Supporting Information

Potentiating Activity of GmhA Inhibitors on Gram-negative Bacteria

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Table of Contents

General Methods	S2		
Synthetic procedures for compounds 17, 20, 24, 25, 27, 31 and 96	S3		
Fig. S1. HILIC reaction control of the final deprotection and stacked ¹ H NMR plot of individual			
fractions of compound 17.	S8		
¹ H, ¹³ C and ³¹ P NMR spectra of compounds 17 , 20 , 24 , 25 , 27 , 31 and 96	S19		
Fig. S2. 1D NOe-difference spectrum of compound 17 with selective saturation of one formyl proton			
Synthetic procedures reference for compounds 41, 54, 68, 76 and 84	S20		
Table S1. X-ray data collection and refinement statistics	S31		
Fig. S3. Role of UhpT phospho-sugar transporter in the transport of D-glycero-D-manno-heptose 7-			
phosphate (H7P)	S34		
Fig. S4 : IC ₅₀ fits of <i>E. coli</i> GmhA in biochemical inhibition assays	S35		
Fig. S5: EC ₅₀ fits of LPS biosynthesis inhibition assays	S36		

Experimental section

General methods

All purchased chemicals were used without further purification unless stated otherwise. Solvents were dried over activated 4 Å (CH₂Cl₂, pyridine, THF, DMF) and 3 Å (CH₃CN) molecular sieves. Cation exchange resin DOWEX 50 H⁺ was regenerated by consecutive washing with HCl (3 M), water and dry MeOH. Aqueous solutions of salts were saturated unless stated otherwise. Concentration of organic solutions was performed under reduced pressure < 40 °C. Optical rotations were measured with a Perkin-Elmer 243 B or Anton Paar MCP100 Polarimeter. Thin layer chromatography was performed on Merck precoated plates: generally on 5 x 10 cm, layer thickness 0.25 mm, Silica Gel 60F₂₅₄; alternatively on HPTLC plates with 2.5 cm concentration zone (Merck). Spots were detected by dipping reagent (anisaldehyde-H₂SO₄) followed by heating. For column chromatography silica gel (0.040 – 0.063 mm) was used. HP-column chromatography was performed on pre-packed columns (YMC-Pack SIL-06, 0.005 mm, 25 x 1 cm and 25 x 2 cm). HPLC (analytical and preparative) was performed on an Agilent 1100 HPLC with diode array detection. Preparative HPLC was performed at 0.7 mL/min on a Thermo Electron, Hypersil BDS C-18 column (250 x 4.6 mm, 5 µm) using a gradient of acetonitrile and water with 0.1% TFA (50% in acetonitrile to 100% and then back to 50%). The tested compounds were determined to be >95% pure via HPLC.

NMR spectra were recorded with a Bruker Avance III 600 instrument (600.22 MHz for ¹H, 150.93 MHz for ¹³C, 242.9 MHz for ³¹P) or a Bruker Avance 300 instrument using standard Bruker NMR software (Topspin3.6.2). ¹H spectra were referenced to 7.26 (CDCl₃), 3.34 (MeOD) and 0.00 (D₂O, external calibration to 2,2-dimethyl-2-silapentane-5-sulfonic acid) ppm unless stated otherwise. ¹³C NMR spectra were referenced to 77.00 (CDCl₃), 49.00 (MeOD) and 67.40 (D₂O, external calibration to 1,4-dioxane) ppm. ³¹P NMR spectra were referenced to external ortho-phosphoric acid (δ 0.0) for solutions in D₂O. Assignments were based on COSY, HSQC, HMBC and TOCSY data. ESI-MS data were obtained on a Micromass Q-TOF Ultima Global instrument.

General procedure for the HILIC based monitoring of final deprotection steps:

Reactions were monitored on a ZIC HILIC (Merck 4.6x 150 mm (CatNo 1.50444.001) analytical HPLC column attached to a Shimadzu LC 10 HPLC system equipped with an Alltech 3300 ELSD and a Shimadzu LC MS 2020 Detector. Acetonitrile/water, both using 0.1% (v/v) as modifier for LCMS was used as solvent system. The gradient employed at a flow rate of 0.75 ml*min⁻¹ a gradient starting from 95% acetonitrile which went after a 2 min plateau at 95% down to 40% acetonitrile over 15 minutes, after which a 40% acetonitrile plateaued was kept for 7 minutes before returning to the

initial 95% acetonitrile concentration. Deprotected compounds usually eluted in or close to the 40% acetonitrile plateau.

General procedure for the purification of compounds via HILIC chromatography:

After final deprotection the compound was dissolved in H₂O and made neutral by adding NaHCO₃. The solution was passed over a short plug of HILIC material (ZIC HILIC SPE SeQuant 500 mg) using 2:1 MeCN-H₂O as eluant. Product containing fractions were determined by TLC, concentrated and lyophilized. The residue was taken up in acetonitrile and water was added slowly until a clear solution for injection was obtained. The sample was injected via a sample loop into a semi preparative SeQuant ZIC HILIC column (Merck, 25x1 cm, Cat No 1.50494.0001) which was connected to a HPLC system equipped with a binary pump system and a fraction collector. The column was equilibrated with acetonitrile/water 85/15. A gradient from 85% to 40% acetonitrile at a flow rate of 3 ml/m over 7 column volumes followed by a short plateau of 1.5 column volumes at 40% acetonitrile with a final backwashing phase to 85% acetonitrile was employed. Fractions were collected at a size of 2 ml. TLC positive (staining with phosphor molybdate) fractions were individually concentrated and freeze dried (SpeedVac) and each fraction was individually checked by ¹H NMR. Pure product fractions were combined and lyophilized.

2,3,4-Tri-O-benzyl-5-O-triisopropylsilyl-D-ribose diethyldithioacetal (11). Compound 10 (300 mg, 0.57 mmol) was dissolved in dry pyridine (2 mL). DMAP (70 mg, 0.57 mmol) and triisopropylsilyl chloride (183 µL, 0.85 mmol) were added and the reaction mixture was stirred at room temperature for 5 h. Additional amounts of DMAP (70 mg, 0.57 mml) and TIPSCI (183 µL, 0.85 mmol) was added and stirring continued for 9 h. The reaction mixture was diluted and co-evaporated with toluene. The residue was dissolved in Et_2O (15 mL) and washed with satd aqu NaHCO₃. The organic phase was dried (MgSO₄), concentrated, and purified via column chromatography on silica gel (toluene-hexane 2:1→5:1) to give **11** (356 mg, 93%) as colorless oil; $[\alpha]_D^{20}$ +20.7 (*c* 1.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.36-7.22 (15 H, Ar-H), 4.98-4.57 (m, 6 H, 3 x OCH₂Ph), 4.26 (d, 1 H, J_{1,2} = 3.3 Hz, H-1), 4.13 (dd, 1 H, J_{3,2} = 7.5 Hz, J_{3,4} = 1.9 Hz, H-3), 4.05 (dd, 1 H, J_{2,1} = 3.3 Hz, J_{2,3} = 7.5 Hz, H-2), 3.99-3.91 (m, 2 H, H-4, H-5a), 3.89-3.79 (m, 1 H, H-5b), 2.70-2.55 (m, 4 H, 2 x SCH₂CH₃), 1.24-1.13 (m, 6 H, 2 x SCH₂CH₃), 1.05-0.95 [m, 21 H, Si(CHMe₂)₃]. ¹³C NMR (150 MHz, CDCl₃): δ 139.00, 138.62, 138.51 (3 x Ar-C_q), 128.25, 128.15, 128.11, 127.82, 127.68, 127.47, 127.80, 127.28,127.26 (Ar-C), 82.78 (C-2), 81.04 (C-4), 78.81 (C-3), 74.75, 73.15, 72.93 (3 x OCH₂Ph), 64.10 (C-5), 54.14 (C-1), 26.21, 24.95 (2 x SCH₂CH₃), 18.02 [Si(CH(CH₃)₂)₃], 14.46, 14.44 (2 x SCH₂CH₃), and 11.91 [Si(CH(CH₃)₂)₃].ESI-HRMS: *m/z* calcd for C₃₉H₅₈O₄S₂Si [M+Na⁺]⁺: 705.3438; found: 705.3437.

