

Supplementary Information

Eph-ephrin signaling couples endothelial cell sorting and arterial specification

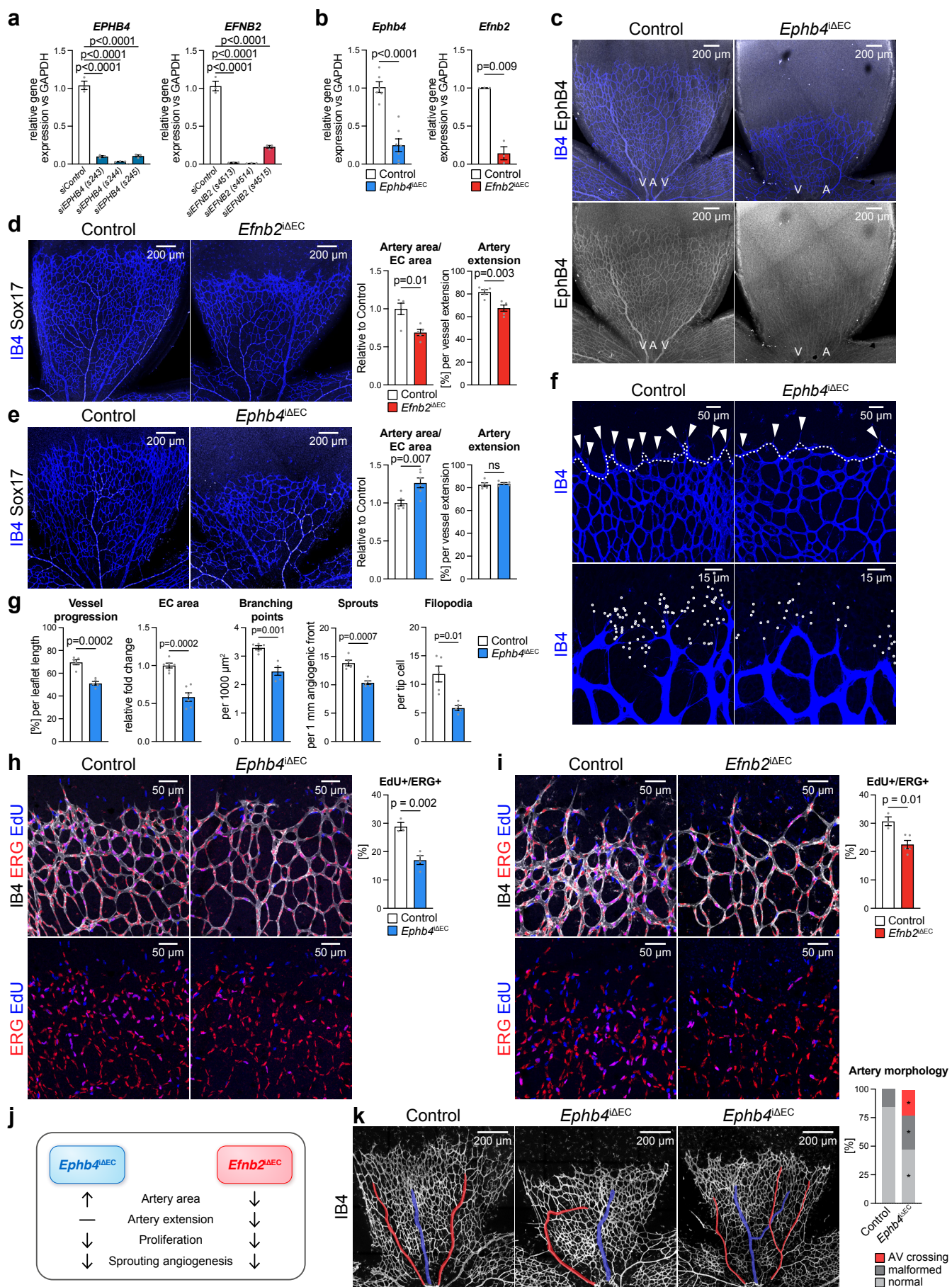
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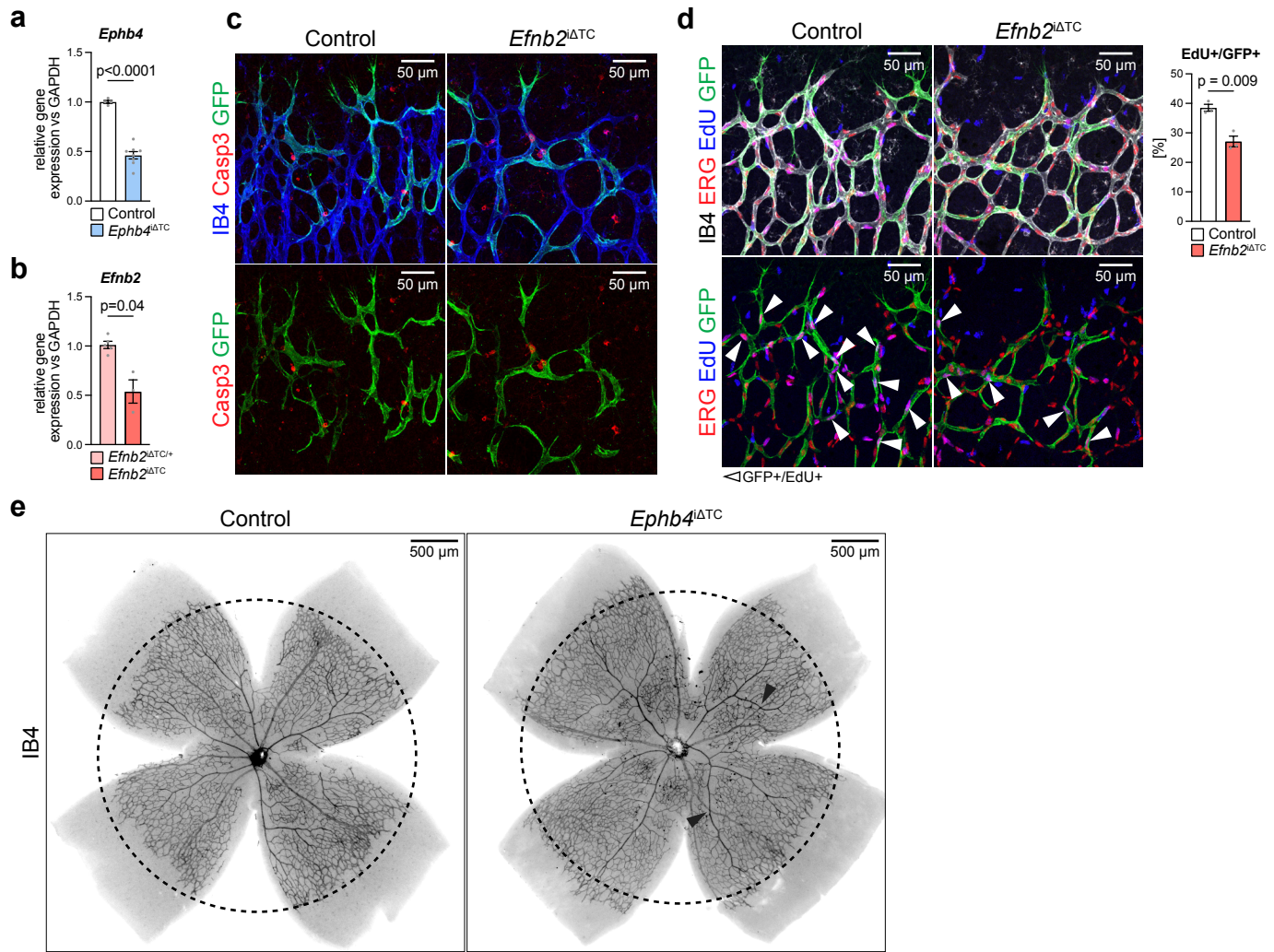
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Supplementary Figure 1: Common and distinct roles of EphB4 and ephrin-B2 in the retinal angiogenesis.

