

## **Supporting Information for**

Vulnerability to APOBEC3G linked to the pathogenicity of deltaretroviruses.

Takafumi Shichijo<sup>1,2</sup>, Jun-ichirou Yasunaga<sup>1,2,\*</sup>, Kei Sato<sup>3,4</sup>, Kisato Nosaka<sup>1</sup>, Kosuke Toyoda<sup>1,2</sup>, Miho Watanabe<sup>1</sup>, Wenyi Zhang<sup>1</sup>, Yoshio Koyanagi<sup>5</sup>, Edward L. Murphy<sup>6,7</sup>, Roberta L. Bruhn<sup>7</sup>, Ki-Ryang Koh<sup>8</sup>, Hirofumi Akari<sup>9</sup>, Terumasa Ikeda<sup>10,11,12</sup>, Reuben S. Harris<sup>11,12</sup>, Patrick L. Green<sup>13</sup>, and Masao Matsuoka<sup>1,2,\*</sup>

### **Corresponding Authors:**

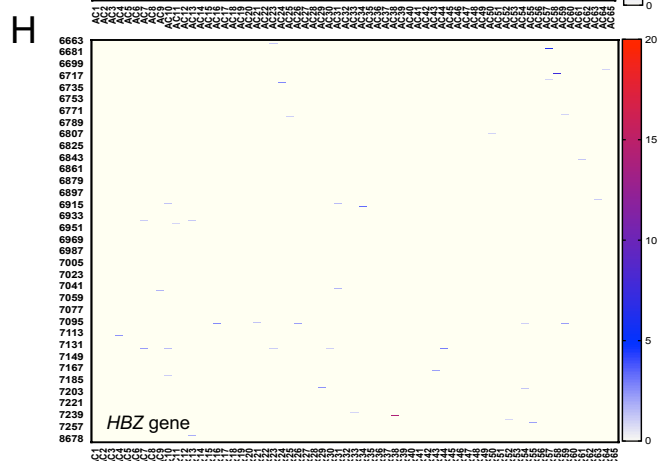
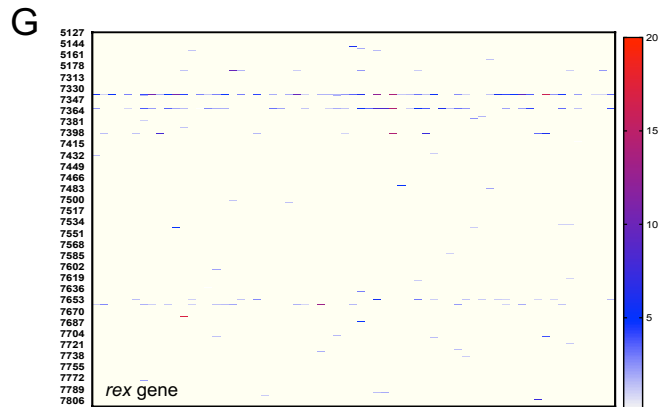
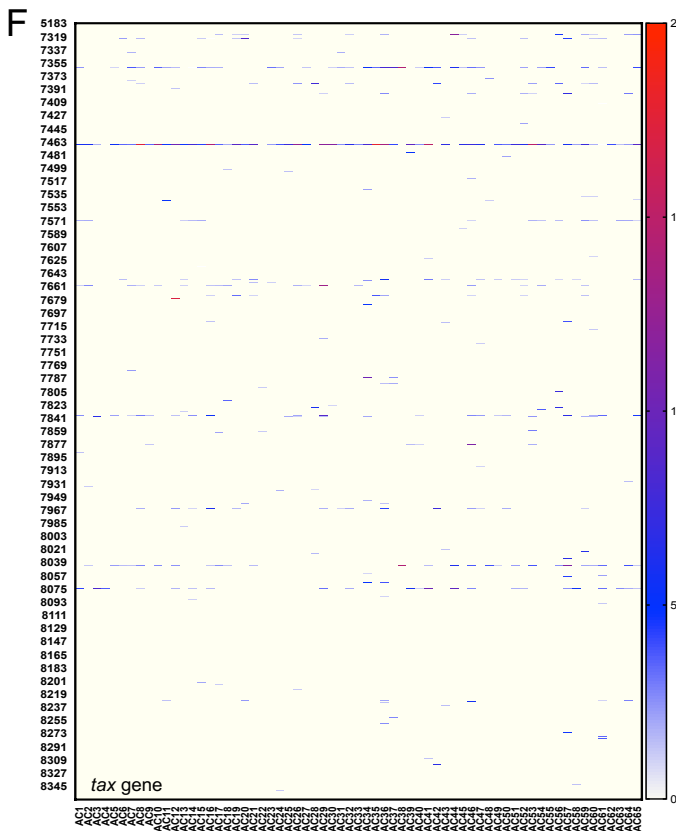
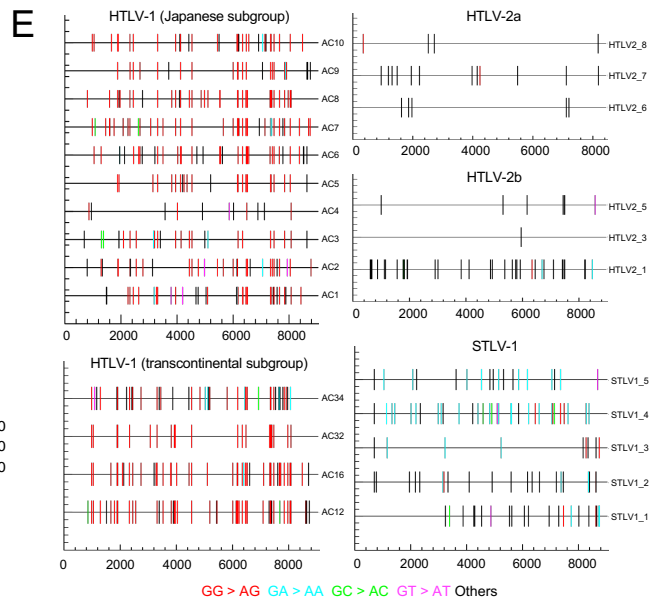
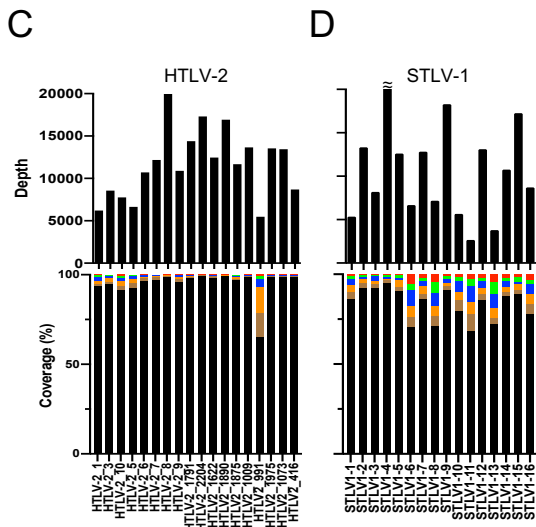
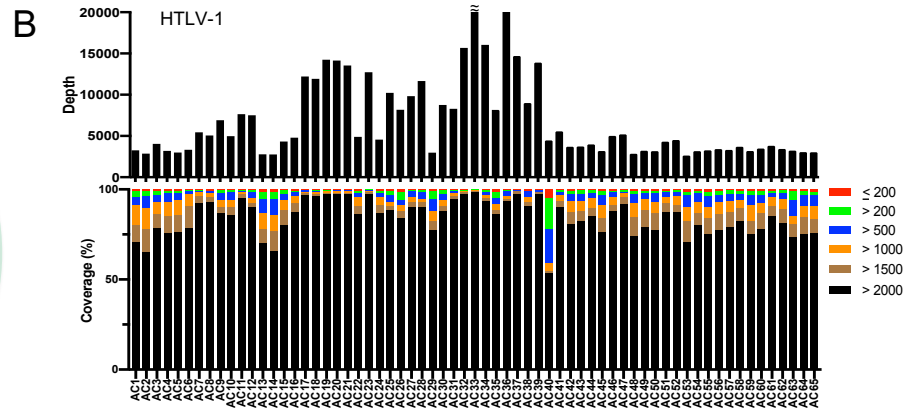
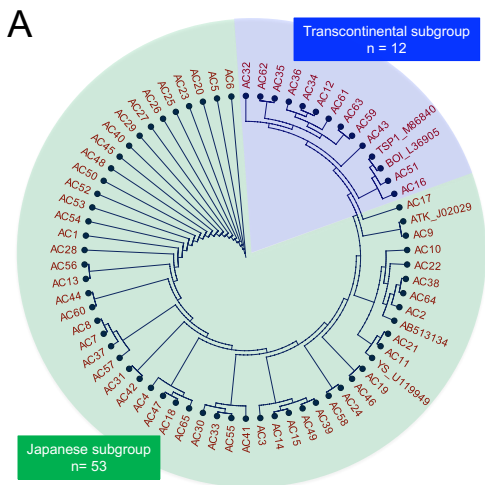
Masao Matsuoka, Jun-ichirou Yasunaga

Email: [mamatsu@kumamoto-u.ac.jp](mailto:mamatsu@kumamoto-u.ac.jp), [jyasunag@kumamoto-u.ac.jp](mailto:jyasunag@kumamoto-u.ac.jp)

