

Supplementary Information File

Shared inflammatory glial cell signature after stab wound injury, revealed by spatial, temporal, and cell-type-specific profiling of the murine cerebral cortex

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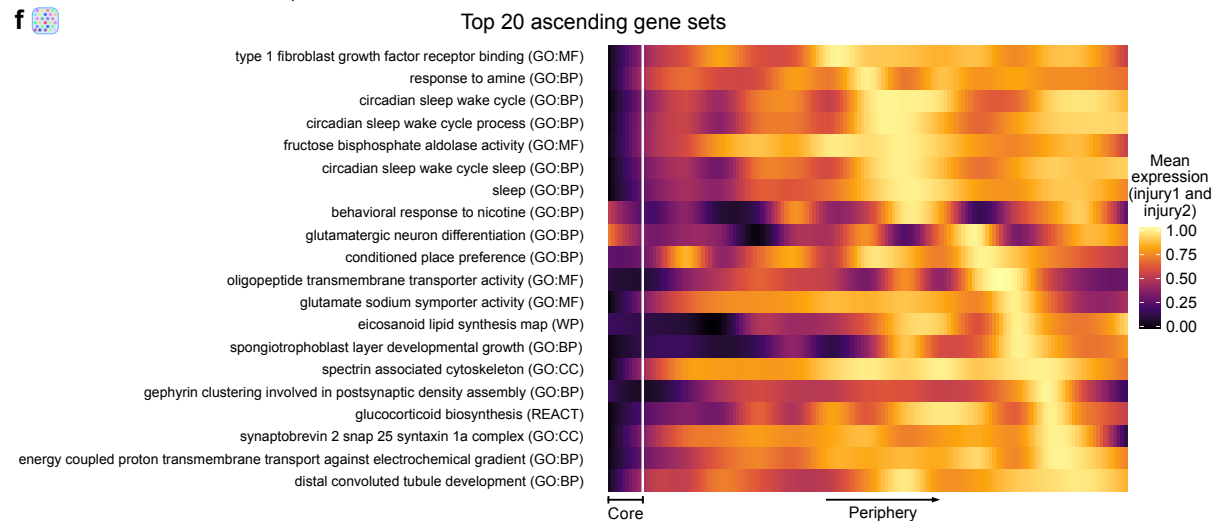
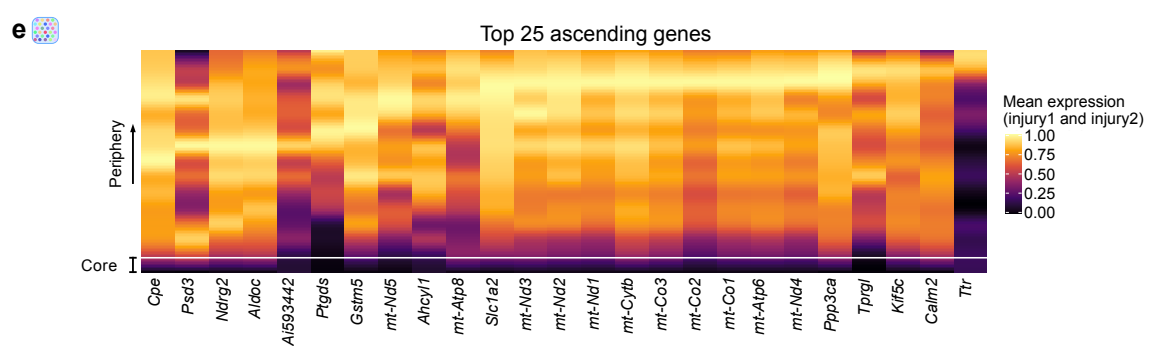
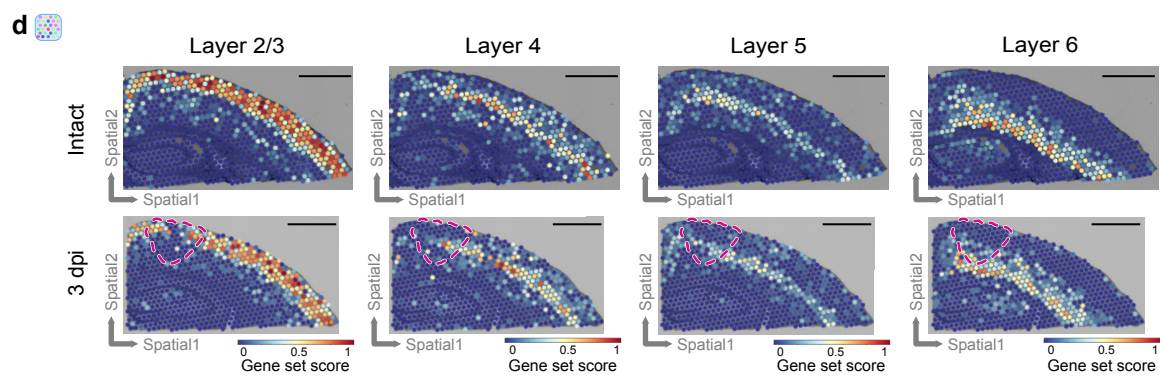
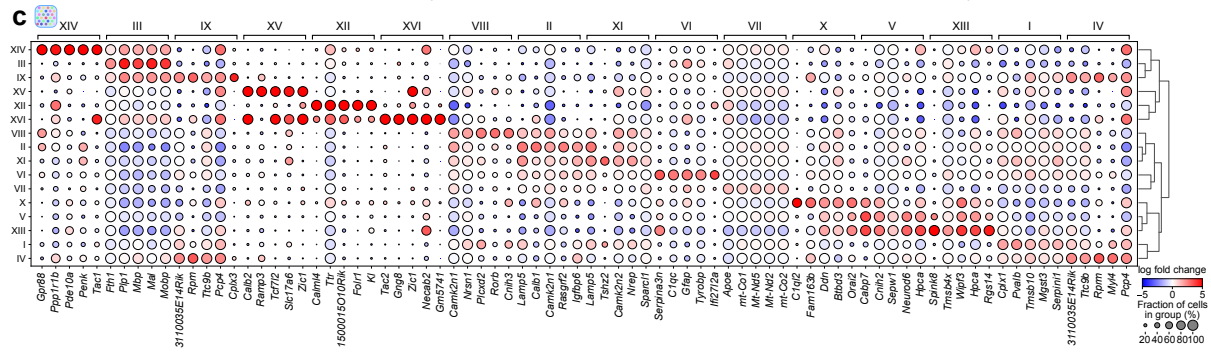
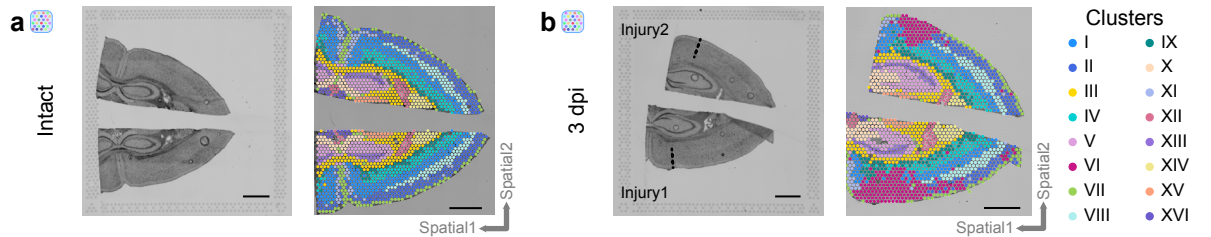
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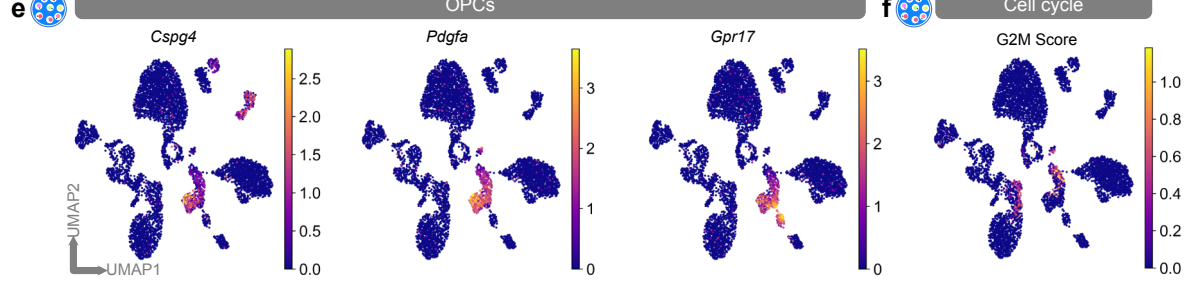
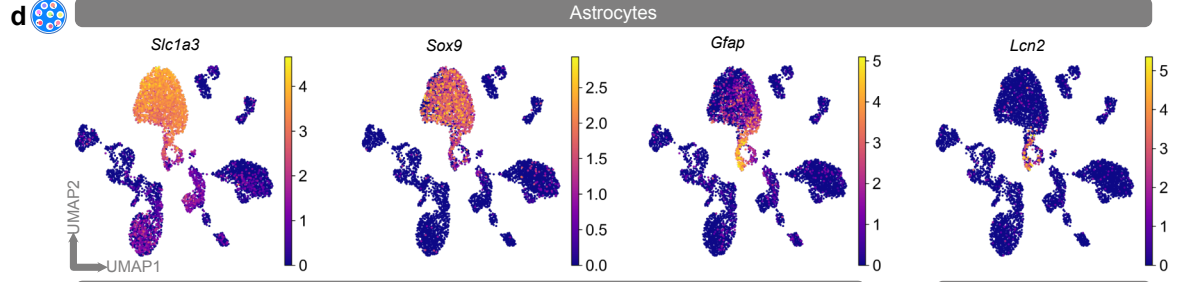
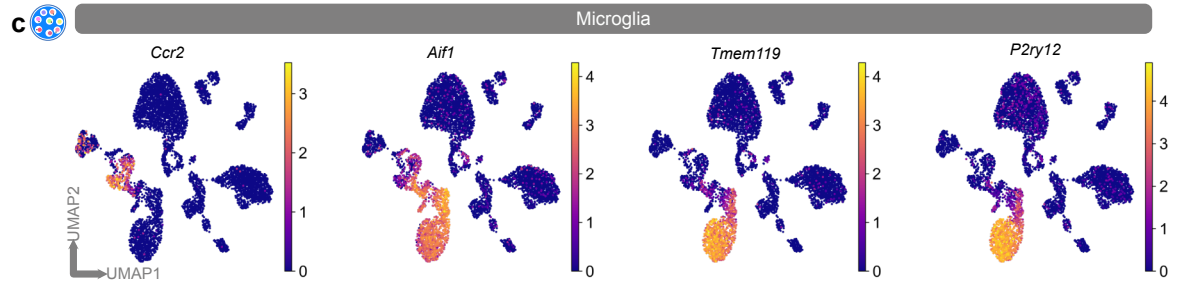
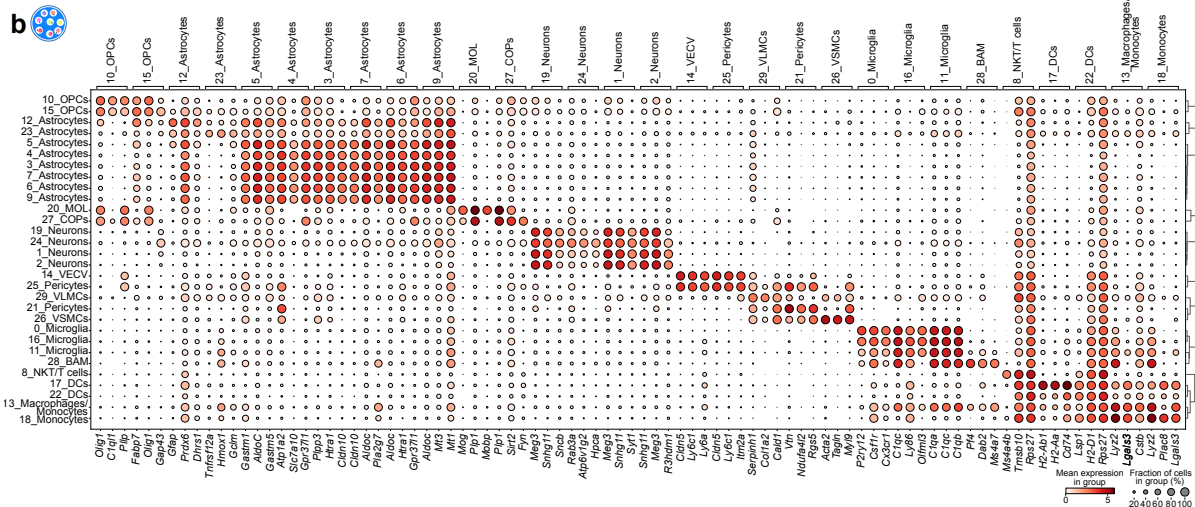
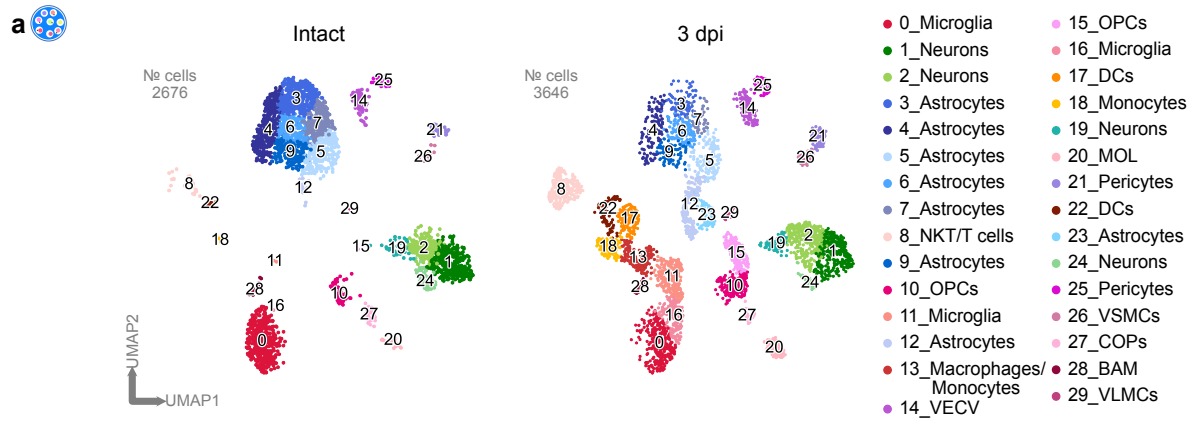
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Supplementary Figure 1. Spatial transcriptome of intact and stab wound-injured mice.

a-b Spatial transcriptomics in intact and stab wound-injured mice (3 dpi). Brains were manually resected and positioned on 10x Visium capture areas. In each capture area, two brain sections of either intact (**a**) or stab wound-injured cortices (**b**) were collected. Clustering of gene expression data on spatial coordinates is based on highly variable genes and subsequent dimensionality reduction in two replicates. The dashed black lines indicate the injury. **c** Dot plot depicting the 5 most enriched genes in each of the 16 identified clusters (Supplementary Table 1). **d** Expression of neuronal layer scores 2/3, 4, 5 and 6 in intact and injured brain sections on spatial coordinates. The dashed pink lines highlight cluster VI borders. Neuronal layer gene set scores are listed in Supplementary Table 2. **e** Heatmap showing the expression of the 25 most ascending genes along the spatial trajectory (see Fig. 2) depicted as mean expression of injury 1 and 2. **f** Heatmap displaying the 20 most enriched ascending gene sets along the spatial trajectory (see Fig. 2) depicted as mean expression of injury 1 and 2. All data shown in this figure are based on n=1 animal per condition over 1 experiment, with two sections being captured. Injury 1 and 2 represent the injuries of each hemisphere in the same animal. Scale bars: **a,b,d**: 1 mm. Abbreviations: BP = biological processes, MF = molecular functions, CC = cellular components, GO = gene ontology, REACT = reactome, WP = WikiPathways, dpi = days post injury.