2,3,4-Tri-O-benzyl-5-O-triisopropylsilyl-D-ribose (12). NBS (464 mg, 2.6 mmol) was added at 0 °C to a solution of **11** (356 mg, 0.52 mmol) in a 5:1 mixture of acetone-water (12 mL). The reaction mixture was stirred for 30 min at 0 °C. Satd aqu NaHCO₃ (5 mL) followed by addition of satd aq Na₂S₂O₃ (5 mL) and stirring was continued for 15 min. The solution was concentrated, and the remaining aqueous phase was extracted with Et₂O (2 x 10 mL). The combined organic phases were dried (MgSO₄) and concentrated. The residue was purified by silica gel chromatography (toluene-hexane 1:1→ toluene) to give **12** (292 mg, 97%) as colorless syrup: $[\alpha]_D^{20}$ +12.2 (*c* 1.5, CHCl₃); ¹H NMR (300 MHz CDCl₃): δ 9.44 (d, 1 H, *J* = 0.9 Hz, CHO), 7.36-7.24 (m, 15 H, Ar-H), 4.85-4.47 (m, 6 H, 3 x OCH₂Ph), 4.10 (dd, 1 H, *J*_{2,3} = 2.1 Hz, H-2), 4.03-3.91 (m, 2 H, H-3, H-5a), 3.84-3.76 (m, 2 H, H-4, H-5b), 1.13-0.96 [m, 21 H, Si(CHMe₂)₃]. ¹³C NMR (75 MHz CDCl₃): δ 201.41 (HC=O), 138.27, 137.77, 137.59 (Ar-C_q), 128.46, 128.39, 128.21, 127.94, 127.89, 127.85, 127.80, 127.44 (Ar-C), 82.60 (C-2), 80.87 (C-3), 78.63 (C-4), 73.17, 73.11, 72.74 (3 x OCH₂Ph), 63.53 (C-5), 18.04,18.03 [Si(CH(CH₃)₂)₃] and 11.95 8 [Si(CH(CH₃)₂)₃]. ESI-MS: *m/z* calcd for C₃₅H₄₈O₅Si [M+H⁺]⁺: 577.33; found 577.45.

1-N-Benzyloxyamino-2,3,4-tri-O-benzyl-1-deoxy-5-O-triisopropylsilyl-D-ribitol (13). Compound 12 (285 mg, 0.49 mmol) and O-benzylhydroxylamine (86 μL, 0.74 mmol) were stirred at 60 °C in 1:6.5 pyridine-MeOH (6 mL) overnight. An additional portion of O-benzylhydroxylamine (20 µL) was added and stirring continued for 12 h. The solution was concentrated, and the residue was subjected to flash-chromatography over a short plug of silica gel (2 g, hexane-EtOAc 10:1) to give the crude oxime (317 mg, 0.47 mol, 96%) as colorless oil. An aliquot of the oxime (150 mg, 0.22 mmol) was dissolved in acetic acid (1.5 ml) followed by portionwise addition of sodium cyanoborohydride (124 mg, 1.98 mmol) and stirring for 20 min at room temperature. The reaction mixture was diluted with water and twice extracted with dichloromethane. The organic phase was made neutral by addition of satd aq NaHCO₃ followed by drying (MgSO₄) and removal of the solvent. The residue was purified by HPLC (hexane-EtOAc 15:1) to give **13** (123 mg, 75% over two steps) as colorless oil; $[\alpha]_D^{20}$ -25.8 (c 1.1, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 7.33-7.21 (20 H, Ar-H), 5.92 (bs, 1 H, NH), 4.80-4.48 (m, 8 H, 4 x OCH₂Ph), 4.12 (dt, 1 H, J_{1b,2} = 8.0 Hz, J_{1a,2} = 3.3 Hz, H-2), 4.00 (dd, 1 H, J_{5a,5b} = 10.7 Hz, J_{5a,4} = 3.7 Hz, H-5a), 3.88 (dd, 1 H, J_{3,2} = 3.4 Hz, J_{3,4} = 6.1 Hz, H-3), 3.83 (dd, 1 H, J_{5b,4} = 6.0 Hz, H-5b), 3.62 (dt, 1 H, H-4), 3.27 (dd, 1 H, J_{1a,1b} = 13.7 Hz, J_{1a,2} = 3.3 Hz, H-1a), 3.11 (dd, 1 H, J_{1b,2} = 8.1 Hz, H-1b), 1.14-0.96 [m, 21 H, Si[CH(CH₃)₂)₃]. ESI-HRMS: *m/z* calcd for C₄₂H₅₇NO₅Si [M+H⁺]⁺ 684.4079; found 684.4078.

1-N-Benzyloxy-N-formylamino-2,3,4-tri-O-benzyl-1-deoxy-5-O-triisopropylsilyl-D-ribitol (14) Formic acid (41 μ L, 1.08 mmol) was added to a solution of carbonyl diimidazole (CDI) (175 mg, 1.0 mmol) in THF (2 mL) at 0 °C was added and stirred at 0 °C for 30 min. Then a solution of **13** (148 mg, 216 μ mol) in THF (1 mL) was added and the reaction mixture was stirred at 0 °C for 1 h. The reaction mixture was concentrated, and the residue was purified on silica gel (hexane-EtOAc 3:1) to give **14** (146 mg, 94 %) as colorless oil; $[\alpha]_D{}^{20}$ -5.4 (*c* 0.9, CHCl₃). ¹H NMR (600 MHz, toluene-d₈, 80 °C): δ 8.07 (bs, 1 H, NCHO), 7.33-7.02 (20 H, Ar-H), 4.79-4.55 (m, 8 H, 4 x OCH₂Ph), 4.23-4.19 (m, 1 H, H-2), 4.03 (dd, 1 H, *J*_{5a,5b} = 10.6 Hz, *J*_{5a,4} = 3.7 Hz, H-5a), 3.93-3.88 (m, 2 H, H-3, H-5b), 3.82-3.71 (m, 3 H, H-4, H-1a, H-1b), 1.08-1.03 [m, 21 H, Si[CH(CH₃)₂)₃]. ¹³C NMR (150 MHz, toluene-d₆, 80 °C, HSQC data): δ 139.54, 139.33, 139.12 (Ar-C_q, overlapped with toluene signals), 129.56, 129.27, 128.74, 128.70, 128.60, 128.55, 128.36, 128.32, 127.84, 127.80 (Ar-C), 81.30 (C-4), 80.10 (C-3), 77.23 (C-2), 77.33 (NOCH₂Ph), 73.98, 73.83, 72.29 (3 x OCH₂Ph), 64.87 (C-5), 48.94 (C-1), 18.42 [Si(CH(*C*H₃)₂)₃], 12.76 [Si(CHMe₂)₃]. ESI-HRMS: *m/z* calcd for C₄₃H₅₇NO₆Si [M+H⁺]⁺ 712.4028; found 712.4037.

