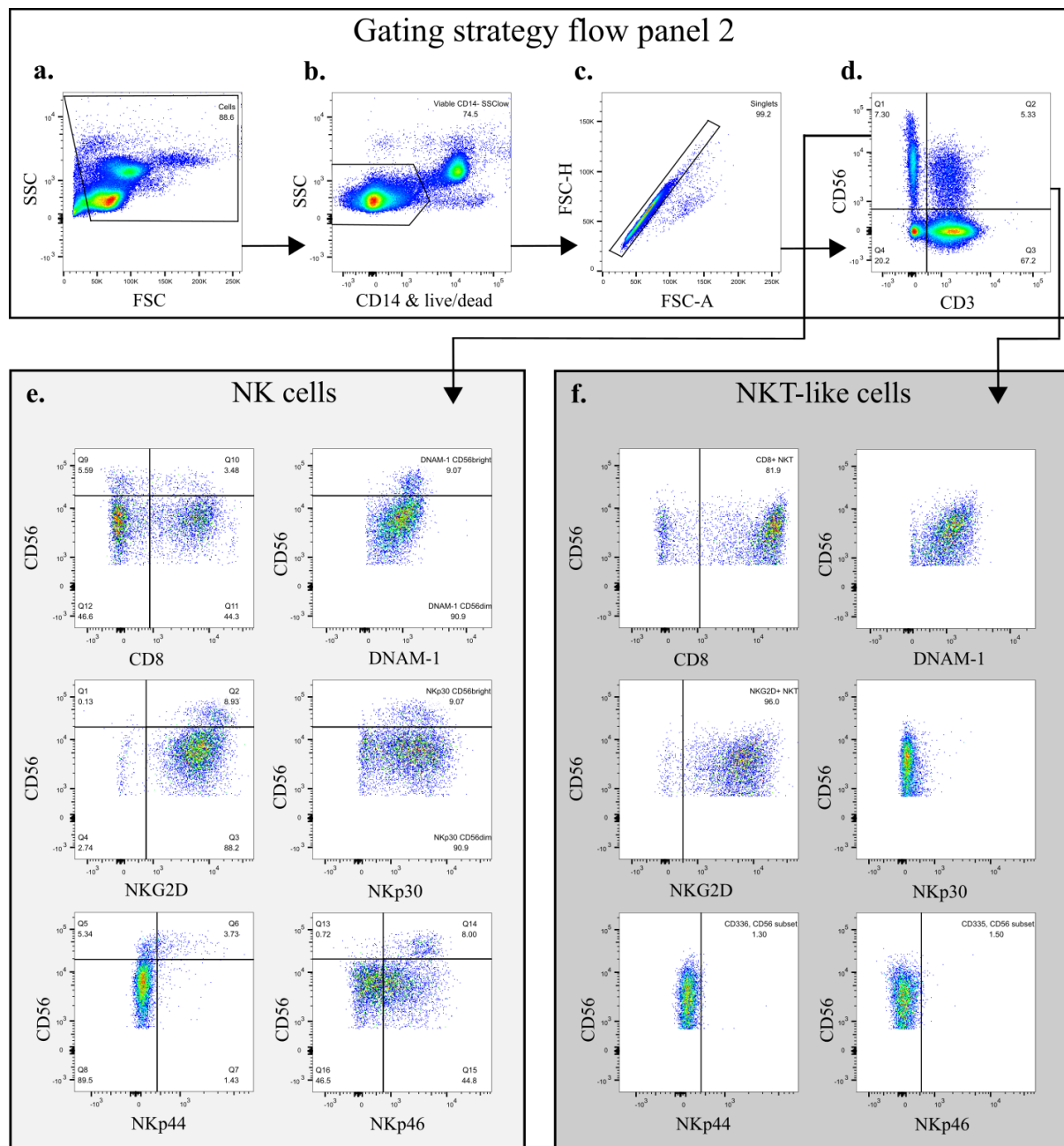


**Supplementary Fig.1** Flow cytometry gating strategy used for the identification of circulating NK cells and NKT-like cells

Two flow cytometry panels were set up in order to study the immunophenotype circulating NK cells and NKT-like cells. This figure shows the standardized gating strategy used for flow cytometry panel 1. **a.** Mononuclear cells (excluding debris (FSC<sup>low</sup>/SSC<sup>low</sup>)). **b.** Viable lymphocytes (excluding CD14<sup>+</sup> monocytes, dead cells, and SSC<sup>high</sup> cells). **c.** Single lymphocytes (excluding doublets). **d.** NK cells (CD3<sup>-</sup>CD56<sup>+</sup> cells) and NKT-like cells (CD3<sup>+</sup>CD56<sup>+</sup> cells). Percentage and expression of immunophenotypic markers were determined on the gated CD56<sup>dim</sup> and CD56<sup>bright</sup> NK cell populations **e.** and **f.** NKT cell-like population.