(a) Validation of *siEPHB4* and *siEFNB2* efficiency by RT-qPCR for 3 different siRNAs (n=3 experiments). (b) Validation of *Ephb4* and *Efnb2* gene deletion using *Cdh5CreERT2*. RT-qPCR of FACS sorted retinal ECs (n=6 control and 7 *Ephb4*^{iΔEC} and n=3 control and 3 *Efnb2*^{iΔEC} mice). (c) Confirmation of *Ephb4* inactivation. IB4 and EphB4 staining of control and *Ephb4*^{iΔEC} retinas. (d, e) Confocal images of isolectin B4 (IB4) and Sox17 stained P6 retinal vessels (n=5 control and 5 *Efnb2*^{iΔEC} mice (d) and n=6 control and 6 *Ephb4*^{iΔEC} mice (e)). Quantitation of arterial area per total EC area and of artery extension defined by Sox17 arterial immunosignal. (f, g) EphB4 is required for sprouting angiogenesis. High magnification pictures of IB4 stained retinas (f). Graphs show quantitation of vessel progression (n=5 control and *Ephb4*^{iΔEC}), EC area (n=6 mice), branching points (n=5 mice), number of sprouts (n=5 mice) and filopodia per angiogenic front line (n=5 mice) (g). (h, i) Reduced proliferation of ECs in pan-endothelial *Ephb4* and *Efnb2* mutants. Graphs show quantitation of EdU+ per total ERG+ ECs (n=3 control and 4 *Ephb4*^{iΔEC} and n=3 control and 5 *Efnb2*^{iΔEC} mice). (j) Scheme summarizes vascular changes in mutant retinas. (k) Representative images of *Ephb4*^{iΔEC} retina with malformed arteries and arteriovenous crossings compared to control. Graph shows quantitation of malformed arteries and normal vascular morphology (n= 19 control and 17 *Ephb4*^{iΔEC}). *P* values were calculated by one-way ANOVA (a), two-tailed unpaired *t*-test (b, d, e, g, h, i) and Chi-square test (k). *In vivo* experiments were performed with tamoxifen injections at P1-P3 following analysis at P6 (c, d, e, f, h, i, k). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.

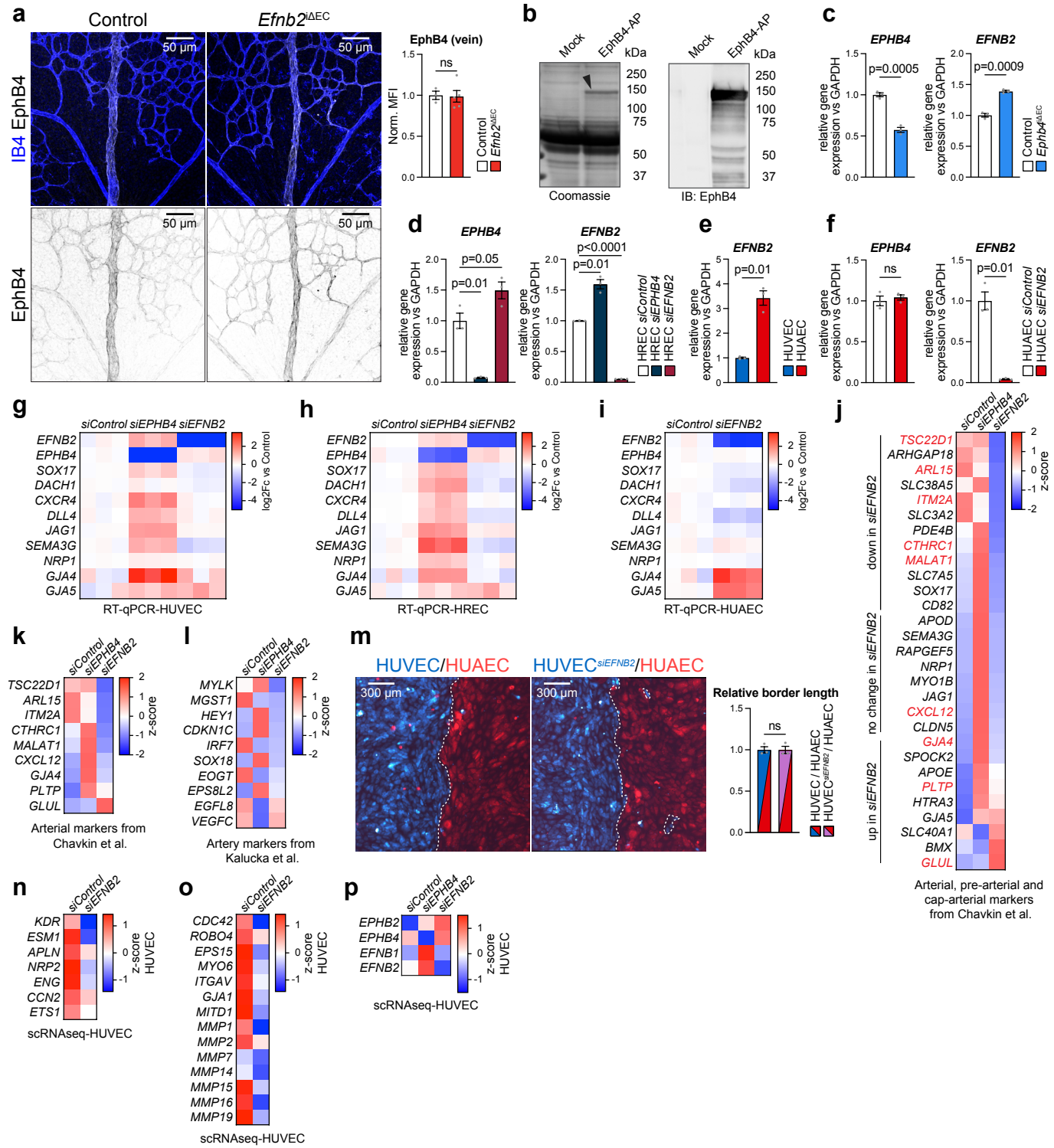
Supplementary Figure 2: Tip cell deletion of *Ephb4* and *Efnb2*.



Supplementary Figure 2: Tip cell deletion of *Ephb4* and *Efnb2*.

