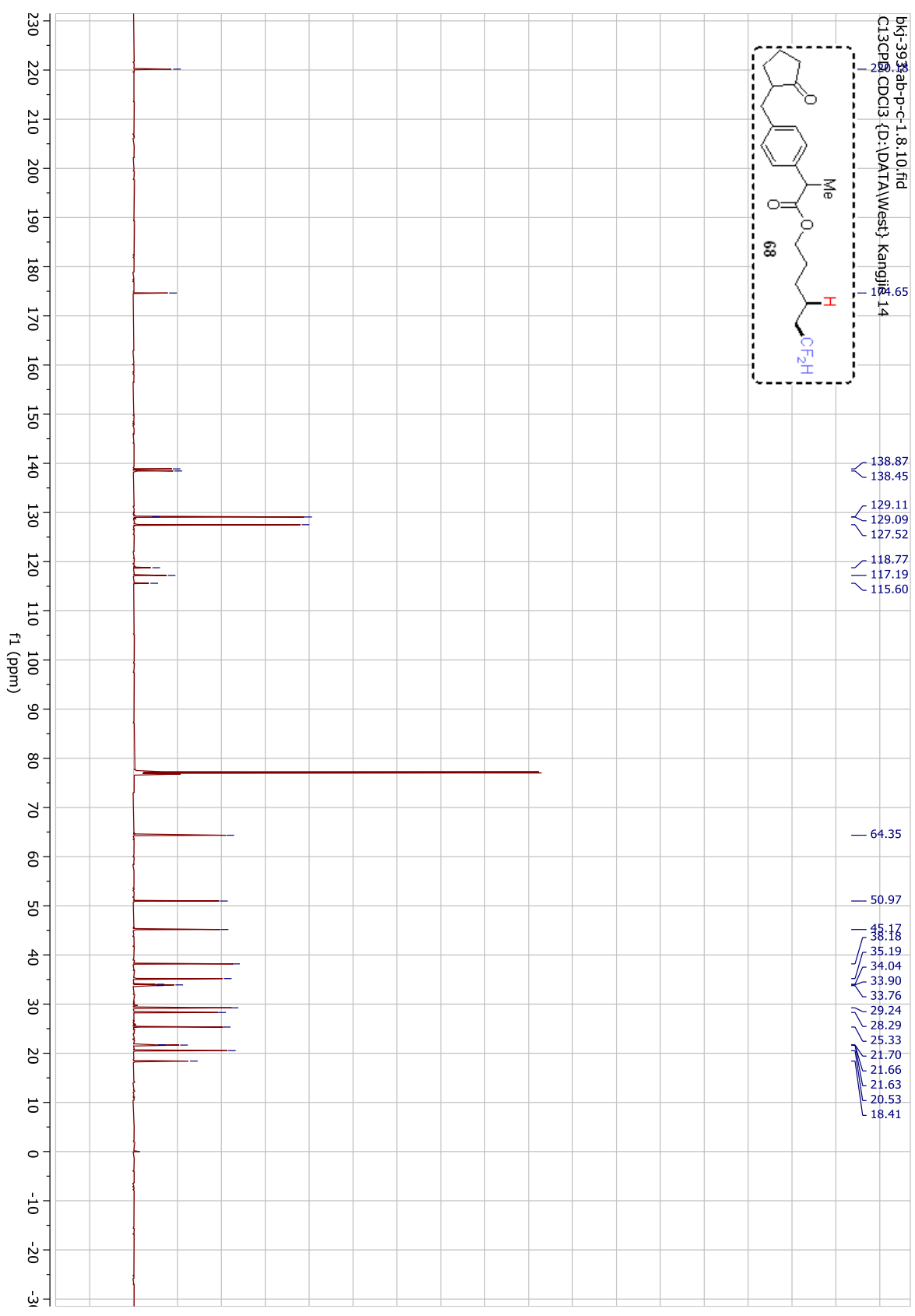
bkj-460-a-p-f-6.5.10.fid
F19

CMC_F19 CDCl3 (D:\DATA\West) Kangjie 22

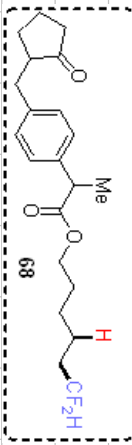


-116.79
-116.82
-116.85
-116.89
-116.92
-116.95

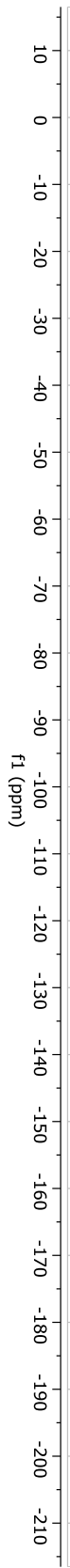




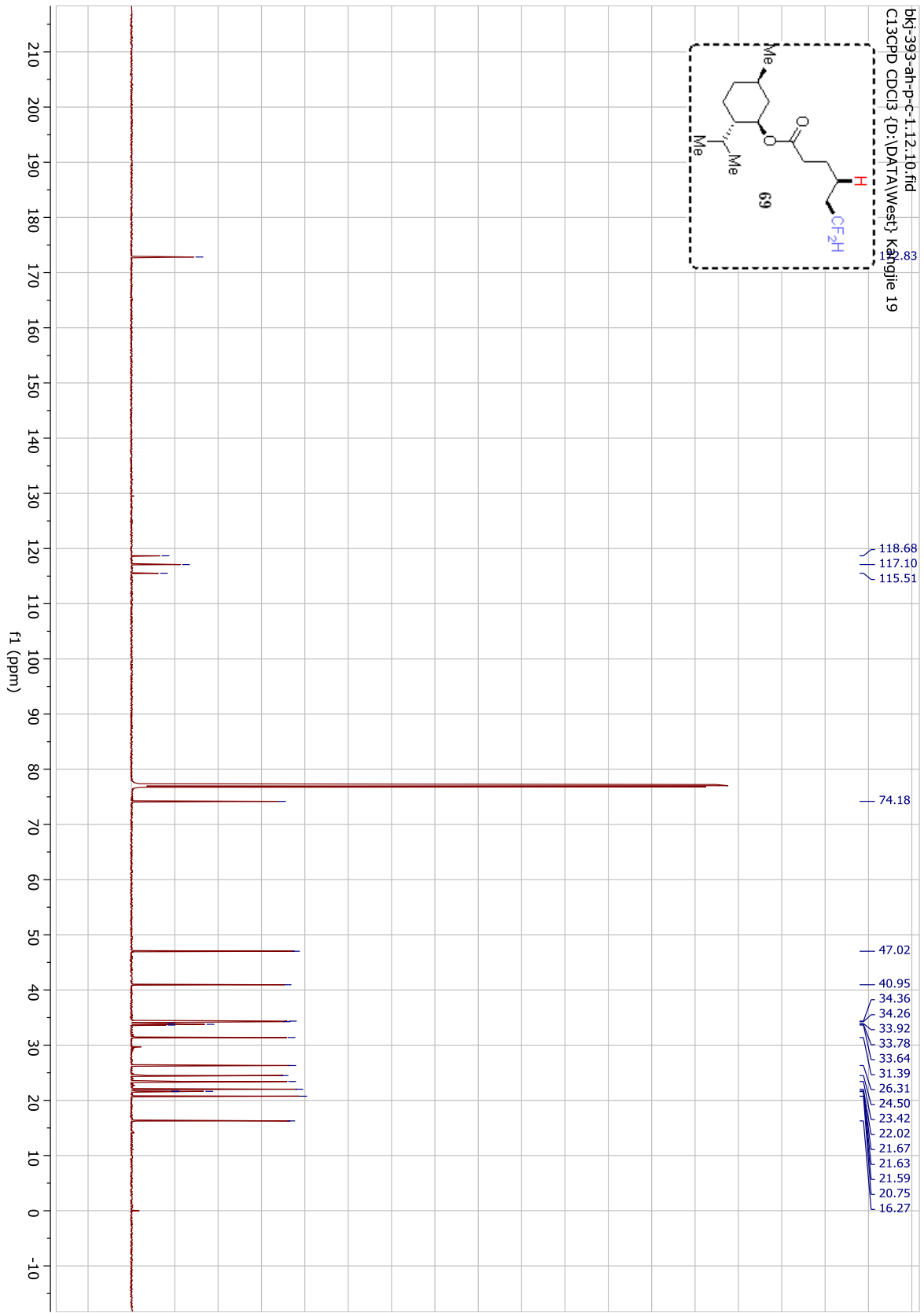
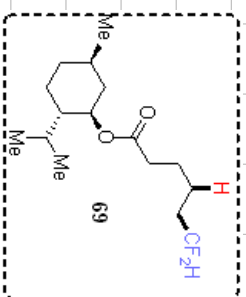
bkj-393-ab-pl-h-1.7.11.ftd
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 8



-115.77
-115.81
-115.84
-115.88
-115.91
-115.94

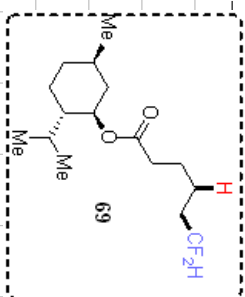


bkt-393-ah-p-c-1.12.10.fid
C13CPD CDCl3 (D:\DATA\Westj k\u00e4ngj\u00e4le 19

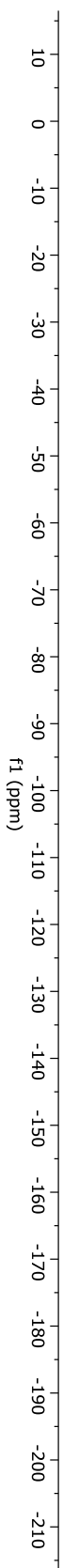


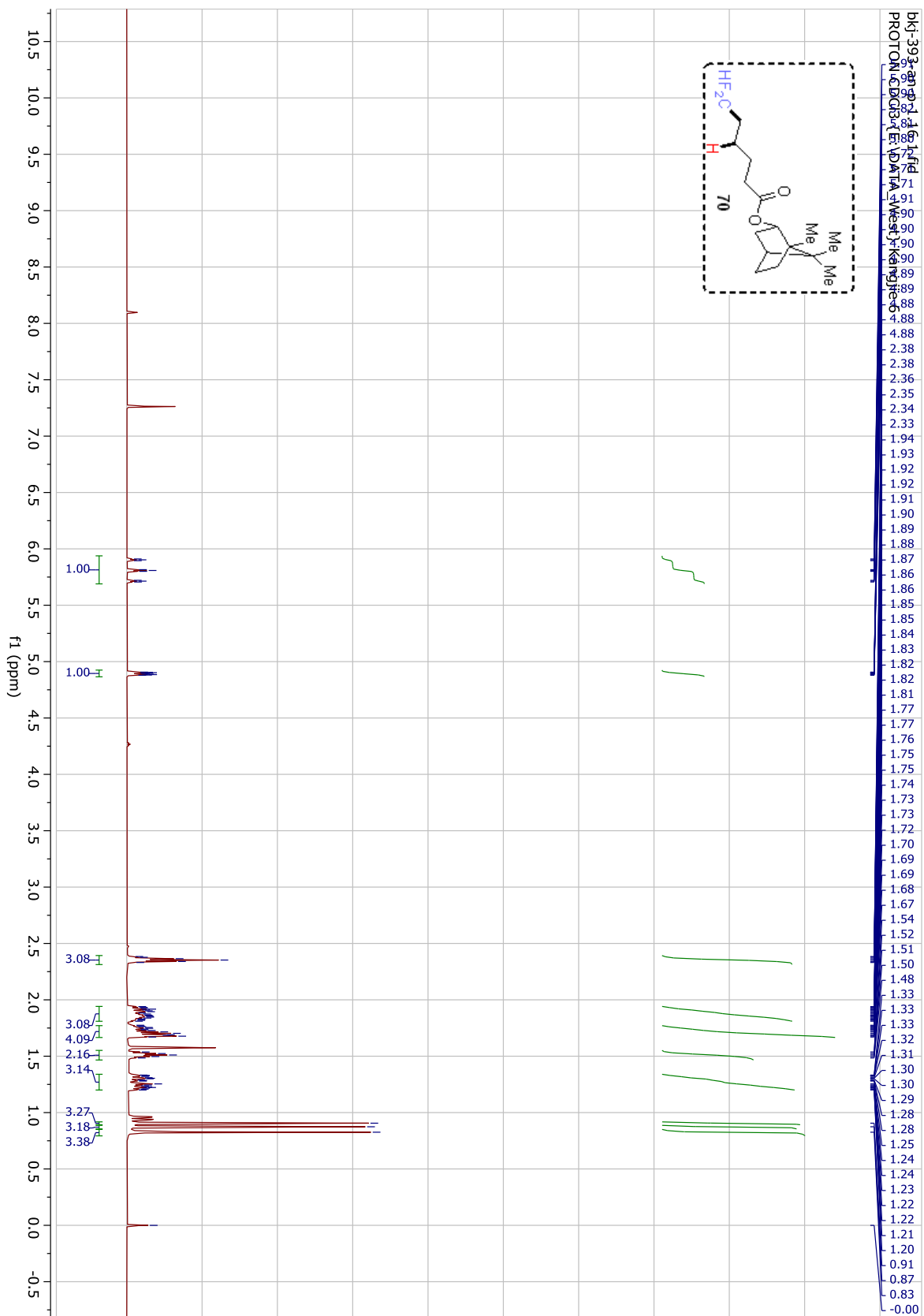
bkj-393-ah-p-f-1.11.10.fid
F19_baseline_correct ID
F19_

AWestf Kangjie 19

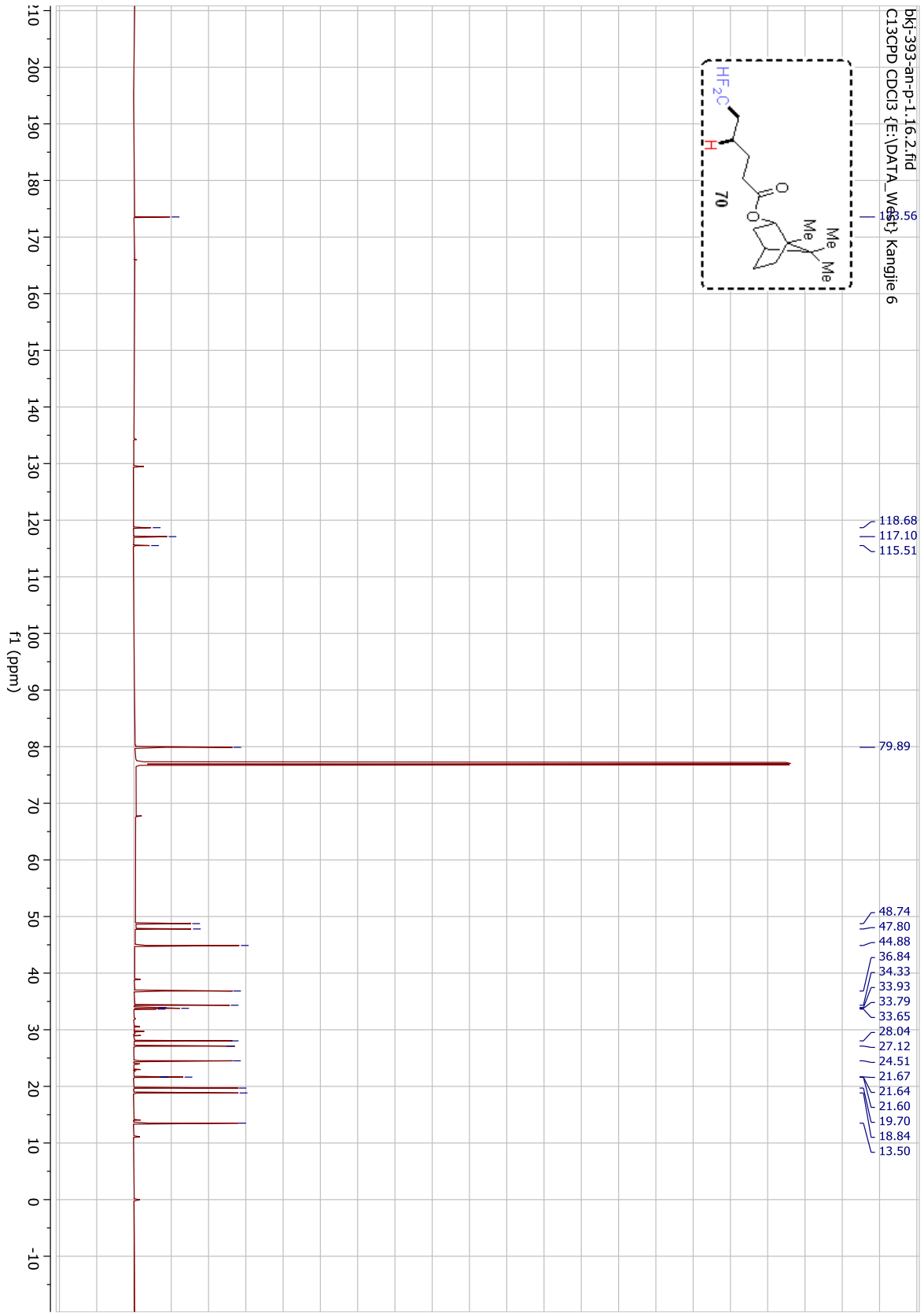
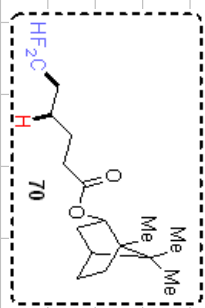


-115.88
-115.91
-115.94
-115.98
-116.01
-116.04

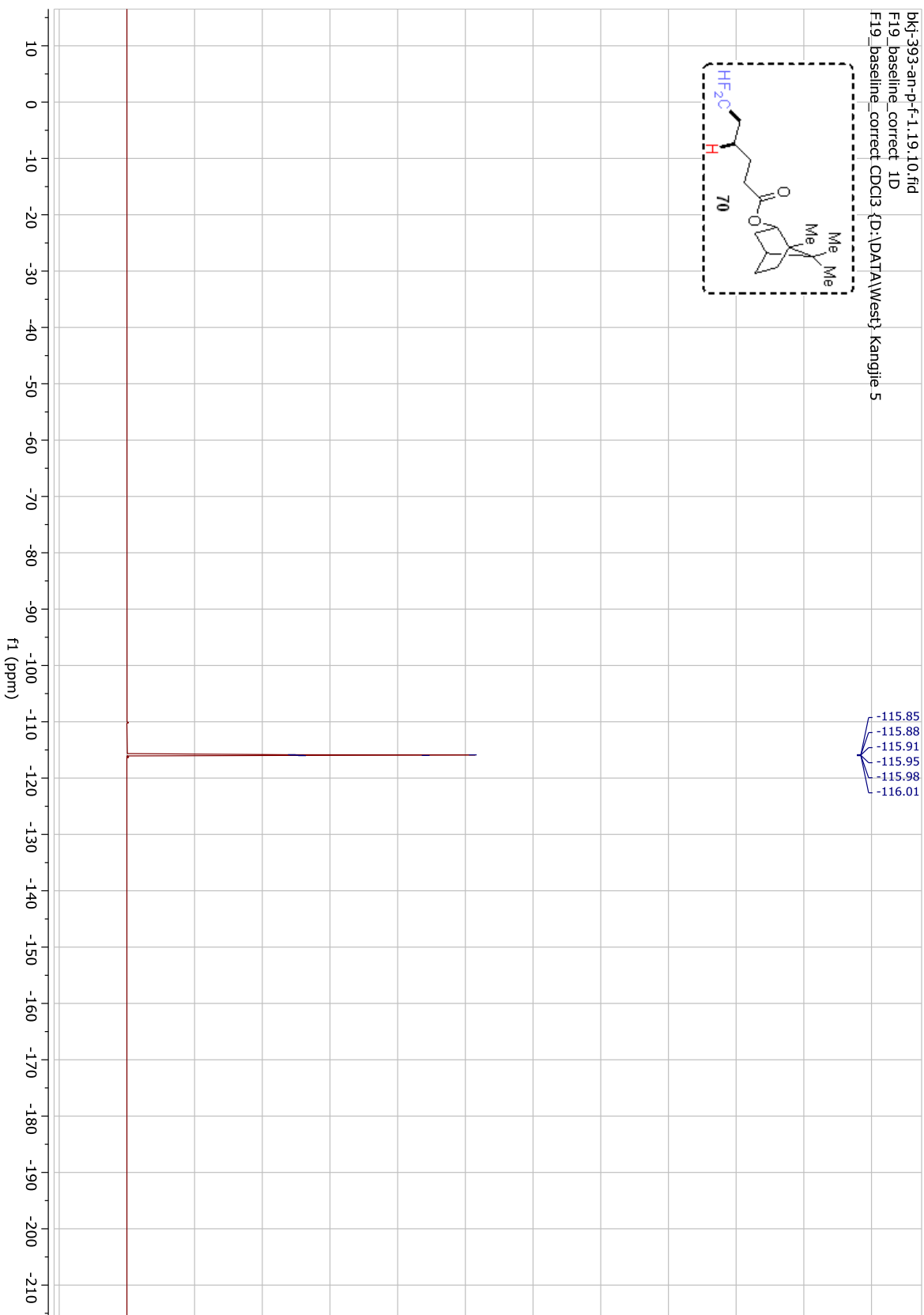
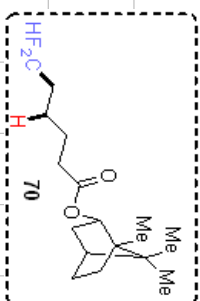




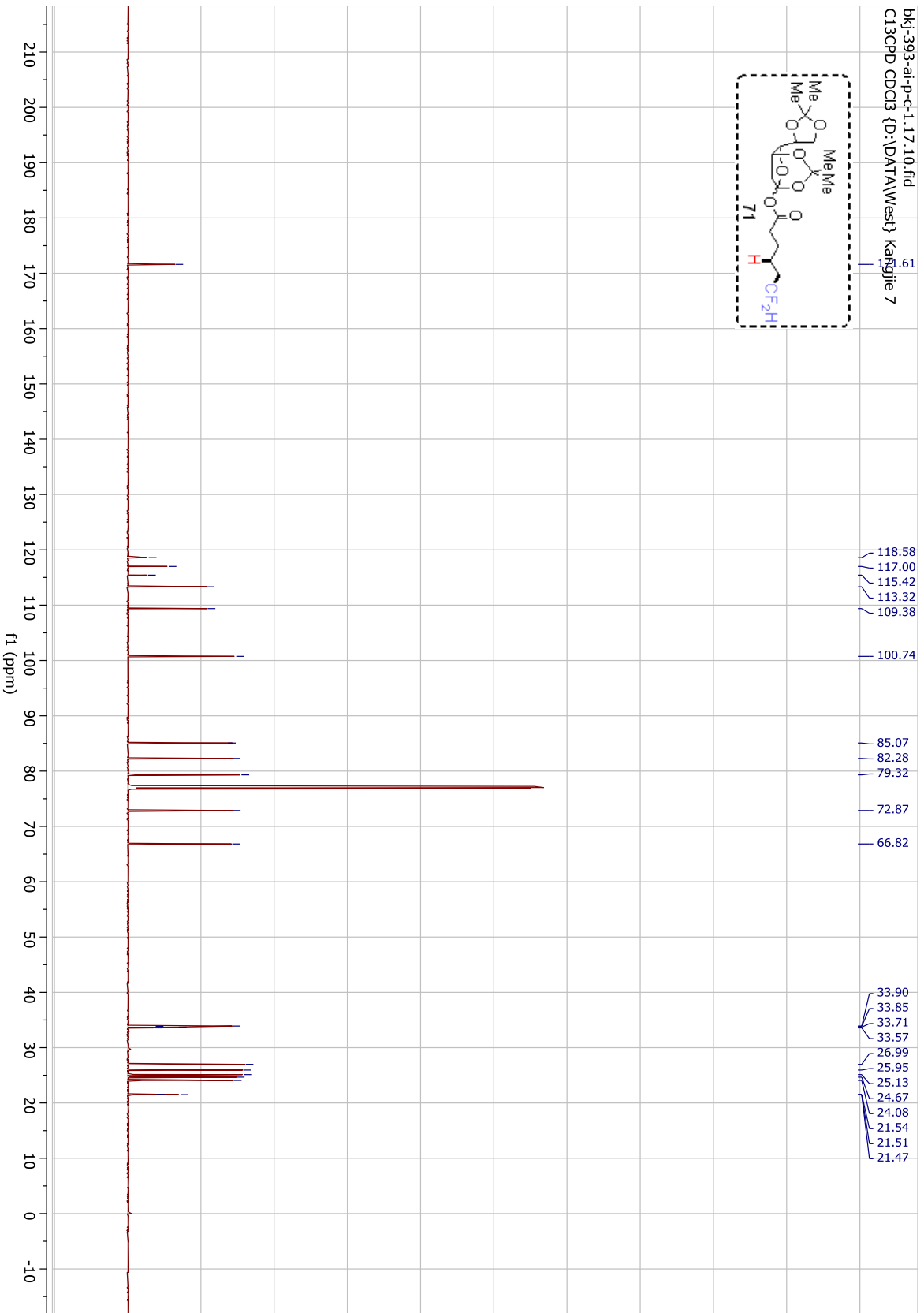
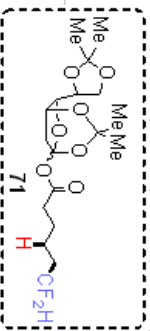
bkt-393-an-p-1.16.2.fid
C13CPD CDCl3 (E:\DATA_West\Kangjie 6



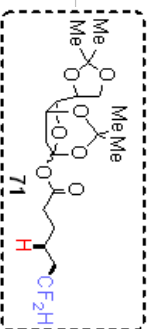
bkj-393-an-p-f-1.19.10.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West\ Kangjie 5



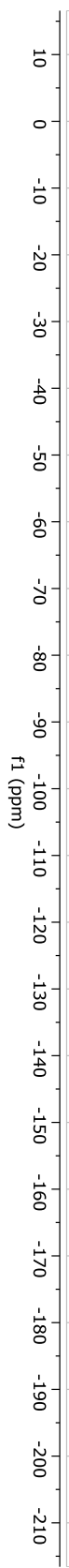
bkj-393-ai-p-c-1.17.10.fid
C13CPD CDCl3 (D:\DATA\Westj_Karjulle 7

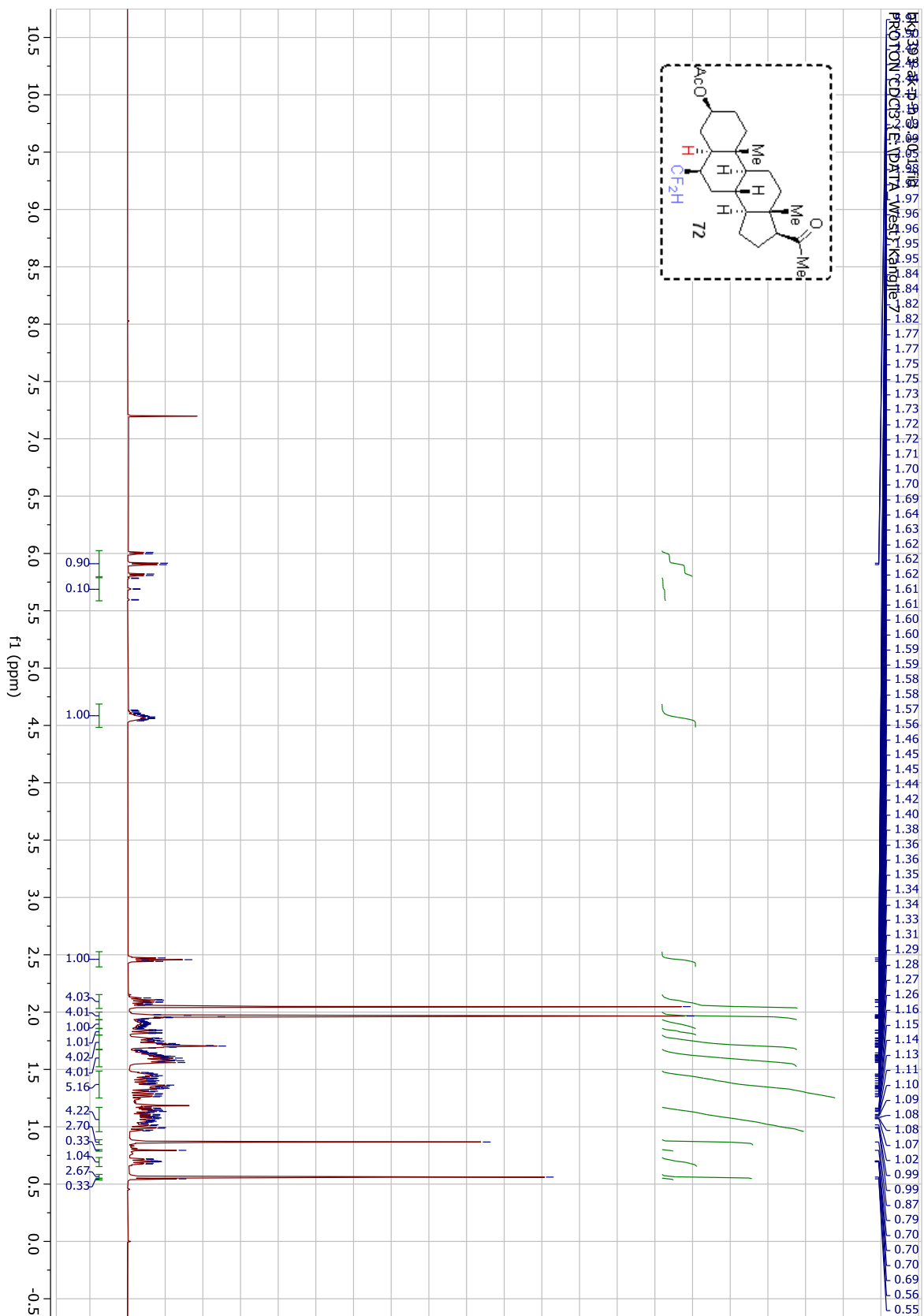


bkj-393-ai-p-f-1.18.1.fid
F19 CDCl3 {D:\DATA\West}\Kangjie 16

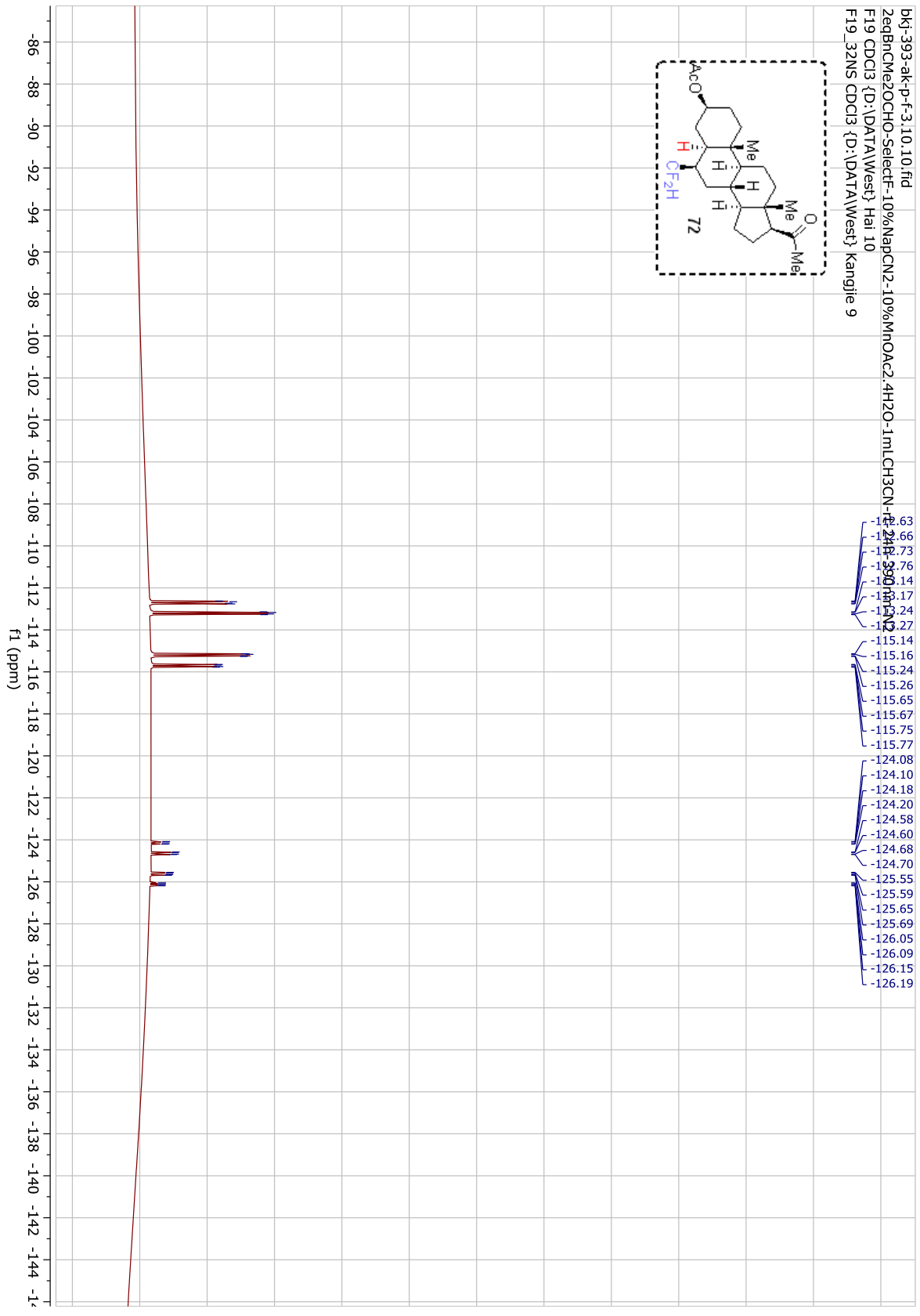
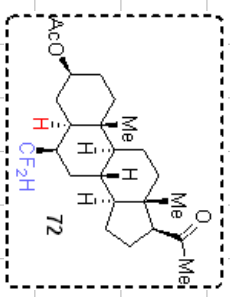


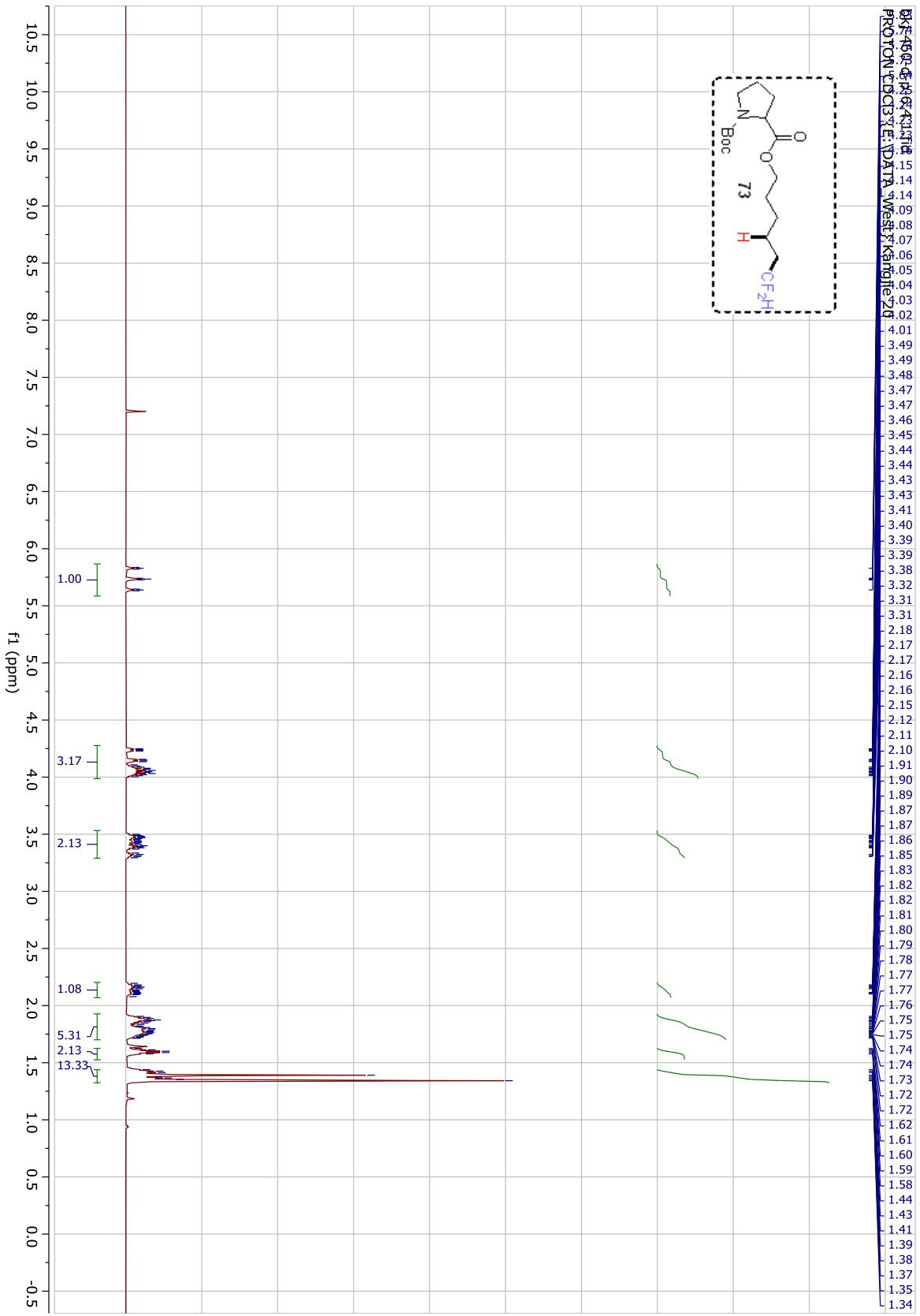
-115.93
-115.96
-115.99
-116.03
-116.03
-116.06
-116.09

