

## Supplementary Material

Supplemental Table 1 | qPCR primer list.

Housekeeping gene	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')
<i>Gapdh</i>	TGGATCTGACGTGCCGC	TGCCTGCTCACCACCTTC

Negative control gene	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')
<i>Gfp</i>	GAAGCGCGATCACATGGT	CCATGCCGAGAGTGATCC

GPCR-encoding genes	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')
<i>Adcyap1r1</i>	TATGGACTTCAAGCACCGGC	TCTTGCTCAGGATGGACAGC
<i>Adora2a</i>	GTTAGGTAGGCAGAGGGACAGG	CTGCGATTGCTTCCCTTCTCTG
<i>Adora2b</i>	GGAACCGAGACTTCCGCTAC	GACTGAGAGTAGACTGCGCC
<i>Adrb1</i>	CTACAACGACCCCAAGTGCT	ACGTAGAAGGAGACGACGGA
<i>Adrb2</i>	TACACAGGGGAGCCAAACAC	TCAACGCTAAGGCTAGGCAC
<i>Adrb3</i>	CAGGCTCTGTGTCTCTGGTTA	GAGGAGACAGGGATGAAACCTC
<i>Agtr1a</i>	CTTAGGGTTGGAACCTGCGG	TCATCCAGTCCCTCCCAACT
<i>Bdkrb1</i>	CCGCTACAGGTTGCTGGTAT	TTGACGGAACGCAGAAGGAA
<i>Cckar</i>	ACTGCCAAGTCCACGTTCAA	TCATCTGGGGCGTTCCAAAA
<i>Chrm2</i>	ACTGCCATTGCGGCTTTCTA	TATTCTGCTCTTGCTCGCCC
<i>Chrm4</i>	GCCTCTGGCTAGTTCCGCC	TCGCCATGCTGAACCCAAC
<i>Drd2</i>	GACACCACTCAAGGGCAACT	ATCCATTCTCCGCCTGTTCA

<i>Drd5</i>	CGAACCTACGCCATCTCCTC	GCGCGTGTAGGTCACTATCA
<i>Gabbr2</i>	ACAGGCGATTCCAGTTCACA	CGTAGGCGGTGGTTTTCTGA
<i>Gpr3</i>	ATCTACGCCTTTTCGCAACCA	CGGGACCGGAATGGAATCTT
<i>Gpr65</i>	CATGGGCTACGCAATACCCT	TGTTTTCCGTGGCTTGGTTG
<i>Gpr68</i>	ACGATACCAGCCCAAGTGTG	CACCTTAACCAGTCCTCTGGC
<i>Grm4</i>	TACCAGTACCAACGTGCGAA	GCATCCGCTCTATTCTGAGGT
<i>Grm8</i>	TGTGCTCCTAACGGGGATTT	GATGATTGTGTCAGGTGCCG
<i>Hcar1</i>	AGTGTGAAGGAAACCGTGGG	CGCTTTTCTCAGCCATGCAA
<i>Hcar2</i>	GCGGCCATCATTCTTGCTT	GCCTCGCCATTTTTGGTCAT
<i>Hrh3</i>	TTAGAGCATCAACCCGGCAG	CACTCCAGTTCACCAACGA
<i>Htr7</i>	GTGGTCAAAATGGGAAACGGA	CCATTCTGCCTCACGGGGTA
<i>Lpar2</i>	GGCAGATGACTTGACTTCGC	GCCTCCCTGAATGTTTGCTC
<i>Oprd1</i>	TGGATGCTTTTGGGGTTCCT	AAACAAAGGGTCTCGGTGCT
<i>Oprl1</i>	TCCTCAGGCACACCAAGATG	GAAGGGCAGTGTGAGCAAGA
<i>P2ry12</i>	AACGCCAGTGTCAATTGCTG	TCTCCTTTTATTCTTGCACTGTGAC
<i>Pthr1</i>	AGCGAGTGCCTCAAGTTCAT	TCCCACGGTGTAGATCATGC
<i>S1pr5</i>	AACTCGCTGCTGAATCCCAT	GGAGGAGTCTTGTTGCAGG
<i>Tacr1</i>	AGGTGTCTGGGGTTTCTTTA	CCTAGAAGTGACAGGTGACCA

