

Supplementary Figure 1. GPR35 expression supports cDC2 maintenance and localization within SED. A. Dot plot showing gene expression levels in PP cell clusters in scRNAseq data

adapted from Xu et al., (16) (Single Cell Portal). Low (blue) and high (red) expression levels are indicated for each gene. Dot size directly correlates with % of expressing cells. B. GPR35 intracellular flow cytometry staining in cDC2, cDC1 and DN from GPR35^{+/-} (top) and GPR35^{-/-} (bottom) mice. Data are representative of 2 independent experiments. C-E. Quantification of cDC2 (C), cDC1 (D) and DN DC (E) % (out of total cDC) in PPs from GPR35^{+/-} and GPR35^{-/-} mice. n=14 (GPR35^{+/-}); n=13 (GPR35^{-/-}). Data are pooled from 3 independent experiments. **F**, **G**. Quantification of PP number (**F**) and PP total cells (**G**) in GPR35^{+/-} and GPR35^{-/-} mice. n=18, GPR35^{+/-} F; *n*=26, GPR35^{-/-} F; *n*=14, GPR35^{+/-} G; *n*=13, GPR35^{-/-} G. **H**, **I**. Quantification of splenic cDC2 % (H) and absolute numbers (I) from GPR35^{+/-} and GPR35^{-/-} mice. *n*=5 (GPR35 ^{+/-}); n=4 (GPR35^{-/-}). Data are pooled from 2 independent experiments. J, K. Quantification of GPR35^{+/+} (J) and GPR35^{-/-} (K) PP cDC2 in vitro migration to 5-HIAA or no chemoattractant (Nil) at the indicated concentrations. Each dot represents a technical replicate (n=3). Data are representative of 3 independent experiments. L. Immunofluorescence micrographs showing PPs from GPR35^{+/-} (left) and GPR35^{-/-} (right) mice. CD11c (green), BST2 (blue), CD11b (red) and IgD (grey) staining (left images) or cDC2 and Lyz DC/Lyz Mac gated populations (right images) are shown. SED, subepithelial dome. White dashed delimits the SED and was depicted based on BST2 and IgD staining. M. Quantification of cDC2 % in SED from GPR35^{+/-} (n=6) or GPR35^{-/-} (n=5) mice, from images shown in **L**. Data are pooled from 2 independent experiments. N, O. Immunofluorescence micrographs (N) and quantification (**O**) of CD11c⁺ CD11b⁺ WT (GFP⁺) or GPR35^{-/-} (GFP⁻) cells within SED and IFA of PPs from CD11c-GFP/ GPR35^{-/-} mixed chimeras. n=3. Data are pooled from 2 independent experiments. * p<0.05; *** p<0.005; *** p<0.0005. Data are presented as mean ± SEM. Twotailed unpaired t-tests were performed (C-I; M) and ordinary one-way ANOVA using Turkey's multiple comparisons test were performed (J, K, O).

Supplementary Fig. 2



Supplementary Figure 2. Bone marrow derived GPR35-expressing cells support intestinal IgA responses. A. ELISA quantification of fecal total free IgA in GPR35^{+/-} (*n*=9), GPR35^{-/-} (*n*=9) and negative controls (RAG1^{-/-}, *n*=4). Data are pooled from 2 independent experiments (distinguished by color). B, C. Quantification of IgA⁺ GC % (out of GL7⁺ CD95⁺ CD38⁻ IgD⁻ CD19⁺ cells, B) and IgA⁺ plasma cells % (out of CD19⁺ IgD⁻ CD138⁺ cells, C) in GPR35^{+/+} BM -> GPR35^{+/+} recipient (wt-wt), GPR35^{-/-} BM -> GPR35^{+/+} recipient (wt-wt) and GPR35^{+/+} BM -> GPR35^{-/-} (wt-ko) chimeric mice. *n*=7. Data are pooled from 2 independent experiments. D, E. Quantification of fecal (D) and small intestine (E) IgA-bound bacteria in wt-wt, ko-wt and wt-ko chimeric mice. *n*=7. Data are pooled from 2 independent experiments. F-H. Quantification of cDC2 % (F, *n*=3), IgA⁺ and IgG1⁺ GC % (G, *n*=4) and IgA⁺ plasma cells % (H, *n*=4) in mesenteric LNs from GPR35^{+/-} and GPR35^{-/-} mice. Data are pooled from 2 independent seperiments. * p<0.05; ** p<0.0005. Data are presented as mean ± SEM. Two-tailed unpaired t-tests (A, F, H) and ordinary one-way ANOVA using Turkey's multiple comparisons test were performed (B-E; G).