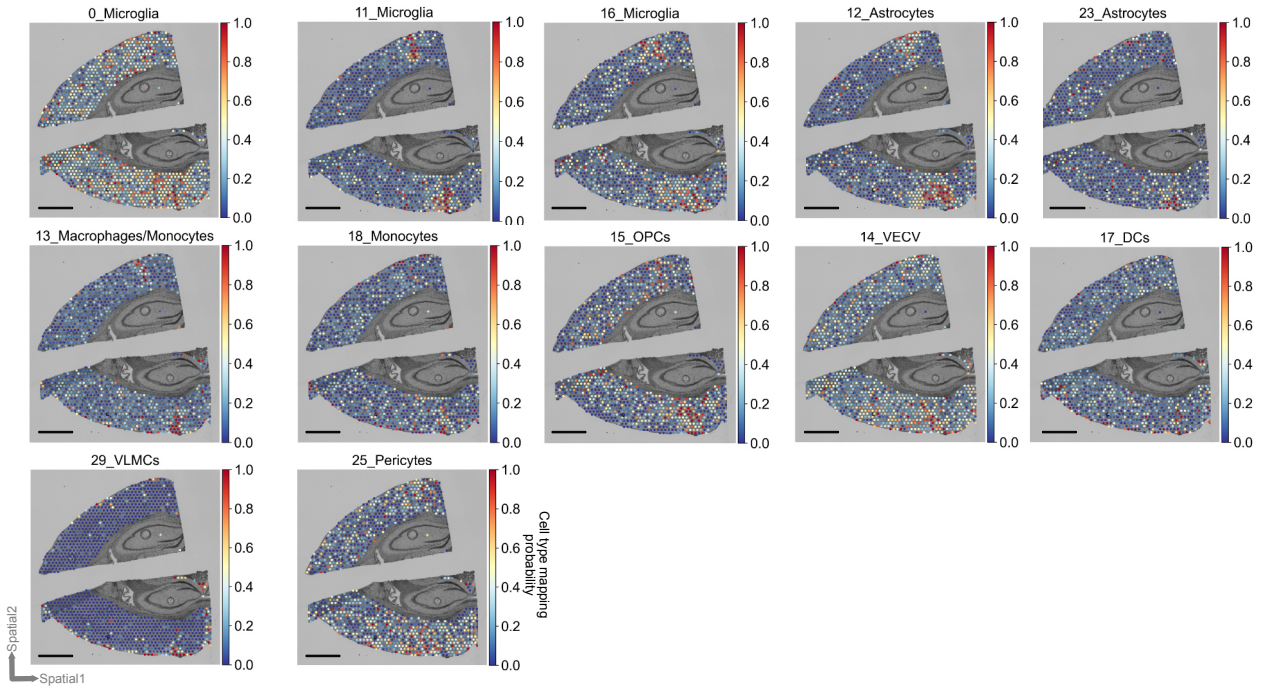


Supplementary Figure 2. scRNA-seq clustering of intact and stab wound-injured mice (3 dpi) and cell-type identification.

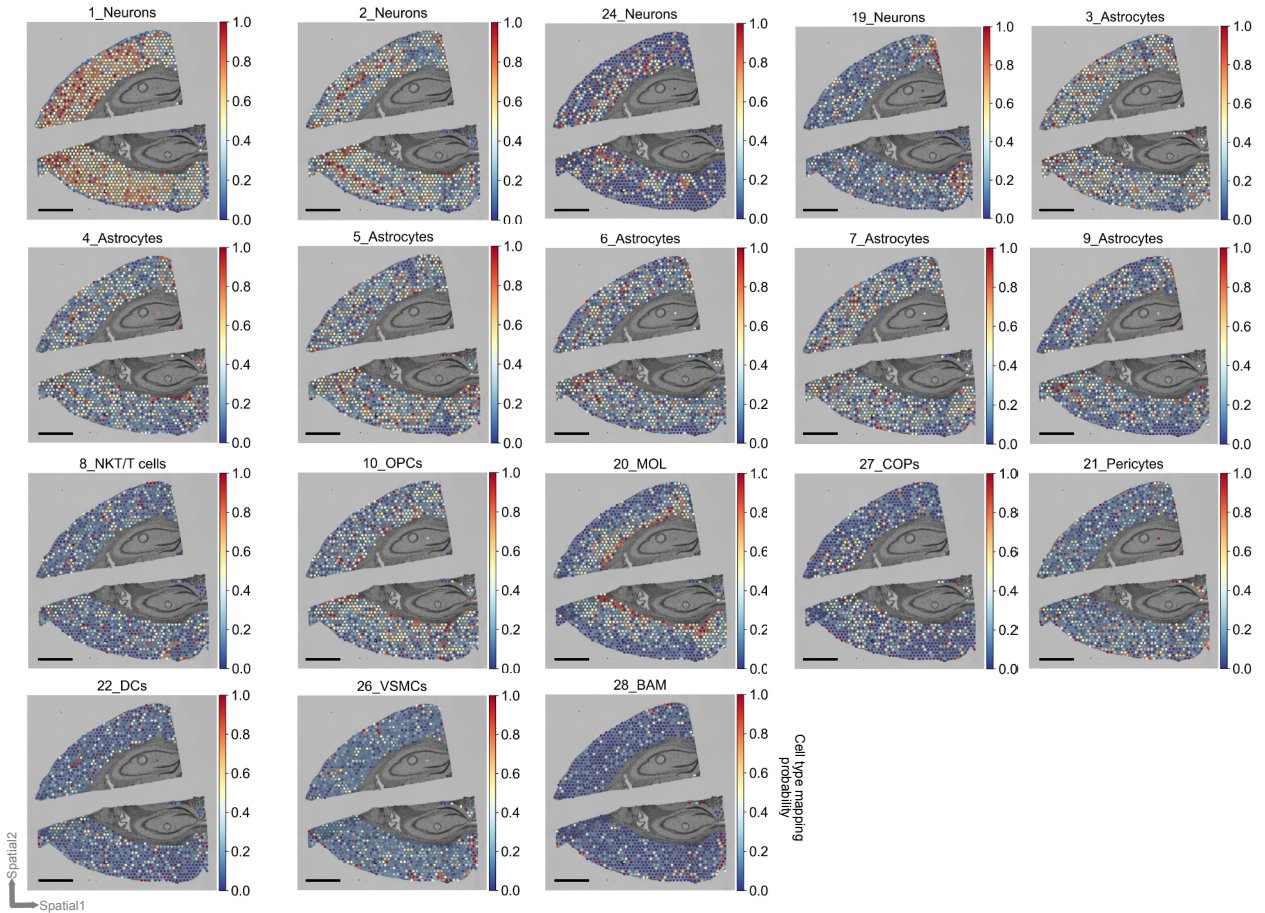
a UMAP plots depicting clustering of cells originating from intact (2676 cells) and injured (3646 cells) conditions among 30 defined clusters. Clusters are color-coded and annotated according to their transcriptional identities. Note that clusters 13_Macrophages/Monocytes, 17_DCs, and 23_Astrocytes are almost absent in the intact condition. **b** Dot plot depicting the expression of the 3 most enriched genes in each of the 30 identified clusters (see Supplementary Data 3). **c-e** UMAPs highlighting example marker genes to identify microglia/macrophages (**c**), astrocytes (**d**) OPCs (**e**) and cycling cells (**f**). G2/M gene set score is listed in Supplementary Table 4. Data shown in this figure are derived from n=3 intact animals over 1 experiment (1 scRNAseq library) and n=3 3dpi animals over 1 experiment (2 scRNAseq libraries). Abbreviations: UMAP = uniform manifold approximation and projection, dpi = days post injury, OPCs = oligodendrocyte progenitor cells, COPs = committed oligodendrocyte progenitors, MOL = mature oligodendrocytes, VECV = vascular endothelial cells (venous), VSMCs = vascular smooth muscle cells, VLMCs = vascular and leptomeningeal cells, BAM = border-associated macrophages, NKT cells = natural killer T cells, DCs = dendritic cells.



Enriched clusters

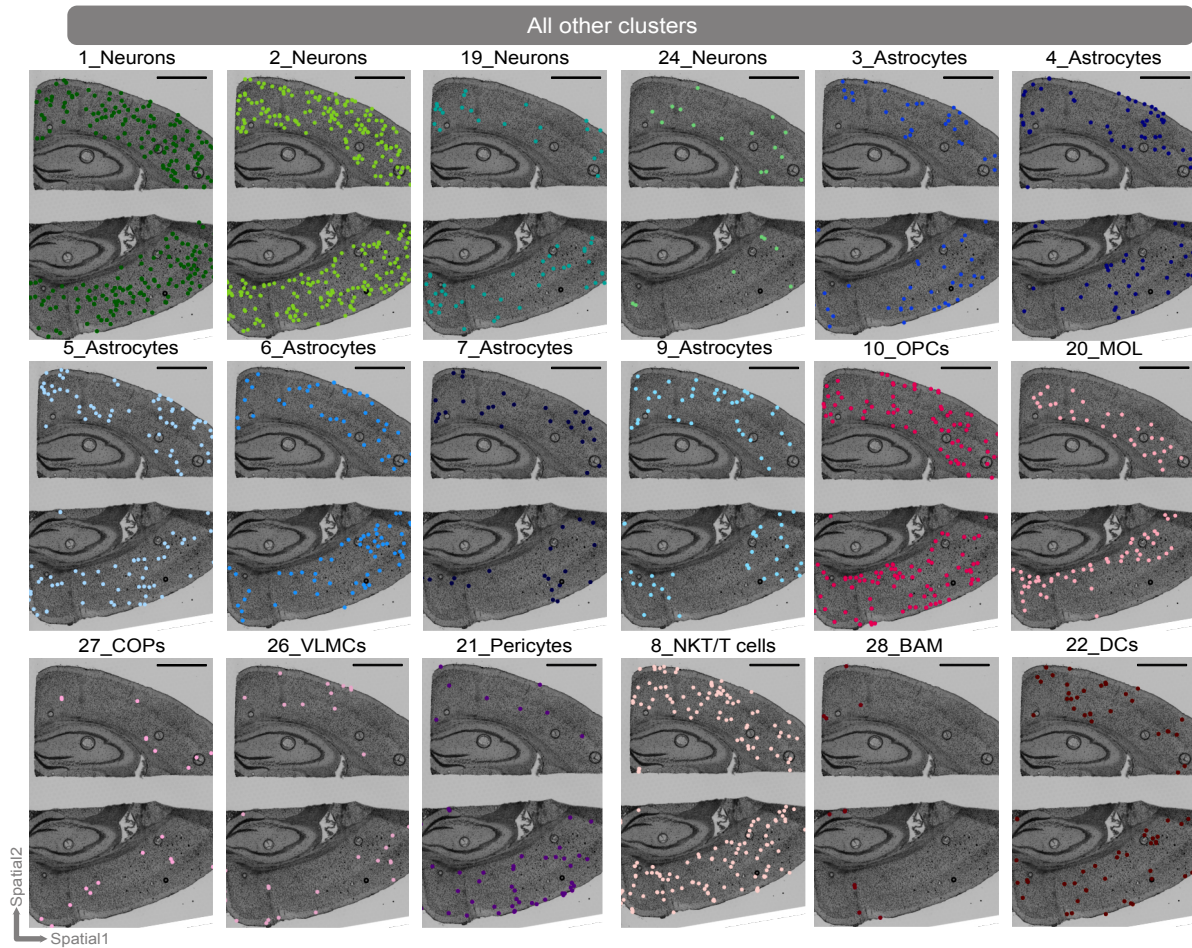
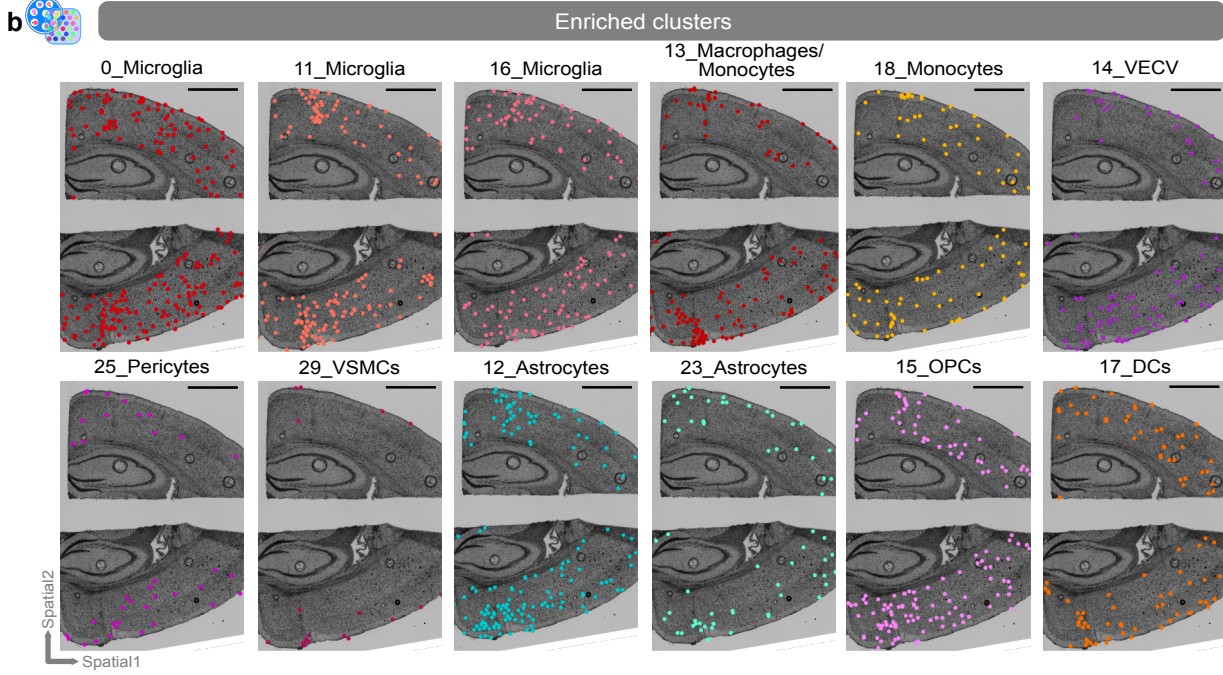
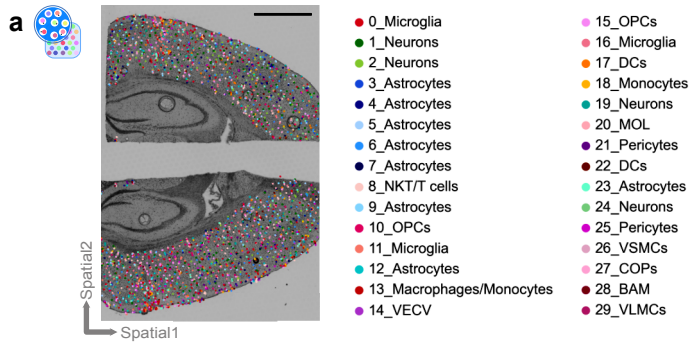