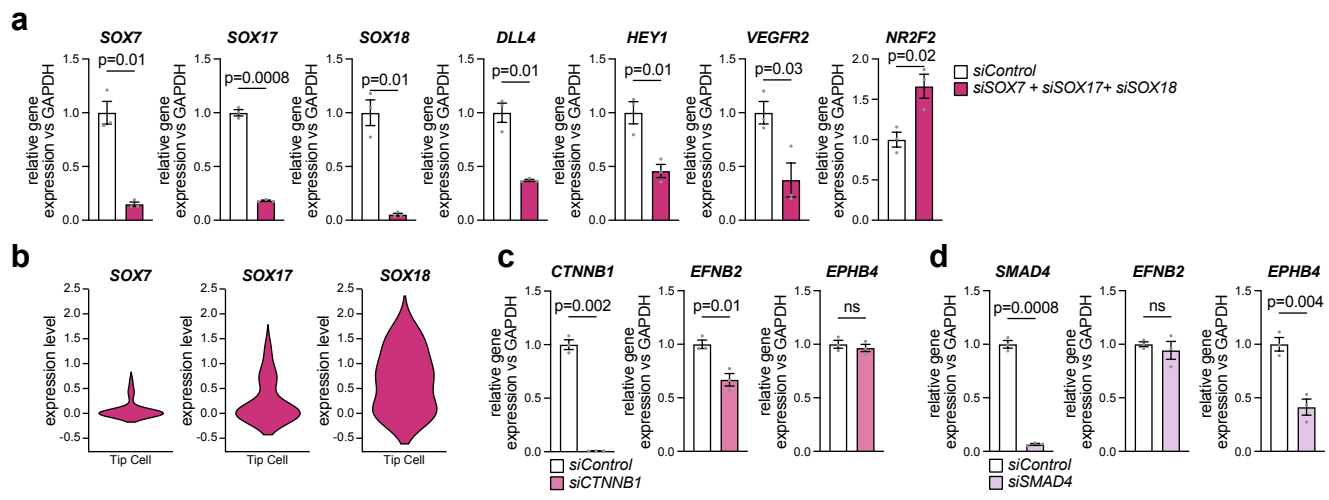
(a, b) Validation of *Ephb4* and *Efnb2* gene inactivation using *Esm1-CreERT2*. RT-qPCR of FACS sorted GFP+ retinal ECs (n=4 control and 8 *Ephb4*^{iΔTC} and n=5 control (*Efnb2*^{iΔTC/+}) and 3 *Efnb2*^{iΔTC} mice). (c) Confocal images of control and *Efnb2*^{iΔTC} P6 retinas showing IB4+, GFP+ and Casp3+ cells. Note that there are no GFP+/Casp3+ ECs (n=3 mice). (d) EC proliferation (arrowheads) is reduced in *Efnb2* depleted tip cell progeny. Confocal images and quantitation of proliferating GFP+ ECs using IB4, ERG, EdU and GFP immunostaining (n=3 control and 3 *Efnb2*^{iΔTC} mice). (e) IB4-stained overview pictures of control and *Ephb4*^{iΔTC} mouse retinas. Arrowheads point to AV crossings. *P* values were calculated using two-tailed unpaired *t*-test (a, b, d). *In vivo* experiments were performed with tamoxifen injections at P1-P3 following analysis at P6 (c, d, e). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.

Supplementary Figure 3: Ephrin-B2 represses EphB4 expression but promotes VEGFR2 signaling.



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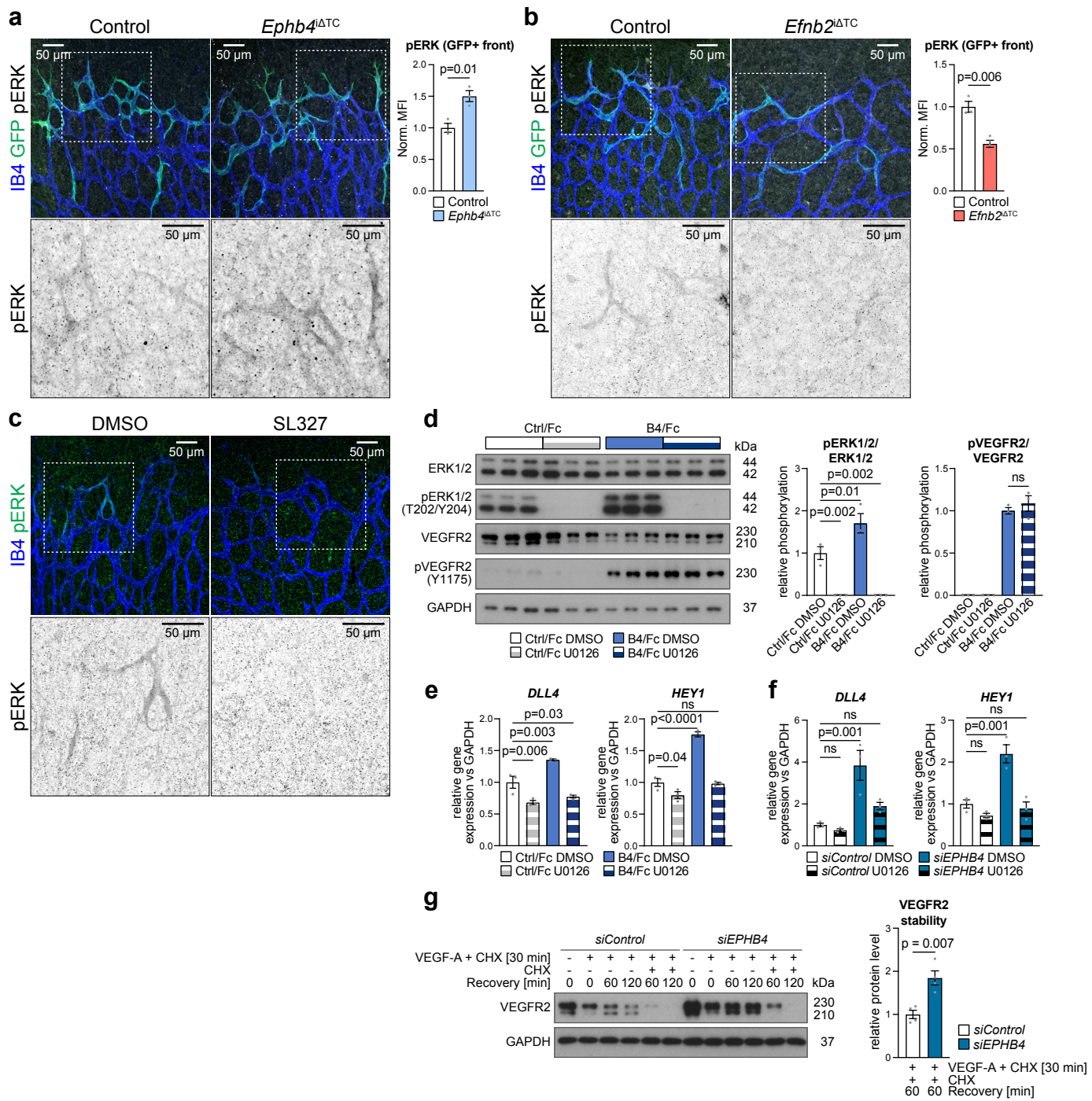
(a) EphB4 is unchanged in *Efnb2*^{iΔEC} mature retinal vessels and central capillary plexus. EphB4 staining and immunosignal quantitation (n=3 control and 5 *Efnb2*^{iΔEC} mice). (b) EphB4-AP protein expression. Coomassie blue and immunoblot detection of overexpressed EphB4-AP protein in HEK293 cell supernatant. (c) RT-qPCR of indicated genes in FACS-isolated P6 brain ECs (n=3 control and 3 *Ephb4*^{iΔEC}). (d, e, f) RT-qPCR of *EPHB4* and *EFNB2* in primary human retinal ECs (HRECs) upon *EPHB4* or *EFNB2* depletion (d), in HUVECs and HUAECs (e) and upon knockdown of *EFNB2* in HUAECs (f) (n=3 experiments). (g, h, i) RT-qPCR of indicated genes upon *EPHB4* and *EFNB2* knockdown in HUVECs (g) and HRECs (h) and upon knockdown of *EFNB2* in HUAECs (i) (n=3 experiments). (j, k, l) Comparative expression analysis of top arterial marker genes from P6 postnatal retina (Chavkin et al.⁵²) and adult mouse brain (Kalucka et al.⁵³) dataset, respectively, in *siControl*, *siEPHB4* and *siEFNB2* HUVEC scRNAseq own dataset. Z-score for tip cell-like main cluster was calculated in (j) for arterial (marked in red), pre-arterial and cap-arterial marker genes and in (k) only for arterial marker genes from Chavkin et al. retina dataset⁵², while in (l) for arterial marker genes from Kalucka et al. brain dataset⁵³. (m) Confrontation assay for heterotypic co-culture of HUVECs and HUAECs, as indicated. Final snapshots of CellTracker-labeled cells imaged for 48 h after removal of Ibidi insert. Quantitation graph for relative border length (n=3 experiments). (n, o) Decreased expression of genes related to angiogenesis (n) and cell migration (o) upon *EFNB2* KD in HUVECs compared to control. Genes selected from pseudobulk DGE analysis comparing *siControl* and *siEFNB2* in tip cell-like main cluster. (p) EphB and ephrin-B family genes are differentially affected by KD of EphB4 and ephrin-B2 in HUVECs. Results are based on pseudobulk DGE analysis comparing *siControl*, *siEPHB4* and *siEFNB2* in tip cell-like main cluster. *P* values were calculated using two-tailed unpaired *t*-test (a, c-i, m). *In vivo* experiments were performed with tamoxifen injections at P1-P3 following analysis at P6 (a). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.



