## **Supporting Information**

## Next generation chemiluminescent probes for antimalarial drug discovery

Angela Hellingman<sup>1,2</sup>, Kleopatra Sifoniou<sup>1,2</sup>, Tamara Buser<sup>1,2</sup>, Basil T. Thommen<sup>1,2</sup>, Annabelle Walz<sup>1,2</sup>, Armin Passecker<sup>1,2</sup>, James Collins<sup>3</sup>, Mario Hupfeld<sup>3</sup>, Sergio Wittlin<sup>\*,1,2</sup>, Kathrin Witmer<sup>\*,1,2,3</sup>, and Nicolas M. B. Brancucci<sup>\*,1,2</sup>

<sup>1</sup> Department of Medical Parasitology and Infection Biology, Swiss Tropical and Public

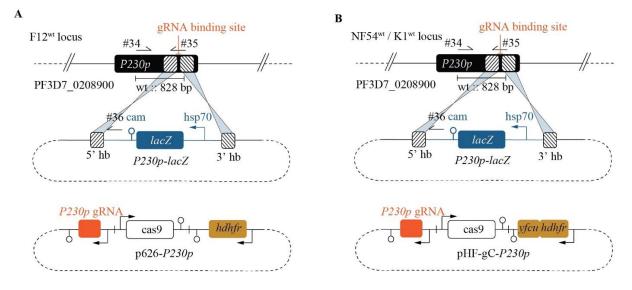
Health Institute, 4123 Allschwil, Switzerland

<sup>2</sup> University of Basel, 4001 Basel, Switzerland

<sup>3</sup>NEMIS Technologies AG, 8804 Au ZH, Switzerland

\* Corresponding authors: Nicolas M. B. Brancucci (nicolas.brancucci@swisstph.ch), Kathrin Witmer

(kathrin.witmer@gmail.com), Sergio Wittlin (sergio.wittlin@swisstph.ch)



**Figure S1.** Overview of the two-plasmid CRISPR/Cas9-based gene editing strategy. (A) Schematic of the *P230p* locus in wild-type F12 parasites and the plasmids used for CRISPR/Cas9-mediated gene editing (*P230p-lacZ* and p626-*P230p*) to generate the F12<sup>lacZ</sup> parasites. (B) Wild-type *P230p* loci in NF54 and K1 parasites and the plasmids used for CRISPR/Cas9-mediated gene editing (*P230p-lacZ* and pHF-gC-*P230p*) to generate the NF54<sup>lacZ</sup> and K1<sup>lacZ</sup> parasites. The heterologous expression cassette inserted into the *P230p* locus consists of an *hsp70* promoter, the *lacZ* gene and a *cam* terminator. Arrows indicate primers used for diagnostic PCRs. Striped boxes represent the homology regions used for CRISPR/Cas9-editing. The guideRNA binding site is indicated. *cas9*, recombinase; *yfcu*, negative selection marker<sup>1</sup> (yeast cytosine deaminase and uridyl phosphoribosyl transferase); hb, homology boxes; wt, wild-type.

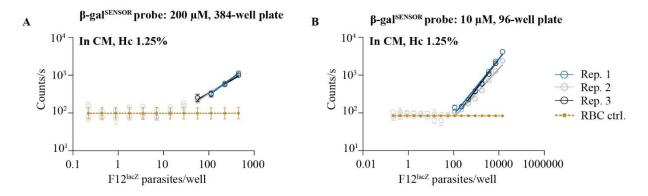


Figure S2. The  $\beta$ -gal<sup>SENSOR</sup> probe allows detecting low numbers of F12<sup>lacZ</sup> parasites under assay conditions. (A) Mixed-stage F12<sup>lacZ</sup> parasites are detected at low numbers (~50 parasites/well) when incubated with  $\beta$ -gal<sup>SENSOR</sup> probe at 200  $\mu$ M in CM with a Hc of 1.25% and emitted chemiluminescence linearly correlates with parasite numbers. (B) Compared to (A), the LOD increases (~200 parasites/well) when reducing the  $\beta$ -gal<sup>SENSOR</sup> probe to 10  $\mu$ M and performing the experiments in 96-wel plates (assay conditions). n = 3; biological replicates (rep.) are shown individually with error bars indicating standard deviations of technical replicates; for the control samples (ctrl.), biological replicates are averaged and error bars indicate standard deviations of the mean.

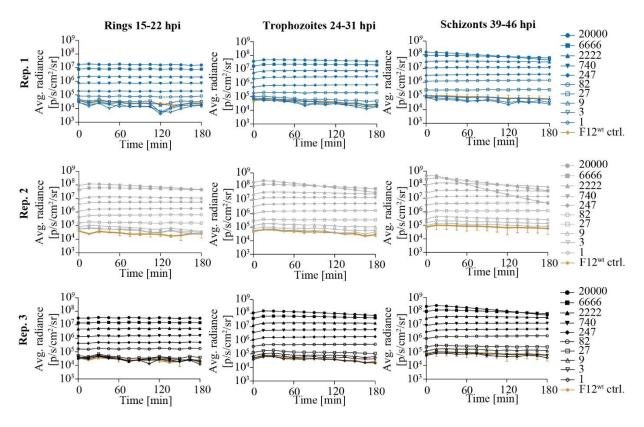


Figure S3. Luminescence emitted from F12<sup>lacZ</sup> parasites remains stable. Following addition of the  $\beta$ -gal<sup>SENSOR</sup> probe, luminescence flux is stable over a wide range of absolute parasite numbers for at least 180 min. Parasite numbers used per condition are indicated. 20`000 wild-type parasites (F12<sup>wt</sup>) were used as a negative control sample. n = 3.