1-*N*-**Benzyloxy-***N***-formylamino-2,3,4-tri-***O***-benzyl-1-deoxy-D-ribitol (15)** Compound 14 (146 mg, 0.20 mmol) was dissolved in dry THF (1 mL) and TBAF (308 μl of a 1 M solution in THF) was added. The mixture was stirred at rt for 30 minutes when TLC showed complete conversion. The reaction mixture was concentrated, and the residue dissolved in EtOAc, filtered through a short plug of silica gel (2 g) and concentrated. Purification by HPLC (hexane-EtOAc 2:1→1:1) gave 15 as colorless syrup (100 mg, 87%). [α]_D²⁰ +44 (*c* 0.5, CHCl₃), ¹H NMR (600 MHz, toluene- d₈, 80 °C): δ 8.06 (bs, 1 H, NCHO), 7.25-7.02 (20 H, Ar-H), 4.65-4.40 (m, 8 H, 4 x OCH₂Ph), 4.12-4.08 (m, 1 H, H-2), 3.81 (dd, 1 H, *J* = 6.5 Hz, *J* = 2.5 Hz, H-3), 3.75-3.62 (m, 4 H, H-5a, H-5b, H-1a,H-1b), 3.60-3.56 (m, 1 H, H-4), ¹³C NMR (150 MHz, toluene- d₈, 80°C): δ 129.19, 128.91, 128.39, 128.36, 128.34, 128.26, 128.10, 128.00, 127.96 (Ar-C), 79.92 (C-4), 79.84 (C-3), 77.11 (NOCH₂Ph), 76.94 (C-2*), 73.96, 72.91, 72.56 (3 x OCH₂Ph), 61.52 (C-5), 48.66* (C-1). *determined by HSQC. ESI-HRMS: *m/z* calcd for C₃₄H₃₇NO₆ [M+H⁺]⁺ 556.2694; found 556.2702.

1-*N*-**Benzyloxy-***N***-formylamino-2,3,4-tri-***O***-benzyl-5-dibenzyloxyphosphoryl-1-deoxy-D-ribitol (16**). Compound **15** (100 mg, 0.18 mmol) was twice evaporated with toluene and the residue was dried under vacuum for 12 h. Dibenzyl *N*,*N*-diisopropylaminophosphoramidite (91 μL, 0.27 mmol) was added to a solution of **15** in anhydrous DCM (1.5 mL) followed by slow addition of 1*H*-tetrazole (0.45 M in MeCN, 600 μL, 0.27 mmol) at rt. The solution was stirred for 40 min and then cooled to -78 °C. A solution of *m*CPBA (70%, 71 mg, 0.28 mmol) in DCM (1 mL) was added slowly to the reaction mixture and stirred for 20 min at -78 °C. Triethylamine (40 μL, 0.28 mmol) was added and the reaction mixture was allowed to warm to rt. The reaction mixture was diluted with DCM (10 mL), washed with sat aqu NaHCO₃, dried (MgSO₄) and concentrated. Purification of the residue on silica gel (hexane-EtOAc 1:2) followed by HPLC (hexane-EtOAc 2:1 → 1:1) gave **16** as colorless oil (137 mg, 93%); [α]_D²⁰ -44 (*c* 0.5, CHCl₃), ¹H NMR (600 MHz, toluene- d₈, 80°C): δ 8.04 (bs, 1 H, NCHO), 7.33-6.99 (30 H, Ar-H), 4.95-4.87 (m, 2 H, P(O)CH₂Ph), 4.66-4.41 (m, 9 H, 4 x OCH₂Ph, H-5a), 4.25-4.20 (m, 1 H, H-5b), 4.12-4.09 (m, 1 H, H-2), 3.84-3.79 (m, 2 H, H-3, H-4), 3.75-3.58 (m, 2 H, CH₂N). ¹³C NMR (150 MHz, toluene- d₈, 80°C): δ 129.59, 129.28, 128.77, 128.68, 128.65, 128.63, 128.53, 128.50, 128.36 (Ar-C), 79.56 (C-3), 78.86 ($J_{P;C}$ = 6.2 Hz, C-4), 77.45 (NOCH₂Ph), 77.17 (C-2), 74.17, 73.33, 73.19 (3 x OCH₂Ph), 69.48 (2 x P(O)CH₂Ph), 67.05 ($J_{P,C}$ = 5.4, C-5), 49.63* (C-1).*determined by HSQC, ESI-HRMS: m/z calcd for C₄₈H₅₀NO₉P [M+Na⁺]⁺ 838.3115; found 838.3116.

1-N-Formyl-N-hydroxyamino-1-deoxy-5-phosphoryl-D-ribitol monosodium salt (17) Pd(OH)₂ (10 mg) was added to a solution of compound 16 (50 mg, 61 μ mol) in 5:1:1 THF-H₂O-AcOH (5 mL,) and stirred under H₂ atmosphere at rt for 10.5 h. The progress of the reaction was monitored by HPLC-MS/ELS (Sequant ZIC HILIC 4.6 x 150 mm, gradient MeCN-H₂O 85 \rightarrow 40 % MeCN). The suspension was filtered through a syringe filter, the syringe filter was washed with 1:1 H₂O-MeOH (10 mL), 1:1 H₂O-THF (10 mL) and the combined filtrates were concentrated. The residue was dissolved in H₂O and made neutral by adding NaHCO₃ (11 mg, 0.1 mmol). The solution was passed over a short plug of ZIC-HILIC silica gel (500 mg) using 2:1 MeCN-H₂O as eluant. Product containing fractions were concentrated and lyophilized. The residue (21 mg) was redissolved in 2:1 MeCN-H₂O (300 μL) and purified by repeated ZIC-HILIC chromatography (Sequant ZIC HILIC 10 x 250 mm, gradient MeCN-H₂O 85 \rightarrow 40%). Fractions were concentrated, individually checked by ¹H NMR analysis and pooled accordingly (Fig.S1 Supporting Information). Lyophilization of the combined fractions gave **17** (5.2 mg, 30%) as colorless solid; $[\alpha]_{D}^{20}$ -15 (c 0.5, H₂O), ¹H NMR (600 MHz, D₂O, rotameric mixture): δ 8.33 (NCHO minor), 7.90 (NCHO major), 4.17-4.13 (m, 1 H, H-2), 3.98-3.89 (m, 2 H, H-5a, H-5b), 3.84-3.80 (m, 1 H, H-4), 3.77 (dd, 1 H, J_{3,4} = 7.2 Hz, J_{3,2} = 5.2 Hz, H-3), 3.71 (dd, 1 H, J_{1a,1b} = 14.9 Hz, J_{1a,2} = 2.8 Hz, H-1a), 3.66 (dd, 1 H, J_{1b,2} = 8.8 Hz, H-1b), ¹³C NMR (150 MHz, D₂O): δ 165.11* (NCHO minor), 161.11* (NCHO, major), 73.10 (C-3), 71.91 (J_{C,P} = 6.3 Hz, C-4), 68.08 (C-2), 65.71 (J_{C,P} = 4.6 Hz, C-5), 53.60 (C-1), ³¹P NMR (242 MHz, D₂O): δ 3.67; ESI-HRMS: m/z calcd for C₆H₁₄NO₉P [M-H]⁻ 274.0333; found 274.0337.

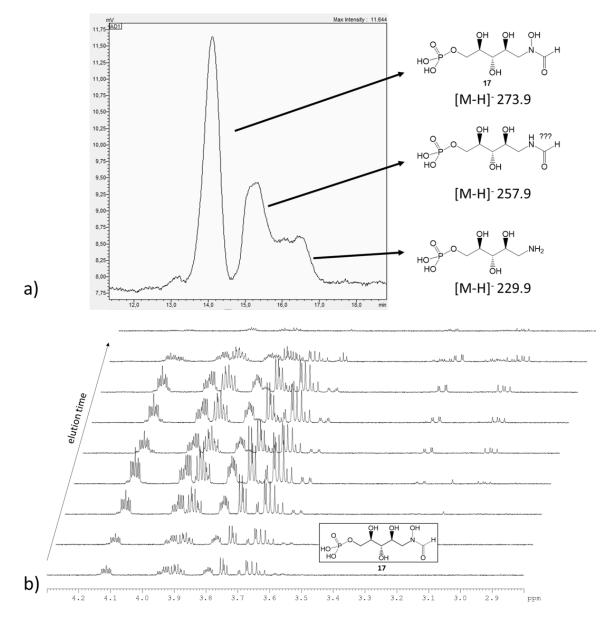


Fig. S1. a) ELSD track of a typical analytical HILIC reaction control of the final deprotection; b) stacked ¹H NMR plot of individual fractions of a preparative HILIC purification of compound **17**.