Supplementary Figure 4: Regulation of *EFNB2* and *EPHB4* expression in HUVECs.

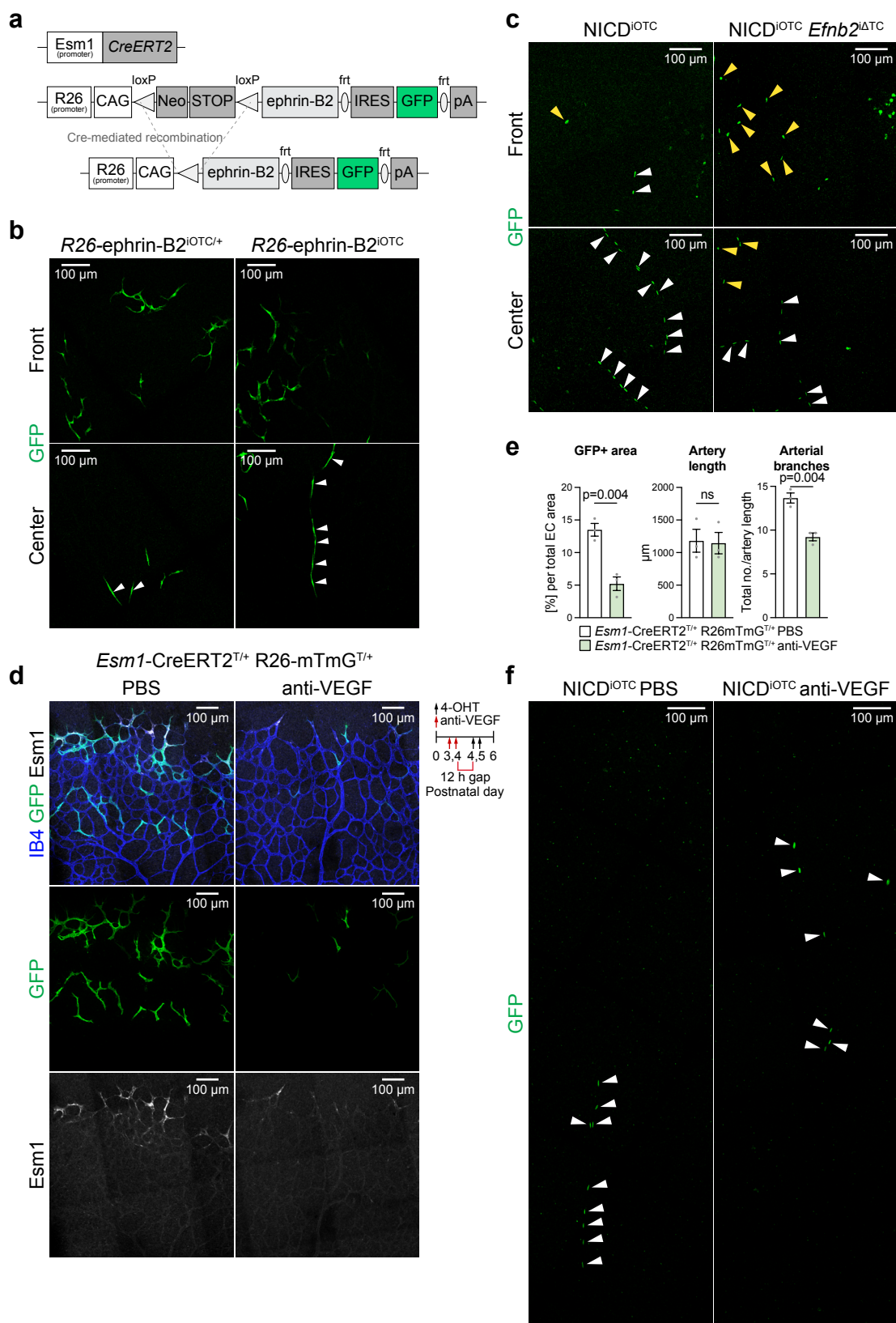
(a) SoxF factors promote arterial and repress venous gene expression. Validation by RT-qPCR of *SOX7* + *SOX17* + *SOX18* triple knockdown in HUVECs (n=3 experiments). (b) Violin plots of *SOX7*, *SOX17* and *SOX18* expression in P6 retinal tip cells. Data extracted from published scRNA-seq results⁵². (c, d) Not all transcription factors involved in vascular development and AV specification mediate reciprocal regulation of *EPHB4* and *EFNB2*. Graphs show RT-qPCR analysis of *siCTNNB1* vs *siControl* HUVECs (c) and *siSMAD4* vs *siControl* HUVECs (d) (n=3 experiments). *P* values were calculated using two-tailed unpaired *t*-test (a, c, d). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.

Supplementary Figure 5: ERK1/2 signaling promotes arterial gene expression.



