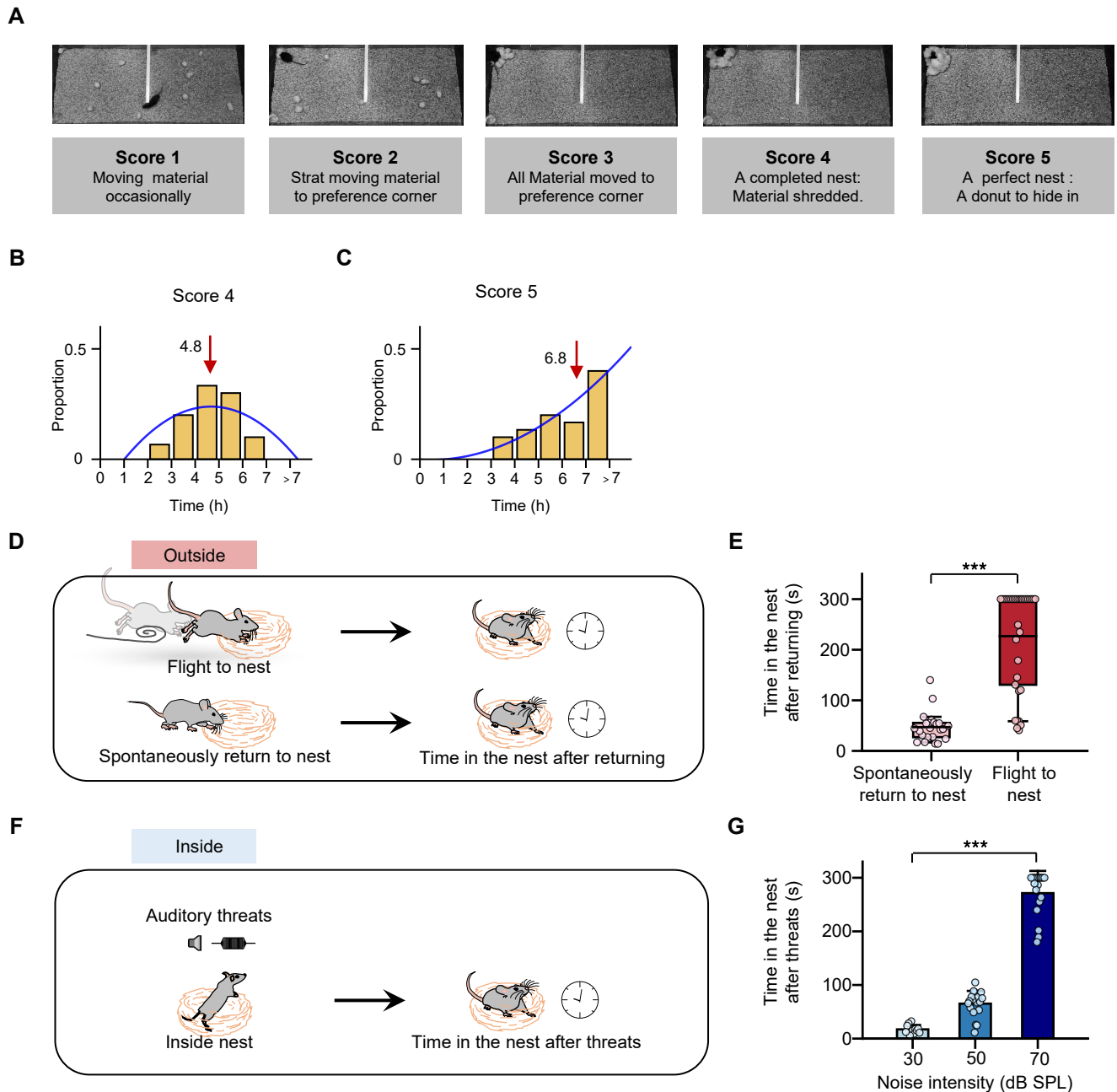


**Distinct Circuits in Anterior Cingulate Cortex Encode Safety Assessment and  
Mediate Flexibility of Fear Reactions**

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Supplementary Figures 1-7



**Figure S1. The scoring standards for nest-building and various behavioral parameters. Related to Figure 1.**

(A) Video frames of nest-building at five typical stages (top) and their scoring standards (bottom).