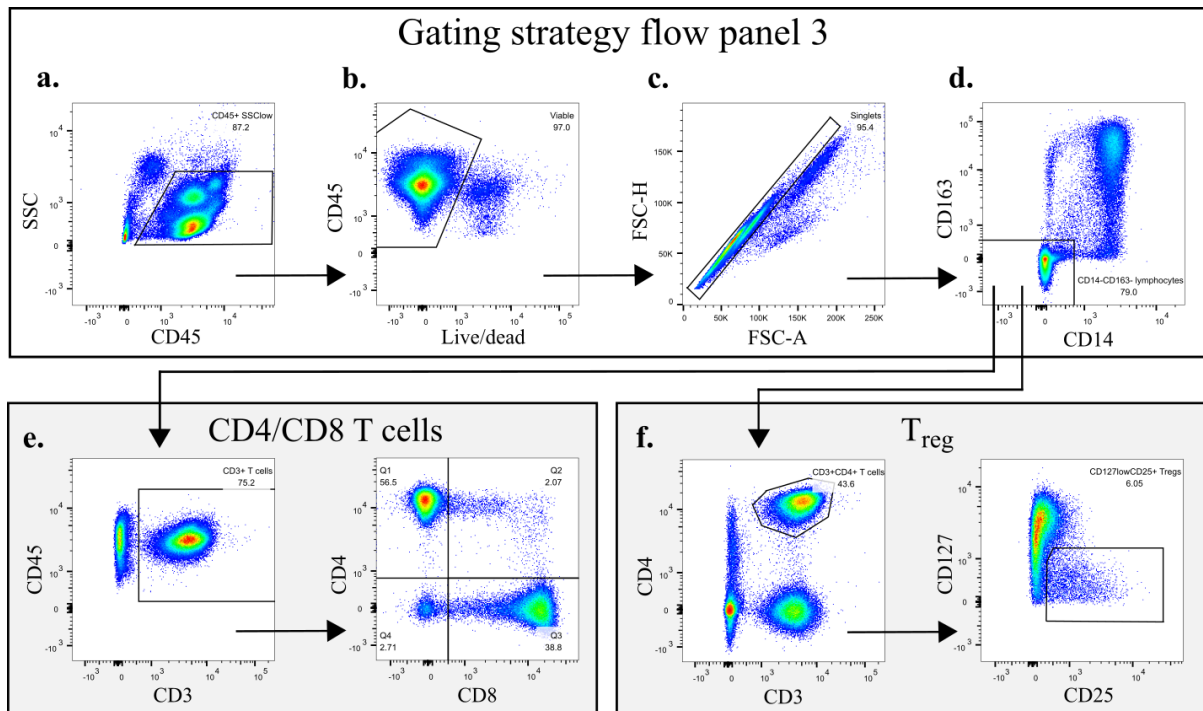
Abbreviations: FSC (forward scatter), SSC (side scatter).



**Supplementary Fig.2** Flow cytometry gating strategy used for the identification of circulating NK cells and NKT-like cells

Two flow cytometry panels were set up in order to study the immunophenotype circulating NK cells and NKT-like cells. This figure shows the standardized gating strategy used for flow cytometry panel 2. **a.** Mononuclear cells (excluding debris (FSC<sup>low</sup>/SSC<sup>low</sup>)). **b.** Viable lymphocytes (excluding CD14<sup>+</sup> monocytes, dead cells, and SSC<sup>high</sup> cells). **c.** Single lymphocytes (excluding doublets). **d.** NK cells (CD3<sup>-</sup>CD56<sup>+</sup> cells) and NKT-like cells (CD3<sup>+</sup>CD56<sup>+</sup> cells). Percentage and expression of immunophenotypic markers were determined on the gated CD56<sup>dim</sup> and CD56<sup>bright</sup> NK cell populations **e.** and **f.** NKT cell-like population.

Abbreviations: FSC (forward scatter), SSC (side scatter).