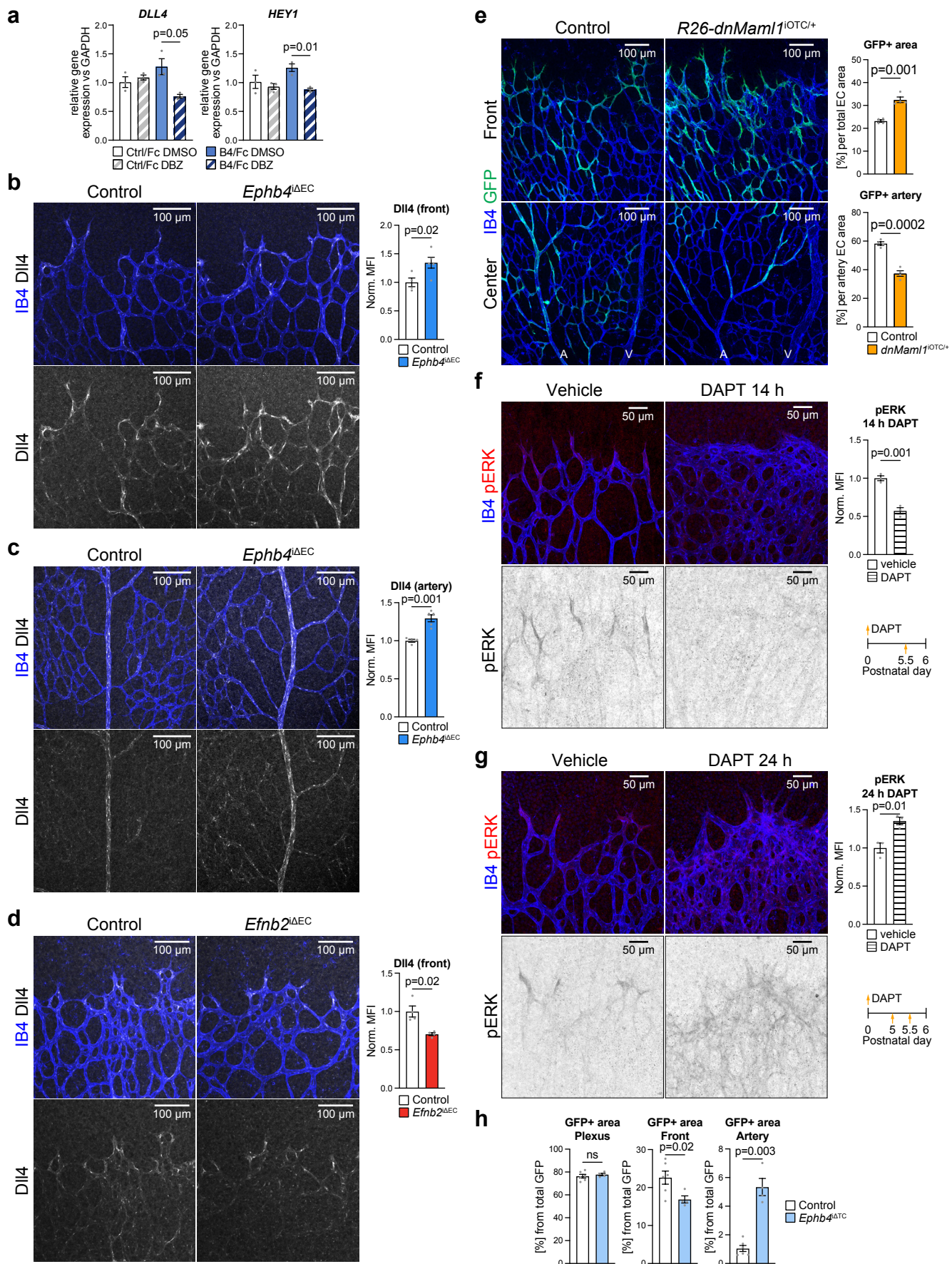
Supplementary Figure 5: ERK1/2 signaling promotes arterial gene expression.

(a) Increased pERK1/2 (pERK) in *Ephb4*-deleted TC-derived progeny (n=3 control and 3 *Ephb4*^{iΔTC}). (b) Decreased pERK in *Efnb2*-deleted TC-derived progeny (n=3 control and 4 *Efnb2*^{iΔTC}). (c) Validation of pERK retinal immunosignal in mice treated with SL327 MEK inhibitor or vehicle. (d) VEGFR2 and ERK1/2 activation upon EphB4/Fc stimulation. Immunoblot for pVEGFR2 (Y1175) and pERK1/2 from stimulated HUVECs, treated with ERK inhibitor (U0126) or vehicle. Immunoblot quantitation of pVEGFR2 and pERK1/2 relative levels upon U0126 inhibitor or vehicle treatment (n=3 experiments). (e, f) ERK signaling is required downstream of ephrin-B2 stimulation (e) and *EPHB4* knockdown (f) to increase *DLL4* and *HEY1* expression. RT-qPCR for *HEY1* and *DLL4* (n=3 experiments). (g) Increased VEGFR2 stability in *EPHB4* KD cells. VEGFR2 immunoblotting of *siControl* and *siEPHB4* HUVECs pre-treated with VEGF-A and cycloheximide (CHX), washed and recovered in the absence or presence of CHX. Graph shows quantitation of VEGFR2 levels at 60 min of recovery (+ CHX) relative to 0 min (VEGF-A+CHX) (n=3 experiments). *P* values were calculated using two-tailed unpaired *t*-test (a, b, g) and one-way ANOVA (d, e, f). *In vivo* experiments were performed with 4-OHT injections at P4.5 following analysis at P6 (a, b). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.