(B-C) Distribution of nest-building times (the time point when a mouse was placed in was hour 0) reaching the score 4 (B) or 5 (C) stage. Arrow indicates the mean value.

(D) Schematic of mice staying inside the self-built nest after threat-triggered returning to the nest.

(E) Time spent in the nest after a spontaneous return vs. after threat-triggered return. Mann-Whitney test ( $P < 0.001$ ,  $U = 49.00$ ).  $n = 23, 34$  bouts for spontaneous return and flight, respectively.

(F) Schematic of mice remaining in the self-built nest after exposure to auditory threats.

(G) Time spent in the nest after experiencing noise sounds at different intensities with the mouse initially inside the self-built nest. Kruskal-Wallis test ( $***P < 0.001$ ,  $H = 40.14$ ),  $n = 11, 19, 18$  trials for 30, 50, 70 dB SPL, respectively.



**Figure S2. The scoring standards for context-dependent behavioral responses to auditory threats and the behavioral summary for male and female mice. Related to Figure 1.**

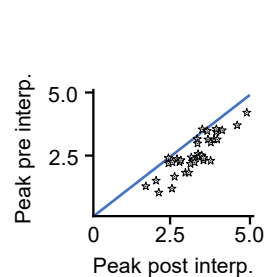
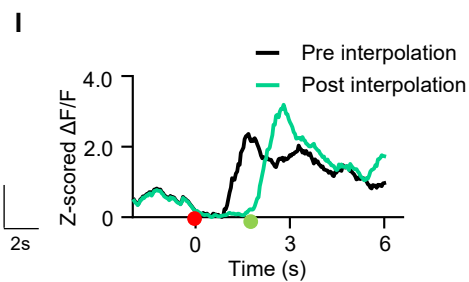
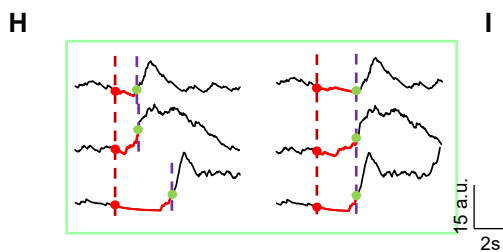
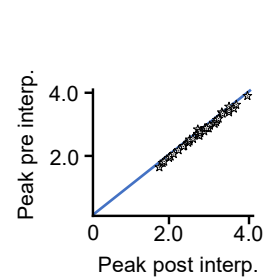
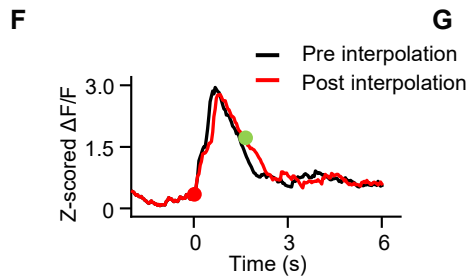
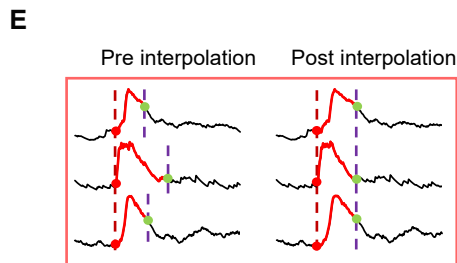
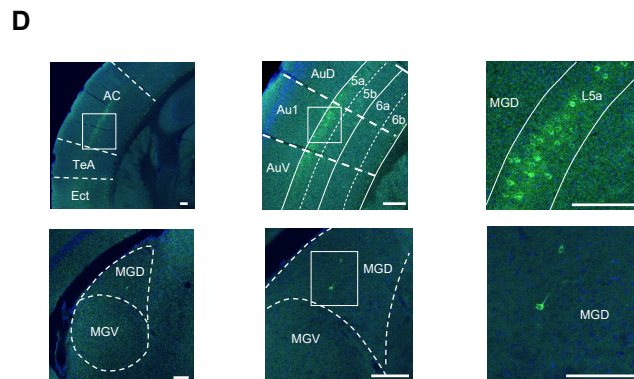
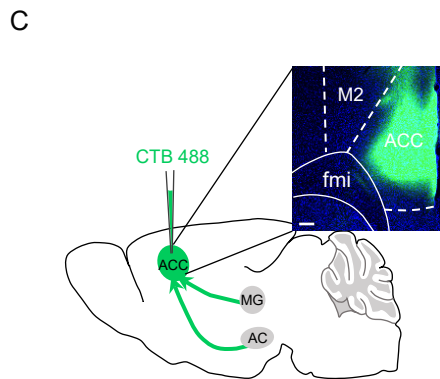
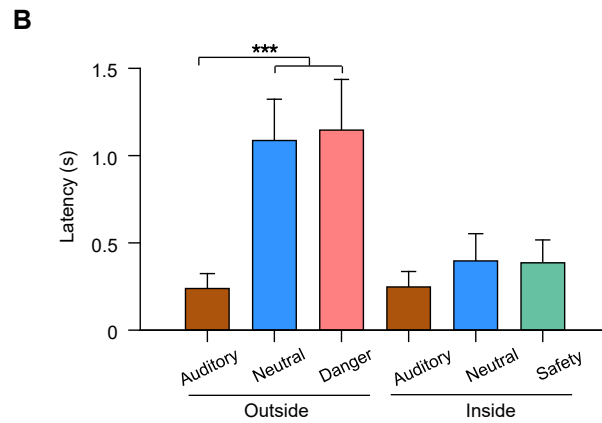
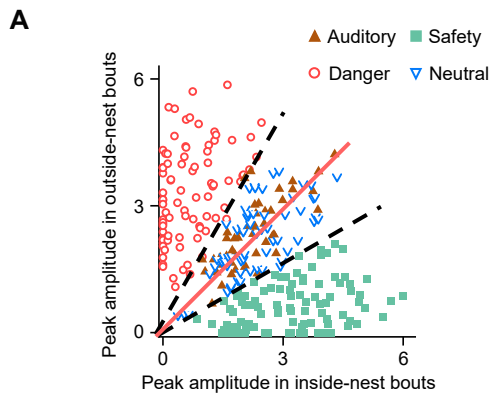
(A) The flow chart for scoring safety/danger-related behaviors both in outside- and inside-nest conditions.