**Supplementary Fig.3** Overview of the flow cytometry gating strategy used for the identification of circulating T cell subsets

A third flow cytometry panel was set up and a standardized gating strategy was created in order to study the immunophenotype circulating T cell subsets. **a.** Mononuclear cells (excluding CD45<sup>-</sup> cells and SSC<sup>high</sup> cells). **b.** Viable mononuclear cells (excluding dead cells). **c.** Single mononuclear cells (excluding doublets). **d.** Lymphocytes (CD14<sup>-</sup>CD163<sup>-</sup> cells). **e.** T cells (CD3<sup>+</sup>), further divided into CD4<sup>+</sup> versus CD8<sup>+</sup> T cells. **f.** Helper T cells (CD3<sup>+</sup>CD4<sup>+</sup>), and their subpopulation T<sub>reg</sub> (CD127<sup>low</sup>CD25<sup>+</sup> cells).

Abbreviations: FSC (forward scatter), SSC (side scatter)



**Supplementary Table 2** *Survival analyses of CRC patients in relation to immune subset distribution and immunophenotype of circulating lymphocyte subsets*

Survival plots and univariate analyses were generated for stage II and III CRC patients (N=49) at risk of developing metastases. Stratifications were based on the median percentage of positive cells or expression of the respective immunophenotypic marker. The below-median group was used as reference group. *P*-values  $\leq 0.05$  were considered statistically significant and indicated in bold.