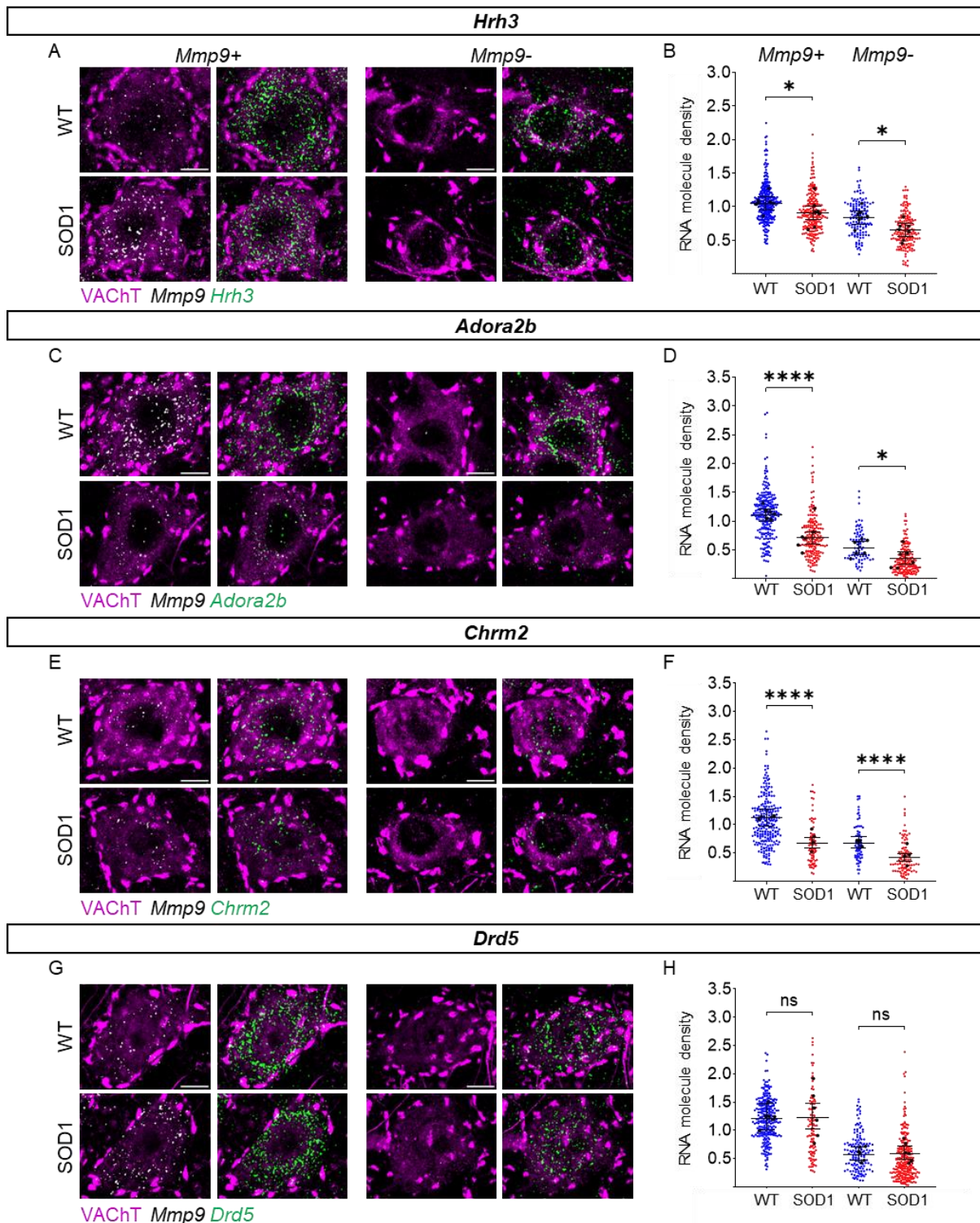
Immediate-early genes	Forward primer sequence (5'-3')	Reverse primer sequence (5'-3')
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<i>ΔFosB</i>	AGGCAGAGCTGGAGTCGGAGAT	GCCGAGGACTTGAACCTCACTCG
<i>Arc</i>	GCACAAAAGCCATGACCCAT	TCTCCCTAGTCCCCAGGGC
<i>c-Fos</i>	CCTGCCCCTTCTCAACGAC	GCTCCACGTTGCTGATGCT
<i>Egr1</i>	GCCGAGCGAACAACCCTAT	TCCACCATCGCCTTCTCATT
<i>Egr2</i>	GTTGACTGTCACTCCAAGAAATGG	AGCGCAGCCCTGTAGGC
<i>NPas4</i>	GCTATACTCAGAAGGTCCAGAAGG C	TCAGAGAATGAGGGTAGCACAGC