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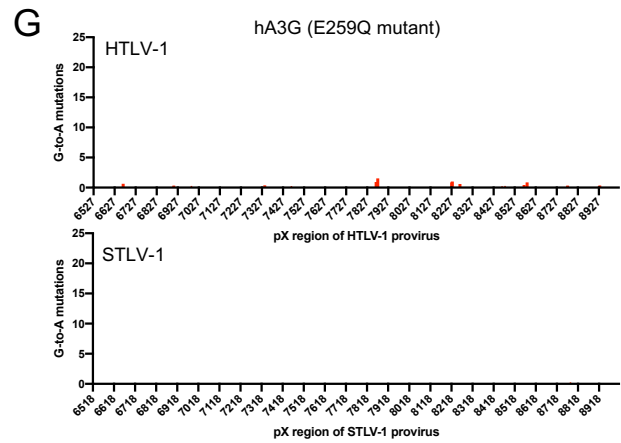
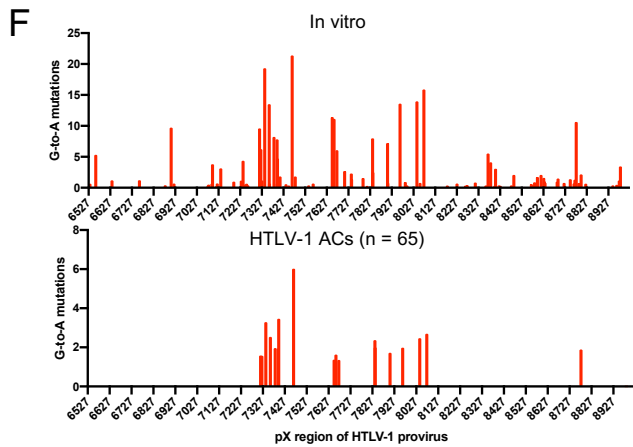
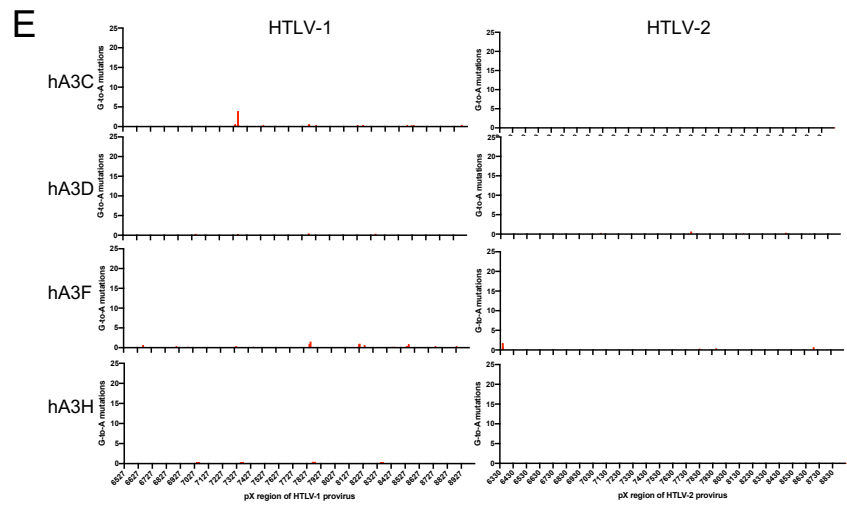
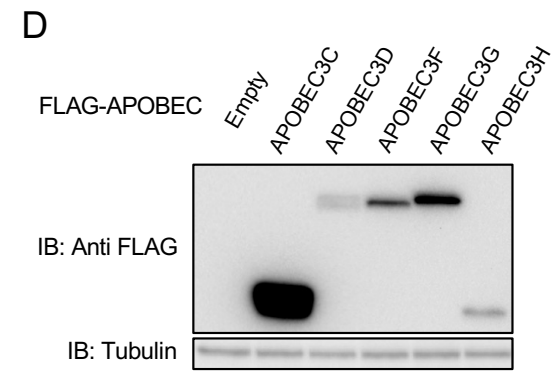
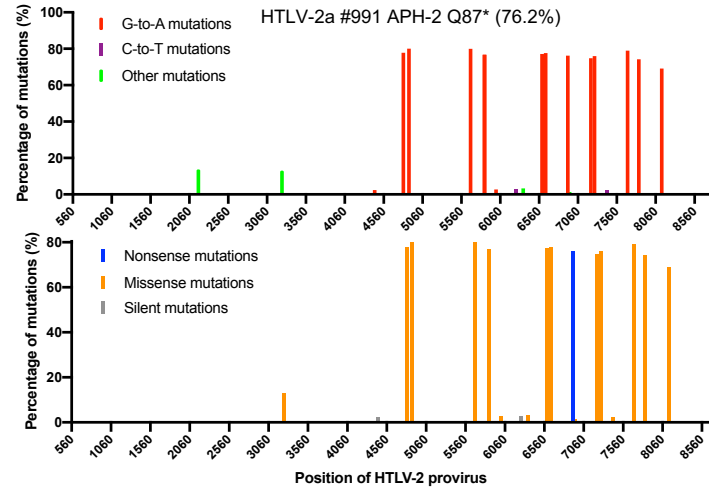
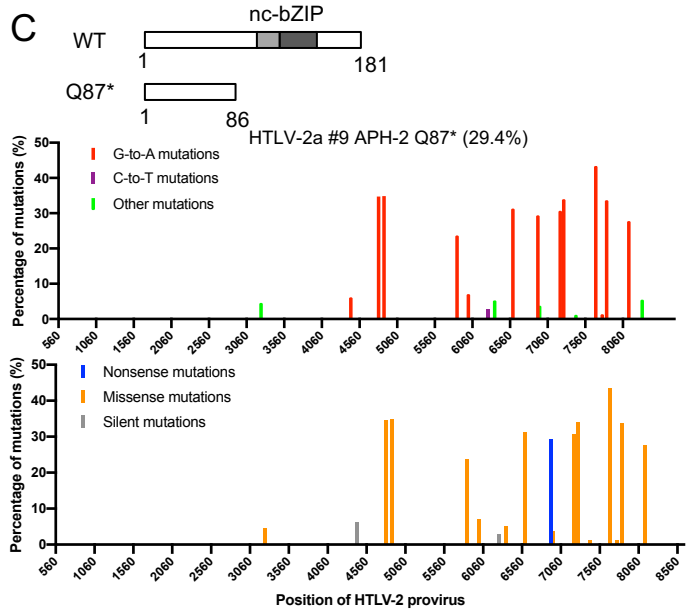
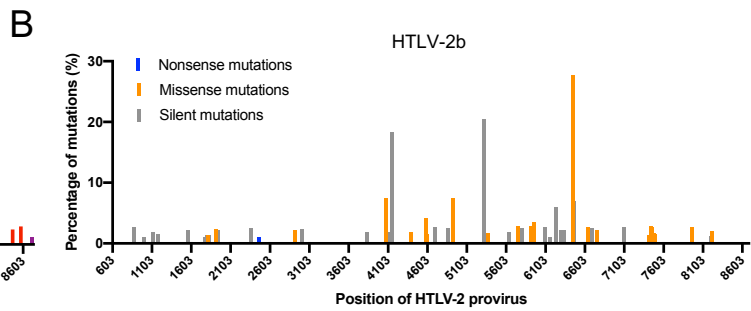
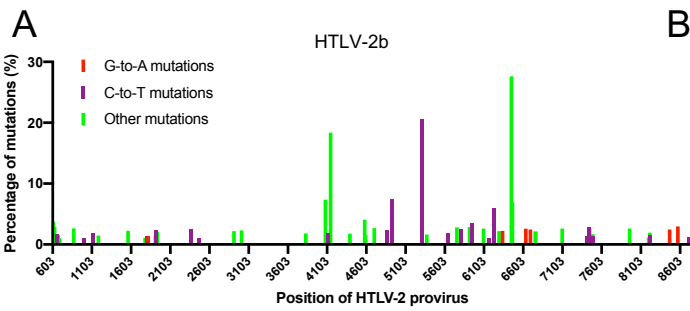
Figures S1 to S10