		<b>Before free</b>	zing	After freezing
Pyrimethamine	150 -		150- 150- 100-	$\frac{1}{10} \frac{1}{100} \frac{1}{1000}$ Concentration [nM]
		IC	<sub>0</sub> β-gal <sup>SENSOR</sup> probe	assay
		Pyrimethamine	Before freezing	After freezing
		Rep. 1	-	30.51
		Rep. 2	-	49.99
		Rep. 3	-	33.44

Figure S4. Freezing of the  $\beta$ -gal<sup>SENSOR</sup> probe assay plates prior to the readout is strongly recommended. For pyrimethamine, it was not possible to calculate the IC<sub>50</sub> values when the assay plates were not frozen prior to the readout with 10  $\mu$ M  $\beta$ -gal<sup>SENSOR</sup> probe. Biological replicates (rep.) are shown individually. IC<sub>50</sub> values [nM] are indicated.

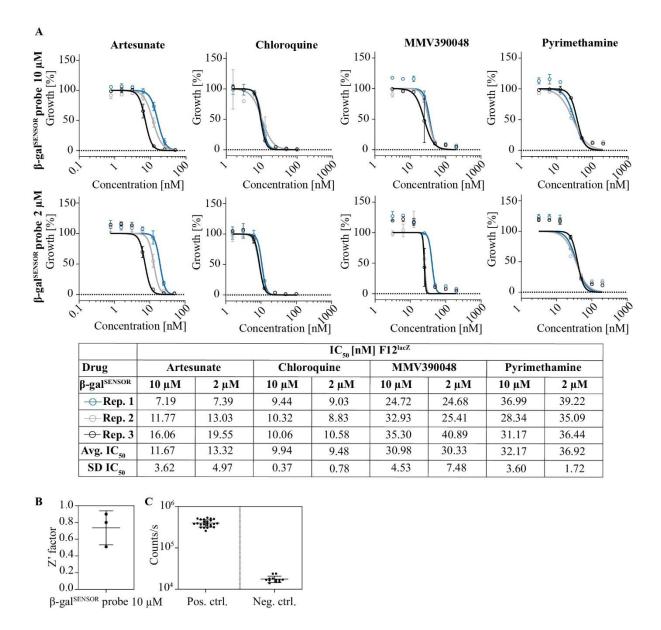
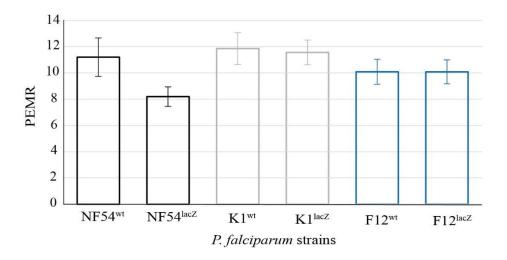


Figure S5. *lacZ*-expressing parasites allow monitoring dose-responses at different  $\beta$ -gal<sup>SENSOR</sup> probe concentrations. (A) Comparison the F12<sup>lacZ</sup>/ $\beta$ -gal<sup>SENSOR</sup> probe assay formats using either 10 or 2  $\mu$ M  $\beta$ -gal<sup>SENSOR</sup> probe. Biological replicates (Rep.) are shown individually. IC<sub>50</sub> values [nM] are indicated. Avg., average; SD, standard deviation. (B) Assay robustness using  $\beta$ -gal<sup>SENSOR</sup> probe at 10  $\mu$ M determined by the Z'-factor. n = 3. (C) Luminescence derived from F12<sup>lacZ</sup> parasite cultures lysed at the start of the experiment (negative control) and untreated F12<sup>lacZ</sup> following the 72-hour incubation period (positive control).



**Figure S6:** Parasitized Erythrocyte Multiplication Rate (PEMR) is comparable among parental *P*. *falciparum* strains and transgenic strains. n = 3. wt, wild type.

**Table S1. The**  $\beta$ -gal<sup>SENSOR</sup> probe IC<sub>50</sub> assay is cost effective. The price of one IC<sub>50</sub> assay plate (96-well format) using the *lacZ*/ $\beta$ -gal<sup>SENSOR</sup> probe 2  $\mu$ M system can be reduced by one third compared to the [<sup>3</sup>H]-hypoxanthine incorporation assay.

	IC <sub>50</sub> assay costs per plate		
Currency: USD	[ <sup>3</sup> H] incorporation	$\beta$ -gal <sup>SENSOR</sup> probe 2 $\mu$ M	
Drug preparation	3.29	3.31	
Plate preparation	9.15	12.85	
Readout reagent	12.86	5.88	
Readout consumables	7.82	-	
Total per plate	33.12	22.04	

## References

1. Manzoni G, Briquet S, Risco-Castillo V, Gaultier C, Topçu S, Ivănescu ML, Franetich JF, Hoareau-Coudert B, Mazier D, Silvie O. A rapid and robust selection procedure for generating drug-selectable marker-free recombinant malaria parasites. Sci Rep. 2014;4:4760. doi:10.1038/srep04760.