Supplementary Fig. 3



Supplementary Figure 3. GPR35 supports intestinal IgA responses via Itgb8. A, B. Representative flow cytometry plots showing itgb8 expression levels from negative control (top) and itgb8-reporter mice (bottom) within cDC2, DN DC, cDC1 and Lyz DC/Lyz Mac (**A**) and relative Itgb8 % quantification (**B**). **C-E**. Quantification of IgA⁺ GC % (out of GL7⁺ CD95⁺ CD38⁻ IgD⁻ CD19⁺ cells, **C**), IgA⁺ plasma cells % (out of CD19⁺ IgD⁻ CD138⁺ cells, **D**) and fecal IgA-bound bacteria (out of DAPI⁺, **E**) in control mice or mice treated for 3 weeks with Itgb8 blocking antibody. *n*=3 (GPR35^{+/+}, **C**, **D**); *n*=6 (GPR35^{+/-} + anti-itgb8, **C-E**); *n*=5 (GPR35^{-/-} + antiitgb8, **C-E**); *n*=2 (GPR35^{+/+}, **E**). Data are pooled from 2 independent experiments. Data are representative of 3 independent experiments. **F-H**. Representative flow cytometry plots from GPR35^{+/-} or GPR35^{-/-} itgb8-tdTomato reporter mice (**F**). Quantification of itgb8 % out of cDC2 (**G**) and itgb8 cDC2 % out of CD19⁻ IgD⁻ CD3⁻ (**H**) of data shown in **F**. *n*=4. Data are pooled from 2 independent experiments. * p<0.05; ** p<0.005; *** p<0.0005. Data are presented as mean ± SEM. Two-tailed unpaired t-tests (**G**, **H**) and ordinary one-way ANOVA using Turkey's multiple comparisons test were performed (**B-E**).



Supplementary Fig. 4

Supplementary Figure 4. GPR35-expressing cDC2 support intestinal IgA B cell responses.

A-D. Quantification of cDC (**A**), Lyz DC (**B**), Lyz Mac (**C**) % out of CD45.1⁺ (WT) or CD45.2+ (Zbtb46-DTR) cells, and IgA⁺ (left, **D**) and IgG1⁺ (right, **D**) GC % (out of CD19⁺) within CD45.1 (WT) or GPR35^{-/-} + Zbtb46 mixed chimeric mice. *n*=7. Data are pooled from 2 independent experiments. **E-I**. Quantification of cDC2 (**E**), DN DC (**F**), cDC1 (**G**), Lyz DC (**H**) and Lyz Mac (**I**) % out of IgD⁻ CD3⁻ CD19⁻ cells in control (CD11c-Cre LTbr^{fl/wt} and LTbr^{fl/fl}) and CD11c-Cre LTbr^{fl/fl} mice. *n*=4. Data are representative of 2 independent experiments. **J-L**. Quantification of IgA⁺ GC (left, **J**), IgG1⁺ GC (right, **J**), IgA⁺ plasma cells (**K**) and fecal IgA-bound bacteria in CD45.1/CD11c-Cre LTbr^{fl/fl} and GPR35^{-/-} / CD11c-Cre LTbr^{fl/fl} mixed chimeric mice (**L**). *n*=6. Data are pooled from 2 independent experiments. **M-O**. Quantification of Lyz DC/Mac (**M**), cDCs (**N**),

IgA⁺ GC (left, **O**), IgG1⁺ GC (right, **O**) and IgA⁺ plasma cells (**P**) from CD45.1 / CCR2^{-/-} and GPR35^{-/-} / CCR2^{-/-} mixed chimeric mice. n=6 (**M-O**); n=4 (**P**). Data are pooled from 2 independent experiments. * p<0.05; ** p<0.005; *** p<0.0005. Data are presented as mean \pm SEM. Two-tailed unpaired t-tests were performed.