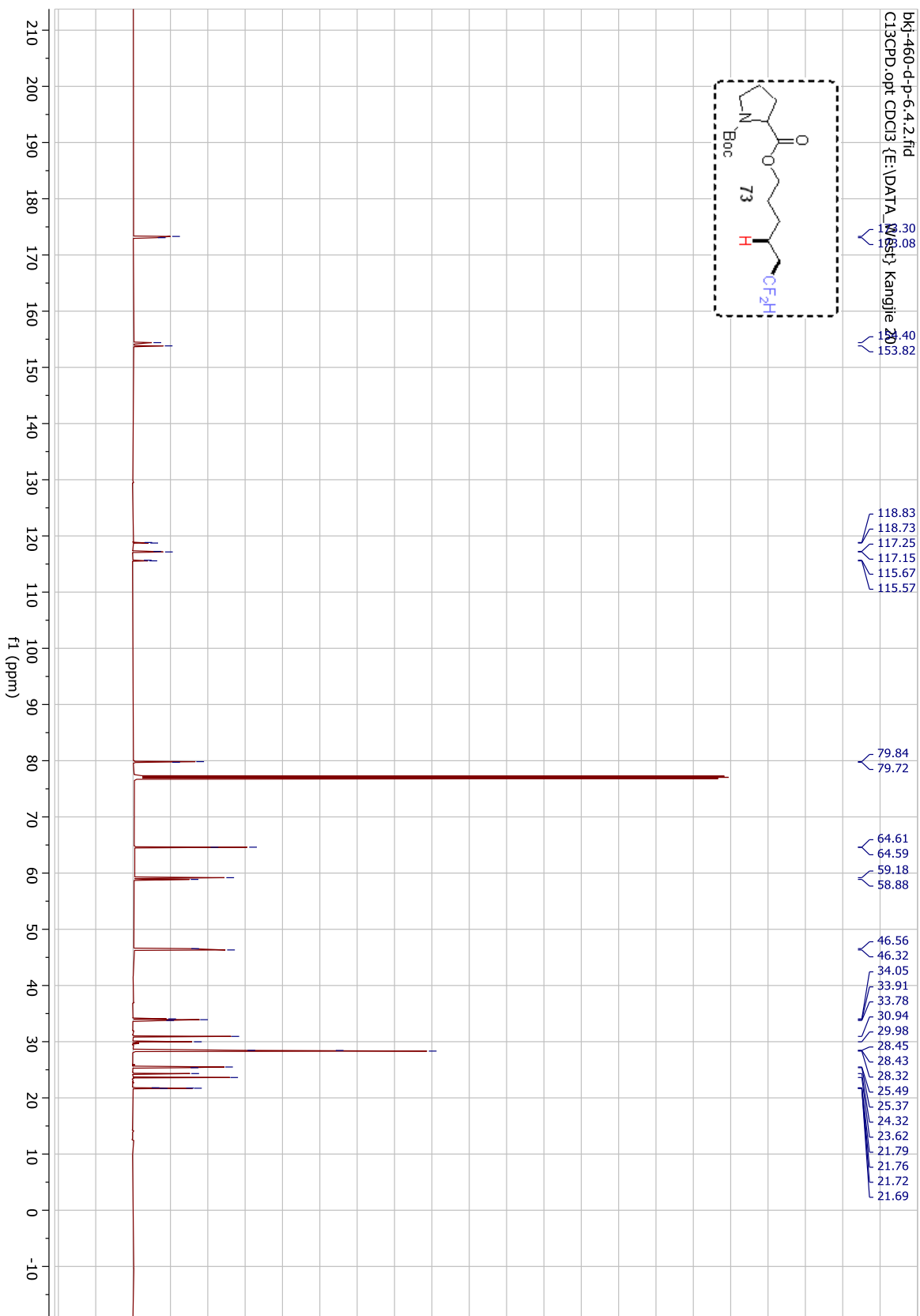




bk1_393-ak-p-f-3.10.10.fid
ZedBnCMz2OCHO-SelectF-10%NappCN2-10%MnOAc2.4H2O-1mLCH3CN-H
F19_CDCl3 {D:\DATA\West\ Hai 10
F19_3ZMS CDCl3 {D:\DATA\West\ Kangjie 9

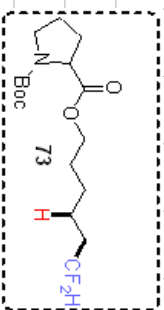




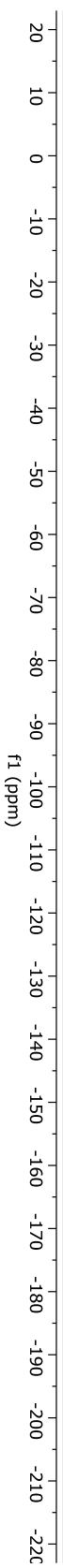


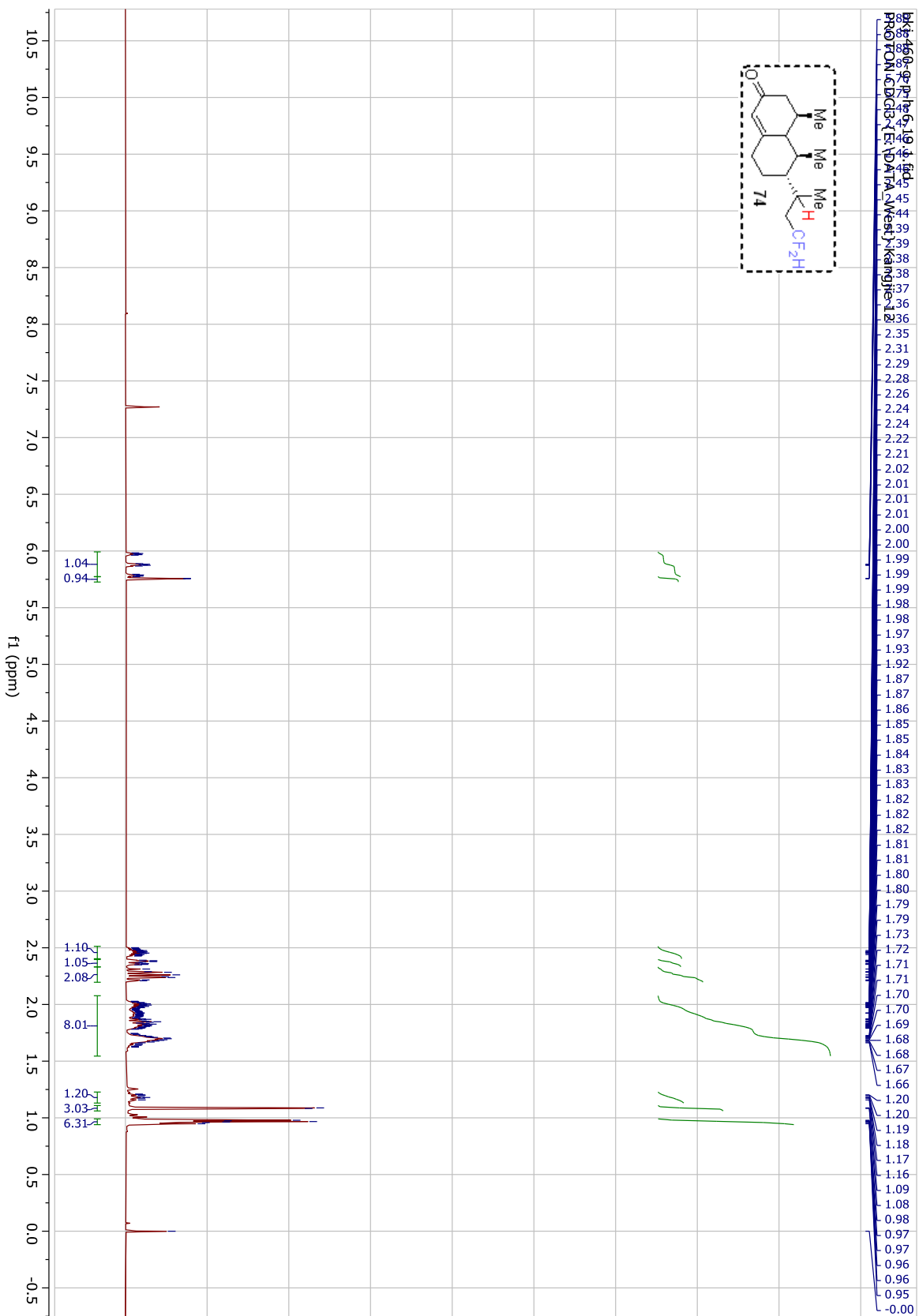
bkj-460-d-p-f-6.5.21.fid
F19

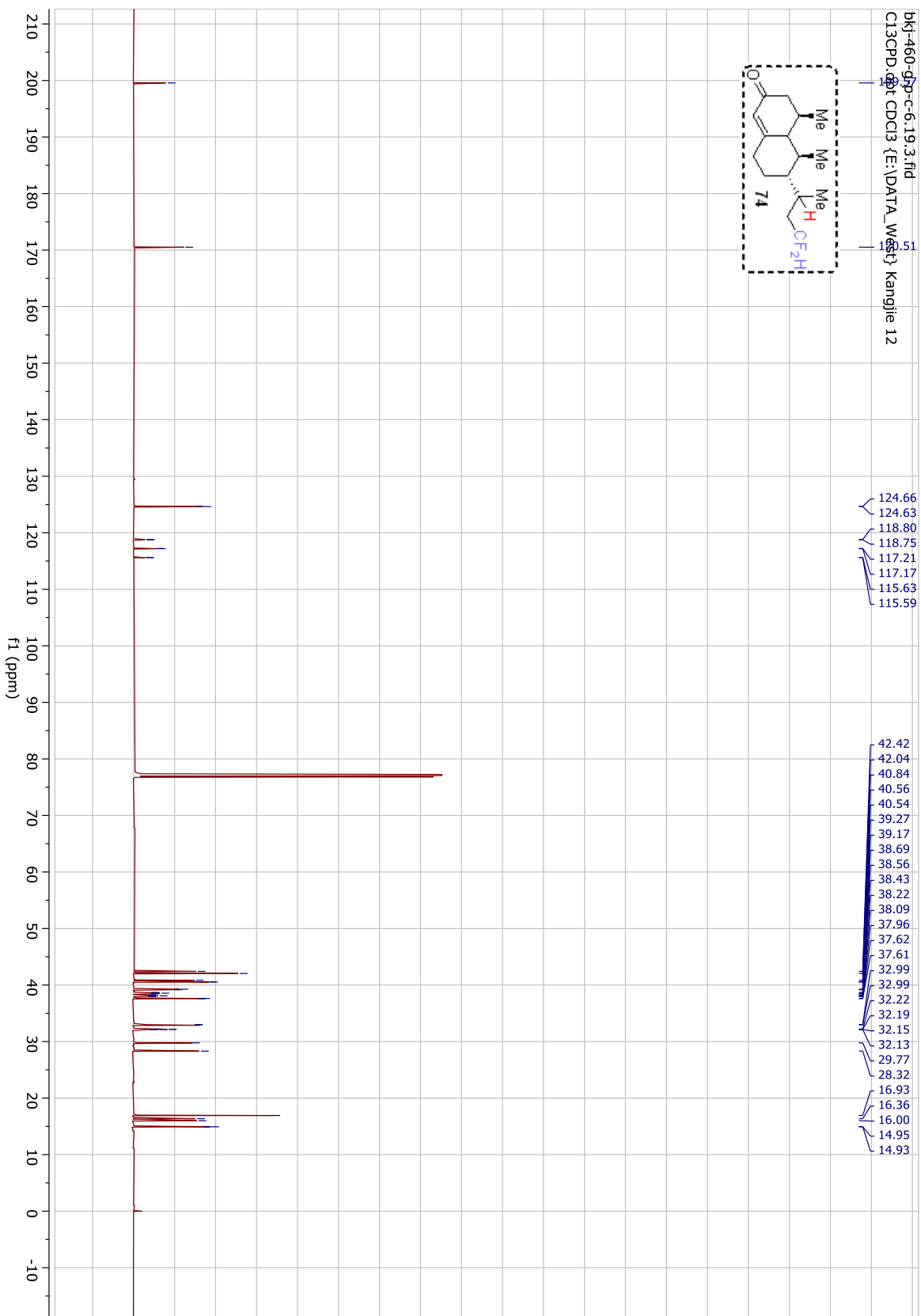
CMC_F19 CDCl3 (D:\DATA\West) Kangjie 23



-115.82
-115.85
-115.87
-115.88
-115.90
-115.92
-115.93
-115.95
-115.97
-115.98
-116.00
-116.03

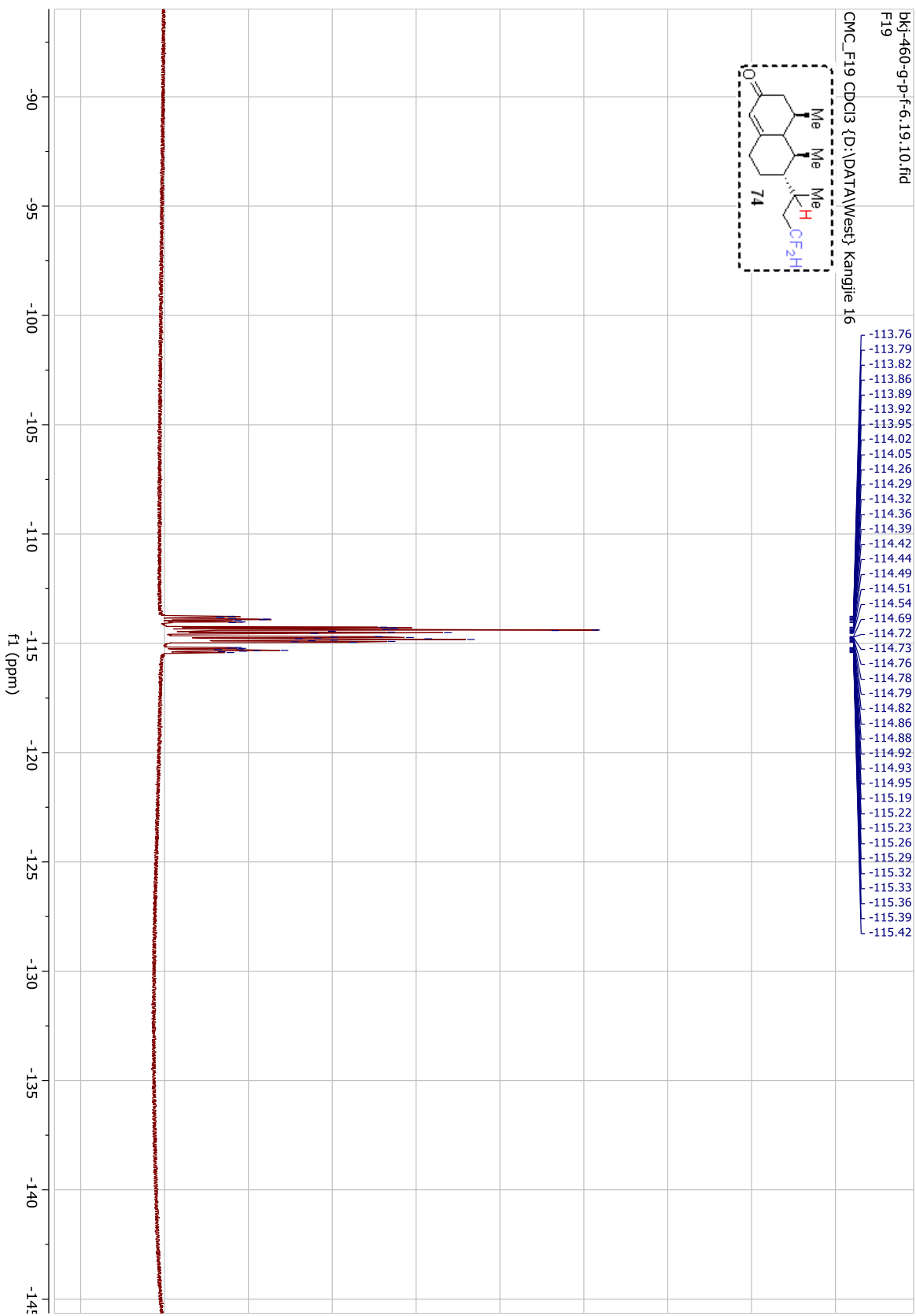
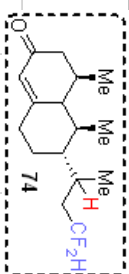


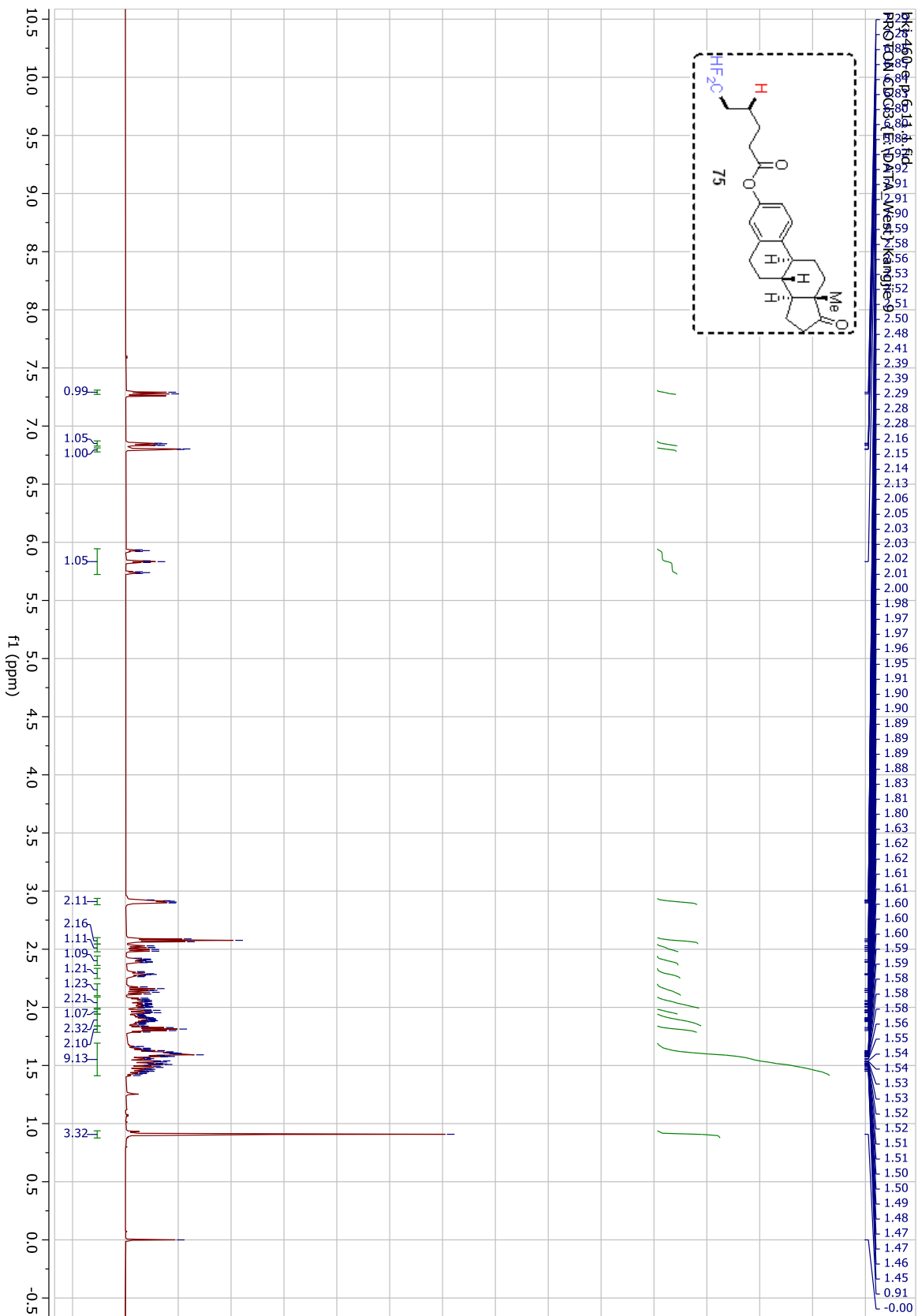


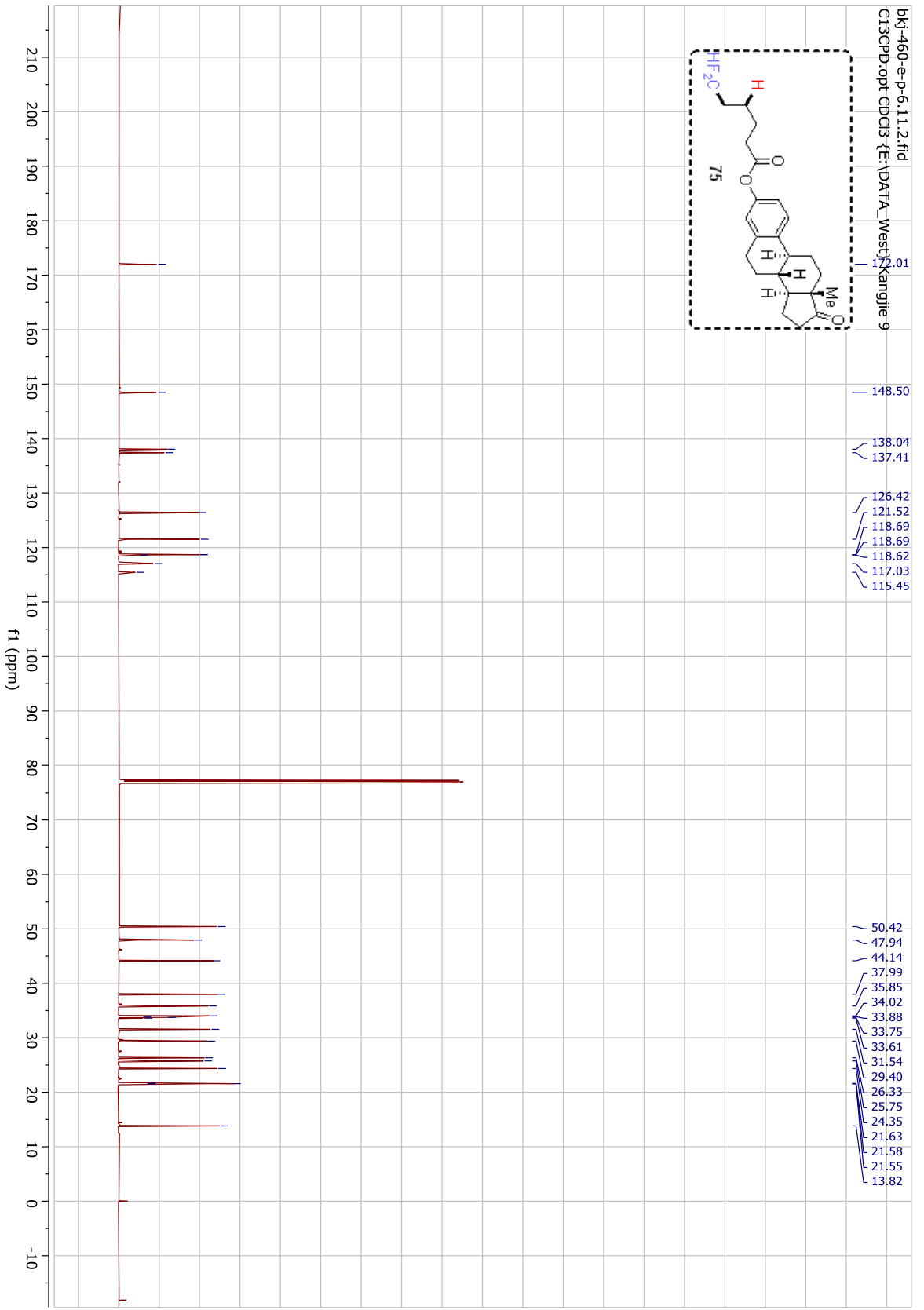


bj-460-g-p-f-6.19.10.ftd
F19

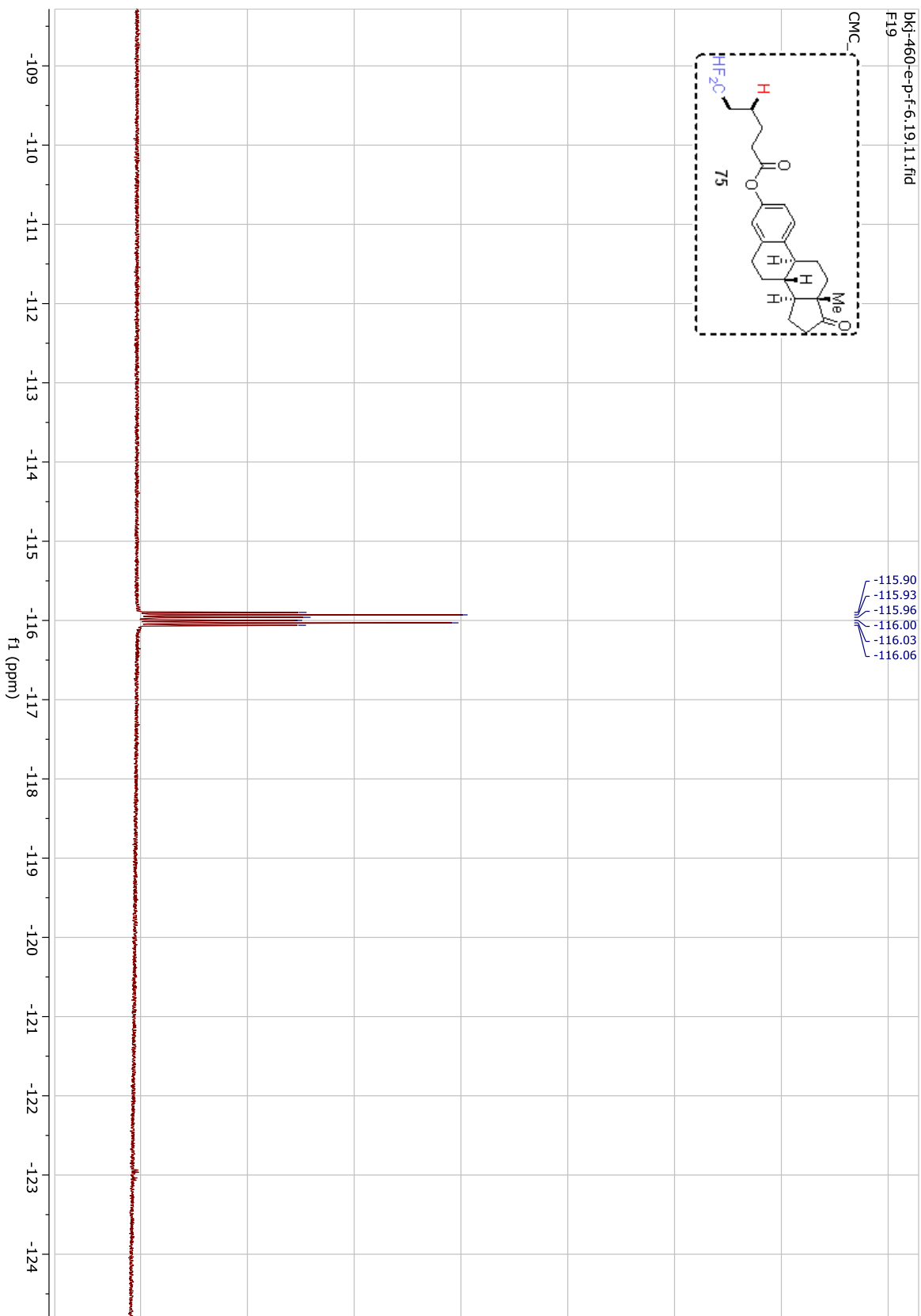
CMC_F19 CDCl3 (D:\DATA\West) Kangjie 16

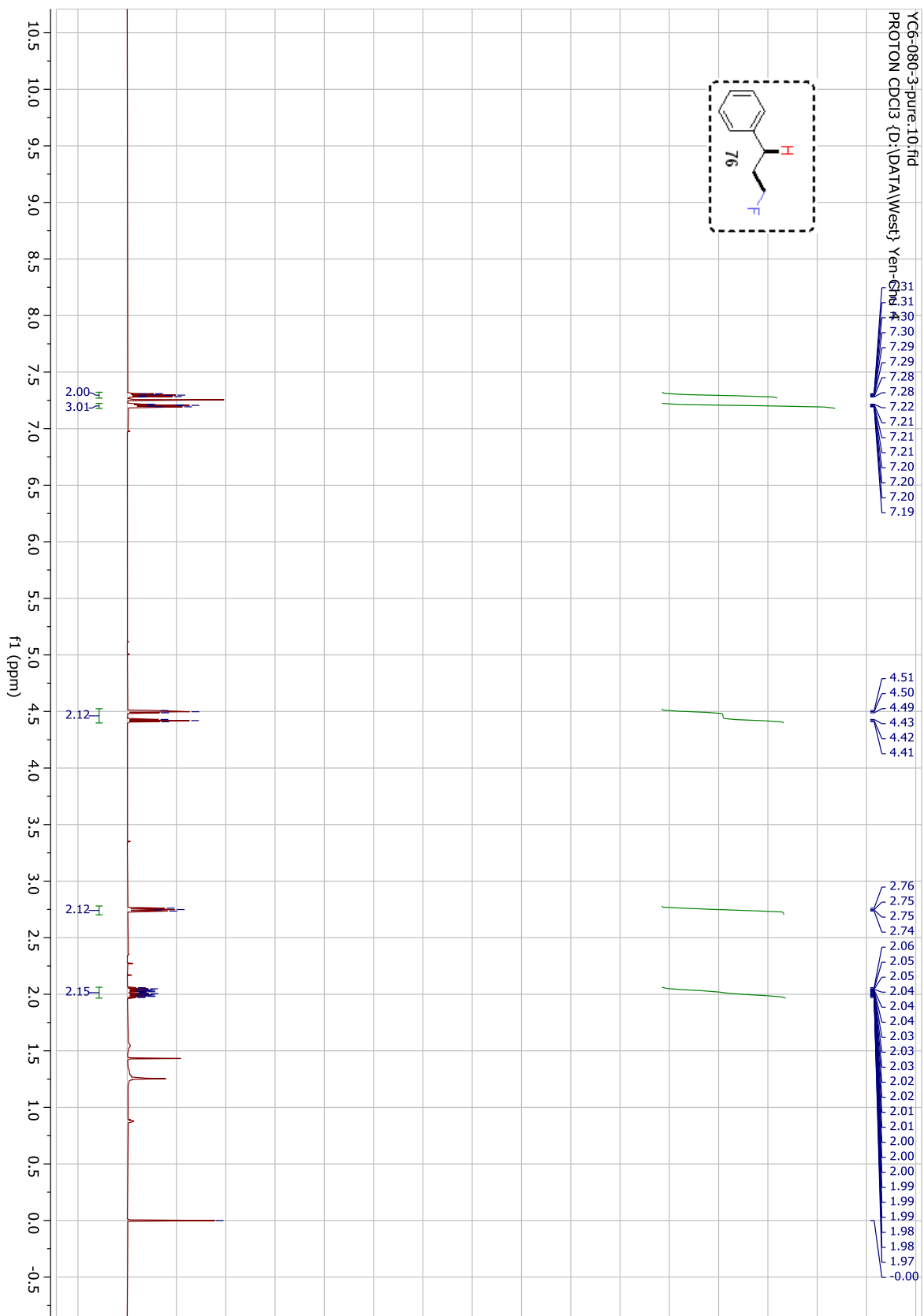




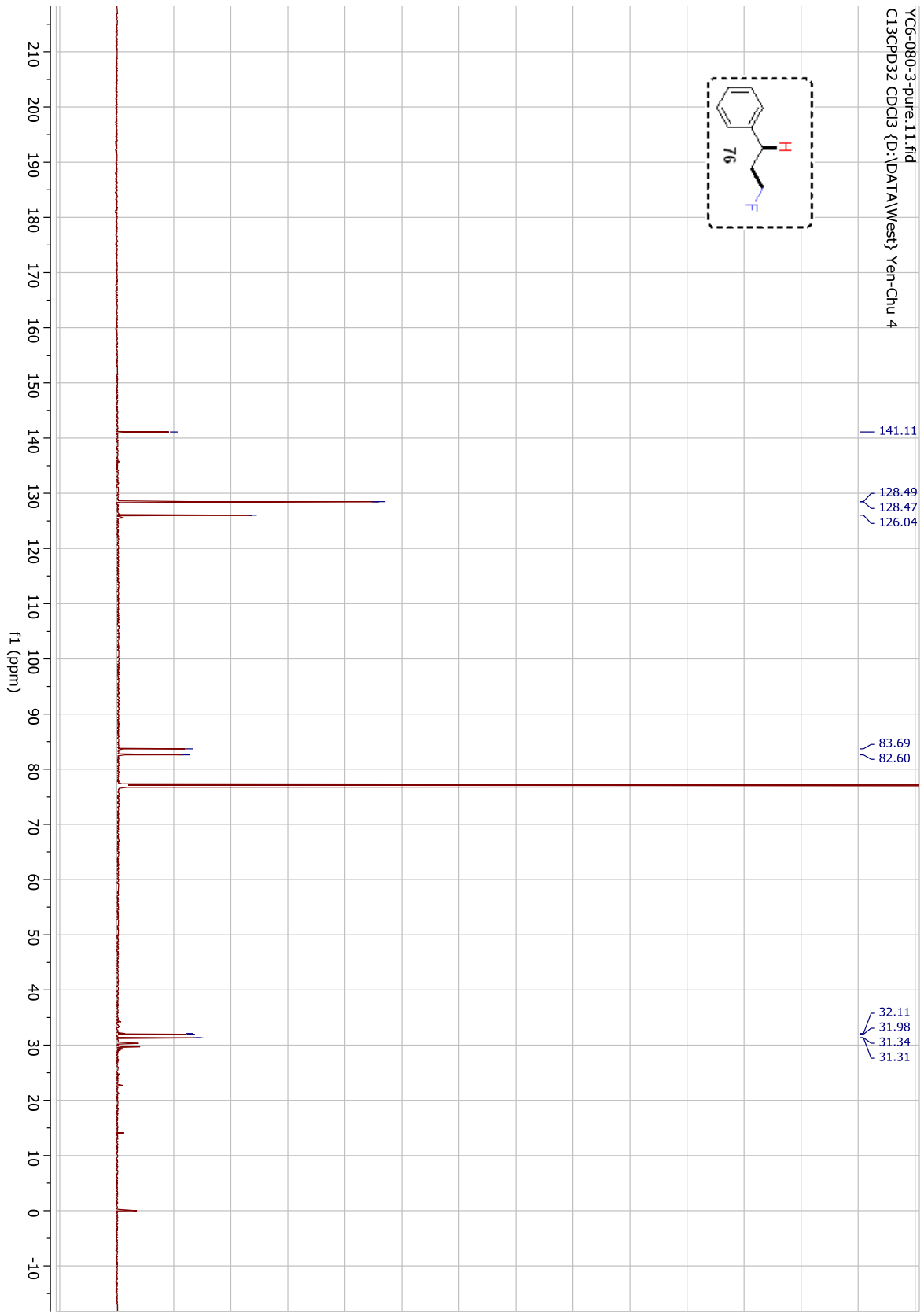
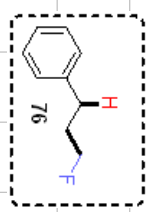


bkj-460-e-p-f-6.19.11.ftd
F19

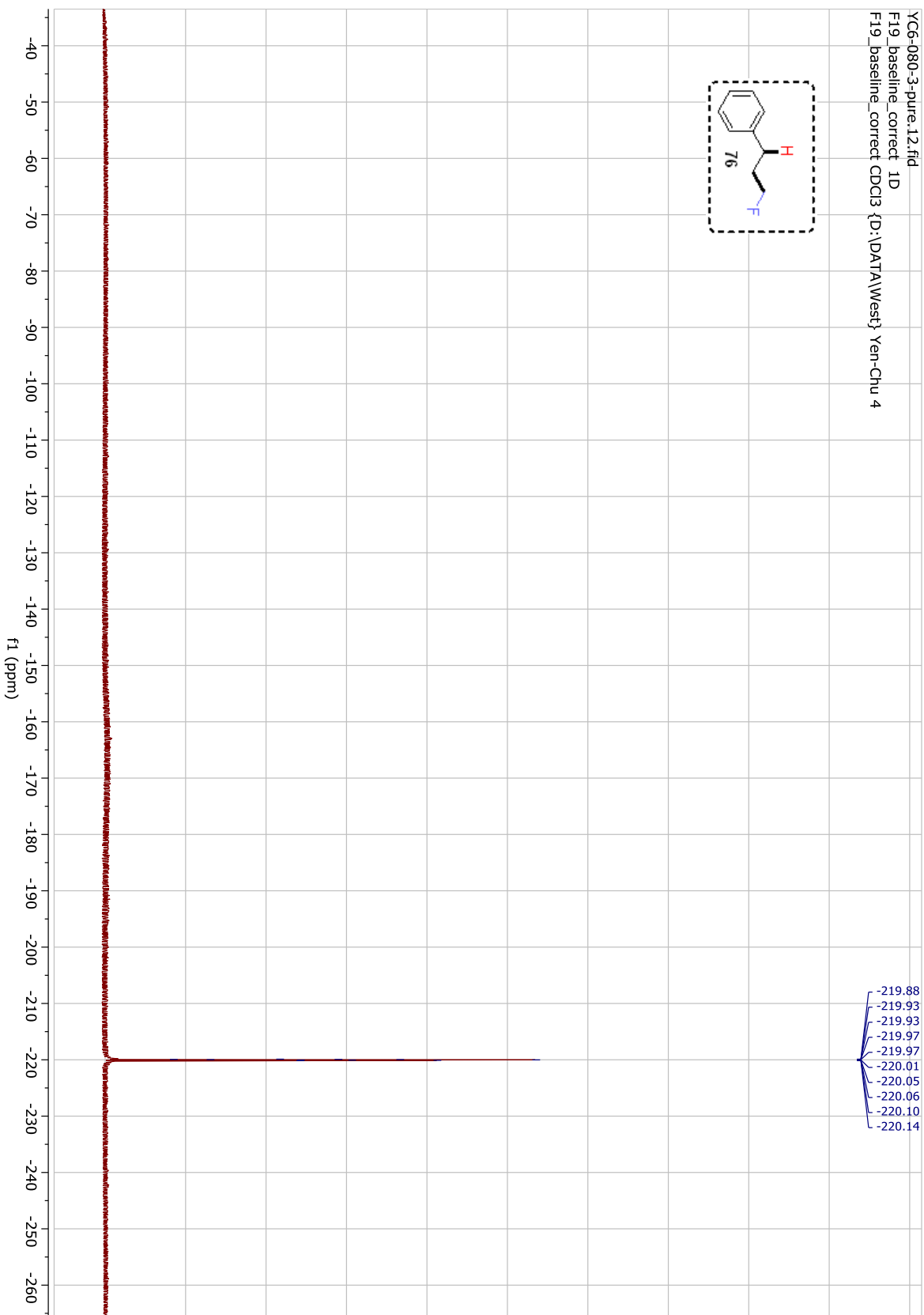
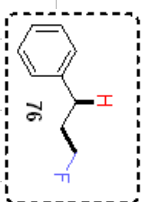