Tables S1 to S2



**Fig. S1. Deep sequencing of HTLV-1, HTLV-2, and STLV-1 proviruses *in vivo*.**

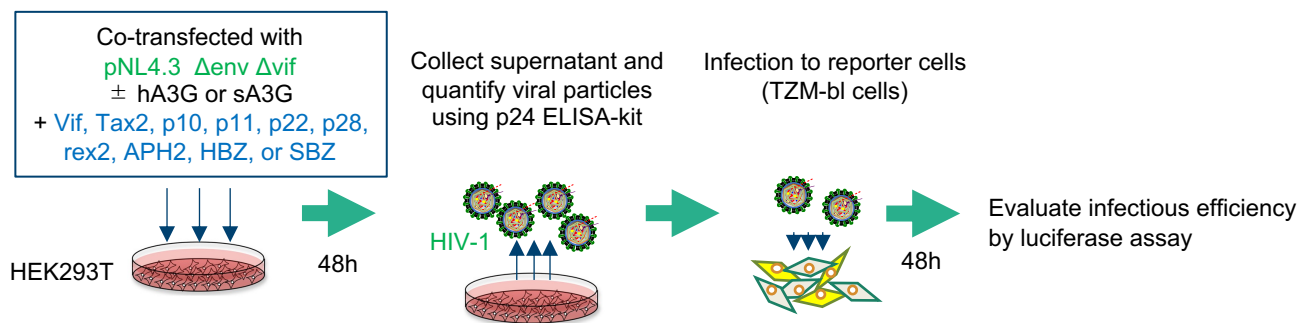
(A) Circular cladogram showing the HTLV-1 subgroups (green, Japanese subgroup [n = 53]; blue, transcontinental subgroup [n = 12]). Reference strains are indicated in the tree: ATK\_J02029, YS\_U119949, and AB513134 are Japanese subgroup; TSP1\_M86840 and BOI\_L36905 are transcontinental subgroup. (B-D) Depth (top) and the percentage of targeted bases covered by  $\leq 200\times$ ,  $> 200\times$ ,  $> 500\times$ ,  $> 1000\times$ ,  $> 1500\times$  and  $> 2000\times$  sequencing reads (bottom) from deep sequencing analysis of the provirus in HTLV-1 asymptomatic carriers (ACs) (n = 65) (B), HTLV-2-infected individuals (n = 18) (C), and STLV-1-infected Japanese macaques (n = 16) (D). (E) Example of mutations in HTLV-1 asymptomatic carriers (ACs; Japanese subgroup, n = 10; transcontinental subgroup, n = 4), HTLV-2-infected individuals (HTLV-2a, n = 3; HTLV-2b, n = 3), and STLV-1-infected Japanese macaques (n = 5), analyzed by HYPERMUT(<https://www.hiv.lanl.gov/content/sequence/HYPERMUT/hypermut.html>). Reference sequences were AB513134 for HTLV-1 Japanese subgroup, L36905 for HTLV-1 transcontinental subgroup, NC\_001488 for HTLV-2a, L20734 for HTLV-2b and MH542226 for STLV-1. (F-H) Heatmaps of mutations observed in the *tax* gene (F), *rex* gene (G), and *HBZ* gene (H) in 65 HTLV-1 ACs.



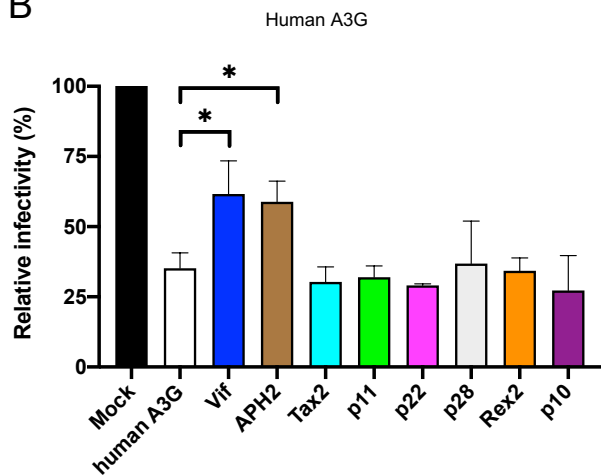
**Fig. S2. Deep sequencing of HTLV-1, HTLV-2, and STLV-1.**

(A) Frequency of mutations detected in the provirus of five HTLV-2b-infected individuals by deep sequencing analysis. (B) Frequency of nonsense mutations detected in the provirus of five HTLV-2b-infected individuals by deep sequencing analysis. (C) Schema of wild-type and mutants with a nonsense mutation (Q87\*) in the *APH-2* gene in two cases (#9 and #991). Frequencies of provirus mutations in these two HTLV-2-infected individuals (#9 and #991): (top) G-to-A mutations and (bottom) nonsense mutations. (D) Immunoblot showing the expression of APOBEC families in transfected HEK293T cells. IB, immunoblot. (E) Frequency of mutations detected in the provirus of HTLV-1-infected cells (left) or HTLV-2-infected cells (right) with hA3C/D/F/H expression *in vitro*. (F) Frequency of mutations detected in HTLV-1-infected cells under hA3G expression *in vitro* (top) corresponds with that in more than 10% of HTLV-1 ACs (n=65) *in vivo* (bottom). (G) Frequency of mutations detected in HTLV-1-infected cells (top) and STLV-1-infected cells (bottom) under E259Q mutant hA3G expression *in vitro*.

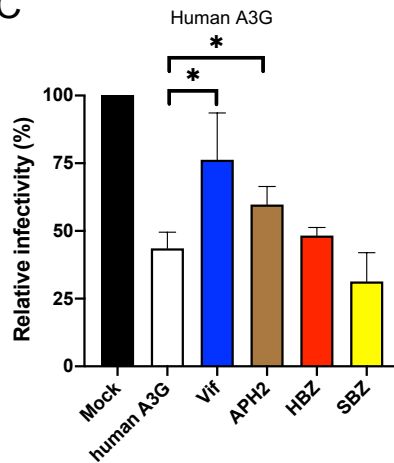
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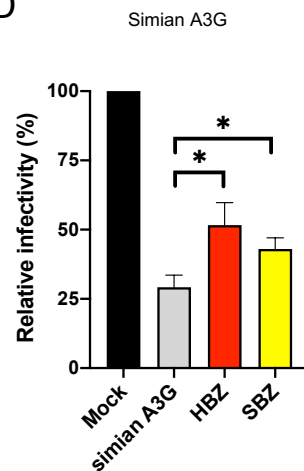
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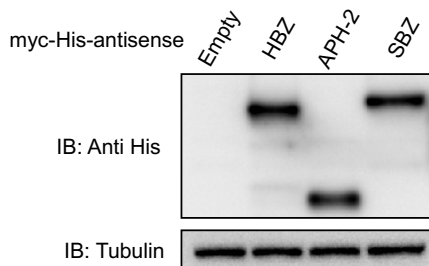
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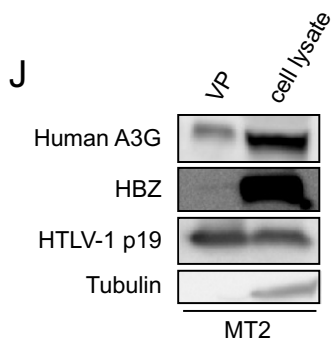
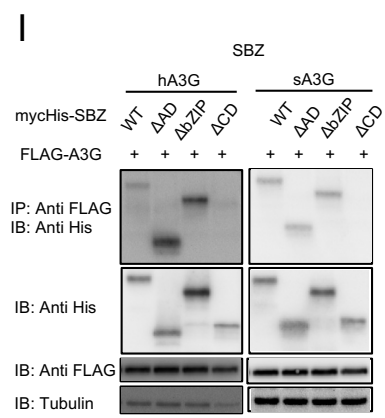
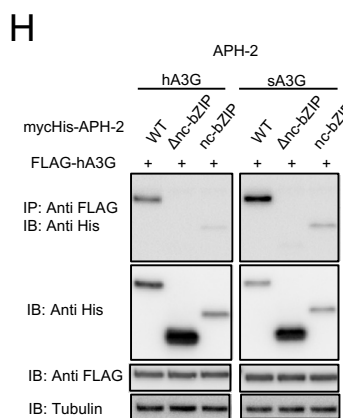
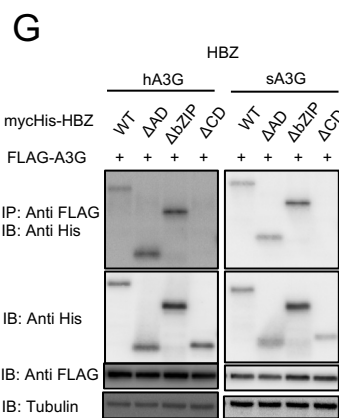
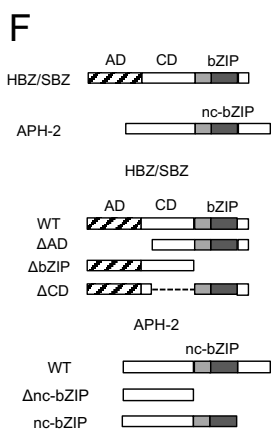
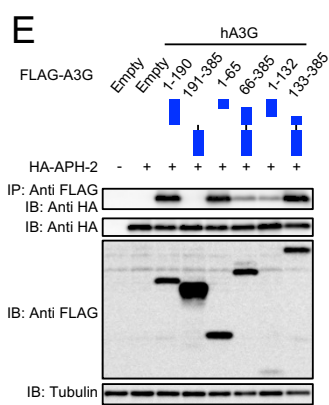
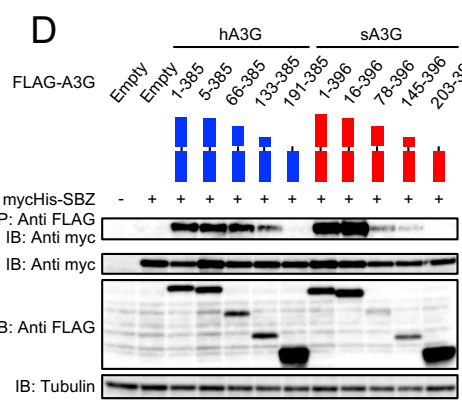
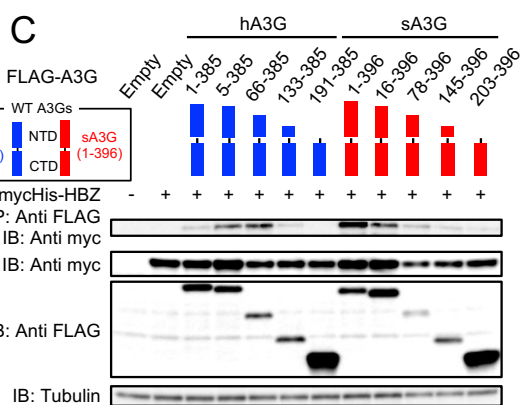
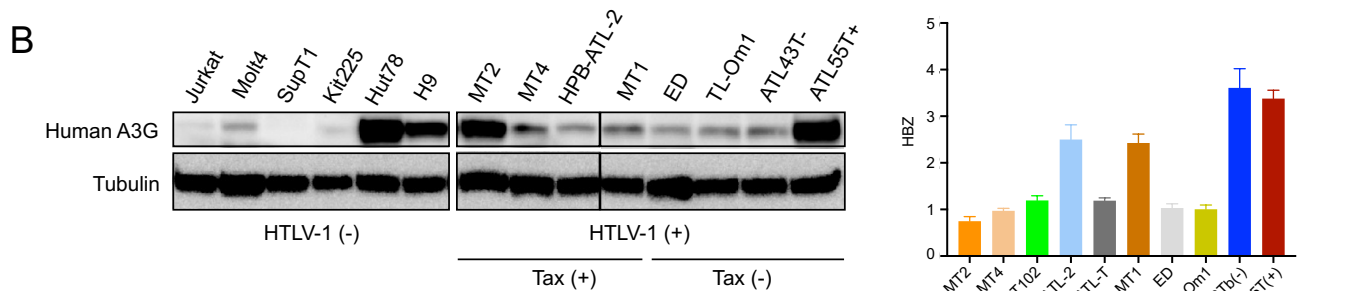
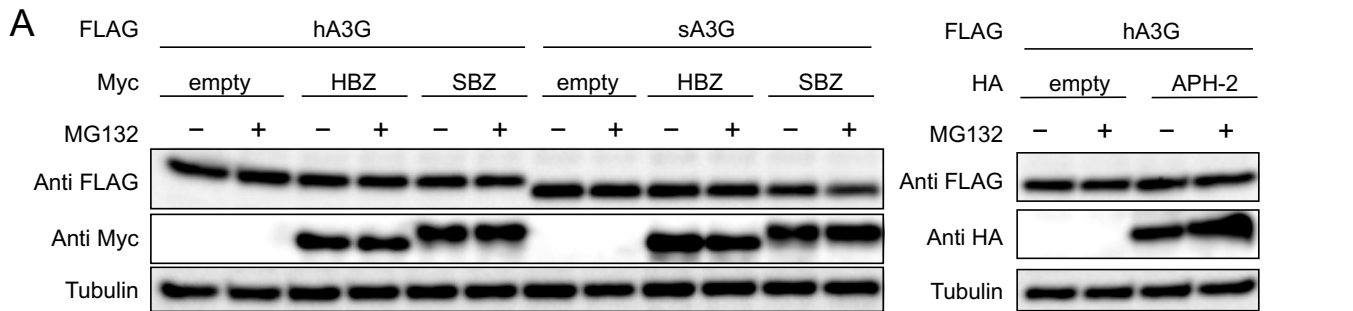