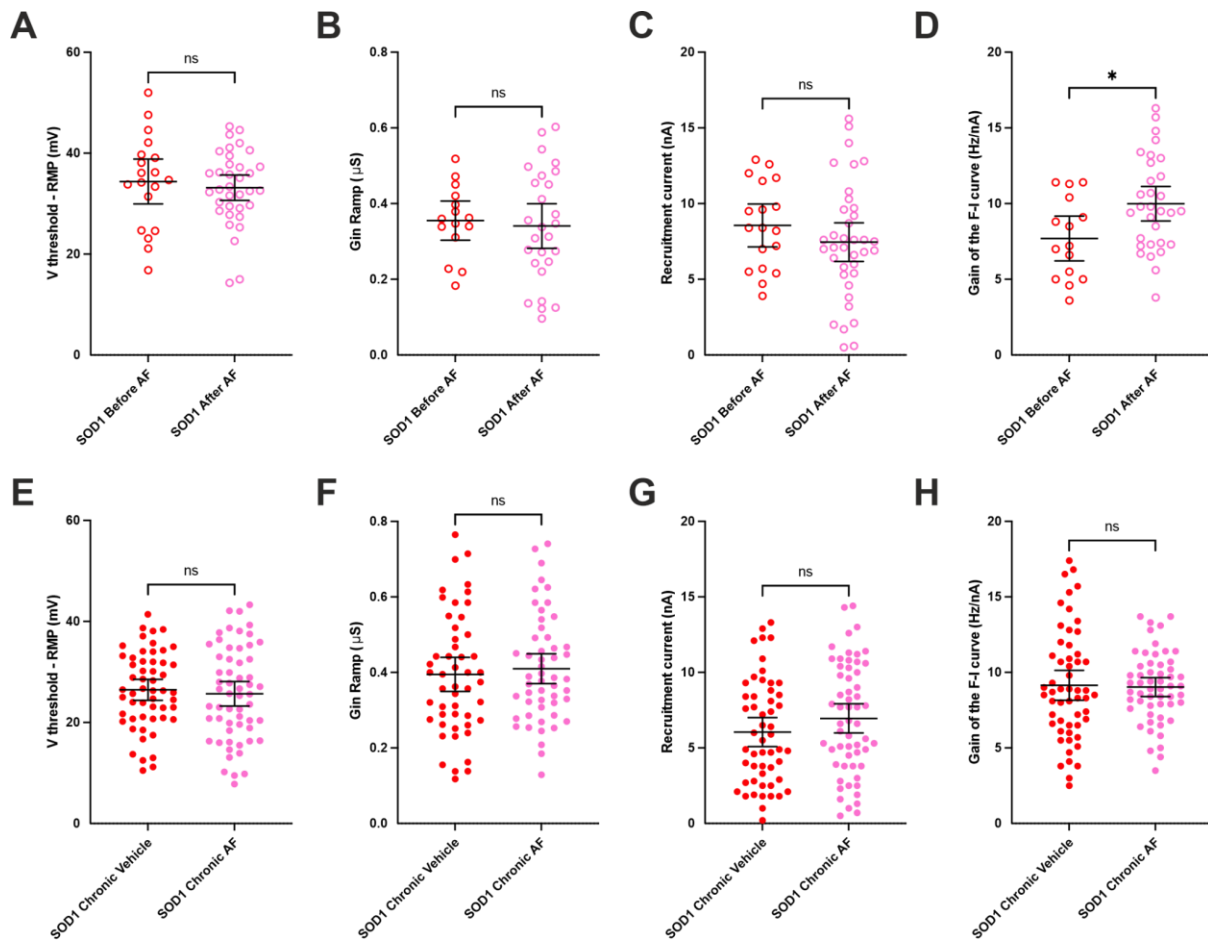
<b>Ion channel-encoding genes</b>	<b>Forward primer sequence (5'-3')</b>	<b>Reverse primer sequence (5'-3')</b>
<i>Cacna1d</i> (isof.1)	GCTCGGTGGCTGTATTTTCAA	CCGTGCTTTCTACCGCACTT
<i>Cacna2d3</i>	GCAGATCGCAGGAAGCTTTG	ACGGGAGATTTCCGCTCATC
<i>Hcn1</i>	CGTGAAGCATGACCGAGAGA	GTAGACTGGCGGAGATTGGG
<i>Hcn2</i>	CATCCACACCAAAGCCATGC	CCCGCCTCCTAAGCTACCTA
<i>Kcna1</i>	TGCTGTGTGTCGCTCAATCT	TCTCCGAACTGGACACTTGC
<i>Kcna2</i>	TACCCATCTGCAAGGGCAACG	CGACTTGAGGAGGAGAGTGGA
<i>Kcnab1</i>	TCTTGGACTGGTCCCCTACC	AGATTCCCCTACCCCAGCAT
<i>Kcnb1</i>	GAGAGGGCGTGGCTAAGAAG	GCCCTCTTGGTCCATTTCCA
<i>Kcnj14</i>	GCCGAGGACAGACCTGAACAC	ACTGGGGGTTCTCTGCTCA
<i>Kcnn3</i>	ATCCACCGTCATCCTGCTTG	GTAGGTCATGGCTATCCGCC
<i>Kcnn2</i>	ACAAGGCGTCGCTGTATTCT	CTGTATTTCCCTGGCGTGGT