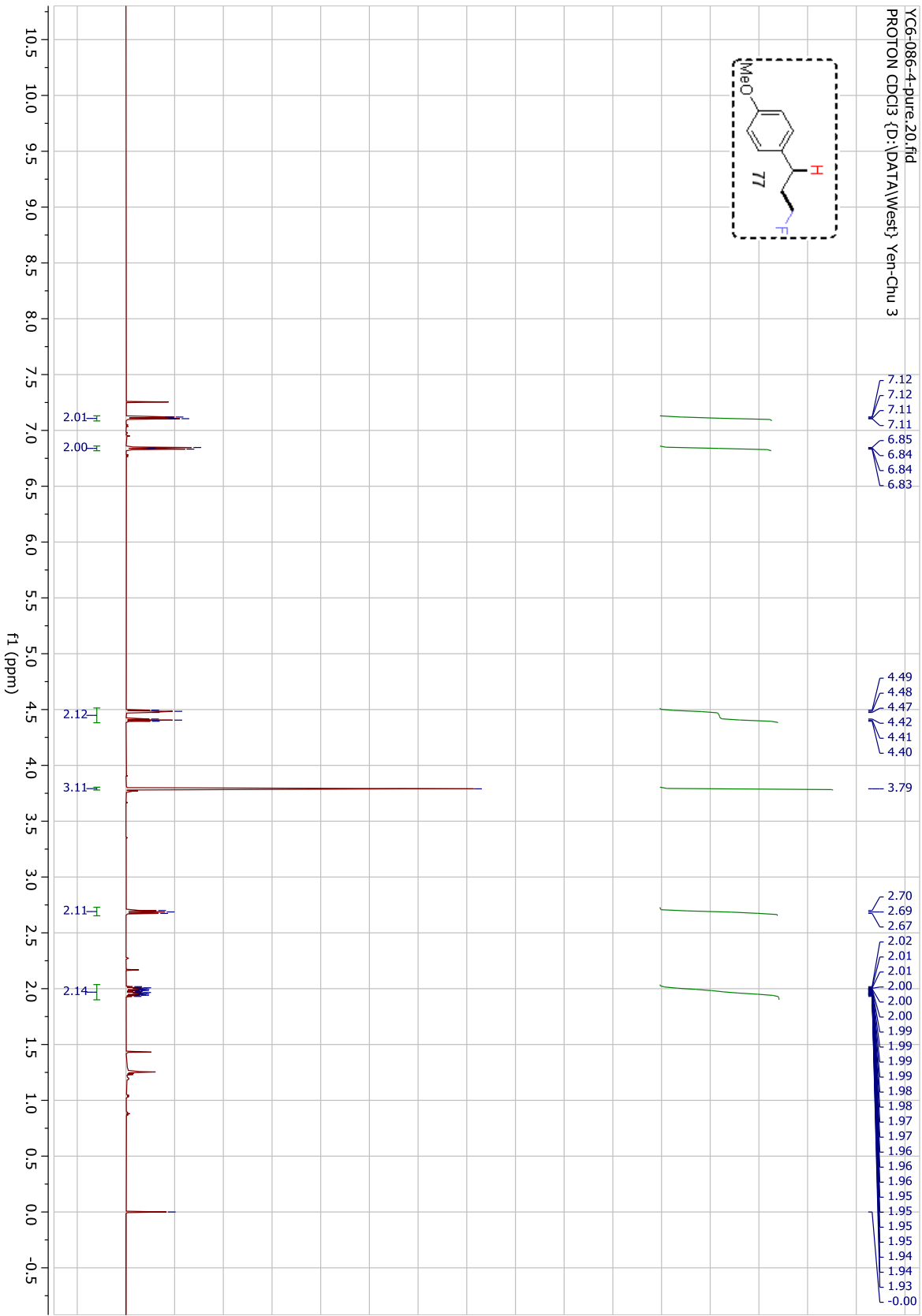
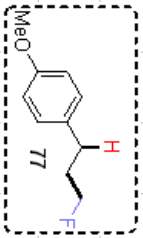
YC6-080-3-pure.11.fid
C13CPD32 CDCl3 {D:\DATA\West}\Yen-Chu 4



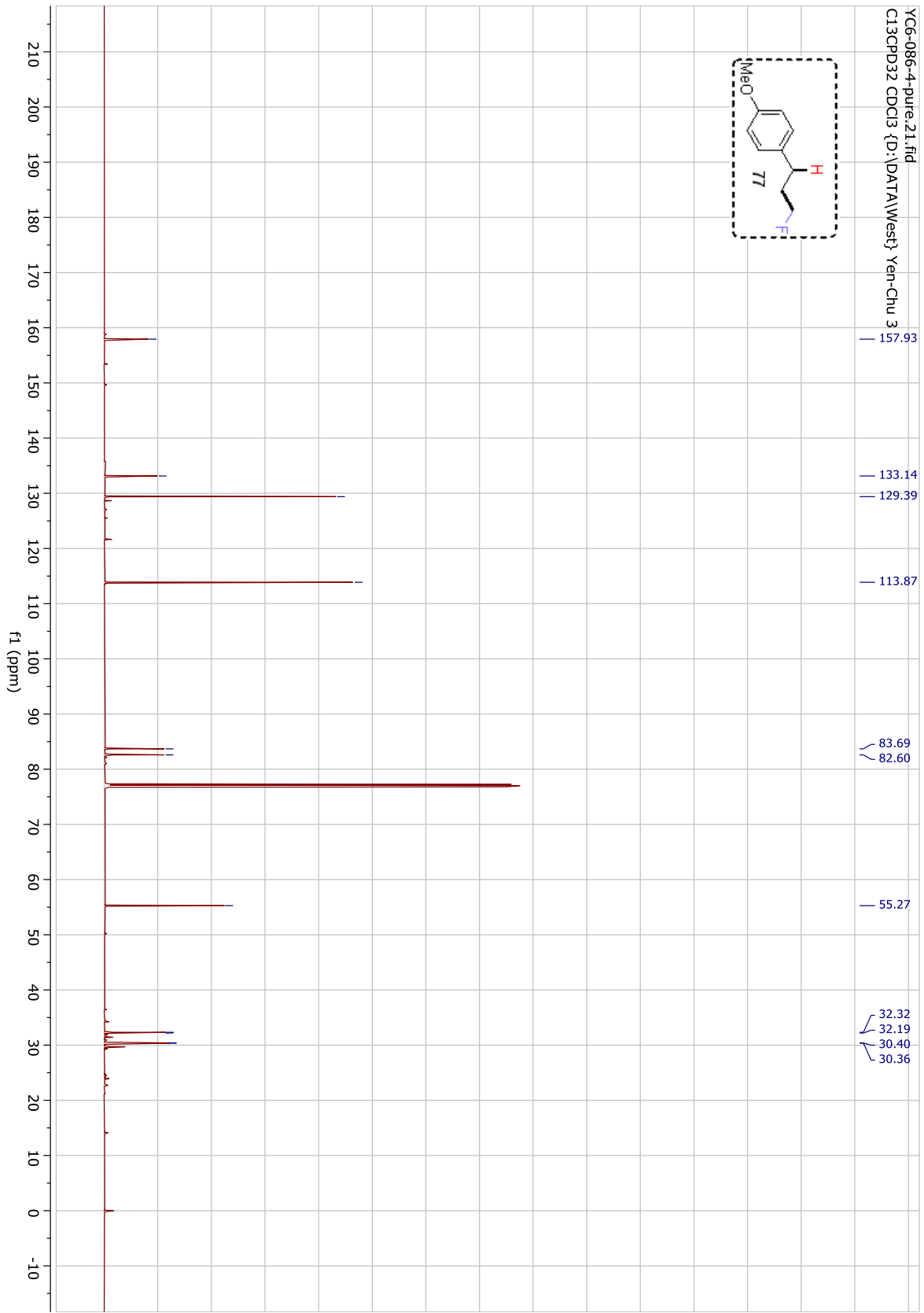
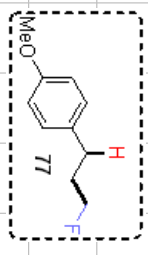
YC6-080-3-pure.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Yen-Chu 4



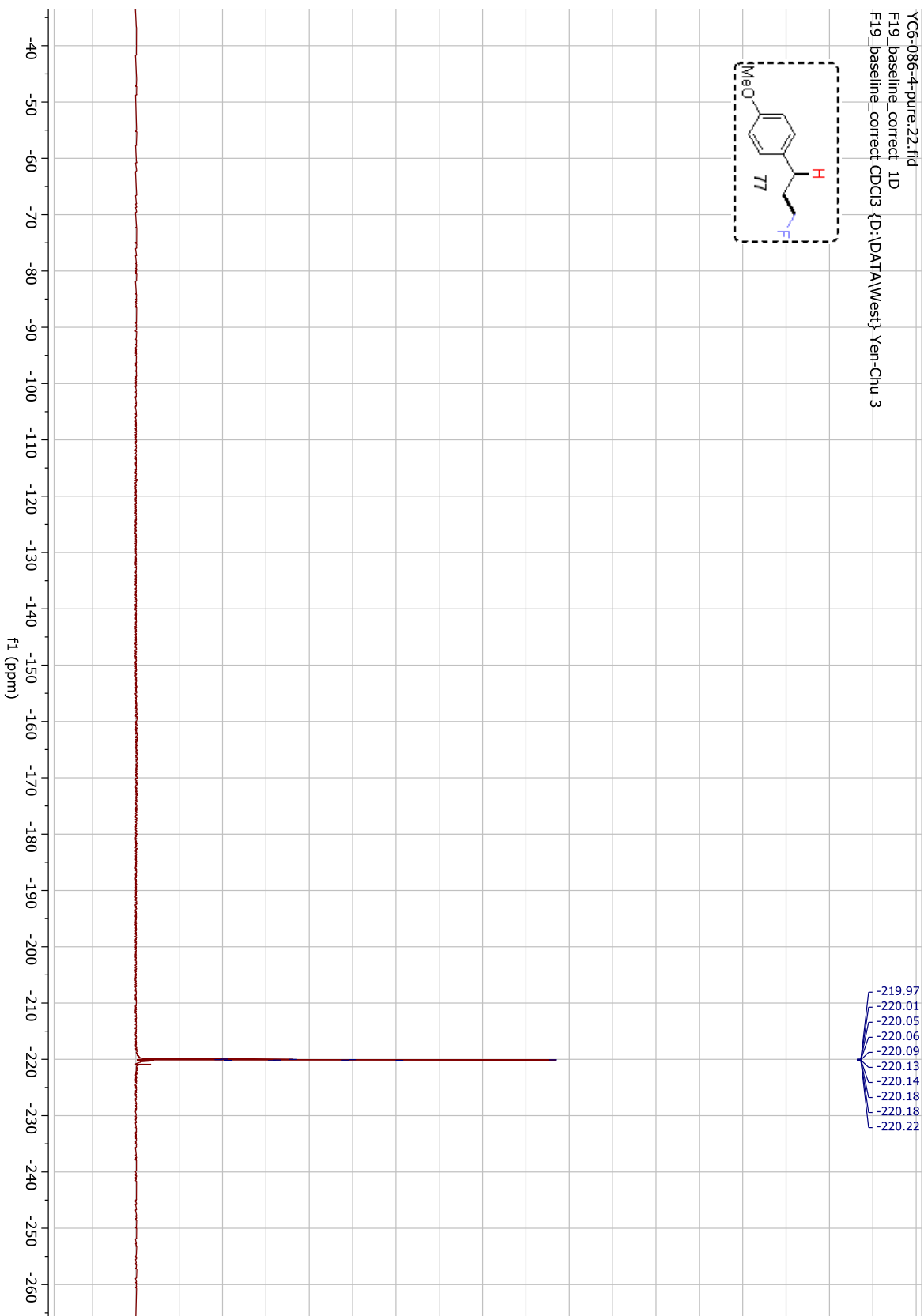
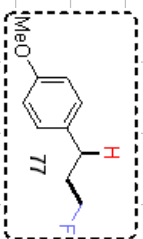
YC6-086-4-pure.20.fid
PROTON CDCl3 {D:\DATA\Westf\Yen-Chu 3



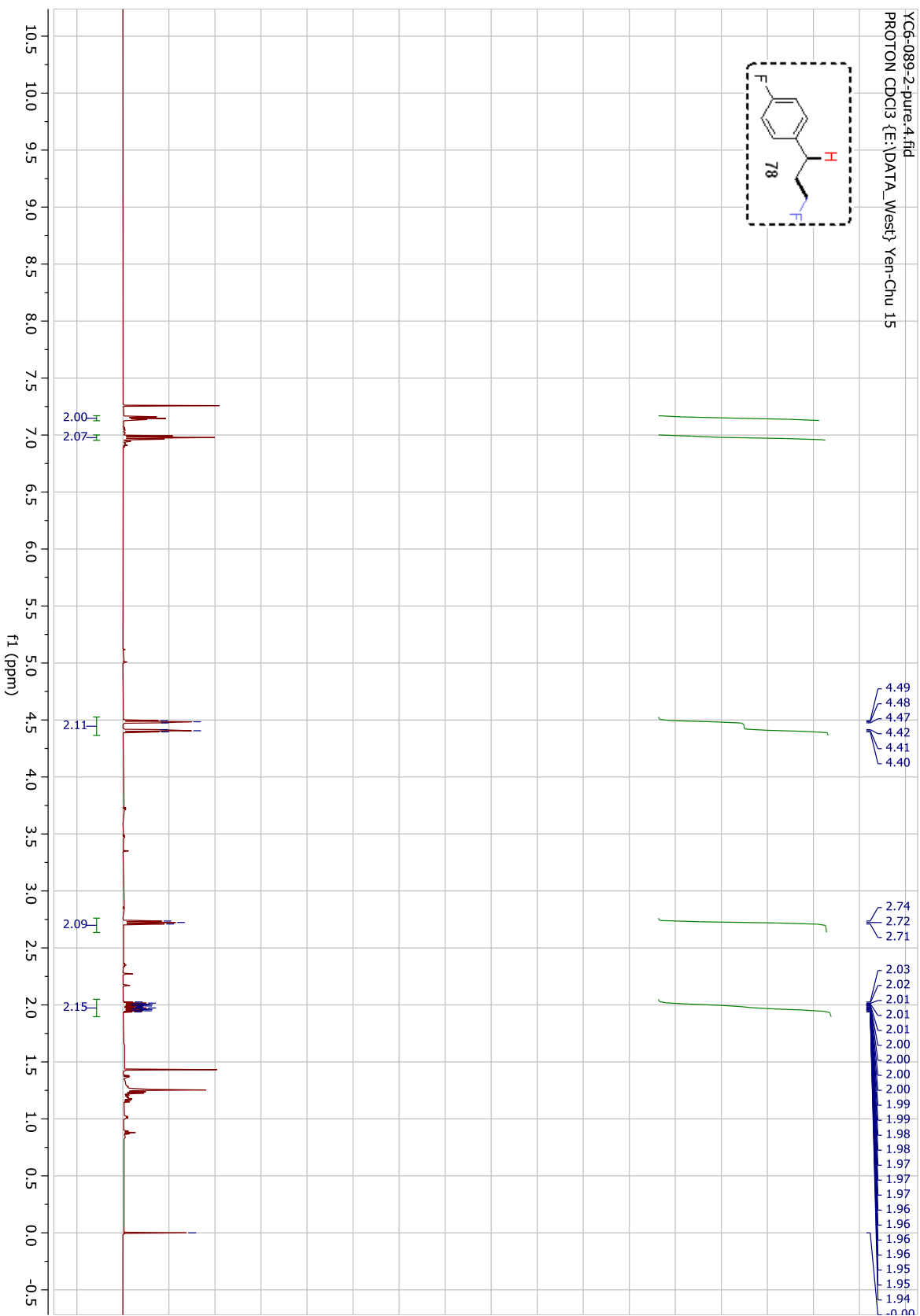
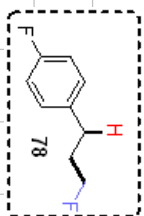
YC6-086-4-pure.21.fid
C13CPD32 CDCl3 {D:\DATA\West}\Yen-Chu 3

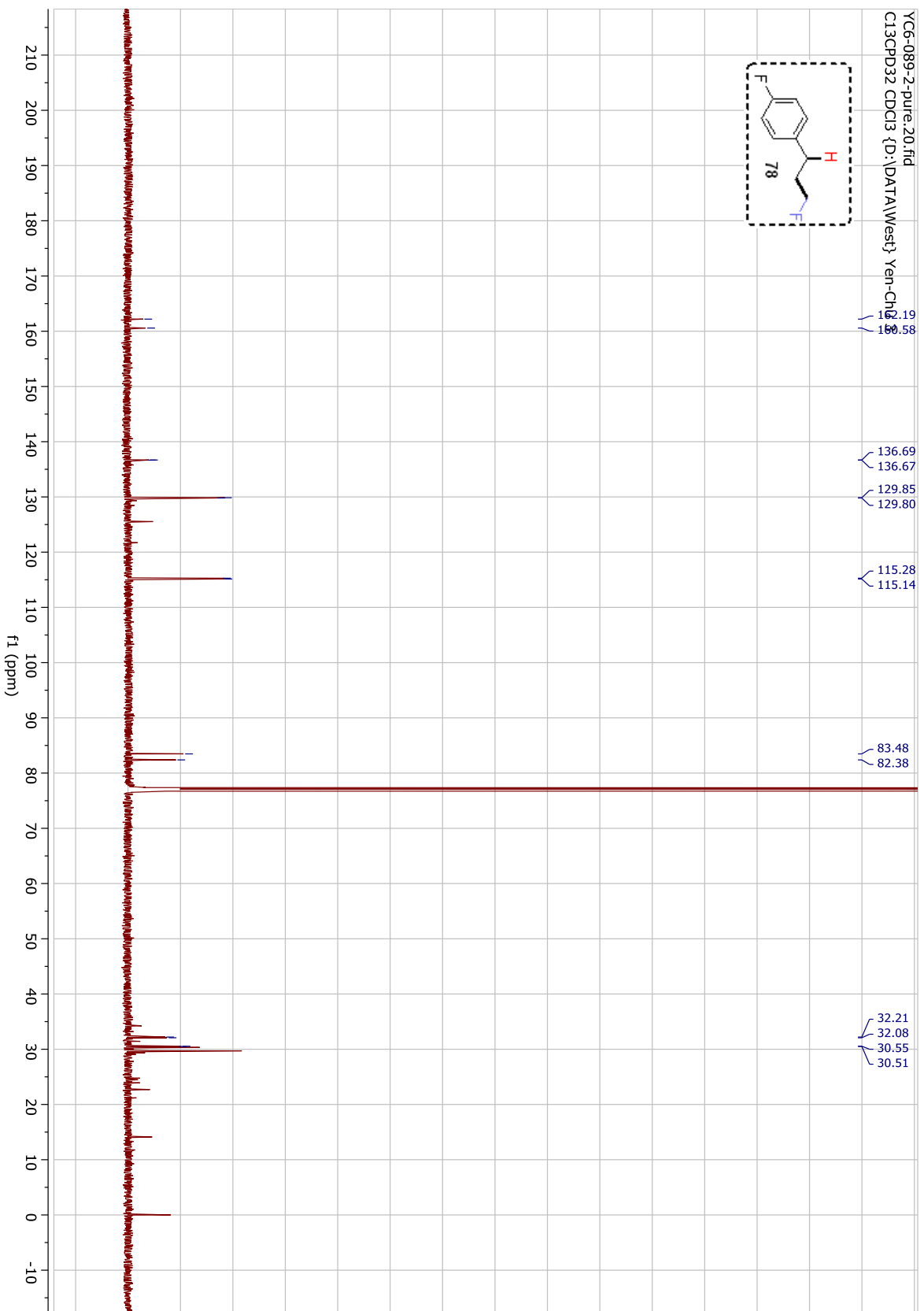


YC6-086-4-pure.22.ftd
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Yen-Chu 3

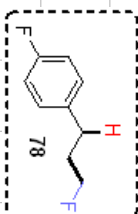


YC6-089-2-pure 4.fid
PROTON CDCl3 {E:\DATA_West\Yen-Chu 15



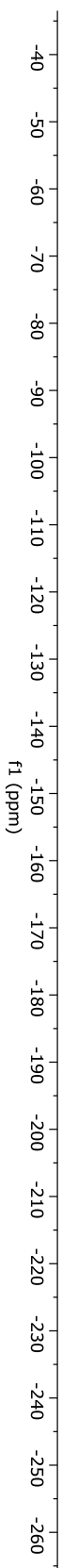


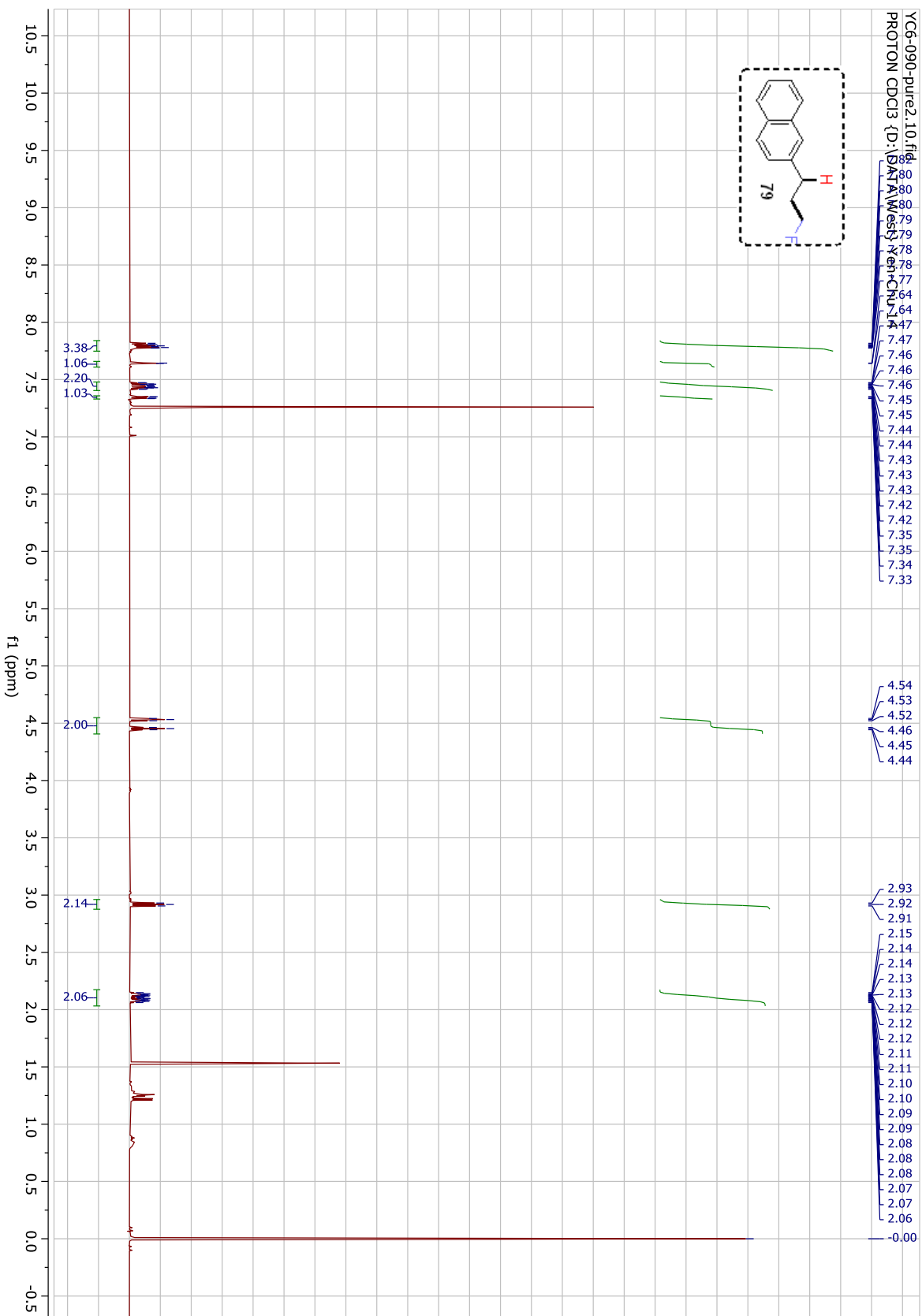
YC6-089-2-pure.60.fid
F19_baseline_correct_1D
F19_baseline_correct_CDCl3 {D:\DATA\West}\Yen-Chu 9



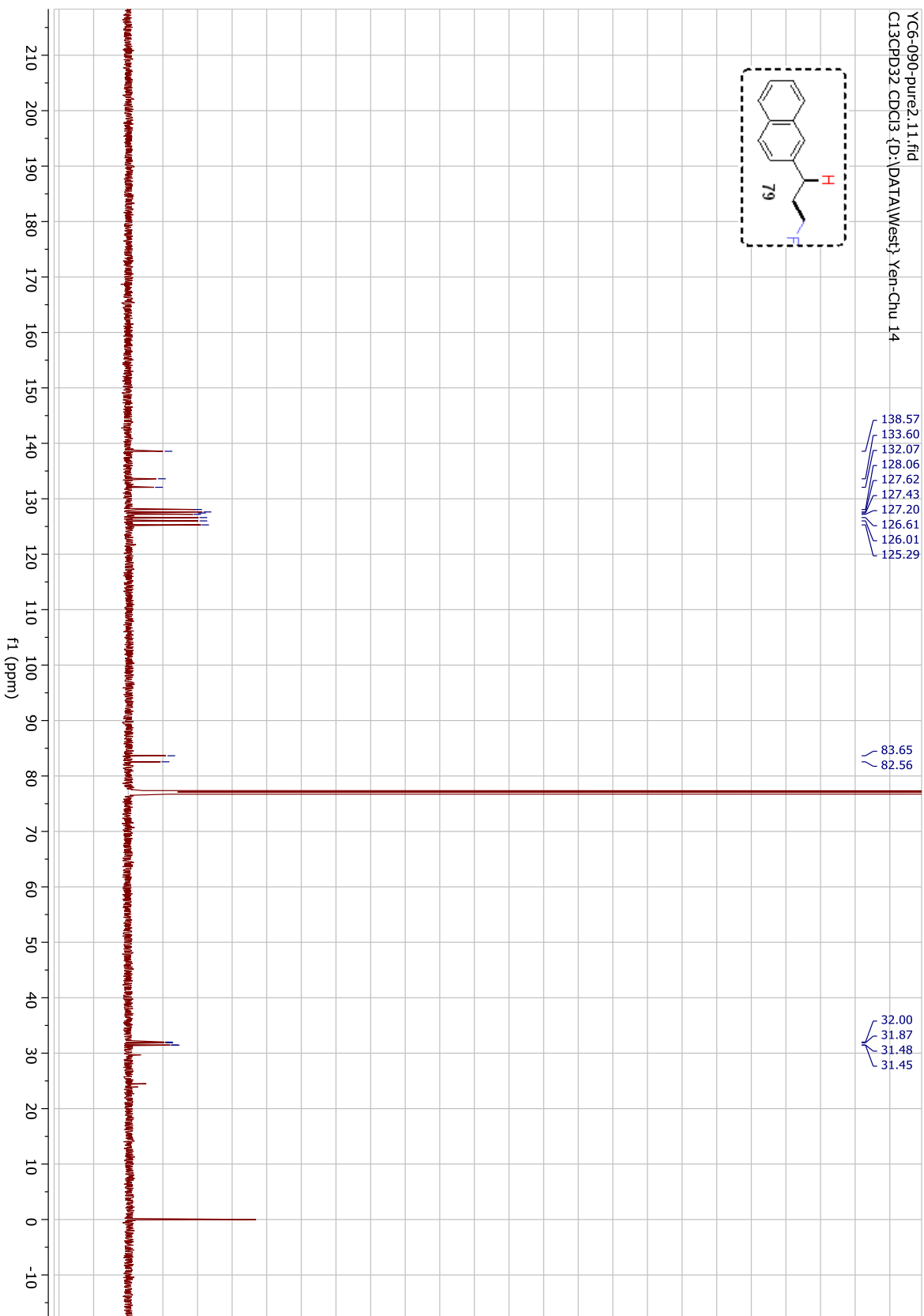
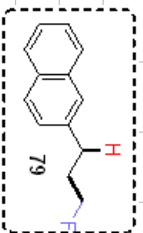
-117.42
-117.43
-117.44
-117.45
-117.46
-117.47
-117.48

-220.19
-220.24
-220.28
-220.29
-220.32
-220.36
-220.37
-220.41
-220.45

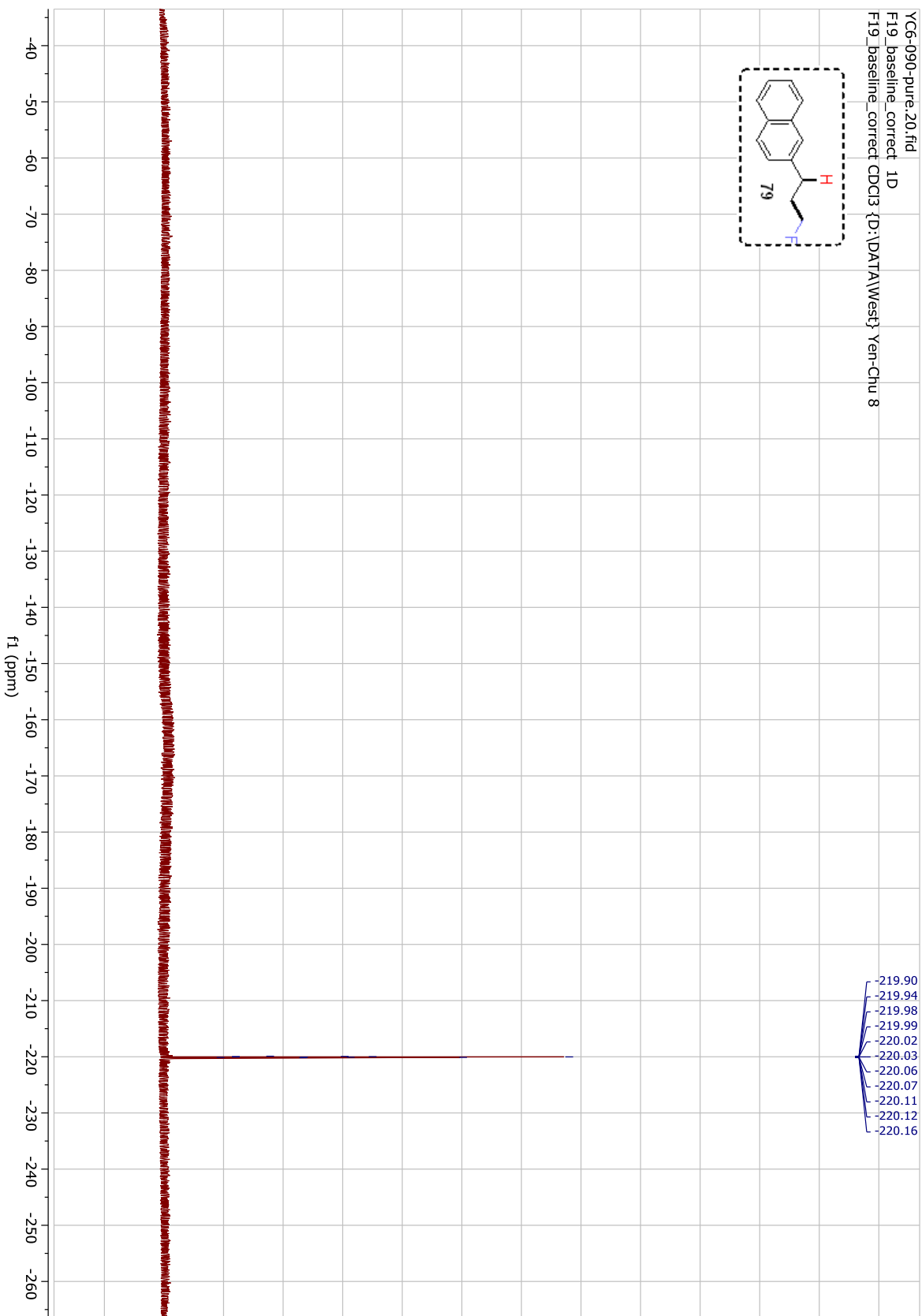
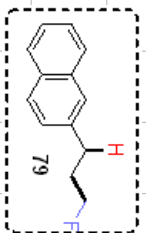


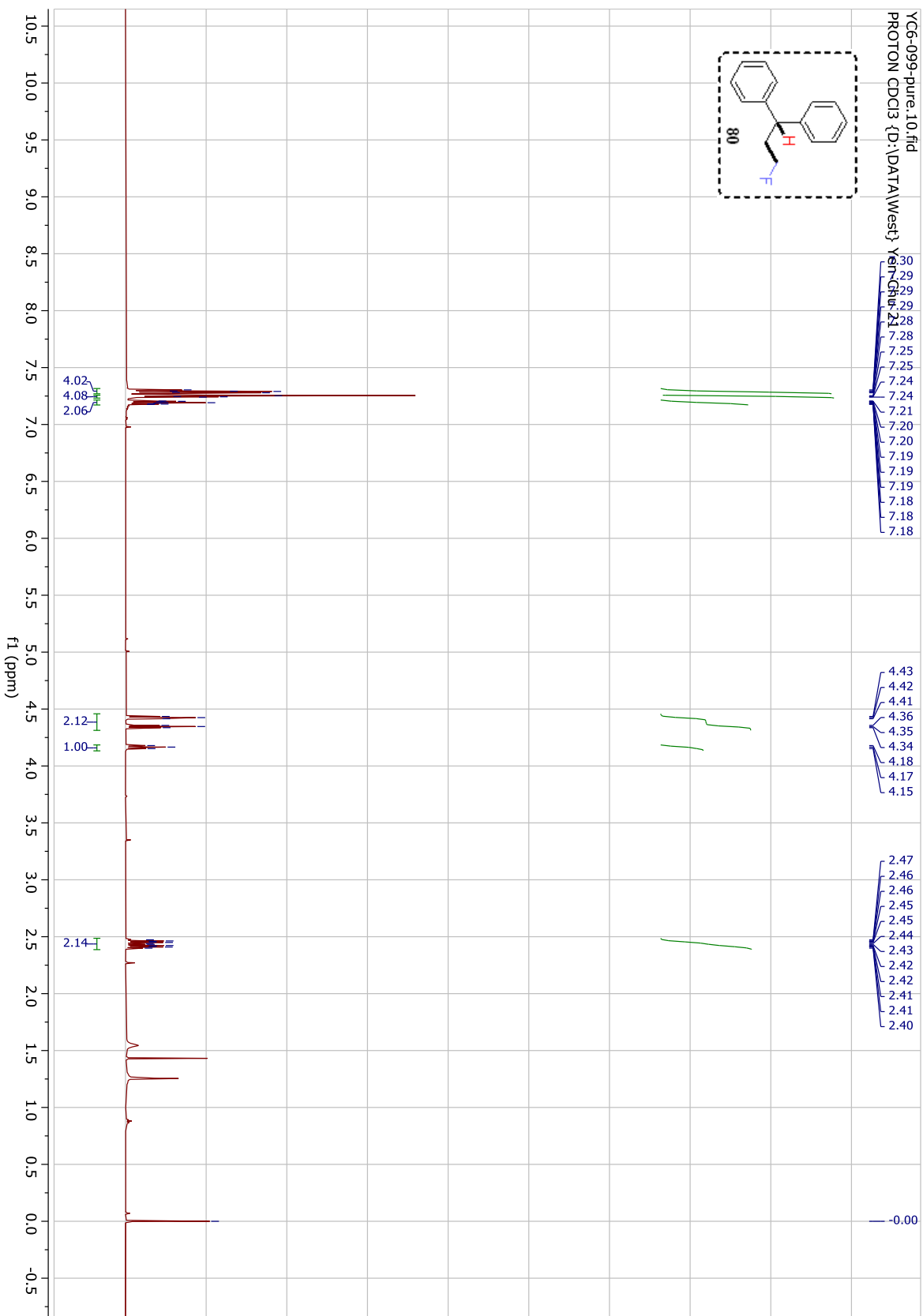


YC6-090-pure2_11.fid
C13CPD32 CDCl3 {D:\DATA\Westf\Yen-Chu 14

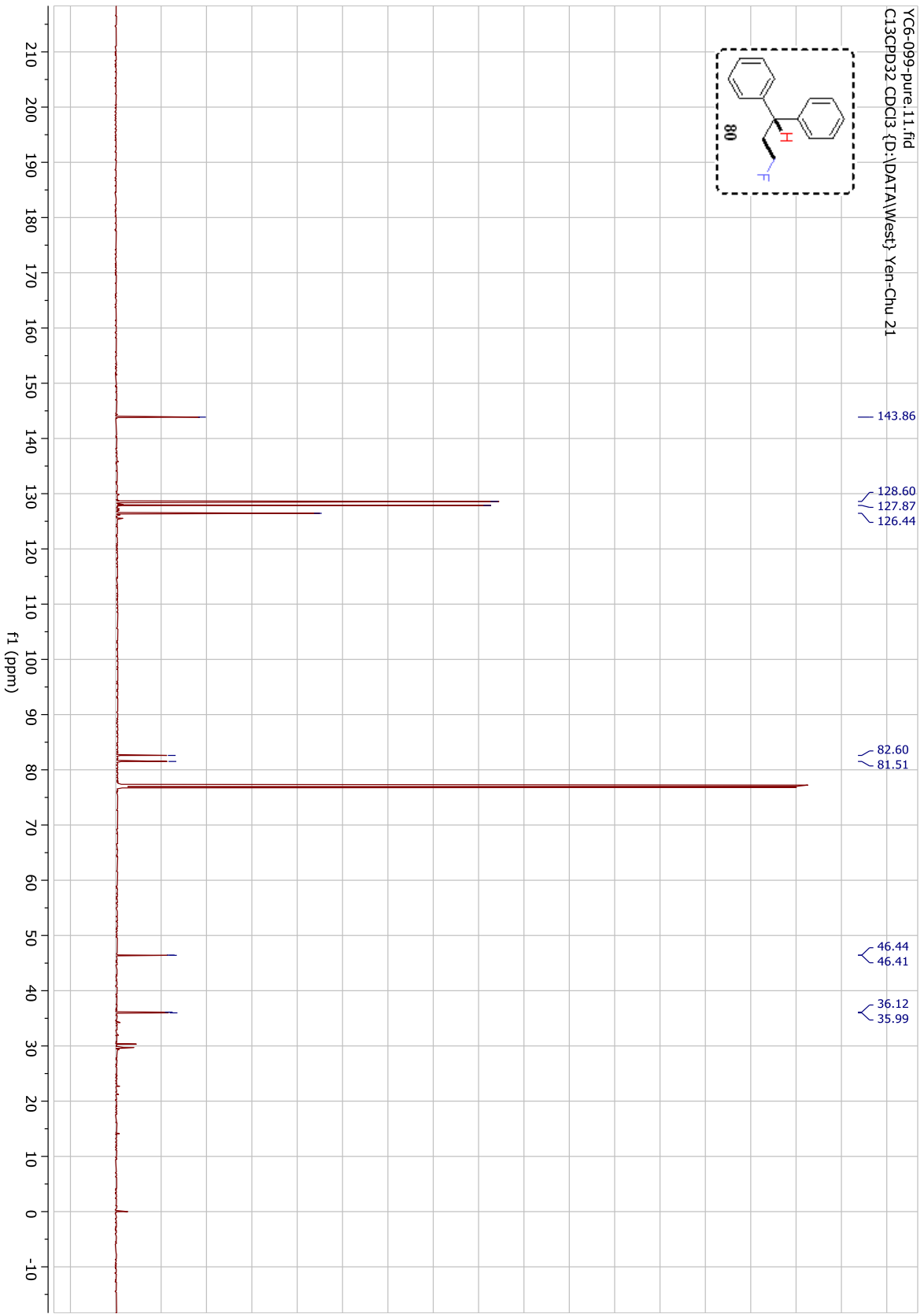
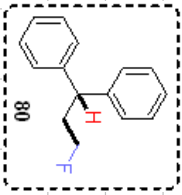


YC6-090-pure.20.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West\ Yen-Chu 8