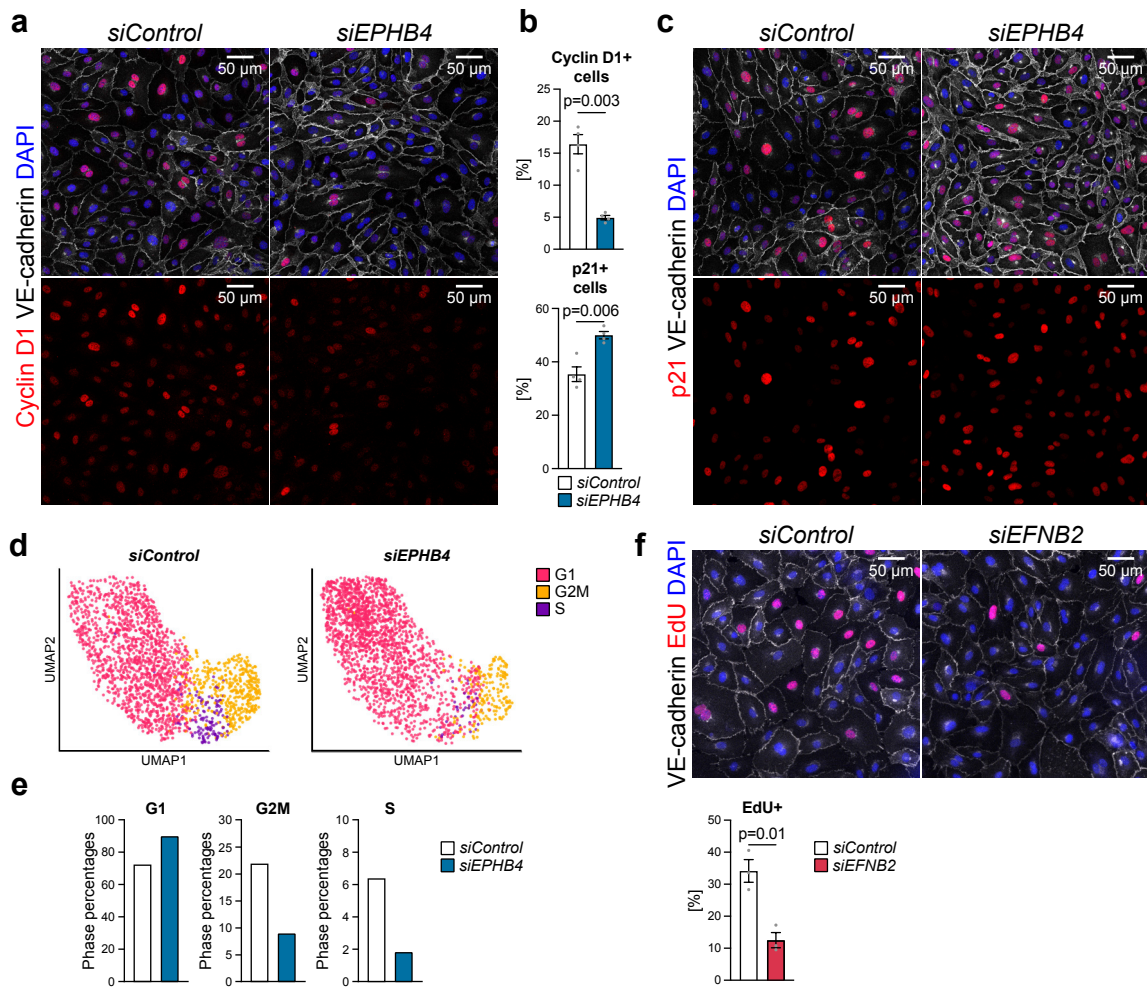
Supplementary Figure 6: Notch and VEGF-A control arterial progenitors in the retina.

(a) Schematic depiction of the *Esm1-CreERT2* transgene and Cre-mediated ephrin-B2 overexpression. (b) Confocal images showing GFP+ ephrin-B2 overexpressing cells. (c) Confocal images of GFP+ (NICD+ cells) ECs in the NICD^{iOTC} P6 retinal vasculature. (d) Induction of *Esm1* expression in tip cells requires VEGF signaling. Confocal images of vehicle or anti-VEGF treated *Esm1-CreERT2 R26-mTmG^{+T}* retinas showing IB4+, *Esm1*+ and GFP+ (*Esm1*-derived) ECs. (e) Quantitation of GFP+ area per total EC area, artery length and arterial branches (n=3 vehicle and n=3 anti-VEGF). (f) Confocal images of vehicle or anti-VEGF treated NICD^{iOTC} retinas showing GFP+ (NICD+) ECs. *P* values were calculated using two-tailed unpaired *t*-test (c). *In vivo* experiments were performed with tamoxifen injections at P1-P3 (b, c), 4-OHT injections at P4-P5 following analysis at P6 (d) or 4-OHT injections at P3-P4 followed by anti-VEGF injections at P5-P6 and analysis at P7 (f). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.



Supplementary Figure 7: Interactions between EphB4 and Notch during artery formation.

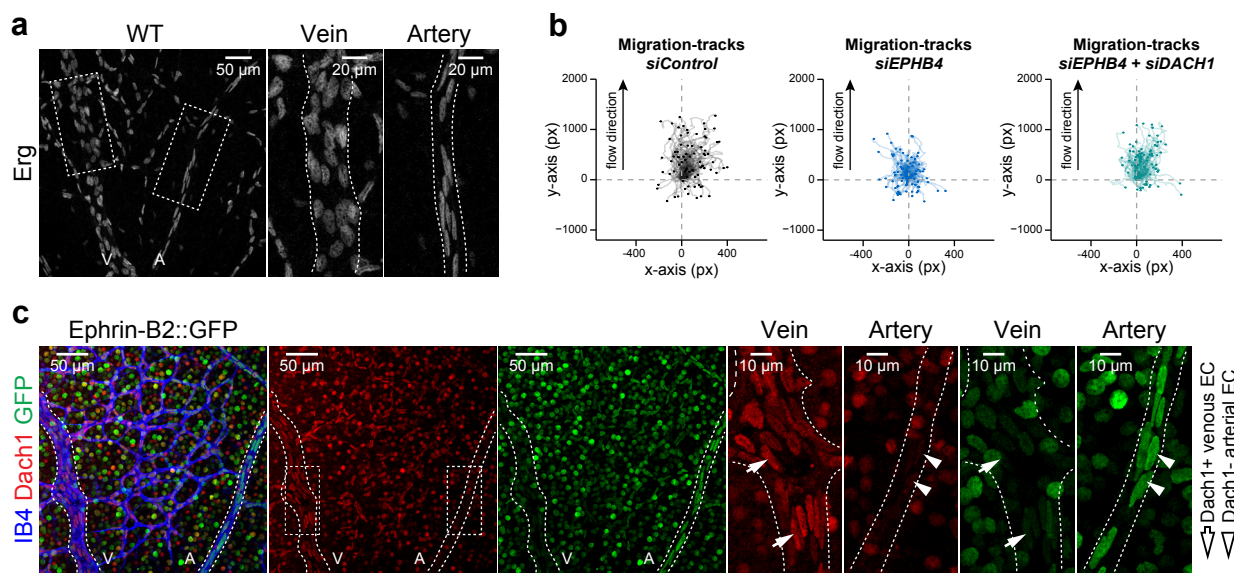
(a) Normalization of *DLL4* and *HEY1* expression upon DBZ treatment of EphB4/Fc stimulated cells (n=3 experiments). (b, c) Quantitation of Dll4 immunosignal in the P6 control (n=4 animals) and *Ephb4*^{i Δ EC} (n= 5) vascular front (b) and retinal arteries (c). (d) Decreased Dll4 capillary plexus expression upon EC-specific *Efnb2* inactivation. Quantitation of Dll4 MFI in the retinal vascular front area (n=4 control and 4 *Efnb2*^{i Δ EC}). (e) dnMaml1 overexpression in tip cells impairs artery growth. Graphs show quantitation of total GFP+ area and GFP+ arterial area (n=4 control and 4 *R26*-dnMaml1^{iOTC/+}). (f) Acute Notch inhibition (DAPT) for 14 h reduces pERK1/2. Graph shows quantitation of pERK MFI in angiogenic front area (n=3 vehicle and n=3 DAPT injected in WT C57BL6 mice). (g) Robust Notch inhibition (DAPT) for 24 h increased pERK. Graph shows quantitation of pERK MFI in angiogenic front area (n=3 vehicle and n=3 DAPT injected in WT C57BL6 mice). (h) Graphs showing the distribution of GFP+ area in capillary plexus, front and artery per total GFP positive area (n=6 control and 4 *Ephb4*^{i Δ TC} mice). *P* values were calculated by one-way ANOVA (a) and two-tailed unpaired *t*-test (b, c, d, e, f, g, h). *In vivo* experiments were performed with tamoxifen injections at P1-P3 following analysis at P6 (b, c, d, e). DAPT or vehicle was administered once, 14 h prior to analysis (f) or twice, at 24 h and 14 h prior to analysis (g). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.