(B) Illustration of the standards of safety scores from 1 to 7 in outside-nest bouts.

(C) Similar to (B), but for inside-nest bouts.

(D) Comparison of safety scores (left) and the distribution of context selectivity indices (CSIs) (right) in different scenarios between male (blue) and female mice (red). Kolmogorov-Smirnov test ( $P = 0.65$ ,  $D = 0.25$ ),  $n = 10$  female mice and 20 male mice.

(E) Similar to (D), but for the sham-nest conditions. Kolmogorov-Smirnov test ( $P = 0.81$ ,  $D = 0.33$ ),  $n = 10$  female mice and 15 male mice.



**Figure S3. The analysis of behavioral-event-related neurons. Related to Figure 2 and 3.**

(A) Peak amplitudes of  $\text{Ca}^{2+}$  responses of auditory-, danger-, safety-responding neurons and neutral cells in the outside- vs. inside-nest condition. Dashed black lines separate different groups of cells. Red line is the unity line.

(B) The average response latency for auditory-, danger-, safety-responding neurons and neutral cells. Kruskal-Wallis test ( $***P < 0.001$ ,  $H = 119.75$ ),  $n = 16, 44, 46, 45$  for auditory-, safety-, danger-responding and neutral cells, respectively.

(C) Schematic of retrograde tracing of inputs to ACC neurons from the medial geniculate body (MG) and auditory cortex (AC). Inset, example image showing the expression of Alexa Fluor 488-CTB (green) at the injection site.