2,3,4-Tri-*O***-benzyl-1-(***N***-benzyloxy-***N***-formylamino)-1,5-dideoxy-5-iodo-D-ribitol (18)**. A solution of imidazole (44 mg, 0.65 mmol), triphenylphosphine (142 mg, 0.54 mmol), I₂ (137 mg, 0.54 mmol) and compound **15** (250 mg, 0.45 mmol) in anhydrous toluene (5 mL) was stirred under argon at 80 °C for 1 h. The dark yellow reaction mixture was cooled to rt and concentrated. The residue was taken up in DCM (10 mL), filtered through cotton, washed with 5% aq Na₂S₂O₃ (2 x 5 mL) and NaHCO₃ (5 mL). The aqueous phases were reextracted with DCM (10 mL) and the combined organic phases were dried (MgSO₄) and filtered. Silica gel (3 g) was added to the filtrate and the solvent was removed. The remaining silica gel was directly applied onto a prepacked silica gel column. Elution with 2:1 hexane-EtOAc afforded the target compound **18** (240 mg, 66%) as colorless oil; R_f 0.6 (hexane-EtOAc 2:1);

[α]_D²⁰ -12 (*c* 0.5, CHCl₃). ¹ H NMR (600 MHz, toluene-d₈, 85 °C) δ: 8.03 (s, 1 H, NCHO), 7.28-6.92 (m, 20 H, Ar-H), 4.67-4.27 (m, 8 H, 4 x OCH₂Ph), 4.09 (m, 1 H, H-2), 3.76 (m, 1 H, H-3), 3.72-3.55 (2 x bs, 2 H, H-1a, H-1b), 3.37 (dd, 1 H, $J_{5a,5b}$ = 10.7 Hz, $J_{5a,4}$ = 4.0 Hz, H-5a), 3.28 (m, 1 H, H-4), 3.21 (dd, 1 H, $J_{5b,5a}$ = 10.6 Hz, $J_{5b,4}$ = 3.7 Hz, H-5b). ¹³C NMR (150 MHz, toluene-d₈, 85°C): δ 138.95, 138.87, 138.50 (Ar-Cq), 136.01 (Cq, NOCH₂Ph), 129.57-128.00 (Ar-C), 81.78 (C-3), 78.06 (C-4), 77.52 (OCH₂Ph), 77.12 (C-2), 74.52, 73.36, 72.72 (3 x OCH₂Ph), 8.25 (C-5). ESI-HRMS: *m/z* calcd for C₃₄H₃₆INO₅ [M+H⁺]⁺ 666.1711; found 666.1716.

Diethyl [2,3,4-tri-O-benzyl-1-(N-benzyloxy-N-formylamino)-1,5-dideoxy-D-ribitol] 5-phosphonate (19) A solution of compound 18 (222 mg, 0.33 mol) and $P(OEt)_3$ (300 μ L) was heated to 150 °C for 3 h. The reaction mixture was cooled to rt and the residual P(OEt)₃ was removed *in vacuo*. The residual oil was chromatographed on silica gel (2:1 hexane-EtOAc). The product containing fractions were pooled, concentrated, and purified by HPLC (2:1 hexane-EtOAc) to give 19 as colorless oil (100 mg, 44%); R_f 0.26 (2:1 hexane-EtOAc); [α]_D²⁰ +0.5 (c 0.9, CHCl₃); ¹H NMR (DMSO, 600 MHz, 90°C): δ 8.14 (bs, 1 H, NCHO), 7.0-7.24 (m, 20 H, Ar-H), 4.92-4.89 (m, 2 H, NOCH₂Ph), 4.74-4.56 (m, 6 H, 2 x OCH₂Ph), 4.14-4.08 (m, 1 H, H-4), 4.03-3.95 (m, 6 H, H-2, H-3, 2 x OCH₂CH₃), 3.88-3.80 (m, 2 H, H-1a, H-1b), 2.23 (ddd, 1 H, J_{5a,5b} = 15.8 Hz, J_{5a,4} = 4.4 Hz, J_{5a,P} = 18.4 Hz, H-5a), 2.15 (ddd, 1 H, J_{5b,5a} = 15.8 Hz, J_{5b,4} = 7.4 Hz, J_{5b,P} = 17.6 Hz, H-5b), 1.24-1.20 (m, 6 H, 2 x OCH₂CH₃); ¹³C NMR (DMSO, 150 MHz, 90 °C): δ 137.64, 137.52, 137.37 (Cq, 3 x C-Ar), 134.38 (Cq, NOCH₂Ph), 128.92, 128.88, 128.26, 127.67, 127.51, 127.39, 127.31, 127.26, 126.85, 126.83, 126.79, 126.64, 126.61, 126.54 (Ar-C), 79.88 (d, J_{C4,P} = 10.3 Hz, C-3), 75.37 (OCH₂Ph), 74.72 (C-2), 74.15 (C-4), 71.94, 71.10, 71.05 (OCH₂Ph), 60.25 (d, J_{P,OCH2CH3} = 6.0 Hz, POCH₂CH₃), 60.15 (d, J_{P,OCH2CH3} = 6.0 Hz, POCH₂CH₃), 46.82* (*det by HSQC, NCH₂), 27.02 (d, J_{P,C-5} = 139 Hz, C-5), 15.31, 15.27, 15.24 (2 x OCH₂CH₃); ³¹P NMR (DMSO, 242 MHz, 90 °C): δ 28.78. ESI-HRMS: *m/z* calcd for C₃₈H₄₆NO₈P [M+Na⁺]⁺ 698.2853; found 698.2582.

Sodium 1-(*N*-benzyloxy-*N*-formylamino)-1,5-dideoxy-D-ribitol] 5-phosphonate (20) Compound 19 (44.5 mg, 64 µmol) was dissolved in anhydrous DCM, pyridine (21 µL, 0.26 mmol) and TMSBr (34 µl, 0.26 mmol) were added under argon atmosphere, and the reaction was stirred at room temperature for 29 h. The reaction was monitored by HPLC MS (C-4, gradient 95:5 H₂O-MeCN 5 \rightarrow MeCN). Upon full conversion to the product the reaction mixture was concentrated, taken up in DCM (10 mL) and washed with citric acid (0.25 M). The aqueous phase was reextracted with DCM. The organic phase was dried (MgSO₄) and concentrated to give a white solid (36 mg) that was dissolved in 5:1 THF-H₂O containing 1% AcOH (2 mL). Pd(OH)₂-C (6.2 mg) was added and the reaction mixture was stirred under hydrogen atmosphere (1 bar) at rt for 7 h when fresh catalyst (6.2 mg) was added and stirring was