E



### Fig. S3. Antisense proteins suppress A3G.

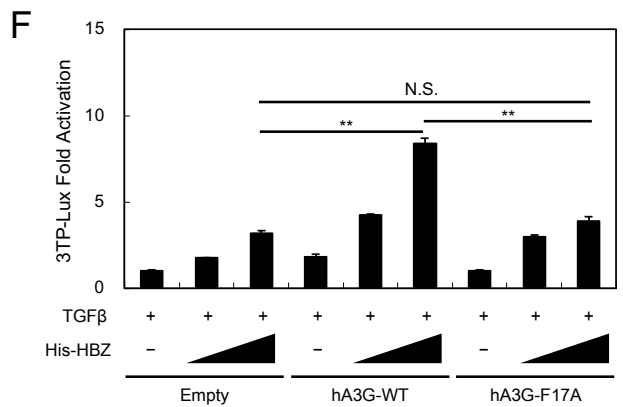
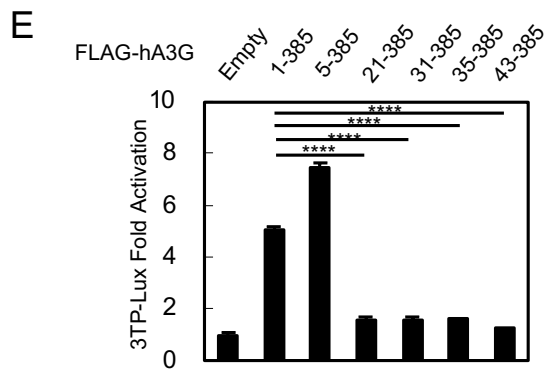
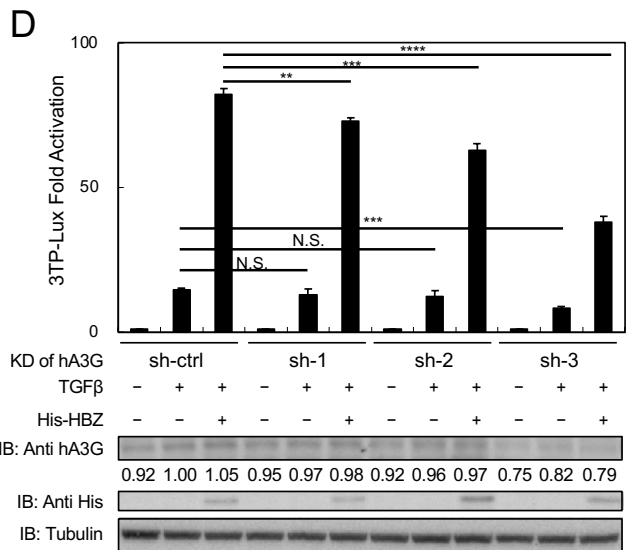
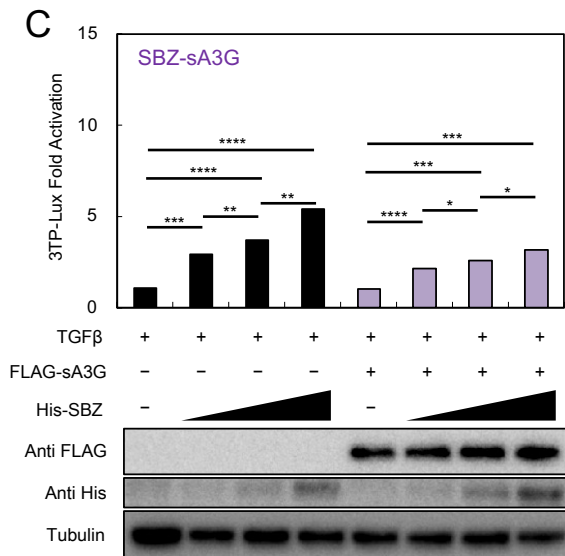
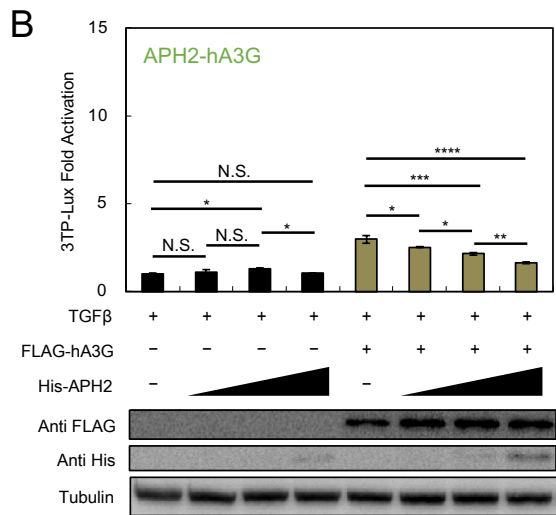
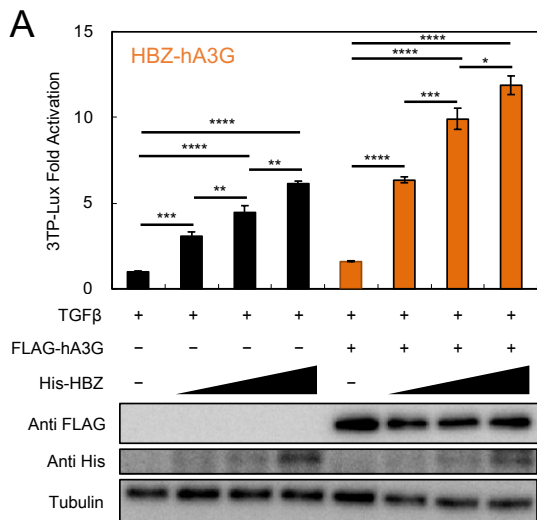
(A) Schema of the experiment to analyze the effects of HTLV-2 encoded genes and anti-sense viral proteins on viral infectivity in the presence of A3G using TZM-bl cells. Pseudotype recombinant HIV-1 (pNL4.3/ $\Delta$ Vif/ $\Delta$ Env) is produced using 293T cells expressing A3G and various viral genes. Vif is used as a positive control. (B) Among all the HTLV-2 coding proteins tested, only APH2 restored viral infectivity in the presence of hA3G (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \* $P < 0.05$ ). (C) Of the antisense proteins of HTLV-2, HTLV-1, or STLV-1, APH2 best inhibits human A3G and restores infectivity (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \* $P < 0.05$ ). (D) The antisense proteins of HTLV-1 or STLV-1 can inhibit simian A3G and restore infectivity (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \* $P < 0.05$ ). (E) Immunoblotting showing the antisense proteins purified from QT6 cells overexpressing HBZ, APH-2, or SBZ. All experiments were performed at least twice.