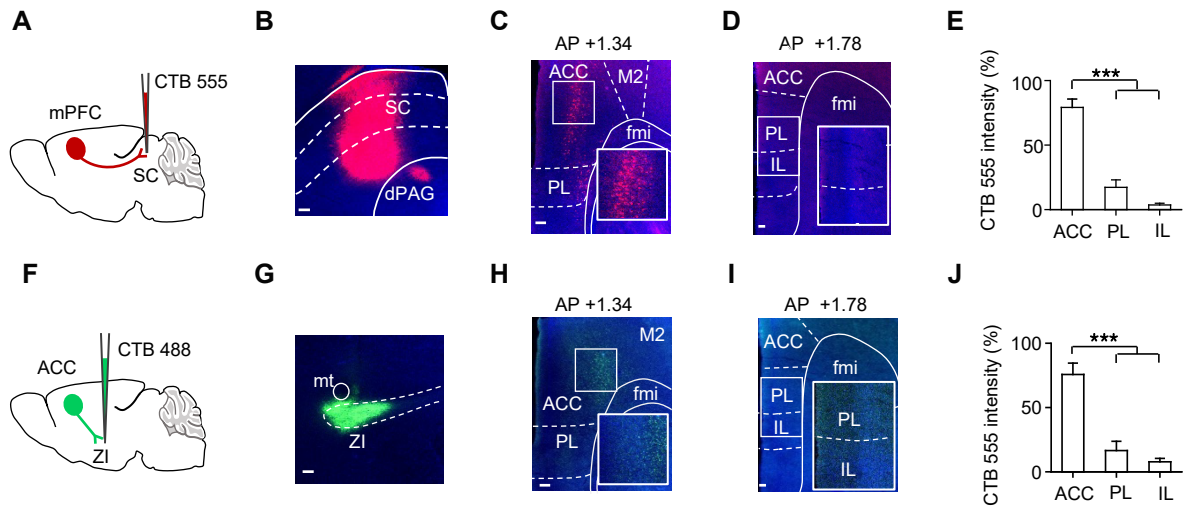
(D) Confocal images showing retrogradely labeled cells in AC and MG with zoom-in images of the boxed areas (right). Scale bar, 100  $\mu\text{m}$ .

(E) Example interpolation for a danger-responding neuron. The  $\text{Ca}^{2+}$  signal trace in the window from the onset of flight (marked by the red dashed line) to the timing of nest-entering (marked by the purple dashed line) of each trial (left panel) was interpolated into a 2 sec long waveform (right panel), and the rest of the waveform was unchanged.

(F) Comparison of average traces of pre- and post-interpolation for the cell shown in (E).

(G) Comparison of peak amplitudes of the average  $\text{Ca}^{2+}$  signal waveform during 0-2 sec after the flight onset between pre- and post-interpolation. Blue line is the unity line.

(H-J) Similar to (E-G), but for a safety-responding cell. Note that after interpolation, the peak amplitude of the average waveform became larger because of a better temporal alignment.