continued for further 15 h for completion. The reaction was monitored by HPLC-MS (Sequant ZIC HILIC 4.6 x 150 mm, Gradient 15:85 MeCN-H₂O \rightarrow 4:6 MeCN-H₂O, R_t of product 13.5 min). The reaction mixture was filtered through a syringe filter and the filter was successively washed with 1:1 MeOH-H₂O (15 mL). The filtrate was concentrated, and the residue was dissolved in H₂O, neutralized with NaHCO₃ (10 mg, 0.12 mmol) and concentrated. The residue was passed (1: 1 MeCN-H₂O) through a short plug of ZIC-HILIC silica gel (500 mg). Product containing fractions were concentrated, dissolved in 3:1 MeCN-H₂O (200 μ L) and purified by repeated ZIC-HILIC chromatography (Column: Sequant ZIC HILIC 10 x 250 mm, Merck, gradient 15:85 MeCN-H₂O \rightarrow 4:6 MeCN-H₂O, flow rate 3 mL, fraction size 2 mL). Fractions were concentrated and pooled according to ¹H NMR analysis (Fig.S2 Supporting Information) to give the target compound **20** (4 mg, 23 %) as white solid. $[\alpha]_D^{20}$ -6.7 (c 0.4, H₂O); ¹H NMR (600 MHz, D₂O): δ 8.33 (s, 0.2 H, NC(=O)H, minor rotamer), 7.89 (s, 0.8 H, NC(=O)H, major rotamer), 4.07-4.02 (m, 1 H, H-4), 4.02-3.98 (dt, 1 H, H-3), 3.75 (dd, 1 H, J_{1a,1b} 14.9, J_{1a,2} 2.2 Hz, H-1a), 3.67-3.61 (m, 2 H, H-2, H-1b), 1.83 (ddd, 1 H, J_{5a,5b} 14.9, J_{5a,4} 3.7, J_{5a,P} 17.8 Hz, H-5a), 1.61 (ddd, 1 H, J_{5b,5a} 15.0, *J*_{5b,4} 9.4, *J*_{5b,P} 15.1 Hz, H-5b).³¹P NMR (D₂O, 242 MHz): δ 20.53 (P(O)(OH)₂). ¹³C NMR (D₂O, 150 MHz): δ 161.08 (CHO), 76.72 (d, J 11.9 Hz, C-3), 69.19 (d, J 3.3 Hz, C-4), 68.44 (C-2, minor rotamer), 67.76 (C-2, major rotamer), 54.12 (C-1, major rotamer), 50.07 (C-1, minor rotamer), 31.53 (d, J 128.39 Hz, C-5); ESI-MS: *m*/*z* calcd. for C₆H₁₄NO₈P [M-H]⁻: 258.0384, found 258.0383.

2,3,4-Tri-O-benzyl-1-(*N***-benzyloxy-***N***-formylamino)-1-deoxy-5-oxo-D-ribitol (21)**. Dess-Martin reagent (200 mg, 0.46 mmol) was added to a solution of **15** (170 mg, 0.31 mmol) in DCM (4 mL). The reaction was stirred at rt for 1 h, then diluted with DCM and washed sequentially with satd aq NaHCO₃ and Na₂S₂O₃. The organic phase was washed with brine, dried and concentrated to give the crude product, which was stored under Ar atmosphere at -78 °C. Yield: 168 mg, (quant.) of aldehyde **21** as a colorless syrup; $[\alpha]_D^{20}$ +4.9 (*c* 0.6, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 9.47 (s, 1H, CHO), 8.28-7.26 (m, 21 H, H-Ar, H-formyl), 5.30 (s, 0.4 H), 5.06-4.40 (br. m, 8 H, CH₂Ar), 4.31- 3.20 (br. m, 5H, H-2, H-3, H-4, H-1a, H-1b); ¹³C NMR (150 MHz, 90 °C, toluene-d8): δ 190.0 (*C*H=O), 161.2 (NCH=O), 137.4–125.2 (24 x C-Ar), 82.1 (C-4), 81.2 (C-3), 76.2 (OCH₂Ph), 75.3 (C-2), 73.1, 73.1, 72.2 (3 x OCH₂Ph), 46.1 (C-1). ESI-MS: *m/z* calcd for C₃₄H₃₅NO₆ [M+Na]⁺: 576.2357; found 576.2365.

Dibenzyl [2,3,4-tri-O-benzyl-1-(N-benzyloxy-N-formylamino)-1-deoxy-D-ribo-5-(E)-hex-enitol)] 6phosphonate (23). The bisphosphonate reagent 22 (450 mg, 0.84 mmol) was dissolved in THF (2 mL) and cooled to -78 °C. Then BuLi (0.35 mL, 0.8 mmol) was added dropwise, and the reaction mixture was stirred at -78 °C for 30 min. Then aldehyde 21 (235 mg, 0.042 mmol) was added in one portion and the solution was stirred for 90 min before warming up to rt and continued stirring for 12 h. The reaction was quenched by adding aq NH₄Cl and the suspension was extracted with EtOAc. The organic layer was washed with brine, dried and concentrated. Purification of the crude by column chromatography (1:2 EtOAc-hexane) gave compound **23** as syrup. Yield: 225 mg, (76 %); $[\alpha]_D^{20}$ +3.3 (*c* 0.7, CHCl₃). ¹ H NMR (toluene-d₈, 70°): δ 8.04 (s, 1H, HC=O), 7.28–6.91 (m, 31 H, 30 Ar-H, H-5), 6.08 (m, 1 H, H-6), 4.95–4.89 (m, 4 H, 2 x OCH₂Ph), 4.60–4.50 (m, 4H, 2 x OCH₂Ph), 4.44 (AB, 2 H, OCH₂Ph), 4.29 (AB, 2 H, OCH₂Ph), 4.13 (bt, 1 H, H-4), 3.98 (bt, 1 H, H-2), 3.67 (m, 3 H, H-3, H-1a, H-1b), ¹³C NMR (toluene-d₈, 70°): δ 149.4 (C-formyl), 137.1 (6 x Ar-C), 129.0–124.3 (24 x Ar-C), 121.5 (C-5), 120.3 (C-6), 81.6 (C-3), 79.6 (C-4), 77.2 (C-2), 76.7 (OCH₂Ph), 73.6, 72.6, 71.9 (3 x OCH₂Ph), 66.9 (OCH₂Ph), 47.3 (C-1). ESI-MS: *m/z* calcd for C₅₆H₄₇N₂O₄P [M+H]⁺: 812.3609; found: 812.3623.

Sodium 1-N-formylamino-1-deoxy-D-*ribo***-hexitol 6-phosphonate (24)**. Compound **23** (35 mg, 0.04 mmol) was dissolved in 1 : 1: 2 THF-MeOH-H₂O (1 mL) and Pd(OH)₂ catalyst was added (8 mg). The reaction was stirred under hydrogen atmosphere for 72 h at rt. TLC (3:1 EtOAc-MeOH and 10:10:3 CHCl-MeOH-water) showed several spots. Every 24 h a new portion of catalyst (8 mg) was added. After 72 h most of the product had been formed according to LC-MS. The suspension was filtered, and the filtrate was concentrated. The residue was treated with 8.3 mg NaHCO₃ and filtered. The filtrate was concentrated and lyophilised. Purification on a HILIC column (2 x) gave 2.1 mg of **24** (16%) as syrup. $[\alpha]_D^{20}$ -34.7 (*c* 0.2, water); ¹H NMR (600 MHz, D₂O, ratio of rotamers 5.5:1): δ 8.42 (s, H-formyl minor rotamer), 7.97 (s, H-formyl major rotamer), 4.15 (ddd, 1 H, *J* = 2.5, *J* = 5.9 Hz, H-2), 3.79 (dd, 1H, *J* = 2.5, *J* = 14.9 Hz, H-1a), 3.76 (ddd, 1 H, *J* = 2.6, *J* = 6.3, *J* = 9.1 Hz, H-4), 3.72 (dd, 1 H, *J* = 9.2, *J* = 15.0 Hz, H-1b), 3.69 (dd, 1 H, H-3), 1.96 (dddd, 1 H, *J* = 2.8, *J* = 4.6, *J* = 11.9 Hz, H-5a), 1.83 (ddd, 1 H, *J* = 4.5, *J* = 11.9 Hz, H-6a), 1.69 (dddd, 1 H, *J* = 4.6, *J* = 9.1 Hz, H-5b), 1.61 (ddd, 1 H, *J* = 4.6, *J* = 11.5 Hz, H-6b); ¹³C NMR (150 MHz, D₂O): δ 160.9 (CHO), 76.0 (C-3), 72.8 (d, C-4), 68.2 (C-2), 54.0 (C-1), 27.2 (C-5), 25.3 (C-6). ³¹P NMR: δ 25.47.ESI-MS: *m/z* calcd for C₇H₁₆NO₈P [M]⁻ 272.0541; found: 272.0552.