All other clusters



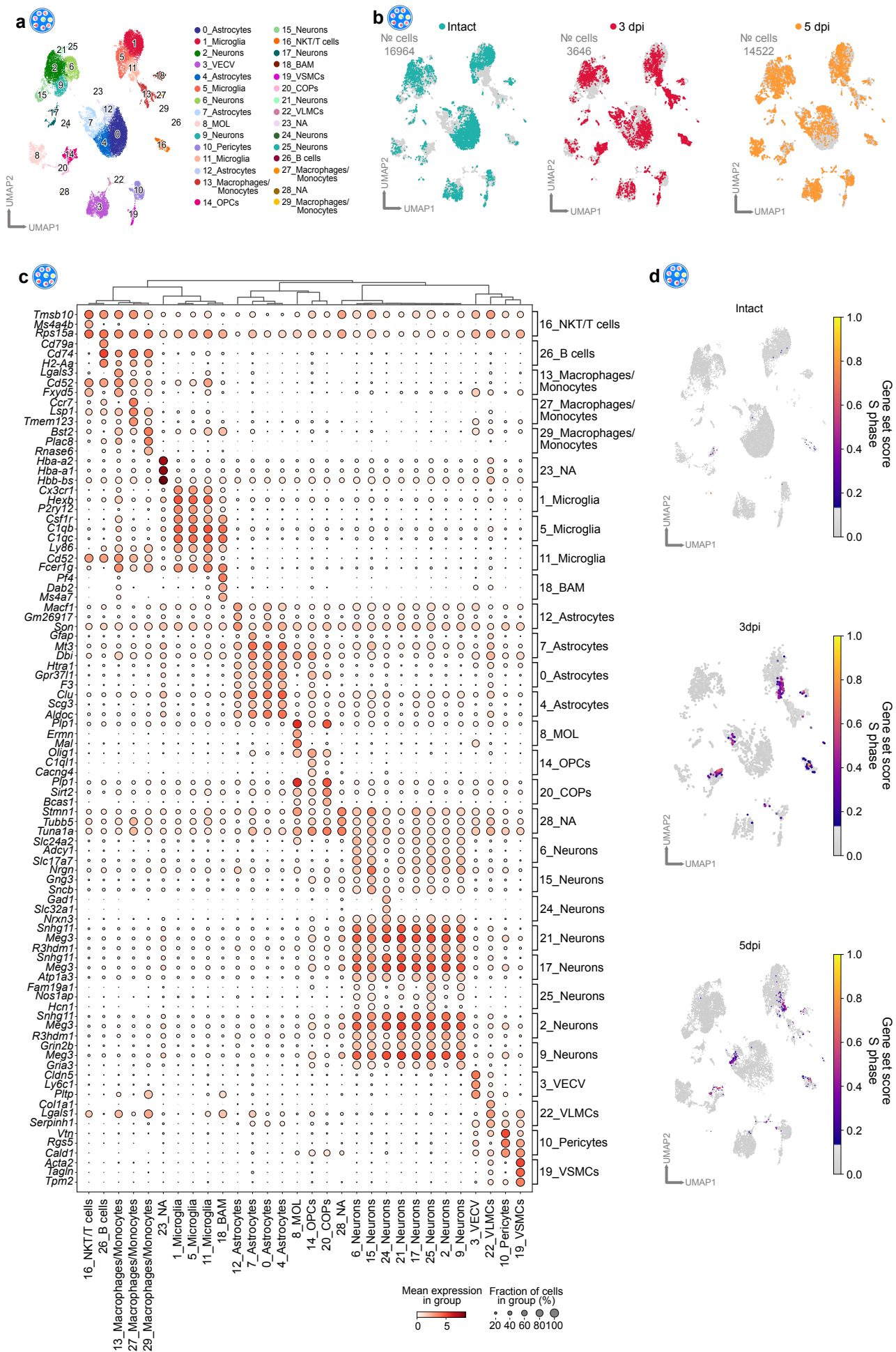
Supplementary Figure 3. Probabilistic mapping of in scRNA-seq identified cellular clusters on Visium dataset.

Probabilistic mapping of in Fig. 3b identified scRNA-seq (3 dpi only) clusters on the spatial transcriptome dataset (3 dpi). Stab wound injury elicits different reaction modes in the injury vicinity. Plots are grouped into injury enriched clusters (upper panel) and all other clusters (lower panel). scRNAseq data shown in this figure are derived from n=3 intact animals over 1 experiment (1 scRNAseq library) and n=3 3dpi animals over 1 experiment (2 scRNAseq libraries). stRNAseq data are based on n=1 animal over 1 experiment with two injured hemispheres being captured. Scale bars: 1 mm. Abbreviations: dpi = days post injury, OPCs = oligodendrocyte progenitor cells, NKT cells = natural killer T cells, DCs = dendritic cells, MOL = mature oligodendrocytes, VSMCs = vascular smooth muscle cells, COPs = committed oligodendrocyte progenitors, BAM = border-associated macrophages, VLMCs = vascular and leptomeningeal cells, VECV = vascular endothelial cells (venous).



Supplementary Figure 4. Single-cell deconvolution-based mapping of in scRNA-seq identified cellular clusters on Visium dataset.

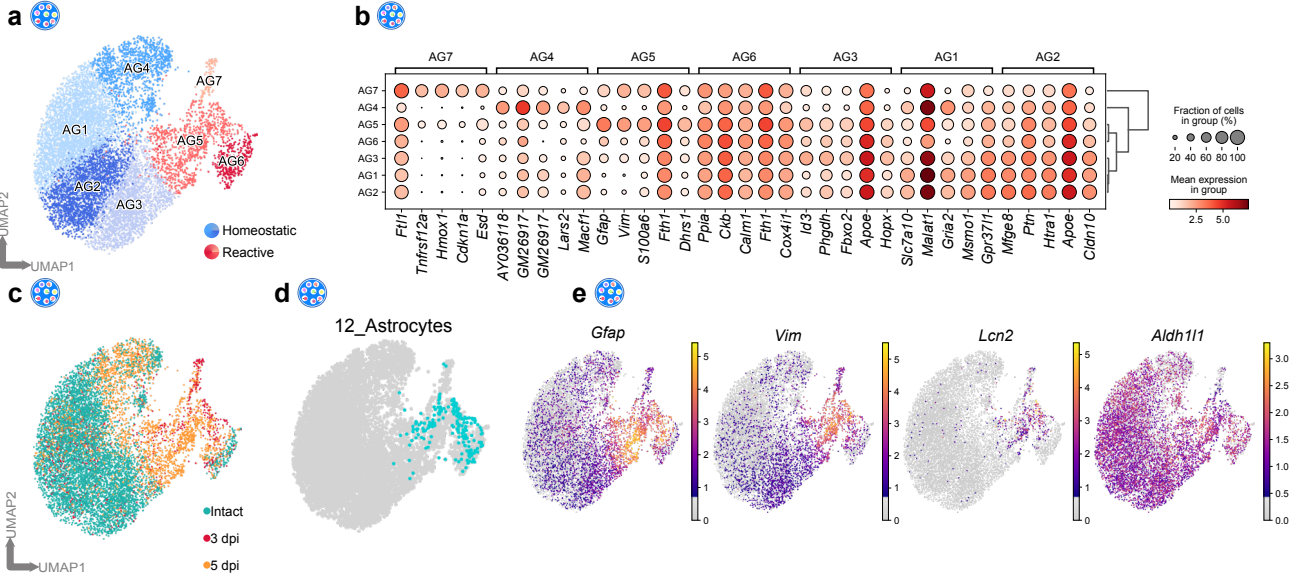
a,b Deconvolution based mapping of in Fig. 3 identified scRNA-seq clusters on the spatial transcriptome dataset (3 dpi). Plots are grouped into injury enriched clusters (upper panel) and all other clusters (lower panel). scRNAseq data shown in this figure are derived from n=3 intact animals over 1 experiment (1 scRNAseq library) and n=3 3dpi animals over 1 experiment (2 scRNAseq libraries). stRNAseq data are based on n=1 animal over 1 experiment with two injured hemispheres being captured. Scale bars: 1 mm. Abbreviations: dpi = days post injury, OPCs = oligodendrocyte progenitor cells, NKT cells = natural killer T cells, DCs = dendritic cells, MOL = mature oligodendrocytes, VSMCs = vascular smooth muscle cells, COPs = committed oligodendrocyte progenitors, BAM = border-associated macrophages, VLMCs = vascular and leptomeningeal cells, VECV = vascular endothelial cells (venous).



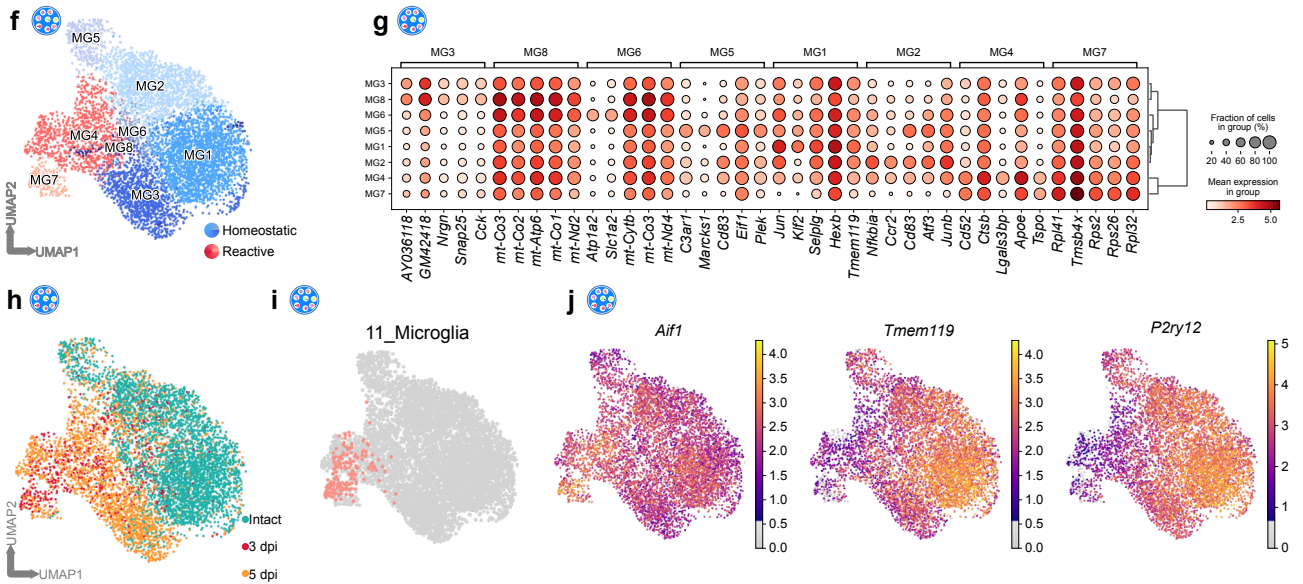
Supplementary Figure 5. Integration of scRNA-seq datasets of intact and stab wound-injured mice (3 and 5 dpi), cluster distribution and cell type identification.

a,b UMAP plots depicting clustering (**a**) and cell distribution of intact (16964 cells), 3 dpi (3646 cells), and 5 dpi (14522 cells) cells among 30 defined clusters (**b**). Clusters were color-coded and annotated according to their transcriptional identities. **c** Dot plot depicting expression of the 3 most enriched genes in each of the 30 identified clusters (see Supplementary Data 4). **d** UMAP plots highlighting proliferating cells entering S-phase in intact and brain injured conditions (see Supplementary Table 4). Data shown in this figure are derived from n=12 intact animals over 3 independent experiments (5 scRNAseq libraries), n=3 3dpi animals over 1 experiment (2 scRNAseq libraries), and n=9 5dpi animals over 3 independent experiments (3 scRNAseq libraries). Abbreviations: dpi = days post injury, UMAP = uniform manifold approximation and projection, dpi = days post injury, VECV = vascular endothelial cells (venous), MOL = mature oligodendrocytes, OPCs = oligodendrocyte progenitor cells, NKT cells = natural killer T cells, BAM = border-associated macrophages, VSMCs = vascular smooth muscle cells, COPs = committed oligodendrocyte progenitors, VLMCs = vascular and leptomeningeal cells, NA = not available.