**Figure S4. SC- and ZI-projecting neurons in mPFC. Related to figure 4.**

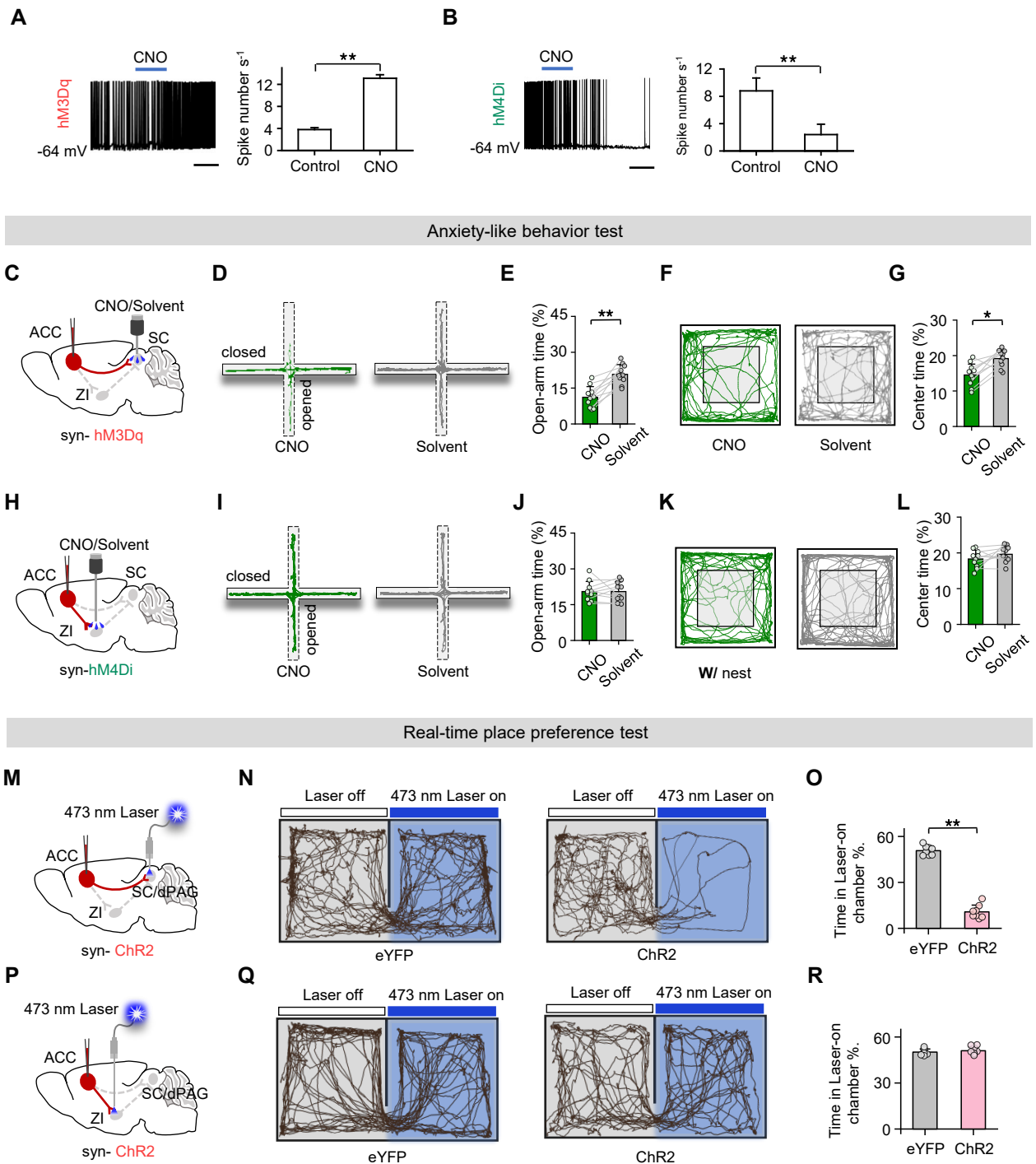
(A) Schematic of retrograde labeling of SC-projecting neurons in mPFC.

(B) CTB 555 fluorescence at the injection site of SC. Scale bar, 100 μm.

(C-D) Retrogradely labeled SC-projecting neurons in the center section of ACC (AP +1.34 mm) (C), PL and IL (AP +1.78 mm) (D), respectively. Inset, zoom-in image of the boxed area. Blue, DAPI staining. Scale bar, 100 μm.

(E) Relative intensity of CTB labeling in ACC, PL and IL. Kruskal-Wallis test ( $***P < 0.001$ ,  $H = 23.14$ ),  $n = 9$  brain slices from 3 mice.

(F-J) similar to (A-E), but for ZI-projecting neurons labeled by CTB 488. Kruskal-Wallis test ( $***P < 0.001$ ,  $H = 20.43$ ),  $n = 9$  brain slices from 3 mice.