Supplementary Figure 8: EphB4 and ephrin-B2 in cell cycle regulation.

(a-c) Decreased Cyclin D1 and increased P21 protein expression in *siEPHB4* cells at 48 h of knockdown (n=3 experiments). (d, e) *siEPHB4* cells show impaired cell cycle progression, as indicated by 2D UMAP representation of cell cycle phase distribution relative to *siControl* cells (d). Graphs represent proportion of cells in G1, G2/M and S phases (e). (f) Reduced proliferation of HUVECs after *EFNB2* KD. Graphs shows quantitation of EdU+ per total cells. *P* values were calculated by two-tailed unpaired *t*-test (b, f). Data are presented as mean values +/- SEM. Source data are provided as a Source Data file.

Supplementary Figure 9: EphB4, ephrin-B2 and Dach1 expression levels correlate with EC nuclear morphology and cell migration behavior under flow.



Supplementary Figure 9: EphB4, ephrin-B2 and Dach1 expression levels correlate with EC nuclear morphology and cell migration behavior under flow.

(a) Arterial ECs align better to flow than venous cells. Example of wild-type (WT) P6 retinal vasculature immunostained for ERG. Boxed areas show representative parts of artery (A) or vein (V). **(b)** Examples of quantified migration tracks for *siControl*, *siEPHB4* and *siEPHB4/DACH1* cells (80 randomly selected tracks/experiment). **(c)** Dach1 is expressed in retinal veins but not in arteries. IB4, GFP and Dach1 staining of the P6 *Efnb2-H2B-GFP* knock-in reporter retinal vasculature. Source data are provided as a Source Data file.

Supplementary Table S1.

Summary of genetic mouse models used in this study

Full name	Short name	Source/Reference
<i>Rosa26-mT/mG</i>	<i>R26-mTmG</i>	Muzumdar et al. 2007
<i>Ephb4</i> ^{lox/lox}	/	Wang et al. 2015
<i>Efnb2</i> ^{lox/lox}	/	Grunwald et al. 2004
<i>Cdh5(PAC)-CreERT2</i>	<i>Cdh5-CreERT2</i>	Wang et al. 2010
<i>Esm1-CreERT2</i>		Rocha et al. 2014
<i>Gli(ROSA)</i> ^{26Sortm1(Natch1)dam}	<i>NICD</i>	Murtaugh et al. 2003
<i>Efnb2-GFP</i>		Davy and Soriano, 2007
<i>C57BL/6J</i>	WT	Janvier Labs
<i>Rosa26-ephrin-B2-GFP</i>	<i>R26-ephrin-B2-GFP</i>	this study
<i>Rosa26-Dach1OE</i>	<i>R26-Dach1</i>	Raffrey et al. 2021
<i>Gli(ROSA)</i> ^{26Sortm1(MAML1)Wsp}	<i>R26-dnMaml1</i>	Tu et al. 2005

Control genotype	Mutant genotype	Tamoxifen treatment	Figure
<i>Efnb2</i> ^{lox/lox}	<i>Efnb2</i> ^{lox/lox} , <i>Cdh5(PAC)-CreERT2</i> ^{+/-}	P1-P3 50 µg	2d, 3c, S1d, S1i, S3a, S7d
<i>Ephb4</i> ^{lox/lox}	<i>Ephb4</i> ^{lox/lox} , <i>Cdh5(PAC)-CreERT2</i> ^{+/-}	P1-P3 50 µg	2f, 3a, 4c, 5d, S1c, S1e, S1f, S1h, S1k, S7b, S7c
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Ephb4</i> ^{lox/lox} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P1-P3 50 µg	1c, 1g, 1i, 6b, 7a, S2e
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Efnb2</i> ^{lox/lox} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P1-P3 50 µg	1d, 1h, 7b, S2c, S2d
<i>NICD</i> ^{lox/lox} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Efnb2</i> ^{lox/lox} , <i>NICD</i> ^{lox/lox} , <i>Esm1-CreERT2</i> ^{+/-}	P1-P3 50 µg	5b
<i>R26-ephrin-B2-GFP</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>R26-ephrin-B2-GFP</i> ^{lox/lox} , <i>Esm1-CreERT2</i> ^{+/-}	P1-P3 50 µg	5a
<i>Ephb4</i> ^{lox/lox} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Ephb4</i> ^{lox/lox} , <i>R26-dnMaml1</i> ^{lox/+} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P1-P3 50 µg	6c
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Ephb4</i> ^{lox/lox} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P4.5 50 µg 4OHT	6g, 9h, S5a
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Ephb4</i> ^{lox/lox} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P4.5 50 µg 4OHT	6h
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>Efnb2</i> ^{lox/lox} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P4.5 50 µg 4OHT	S5b
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>R26-dnMaml1</i> ^{lox/+} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P1-P3 50 µg	S7e
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>R26-Dach1</i> ^{lox/+} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P4.5 50 µg 4OHT	9i
<i>NICD</i> ^{lox/lox} , <i>Esm1-CreERT2</i> ^{+/-} vehicle	<i>NICD</i> ^{lox/lox} , <i>Esm1-CreERT2</i> ^{+/-} anti-VEGF	P3, 4 50 µg 4OHT	5c, S6d
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	<i>R26-dnMaml1</i> ^{lox/+} , <i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-}	P4.5 50 µg 4OHT	6e
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-} vehicle	<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-} DAPT	P4.5 50 µg 4OHT	6f
<i>Efnb2-GFP</i>	/	/	9c, S9c
<i>Efnb2-GFP</i> , <i>Esm1-CreERT2</i> ^{+/-} , <i>Rosa26-RFP</i> ^{lox/+}		P5 50 µg 4OHT	9e
<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-} vehicle	<i>Rosa26-mT/mG</i> ^{lox/+} , <i>Esm1-CreERT2</i> ^{+/-} anti-VEGF	P4, 5 50 µg 4OHT	S6b

Supplementary Table S2. Antibodies used for retina immunostaining

a

Primary antibodies	Species	Clone	Company	Catalog #	Dilution
α-Dach1	Rabbit	Polyclonal	Proteintech	10914-1-AP	1:200
α-DLL4	Goat	Polyclonal	R&D Systems	AF1389	1:50
α-EphB4	Goat	Polyclonal	R&D Systems	AF446	1:50
α-ERG	Rabbit	Monoclonal	Abcam	ab110639	1:200
α-GFP	Chicken	Polyclonal	Abcam	ab13970	1:1000
α-GFP-AF-488	Rabbit	Polyclonal	Invitrogen	A21311	1:500
α-IB4-biotinylated	Griffonia simplicifolia		Vector	B-1205	1:50
α-pERK	Rabbit	Monoclonal	Cell Signaling	4370	1:200
α-Sox17	Goat	Polyclonal	R&D Systems	AF1924	1:100
α-Casp3	Rabbit	Polyclonal	Cell Signaling	9661	1:100

b

Secondary antibodies	Species	Clone	Company	Catalog #	Dilution
α-chicken-AF-488	Donkey	Polyclonal	Jackson ImmunoResearch	703-545-155	1:500
α-goat-AF-647	Donkey	Polyclonal	Invitrogen	A21447	1:500
α-goat-AF-647	Donkey	Polyclonal	Thermo Scientific	A32849	1:500
α-rabbit-AF-488	Donkey	Polyclonal	Invitrogen	A21206	1:500
α-rabbit-AF-546	Donkey	Polyclonal	Invitrogen	A10040	1:500
α-rabbit-AF-594	Donkey	Polyclonal	Invitrogen	A21207	1:500
Streptavidine-AF-405			Invitrogen	S32351	1:100
Streptavidine-AF-546			Invitrogen	S11225	1:100

Supplementary Table S3.