**Fig. S4. Interaction between A3G and antisense viral proteins.**

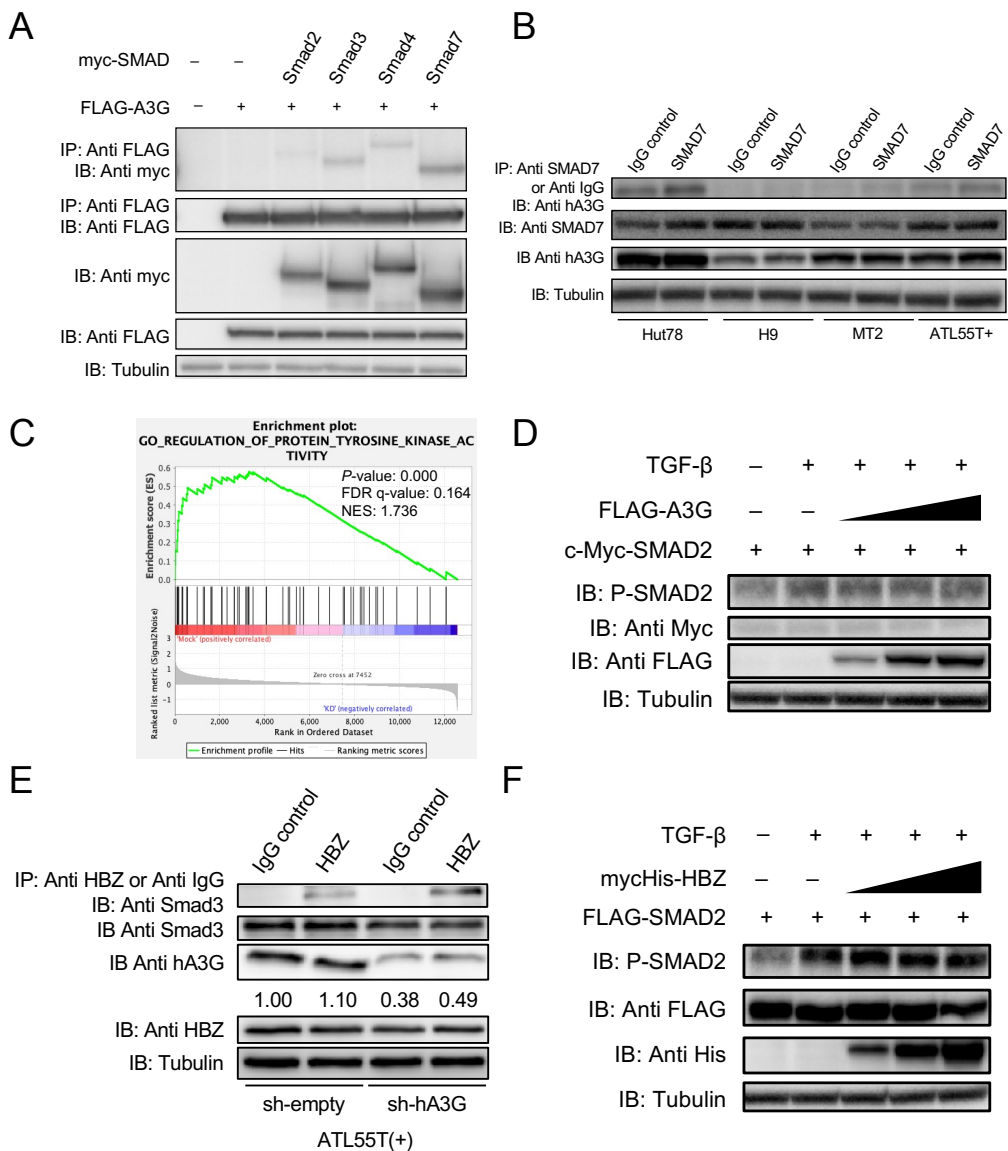
(A) Immunoblot showing that the amount of A3G is unchanged in the presence or absence of antisense proteins with or without MG132 treatment. (B) Immunoblot showing the expression of endogenous hA3G in HTLV-1-negative T-cell lines (Jurkat, Molt4, SupT1, Kit225, Hut78, and H9), HTLV-1-infected T-cell lines (MT-2, and MT-4), and ATL cell lines (HPB-ATL-2, MT-1, ED, TL-Om1, ATL43Tb(-), and ATL55T+)(left). Expression of mRNA of *HBZ* in HTLV-1-infected T-cell lines (MT-2, MT-4, and Hut102), and ATL cell lines (HPB-ATL-2, HPB-ATL-T, MT-1, ED, TL- Om1, ATL43Tb(-), and ATL55T(+)) by RT-qPCR (right). (C-E) Co-immunoprecipitation experiments showing the interaction of deletion mutants of A3Gs with HBZ (C), SBZ (D) and APH-2 (E) in transfected HEK293T cells. IP, immunoprecipitation; IB, immunoblot. (F) Schema of wild-type and deletion mutants of HBZ/SBZ and APH-2. (G-I) Co-immunoprecipitation experiment showing the interaction of A3Gs with deletion mutants of HBZ (G), APH-2 (H) and SBZ (I) in transfected HEK293T cells. IP, immunoprecipitation; IB, immunoblot. (J) Immunoblot showing the incorporation of both hA3G and HBZ into viral particles in MT-2 cells. Experiments were performed at least twice.





