## Supplementary Information for

# Multiscale architecture design of 3D printed biodegradable Zn-based porous scaffolds for immunomodulatory osteogenesis

#### **Authors list**

Shuang Li<sup>1†</sup>, Hongtao Yang<sup>1,2†,\*</sup>, Xinhua Qu<sup>3†</sup>, Yu Qin<sup>2</sup>, Aobo Liu<sup>4</sup>, Guo Bao<sup>5</sup>, He Huang<sup>6</sup>, Chaoyang Sun<sup>1</sup>, Jiabao Dai<sup>4</sup>, Junlong Tan<sup>1</sup>, Jiahui Shi<sup>2</sup>, Yan Guan<sup>7</sup>, Wei Pan<sup>7</sup>, Xunan Gu<sup>1</sup>, Bo Jia<sup>4</sup>, Peng Wen<sup>4\*</sup>, Xiaogang Wang<sup>1\*</sup>, Yufeng Zheng<sup>2\*</sup>

### Affiliations

<sup>1</sup>School of Engineering Medicine, School of Biological Science and Medical Engineering, Beihang University, Beijing 100191, China
<sup>2</sup>School of Materials Science and Engineering, Peking University, Beijing, 100871, China
<sup>3</sup>Department of Bone and Joint Surgery, Department of Orthopedics, Renji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai 200001, China
<sup>4</sup>Department of Mechanical Engineering, Tsinghua University, Beijing, 100084, China.
<sup>5</sup>Department of Reproduction and Physiology National Research Institute for Family Planning Beijing 100081, China
<sup>6</sup>School of Materials Science and Engineering, Zhengzhou University, Zhengzhou, 450003, China
<sup>7</sup>College of Chemistry and Molecular Engineering, Peking University, Beijing, 100871, China

<sup>†</sup> These authors contributed equally to this work. **Corresponding author: Hongtao Yang, Yufeng Zheng** 

Email address: <u>yang276070@buaa.edu.cn</u>, yfzheng@pku.edu.cn

### This PDF file includes:

Figures. S1 to S9 Tables S1 to S5



Fig. S1. Immunofluorescence staining of iNOS, CD206, and DAPI of RAW264.7 after co-culture with material extracts for 48h. Each image was acquired independently three times, with similar results.



Fig. S2. SEM images of RAW264.7 cells after 6h attachment on different surfaces. Each image was acquired independently three times, with similar results.



Fig. S3. Diffusion coefficient of Zn ions in scaffolds with BCC and G unit over 120 minutes (n=3, independent experiments). Data are presented as mean  $\pm$  standard deviation. OA: orientation A, OB: orientation B. P-values are calculated using one-way ANOVA with Tukey's post hoc test, \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.005.



Fig. S4. Enriched KEGG pathway of G scaffold versus control.



Fig. S5. Runx2 and Osx immunofluorescence staining of MC3T3-E1 cultured in conditioned medium at 7 days. Each image was acquired independently three times, with similar results.



Fig. S6. Comparison of new bone regeneration and degradation between Zn-Li alloy bulk sample and scaffolds at 3 months. Red asterisks indicate scaffold degradation. Each image was acquired independently three times, with similar results.



Fig. S7. Bone regeneration comparison between Zn-Li scaffold and Ti scaffold in a critical bone defect rabbit model at 2 months. A Micro-CT reconstruction of new bone tissue and metallic implants with quantitative analysis (n=3, independent experiments) of bone volume/tissue volume (BV/TV), bone mineral density (BMD), trabecular thickness (Tb. Th), and trabecular separation (Tb. Sp). New bone is marked in yellow, implants are marked in white. B Methylene blue acid fuchsin staining of bone defect regions. Yellow asterisks indicate newly formed bone, white asterisks are scaffold struts. Data are presented as mean  $\pm$  standard deviation. P-values are calculated using one-way ANOVA with Tukey's post hoc test, \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.005. Each image was acquired independently three times, with similar results.



Fig. S8. Schematic diagram of diffusion device.



Fig. S9. Zn concentrations of BCC and G scaffold extracts determined by ICP (n=4 independent experiments). Data are presented as mean  $\pm$  standard deviation. P-values are calculated using one-way ANOVA unpaired t-test with a Mann-Whitney test.