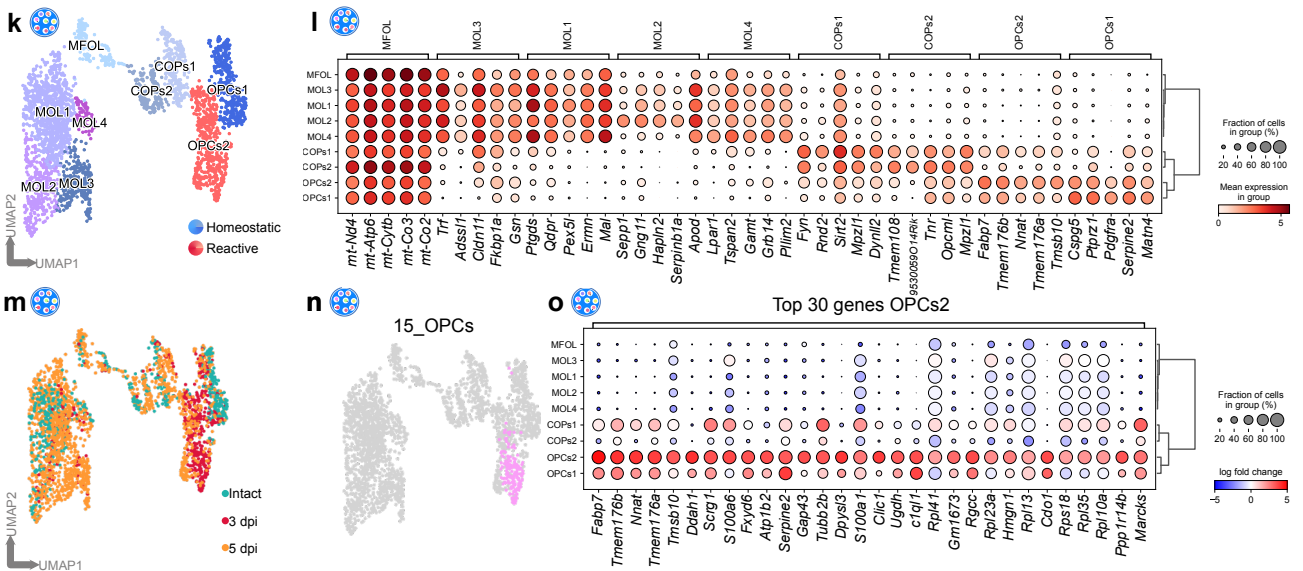
Astrocytes



Microglia



Oligodendrocytes

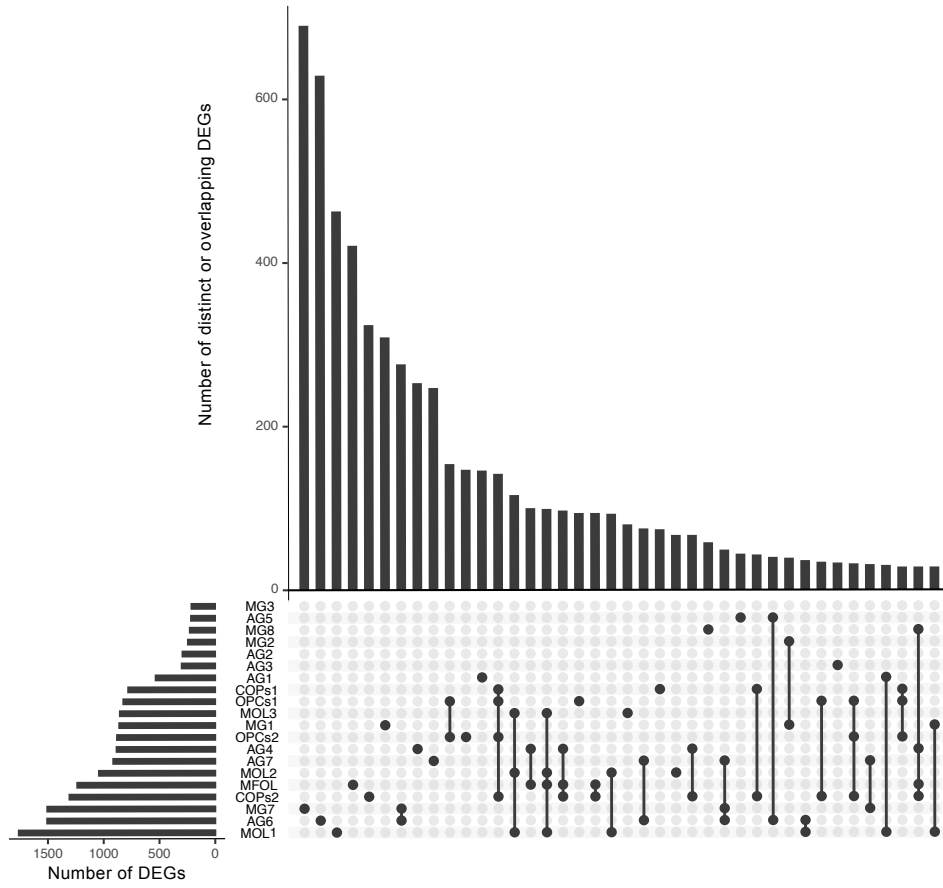


Supplementary Figure 6. Stab wound injury elicits unique gene expression in distinct glial subpopulations.

a UMAP depicting subclustered astrocytes of integrated and batch-corrected intact, 3 dpi, and 5 dpi datasets. **b** Dot plot depicting expression of the 5 most enriched genes in each of the 7 identified clusters. **c** UMAP displaying cell distribution of intact (green), 3 dpi (red) and 5 dpi (orange) cells among all 7 astrocytic clusters. **d** UMAP displaying localization of injury-enriched cluster 12_Astrocytes (turquoise, Supplementary Fig. 4b) on subclustered astrocytes. **e** UMAPs highlighting expression of example marker genes *Gfap*, *Vim*, *Lcn2* and *Aldh1l1* to identify reactive astrocyte clusters. **f** UMAP depicting subclustered microglia of integrated and batch-corrected intact, 3 dpi, and 5 dpi datasets. **g** Dot plot depicting expression of the 5 most enriched genes in each of the 8 identified clusters. **h** UMAP displaying cell distribution of intact (green), 3 dpi (red), and 5 dpi (orange) cells among all 8 microglial clusters. **i** UMAP displaying localization of injury-enriched cluster 11_Microglia (peach, Supplementary Fig. 4b) on subclustered microglia. **j** UMAPs highlighting expression of example marker genes *Aif1*, *Tmem119* and *P2ry12* to identify reactive microglial clusters. **k** UMAP depicting subclustered oligodendrocytes of integrated and batch-corrected intact, 3 dpi, and 5 dpi datasets. **l** Dot plot depicting expression of the 5 most enriched genes in each of the 9 identified clusters. **m** UMAP displaying cell distribution of intact (green), 3 dpi (red), and 5 dpi (orange) cells among all 9 clusters. **n** UMAP displaying localization of injury-enriched cluster 15_OPCs (pink, Supplementary Fig. 4b) on subclustered oligodendrocytes. **o** Dot plot depicting expression of the 30 most enriched genes in cluster OPCs2. Note comparable gene expression of clusters OPCs1 and OPCs2 prevents the identification of unique reactive OPC markers. Data shown in this figure are derived from n=12 intact animals over 3 independent experiments (5 scRNAseq libraries), n=3 3dpi animals over 1 experiment (2 scRNAseq libraries), and n=9 5dpi animals over 3 independent experiments (3 scRNAseq libraries). Abbreviations: UMAP = uniform manifold approximation and projection, AG = astroglia, dpi = days post injury, MG = microglia, OPCs = oligodendrocyte progenitor cells, COPs = committed oligodendrocyte progenitors, MFOL = myelin-forming oligodendrocytes, MOL = mature oligodendrocytes.

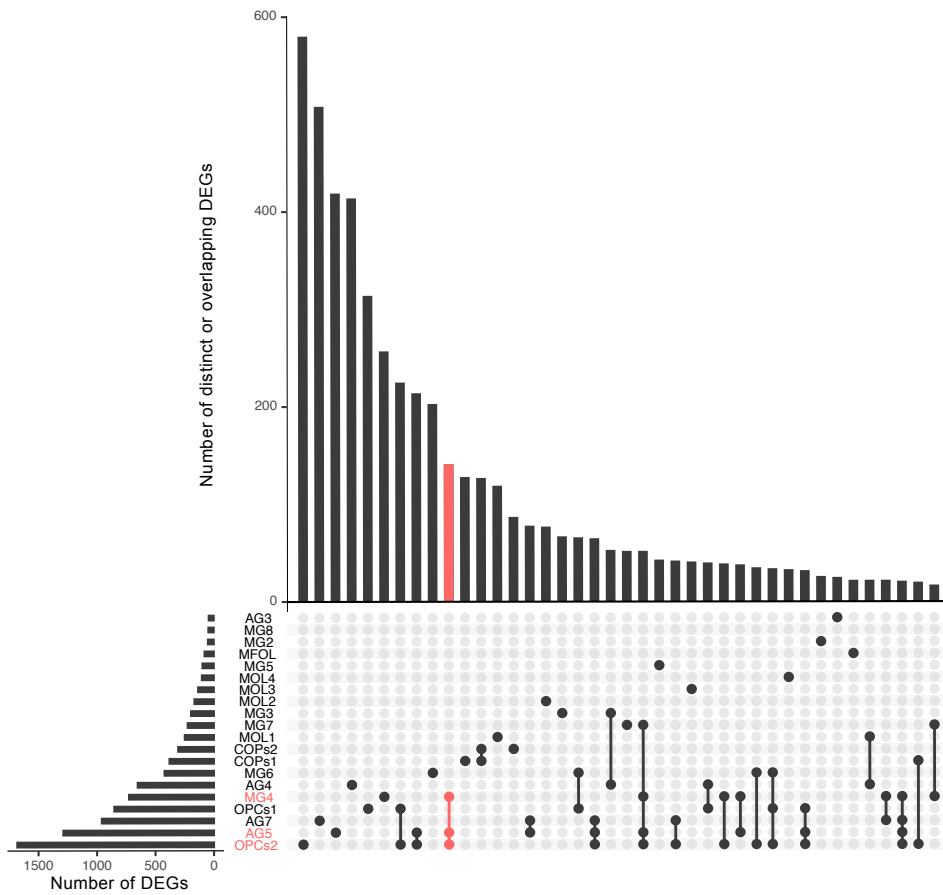
a

UpSet plot downregulated genes



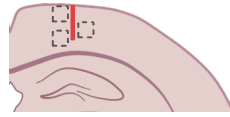
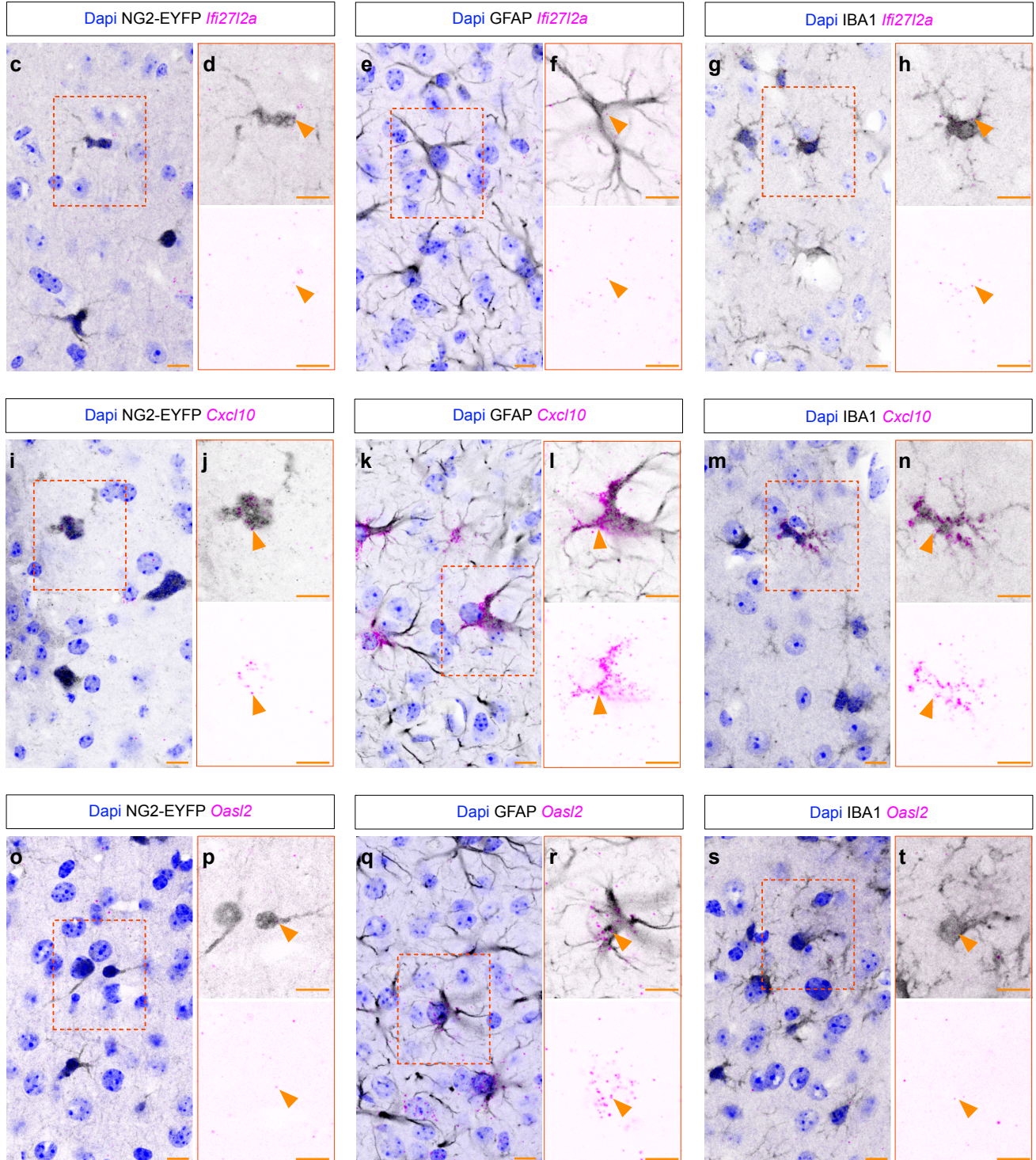
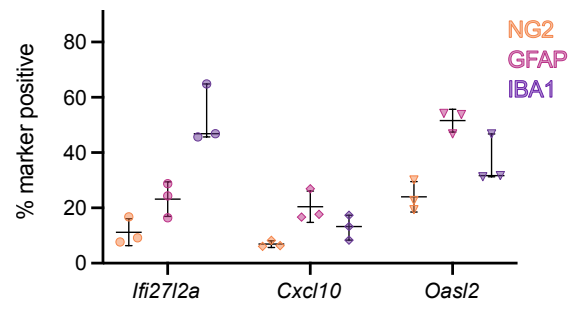
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UpSet plot upregulated genes



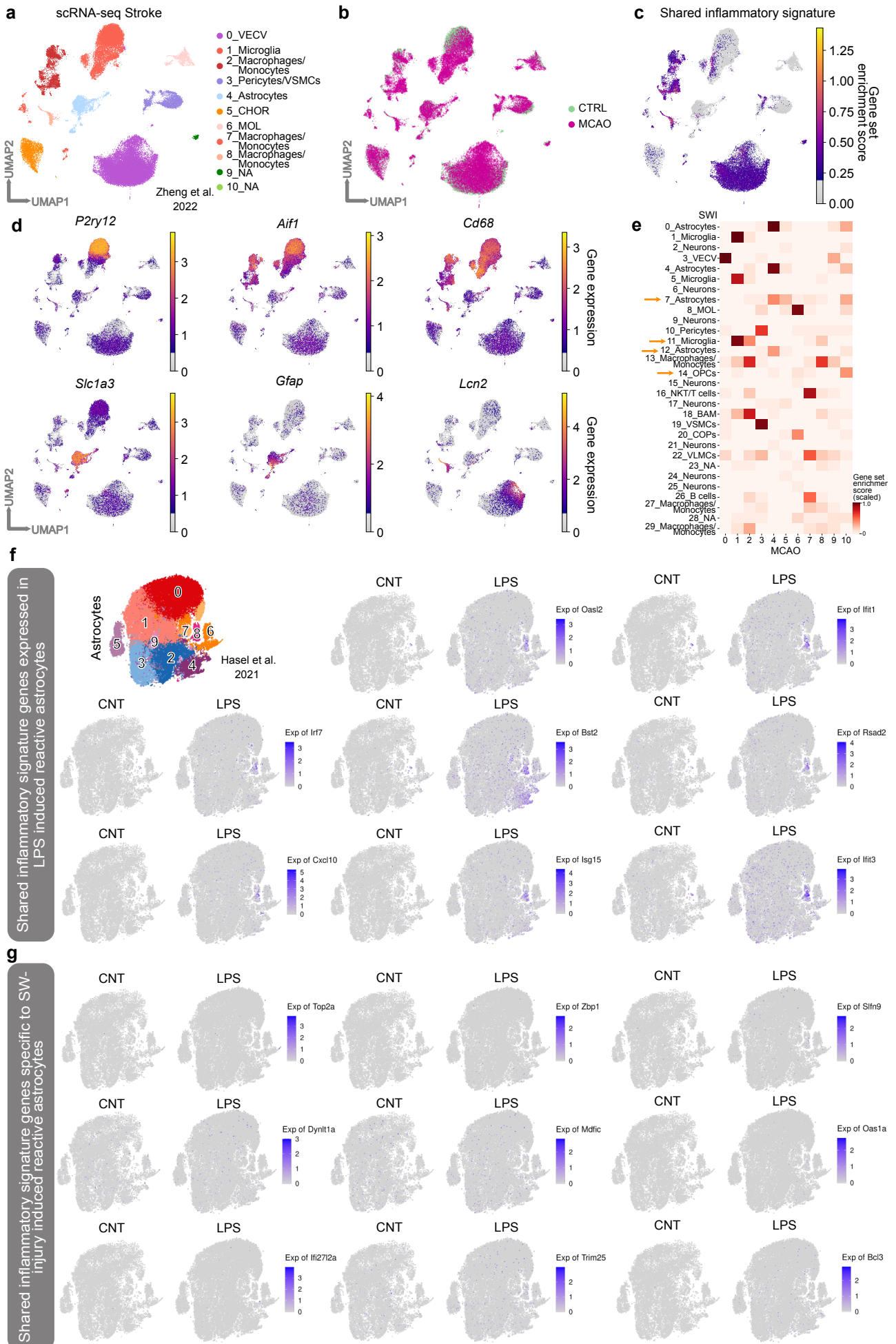
Supplementary Figure 7. Reactive glial subpopulations share injury-induced transcriptomic signature.