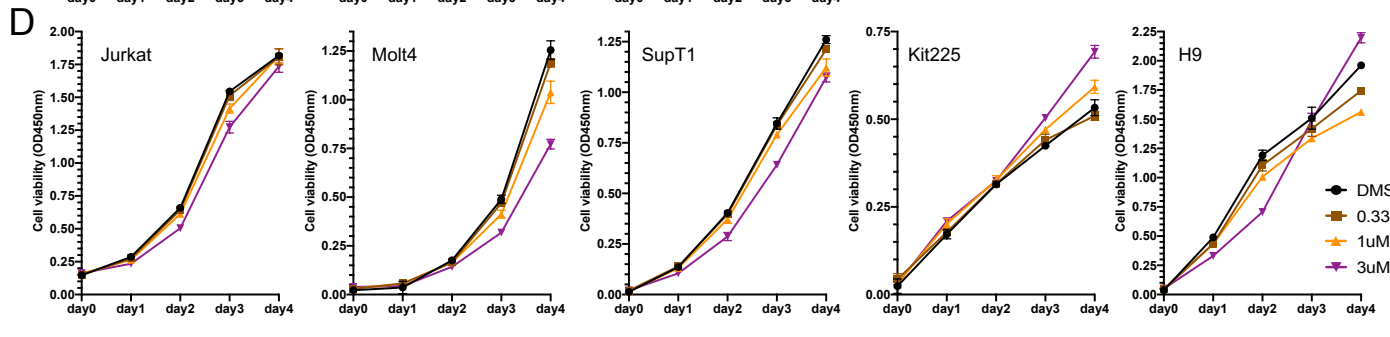
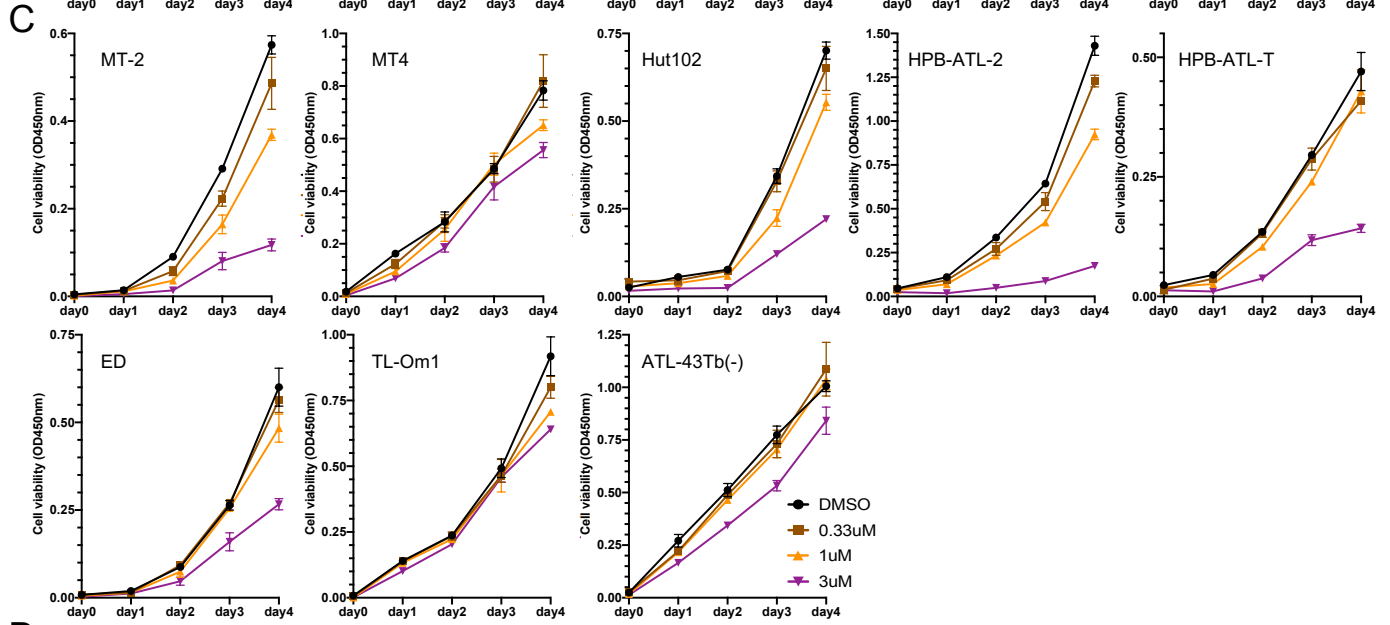
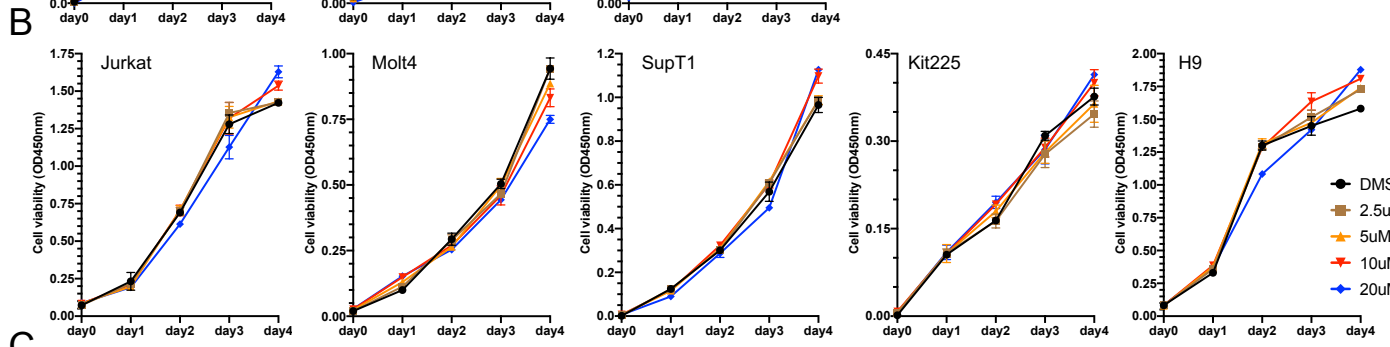
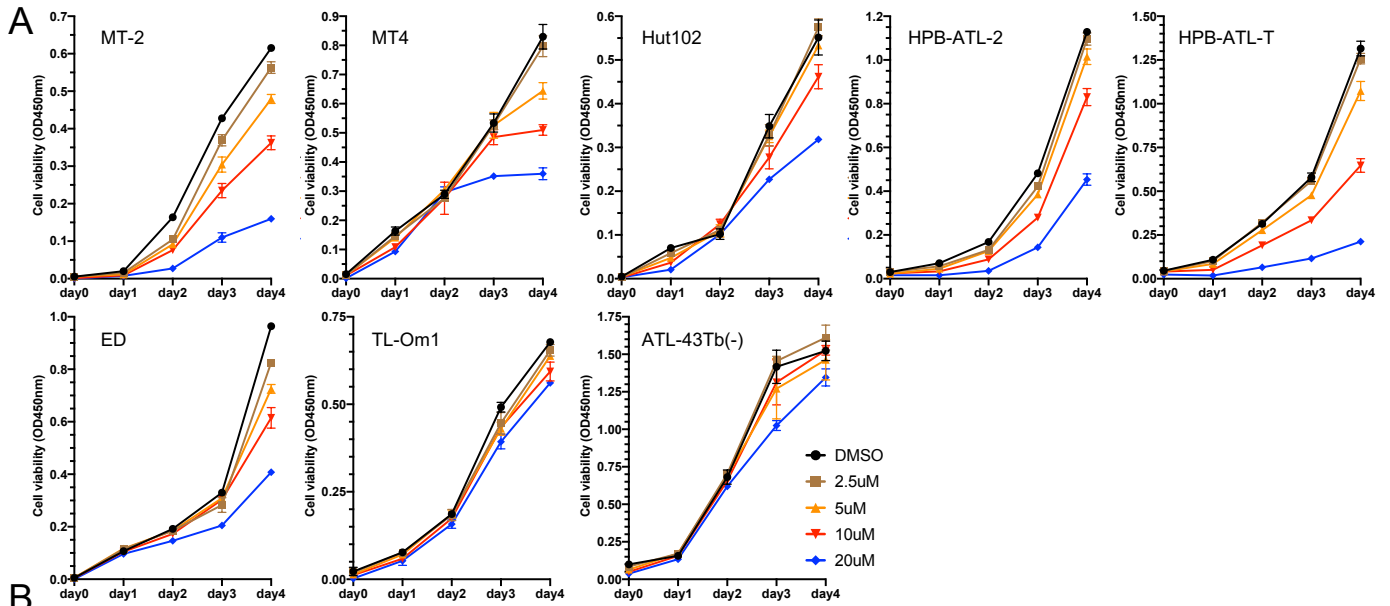
**Fig. S5. hA3G enhances the activation of the TGF- $\beta$ /Smad pathway by HBZ.**

(A-C) Luciferase activity of 3TP-Lux under the control of a TGF- $\beta$  responsive element in cells co-expressing hA3G with HBZ (A), hA3G with APH-2 (B), or sA3G with SBZ (C) (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001; \*\*\*\*P < 0.0001). (D) Luciferase activity of 3TP-Lux under the control of a TGF- $\beta$  responsive element in hA3G knockdown HepG2 cells expressing HBZ (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \*\*P < 0.01; \*\*\*P < 0.001; \*\*\*\*P < 0.0001). The hA3G expression values were analyzed by ImageJ. (E) Luciferase activity of 3TP-Lux under the control of a TGF- $\beta$  responsive element in cells expressing deletion mutants of hA3G (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \*\*\*\*P < 0.0001). (F) Luciferase activity of 3TP-Lux under the control of a TGF- $\beta$  responsive element in cells co-expressing HBZ with empty, hA3G-WT, or hA3G-F17A mutant (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \*\*P < 0.01). All experiments were performed at least twice.



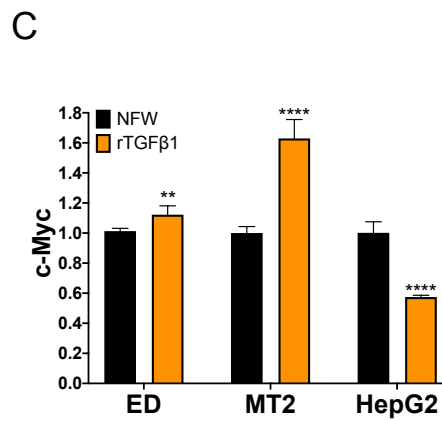
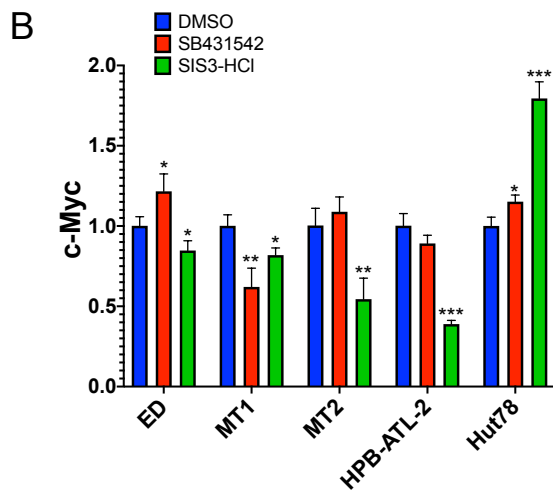
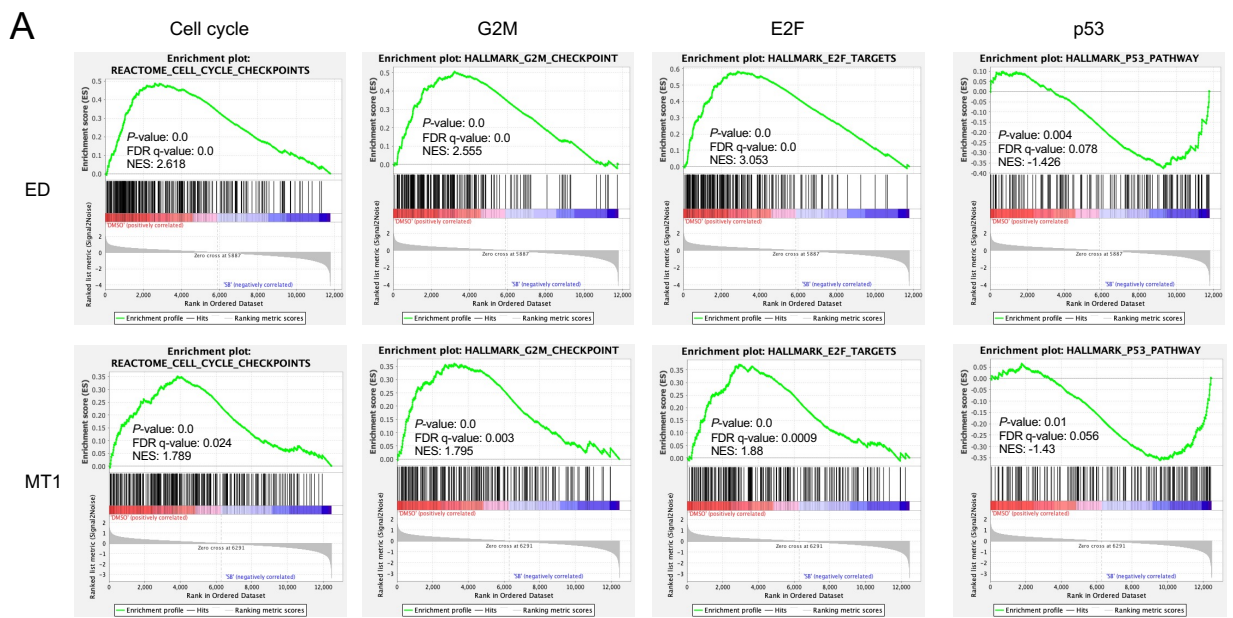
**Fig. S6. Interaction of hA3G with Smad proteins.**