Nominal composition (wt.%)	Actual composition (wt.%)
Zn-0.2Li	0.239
Zn-0.5Li	0.494
Zn-0.8Li	0.779
Zn-1.2Li	1.36

Table S1 Chemical composition of Zn-Li alloys

Products	Component	Binding energy (eV)		
		Measured	Ref.	
ZnO	Zn $2p_{3/2}$	1021.8	1021.9	
	O 1 <i>s</i>	530.6	530.8	
	Li 1s	55.0	55.1	
Li <sub>2</sub> CO <sub>3</sub>	C 1 <i>s</i>	289.6	289.6	
	O 1 <i>s</i>	530.6	531.4	

Table S2. Binding energy of corrosion products detected in Zn-Li alloys after immersion in SBF

Size	Structure characteristics	BCC	G
	Porosity (%)	88	86
	Pore size (mm)	0.75	0.4
$\Phi$ 3×4	Strut thickness (mm)	0.4	0.5
mm	Surface area (mm <sup>2</sup> )	78.7	76.08
	Scaffold volume (mm <sup>3</sup> )	3.58	3.99
	Specific surface area (scaffold) (mm <sup>-1</sup> )	21.99	19.04
	Porosity (%)	90.48	90.34
Φ10×2	Surface area (mm <sup>2</sup> )	404.12	353.59
mm	Volume (mm <sup>3</sup> )	15.20	15.14
	Specific surface area (scaffold)	26.6	23.35

Table S3. CT-measured structure parameters of Zn-Li scaffolds with different pore unit and size.

Materials	Mechanical properties			Biodegrad	Bioactivity	Printability	
	$\mathrm{UTS}^{\mathrm{b}}$	UCS <sup>c</sup>	Elongation	Elastic	ability		
	(MPa)	(MPa)	%	Modulu			
				s (GPa)			
Cortical bone <sup>1</sup>	50-151	130-200	-	7-30	No	Yes	No
Pure Ti (Grade	240-550	-	15-24	110	No	No	Yes
1-4) <sup>a</sup>							
Zn-Li alloys	252-780	790-1100 <sup>d2</sup>	0-26	100	Yes	Yes	Yes

Table S4 Comparison of key properties between pure Ti, autologous bone and Zn-Li alloys

<sup>a</sup>ASTM-F67

<sup>b</sup> Ultimate tensile strength

<sup>c</sup> Ultimate compressive strength

<sup>d</sup>Zn-Li alloys have compression super plasticity, the maximum stress before 50% compressive strain was defined as ultimate compressive strength

#### References

1. Gerhardt, L. C., Boccaccini, A. R. Bioactive glass and glass-ceramic scaffolds for bone tissue engineering. *Materials* **3**, 3867-3910 (2010).

2. Yang, H., Jia, B., Zhang, Z., Qu, X., Li, G., Lin, W., Zhu, D., Dai, K., Zheng, Y. Alloying design of biodegradable zinc as promising bone implants for load-bearing applications. *Nat. commun.* **11**, 1-16 (2020).

Primer	Sequences
iNos	GTTCTCAGCCCAACAATACAAGA
	GTGGACGGGTCGATGTCAC
11 10	GCAACTGTTCCTGAACTCAACT
11 <b>-</b> 1 <i>þ</i>	ATCTTTTGGGGTCCGTCAACT
Tnf-α	CCCTCACACTCAGATCATCTTCT
	GCTACGACGTGGGCTACAG
Argl	CTCCAAGCCAAAGTCCTTAGAG
	AGGAGCTGTCATTAGGGACATC
<i>Il-4</i>	GGTCTCAACCCCCAGCTAGT
	GCCGATGATCTCTCTCAAGTGAT
Il-10	GCTCTTACTGACTGGCATGAG
	CGCAGCTCTAGGAGCATGTG
Collal	GCTCCTCTTAGGGGGCCACT
	CCACGTCTCACCATTGGGG
Ono	ACCCAGAAACTGGTCATCAGC
Opg	CTGCAATACACACACTCATCACT
Onn	ACCCAGAAACTGGTCATCAGC
Opn	CTGCAATACACACACTCATCAC
Alp	CCAACTCTTTTGTGCCAGAGA
	GGCTACATTGGTGTTGAGCTTTT
Gapdh	AGGTCGGTGTGAACGGATTTG
	TGTAGACCATGTAGTTGAGGTCA

Table S5 Primer sequences of migration-related genes for qRT-PCR