Sodium 1-*N*-formylamino)-1-deoxy-D-*ribo*-5-(*E*)-hexenitol 6-phosphonate (25). Compound 23 (20 mg, 0.025 mmol) was dissolved in dry DCM (0.5 mL) and a 1 M sol. of SnCl₄ in DCM (0.17 mmol) was added dropwise. The solution was stirred at rt overnight. After this time another aliquot (0.1 mmol) of the reagent was added and stirring was continued for another 3 h. Then the mixture was poured into a solution of NaHCO₃ (61 mg, 0.73 mmol) in water (2 mL) and stirred for 10 min. DCM was evaporated and the residue containing insoluble Sn(IV) salts was filtered through cotton and a micro filter (2 x). The filtrate was concentrated and purified by ZIC-HILIC chromatography (column: Sequant ZIC HILIC 10 x 250 mm, Merck, 15:85 MeCN-H₂O \rightarrow 4:6 MeCN-H₂O, flow rate 3 mL, fraction size 2 mL). Purity of fractions was checked by ¹H NMR and product containing fractions were pooled and

Iyophilized. To remove residual sodium bicarbonate, the material was passed over DOWEX-50 cation exchange resin (H⁺-form) and the pH of the filtrate was adjusted to 7.0 by addition of 0.01 M NaOH followed by lyophilization to give **25** as syrup. Yield: 3.6 mg (50%). [α]_D²⁰+4.1 (*c* 0.4, D₂O); ¹H NMR (D₂O 600 MHz, ratio of rotamers 7.5:1): δ 8.49 (s, CH=O, minor rotamer), 7.81 (s, 1 H, CH=O, major rotamer), 6.25 (dt, 1 H, *J* = 6.5, *J* = 17.4 Hz, H-5), 6.08 (dt, 1 H, *J* = 16.1 Hz H-6), 4.36 (bt, 1 H, *J* = 6.1 Hz, H-4), 4.06 (ddd, 1 H, *J* = 2.6 Hz, *J* = 6.7 Hz, *J* = 8.3 Hz, H-2), 3.75-3.70 (m, 2 H, H-3, H-1a), 3.64 (ddd, 1 H, *J* = 13.6, *J* = 7.3 Hz, H-1b). ¹³C NMR (D₂O, 125 MHz): δ 156.07 (C=O), 137.09 (C-5), 131.68 (d, *J* = 167.0 Hz, C-6), 75.09 (C-3), 72.99 (d, *J* = 17.0 Hz, C-4), 67.46 (C-2), 54.34 (C-1). ³¹P NMR (242 MHz, D₂O): δ 9.62. ESI-MS: *m/z* calcd for C₇H₁₆NO₈P [M-H]⁻ 272.0541; found: 272.0552.

Dibenzyl [2,3,4-tri-O-benzyl-1-(N-benzyloxy-N-formylamino)-1-deoxy-5,6-dihydroxy-D-ribohexitol)]-6-phosphonate (26). Hexenitol 23 (50 mg, 0.06 mmol) was dissolved in 2:1 acetone-water (0.75 mL) and NMO (10 mg, 0.1 mmol) was added. Then potassium osmate (2.25 mg, 6.2 µmol) was added and the reaction was stirred overnight at rt. Since TLC (1:2 EtOAc-hexane) showed only minor conversion, 0.1 equivalents of potassium osmate and 1.6 eq. of NMO were added. This was repeated 3 times until an acceptable conversion was obtained (5 days). The reaction was quenched by adding satd aq Na₂SO₃ followed by extraction with EtOAc. The organic phase was washed with aq 4 % HCl, satd aq NaHCO₃, and brine. The organic layer was dried (MgSO₄), concentrated and the product was isolated by column chromatography (1:2 EtOAc-hexane) to give 26 as syrup. Yield: 41 mg (79 %). [α]_D²⁰ +11.9 (*c* 0.56, CHCl₃); ¹H NMR (600 MHz, toluene-d₈ 80°C): δ 8.09 (s, 1H, H-formyl), 7.30–6.96 (m, 30 H, Ar-H), 5.02 (d, 2 H, OCH₂Ph), 4.87 (d, 2 H, OCH₂Ph) 4.70–4.62 (m, 6 H, 3 x OCH₂Ph), 4.54 (s, 2H, OCH₂Ph), 4.44 (t, 1 H, J = 6.9 Hz, H-5), 4.41 (d, 1 H, J = 9.7 Hz, H-6), 4.19 (dd, 1 H, J = 4.2 Hz, J = 2.5 Hz, H-3), 4.08 (m, 2 H, H-3, H-2), 3.80 (bs, 2 H, H-1a, H-1b); ¹³C NMR (150 MHz): δ 138.5 –124.5 (30 x Ar-C), 80.4 (C-4), 79.4 (C-2), 76.8 (C-3), 76.4 (OCH₂Ph), 74.5 (OCH₂Ph), 73.5, 72.7 (2 x OCH₂Ph), 70.6 (C-5), 68.8 (OCH₂Ph), 68.5 (C-6), 67.5 (OCH₂Ph), 48.9 (C-1). ESI HR-MS: *m/z* cald for C₇H₁₆NO₈P [M+H]⁺: 846.3402; found: 846.3401.

Sodium 1-(N-benzyloxy-N-formylamino)-1-deoxy-5,6-dihydroxy-D-ribo-hexitol)] 6-phosphonate

(27). Compound 26 (24 mg, 0.03 mmol) was dissolved in 1:2 THF-water and Pd(OH)₂ catalyst (4 mg) was added. The suspension was stirred overnight and checked by TLC (CHCl₃-MeOH-water 10:10:3). After completeness, solid sodium bicarbonate (4 mg, 0.09 mmol) was added, and all volatiles were removed. Then the crude was purified by repeated ZIC-HILIC chromatography (column: Sequant ZIC HILIC 10 x 250 mm, Merck, Gradient MeCN-H₂O 85 to 40 %, flow rate 3 mL, fraction size 2 mL). Purity of fractions was checked by ¹H NMR and product containing fractions were pooled and lyophilized to give 27 (2.4 mg, 24%) as colorless solid. [α]_D²⁰ –20.5 (*c* 0.24, water). ¹H NMR (D₂O, 600 MHz, pD =

7.4): δ 8.29 (s, 0.2 H, NC(O)H, minor rotamer), 7.84 (s, 0.8 H, NC(O)H, major rotamer), 4.22 (m, 1H, *J* = 5 Hz, H-2), 4.01 (dt, 1H, H-5), 3.88 (m, 2H, H-3, H-4), 3.83 (dd, 1H, *J*_{6,5} = 2.5 Hz, *J*_{6,P} = 10 Hz, H-6), 3.72 (dd, 1H, *J*_{1a,2} = 2 Hz, *J*_{1b,1a} = 15 Hz, H-1a), 3.65 (dd, 1H, *J*_{1b,2} = 9 Hz, H-1b). ¹³C NMR (150 MHz): δ 159.7 (CH=O), 74.65 (C-3), 72.85 (C-5), 72.1 (C-4), 69.3 (C-6), 68.4 (C-2) and 54.4 (C-1). ESI HR-MS: *m/z* calcd for C₇H₁₆NO₁₀P [M-H]⁻: 304.0439; found 304.0440.