(A) Co-immunoprecipitation experiment showing the interaction of hA3G with Smad2/3/4/7 in transfected HEK293T cells. IP, immunoprecipitation; IB, immunoblot. (B) Interaction between hA3G and endogenous Smad7 is analyzed by immunoprecipitation assay in HTLV-1-negative T-cell lines (Hut78, H9), an HTLV-1-infected T-cell line (MT-2), and an ATL cell line (ATL55T+). IP, immunoprecipitation; IB, immunoblot. (C) GSEA with RNA-seq showing a category of significantly enriched gene signatures related to protein tyrosine kinase activity in hA3G knockdown ATL cells. (D) Immunoblot analysis reveals no dose-dependent phosphorylation of Smad2 under expression of hA3G in transfected HepG2 cells. (E) Endogenous interaction between HBZ and Smad3 in ATL55T(+) and hA3G-knockdowned ATL55T(+) cells is analyzed by immunoprecipitation assay. IP, immunoprecipitation; IB, immunoblot. The hA3G expression values were analyzed by ImageJ. (F) Immunoblot analysis reveals no dose-dependent phosphorylation of Smad2 under expression of HBZ in transfected HepG2 cells. Experiments were performed at least twice (A-B, D, and F).

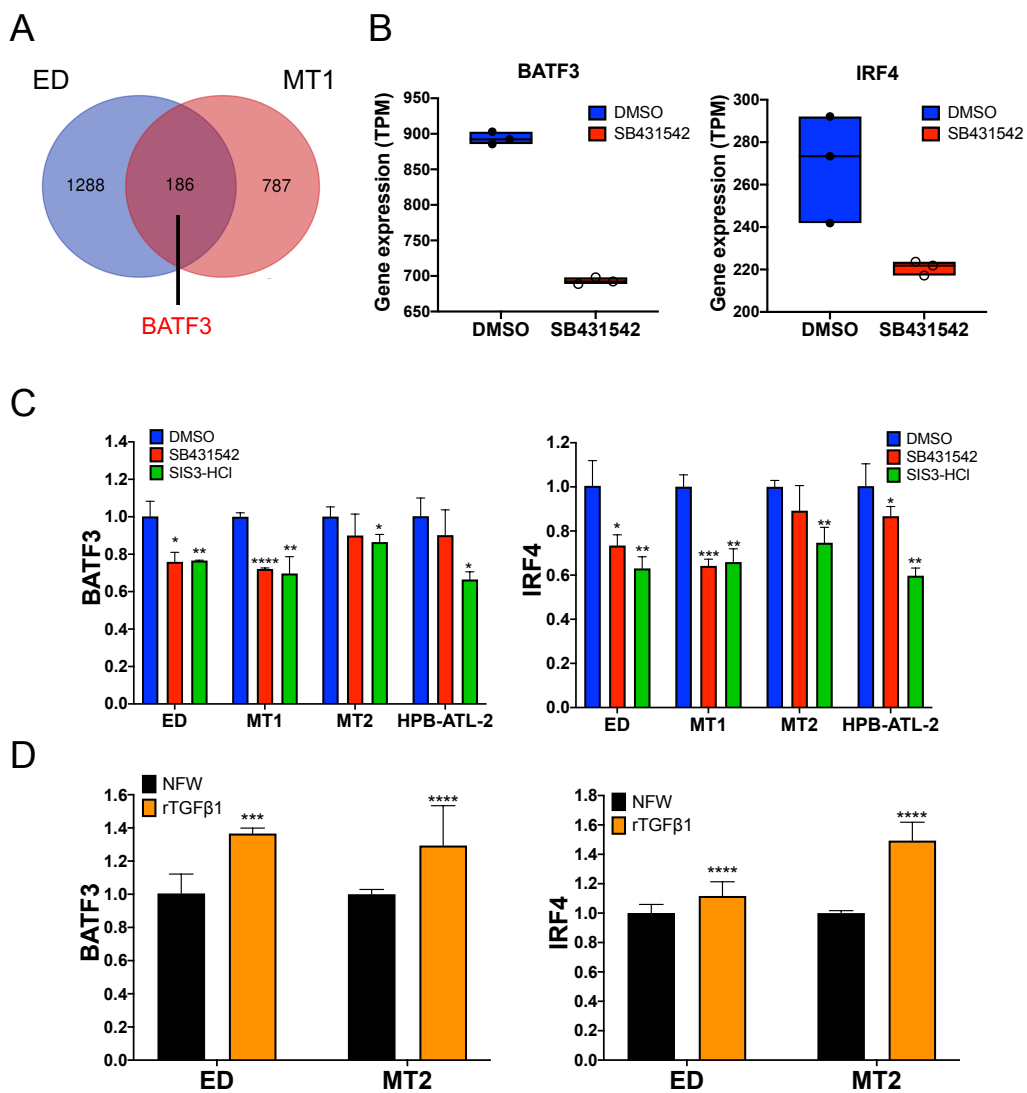


**Fig. S7. Cell growth assays in HTLV-1-non-infected and -infected T-cell lines.**

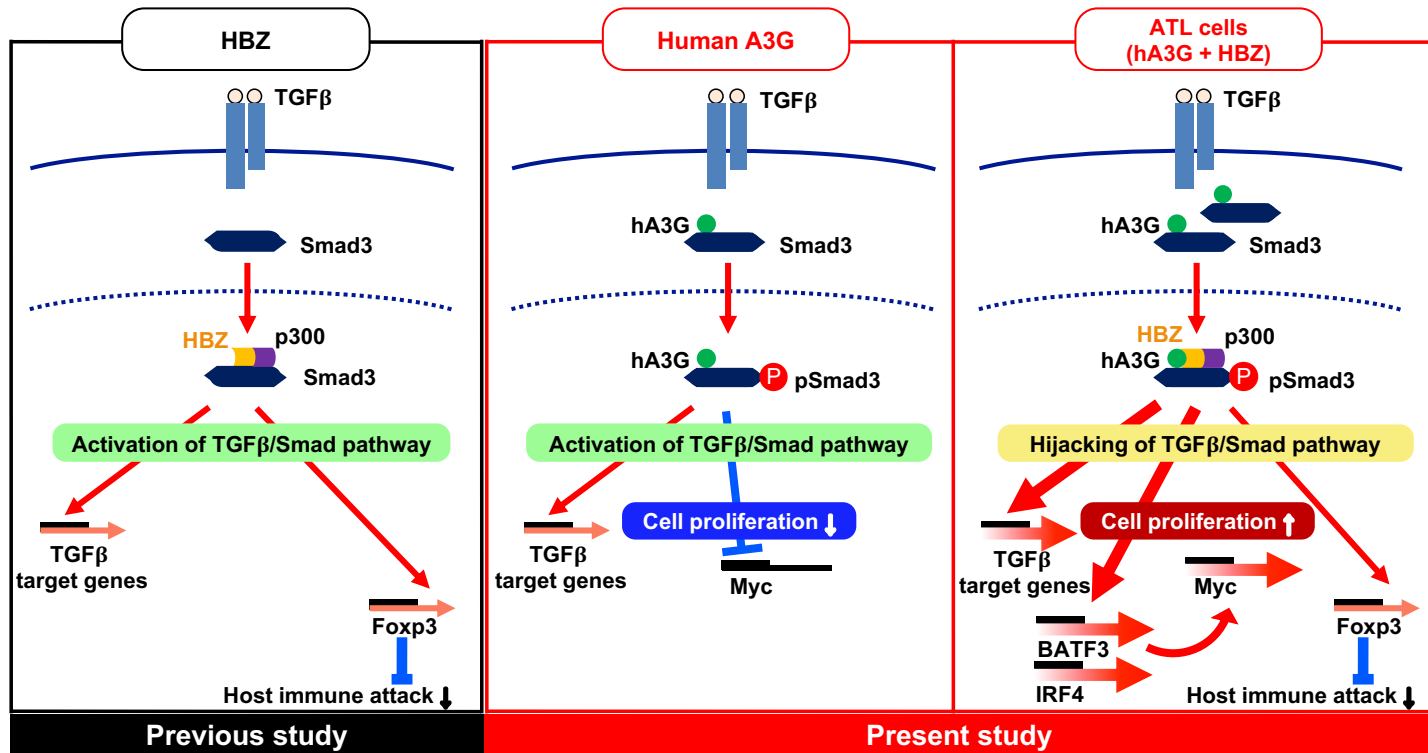
(A) Cell growth of HTLV-1-infected T-cell lines/ATL cell lines treated with SB431542 (normalized mean  $\pm$  s.d. of triplicate experiments). (B) Cell growth of HTLV-1-non-infected T-cell lines treated with SB431542 (normalized mean  $\pm$  s.d. of triplicate experiments). (C) Cell growth of HTLV-1-infected T-cell lines/ATL cells lines treated with SIS3-HCl (normalized mean  $\pm$  s.d. of triplicate experiments). (D) Cell growth of HTLV-1-non-infected T-cell lines treated with SIS3-HCl (normalized mean  $\pm$  s.d. of triplicate experiments).



