



Figure S1

Figure S1. Summary. In age, ATA B increased in PBL and CD5 can decrease, and increase of CD11b in ATA B, aMyIIA, and aPtC. V8-12 generation in MZ B cell.

(A) 2 mo ATA μ Tg mice with ATA B ($V_{H}8-12/V_k21-5$) show BM and most Spl with immature AA4 $^{+}$, different most mature in PerC, mLN, and intestinal PP. In PBL, 2 mo ATA B cells are immature, then, 8 mo B1a cells increase with higher mature (AA4 $^{-}$) ATA B cells (1), and further increase mature ATA B in 14 mo, contrast low ATA B cells in originally Pla2g2a negative ATA μ Tg mice (1). Serum IgM, IgG1, IgG2, IgG3 also increase from 2 mo to 12 mo, and IgA originally 2 mo high. **(B)** Summary (2). Left: CD5 down-regulation after stimulation of ATA B pB1 cells in ATA μ Tg mice by cell proliferation by LPS or CpG in 3 days. Middle: Increased ATA B cell frequency in PBL B cells in ATA μ Tg mouse at 12 mo, with further B220 down and CD5 reduction. Right: CD5 down-regulation occurrence at CLL stage also in TC $^{+}$ ATA μ Tg mice and CD5 $^{+}$ and CD5 $^{-}$ CLL in PBL found in two littermates at 14 mo. **(C)** Middle: aMyIIA B1 B cells showed CD5 down and CD11b $^{+}$ in old aged, summary of aMyIIA B (3). Right: ZAP70 expression determined by quantitative PCR in aMyIIA B cells from the spleen (s) and peritoneal cavity (p), after the tumor stage. Relative messenger RNA transcript levels with FO B was set to 1.0. Down: Summary of $V_{H}11t$ (4). Left: Three days after indicated CD11b stimulation, LPS (low), CpG, CpG+IL-10 (high), of spleen B1a cells from an adult 2 mo $V_{H}11t$ mice. Middle: In PBL, 12 mo aPtC B cells are 22 % and increased 27 mo 86.7% (#952) in aPtC ($V_{H}11/V_k9$) in CD19 $^{+}$ B cells as high MBL. Right: Comparison of CD11b expression levels by aPtC B1a cells in Spl between day 10 neonate as CD11b $^{-}$, and 27 mo as CD11b $^{+}$. **(D)** Summary about AGcA μ Tg and AGcA μ kTg mice with MZ B (5). Left: Analysis of B cell subsets in the spleen. In FO B and MZ B cells, $V_{H}3609$ (V8-12) id $^{+}$ cells marked by a vertical line. The majority of 13H8K ($V_k19-17/Jk1$) $^{hi/med}$ Igk B cells are present in low FOB, and MZ B cells are higher. Right: AGcA μ kTg mice, as EP69. 3 wk of EP69 μ kTg mice showed dominantly in MZ B cells with the $V_{H}8-12/V_k19-17$.

1. Susan A.Shinton, Joni Brill-Dashoff, Kyoko Hayakawa. (2022). Pla2g2a promotes innate Th2-type immunity lymphocytes to increase B1a cells. *Scientific Reports* 12:14899
2. Hayakawa K, Formica AM, Brill-Dashoff J, Shinton SA, Ichikawa D, Zhou Y, Morse III HC, Hardy RR. (2016). Early generated B1 B cells with restricted BCRs become chronic lymphocytic leukemia with continued c-Myc and low Bmf expression. *J. Exp. Med.* 213:3007.
3. Hayakawa, K., Formica, A.M., Colombo, M.J., Shinton, S.A., Brill-Dashoff, J., Morse III, H.C., Li, Y.S., Hardy, R.R. (2016). Loss of a chromosomal region with syntheny to human 13q14 occurs in mouse chronic lymphocytic leukemia that originates from early-generated B-1 B cells. *Leukemia* 30:1510.
4. Hayakawa K, Formica A.M, Nakao Y, Ichikawa D, Shinton, S.A, Brill-Dashoff J, Smith, M.R, Morse III, H.C, Hardy, R.R. (2018). Early generated B-1 derived B cells have the capacity to progress to become mantle cell lymphoma-like neoplasia in aged mice. *J. Immunol.* 201: 804.
5. Ichikawa, D., Asano, A., Shinton, S.A., Brill-Dashoff, J., Formica, A.M., Velcich, A., Hardy, R.R., and Hayakawa K. (2015). Natural anti-intestinal goblet cell autoantibody production from Marginal zone B cells. *J. Immunol.* 194:606.