Supplementary Figure 5. Mast cells support intestinal IgA B cell responses. A-G.

Quantification of cDC2 (A), cDC1 (B), DN DC (C), total GC (D), IgA⁺ GC (E) and IgA⁺ plasma cells % (F) and numbers (G) in WT and mast cell deficient (Kit/v x Kit/W) mice. n=8 (WT, A-C); n=5 (WT, D-G) n=11 (Mast cell-deficient, A-C); n=8 (Mast cell deficient, D-G). Data are pooled from 3 (A-C) or 2 (D-G) independent experiments. H. Immunofluorescence micrographs showing PPs from Cpa3-Cre Ai14^{fl/fl} mice. BST2 (green), CD11c (white), Cpa3 (red) staining and signal are shown. SED= subepithelial dome. White dashed lines identify the SED and were depicted based on BST2, CD11c and IgD (not shown) staining. Images are representative of multiple sections from at least 4 mice and 2 independent experiments. I. Flow cytometry plots showing Cpa3-Ai14 (tdTomato), Fccr1 and CD49b levels in negative control (left) and in Cpa3-Cre Ai14^{fl/fl} mice. J. Representative flow cytometry plots of PP mast cells identified as FccR1⁺CD117⁺, showing CD63 (right) and fluorescence-minus-one (Fmo) control (left). Gate used to determine % CD63⁺ cells is shown. Data are representative of at least 2 independent experiments. K, L. Quantification of CD63⁺ mast cell % (K) and cDC2s % (L, left) and absolute numbers (L, right) in germ free PPs. n=9 (WT); n=6 (germ free). Data are pooled from 2 independent experiments. M. Immunofluorescence microscopy of WT (left) and germ free (right) PPs to detect IgD (blue), CD11c (red), BST2 (green), and CD11b (white). Data representative of multiple sections from each of 6 mice. * p<0.05; ** p<0.005; *** p<0.0005. Data are presented as mean ± SEM. Two-tailed unpaired t-tests were performed.



Supplementary Figure 6. Mast cell-derived serotonin metabolite(s) sustain cDC2 maintenance and localization within SED to drive intestinal IgA responses. Dot plot showing *Cpa3* and *Tph1* gene expression levels in PP cell clusters from published scRNAseq data (adapted from *Xu et al., Immunity 2019*, Single Cell Portal). Low (blue) and High (red)

expression levels are indicated for each gene. B. ELISA quantification of 5-HIAA in supernatants from LPS-activated Control (Tph1^{fl/fl}) and Cpa3-Cre x Tph1^{fl/fl} peritoneal mast cells. n=2 (Control); n=3 (Cpa3-Cre x Tph1^{fl/fl}). **C**. Transwell migration assay with GPR35overespressing WEHI231 cells to supernatants from Control (Tph1^{fl/fl}) or Cpa3-Cre Tph1^{fl/fl} activated peritoneal mast cell supernatant, or to medium alone (Nil). n=4 (Control and Nil); *n*=3 (Cpa3-Cre Tph1^{fl/fl}). **D**. ELISA quantification of 5-HIAA in extracts of PPs isolated from Tph1^{fl/fl} and Cpa3-Cre Tph1^{fl/fl} mice. *n*=4. Each dot represents supernatants obtained from independent mast cell pools (B, C) or independent PP pool (D). Data are pooled from 2 independent experiments. E. Representative histogram overlay plot of surface CD63 on Fccr1⁺CD117⁺ PP mast cells in control and Cpa3-Cre Tph1^{fl/fl} mice (n=2 of each). Fmo, fluorescence-minus-one refers to sample stained without CD63 antibody. F-H. Quantification of cDC2 (F), cDC1 (G) and LyZ DC/Mac (H) in PP from control (Cpa3-Cre Tph1^{fl/+} and Tph1^{fl/fl}) and Cpa3-Cre Tph1^{fl/fl} mice. *n*=11 (control); *n*=12 (Cpa3-Cre x Tph1^{fl/fl}). Data are pooled from 3 independent experiments. I-M. Quantification of cDC2 absolute numbers (I), total GC % (J), IgA⁺ GC % (K) and IgA⁺ plasma cells % (out of total plasma cells, L; out of total cells, **M**) in PPs from Tph1^{+/-} and Tph1^{-/-} chimeric mice. n=8 (I); n=7 (J-M). Data are pooled from 3 independent experiments. * p<0.05; ** p<0.005; *** p<0.0005. Data are presented as mean ± SEM. Two-tailed unpaired t-tests (B-D; F-M) and ordinary one-way ANOVA using Turkey's multiple comparisons test were performed (C).