**Figure S5. Validation of the efficacy of chemo/optogenetic manipulations and associated behavioral side effects. Related to figure 6.**

(A) Left, raw recorded trace showing an increase of firing rate in a hM3Dq-expressing ACC neuron after CNO application. Whole-cell current-clamp recording was performed in the slice preparation. Right, average firing rates before and after perfusion of CNO. Temporal scale, 3 min. Wilcoxon signed rank test (\*\*P = 0.004, W = 45.00), n = 9 brain slices from 3 mice.

(B) Similar to (A), but for verifying the effectiveness of CNO application in suppressing hM4Di-expressing neurons in ACC. Wilcoxon signed rank test (\*\*P = 0.004, W = 45.00), n = 9 brain slices from 3 mice.

(C) Schematic of chemogenetic activation of ACC axon terminals in the SC.

(D) Movement tracks of an example mouse in the EPM test with (left) and without (right) chemogenetic activation.

(E) Percentage of time spent in open arms of EPM. Two-tailed Wilcoxon matched-pairs signed rank test (CNO vs. Solvent: \*\*P = 0.002, W = 55.00, n = 10 mice each for CNO and Solvent groups).

(F) Movement tracks of an example mouse in the OF test.

(G) Percentage of time spent in the center of OF. Two-tailed Wilcoxon matched-pairs signed rank test (CNO vs. Solvent: \*P = 0.014, W = 51.00, n = 10 mice each for CNO and Solvent groups).

(H-L) Similar to (C-G), but for chemogenetic inhibition of ACC axon terminals in ZI. (J) Two-tailed Wilcoxon matched-pairs signed rank test (CNO vs. Solvent: P = 0.63, W = 22.00, n = 10 mice each for CNO and Solvent).

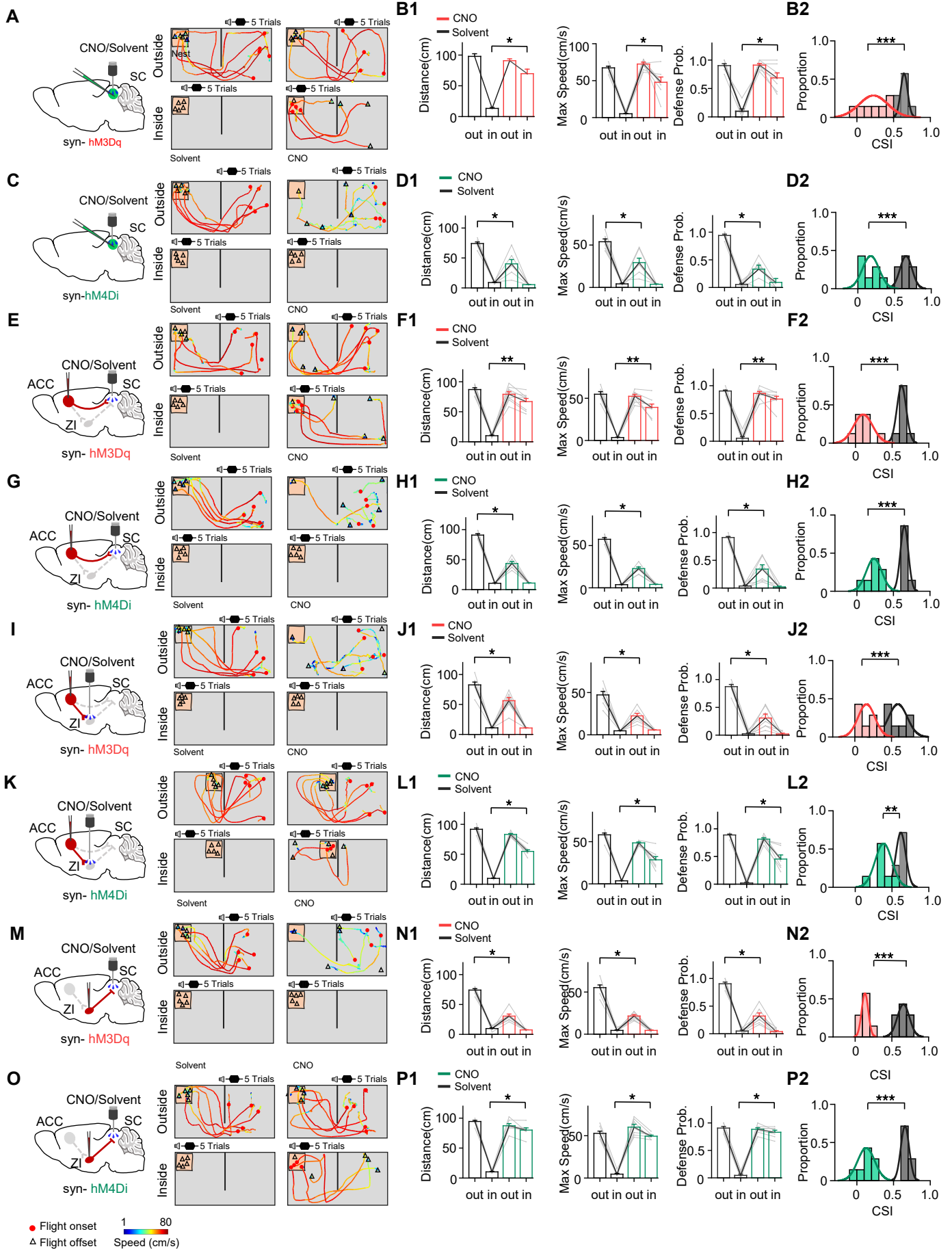
(L) Two-tailed Wilcoxon matched-pairs signed rank test (CNO vs. Solvent: P = 0.23, W = 15.00, n = 10 mice each for CNO and Solvent).