a,b UpSet plot illustrating downregulated (**a**) and upregulated (**b**) DEGs ($p_{\text{val}} < 0.05$, \log_2 fold change > 1.6 or \log_2 fold change < -1.6 , method: t-test overestimated_variance) across different glial subclusters of integrated and batch-corrected intact, 3 dpi, and 5 dpi datasets (Fig. 4c-e). Each horizontal bar represents number of DEGs within a specific cluster, and vertical bars indicate the number of unique or shared genes between clusters. Connected dots highlight shared DEGs across clusters, while single dots represent uniquely regulated DEGs. Wherever applicable, DEGs are determined by comparing each glial subcluster to all other subclusters within the respective cell type (further details in Methods). The red bar in (**b**) highlights commonly shared genes between reactive astrocytic, microglial, and oligodendroglial subclusters. Data shown in this figure are derived from $n=12$ intact animals over 3 independent experiments (5 scRNAseq libraries), $n=3$ 3dpi animals over 1 experiment (2 scRNAseq libraries), and $n=9$ 5dpi animals over 3 independent experiments (3 scRNAseq libraries). Abbreviations: DEGs = differentially expressed genes, AG = astroglia, MG = microglia, OPCs = oligodendrocyte progenitor cells, COPs = committed oligodendrocyte progenitors, MFOL = myelin-forming oligodendrocytes, MOL = mature oligodendrocytes.

a**b**

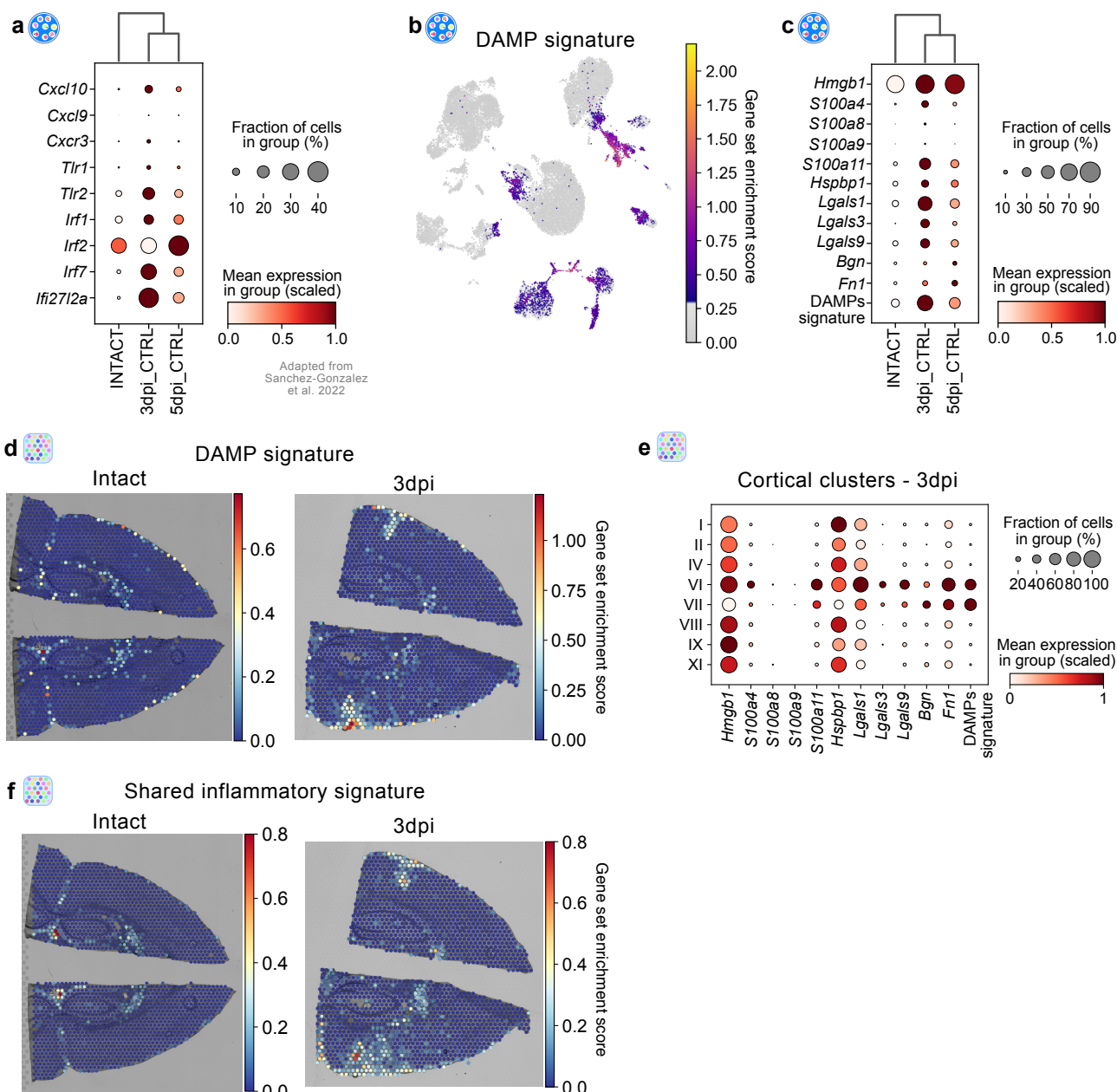
Supplementary Figure 8. Reactive glial subpopulations display shared gene expression following injury.

a Experimental paradigm for assessing shared gene expression in astrocytes, microglia, and OPCs (NG2-EYFP). The dashed gray boxes on the mouse brain scheme indicate the example imaging areas. The red line indicates the injury core. **b** Dot plot depicting the proportion of OPCs (NG2-EYFP, yellow), astrocytes (GFAP, pink) and microglia (IBA1, purple) expressing the selected shared innate immunity- associated genes *lfi27l2a*, *Cxcl10*, and *Oasl2*. Data are shown as median \pm 95% confidence interval. Each data point represents one animal (n = 3). Source data are provided as Source Data file. **c-t** Representative images of RNAscope for *lfi27l2a* (**c-g**), *Cxcl10* (**i-m**), and *Oasl2* (**o-s**) (magenta) combined with GFP (NG2 glia) (black) (**c,i,o**), GFAP (black) (**e,k,q**), IBA1 (black) (**g,m,s**) immunostaining to identify cell types. Framed micrographs are magnifications of the red boxed areas in corresponding overview images. The orange arrowheads in micrograph depict double positive cells. All images are single z-plane of confocal z-stacks. Scale bars: **c-t**: 10 μ m.



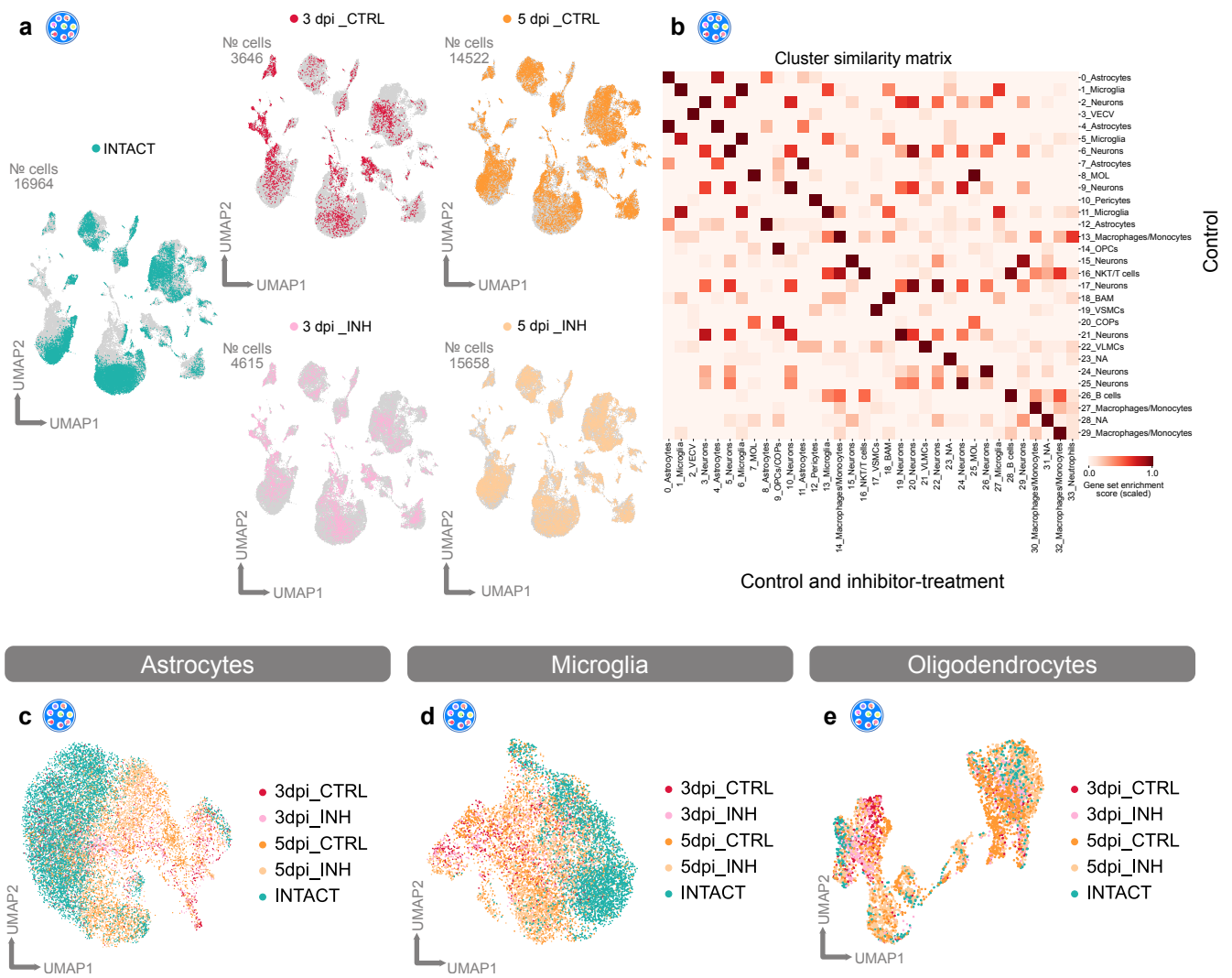
Supplementary Figure 9. Common inflammatory gene signature in murine brain cells following MCAO and LPS-induced reactive astrocytes

a-d Analysis of MCAO scRNA-seq dataset⁵¹. **a** UMAP plot illustrating single cells distributed among 11 distinct clusters. **b** UMAP plot depicting the distribution of cells isolated from CTRL and MCAO cerebral cortices. **c** UMAP plot depicting localization of the gene set scores related to the shared inflammatory signature (see Fig. 5c) among all defined cell clusters depicted in Supplementary Fig. 9a. **d** UMAP plots highlighting expression of example marker genes *P2ry12*, *Aif1*, *Cd68* to identify reactive microglia clusters and *Slc1a3*, *Gfap* and *Lcn2* to identify reactive astrocyte clusters. **e** Heatmap displaying cluster similarity of SWI dataset (y-axis), and MCAO dataset (x-axis). Orange arrows highlight clusters of interest. **f-g** UMAPs illustrating selective marker gene expression of shared inflammatory signature (see Fig. 5c) in subclustered astrocytes based on Hasel et al.³⁸. Plots depict presence (**f**) and absence (**g**) of several shared inflammatory genes in reactive astrocyte cluster 8. Abbreviations: UMAP = uniform manifold approximation and projection, MCAO = middle cerebral artery occlusion, CTRL = MCAO sham group, VECV = vascular endothelial cells (venous), CHOR = choroid plexus epithelial cells, MOL = mature oligodendrocytes, OPCs = oligodendrocyte progenitor cells, NKT cells = natural killer T cells, BAM = border-associated macrophages, VSMCs = vascular smooth muscle cells, COPs = committed oligodendrocyte progenitors, VLMCs = vascular and leptomeningeal cells, NA = not available, CNT = control, LPS = lipopolysaccharide.



Supplementary Figure 10. Expression of genes associated with Tlr1/2 and Cxcr3 signaling pathways and DAMP signature upon stab wound injury.

a Dot plot depicting gene expression of components associated with the Tlr1/2 and Cxcr3 signaling pathways. Genes are adapted and complemented based on Sanchez-Gonzalez et al.⁵³. **b** UMAP plot illustrating the expression of the gene set scores associated with DAMP signature. **c** Dot plot depicting gene expression of components associated with DAMP signature⁵² and used to calculate the score in Supplementary Fig. 10b. **d, f** Expression of the gene set scores of DAMP (**d**) and shared inflammatory (**f**) signature in spatial context. **e** Dot plot depicting gene expression of components associated with DAMP signature within cortical clusters. Data shown in this figure are derived from n=12 intact animals over 3 independent experiments (5 scRNAseq libraries), n=3 3dpi animals over 1 experiment (2 scRNAseq libraries), and n=9 5dpi animals over 3 independent experiments (3 scRNAseq libraries). stRNAseq data are based on n=1 animal over 1 experiment with two injured hemispheres being captured. Abbreviations: UMAP = uniform manifold approximation and projection, CTRL = stab wound-injured control animals, DAMPs = Damage-associated molecular patterns.