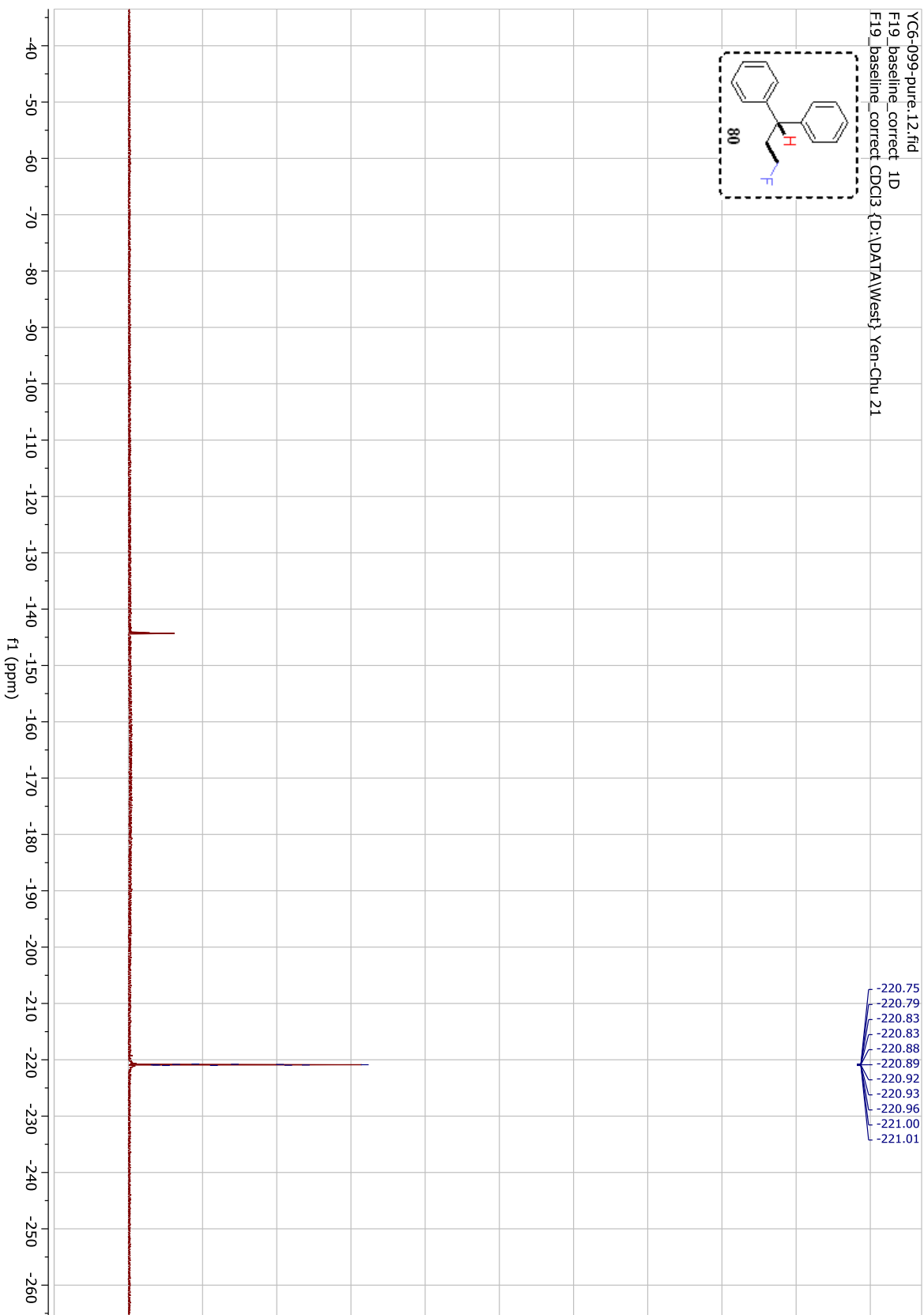
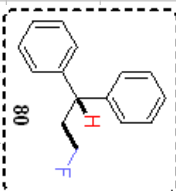


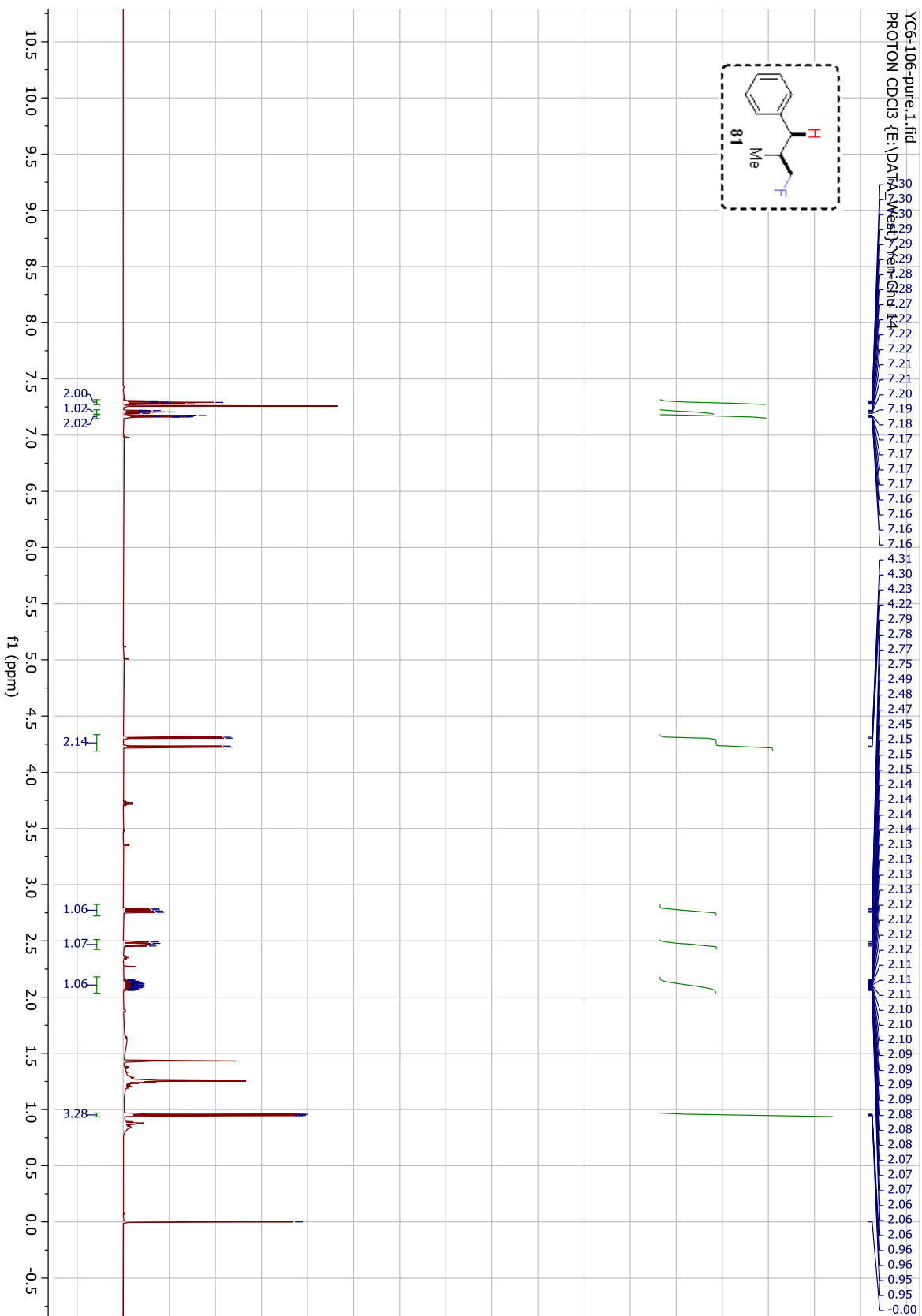


YC6-099-pure1.11.fid
C13CPD32 CDCl3 {D:\DATA\Westfj-Yen-Chu 21

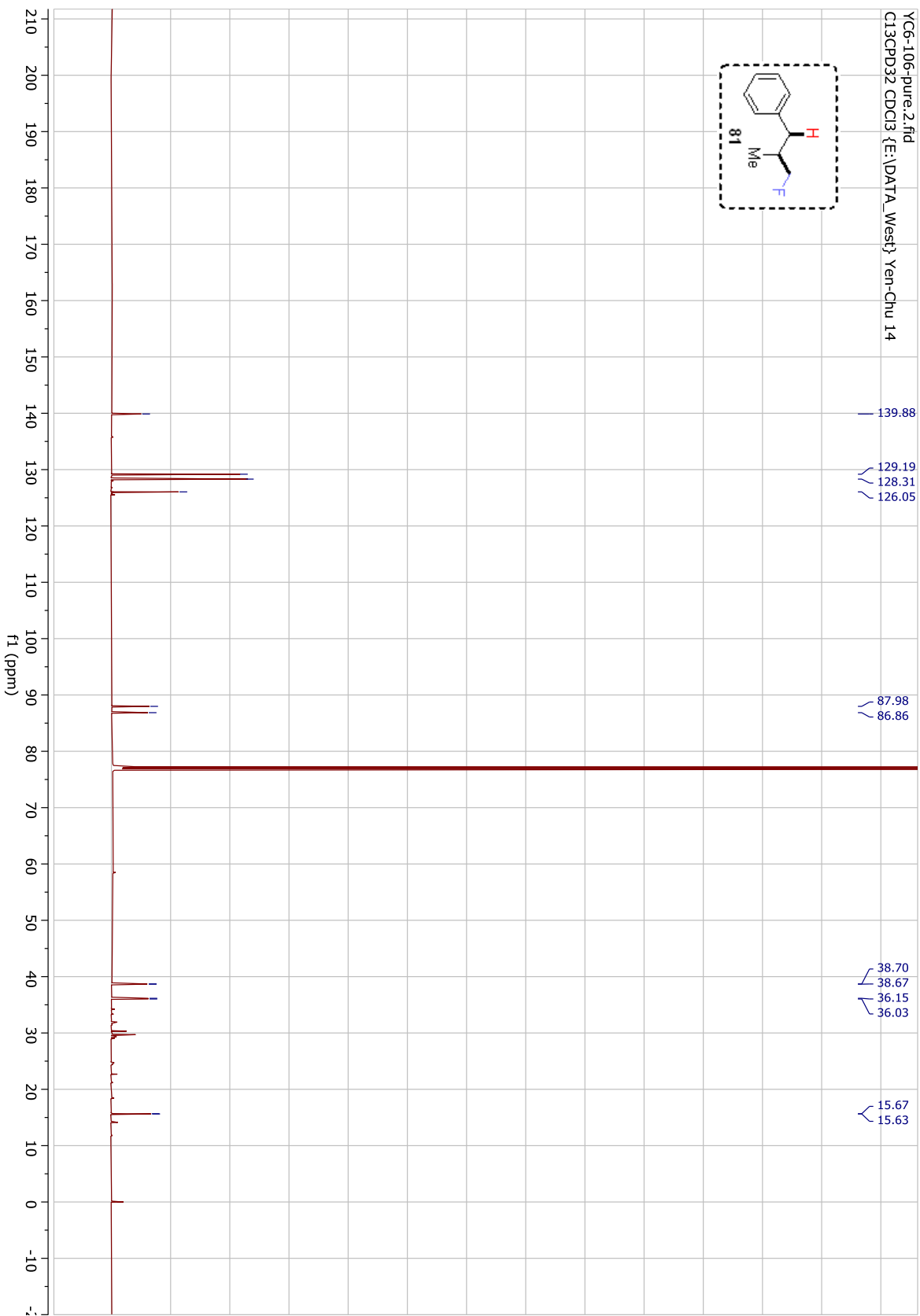
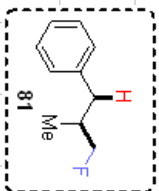


YC6-099-pure.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Yen-Chu 21

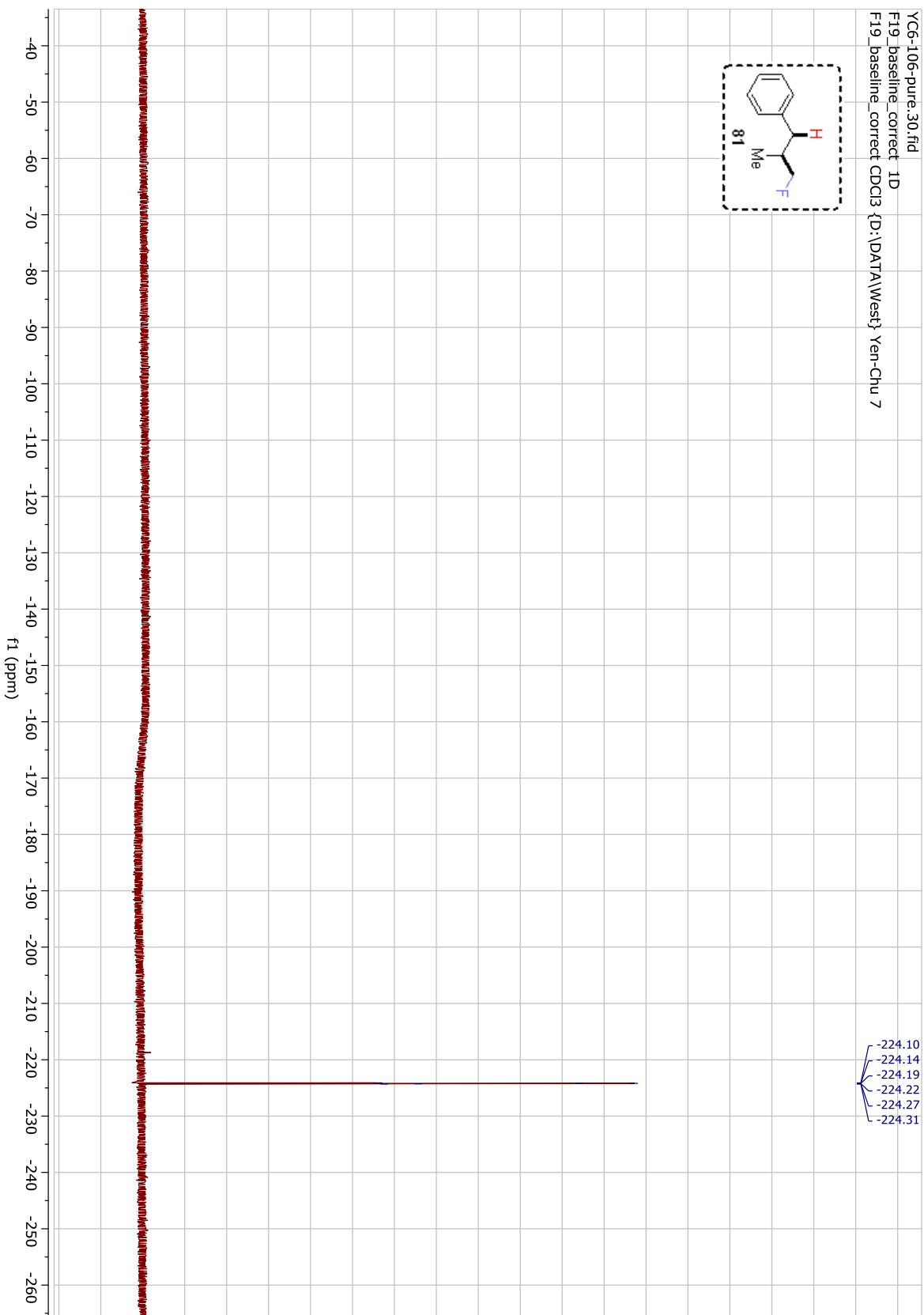
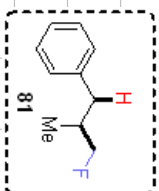


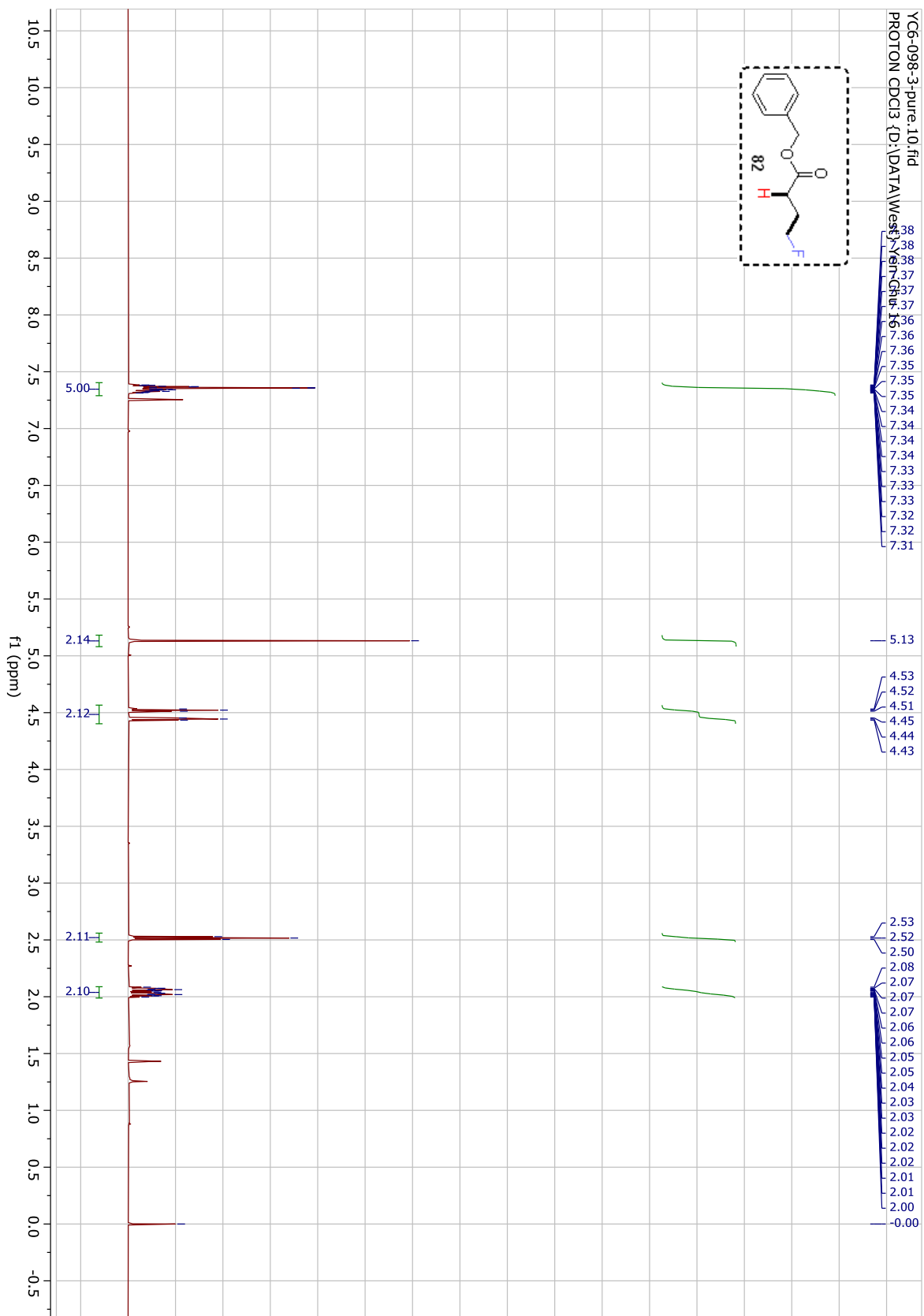


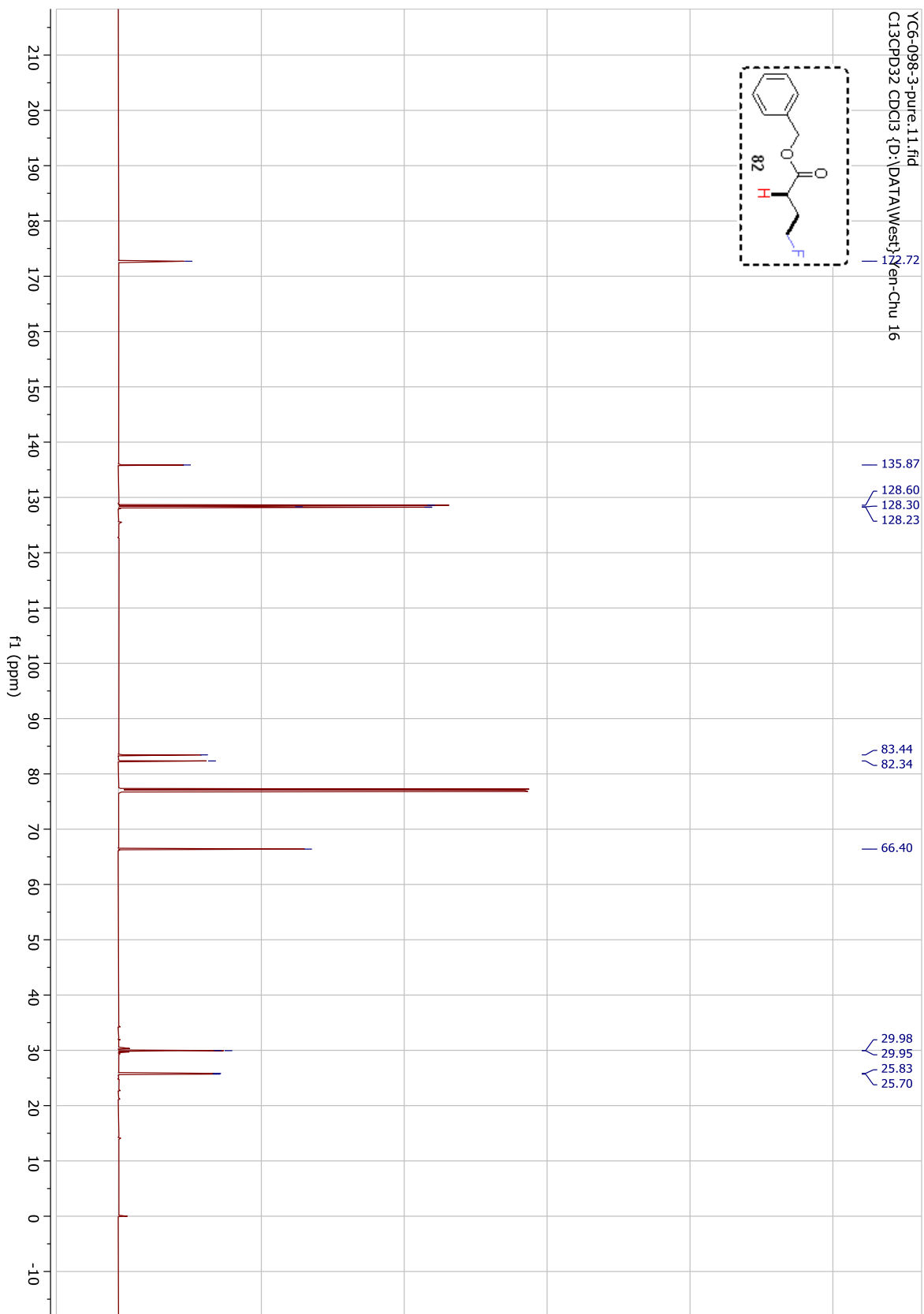
YC6-106-pure.2.fid
C13CPD32 CDDQ3 {E:\DATA_Westf\Yen-Chu 14



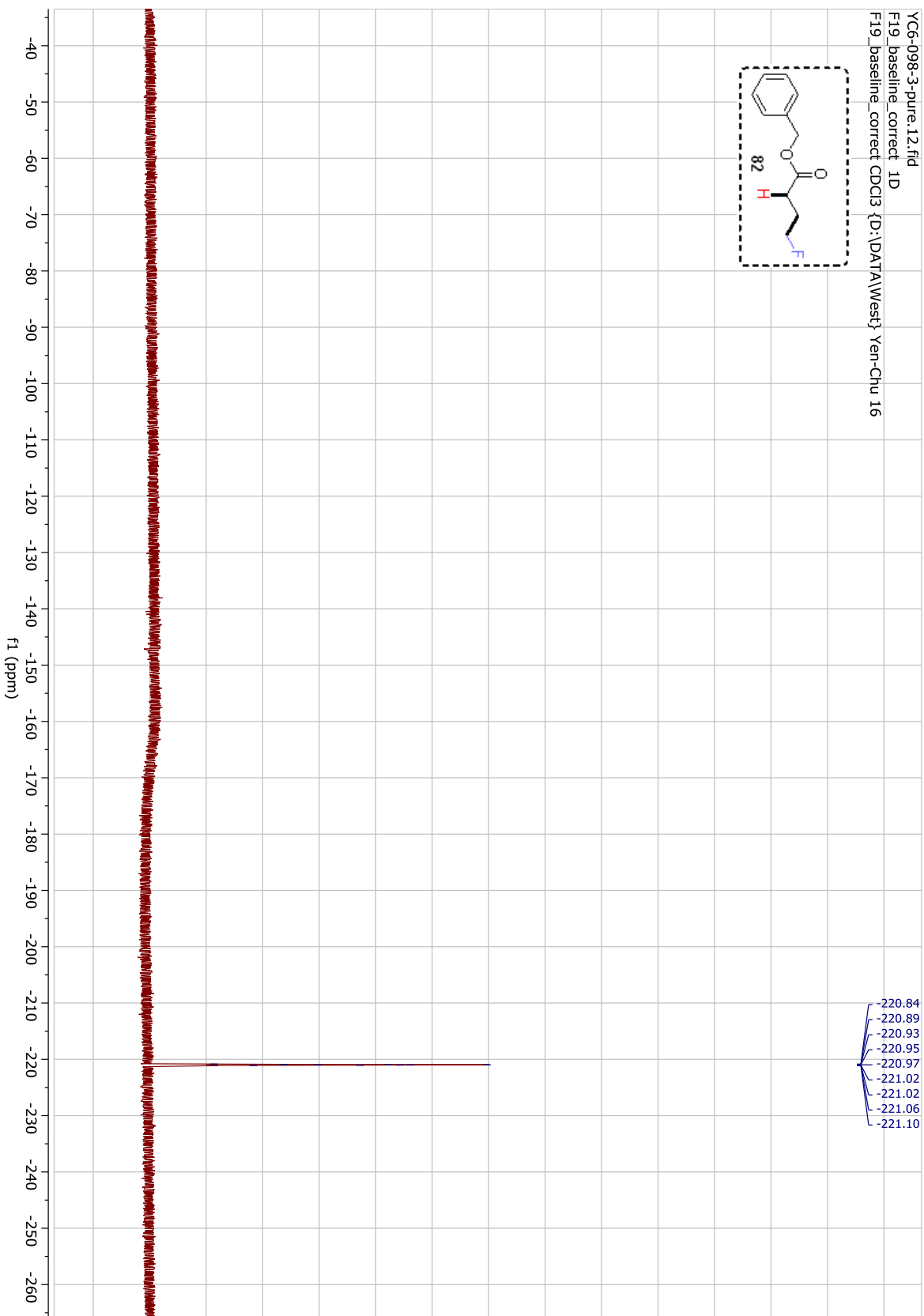
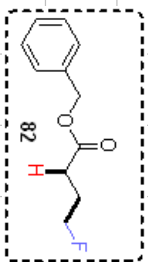
YC6-106-pure.30.fid
F19_baseline_correct_ID
F19_baseline_correct CDC13 {D:\DATA\West} Yen-Chu 7

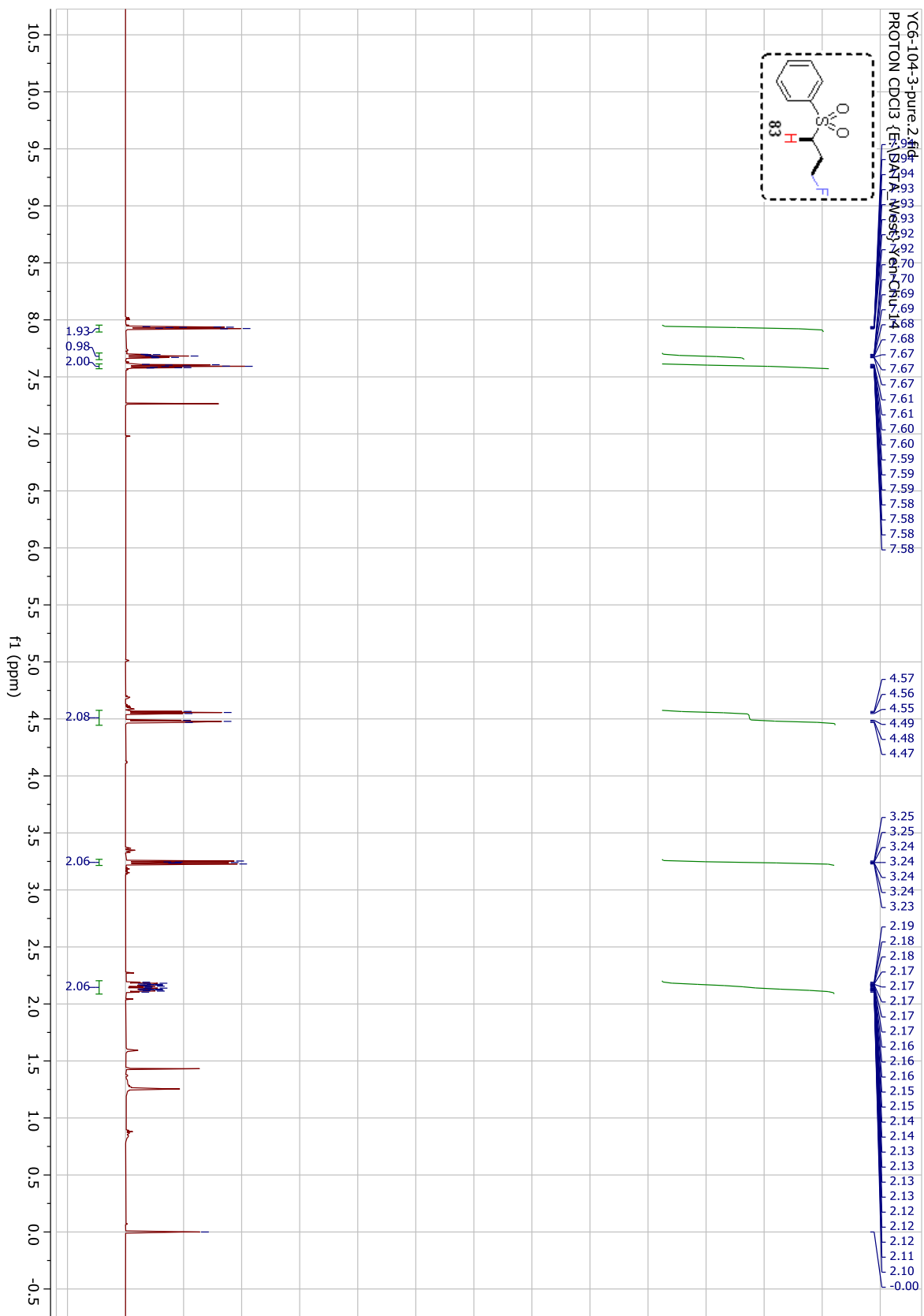




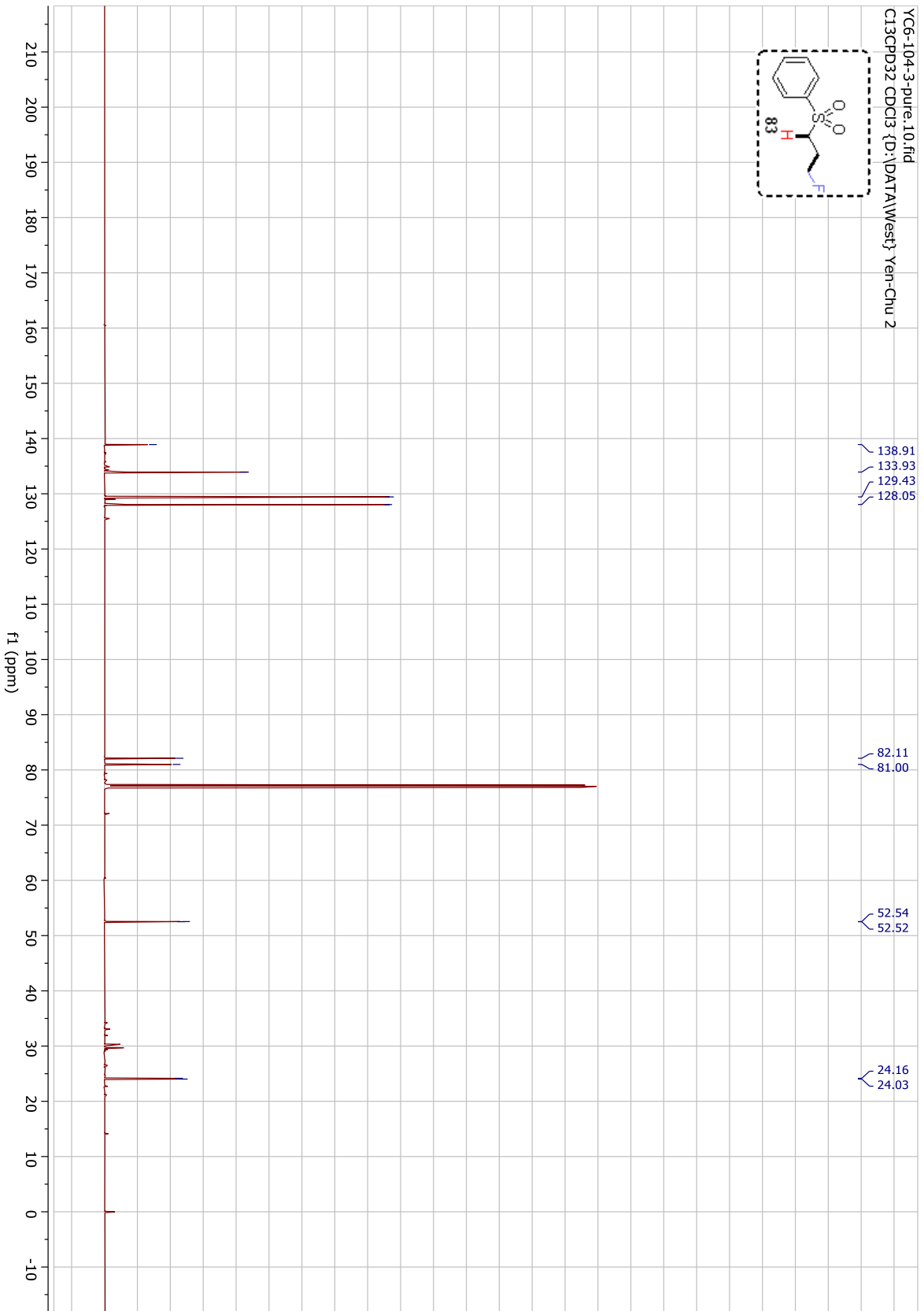
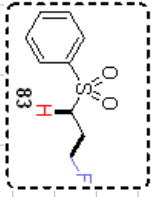


YC6-098-3-pure.12.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West\} Yen-Chu 16

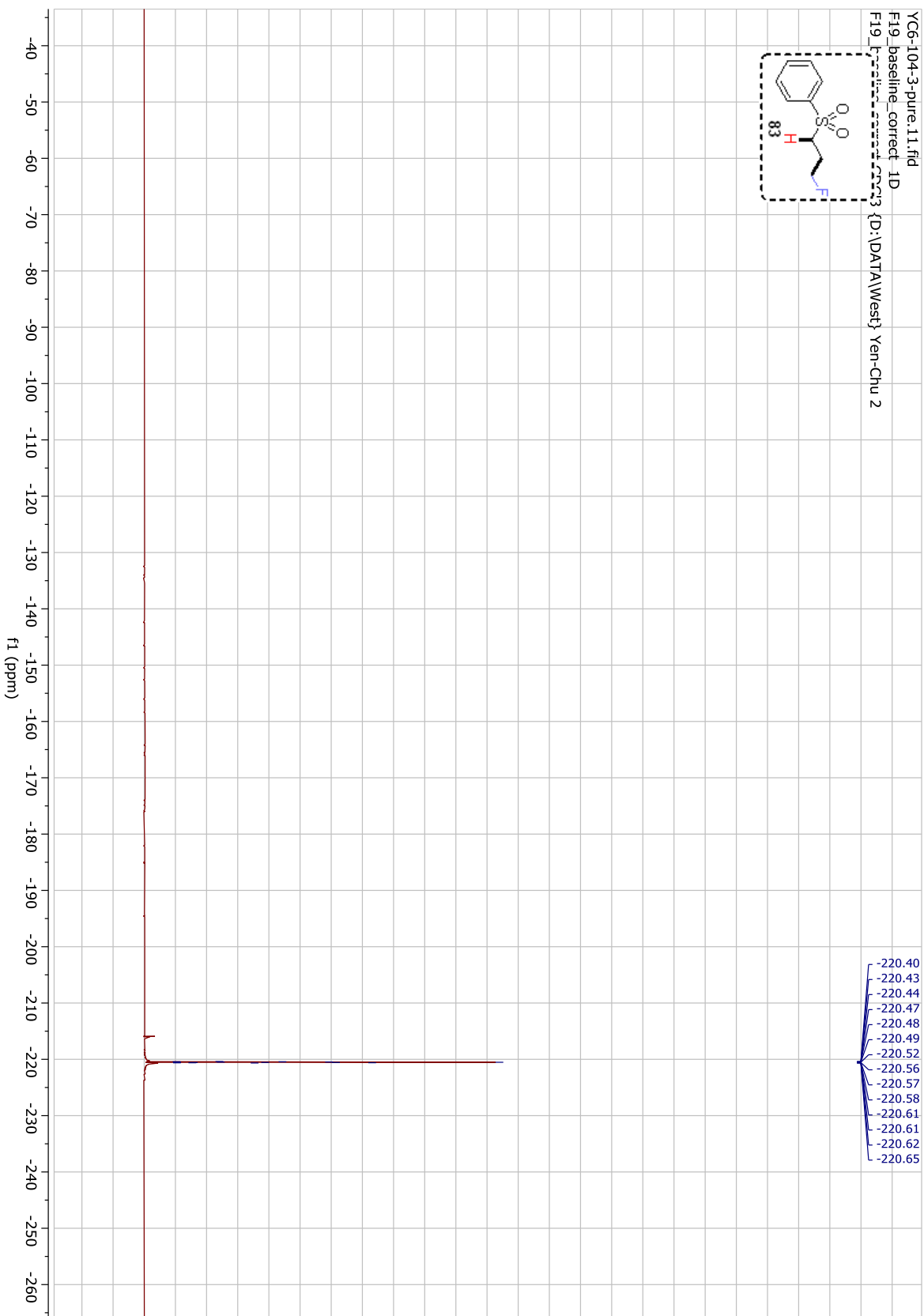
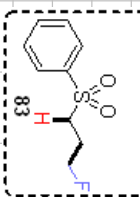