Taqman probes used for qRT-PCR

Name/Gene	Target Species	Company	Assay ID/Cat#
CXCR4 FAM	Human	Thermo Fisher Scientific	Hs00607978_s1
DACH1 FAM	Human	Thermo Fisher Scientific	Hs00974297_m1
DLL1 FAM	Human	Thermo Fisher Scientific	Hs01011330_m1
DLL4 FAM	Human	Thermo Fisher Scientific	Hs00184092_m1
EFNB2 FAM	Human	Thermo Fisher Scientific	Hs00187950_m1
EPHB4 FAM	Human	Thermo Fisher Scientific	Hs00174752_m1
HEY1 FAM	Human	Thermo Fisher Scientific	Hs01114113_m1
JAG1 FAM	Human	Thermo Fisher Scientific	Hs01070036_m1
NR2F2 FAM	Human	Thermo Fisher Scientific	Hs00819630_m1
SOX17 FAM	Human	Thermo Fisher Scientific	Hs00751752_s1
SOX18 FAM	Human	Thermo Fisher Scientific	Hs00746079_s1
SOX7 FAM	Human	Thermo Fisher Scientific	Hs00846731_s1
VEGFR2 FAM	Human	Thermo Fisher Scientific	Hs00911700_m1
GAPDH Endogenous Control VIC	Human	Thermo Fisher Scientific	4326317E
SEMA3G FAM	Human	Thermo Fisher Scientific	Hs00928870_g1
NRP1 FAM	Human	Thermo Fisher Scientific	Hs01546494_m1
GJA4 FAM	Human	Thermo Fisher Scientific	Hs01098016_m1
GJA5 FAM	Human	Thermo Fisher Scientific	Hs00979198_m1
Ephb4 FAM	Mouse	Thermo Fisher Scientific	Mm01201156_m1
Efnb2 FAM	Mouse	Thermo Fisher Scientific	Mm00438670_m1
Gapdh Endogenous Control VIC	Mouse	Thermo Fisher Scientific	4352339E

Supplementary Table S4. Antibodies used for western blotting

a

Primary antibodies	Species	Clone	Company	Catalog #	Dilution
α-Akt	Rabbit	Polyclonal	Cell Signaling	9272	1:1000
α-pAkt(S473)	Rabbit	Monoclonal	Cell Signaling	4060	1:500
α-bTAN20	Rat	Monoclonal	DSHB	bTAN 20	1:700
α-Dll4	Rabbit	Polyclonal	Cell Signaling	2589	1:1000
α-EphB4	Goat	Polyclonal	R&D Systems	AF446	1:2000
α-Ephrin-B2	Goat	Polyclonal	R&D Systems	AF496	1:2000
α-Epsin1	Rabbit	Monoclonal	Abcam	ab75879	1:1000
α-ERK1/2	Rabbit	Monoclonal	Cell Signaling	4695	1:1000
α-pERK1/2(T202/Y204)	Rabbit	Monoclonal	Cell Signaling	4370	1:1000
α-GAPDH	Rabbit	Monoclonal	Cell Signaling	2118	1:10000
α-Notch1	Rabbit	Monoclonal	Cell Signaling	4380	1:1000
α-PLCy	Rabbit	Monoclonal	Cell Signaling	5690	1:1000
α-pPLCy(Y783)	Rabbit	Polyclonal	Cell Signaling	2821	1:500
α-Sox17	Goat	Polyclonal	R&D Systems	AF1924	1:1000
α-Tubulin	Mouse	Polyclonal	Sigma	T5168	1:6000
α-VEGFR2	Rabbit	Monoclonal	Cell Signaling	2479	1:1000
α-pVEGFR2(Y1175)	Rabbit	Monoclonal	Cell Signaling	2478	1:500
α-pVEGFR2(Y951)	Rabbit	Monoclonal	Cell Signaling	4991	1:500

b

Secondary antibodies	Species	Clone	Company	Catalog #	Dilution
α-goat HRP-linked	Bovine	Polyclonal	Jackson Immuno Research	805-035-180	1:15000
α-mouse HRP-linked	Sheep		Amersham	NA931	1:40000
α-rabbit HRP-linked	Goat		Cell Signaling	7074	1:15000
α-rat-HRP-linked	Goat		Amersham	NA935	1:15000

Supplementary Table S5. Antibodies used for HUVEC immunostaining and FACS of murine ECs

a	Primary antibodies	Species	Clone	Company	Catalog #	Dilution
	α -Dach1	rabbit	polyclonal	Proteintech	10914-1-AP	1:200
	α -CyclinD1	rabbit	monoclonal	Cell Signaling	2978	1:50
	α -GOLPH4	rabbit	polyclonal	Abcam	28049	1:500
	α -p21	rabbit	monoclonal	Cell Signaling	2947	1:100
	α -Phalloidin			Invitrogen	A22287	1:40
	α -VE-Cadherin	rabbit	monoclonal	Cell Signaling	3195	1:100
	α -VE-Cadherin	mouse	monoclonal	Santa Cruz	sc-9989	1:100
	α -VEGFR2	goat	polyclonal	R&D Systems	AF644	1:100
	DAPI			Sigma	D9542	1:500

b	Secondary antibodies	Species	Clone	Company	Catalog #	Dilution
	α -goat-AF-488	donkey	polyclonal	Invitrogen	A11057	1:500
	α -mouse-AF-488	donkey	polyclonal	Invitrogen	A21202	1:500
	α -mouse-AF-546	donkey	polyclonal	Invitrogen	A10036	1:500
	α -rabbit-AF-546	donkey	polyclonal	Invitrogen	A10040	1:500
	α -rabbit-AF-647	donkey	polyclonal	Invitrogen	A31573	1:500

c	Conjugated primary antibodies	Species	Clone	Company	Catalog #	Dilution
	α -CD45-BV421	rat	monoclonal	Biolegend	103134	1:100
	α -Ter119-BV605	rat	monoclonal	Biolegend	116239	1:100
	α -CD31-PerCP/ Cy5.5	rat	monoclonal	Biolegend	102420	1:50
	α -CD326-PE/Cy7	rat	monoclonal	Biolegend	118216	1:50
	α -CD140a-PE/Cy7	rat	monoclonal	eBioscience	25-1401-82	1:100
	α -CD140b-APC	rat	monoclonal	eBioscience	17-1402-82	1:25
	α -Podoplanin-eFluor660	hamster	monoclonal	eBioscience	50-5381	1:100
	α -CD208-A647	rat	monoclonal	Dentric	DDX0192A647-100	1:50