<i>Kcnq2</i>	GCCATTTTGTACGTGCCCTT	TAGAAGACAGCGTCGTGTGC
<i>Kcnq3</i>	AGTCTTGCTTCCCTGGTGATTC	TCGTCCTGCATTTGGCTGATA
<i>Kcnq5</i>	GCAGCCACCAGACTAAAGGA	CTGCCGCTTCCAATTCCAAA
<i>Kcnt2</i>	CTGTGCACTTAAAAGCAATACAGT	AGCATTTTCCACATCCATGACT
<i>Scn1a</i>	TACAGAAGCAGACCGTAGGC	TGTGATTAGCATCATTTTGGGCT
<i>Scn8a</i>	CCTTCTTACGAGACCCGTGG	ACCCTGAAAGTGCGTAGAGC
<i>Tmem16f</i>	TGGAACCCTGATCTTCGCTG	TTGCTGTAGCTCAACGGTGT
<i>Trpm5</i>	GAATGGGGACTACAGAGGCTG	CGAATGTTTCCTGTGGAGGC

## Supplementary Figure



**Supplemental Figure 1 | Additional disease-driven transcriptional changes in pre-symptomatic SOD1 MNs at single-molecule single-MN resolution.** **A, C, E, G**) Representative confocal images of individual p45 WT and SOD1 *Mmp9+* and *Mmp9-* MNs for *Hrh3*, *Adora2b*, *Chrm2* and *Drd5* ISH, respectively. MNs are identified by VACHT+ C-boutons and diffuse cytoplasmic signal (magenta); cytoplasmic spots define single mRNA molecules of *Mmp9* (white) or the GPCRs of interest (green). Scale bar=10 $\mu$ m. **B, D, F, H**) Quantification of the density of single mRNA molecules of *Hrh3* (**B**), *Adora2b* (**D**), *Chrm2* (**G**) and *Drd5* (**H**) per MN cross-section area. In each dot plot, small dots correspond to individual MN, whereas large dots indicate individual mouse means; the error bar delineates the overall mean and corresponding 95% confidence interval (for *Chrm2* and *Drd5* it is back-propagated from the log scale). N = 4-6 mice per genotype group. \* $p$ <0.05, \*\* $p$ <0.01, \*\*\* $p$ <0.001, \*\*\*\* $p$ <0.0001, ns - non significant.



**Supplemental Figure 2 | Replication of acute and prolonged delivery of adrenergic  $\beta$ 2/ $\beta$ 3 agonists on a different cohort. A-H) Electrophysiological properties were obtained from slow ramps of current, as in Figure 3 and 7 but were performed in Poznan. A-D) Effect of the acute treatment on Voltage threshold - resting membrane potential (A), ramp input conductance (B), recruitment current (C), gain of the F-I relationship (D), in MNs from SOD1 mice. E-H) Effect of the chronic treatment on Voltage threshold - resting membrane potential (E), ramp input conductance (F), recruitment current (G), gain of the F-I relationship (H), in MNs from SOD1 mice. In all graphs, each point represents one MN and the mean  $\pm$  95% CI are shown. Significances on top bars are for treatment effects. N = 7 Acute SOD1 mice and N = 11 Chronic SOD1 mice. \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , \*\*\*\* $p < 0.0001$ , ns - non significant.**