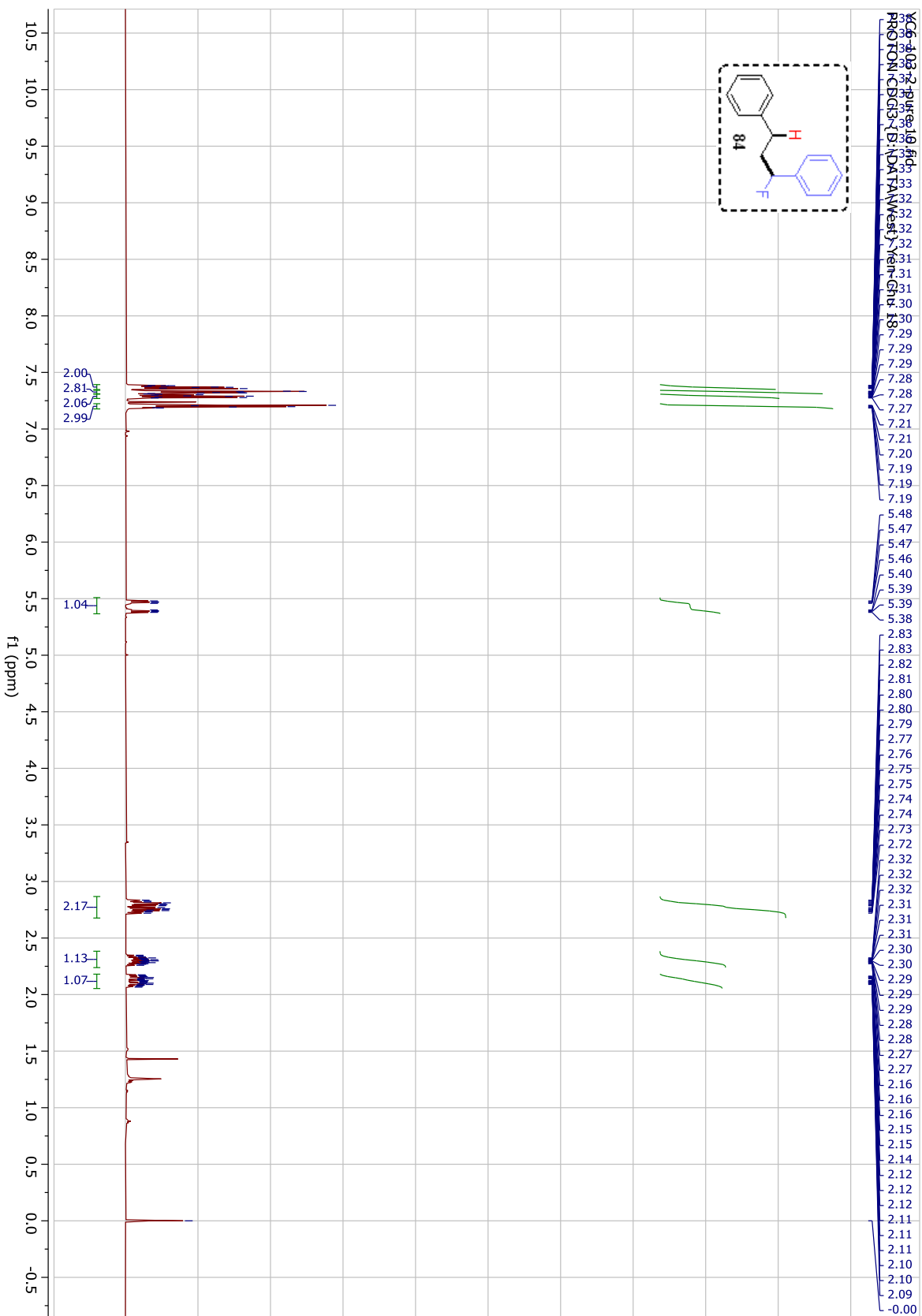


YC6-104-3-pure.10.fid
C13CPD32 CDCl3 {D:\DATA\Westf}\Yen-Chu 2

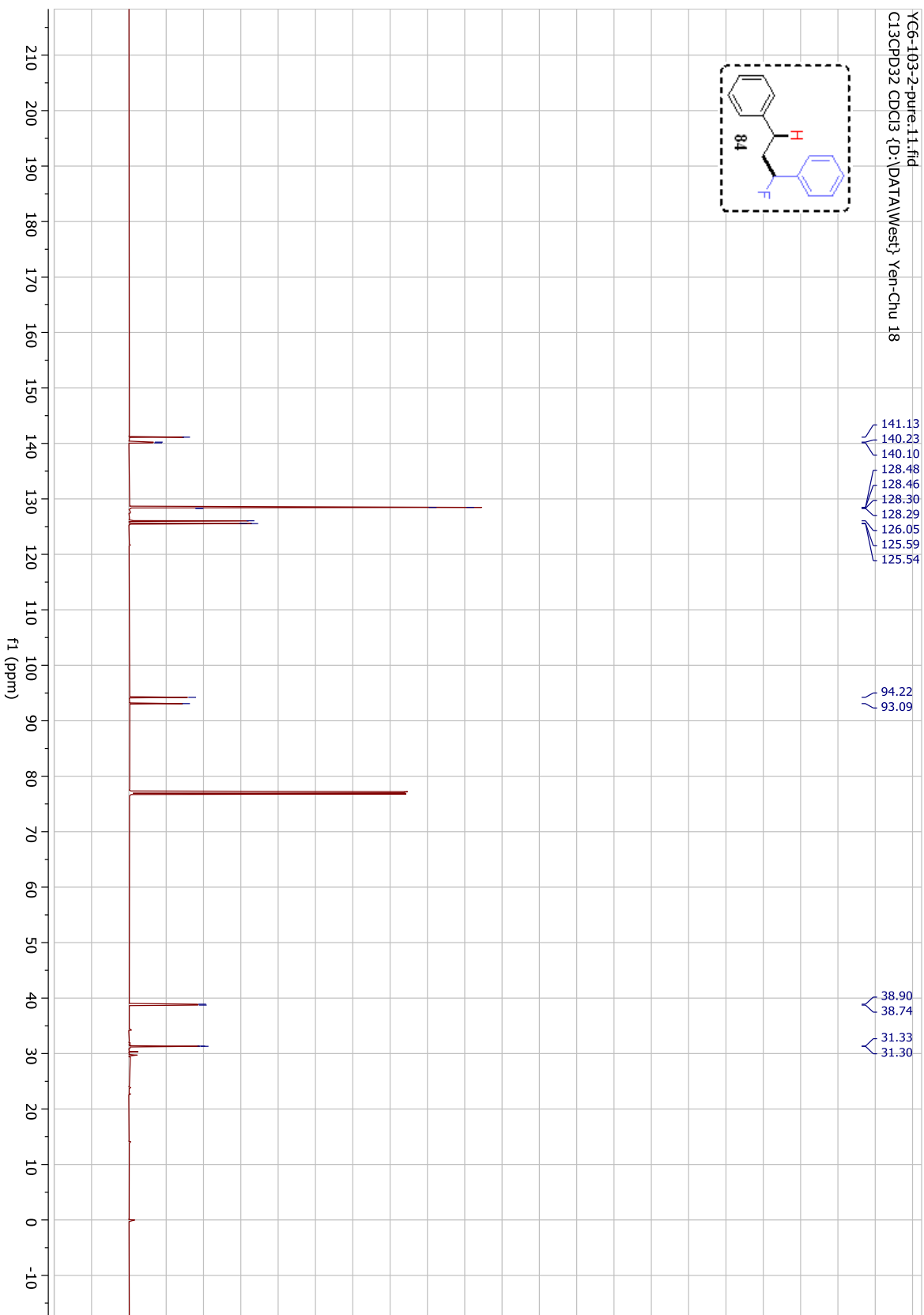
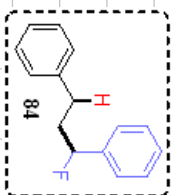


YC6-104-3-pure.11.fid
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F19_17333 {D:\DATA\West} Yen-Chu 2

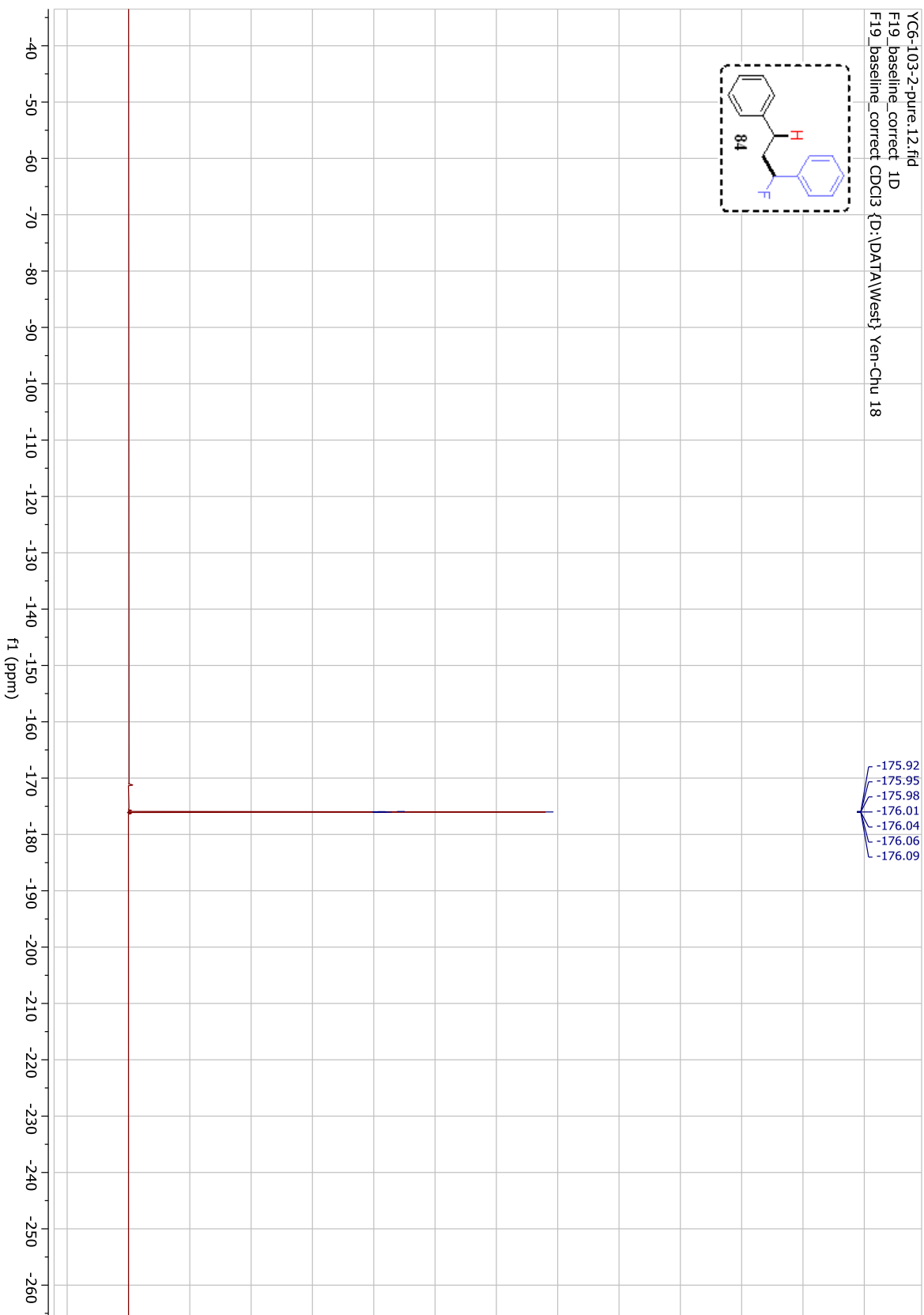
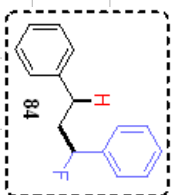


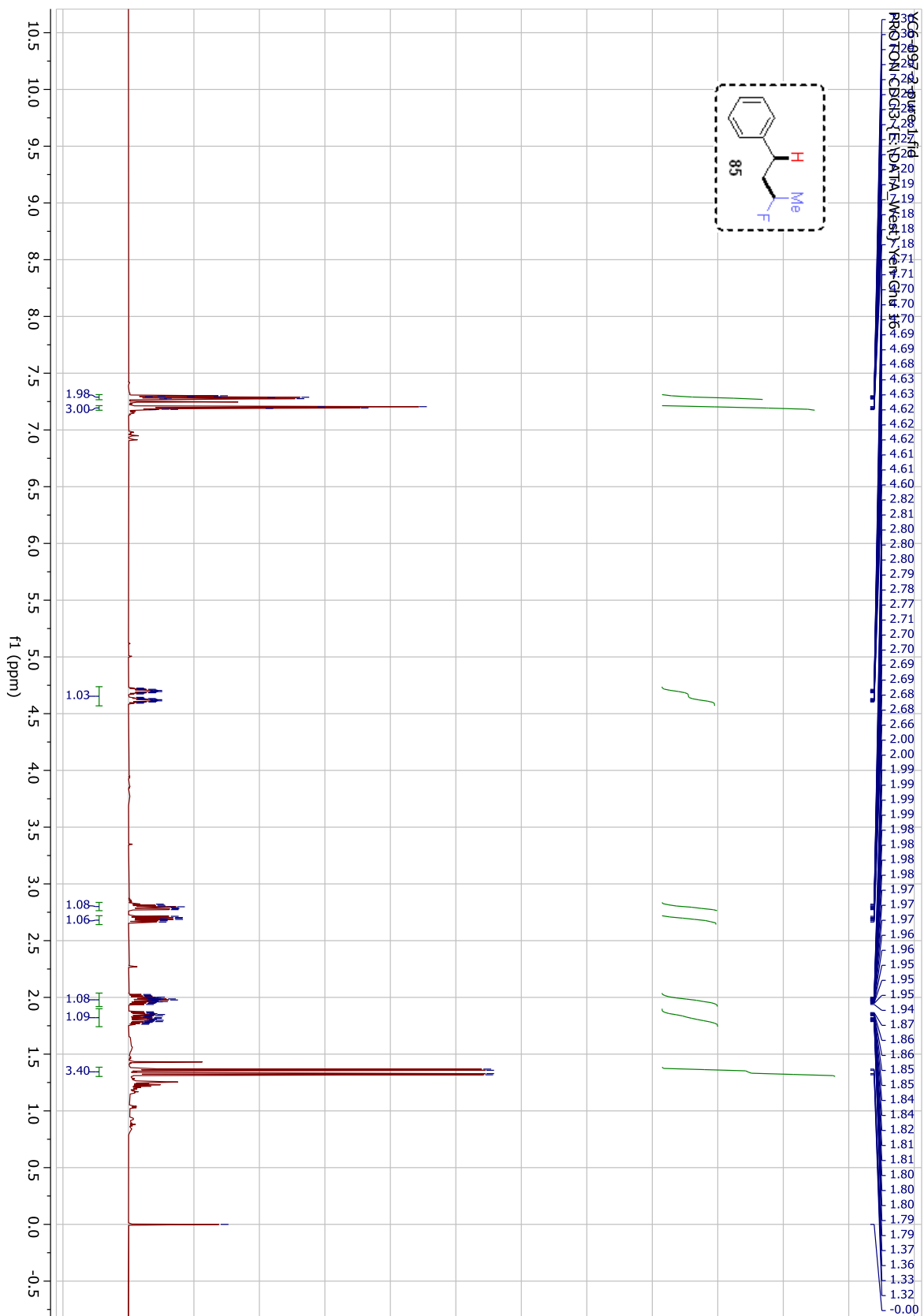


YC6-103-2-pure.11.fid
C13CPD32 CDCl3 {D:\DATA\West}\Yen-Chu 18

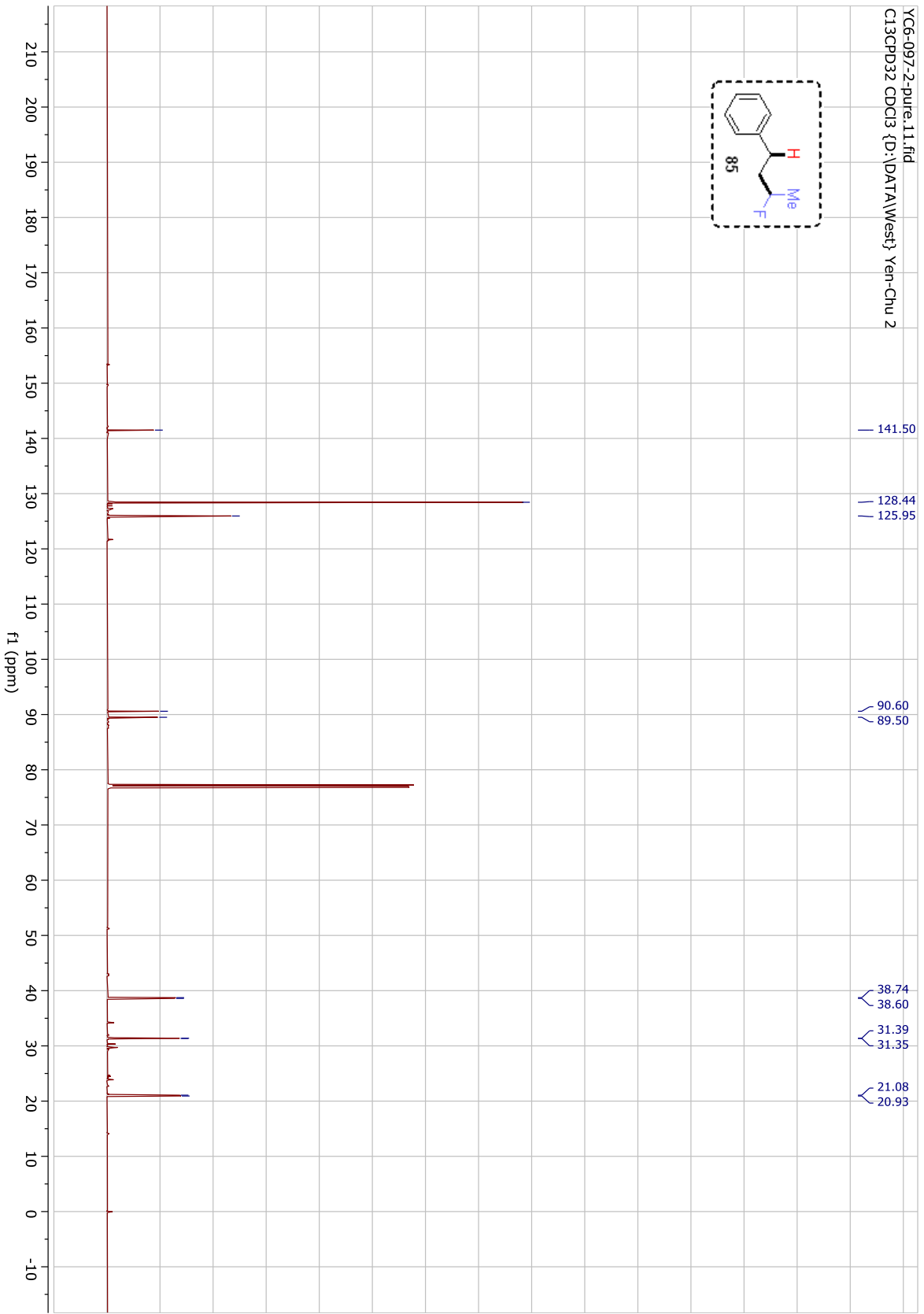
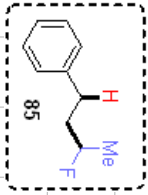


YC6-103-2-pure.12.ftd
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Yen-Chu 18

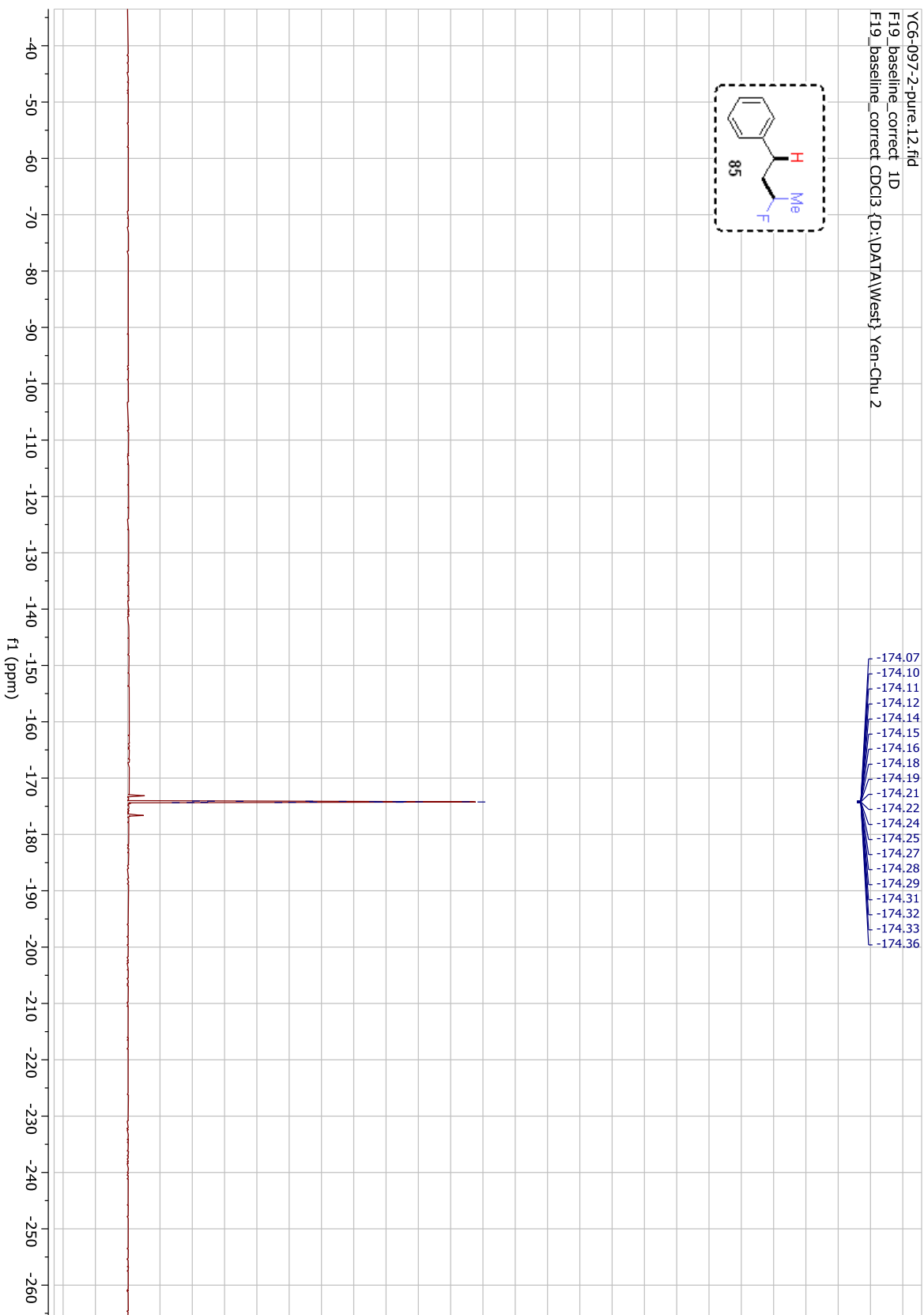
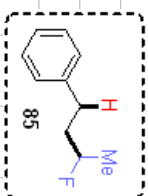


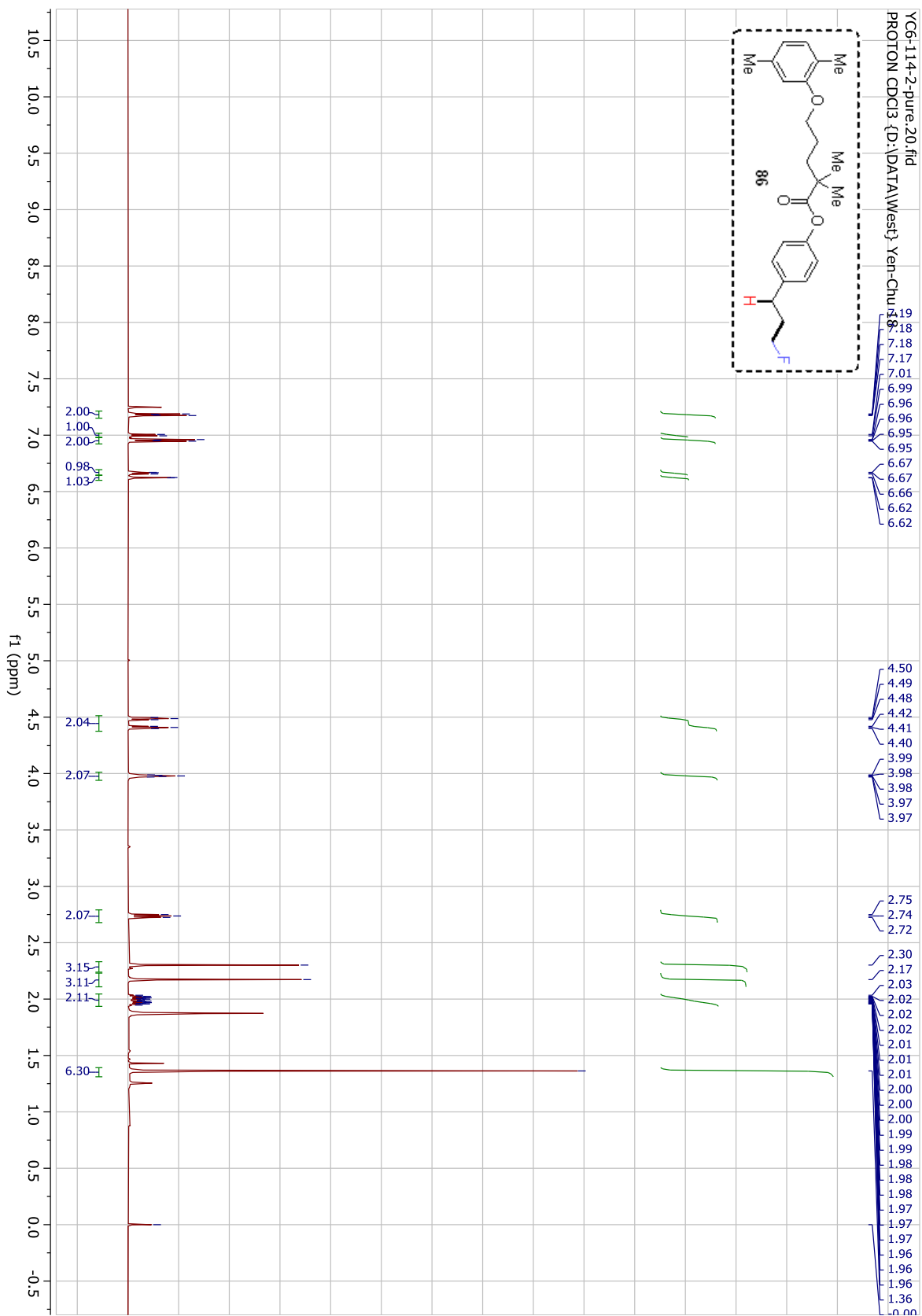


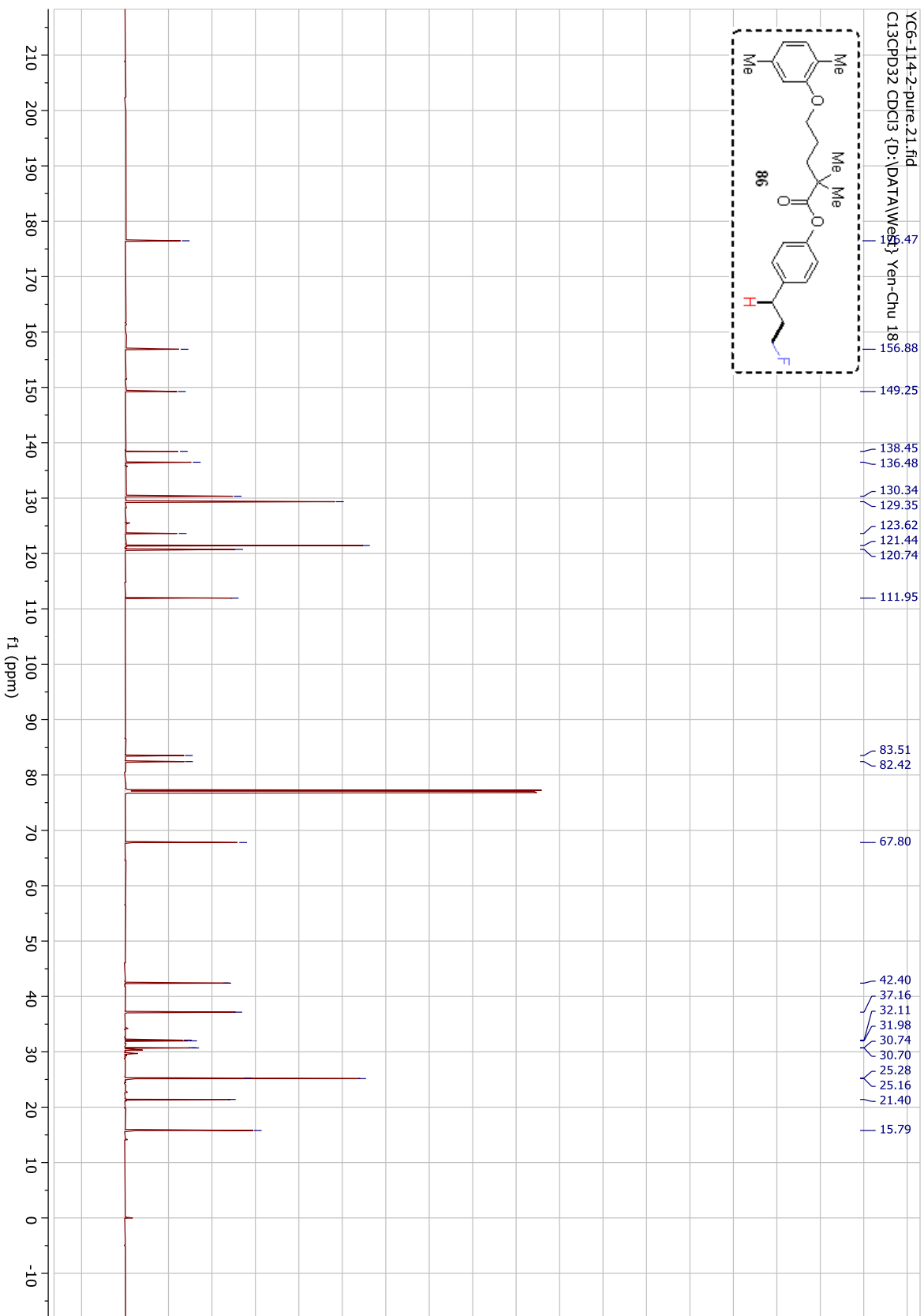
YC6-097-2-pure.11.fid
C13CPD32 CDCl3 {D:\DATA\West}\Yen-Chu 2



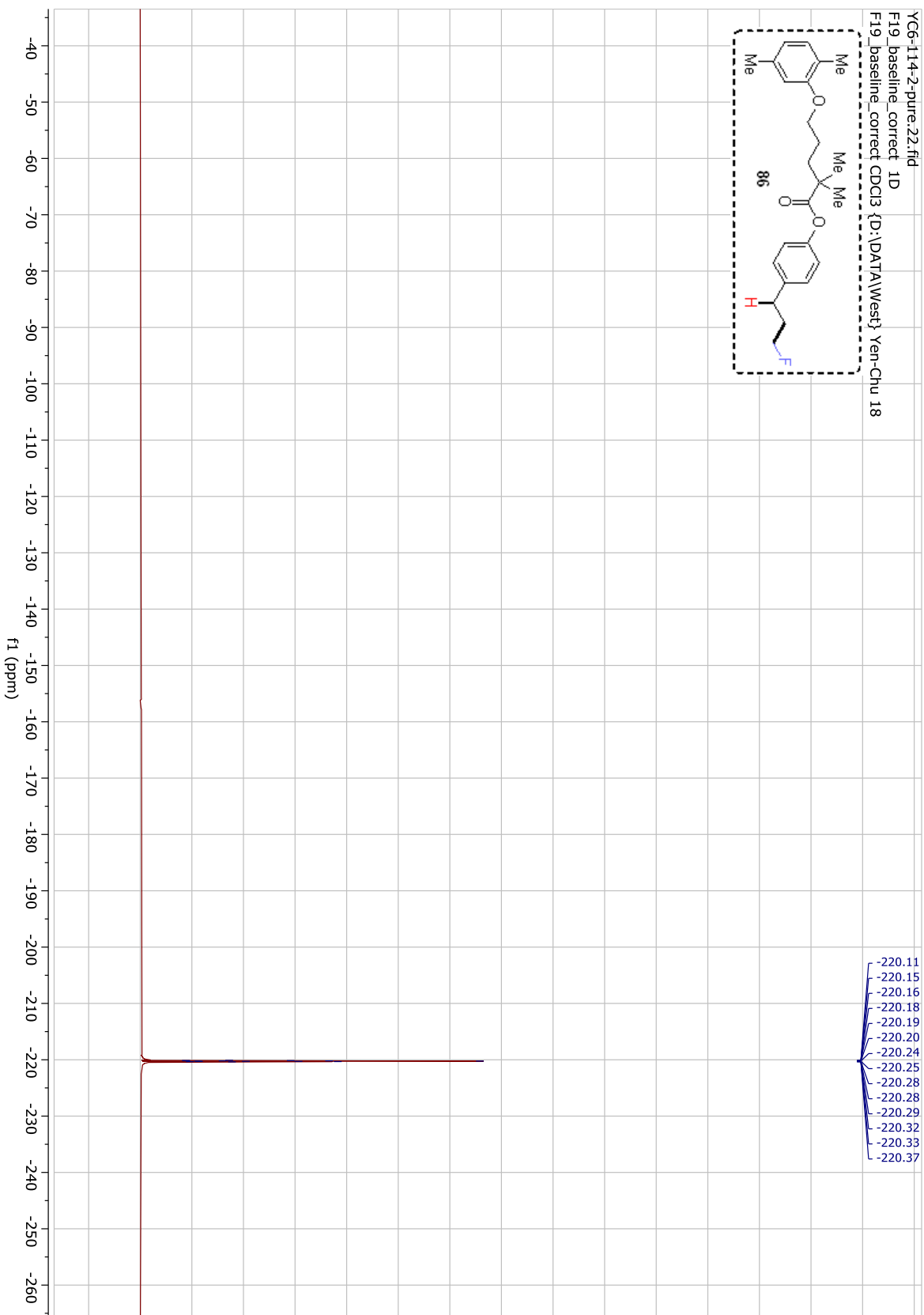
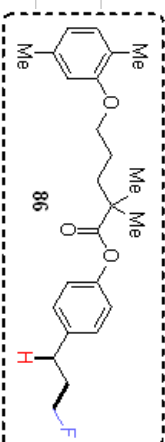
YC6-097-2-pure.12.fid
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West\ Yen-Chu 2

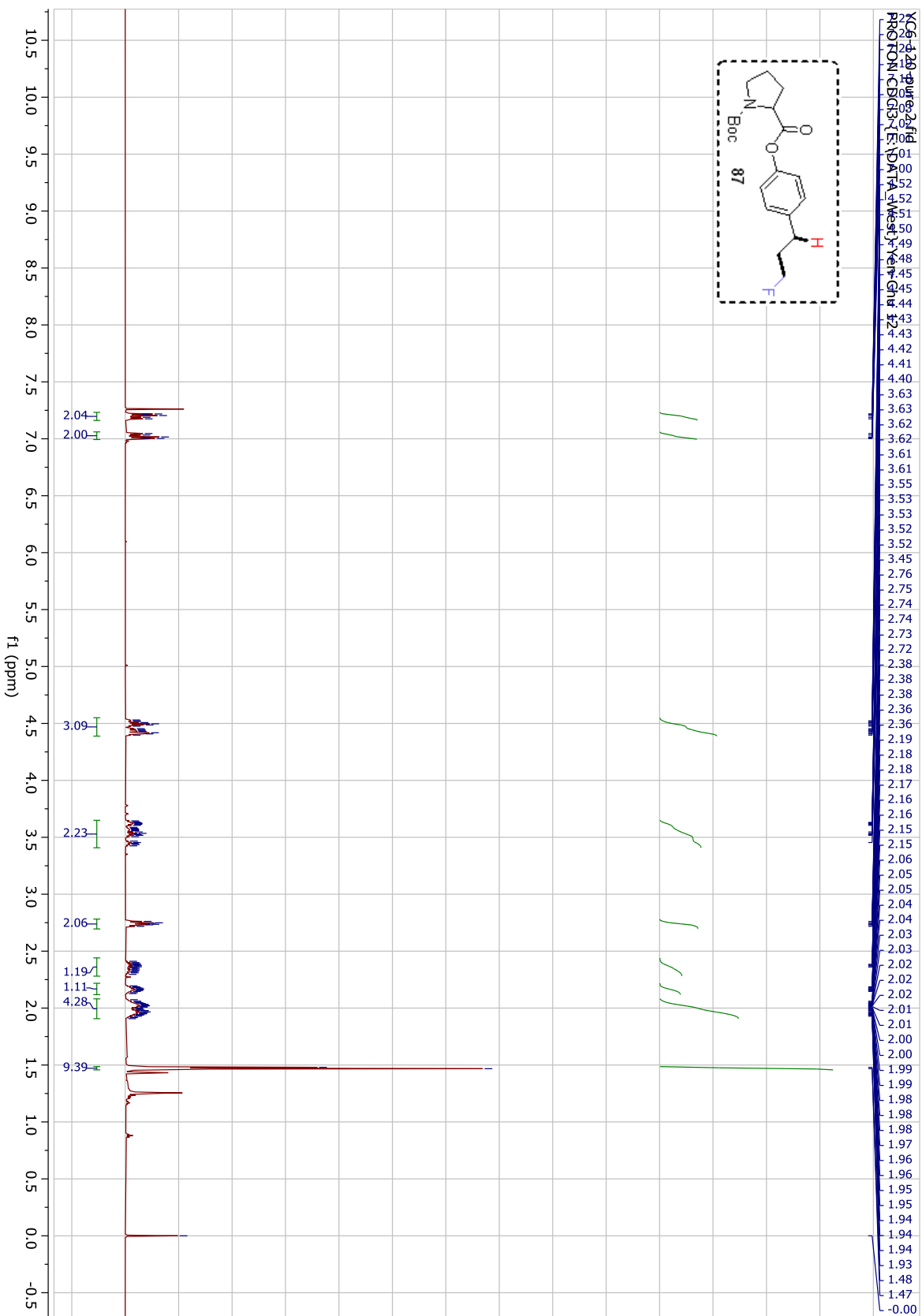


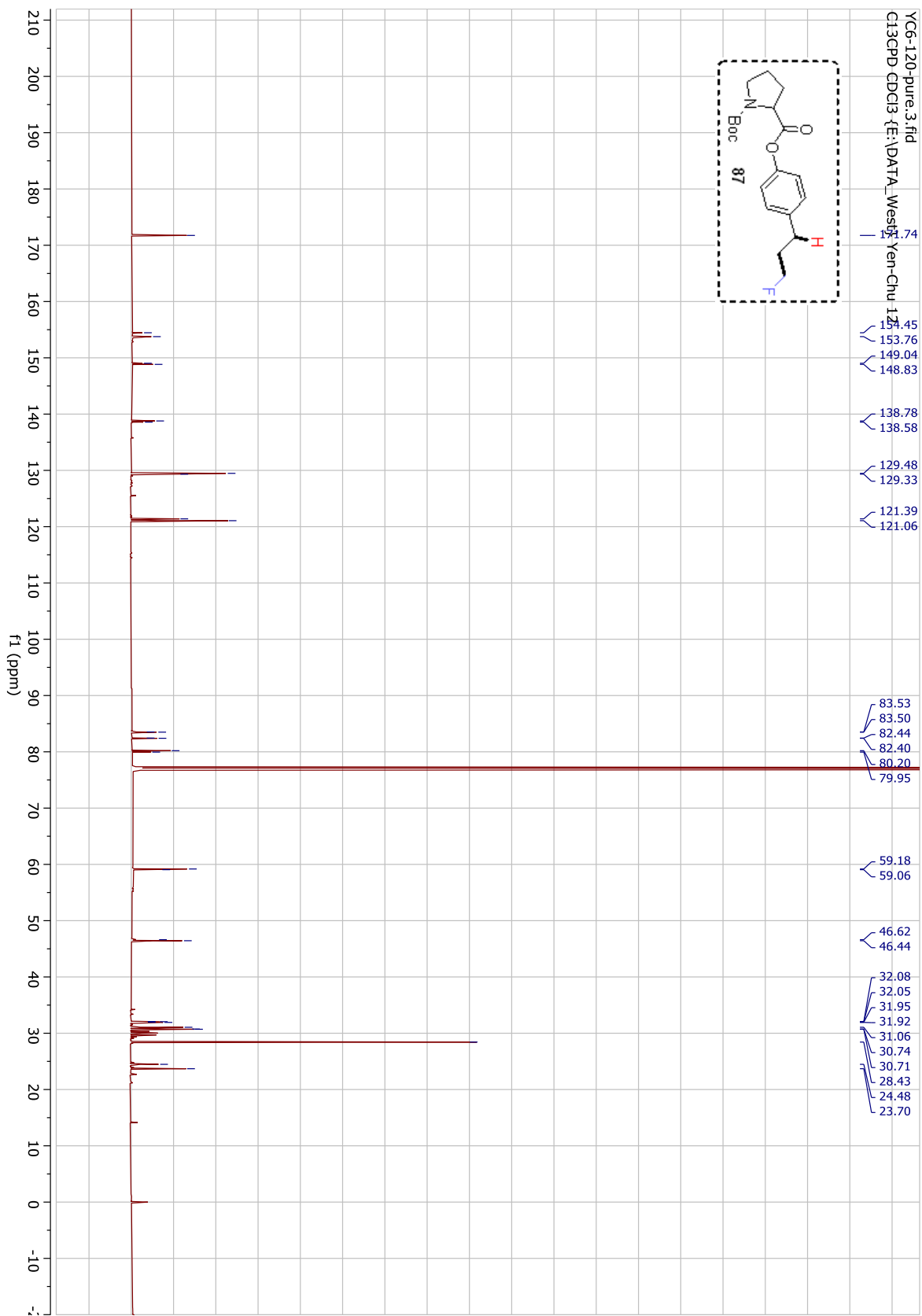




YC6-114-2-pure.22.ftd
F19_baseline_correct ID
F19_baseline_correct CDCl3 {D:\DATA\West} Yen-Chu 18