Supplementary Fig. 7



Supplementary Figure 7. Mast cell-derived serotonin metabolite(s) support cell positioning within SED. A. Immunofluorescence micrographs showing PPs from Tph1^{fl/fl} (left), mast cell deficient Kit/v x Kit/W (middle) and Cpa3-Cre Tph1^{fl/fl} (right) mice. CD11c (green), BST2 (blue), CD11b (red) and IgD (grey) staining (top), or cDC2 (green, bottom) and

Lyz DC/Mac (blue, bottom) are shown. SED= subepithelial dome. White dashed lines identify the SED and were depicted based on BST2 and IgD staining. B. Quantification of cDC2 % in the SED of Tph1^{fl/fl}, mast cell deficient and Cpa3-Cre x Tph1^{fl/fl} mice from images shown in **A**. *n*=4. Data are pooled from 2 independent experiments. **C**. Immunofluorescence micrographs showing PPs from control (left) or phenelzine treated (right) GPR35-GFP overexpressing chimeras. GFP high (green), GFP low (white), CD11c (purple) staining are shown. D, E. Quantification of GPR35-GFP high and low cell % in SED of control (D) or phenelzine treated (E) GPR35-GFP overexpressing chimeras, from images of the type shown in C. n=3. Data are pooled from 2 independent experiments. F. Immunofluorescence micrographs showing PPs from Tph1^{fl/fl} (left) and Cpa3-Cre x Tph1^{fl/fl} (right) that received Deep Red-labeled GPR35-GFP overexpressing cells, 60 hrs after transfer. Deep Red⁺ GFP⁺ (green), Deep Red⁺ GFP⁻ (untransduced, white) and CD11c (blue) staining are shown. G, H. Quantification of GPR35-GFP⁺ and GFP⁻ % (out of total CD11C⁺ CD11b⁺ cells) in SED of Tph1^{fl/fl} (G) and Cpa3-Cre Tph1^{fl/fl} (H) mice from images of the type shown in F. n=4. Data are pooled from 2 independent experiments. * p<0.05; ** p<0.005; *** p<0.0005. Data are presented as mean \pm SEM. Paired-t-tests (**D-H**) and ordinary one-way ANOVA using Turkey's multiple comparisons test (**B**) were performed.



Supplementary Figure 8. Flow cytometry gating strategies.

A-G. Representative examples of cDC (A), Lyz DC and Lyz MAC (B), IgA GC and plasma cell (C), lamina propria plasma cells (D), IgA-coated intestinal bacteria (E), itgb8-expressing cDC2 (F), and CD63⁺ mast cell (G) gating strategies are shown.

Supplementary Movie Legends

Supplementary movie 1.

3D reconstruction of whole-mount SED imaging showing CD45.1 (red), CD45.2 (blue), CD11c (white) and BST2 (green) staining (left); or cDC WT (red), cDC KO (blue), BST2 (green) gated populations (right), from CD45.1 WT/ CD45.2 GPR35^{+/-} control mixed chimeras.

Supplementary movie 2.

3D reconstruction of whole-mount SED imaging showing CD45.1 (red), CD45.2 (blue), CD11c (white) and BST2 (green) staining (left); or cDC WT (red), cDC KO (blue), BST2 (green) gated populations (right), from CD45.1 WT/ CD45.2 GPR35^{-/-} mixed chimeras.