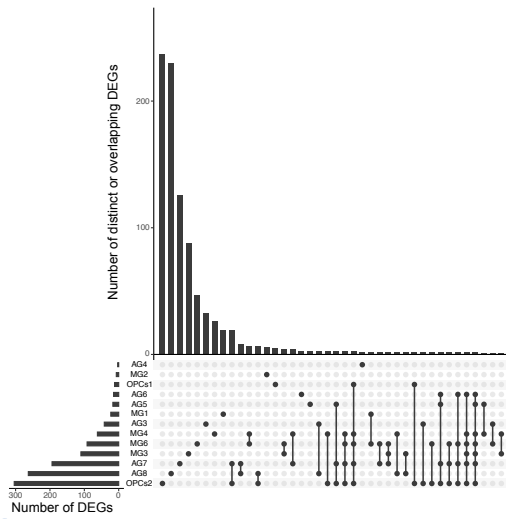
Supplementary Figure 11. scRNA-seq data integration of intact and stab wound-injured cortices.

a UMAP plots depicting cell distribution of integrated and batch-corrected intact (green, 16964 cells), 3 dpi CTRL (red, 3646 cells), 3 dpi INH (pink, 4615 cells), 5 dpi CTRL (orange, 14522 cells), and 5 dpi INH (peach, 15658 cells) datasets. **b** Heatmap displaying cluster similarity between dataset containing integrated control conditions (INT, injured CTRL, y-axis), and dataset containing integrated control and inhibitor treatment conditions (INT, injured CTRL + INH, x-axis). **c** UMAP displaying cell distribution of intact (green), 3 dpi CTRL (red), 3 dpi INH (pink), 5 dpi CTRL (orange), and 5 dpi INH (peach) cells among the 8 identified astrocytic clusters as depicted in Fig. 6c. Clusters AG5, AG6, AG7 and AG8 are mainly formed by cells originating from injured CTRL and INH animals. **d** UMAP displaying cell distribution of intact (green), 3 dpi CTRL (red), 3 dpi INH (pink), 5 dpi CTRL (orange), and 5 dpi INH (peach) cells among the 8 identified microglial clusters as depicted in Fig. 6d. Clusters MG4 and MG6 are mainly formed by cells originating from injured CTRL and INH animals. **e** UMAP displaying cell distribution of intact (green), 3 dpi CTRL (red), 3 dpi INH (pink), 5 dpi CTRL (orange), and 5 dpi INH (peach) cells among the 11 identified oligodendroglial clusters as depicted in Fig. 6e. Cluster OPCs2 is mainly formed by cells originating from injured CTRL and INH animals. Data shown in this figure are derived from n=12 intact animals over 3 independent experiments (5 scRNAseq libraries), n=3 3dpi CTRL animals over 1 experiment (2 scRNAseq libraries), n=3 3dpi INH animals over 1 experiment (2 scRNAseq libraries), n=9 5dpi CTRL animals over 3 independent experiments (3 scRNAseq libraries) and n=6 5dpi INH animals over 2 independent experiments (3 scRNAseq libraries). Abbreviations: UMAP = uniform manifold approximation and

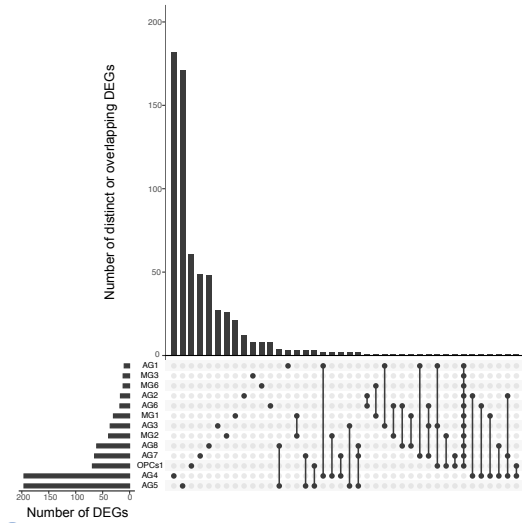
projection, dpi=days post injury, CTRL = stab wound-injured control mice, INH = stab wound-injured inhibitor-treated mice, VECV = vascular endothelial cells (venous), MOL = mature oligodendrocytes, OPCs = oligodendrocyte progenitor cells, NKT cells = natural killer T cells, BAM = border-associated macrophages, VSMCs = vascular smooth muscle cells, COPs = committed oligodendrocyte progenitors, VLMCs = vascular and leptomeningeal cells, NA = not available.



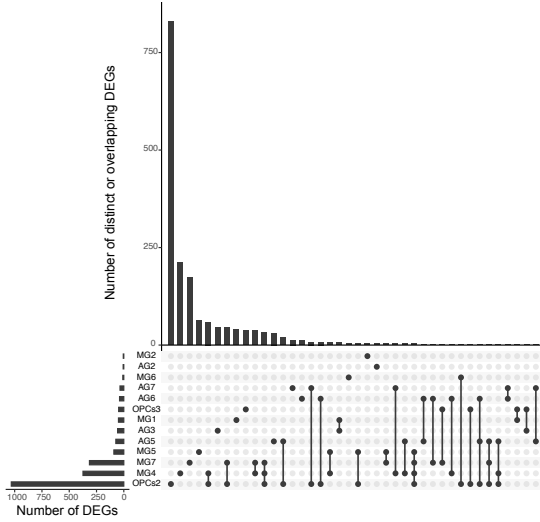
3 dpi downregulated genes after treatment



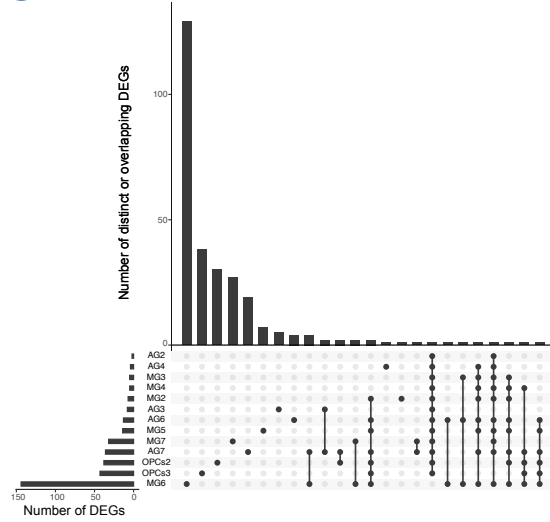
3 dpi upregulated genes after treatment



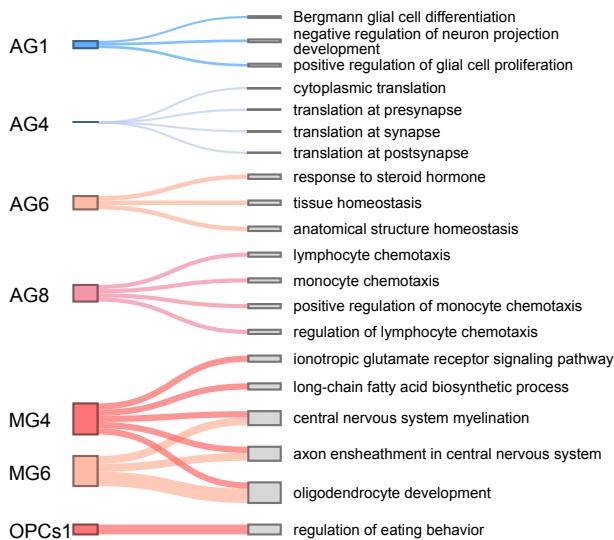
5 dpi downregulated genes after treatment



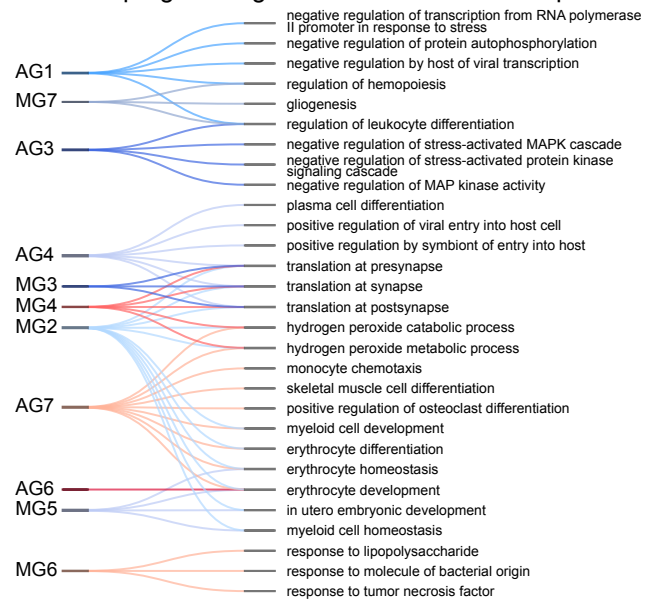
5 dpi upregulated genes after treatment



Enriched biological processes of upregulated genes after treatment - 3 dpi



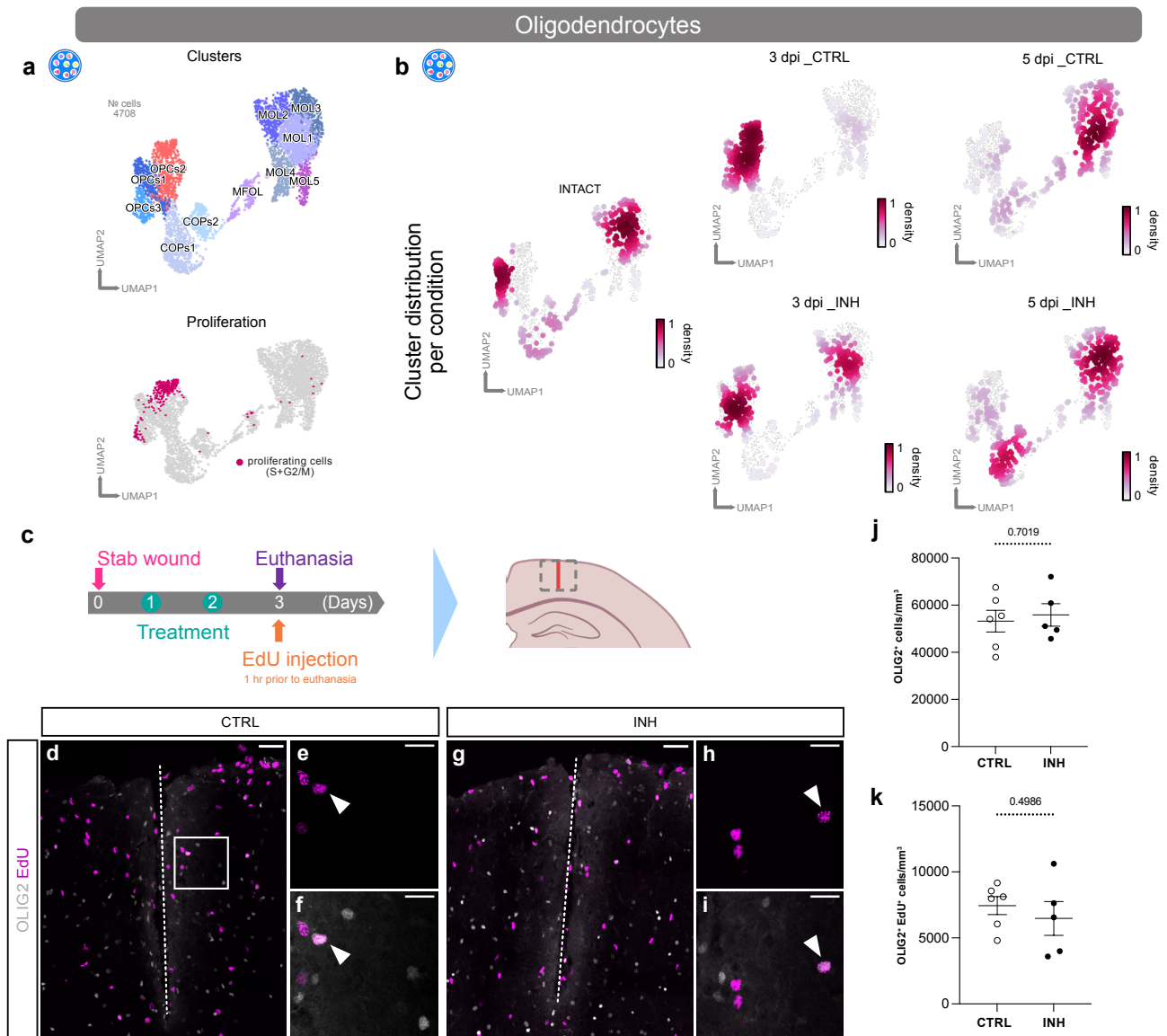
Enriched biological processes of upregulated genes after treatment - 5 dpi



Supplementary Figure 12. The Cxcr3 and Tlr1/2 pathway inhibition after stab wound injury induces subcluster specific changes.

a-d UpSet plots depicting unique and overlapping downregulated (**a,c**) and upregulated (**b,d**) DEGs ($p\text{val} < 0.05$, \log_2 fold change > 0.7 or \log_2 fold change < -0.7 , method: Wald test) between different glial subclusters in response to the Cxcr3 and Tlr1/2 inhibition at 3 dpi (**a,b**) and 5 dpi (**c,d**). Each horizontal bar represents the number of DEGs within a specific cluster, and vertical bars indicate the number of unique or shared genes between clusters. Connected dots highlight shared DEGs across clusters, while single dots represent uniquely regulated DEGs.

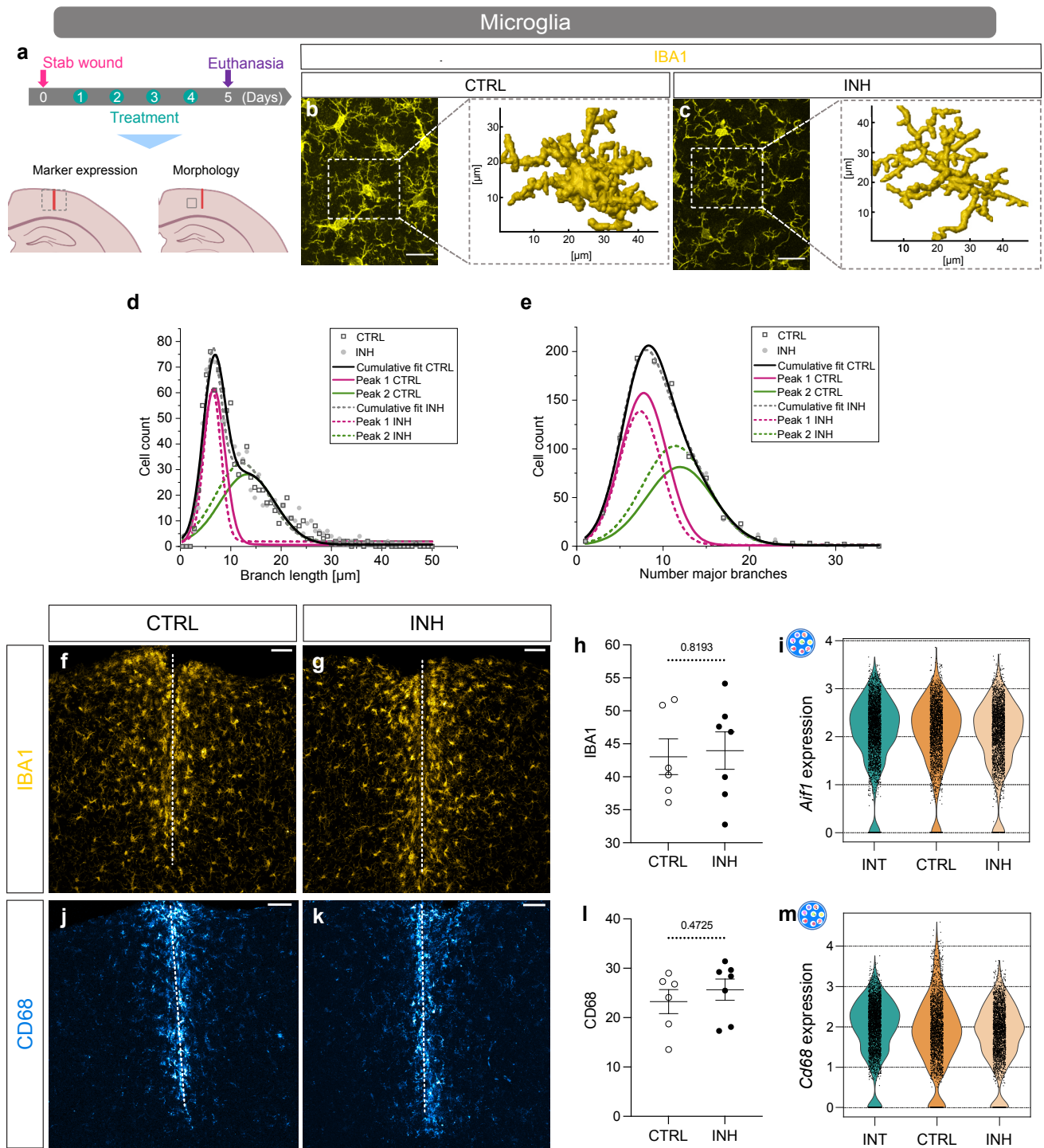
e,f Shankey diagram of GO terms linked to glial subclusters illustrating common and unique upregulated biological processes in response to Cxcr3 and Tlr1/2 pathway inhibition at 3 dpi (**e**) and 5 dpi (**f**) (Supplementary Data 7). The top 3 GO terms (based on adjusted p-value) for each glial subcluster are illustrated. The line width indicates the adjusted p-value. Significance was determined by performing GO analysis on gene set. Data shown in this figure are derived from n=12 intact animals over 3 independent experiments (5 scRNAseq libraries), n=3 3dpi CTRL animals over 1 experiment (2 scRNAseq libraries), n=3 3dpi INH animals over 1 experiment (2 scRNAseq libraries), n=9 5dpi CTRL animals over 3 independent experiments (3 scRNAseq libraries) and n=6 5dpi INH animals over 2 independent experiments (3 scRNAseq libraries). Abbreviations: DEGs = differentially expressed genes, dpi = days post injury, AG = astroglia, MG = microglia, OPCs = oligodendrocyte progenitor cells.