(M) Schematic of optogenetic activation of ACC axon terminals in the SC/dPAG.

(N) Locomotion tracking for an example animal in the eYFP control or ChR2 group during a 10-min session of RTPP test with optogenetic activation of ACC glutamatergic axon terminals in SC/dPAG.

(O) Comparison of percentage time spent in the laser-on chamber between the eYFP (n = 8 mice) and ChR2 (n = 8 mice) groups. Wilcoxon signed rank test (\*\*P = 0.003, W = 36.00).

(P-R) Similar to (M-O), but for optogenetic activation of ACC glutamatergic axon terminals in ZI. Wilcoxon signed rank test (P = 0.56, W = 28.00, n = 7 mice for both eYFP and ChR2 groups).



**Figure S6. Detailed experimental results for chemogenetic manipulations of ACC→SC and ACC→ZI→SC pathways. Related to figure 6.**

**(A)** Left, schematic of chemogenetic activation of SC. Middle and right, trajectory-speed map for an example mouse under two scenarios (top, outside-nest bouts; bottom, inside-nest bouts) with solvent (middle) or CNO (right) infusion in SC.

**(B1)** Comparison of the total flight distance (left), maximum speed (middle) and defense probability (right) under two scenarios between solvent and CNO groups, respectively. Wilcoxon signed rank test (for distance, \*P = 0.016, W = 28.00; for max. speed, \*P = 0.016, W = 28.00; for flight probability, \*P = 0.016, W = 28.00).

**(B2)** Distribution of CSIs without (grey) and with (color) chemogenetic activation. Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 7 mice.

**(C-D)** Similar to **(A-B)**, but for chemogenetic inhibition of SC. **(D1)** Wilcoxon signed rank test (for distance, \*P = 0.016, W = 28.00; for max speed, \*P = 0.031, W = 27.00; for flight probability, \*P = 0.016, W = 28.00). **(D2)** Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 7 mice.

**(E)** Left, schematic of chemogenetic activation of ACC glutamatergic axon terminals in the SC. Middle and right, trajectory-speed map for an example mouse under two scenarios (top, outside-nest bouts; bottom, inside-nest bouts) with solvent (middle) or CNO (right) infusion in SC.