**Fig. S8. MYC is upregulated by the TGF- $\beta$ /SMAD pathway in HTLV-1-infected cells.** (A) Significantly enriched gene signatures for cell cycle related gene sets in ED cells (top) and MT-1 cells (bottom) treated with SB431542, by GSEA with RNA-seq. (B) *c-Myc* expression in an HTLV-1-negative T-cell line (Hut78), an HTLV-1-infected T-cell line (MT-2), and ATL cell lines (ED, MT1, HPB-ATL-2) treated with SB431542 or SIS3-HCl, by RT-qPCR (normalized mean  $\pm$  s.d. of triplicate experiments; one-way ANOVA with Tukey correction; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ ). (C) *c-Myc* expression in cell lines treated with or without recombinant TGF- $\beta$ 1 (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \*\* $P < 0.01$ ; \*\*\*\* $P < 0.0001$ ). Experiments were performed at least twice (B-C).



**Fig. S9. BATF3 and IRF4 are upregulated by activation of the TGF- $\beta$ /SMAD pathway.** (A), Venn diagram of differentially expressed genes (DEG) by RNA-seq in ED cells and MT-1 cells treated with SB431542. (B), Transcripts per million (TPM) of *BATF3* (left) and *IRF4* (right) expression in MT-1 cells treated with SB431542, analyzed by RNA-seq. (C), RT-qPCR showing *BATF3* (left) and *IRF4* (right) expression in an HTLV-1-infected T-cell line (MT-2) and ATL cell lines (ED, MT1, HPB-ATL-2) treated with SB431542 or SIS3-HCl (normalized mean  $\pm$  s.d.; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ ; \*\*\*\* $P < 0.0001$ ). (D), RT-qPCR showing *BATF3* (left) and *IRF4* (right) expression in ED cells and MT-2 cells treated with or without recombinant TGF- $\beta$ 1 (normalized mean  $\pm$  s.d. of triplicate experiments; two-tailed unpaired Student's t test; \*\*\* $P < 0.001$ ; \*\*\*\* $P < 0.0001$ ). Experiments were performed at least twice (C-D).



**Fig. S10. Schema of HBZ/hA3G-induced ATL cell proliferation.**

In ATL cells, HBZ and hA3G potentiate the TGF-β/Smad signaling. Enhanced TGF-β/Smad signaling through HBZ/hA3G promotes the proliferation of ATL cells by upregulation of *BATF3/IRF4* and *MYC*.

**Fig. S10**



Table S1. Proviral loads in HTLV-1 asymptomatic carriers, HTLV-2-infected individuals, and STLV-1-infected Japanese macaques

Cases	PVL (%)	Cases	PVL (%)	Cases	PVL (%)
HTLV-1 AC1	5.4	HTLV-1 AC34	0.5	HTLV-2#3	25.7
HTLV-1 AC2	8.2	HTLV-1 AC35	0.2	HTLV-2#10	0.24
HTLV-1 AC3	0.5	HTLV-1 AC36	6.8	HTLV-2#5	0.009
HTLV-1 AC4	0.1	HTLV-1 AC37	0.8	HTLV-2#6	1.545
HTLV-1 AC5	24.9	HTLV-1 AC38	0.03	HTLV-2#7	0.867
HTLV-1 AC6	1.2	HTLV-1 AC39	8.0	HTLV-2#8	8.927
HTLV-1 AC7	2.5	HTLV-1 AC40	12.5	HTLV-2#9	16.38
HTLV-1 AC8	20.0	HTLV-1 AC41	0.4	HTLV-2#1791	NA
HTLV-1 AC9	10.5	HTLV-1 AC42	1.3	HTLV-2#2204	NA
HTLV-1 AC10	4.5	HTLV-1 AC43	5.3	HTLV-2#1622	NA
HTLV-1 AC11	9.3	HTLV-1 AC44	0.9	HTLV-2#1890	NA
HTLV-1 AC12	6.1	HTLV-1 AC45	12.4	HTLV-2#1875	NA
HTLV-1 AC13	3.8	HTLV-1 AC46	3.1	HTLV-2#1009	NA
HTLV-1 AC14	4.6	HTLV-1 AC47	1.5	HTLV-2#991	NA
HTLV-1 AC15	1.5	HTLV-1 AC48	0.2	HTLV-2#1975	NA
HTLV-1 AC16	91.3	HTLV-1 AC49	5.5	HTLV-2#1073	NA
HTLV-1 AC17	4.2	HTLV-1 AC50	3.8	HTLV-2#416	NA
HTLV-1 AC18	1.2	HTLV-1 AC51	2.5	STLV1-1	1.3
HTLV-1 AC19	10.0	HTLV-1 AC52	3.2	STLV1-2	1.5
HTLV-1 AC20	14.1	HTLV-1 AC53	3.5	STLV1-3	5.9
HTLV-1 AC21	1.9	HTLV-1 AC54	7.3	STLV1-4	6.5
HTLV-1 AC22	0.2	HTLV-1 AC55	2.2	STLV1-5	9.8
HTLV-1 AC23	11.0	HTLV-1 AC56	2.4	STLV1-6	11.5
HTLV-1 AC24	1.3	HTLV-1 AC57	1.5	STLV1-7	11.6
HTLV-1 AC25	22.6	HTLV-1 AC58	0.3	STLV1-8	13.7
HTLV-1 AC26	7.7	HTLV-1 AC59	3.1	STLV1-9	14.1
HTLV-1 AC27	3.5	HTLV-1 AC60	1.2	STLV1-10	17.1
HTLV-1 AC28	1.9	HTLV-1 AC61	17.7	STLV1-11	53.2
HTLV-1 AC29	2.1	HTLV-1 AC62	0.6	STLV1-12	0.2
HTLV-1 AC30	10.4	HTLV-1 AC63	3.5	STLV1-13	0.4
HTLV-1 AC31	1.3	HTLV-1 AC64	7.8	STLV1-14	1.2
HTLV-1 AC32	6.1	HTLV-1 AC65	15.8	STLV1-15	3.3
HTLV-1 AC33	0.1	HTLV-2#1	6.0	STLV1-16	0.3

Abbreviation: HTLV-1, human T-cell leukemia virus type 1; HTLV-2, human T-cell leukemia virus type 2; STLV-1, simian T-cell leukemia virus type 1; AC, asymptomatic carrier; PLV, proviral load; NA, not available.

Table S2. Primers for quantitative RT-PCR and deep-sequencing analysis

Gene/target		Sequence
Human APOBEC3G	F	5'-CCGAGGACCCGAAGGTAC-3'
	probe	5' FAM-ccaggagg-TAMRA 3'
	R	5'-TCCAACAGTGCTGAAATTCG-3'
SMAD2	F	5'-CTCAAGGCAATTGAAAAGTGGC-3'
	R	5'-GGCGGAAGTTCTGTTAGG-3'
SMAD3	F	5'-GTGACCACCAGATGAACCAC-3'
	R	5'-GTAGTAGGAGATGGAGCACC-3'
SMAD7	F	5'-CTGTGCCTTCCCTCCGCTG-3'
	R	5'-CACTCTCGTCTTCTCCTC-3'
BATF3	F	5'-CGGAAGAAGCAGACCCAGAA-3'
	R	5'-CATCTTCTCGTGCTCCTCAGT-3'
IRF4	F	5'-GACTTTGAGGAACTGGTTGAGC-3'
	R	5'-GTAAGGCGTTGTCATGGTGTAG-3'
MYC	F	5'-CCTGGTGCTCCATGAGGAGAC-3'
	R	5'-CAGACTCTGACCTTTGCCAGG-3'
HBZ	F	5'-CGACCTGAGCTTAAACTTACC-3'
	R	5'-GCCCGTCCACCAATTCCTCC-3'
HTLV-1 provirus	F	5'-TACGTCTTTGTTTTCTGTTCTGCGCCG-3'
	R	5'-AGAGCCGGCTGAGCTAGGTAGGCT-3'
HTLV-2a provirus	F	5'-CTCGGCTAGACTCTGCCTTAAACT-3'
	R	5'-TCGACCTGAGAGGAGACTTACCTT-3'
HTLV-2b provirus	F	5'-GTTCTTTCCTTTCGTCGTCAC-3'
	R	5'-GTGACGACGAAGAGAAAGAAC-3'
STLV-1 provirus	F	5'-CCGCTGCAGATCGAAAGTTCC-3'
	R	5'-AGAGCCGGCCGAATCTAGG-3'
pX1MT-M pX region	F	5'-GGCCTTACAACTGGAATCACC-3'
	R	5'-AGAGCCGGCTGAGCTAGGCAGGCT-3'
pH6neo pX region	F	5'-GAGGATTAGACCTCCTATTCTGGG-3'
	R	5'-CTTCCCCGGAAGACAATGC-3'
pWK1699 pX region	F	5'-GGCCTTACAACTGGAATCACC-3'
	R	5'-GAGAGAGTTGTAATGGGCTGCT-3'

Abbreviation: RT-PCR, reverse transcription polymerase chain reaction; F, forward; R, reverse; BATF3, basic leucine zipper ATF-like transcription factor 3; IRF4, interferon regulatory factor 4; HBZ, HTLV-1 bZIP factor.