Supplementary Figure 13. OPC reactivity after injury is not altered by the Cxcr3 and Tlr1/2 pathway inhibition.

a,b UMAPs illustrating subclusters of oligodendrocytes (**a**) and cell distributions among these subclusters (**b**) at all time points (intact, 3 dpi, and 5 dpi) and conditions (CTRL and INH). Data in (**b**) are downsampled to an equal number of cells between time points and conditions. Data shown in this figure are derived from $n=12$ intact animals over 3 independent experiments (5 scRNAseq libraries), $n=3$ 3dpi CTRL animals over 1 experiment (2 scRNAseq libraries), $n=3$ 3dpi INH animals over 1 experiment (2 scRNAseq libraries), $n=9$ 5dpi CTRL animals over 3 independent experiments (3 scRNAseq libraries) and $n=6$ 5dpi INH animals over 2 independent experiments (3 scRNAseq libraries). **c** Experimental paradigm for assessing number of oligodendrocytes and OPC proliferation in injury vicinity. The dashed gray box on mouse brain scheme indicates the analyzed area. The red line highlights the injury core. **d-g** Representative overview images of proliferating OLIG2⁺ (gray) and EdU⁺ (magenta) cells in CTRL (**d**), and INH-treated (**g**) animals. The white dashed lines highlight injury cores. Micrographs in (**e,f,h,i**) are magnifications of the white boxed areas in (**d**) and (**g**), respectively. The white arrowheads in micrographs depict colocalization of EdU (**e,h**) with OLIG2⁺ (**f,i**) cells. All images are full z-projections of confocal z-stacks. **j,k** Dot plots depicting number of oligodendrocytes (OLIG2⁺ cells) (**j**) and proliferating OPCs (OLIG2⁺ and EdU⁺) (**k**) in CTRL and INH-treated animals. Data are shown as mean \pm standard error of the mean. Each data point represents one animal. Source data are provided

as Source Data file. Statistics in **(j)** and **(k)** have been derived from $n_{CTRL} = 6$ and $n_{INH} = 5$ animals. p-values were determined with unpaired t-test (two-tailed). Scale bars: **d,g**: 50 μm , **e, f, h, i**: 20 μm . Abbreviations: UMAP = uniform manifold approximation and projection, dpi = days post injury, EdU = 5-Ethynyl-2'-deoxyuridine, i.p. = intraperitoneal injection, CTRL = stab wound-injured control animals, INH = stab wound-injured inhibitor-treated animals, OPCs = oligodendrocyte progenitor cells, COPs = committed oligodendrocyte progenitors, MFOL = myelin-forming oligodendrocytes, MOL = mature oligodendrocytes.

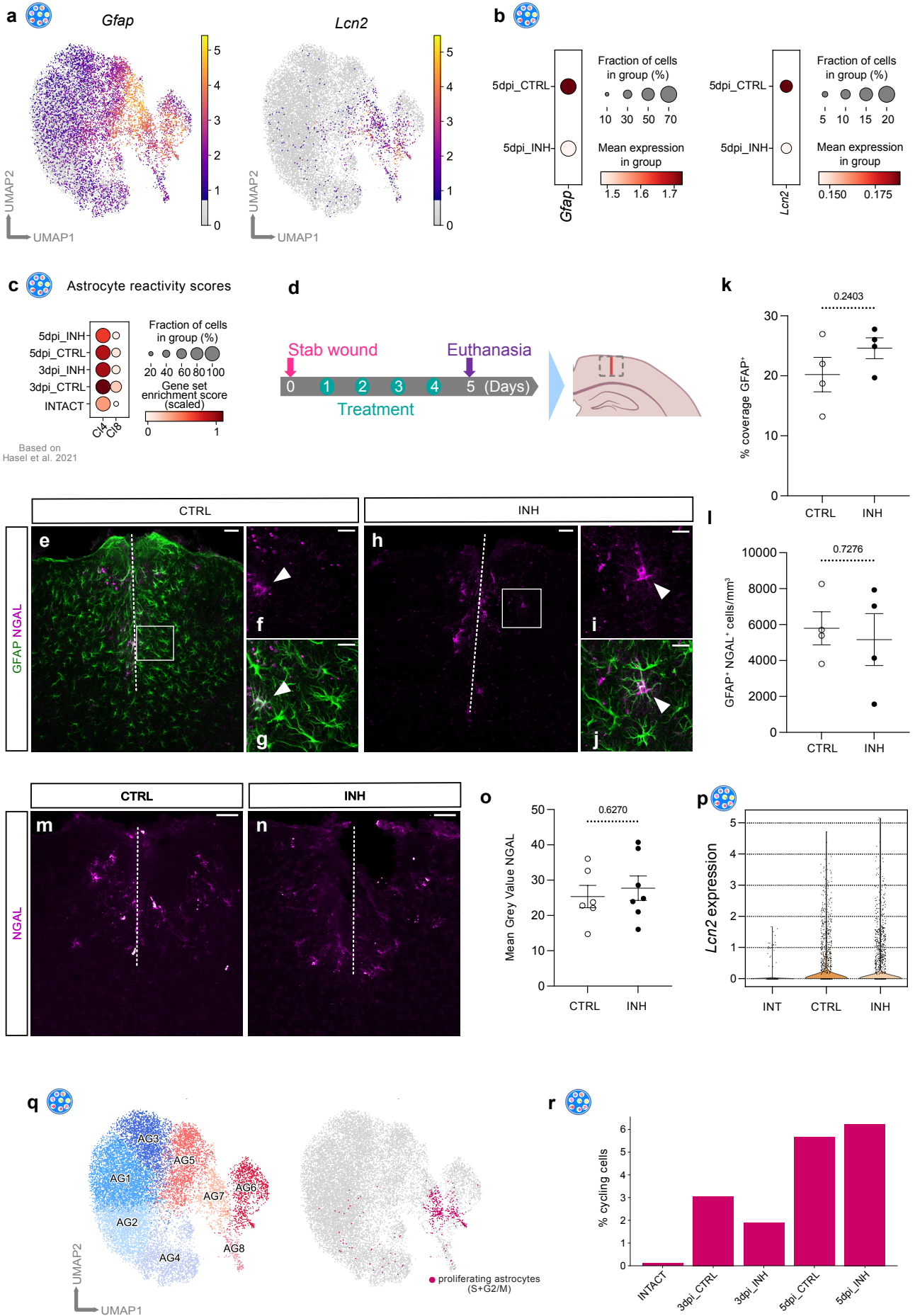


Supplementary Figure 14. Microglia reactivity upon Cxcr3 and Tlr1/2 pathway inhibition.

a Experimental paradigm for assessing microglial reactivity. The gray boxes on the mouse brain scheme highlight the analyzed areas. The red lines indicate the injury cores. **b,c** Representative images with 3D reconstruction of IBA1⁺ microglia (yellow) in CTRL (**b**) and INH-treated (**c**) mice. Dashed white boxes indicate selected microglia used for 3D reconstruction. **d,e** Plots depict the distribution of microglial branch length (**d**) and number of major branches (**e**) of $n=5$ CTRL and $n=6$ INH-treated animals. Source data are provided as Source Data file. Data are fitted with multiple peak functions. The parameters are depicted in Supplementary Data 8 for each fit. **f,g,j,k** Representative

images of IBA1 (**f,g**) and CD68 (**j,k**) staining in CTRL (**f,j**) and INH-treated (**g,k**) mice. Dashed white lines indicate the injury cores. All images are full z-projections of confocal z-stacks. **h,i** Dot plots depict the mean grey values of IBA1⁺ (**h**) and CD68⁺ (**i**) signal in the injury vicinity of CTRL and INH-treated mice. Data are shown as mean \pm standard error of the mean. Each data point represents one animal. Source data are provided as Source Data file. Statistics in (**h**) and (**i**) have been derived from $n_{CTRL} = 6$ and $n_{INH} = 7$ animals. p-values were determined with unpaired t-test (two-tailed). **i,m** Violin plots depicting expression levels of *Aif1* (**i**) and *Cd68* (**m**) in microglia (scRNA-seq) in intact, 5 dpi CTRL and 5 dpi INH condition. Data shown in violin plot are derived from $n=12$ intact animals over 3 independent experiments (5 scRNAseq libraries), $n=9$ 5dpi CTRL animals over 3 independent experiments (3 scRNAseq libraries) and $n=6$ 5dpi INH animals over 2 independent experiments (3 scRNAseq libraries). Scale bars: **f,g,j,k**: 50 μ m, **b,c**: 20 μ m. Abbreviations: dpi = days post injury, CTRL = stab wound- injured control animals, INH = stab wound-injured inhibitor-treated, INT = intact mice.

Astrocytes



Supplementary Figure 15. Reaction of astrocytes to stab wound injury in absence of *Cxcr3* and *Tlr1/2* signaling.

a,b UMAP (**a**) and dot plots (**b**) depicting the expression of *Gfap* and *Lcn2* among astrocytic subclusters. **c** Dot plot depicting astrocyte reactivity scores between all time points and conditions. Genes defining astrocyte reactivity scores were extracted from cluster_4 and 8 astrocytes of Hasel et al.³⁸ (Supplementary Table 7) and are plotted on the reactive subclusters. **d** Experimental paradigm for assessing astrocyte reactivity in the injury vicinity. Dashed gray box on mouse brain scheme indicates analyzed area. Red line highlights the injury core. **e-j** Representative images of GFAP⁺ astrocytes (green) and NGAL⁺ cells (magenta) in CTRL (**e**) and INH-treated (**h**) animals. White dashed lines highlight injury cores. Micrographs in (**f,g,i** and **j**) are magnifications of white boxed areas in (**e**) and (**h**). The white arrowheads in micrographs depict colocalization of NGAL with GFAP. All images are full z-projections of confocal z-stacks. **k,l** Dot plots depicting percentage of area covered with GFAP⁺ signal (**k**) and density of GFAP⁺ NGAL⁺ (protein of *Lcn2*) double positive astrocytes (**l**) in CTRL and INH-treated mice. Data are shown as mean \pm standard error of the mean. Each data point represents one animal. Source data are provided as Source Data file. Statistics have been derived from $n_{CTRL} = 4$ and $n_{INH} = 4$ animals. p-values were determined with unpaired t-test (two-tailed). **m,n** Representative overview images of NGAL distribution (magenta) in CTRL (**m**) and INH-treated (**n**) animals. **o** Dot plot depicting mean grey values of NGAL⁺ signal of CTRL and INH- treated mice. Data are shown as mean \pm standard error of the mean. Each data point represents one animal. Source data are provided as Source Data file. Statistics have been derived from $n_{CTRL} = 6$ and $n_{INH} = 7$ animals. p-values were determined with unpaired t-test (two-tailed). **p** Violin plots depicting expression levels of *Lcn2* in astrocytes (scRNA-seq). **q** UMAPs highlighting localization of proliferating astrocytes (pink). **r** Histogram illustrating percentage of proliferating astrocytes (based on S+G2/M score expression (Supplementary Table 4)) between all time points and condition. Data shown in this figure are derived from $n=12$ intact animals over 3 independent experiments (5 scRNAseq libraries), $n=3$ 3dpi CTRL animals over 1 experiment (2 scRNAseq libraries), $n=3$ 3dpi INH animals over 1 experiment (2 scRNAseq libraries), $n=9$ 5dpi CTRL animals over 3 independent experiments (3 scRNAseq libraries) and $n=6$ 5dpi INH animals over 2 independent experiments (3 scRNAseq libraries). Scale bars: **e,h,m,n**: 50 μ m, **f, g, i, j**: 20 μ m. Abbreviations: UMAP = uniform manifold approximation and projection, dpi = days post injury, INT = intact mice, CTRL = stab wound-injured control animals, INH = stab wound-injured inhibitor-treated animals.