**(F1)** Comparison of the total flight distance (left), maximum speed (middle) and defense probability (right) under two scenarios between solvent and CNO groups, respectively. Wilcoxon signed rank test (for distance, \*\*P = 0.008, W = 36.00; for max. speed, \*\*P = 0.008, W = 36.00; for flight probability, \*\*P = 0.008, W = 36.00), n = 8 mice.

**(F2)** Distribution of CSIs without (grey) and with (color) chemogenetic activation. Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 8 mice.

**(G-H)** Similar to **(E-F)**, but for chemogenetic inhibition of ACC axon terminals in SC. **(G1)** Wilcoxon signed rank test (for distance, \*P = 0.016, W = 28.00; for max. speed, \*P = 0.016, W = 28.00; for flight probability, \*P = 0.016, W = 28.00), n = 7 mice. **(G2)** Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 7 mice.

**(I-J)** Similar to **(E-F)**, but for chemogenetic activation of ACC axon terminals in ZI. **(J1)** Wilcoxon signed rank test (for distance, \*P = 0.031, W = 27.00; for max. speed, \*P = 0.016, W = 28.00; for flight probability, \*P = 0.016, W = 28.00), n = 7 mice. **(J2)** Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 7 mice.

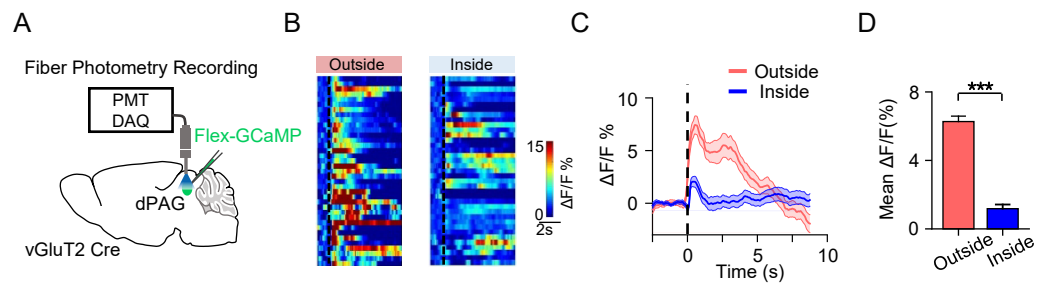
**(K-L)** Similar to **(I-J)**, but for chemogenetic inhibition of ACC axon terminals in ZI. **(L1)** Wilcoxon signed rank test (for distance, \*P = 0.016, W = 28.00; for max. speed, \*P = 0.016, W = 28.00; for flight probability, \*P = 0.016, W = 28.00), n = 7 mice. **(L2)** Kolmogorov-Smirnov test (\*\*P = 0.008, D = 0.86), n = 7 mice.

**(M-N)** Chemogenetic activation of ZI axon terminals in SC. **(N1)** Wilcoxon signed rank test (for distance, \*P = 0.016, W = 28.00; for max. speed, \*P = 0.016, W = 28.00; for flight probability, \*P = 0.016, W = 28.00), n = 7 mice.

**(N2)** Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 7 mice.

**(O-P)** Chemogenetic inhibition of ZI axon terminals in SC. **(P1)** Wilcoxon signed rank test (for distance, \*P = 0.016, W = 28.00; for max speed, \*P = 0.016, W = 28.00; for flight probability, \*P = 0.016, W = 28.00), n = 7 mice.

**(P2)** Kolmogorov-Smirnov test (\*\*\*P < 0.001, D = 1.00), n = 7 mice.



**Figure S7. Fiber photometry recording of dPAG activity under two scenarios. Related to Figure 7.**

(A) Schematic of fiber photometry recording of vGlu2<sup>+</sup> neurons in dPAG.

(B-C) Heat maps (B) and average traces (C) of Ca<sup>2+</sup> responses under two scenarios, aligned to the flight onset (outside-nest bouts) or noise onset (inside-nest bouts).

(D) Mean ΔF/F during 0–3 sec window after the flight onset (outside nest bouts) or noise onset (inside nest bouts). Wilcoxon signed rank test (\*\*P < 0.001, W = 656.00), n = 37 bouts.