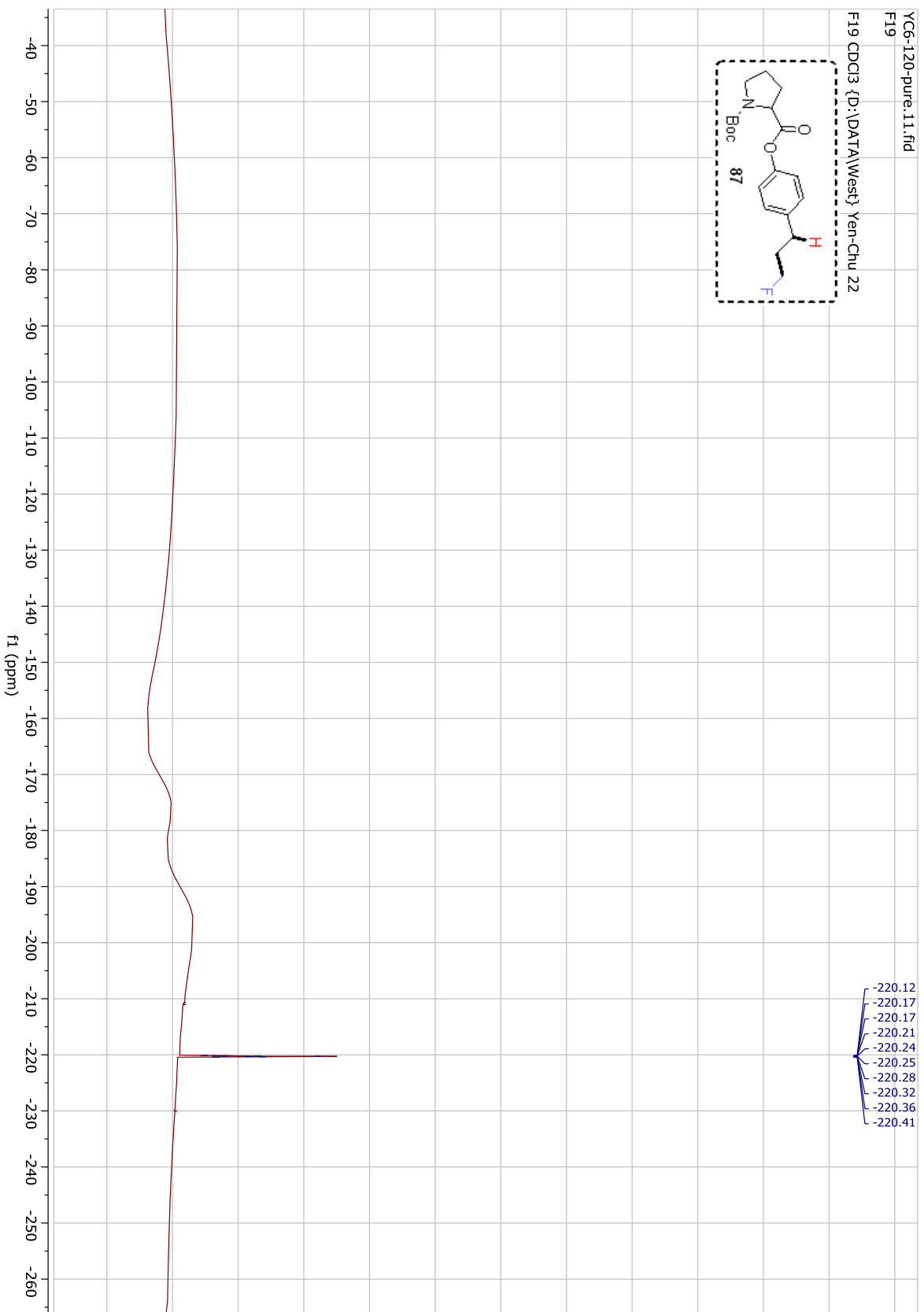
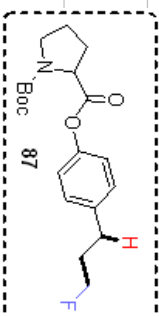


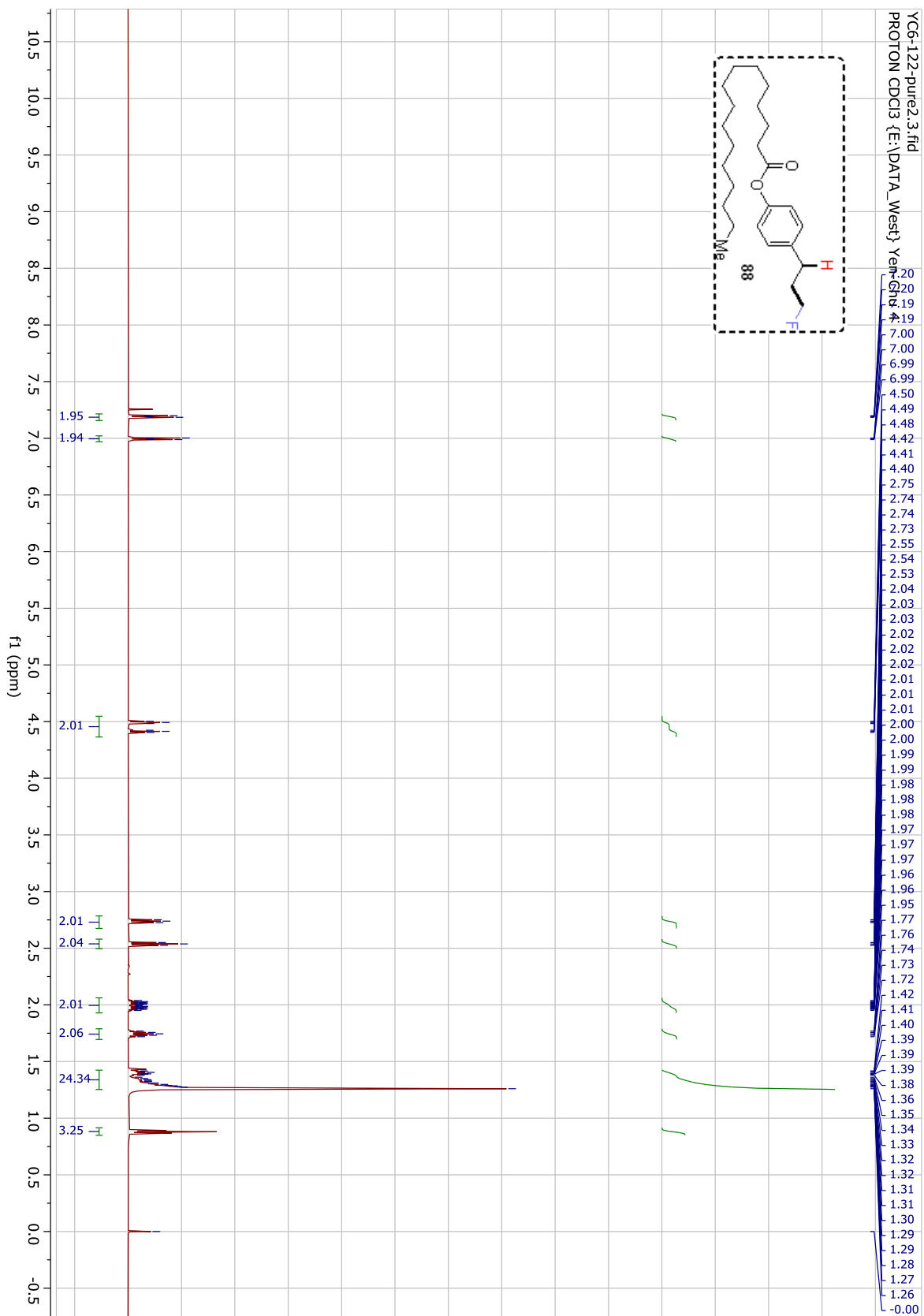




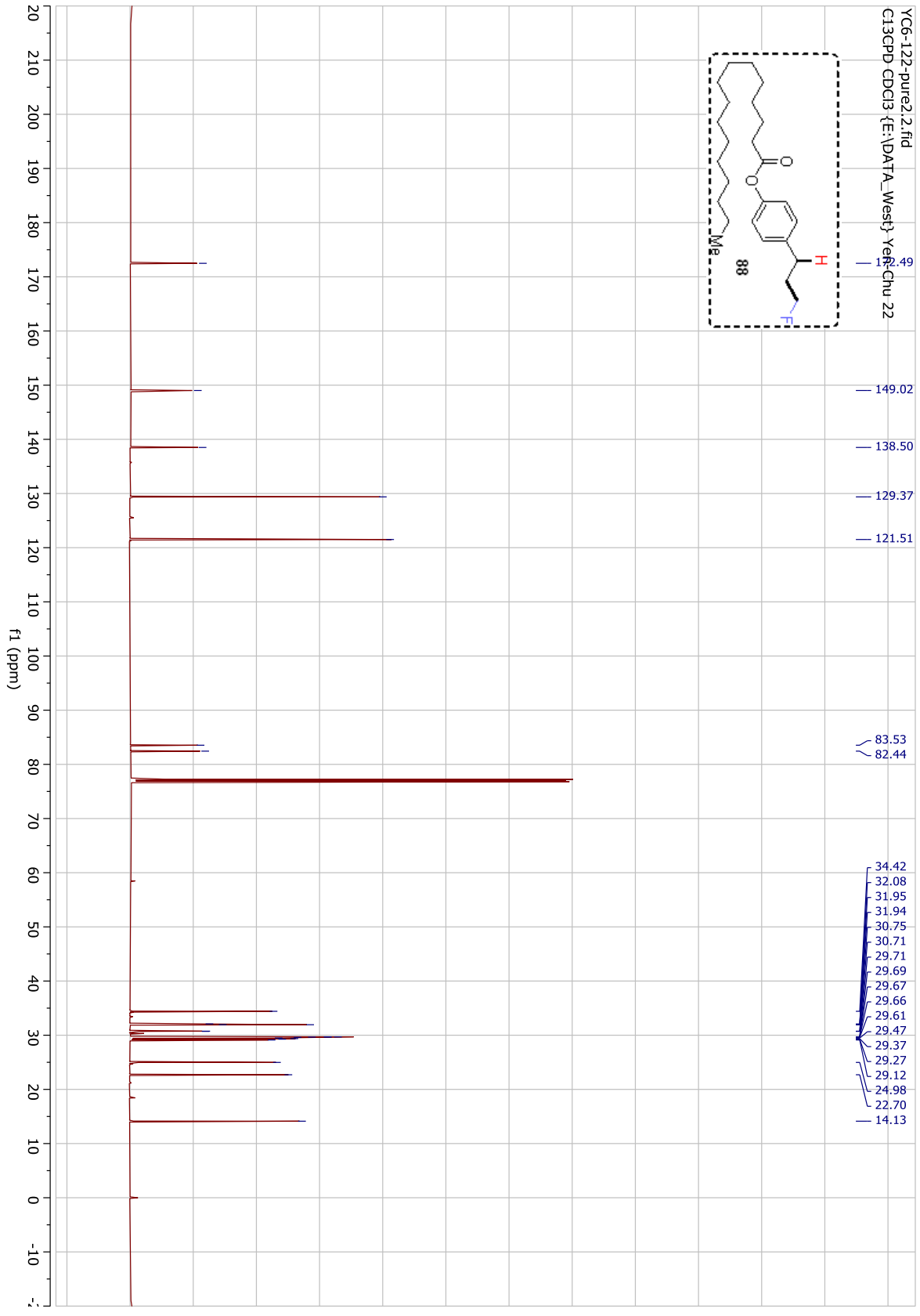
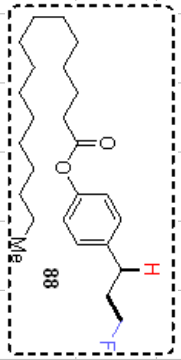
YC6-120-pure.11.fid
F19

F19 CDQ3 {D:\DATA\West\ Yen-Chu 22





YC6-122 -pure2,2.fid
C13CPD CDCl3 (E:\DATA_Westj\YeR\Chu-22

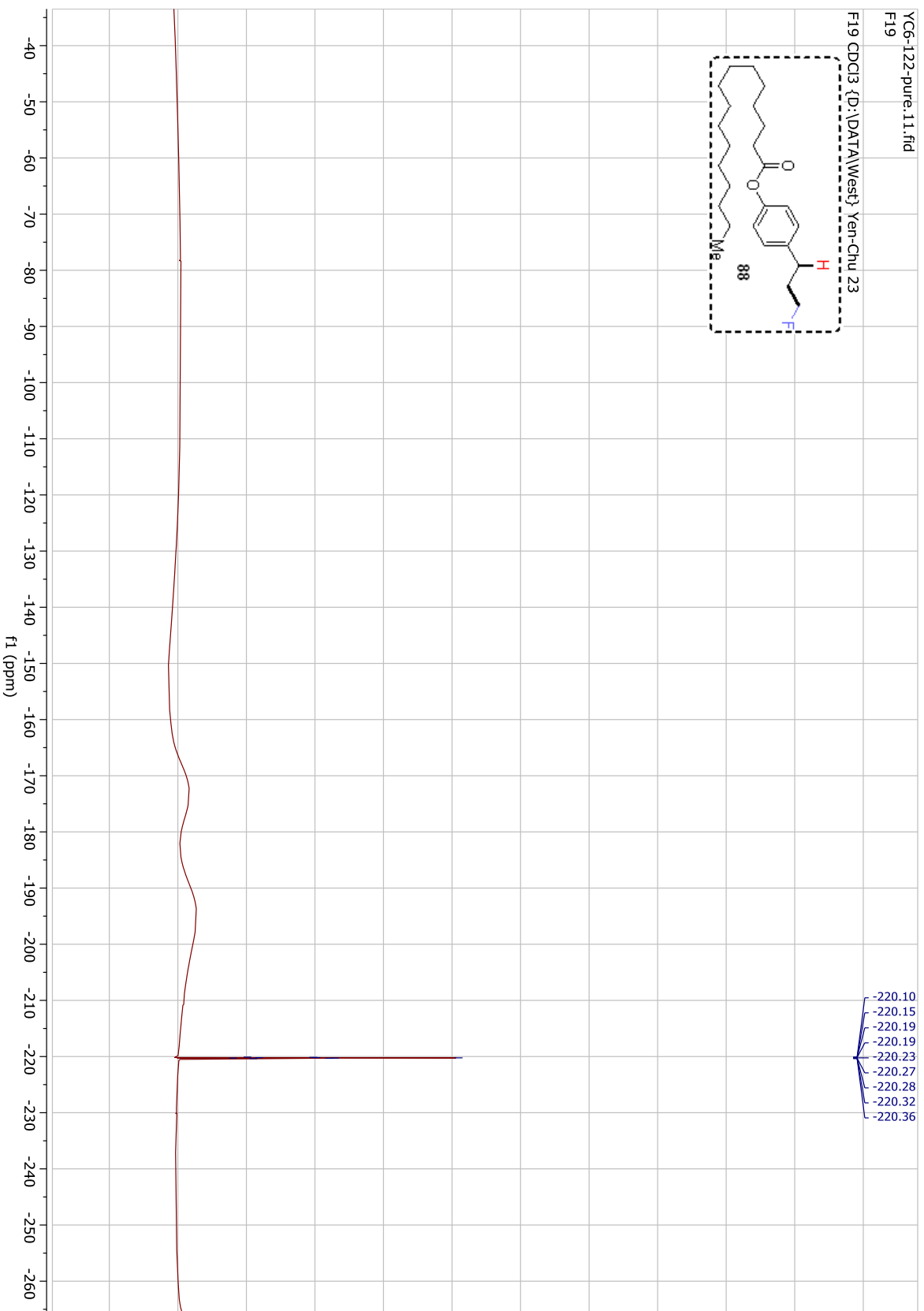
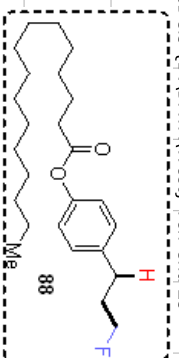


340

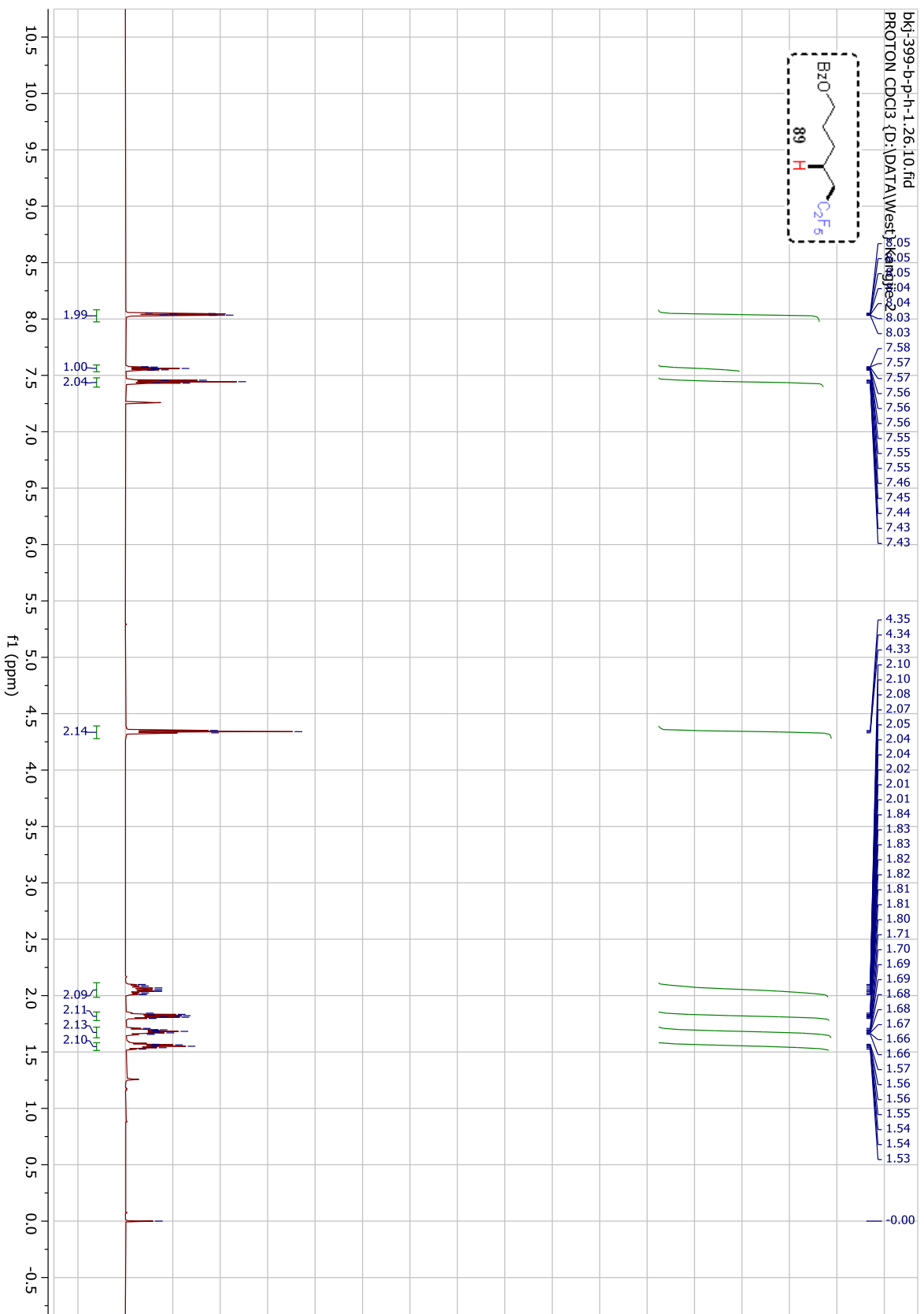
YC6-122-pure.11.fid

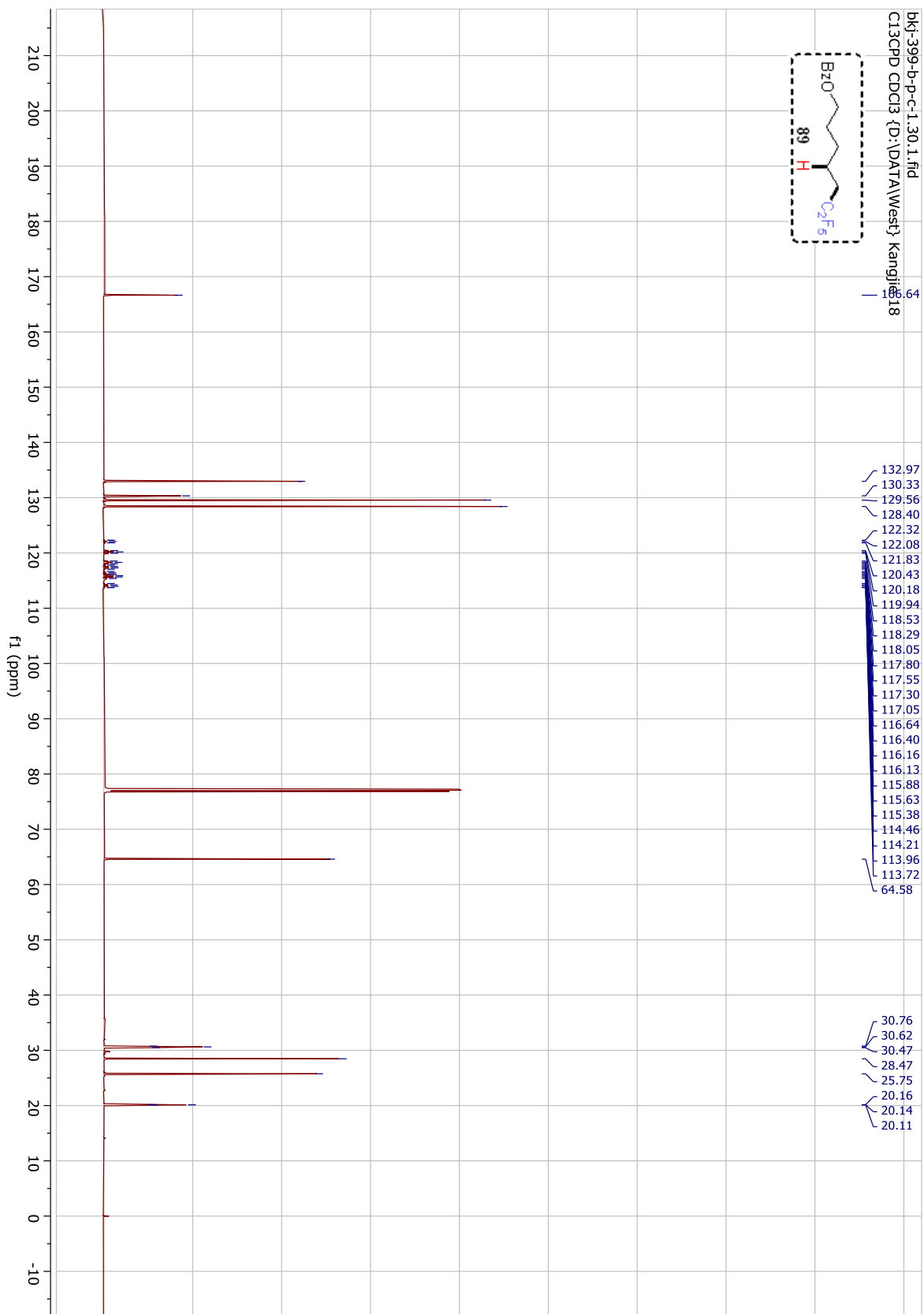
F19

F19 CDCl3 {D:\DATA\West\ Yen-Chu 23

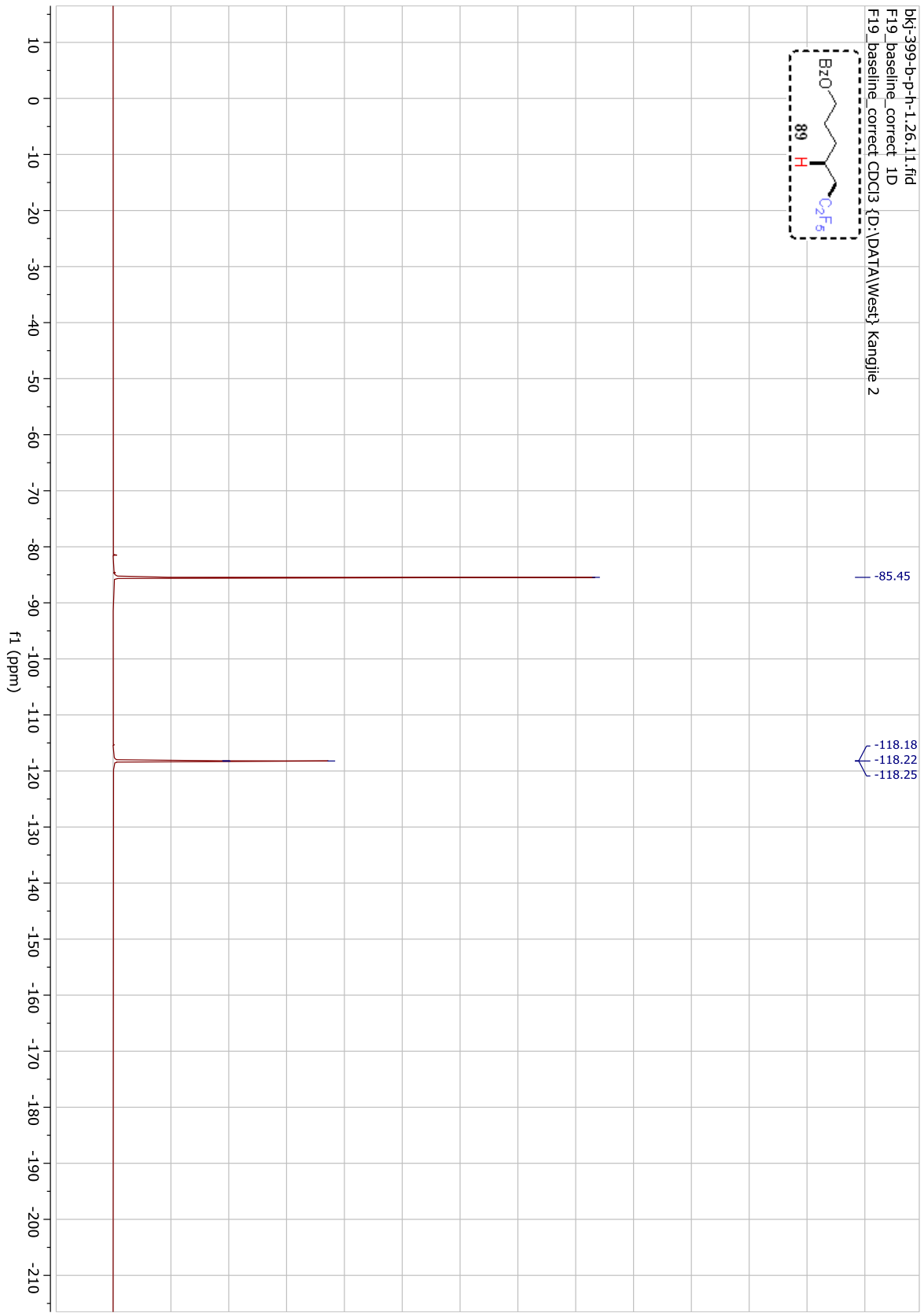
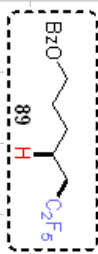


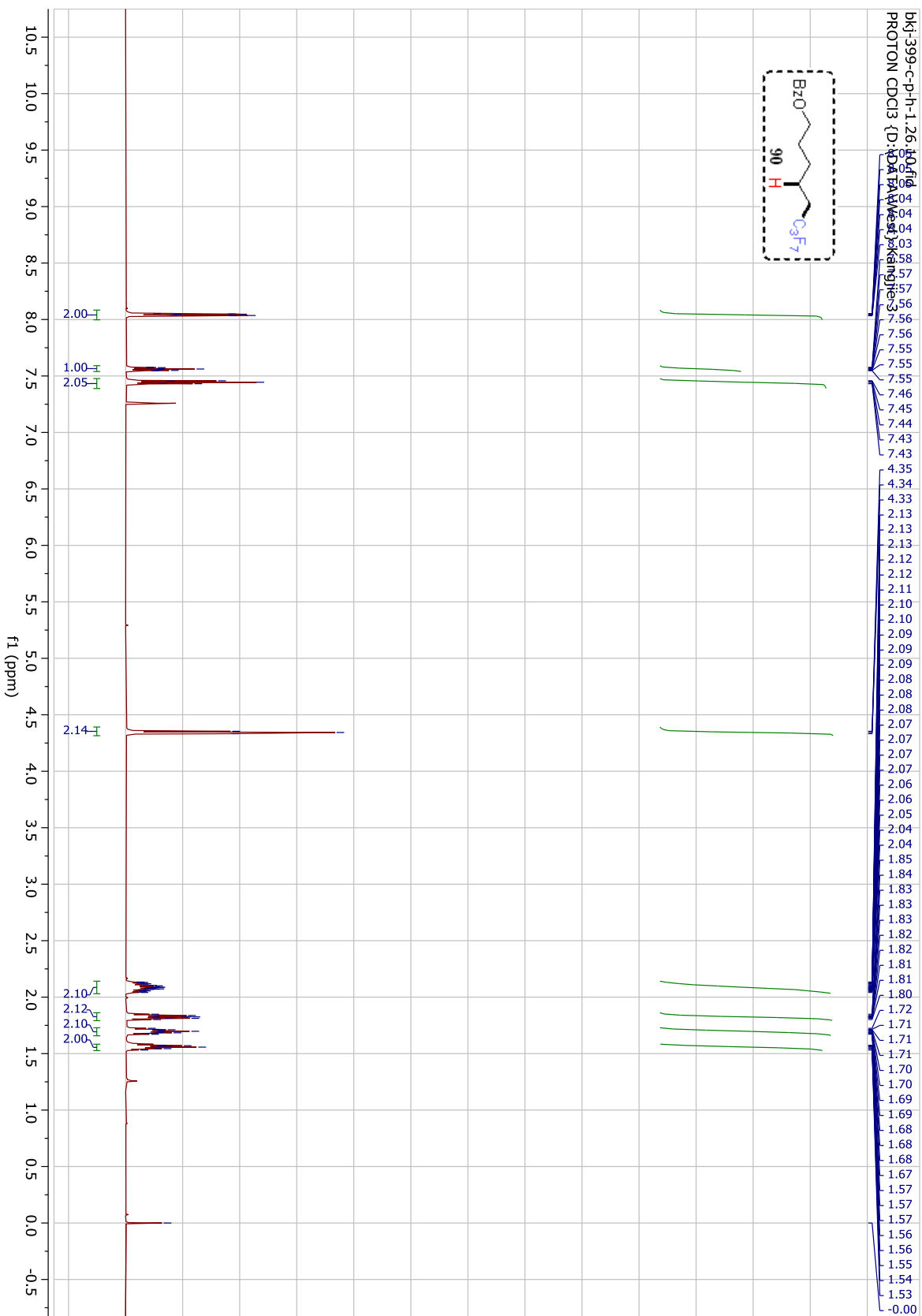
341



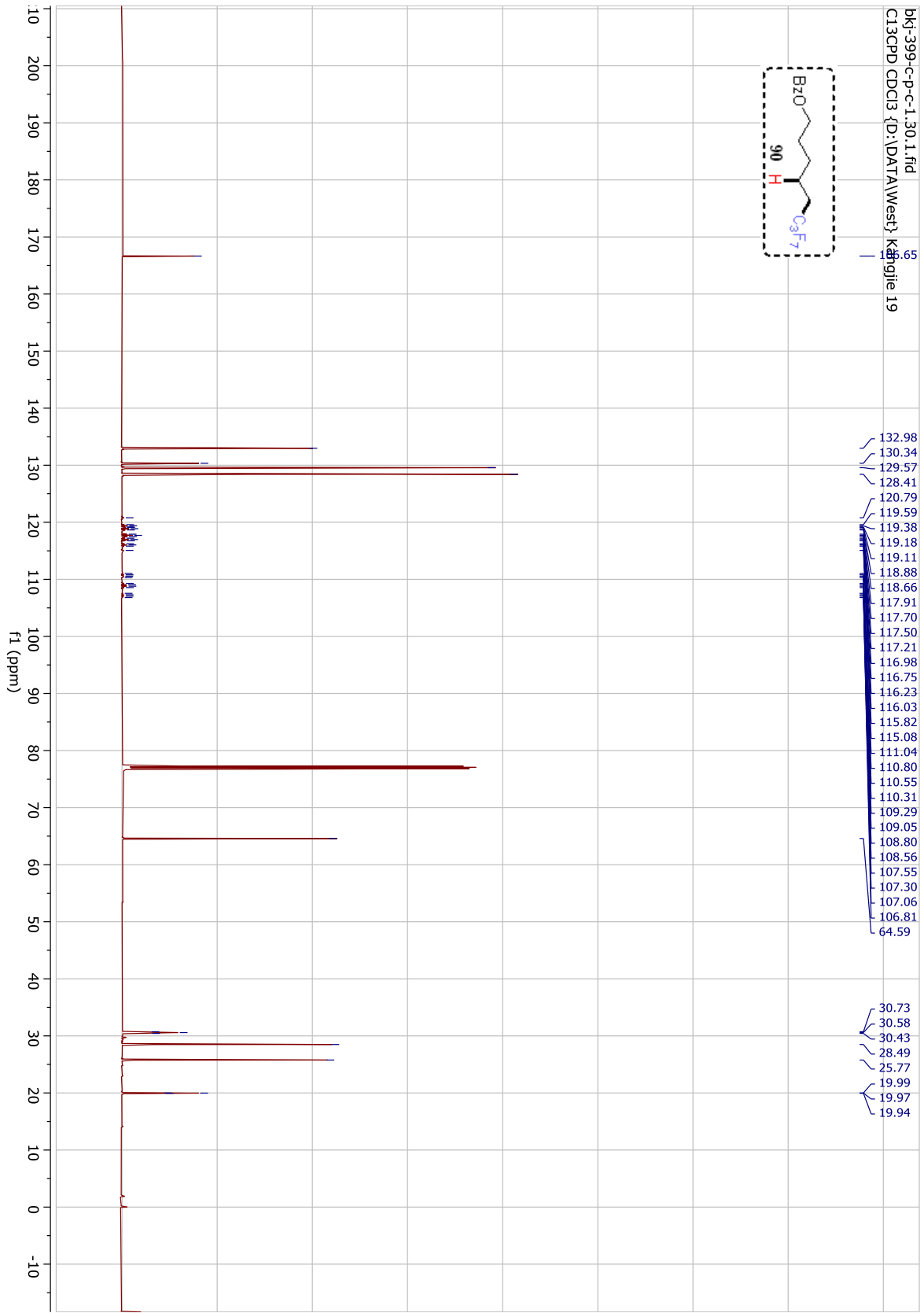
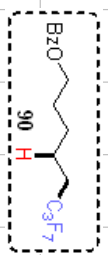


bkj-399-b-p-h-1.26.11.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 2

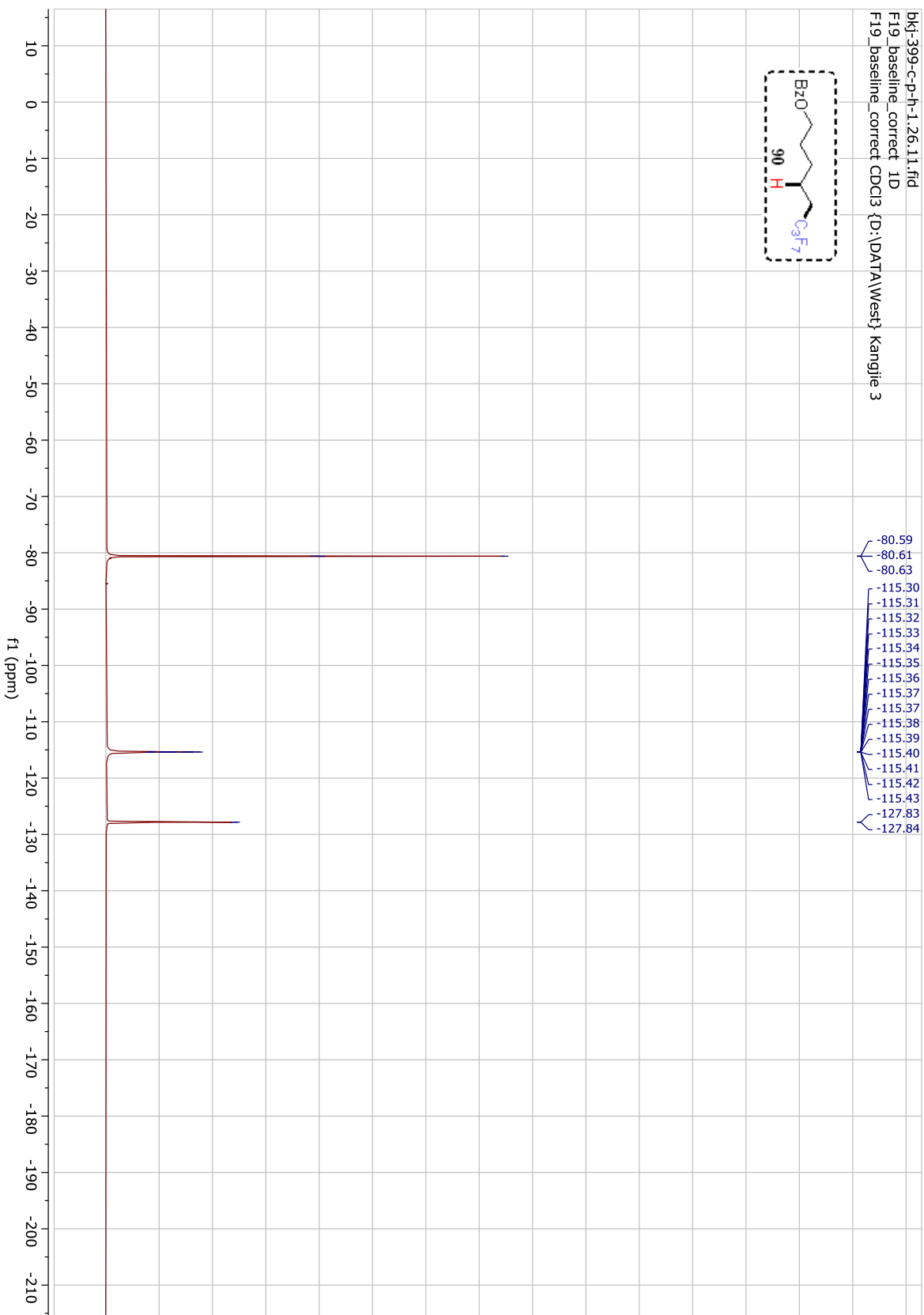
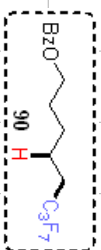