Cluster	names	pvals	logFC	Cluster	names	pvals	logFC
I	<i>Cplx1</i>	1.89E-124	1.76	IX	<i>3110035E14Rik</i>	1.18E-54	2.25
	<i>Pvalb</i>	1.85E-106	2.16		<i>Rprm</i>	8.12E-45	3.03
	<i>Tmsb10</i>	1.80E-78	1.36		<i>Ttc9b</i>	1.90E-40	1.75
	<i>Mgst3</i>	8.69E-74	1.17		<i>Pcp4</i>	1.25E-37	2.17
	<i>Serpini1</i>	1.03E-73	1.41		<i>Cplx3</i>	1.11E-36	4.39
II	<i>Lamp5</i>	4.32E-130	2.63	X	<i>C1ql2</i>	5.68E-56	5.80
	<i>Calb1</i>	9.14E-134	2.79		<i>Fam163b</i>	1.07E-55	3.00
	<i>Camk2n1</i>	1.15E-111	1.76		<i>Ddn</i>	4.44E-49	2.12
	<i>Rasgrf2</i>	5.02E-117	2.58		<i>Btbd3</i>	7.11E-51	2.25
	<i>Igfbp6</i>	2.23E-86	2.00		<i>Orai2</i>	8.37E-49	2.81
III	<i>Fth1</i>	4.75E-311	2.21	XI	<i>Lamp5</i>	3.16E-24	2.08
	<i>Plp1</i>	4.13E-208	4.56		<i>Tshz2</i>	8.22E-23	3.10
	<i>Mbp</i>	7.94E-204	3.72		<i>Camk2n2</i>	1.70E-21	1.61
	<i>Mal</i>	3.80E-238	4.44		<i>Nrep</i>	7.83E-22	1.90
	<i>Mobp</i>	8.61E-205	3.96		<i>Sparcl1</i>	3.24E-20	0.85
IV	<i>3110035E14Rik</i>	7.17E-97	2.24	XII	<i>Calml4</i>	4.99E-91	7.53
	<i>Ttc9b</i>	6.56E-92	2.04		<i>Ttr</i>	4.93E-111	7.60
	<i>Rprm</i>	7.36E-93	3.31		<i>1500015O10Rik</i>	1.15E-92	7.05
	<i>Myl4</i>	4.83E-90	2.85		<i>Folr1</i>	4.86E-80	6.67
	<i>Pcp4</i>	7.09E-73	2.31		<i>Kl</i>	1.91E-70	6.94
V	<i>Cabp7</i>	2.12E-130	3.63	XIII	<i>Spink8</i>	1.93E-111	5.95
	<i>Cnih2</i>	7.44E-99	2.21		<i>Tmsb4x</i>	6.09E-68	2.01
	<i>Sepw1</i>	2.34E-72	0.91		<i>Wipf3</i>	2.23E-64	3.94
	<i>Neurod6</i>	6.58E-69	2.54		<i>Hpca</i>	4.97E-55	3.62
	<i>Hpca</i>	6.12E-63	2.06		<i>Rgs14</i>	1.07E-65	4.47
VI	<i>Serpina3n</i>	4.79E-72	3.30	XIV	<i>Gpr88</i>	2.84E-73	5.57
	<i>C1qc</i>	4.39E-66	2.65		<i>Ppp1r1b</i>	4.65E-70	4.64
	<i>Gfap</i>	2.99E-64	2.65		<i>Pde10a</i>	3.90E-62	5.29
	<i>Tyrobp</i>	6.83E-59	2.38		<i>Penk</i>	1.17E-61	5.64
	<i>Ifi2712a</i>	1.82E-55	3.78		<i>Tac1</i>	2.76E-50	5.08
VII	<i>ApoE</i>	1.18E-70	1.30	XV	<i>Calb2</i>	2.80E-48	6.18
	<i>mt-Co1</i>	1.41E-58	0.89		<i>Ramp3</i>	1.10E-41	4.92
	<i>mt-Nd5</i>	1.22E-58	1.17		<i>Tcf7l2</i>	7.64E-35	5.87
	<i>mt-Nd2</i>	1.35E-57	1.02		<i>Slc17a6</i>	3.02E-35	4.62
	<i>mt-Co2</i>	4.83E-55	1.02		<i>Zic1</i>	2.69E-32	5.21
VIII	<i>Camk2n1</i>	4.14E-70	1.90	XVI	<i>Tac2</i>	7.48E-42	8.23
	<i>Nrsn1</i>	6.58E-66	1.80		<i>Gng8</i>	8.03E-34	7.97
	<i>Plcxd2</i>	3.53E-73	2.61		<i>Zic1</i>	6.28E-43	6.42
	<i>Rorb</i>	1.36E-66	2.90		<i>Necab2</i>	3.60E-41	5.99
	<i>Cnih3</i>	5.63E-65	2.69		<i>Gm5741</i>	2.54E-23	9.60

Supplementary Table 1

Top 5 enriched genes in different stRNA-seq identified clusters. Significance was determined using the method t-test overestimated_variance.

Neuronal layers markers ³⁴	
Layer 2/3	<i>A830009L08Rik, Gm12371, Lamp5, Calb1, Igfbp6, A830009L08Rik</i>
Layer 4	<i>Krt12, Tcap, Dkk1, Rorb, Tnnc1, A830036E02Rik</i>
Layer 5	<i>Hs3st2, Vipr1, Pde1a, Scube1, Ml4, Dkk1, Fezf2, RP24-134N2.1, Nov, Coro6, Lamp5, Kcng1</i>
Layer 6	<i>Rprm, Rel1, Tbr1, 3110035E14Rik, Nxph3, Hs3st4, Myl4, Cpne4, Fezf2, Tshz2, Lypd1</i>

Supplementary Table 2

List of genes used for each of the cortical neuronal layer gene set scores.

Cell type	Gene	Reference
Microglia	<i>Tmem119, Cx3cr1, P2ry12, P2ry13, Gpr34, Olfm13, Selp1g, Sparc, Fcrls, Siglech, Slc2a5, Mafk, Pmepa1, Cd14</i>	45, 98, 99,100,101
Disease associated microglia (DAM)	<i>Lpl, Cst7</i>	45, 99, 102, 103
Macrophages	<i>Cd163, Mrc1, Lyve1, Siglec1, Itga4, Tgfbi, Ifitm2, Ifitm3, Tagln2, F13a1, Kynu, S100a11, S100a6</i>	45,104–106
Border Associated Macrophages (BAM)	<i>Cd163, Mrc1, Lyve1, Siglec1</i>	45, 107,108
Monocytes	<i>Ly6c2, Ccr2, Spn</i>	109, 110
Dendritic cells (DC)	<i>Cd24a, Itgax, Bst2, Cd209a, Xcr1, Ccr7, Ccr9, Flt3, Itgae, Itgam, Nudt17, Zbtb46, Ccr9, H2-Oa, Tnni2, Kctd14, Amica1, Kmo, Hepacam2</i>	45,111, 112, 113, 114
NKT cells	<i>Cd3e, Klrb1c, Ncr1</i>	45, 114
T cells	<i>Cd3e, Lat, Bcl11b</i>	45, 114
B cells	<i>Cd19, Ms4a1, Pax5</i>	45, 114
Neutrophils	<i>S100a9</i>	103, 114
Neurons	<i>Rbfox3, Slc17a7, Gabra1, Cux1, Pou3f2, Bcl11b, Foxp2</i>	34
OPCs	<i>Pdgfra, C1ql1</i>	34
COPs	<i>Neu4, Brca1</i>	34
MOL	<i>Opalin, Ninj2</i>	34
Astrocytes	<i>Sox9, Gfap, Aldh1l1, Atp1b2, Vim, Gfap, Slc1a3, Aldoc</i>	34
Pericytes	<i>Pecam1, Pdgfrb, Higd1b, Ndufa4l2, Rgs5, Kcnj8, Flt1</i>	34
VSMCs	<i>Acta2, Tagln, Myh11, Mustn1, Map3k7cl</i>	34
VLMCs	<i>Mgp, Slc47a1, Thbd, Col1a1, Col1a2</i>	34
VECV	<i>Ly6c1, Cldn5, Ly6a, Pltp, Pglyrp1, Scgb3a1</i>	34

Supplementary Table 3

List of marker genes of well-characterized cell populations, employed for cluster identification.

Cell Cycle genes ^{46, 47}			
S-phase		G2M-phase	
<i>Mcm5</i>	<i>Atad2</i>	<i>Hmgb2</i>	<i>Hjurp</i>
<i>Pcna</i>	<i>Rad51</i>	<i>Cdk1</i>	<i>Cdca3</i>
<i>Tyms</i>	<i>Rrm2</i>	<i>Nusap1</i>	<i>Hn1</i>
<i>Fen1</i>	<i>Cdc45</i>	<i>Ube2c</i>	<i>Cdc20</i>
<i>Mcm2</i>	<i>Cdc6</i>	<i>Birc5</i>	<i>Ttk</i>
<i>Mcm4</i>	<i>Exo1</i>	<i>Tpx2</i>	<i>Cdc25c</i>
<i>Rrm1</i>	<i>Tipin</i>	<i>Top2a</i>	<i>Kif2c</i>
<i>Ung</i>	<i>Dscc1</i>	<i>Ndc80</i>	<i>Rangap1</i>
<i>Gins2</i>	<i>Blm</i>	<i>Cks2</i>	<i>Ncapd2</i>
<i>Mcm6</i>	<i>Casp8ap2</i>	<i>Nuf2</i>	<i>Dlgap5</i>
<i>Cdca7</i>	<i>Usp1</i>	<i>Cks1b</i>	<i>Cdca2</i>
<i>Dtl</i>	<i>Clspn</i>	<i>Mki67</i>	<i>Cdca8</i>
<i>Prim1</i>	<i>Pola1</i>	<i>Tmpo</i>	<i>Ect2</i>
<i>Uhrf1</i>	<i>Chaf1b</i>	<i>Cenpf</i>	<i>Kif23</i>
<i>Mlf1ip</i>	<i>Brip1</i>	<i>Tacc3</i>	<i>Hmmr</i>
<i>Hells</i>	<i>E2f8</i>	<i>Fam64a</i>	<i>Aurka</i>
<i>Rfc2</i>		<i>Smc4</i>	<i>Psrc1</i>
<i>Rpa2</i>		<i>Ccnb2</i>	<i>Anln</i>
<i>Nasp</i>		<i>Ckap2l</i>	<i>Lbr</i>
<i>Rad51ap1</i>		<i>Ckap2</i>	<i>Ckap5</i>
<i>Gmnn</i>		<i>Aurkb</i>	<i>Cenpe</i>
<i>Wdr76</i>		<i>Bub1</i>	<i>Ctcf</i>
<i>Slbp</i>		<i>Kif11</i>	<i>Nek2</i>
<i>Ccne2</i>		<i>Anp32e</i>	<i>G2e3</i>
<i>Ubr7</i>		<i>Tubb4b</i>	<i>Gas2l3</i>
<i>Pold3</i>		<i>Gtse1</i>	<i>Cbx5</i>
<i>Msh2</i>		<i>Kif20b</i>	<i>Cenpa</i>

Supplementary Table 4

List of genes used to calculate the cell cycle phase score.

	Shared inflammatory genes	
	3dpi - downregulated	5dpi- downregulated
AG1	-	-
AG2	-	<i>Ifitm3</i>
AG3	-	-
AG4	-	-
AG5	<i>Rsad2, Ifit3, Ifit3b, Bst2</i>	<i>Ifitm3, Bst2</i>
AG6	<i>Isg15, Bst2, Rtp4, Ifit3, Ifit3b</i>	<i>Isg15</i>
AG7	<i>Isg15, Cxcl10, Oasl2, Ifitm3, Irf7, Bst2, Trim25, Rsad2, Rtp4, Ifit3, Ifit3b, Ifit1</i>	<i>Isg15, Irf7, Ifi2712a, Rtp4</i>
AG8	<i>Rsad2, Ifi2712a</i>	-
MG1	<i>Ifitm3</i>	-
MG2	-	<i>Ifitm3</i>
MG3	<i>Isg15, Ifitm3, Bst2, Ifi2712a, Rtp4, Ifit3</i>	
MG4	<i>Zbp1, Isg15, Cxcl10, Oasl2, Oasl1, Oas1a, Ifitm3, Irf7, Bst2, Rsad2, Rtp4, Ifit3, Ifit3b</i>	<i>Isg15, Cxcl10, Ifitm2, Irf7, Bst2, Rsad2, Lgals1, Ifit3, Ifit1</i>
MG5	-	<i>Isg15, Ifitm3, Irf7</i>
MG6	<i>Zbp1, Isg15, Oasl2, Oas1a, Ifitm3, Irf7, Bst2, Dhx58, Ifi2712a, Rtp4, Ifit3</i>	-
MG7	-	<i>Bst2, Ifi2712a, Lgals1</i>
MG8	-	-
OPCs1	<i>Ifitm3</i>	-
OPCs2	<i>Isg15, Oasl2, Oas1a, Ifitm3, Irf7, Bst2, Rsad2, Ifi2712a, Lgals1, Rtp4, Ifit3, Ifit3b, Ifit1</i>	<i>Bcl3, Ifitm2, Ifitm3, Bst2, Ifi2712a, Lgals1</i>
OPCs3	-	-

Supplementary Table 5

List of shared inflammatory genes which are downregulated after INH treatment in each subcluster.

Sample-id (Matrix)	FastQ	Condition	Chromium Kit	Sequencing platform	cellranger	number of cells	mean genes	mean counts
21L008532	21L008532, 22L000025	5dpi_CTRL	V3.1	Novaseq6000	6.0.0	7440	3212	10960
21L008533	21L008533, 22L000026	5dpi_INH	V3.1	Novaseq6000	6.0.0	7403	3334	11696
MUC29190	MUC29190	Intact	V3.1	Novaseq6000	6.0.0	4357	2595	7301
MUC18415	MUC18415	5dpi_CTRL	V2	Novaseq6000	3.0.2	4615	2759	9150
MUC13721	MUC13721	Intact	V2	HiSeq 3000/4000	3.0.2	2674	2121	5674
MUC13722	MUC13722	3dpi_CTRL	V2	HiSeq 3000/4000	3.0.2	1848	2306	7934
MUC13723	MUC13723	3dpi_CTRL	V2	HiSeq 3000/4000	3.0.2	1795	2403	8731
MUC13724	MUC13724	3dpi_INH	V2	HiSeq 3000/4000	3.0.2	1865	2377	8036
MUC13725	MUC13725	3dpi_INH	V2	HiSeq 3000/4000	3.0.2	2748	2420	8259
MUC13726	MUC13726	Intact	V2	HiSeq 3000/4000	3.0.2	2471	2069	5360
MUC13727	MUC13727	5dpi_CTRL	V2	HiSeq 3000/4000	3.0.2	1711	2385	7665
MUC13729	MUC13729	5dpi_INH	V2	HiSeq 3000/4000	3.0.2	3831	2199	6306
MUC13730	MUC13730	5dpi_INH	V2	HiSeq 3000/4000	3.0.2	3908	2125	6063
MUC13731	MUC13731	Intact	V2	HiSeq 3000/4000	3.0.2	3385	1764	4443
MUC13732	MUC13732	Intact	V2	HiSeq 3000/4000	3.0.2	3762	1782	4505

Supplementary Table 6

Specification of datasets used for the scRNA-seq analysis.

Genes defining astrocyte reactivity scores - extracted from cluster_4 and 8 astrocytes of Hasel et al. ³⁸	
Ci4	Ci8
<i>Lcn2</i>	<i>Igtp</i>
<i>Ifitm3</i>	<i>Gm4951</i>
<i>H2-D1</i>	<i>Iigp1</i>
<i>Timp1</i>	<i>Irgm1</i>
<i>B2m</i>	<i>Tap1</i>
<i>Vim</i>	<i>Ifit3</i>
<i>Gfap</i>	<i>Gbp2</i>
<i>Psmb8</i>	<i>F830016B08Rik</i>
<i>Bst2</i>	<i>Gbp7</i>
<i>H2-T23</i>	<i>Tap2</i>
<i>Cd63</i>	<i>Ifi47</i>
<i>Serping1</i>	<i>B2m</i>
<i>Tspo</i>	<i>Ifit1</i>
<i>Rps4x</i>	<i>Psmb8</i>
<i>S100a6</i>	<i>Stat1</i>
<i>Tuba1a</i>	<i>H2-T23</i>
<i>H2-K1</i>	<i>Usp18</i>
<i>Mt1</i>	<i>Ifit3b</i>
<i>Gpx1</i>	<i>Rtp4</i>
<i>S100a11</i>	<i>Psmb9</i>

Supplementary Table 7

List of genes used to calculate the gene set score in Supplementary Fig. 15c.

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