Dibenzyl [2,3,4-tri-O-benzyl-1-(N-benzyloxyamino)-1,6-dideoxy-D-allo/L-talo-hexitol)] 6phosphonate (29) and dibenzyl [2,3,4-tri-O-benzyl-1-(N-benzyloxy-N-formylamino)-1,6-dideoxy-Dallo/L-talo-hexitol)]-6-phosphonate (30) Dibenzyl methanephosphonate 28 (134 mg, 0.48 mmol) was dissolved in THF (1.5 mL) and cooled to -78 °C. Then a 2 M solution of BuLi in hexane (190 μ L) was added dropwise and the reaction was stirred at the same temperature for 30 min. Subsequently aldehyde 21 (180 mg, 0.32 mmol) was added dropwise and the solution was stirred at -78 °C. The reaction was quenched after 90 min by adding aq NH₄Cl. The suspension was extracted with EtOAc, washed with brine. dried (MgSO₄), concentrated. The residue was purified by column chromatography (1:2 EtOAc-hexane) to give first 50 mg (28 %) of recovered aldehyde 21 followed by the deformylated product **30** (55 mg, 22 %) and **29** (25 mg, 10%) as syrup; $[\alpha]_{D}^{20}$ +8.6 (*c* 0.45, CHCl₃); ¹H NMR (toluene-d₈ , 90 °C): δ 8.06 (s, 1 H, H-formyl), 7.30–6.94 (m, 30 H, Ar-H), 4.95–4.44 (13 H, 6 x OCH₂Ph, H-2), 4.20 (bt, 1 H, H-5), 3.93 (bt, 1 H, H-4), 3.83 (bt, 1 H, H-3), 3.77 (m, 2 H, H-6a, H-6b), 2.21 (ddd, 1 H, J = 15.0 and 2.5 Hz, H-1a), 2.12 (ddd, 1 H, H-1b). ¹³ C NMR (150 MHz, toluene-d₈ 90 °C): δ 161.0 (C=O), 137.4–124.6 (36 Ar-C), 82.5 (d, C-P coupling, J = 14 Hz, C-4), 79.8 (C-3), 76.8 (C-2), 76.7 (OCH₂Ph), 74.2, 73.5, 72.7 (3 x OCH₂Ph), 67.5 (d, C-P coupling, J = 6 Hz, C-5), 67.0, 67.0 (2 x OCH₂Ph), 48.3 (C-1), 29.8 (d, J_{C-P} = 138 Hz, C-6).³¹ P NMR (242 MHz, toluene-d₈ 90 °C) : δ 25.2 (s). ESI HR-MS: *m*/*z* calcd for C₄₉H₅₂NO₉P [M+H]⁺: 830.3452; found 830.3448. ¹H NMR for **30** (toluene-d₈ 90 °C, major isomer): δ 7.27–6.95 (m, 30 H, Ar-H), 4.92–4.80 (m, 4 H, 2 x OCH₂Ph), 4.68–4.48 (m, 9 H, 4 x OCH₂Ph, H-5), 4.13 (ddd, 1 H, H-2), 3.94 (dd, 1 H, J = 4.6 Hz, H-3), 3.82 (dd, 1 H, J = 4.9 Hz, H-4), 3.36 (dd, 1 H, J = 4 Hz, J = 14 Hz, H-1a), 3.21 (dd, 1 H, J = 7 Hz, H-1b), 2.26

(dd, 1 H, *J* = 3, *J* = 15 Hz, H-6a), 2.14 (ddd, 1 H, *J* = 7 Hz, H-6b). ¹³C NMR (toluene-d₈ 90 °C): δ 137.4– 124.4 (36 Ar-C), 82.4 (d, *J*_{C-P} = 15 Hz, C-4), 80.2 (C-3), 77.1 (C-2), 75.7, 74.0, 73.5, 72.4 (4 x OCH₂Ph), 67.6 (d, *J*_{C-P} = 5 Hz, C-5), 67.0, 66.9 (2 x OCH₂Ph), 52.8 (C-1), 29.9 (d, *J*_{C-P} = 140 Hz, C-6). ³¹ P NMR (242 MHz, toluene-d₈ 90 °C): δ 32.5. ESI HR-MS: *m/z* calcd for C₄₈H₅₂NO₈P [M+H]⁺: 802.3503; found 802.3525.

Formylation reaction of 30. Carbonyldiimidazole (62 mg, 0.38 mmol) was dissolved in THF (1 mL) and cooled to 0 °C. Then formic acid (15 μ L, 0.38 mmol) was added and the mixture was stirred at 0° C for 30 min under Ar. Meanwhile a solution of **30** (60 mg, 0.075 mmol) in THF (0.5 mL) was

prepared. The solution of CDI and formic acid was then added, and the mixture was stirred at rt for 36 h. Then all volatiles were evaporated, the residue was coevaporated with toluene and purified by column chromatography (1:2 EtOAc-hexane) to give unreacted **30** (15 mg, 24%) followed by **29** (45 mg, 73%).

Sodium [1-(*N*-hydroxy-*N*-formylamino)-1,6-dideoxy-D-*allo*/L-*talo*-hexitol}]-6-phosphonate (31) Compound 29 (20 mg, 0.024 mmol) was dissolved in 1:2 THF-water and Pd(OH)₂ catalyst (4 mg) was added. The suspension was stirred overnight and checked by TLC (10:10:3 CHCl₃-MeOH-H₂O). The catalyst was filtered off and the filtrate was washed with 1:1 MeOH-H₂O and purified twice by HILIC separation (column: Sequant ZIC HILIC 10 x 250 mm, Merck, gradient 15:85 MeCN-H₂O \rightarrow 4:6 MeCN-H₂O, flow rate 3 mL, fraction size 2 mL). Purity of fractions was checked by ¹H NMR and product containing fractions were pooled and lyophilized to give **31** (0.65 mg, 10%) as colorless solid; $[\alpha]_D^{20}$ +17.4 (*c* 0.2, water); ¹H NMR (D₂O, 600 MHz, ratio of rotamers 4.1:1): δ 8.31 (s, CHO, minor form), 7.86 (s, 1 H, CHO major isomer), 4.27 (ddd, 1 H, *J* = 8.7, *J* = 4.7 Hz, H-2), 4.19 (dt, 1 H, *J* = 3.6, *J* = 3.6, *J* = 9.8 Hz, H-5), 3.80 (dd, 1H, *J* = 8.3, *J* = 4.0 Hz, H-3), 3.76 (dd, 1H, *J* = 8.2, *J* = 4.4 Hz, H-4), 3.72-3.70 (m, 2H, H-1a, H-1b),1.11 (ddd, 1 H, *J* = 3.5, *J* = 14.6 Hz, H-6a), 1.65 (ddd, 1 H, *J* = 9.3, *J* = 14.9 Hz, H-6b); ¹³C NMR (D₂O, 150 MHz): δ 75.13 (d, *J* = 9.3 Hz, C-4), 73.62 (C-3), 69.82 (C-5), 68.53 (C-2), 54.06 (C-1), 30.79 (d, *J* = 104.1 Hz, C-6); ³¹P NMR (242 MHz, D₂O): δ 29.4. ESI HR-MS: *m/z* calculated for C₇H₁₆NO₉P [M-H]: 288.0490; found 288.0495.