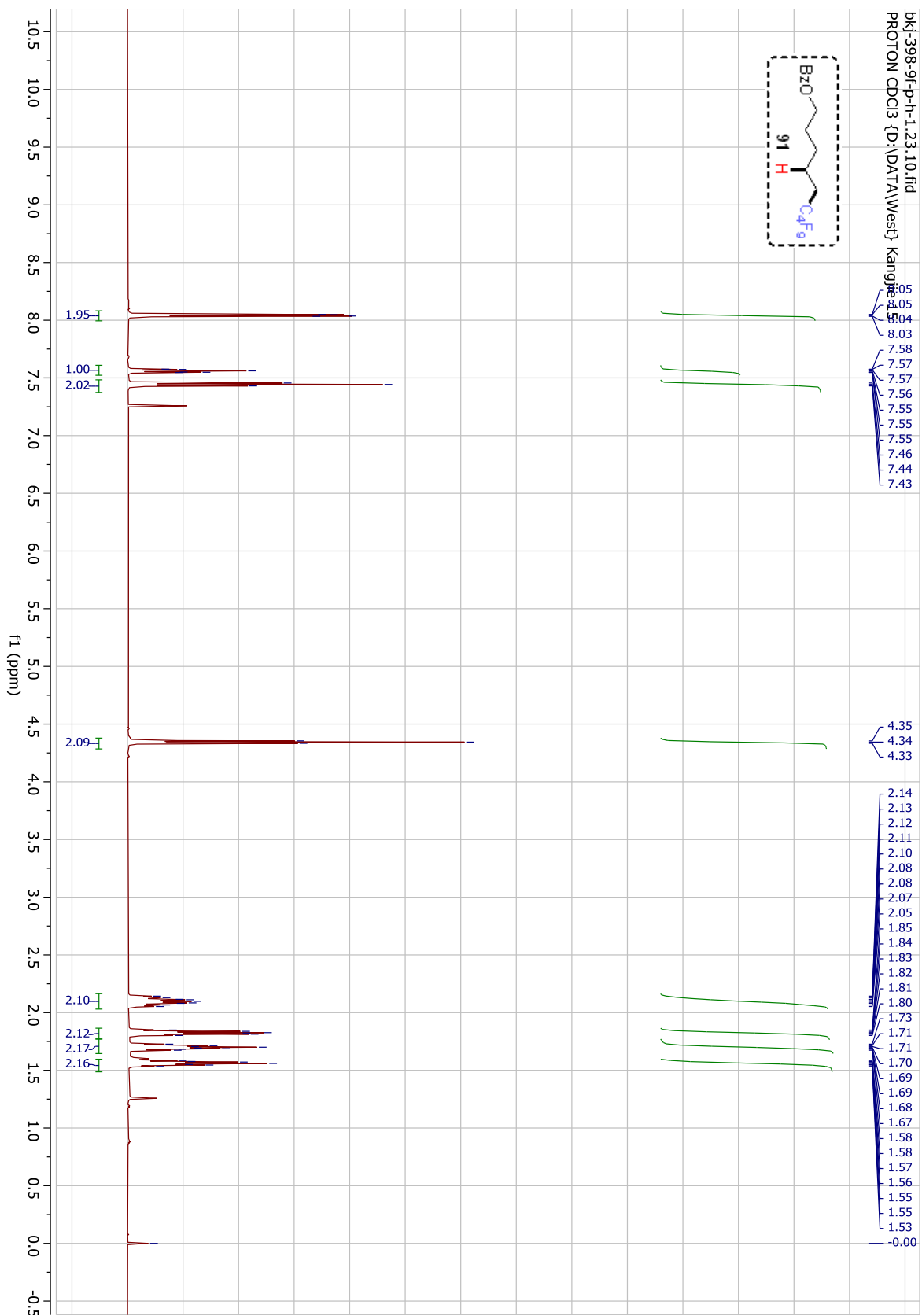


bkj-399-c-p-c-1_30.1.fid
C13CPD CDCl3 (D:\DATA\Westj_k\figlie_19

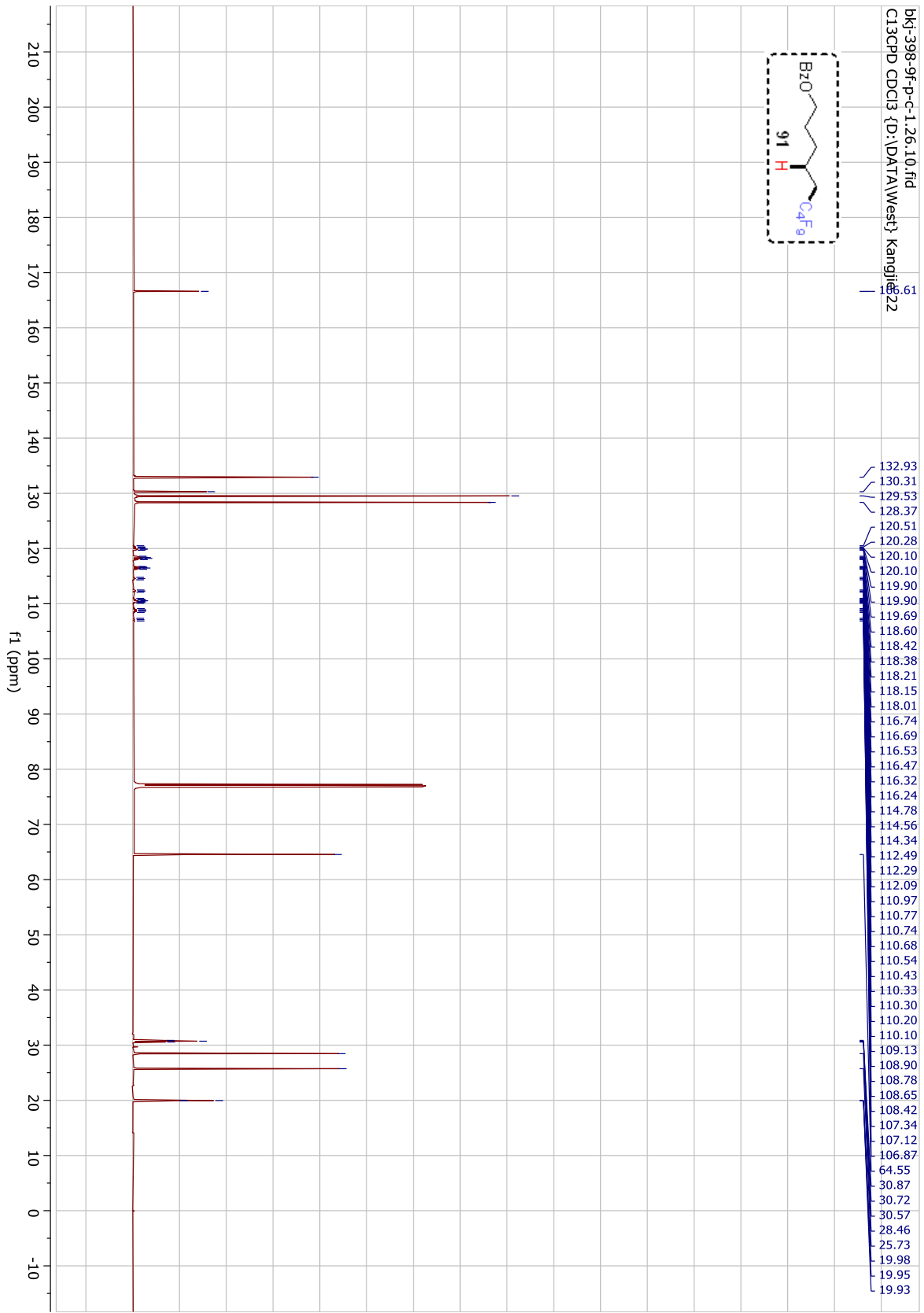
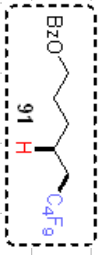


bjl-399-c-p-h-1.26.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 3

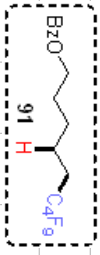




bk1-398-9f-p-c-1.26.10.fid
C13CPD CDCl3 (D:\DATA\Westj Kanji)\822

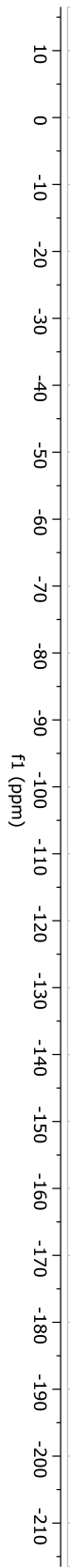


bkj-398-9f-p-h-1.23.11.fid
F19_baseline_correct 1D
F19_baseline_correct CDCl3 {D:\DATA\West\ Kangjie 15

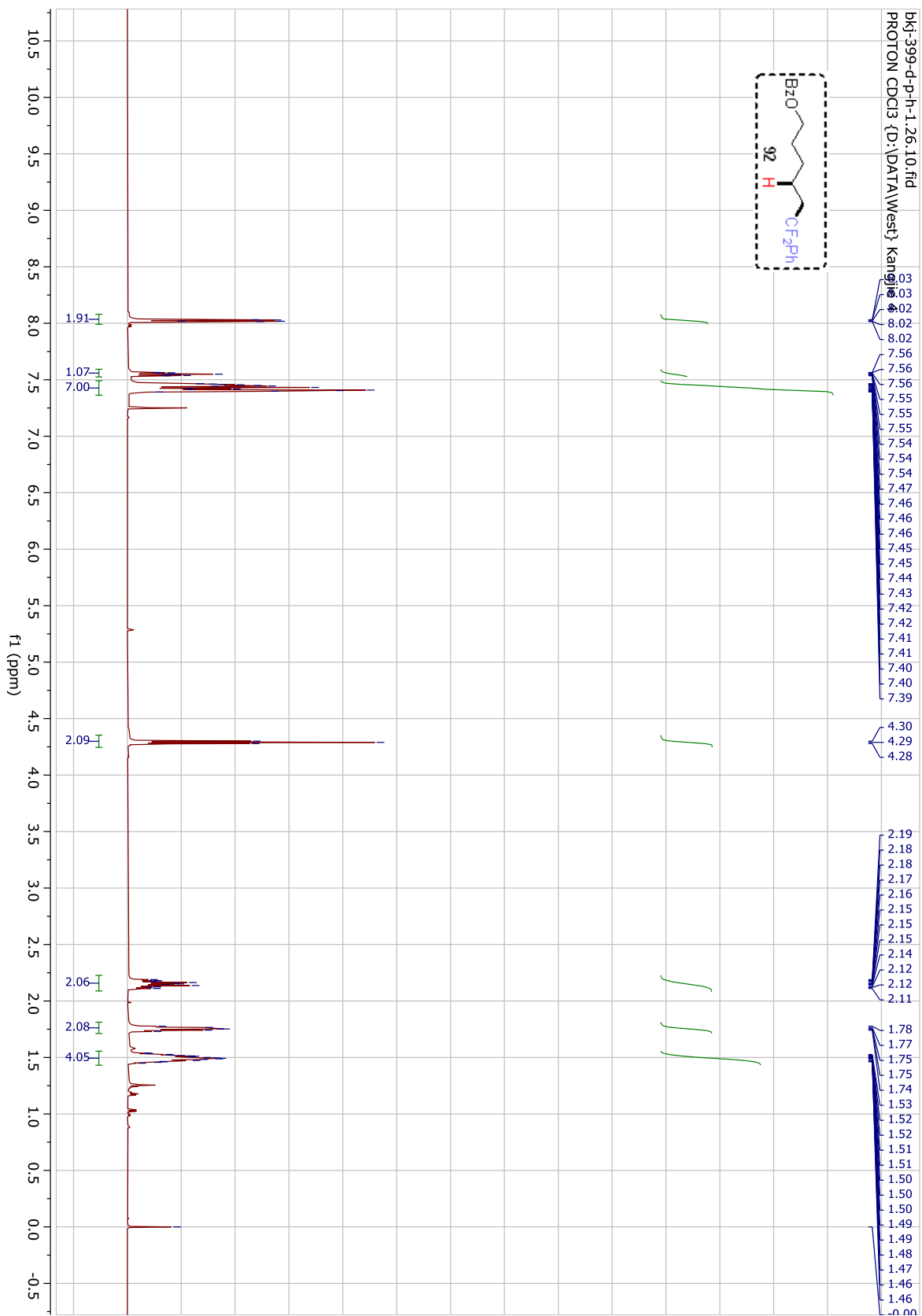


-81.07
-81.07
-81.09
-81.11
-81.12

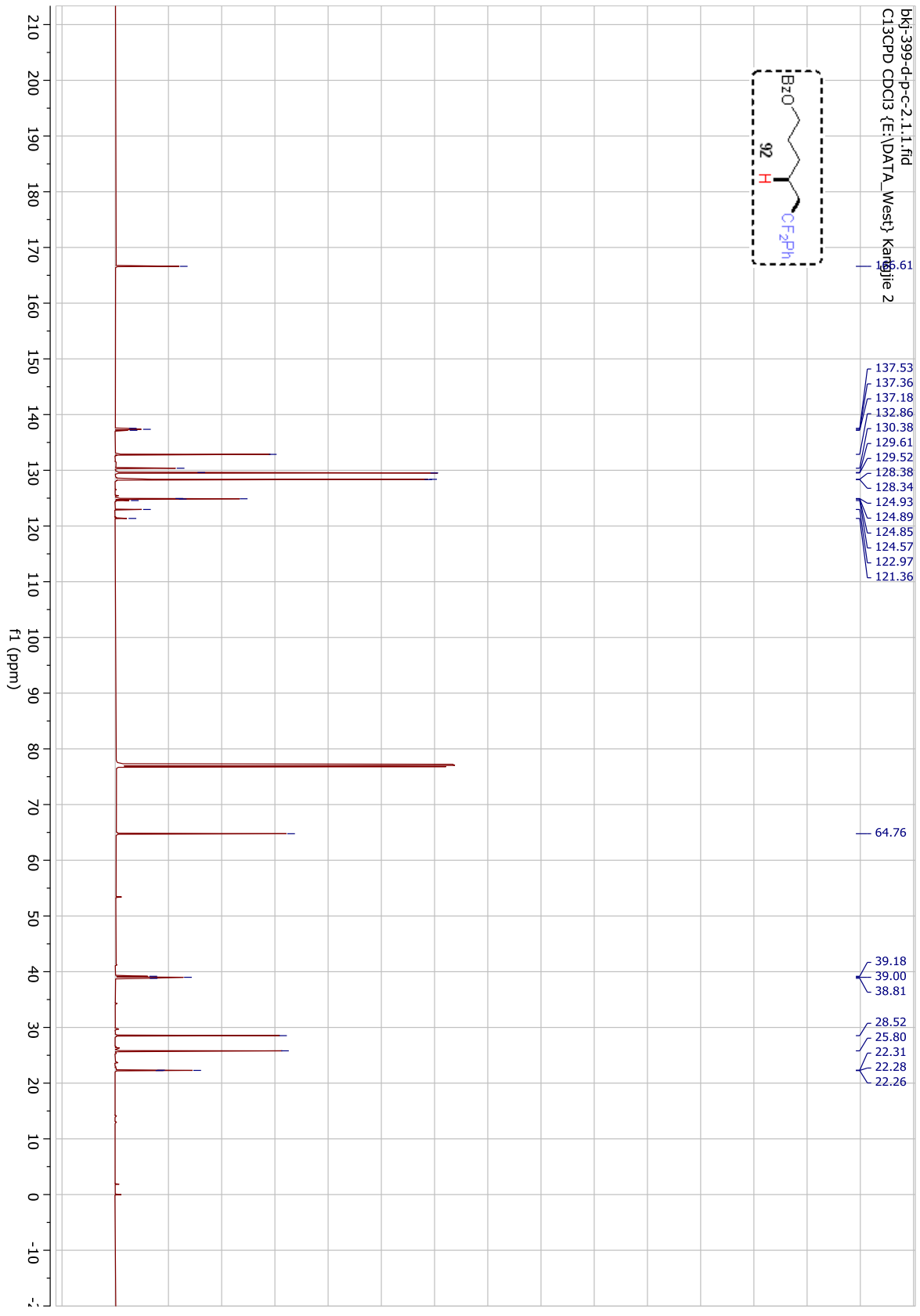
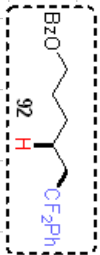
-114.55
-114.58
-114.61
-114.64
-114.68
-124.49
-124.51
-126.03
-126.04
-126.05
-126.07
-126.08
-126.10
-126.10
-126.11



350

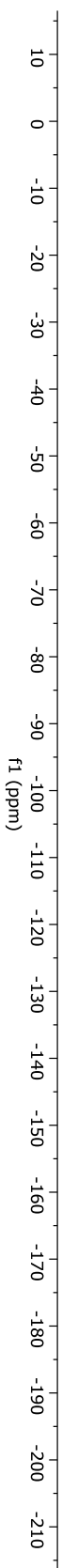
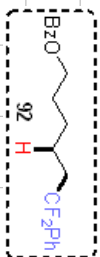


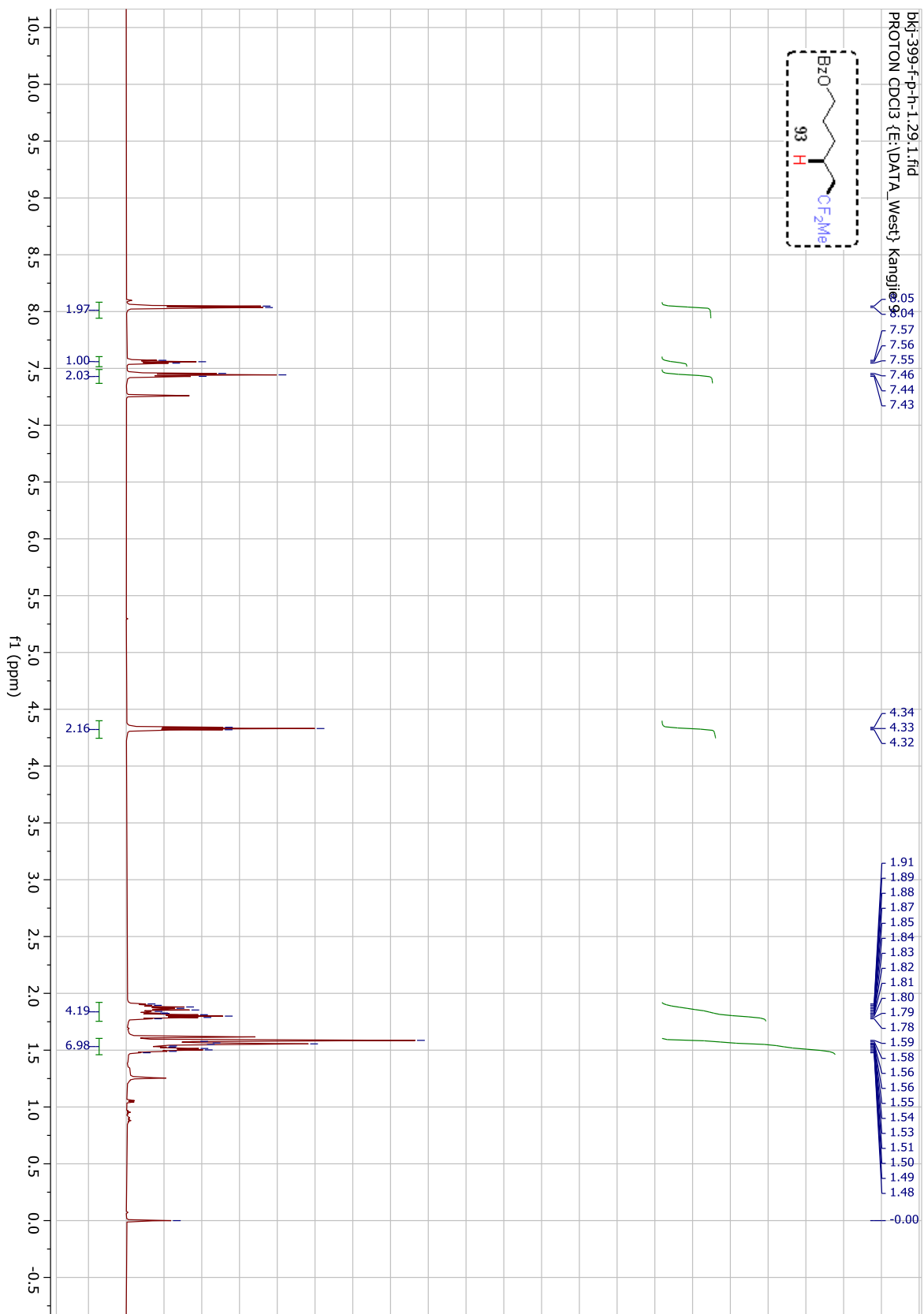
bkj-399-d-p-c-2-1.1.fid
C13CPD CDCl3 {E:\DATA_West\} Kanigie 2



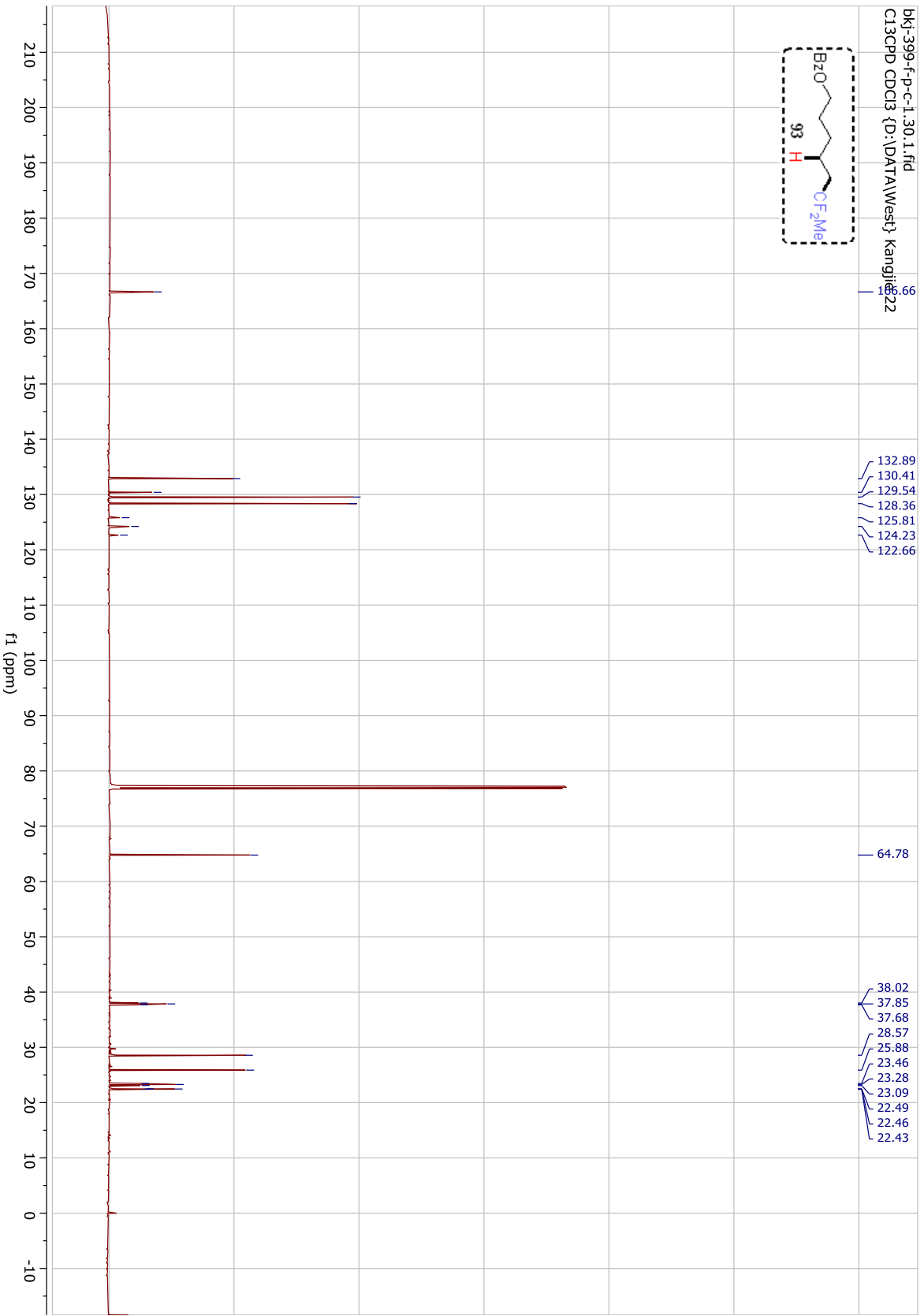
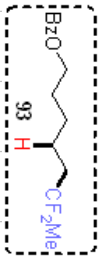
bkj-399-d-p-h-1.26.11.fid
F19_baseline_correct ID
F19_baseline_correct CDC13 {D:\DATA\West} Kangjie 4

-95.48
-95.51
-95.54

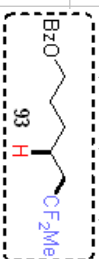




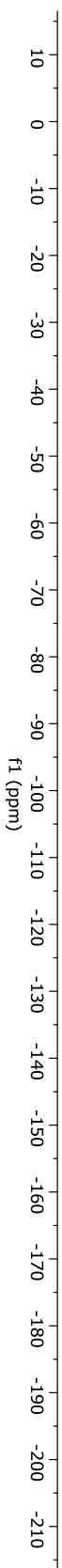
bkt-399-f-p-c-1.30.1.fid
C13CPD CDCl3 (D:\DATA\Westj Kanji)\822

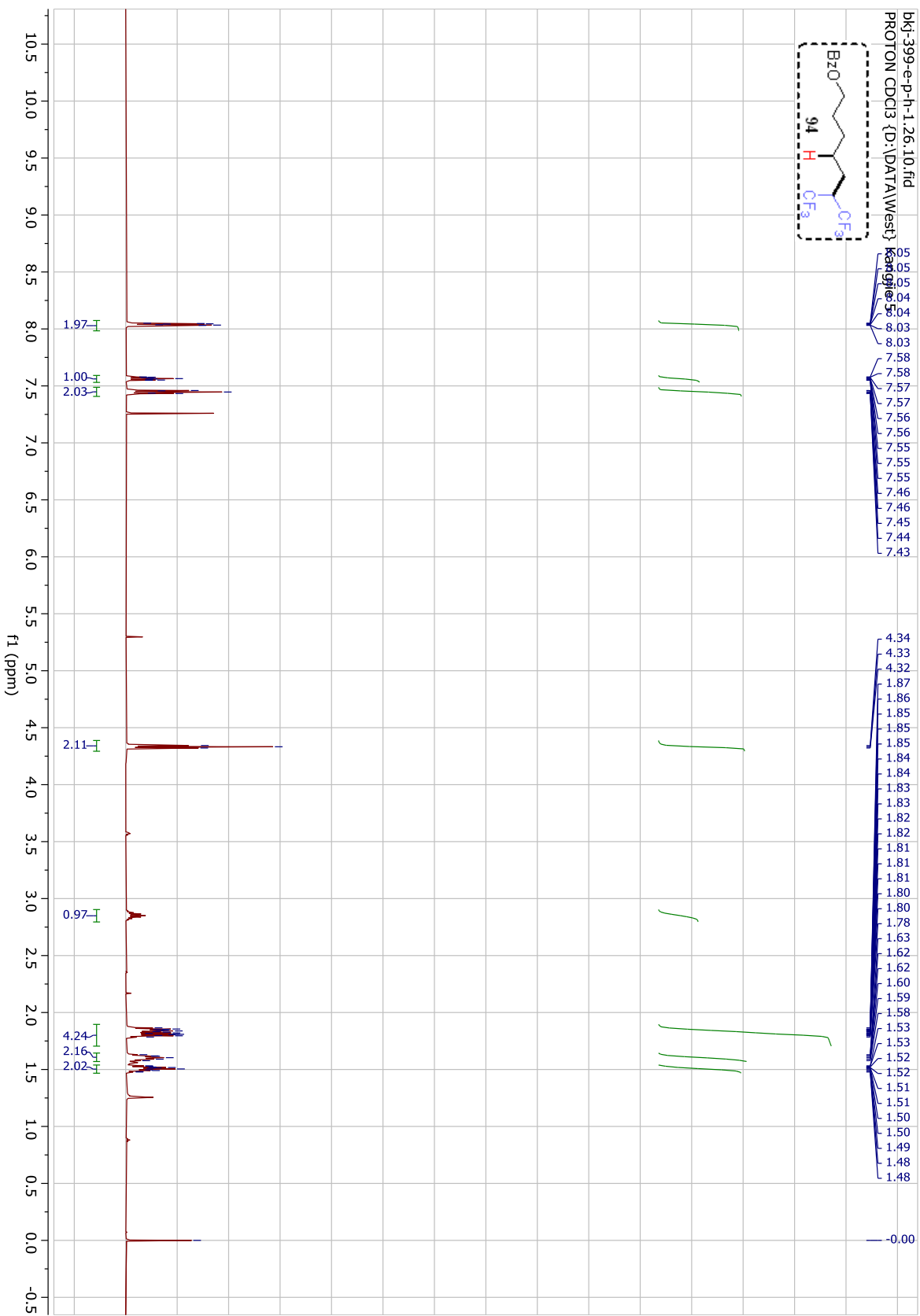


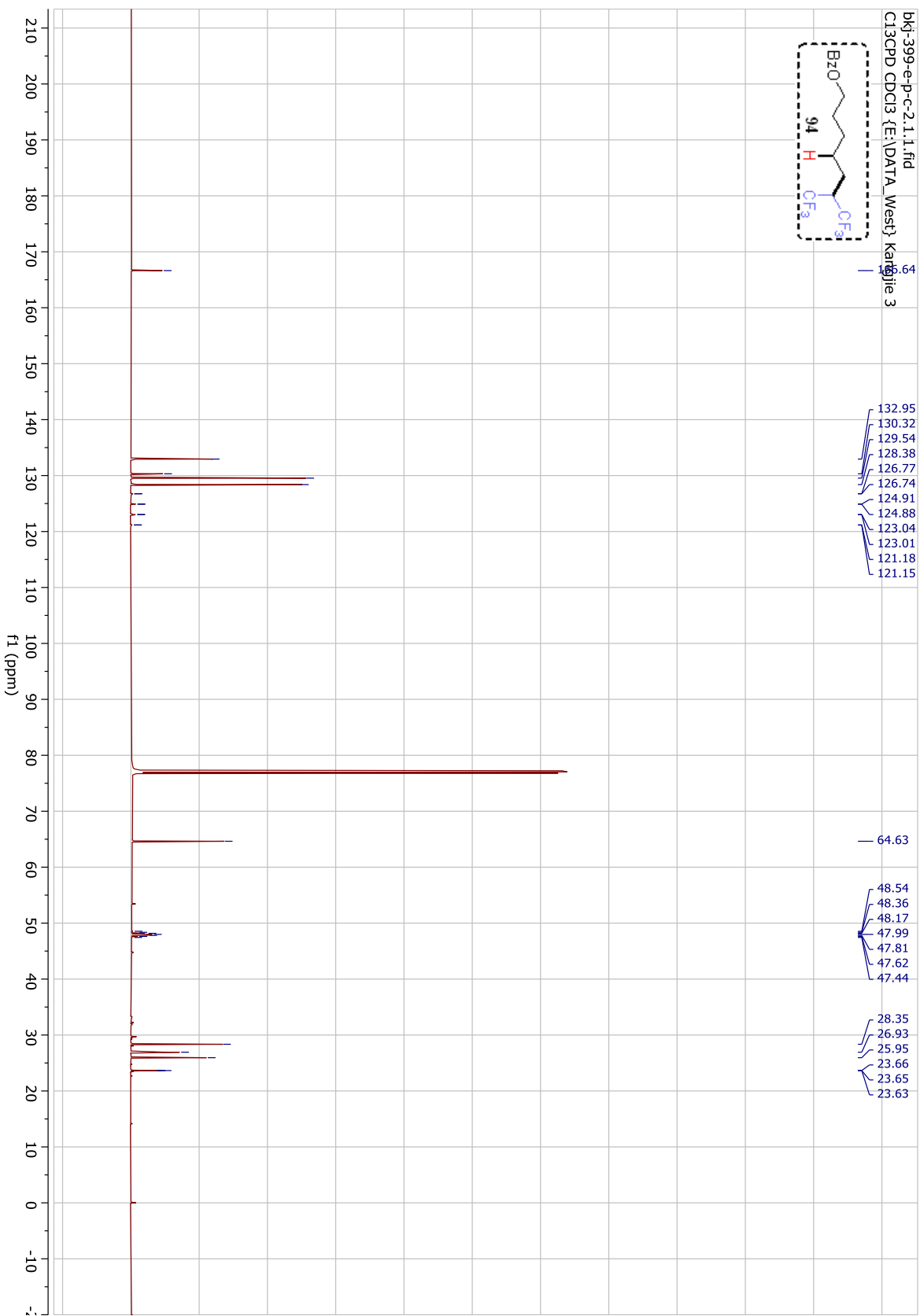
bkt-399-f-p-f1.30.1.fid
F19 CDCl3 {D:\DATA\West}\Kangjie 22



-90.55
-90.58
-90.61
-90.64

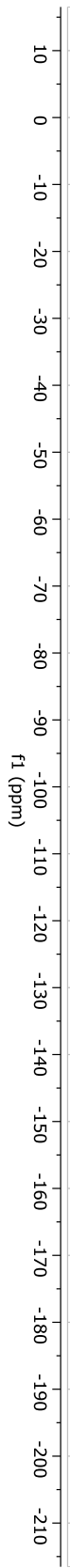
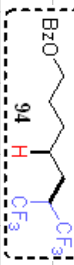