	Kaplan Meier		Univariate analysis		
	Median	P-value	HR	CI	P-value
<b>Subset distribution</b>					
<i>T</i> cells (%) <sup>*</sup>	54.6	0.399	0.655	0.243-1.765	0.403
<i>CD8</i> <sup>+</sup> <i>T</i> cells (%) <sup>*</sup>	16.6	0.222	0.536	0.194-1.481	0.230
<i>CD4</i> <sup>+</sup> <i>T</i> cells (%) <sup>*</sup>	42.6	0.980	1.013	0.379-2.702	0.980
<i>CD127</i> <sup>low</sup> <i>CD25</i> <sup>+</sup> <i>T</i> <sub>reg</sub> (%) <sup>*</sup>	6.9	0.062	2.551	0.921-7.069	0.072
<i>NK</i> cells (%)	13.3	0.170	1.661	0.798-3.459	0.175
<i>CD56</i> <sup>dim</sup> <i>NK</i> cells (%)	96.7	0.585	1.226	0.589-2.550	0.586
<i>CD56</i> <sup>bright</sup> <i>NK</i> cells (%)	3.3	0.380	0.719	0.343-1.507	0.382
<i>NKT</i> -like cells (%)	4.4	0.436	1.335	0.644-2.767	0.438
<b><i>CD56</i><sup>dim</sup> <i>NK</i> cells</b>					
<i>CD16</i> <sup>+</sup> (%)	83.4	0.412	1.358	0.652-2.830	0.413
<i>CD158a</i> <sup>+</sup> (%)	29.9	0.832	0.832	0.443-1.926	0.832
<i>CD158b</i> <sup>+</sup> (%)	34.6	0.462	1.316	0.632-2.742	0.463
<i>NKG2A</i> <sup>+</sup> (%)	47.2	0.948	0.976	0.470-2.025	0.948
<i>NKG2A</i> <sup>+</sup> (MFI)	7212	0.578	0.813	0.391-1.689	0.578
<i>NKG2C</i> <sup>+</sup> (%)	12.9	0.559	0.804	0.386-1.675	0.560
<i>NKG2C</i> <sup>+</sup> (MFI)	2015	0.605	0.824	0.396-1.716	0.606
<i>CD161</i> (MFI)	2670	0.606	0.825	0.397-1.716	0.606
<i>CD8</i> <sup>+</sup> (%)	25.9	0.645	0.843	0.406-1.749	0.646
<i>CD8</i> <sup>+</sup> (MFI)	3091	0.894	1.052	0.500-2.214	0.894
<i>DNAM-1</i> (MFI)	555	0.464	1.313	0.632-2.725	0.465
<i>NKG2D</i> <sup>+</sup> (%)	91.3	0.120	1.784	0.851-3.739	0.125
<i>NKG2D</i> <sup>+</sup> (MFI)	3414	0.841	1.078	0.517-2.248	0.841
<i>NKp30</i> (MFI)	1390	0.180	1.663	0.785-3.525	0.184
<i>NKp44</i> <sup>+</sup> (%)	0.5	0.910	1.043	0.500-2.175	0.910
<i>NKp44</i> (MFI)	111	0.240	1.581	0.732-3.412	0.243
<i>NKp46</i> <sup>+</sup> (%)	33.7	0.343	1.434	0.679-3.029	0.345
<i>NKp46</i> (MFI)	452	0.141	1.768	0.821-3.806	0.146
<b><i>CD56</i><sup>bright</sup> <i>NK</i> cells</b>					
<i>CD16</i> <sup>+</sup> (%)	1.6	0.358	0.708	0.338-1.484	0.360
<i>CD158a</i> <sup>+</sup> (%)	5.2	0.244	1.540	0.741-3.200	0.248
<i>CD158b</i> <sup>+</sup> (%)	7.6	0.641	1.189	0.574-2.466	0.641
<i>NKG2A</i> <sup>+</sup> (%)	3.2	0.734	0.881	0.423-1.834	0.734
<i>NKG2A</i> <sup>+</sup> (MFI)	17899	0.770	1.115	0.537-2.314	0.770
<i>NKG2C</i> <sup>+</sup> (%)	0.9	0.414	0.735	0.351-1.541	0.415
<i>NKG2C</i> <sup>+</sup> (MFI)	2208	0.303	0.679	0.323-1.426	0.306
<i>CD161</i> (MFI)	1616	0.683	0.858	0.413-1.786	0.683
<i>CD8</i> <sup>+</sup> (%)	1.2	0.655	0.846	0.406-1.761	0.655
<i>CD8</i> <sup>+</sup> (MFI)	2897	0.626	1.202	0.573-2.518	0.627
<i>DNAM-1</i> (MFI)	774	0.232	1.561	0.748-3.259	0.236
<i>NKG2D</i> <sup>+</sup> (%)	3.2	0.380	0.719	0.343-1.507	0.382
<i>NKG2D</i> <sup>+</sup> (MFI)	6342	0.376	1.394	0.666-2.918	0.378
<i>NKp30</i> (MFI)	2327	<b>0.042</b>	2.143	1.009-4.551	<b>0.047</b>
<i>NKp44</i> <sup>+</sup> (%)	0.9	0.152	0.581	0.274-1.232	0.157
<i>NKp44</i> (MFI)	238	0.776	1.112	0.535-2.311	0.776
<i>NKp46</i> <sup>+</sup> (%)	85.4	0.287	1.487	0.713-3.100	0.290
<i>NKp46</i> (MFI)	2174	0.822	0.919	0.442-1.914	0.822
<b><i>NKT</i>-like cells</b>					
<i>CD16</i> <sup>+</sup> (%)	20.3	<b>0.000</b>	4.697	2.046-10.783	<b>0.000</b>
<i>CD158a</i> <sup>+</sup> (%)	5.1	0.068	1.969	0.937-4.136	0.074
<i>CD158b</i> <sup>+</sup> (%)	9.2	0.163	1.684	0.803-3.531	0.167
<i>NKG2A</i> <sup>+</sup> (%)	18.3	0.108	1.823	0.868-3.829	0.113
<i>NKG2A</i> <sup>+</sup> (MFI)	2472	0.780	1.110	0.533-2.315	0.780
<i>NKG2C</i> <sup>+</sup> (%)	10.7	0.462	1.314	0.634-2.726	0.463
<i>NKG2C</i> <sup>+</sup> (MFI)	1455	0.459	1.318	0.633-2.742	0.460
<i>CD161</i> (MFI)	1630	0.682	0.858	0.413-1.784	0.682
<i>CD8</i> <sup>+</sup> (%)	75.9	0.442	1.332	0.640-2.775	0.444
<i>CD8</i> <sup>+</sup> (MFI)	12934	0.523	1.268	0.611-2.632	0.524
<i>DNAM-1</i> (MFI)	765	0.603	1.215	0.582-2.534	0.604
<i>NKG2D</i> <sup>+</sup> (%)	92.9	0.696	1.157	0.556-2.409	0.697
<i>NKG2D</i> <sup>+</sup> (MFI)	4704	0.394	1.385	0.653-2.940	0.396
<i>NKp30</i> (MFI)	182	0.784	1.108	0.532-2.308	0.784
<i>NKp44</i> <sup>+</sup> (%)	1.2	0.221	1.591	0.751-3.370	0.225
<i>NKp44</i> (MFI)	114	0.380	1.394	0.662-2.937	0.382

<i>NKp46<sup>+</sup> (%)</i>	3.6	0.316	0.319	0.698-3.024	0.319
<i>NKp46 (MFI)</i>	53.4	0.563	1.240	0.598-2.572	0.564

\*T cells were investigated in 25 CRC patients at risk of developing metastases