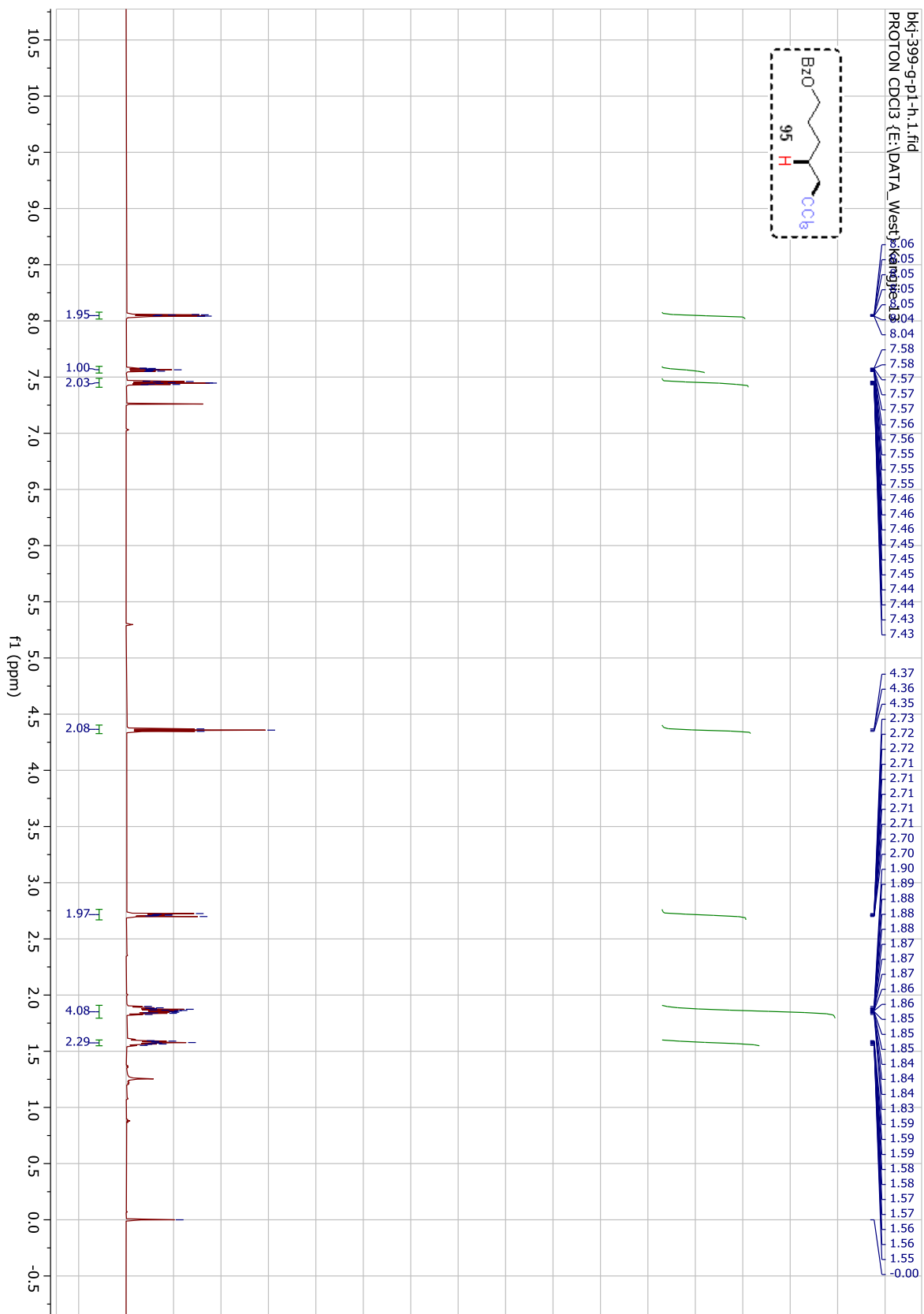




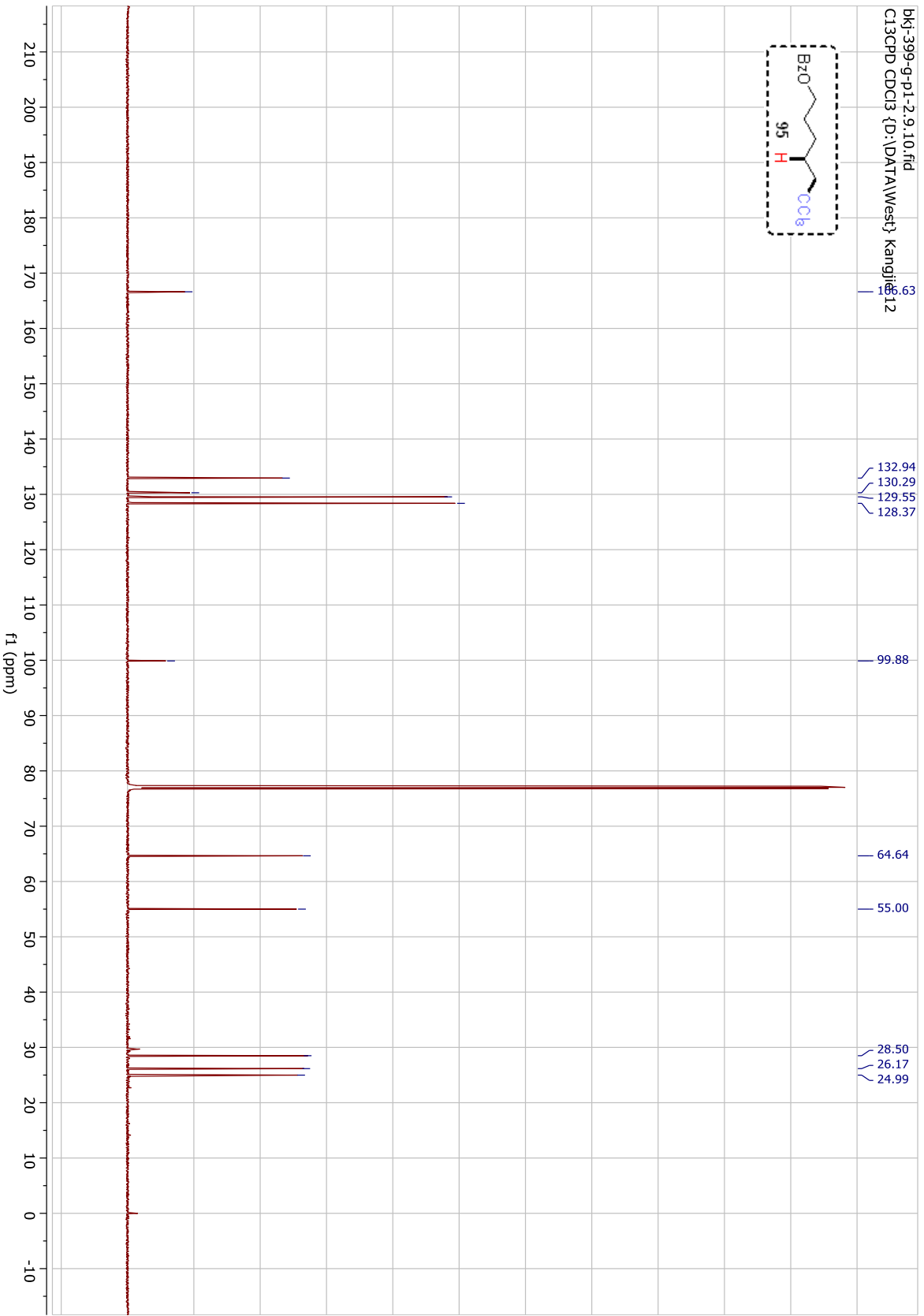
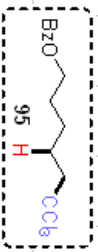
bkj-399-e-p-h-1.26.11.fid
F19_baseline_correct ID
F:\baseline_correct\11712\DATA\West\Kangjie 5

-67.05
-67.06

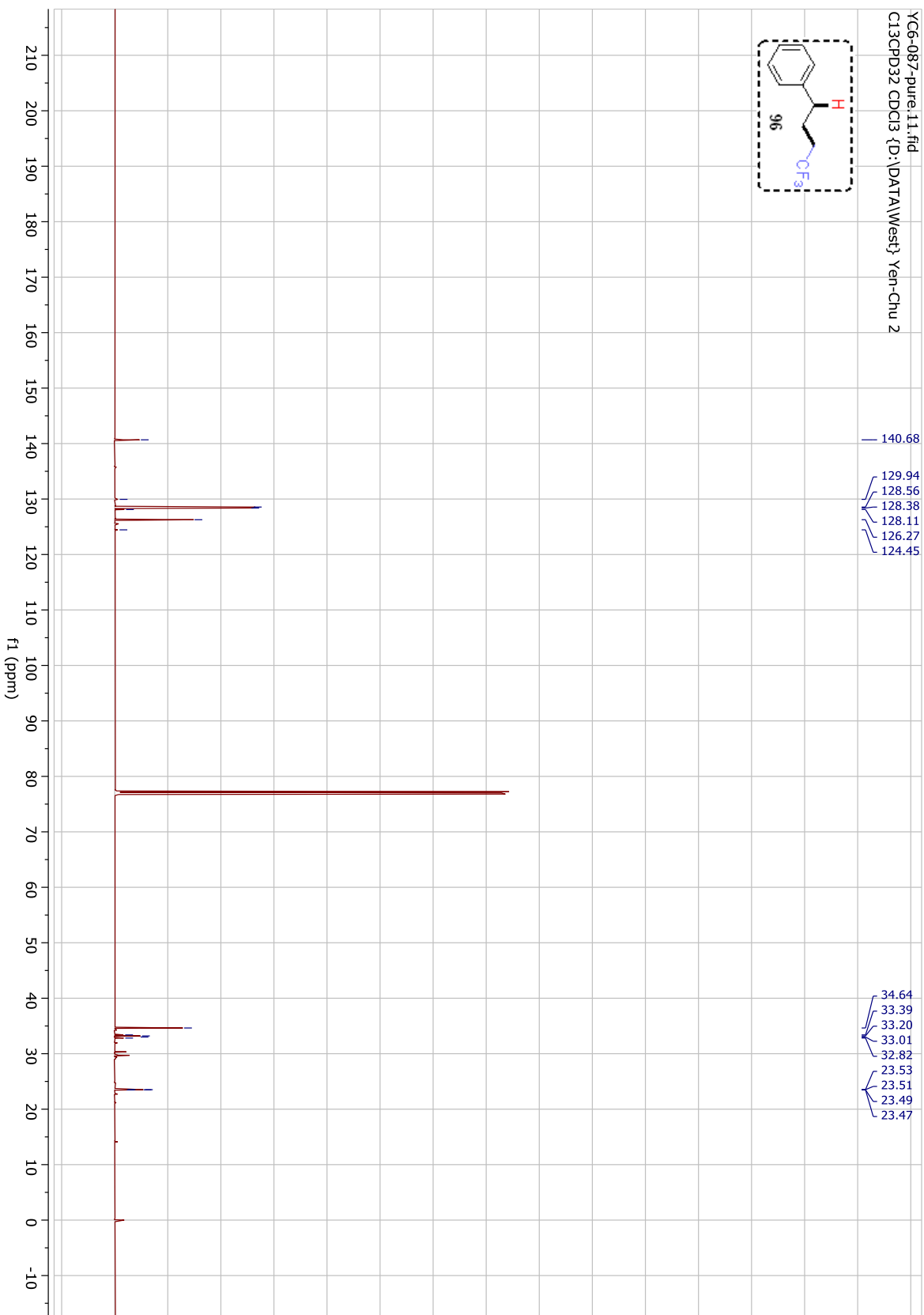
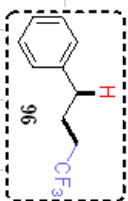




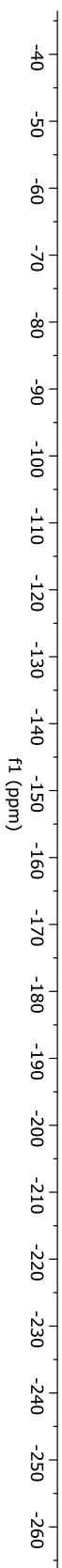
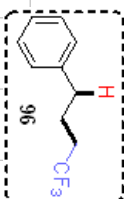
bkt-399-g-p1-2-9.10.fid
C13CPD CDCl3 (D:\DATA\Westj Kanji)

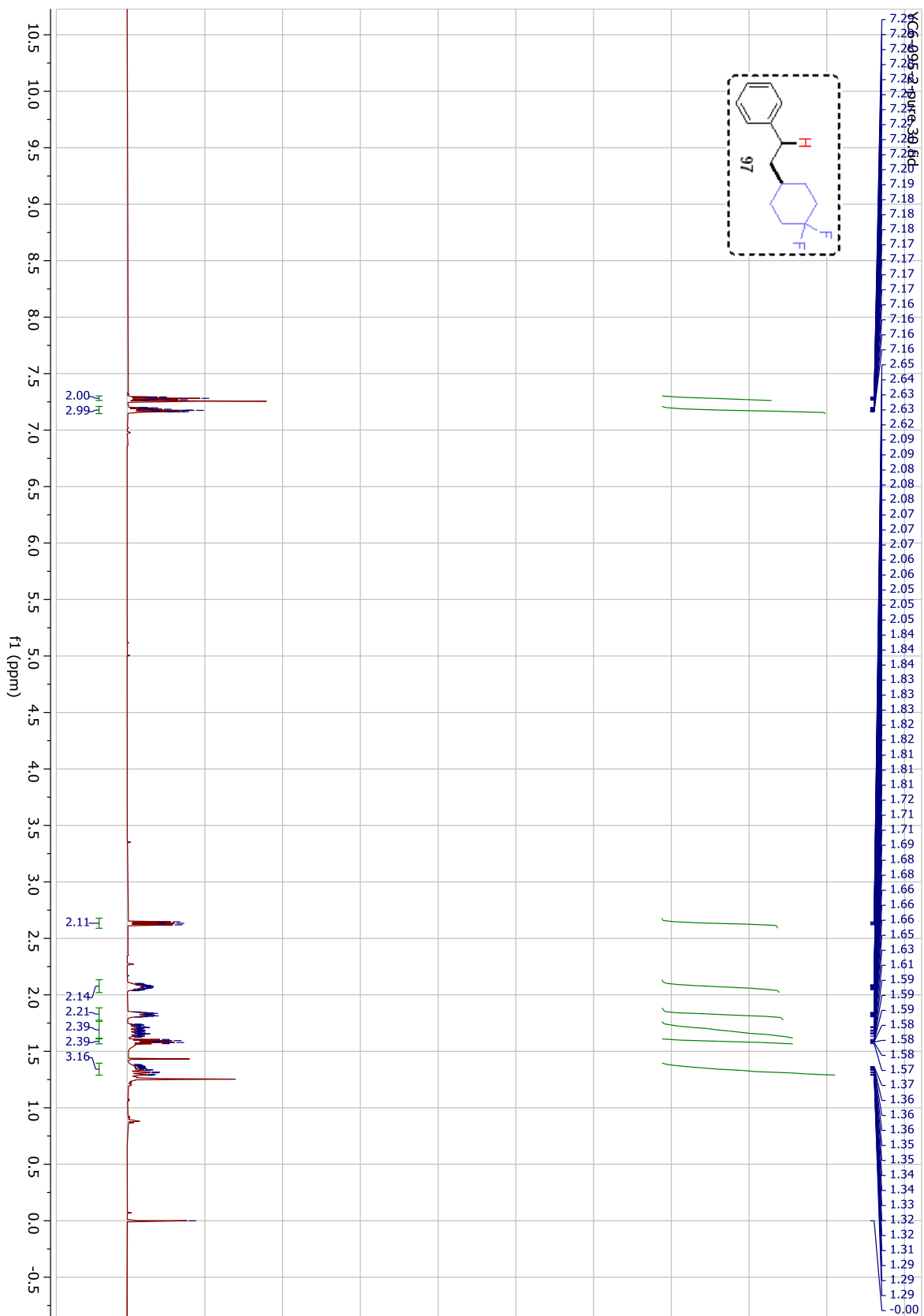


YC6-087-pure1.1.fid
C13CPD32 CDCl3 {D:\DATA\West}\Yen-Chu 2

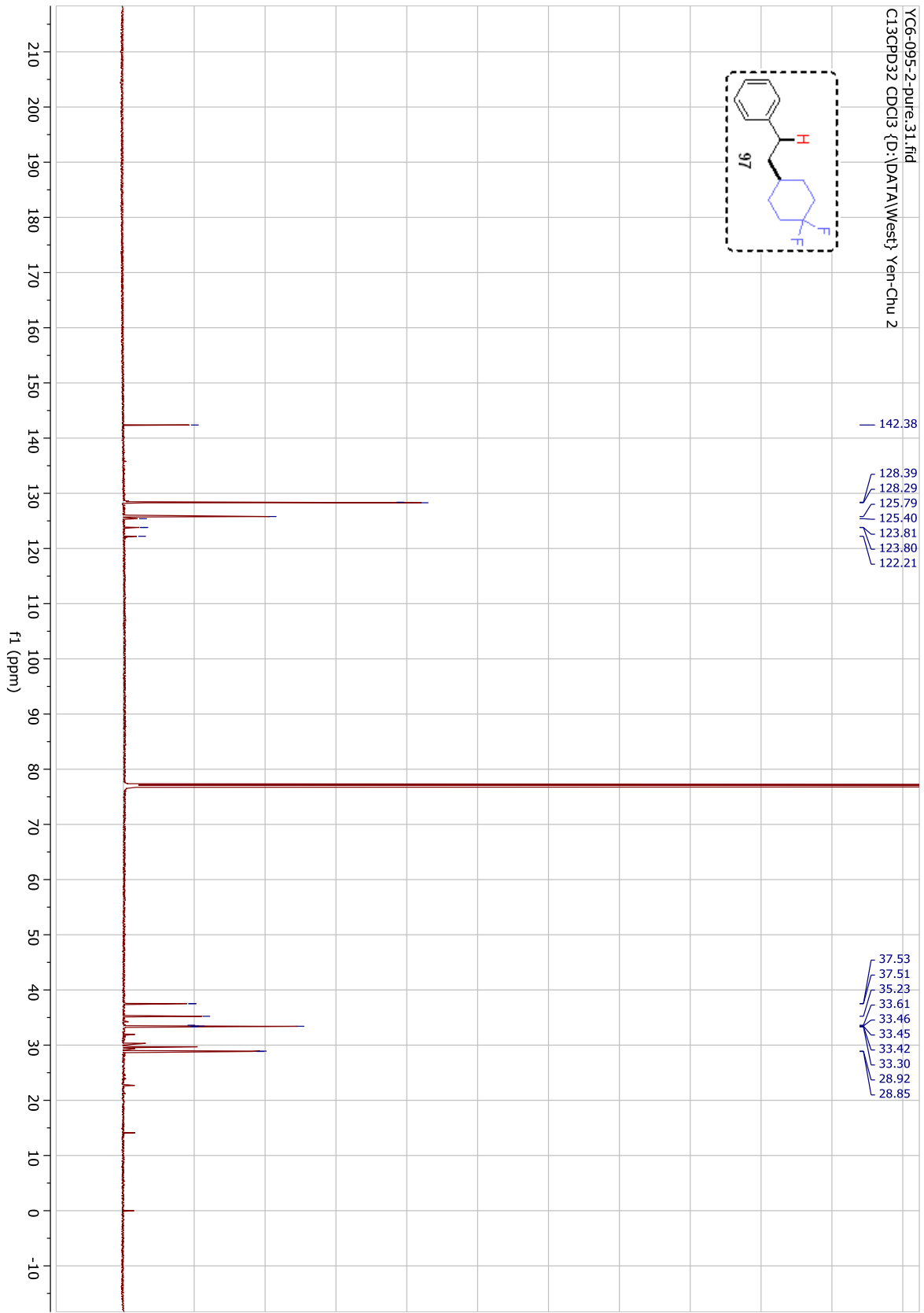
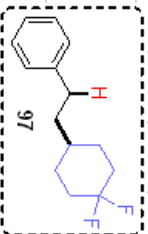


YCG-087-pure.12.fid 91.89 92.02
F19_baseline_correct 80.89
F19_baseline_correct CDCl3 {D:\DATA\West\ Yen-Chu 2

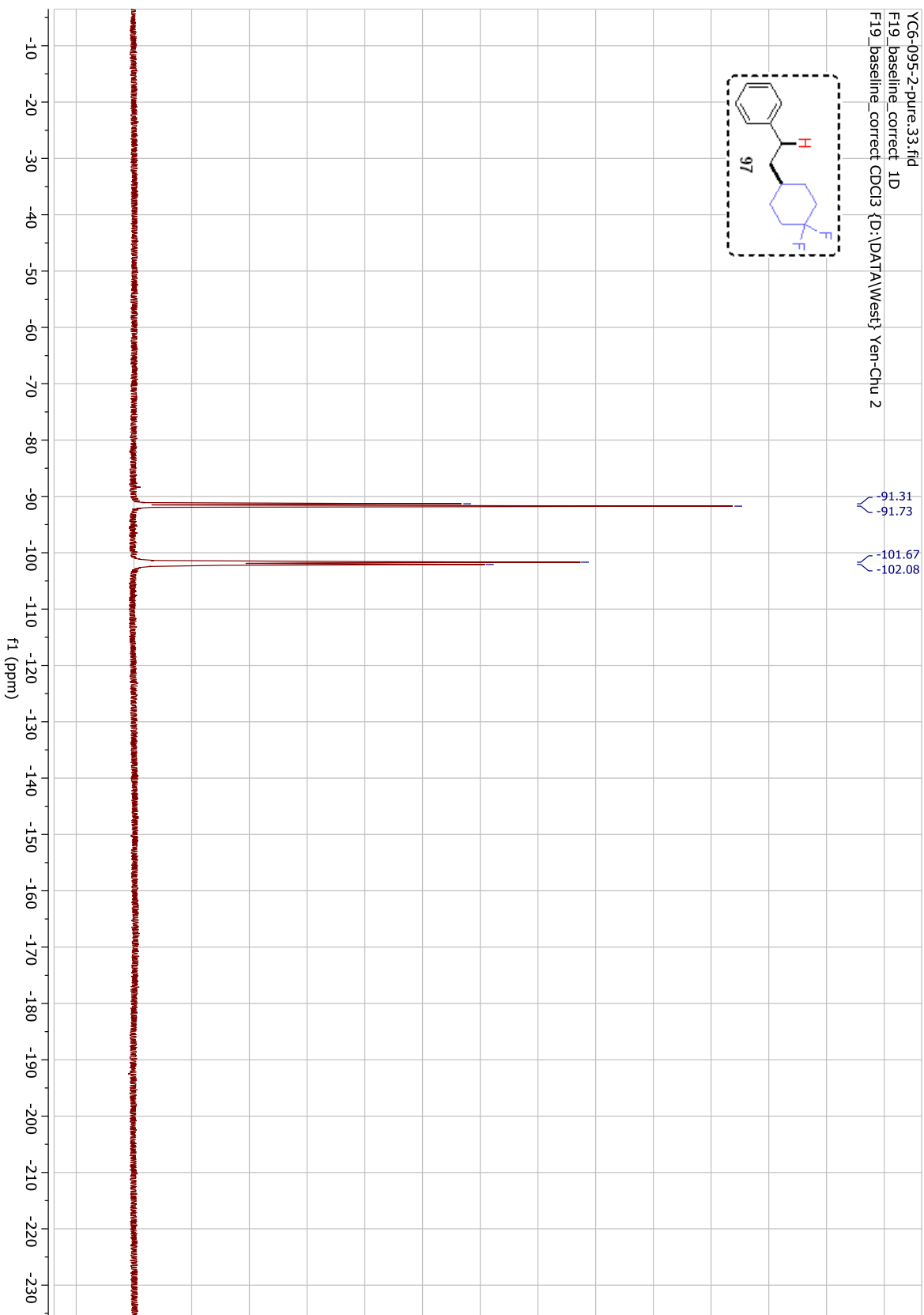
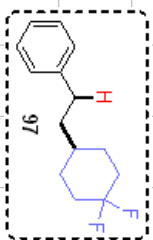


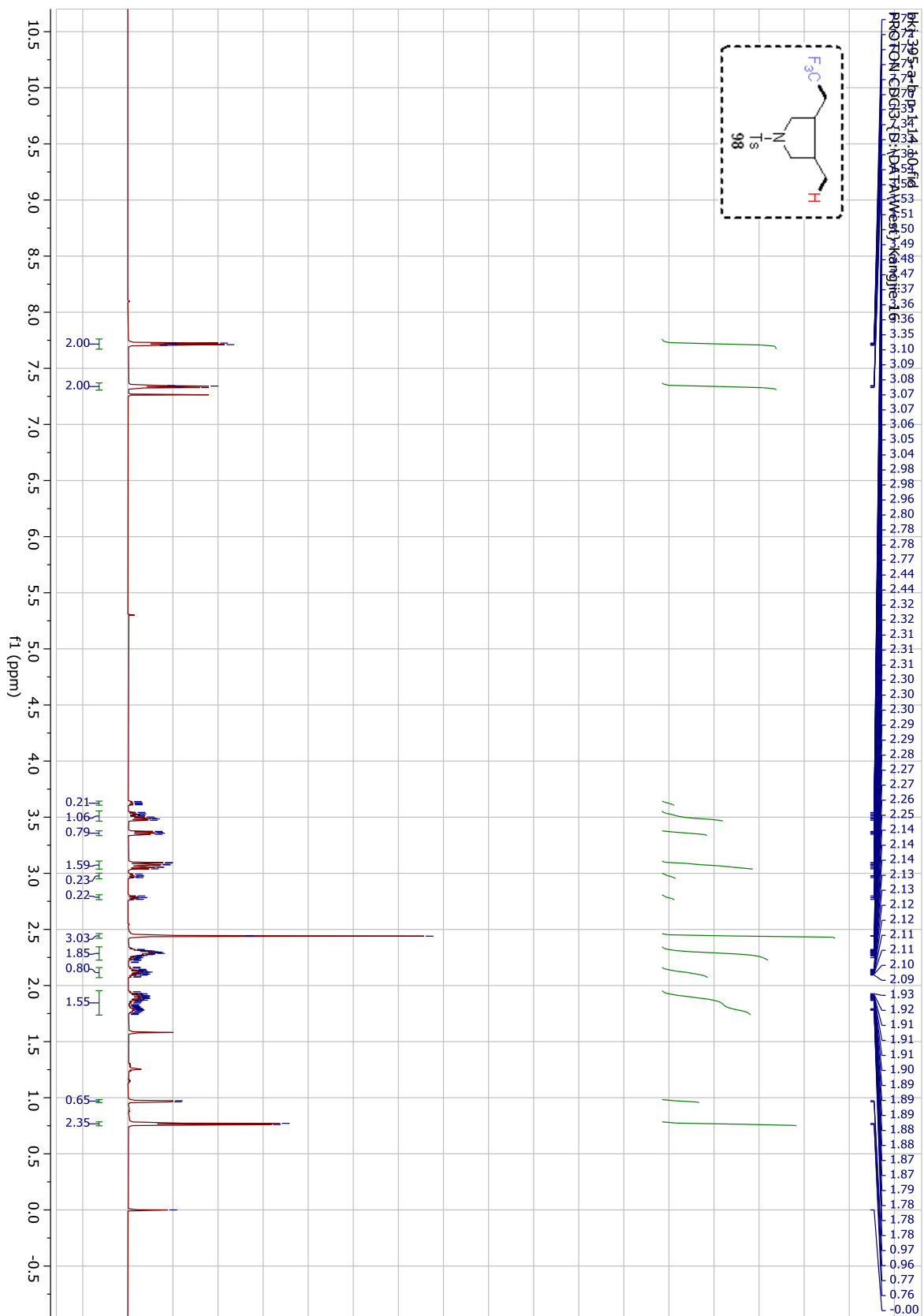


YC6-095-2-pure_31.fid
C13CPD32 CDCl3 {D:\DATA\West\} Yen-Chu 2

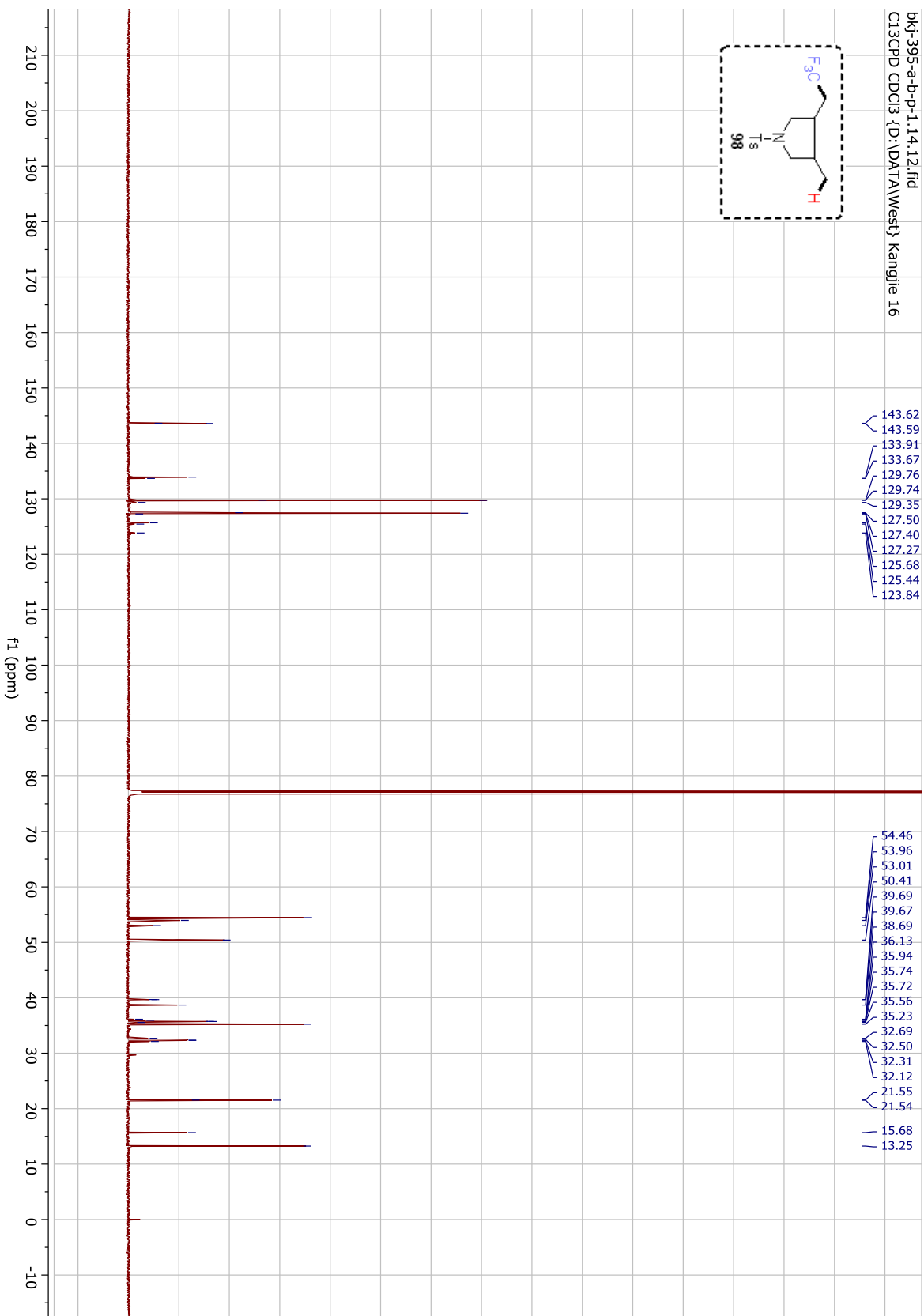
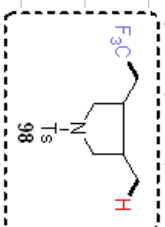


YC6-095-2-pure.33.fid
F19_baseline_correct 1D
F19_baseline_correct CDC13 {D:\DATA\West\} Yen-Chu 2

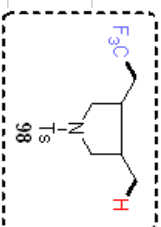




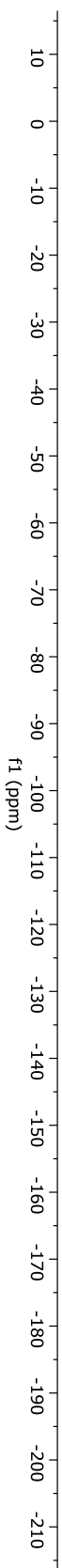
hkl-395-a-b-p-1-14.12.fid
C13CPD CDCl3 (D:\DATA\Westj Kangjie 16



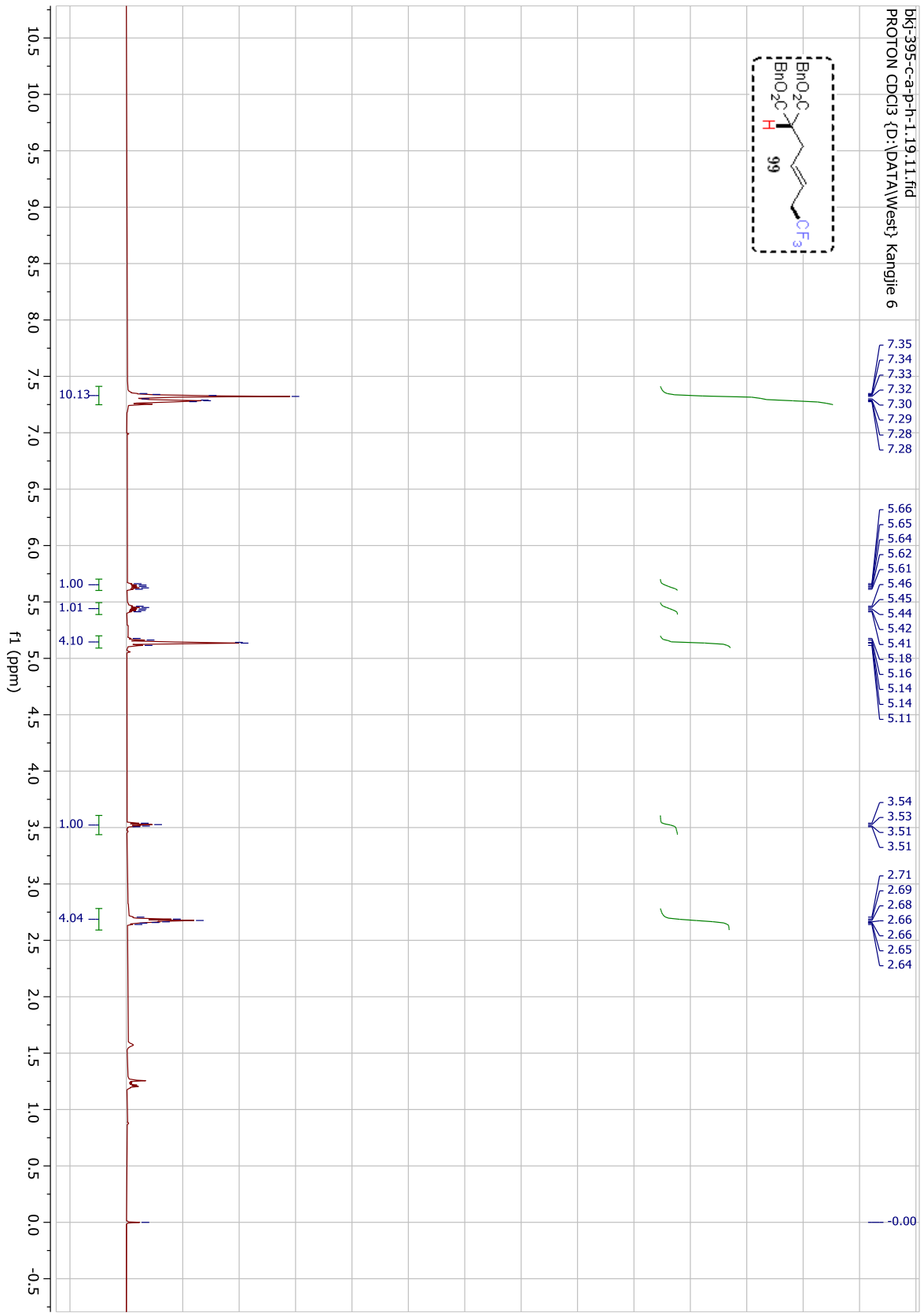
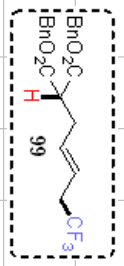
bkj-395-a-b-p-1.14.11.fid
F19_baseline_correct ID
F19_baseline_correct CDCI3 {D:\DATA\West} Kangjie 16



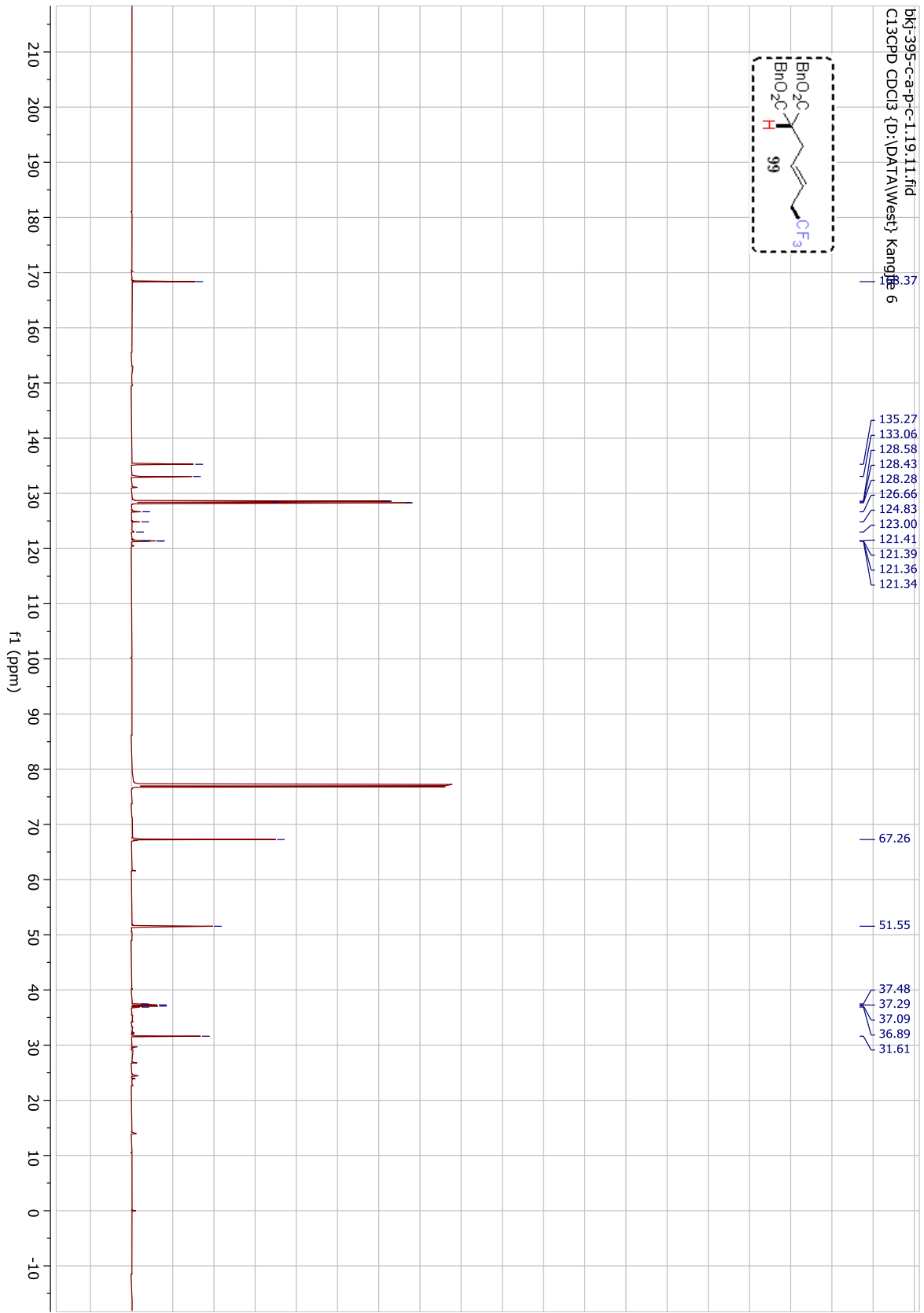
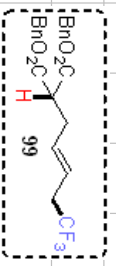
64.97
64.99
65.01
65.03
65.05



bk1-395-c-a-p-h-1.19.11.fid
PROTON CDCl3 {D:\DATA\Westf\ Kangjie 6

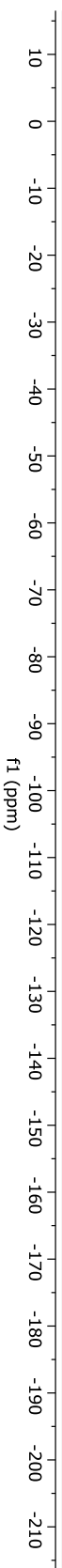
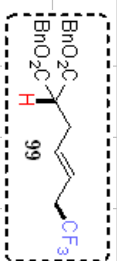


bjl-395-c-a-p-c-1.19.11.fid
C13CPD CDCl3 (D:\DATA\Westj_Kang) 6



bkj-395-c-a-p-f-1.19.11.ftd
F19_baseline_correct CDCl3 {D:\DATA\Westfj Kangjie 6

66.49
66.51
66.53



IV. Supplemental Reference

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