Phenyl 2,3,4,6-tetra-*O***-acetyl-1-thio-D-altropyranoside (87)**. Acetic anhydride (50 mL) and a catalytic amount of DMAP were added to a solution of D-altrose **85** (2 g, 11 mmol) in dry pyridine (60 mL) at ice-bath temperature. The mixture was then stirred at rt for 30 min, whereupon TLC (3:2 hexane-EtOAc) showed complete conversion. EtOAc (150 mL) was added and the mixture was washed with 1 M aq HCl (200 mL). The aqueous phase was extracted with EtOAc (3 × 50 mL). The combined organic layers were sequentially washed with water and satd aq NaHCO₃, then dried (MgSO₄), filtered and concentrated. Diethyl ether was added to the residue and the solution was filtered through a short Florisil pad. The eluate was concentrated to afford penta-*O*-acetyl-D-altrose **86** (ratio α/β/furanose: 1:0.07:0.08) as a pale yellow oil (4.3 g, ~quant.). Thiophenol (16.5 mmol, 1.7 mL) was then added to a solution of **86** (4.3 g, 11 mmol) in anhydrous dichloromethane (60 mL) under argon. Then BF₃·Et₂O (55 mmol, 6.8 mL) was added dropwise at 0 °C. The reaction mixture was allowed to warm to rt and was stirred overnight. Additional thiophenol (8.8 mmol, 0.9 mL) and BF₃ ·Et₂O (0.04 mmol, 5.1 mL) were added and stirring was continued for 4 h. The reaction mixture was diluted with CH₂Cl₂, cooled to 0 °C, washed with satd aq NaHCO₃ followed by re-extraction of the aqueous phase with CH₂Cl₂ (3 x). The combined organic phases were then washed with water, dried

(MgSO₄) and concentrated. The crude was purified by column chromatography on silica gel (7:3 → 3:2 hexane-EtOAc) to afford **87** as a colorless oil (4.0 g, 82%, ratio of α-pyranose/β-pyranose/furanose = 1 : 0.1 : 0.15), along with recovered starting material **86** (244 mg, 6%).¹H NMR (CDCl₃, 300 MHz, α-anomer): δ 7.57-7.45 (m, 2 H, Ph), 7.35-7.23 (m, 3 H, Ph), 5.41 (br d, 1 H, H-1), 5.33 (td, 1 H, *J*_{1,3} = 0.9, *J*_{2,3} = *J*_{3,4} = 3.4 Hz, H-3), 5.24-5.17 (m, 2 H, H-2, H-4), 4.81 (ddd, 1 H, *J*_{4,5} = 10.2, *J*_{5,6a} = 5.7, *J*_{5,6b} = 2.2 Hz, H-5), 4.36 (dd, part A of ABX system, 1 H, *J*_{6a,6b} = 12.2 Hz, H-6a), 4.20 (dd, part B of ABX system, 1 H, H-6b), 2.23, 2.14, 2.08 and 2.05 (4 x s, each 3 H, 4 x Ac). ¹³C NMR (CDCl₃, 75 MHz, α-anomer): δ 170.7, 169.6, 169.4, 169.4 (C=O), 135.2 (Cq, Ph), 131.9, 129.2, 127.9, (*C*H, Ph), 86.1 (C-1), 71.4 (C-2), 66.9 (C-3), 65.6, 65.3 (C-4, C-5), 62.8 (C-6), 20.9, 20.8, 20.7 and 20.6 (*C*H₃CO).

Phenyl 1-thio-D-altropyranoside (88). A solution of 87 (1.55 g, 3.52 mmol) in 8:1:1 MeOH-H₂O-Et₃N was stirred at rt overnight. The solution was concentrated and the residue was dissolved in water and lyophilized to give 88 as colorless amorphous solid. Yield: 873 mg (91%); $[\alpha]_D^{23}$ +62 (*c* 1.0, MeOH); ¹H NMR (MeOD, 300 MHz): δ 7.58-7.50 (m, 2 H, Ph), 7.33-7.20 (m, 3 H, Ph), 5.30 (brd, 1 H, *J*_{1,2} = 1.6 Hz, H-1), 4.33 (ddd, 1 H, *J*_{5,6a} = 2.9, *J*_{5,6b} = 5.1 Hz, H-5), 4.08 (dd, 1 H, *J*_{1,2} = 1.6, *J*_{2,3} = 3.6 Hz, H-2), 3.98-3.90 (m, 2 H, H-3, H-4), 3.88 (dd, 1 H, *J*_{5,6a} = 2.9, *J*_{6a,6b} = 11.9 Hz, H-6a), 3.81 (dd, 1 H, *J*_{5,6b} = 5.2 Hz, H-6b).¹³C NMR (MeOD, 75 MHz): δ 139.0 (Cq, Ph), 132.2, 129.8, 127.8, (*C*H, Ph), 89.7 (C-1), 74.4 (C-2), 71.7, 71.4 (C-3, C-5), 66.0 (C-4), 63.0 (C-6). HRMS (ESI⁺): *m/z* calcd for C₁₂H₁₆O₅S [M + NH₄]⁺ 290.1057; found: 290.1056.

Phenyl 6-*O*-triisopropylsilyl-1-thio-α-D-altropyranoside (89). DMAP (cat. amount) was added to a solution of **88** (823 mg, 3.03 mmol, ratio: α-pyranoside/furanoside = 1 : 0.11) in dry pyridine (30 mL) under argon and cooled to 0 °C. TIPSCI (2 equiv., 6.05 mmol, 1.16 g, 1.3 mL) was then added dropwise, the mixture was warmed to rt and stirred for 3 h. Second portions of TIPSCI (1 equiv. 0.7 mL) and DMAP (cat. amount) were added and the mixture was stirred overnight, followed by a third addition of TIPSCI (1 equiv., 0.7 mL) and DMAP. The mixture was stirred for 7 h, when TLC (1:9 EtOAc-MeOH,) showed virtually complete conversion. EtOAc was added and the organic phase was washed with water. The aqueous phase was extracted with EtOAc (3 ×). The combined organic phases were washed with water, dried (MgSO₄) and concentrated. The residue was subjected to chromatography on silica-gel (2:3 EtOAc-hexane) to give first the furanoside (60 mg) followed by a 10 : 0.7 mixture of α-pyranoside/furanoside (963 mg) and pure **89** (199 mg) as a colorless oil; [α]_D²²+154 (*c* 1.2, CHCl₃). ¹H NMR (CDCl₃, 300 MHz): δ 7.52-7.44 (m, 2 H, Ph), 7.33-7.20 (m, 3 H, Ph), 5.31 (d, 1 H, *J*_{1,2} = 2.4 Hz, H-1), 4.37 (ddd, 1 H, H-5), 4.18 (ddd, 1 H, H-2), 4.11-3.98 (m, 3 H, *J*_{5,6a} = 4.8, *J*_{6a,6b} = 10.0 Hz, H-3, H-4, H-6a), 3.93 (dd, part B of ABX system, 1 H, *J*_{5,6b} = 7.7 Hz, H-6b), 3.71 (d, 1 H, *J* = 1.6 Hz, OH), 2.85 (d, 1 H, *J* = 1.6 Hz, OH), 2.18 (d, 1 H, *J* = 6.3 Hz, OH-2), 1.14-1.05 (m, 21 H, 3 × *i*-Pr, TIPS). ¹³C NMR (CDCl₃,

75 MHz): δ 136.3 (Cq, Ph), 131.2, 129.1, 127.3, (*C*H, Ph), 87.9 (C-1), 71.9 (C-3), 70.1 (C-2), 69.5 (C-4), 68.4 (C-5), 66.3 (C-6), 18.0 (Me, *i*-Pr, TIPS), 11.9 (*CH*, i-Pr, TIPS). HRMS (ESI⁺): *m/z* calcd for C₂₁H₃₆O₅SSi [M + Na]⁺ 451.1945; found: 451.1958.

Phenyl 2,3,4-tri-O-benzyl-6-O-triisopropylsilyl-1-thio- α -D-altropyranoside (90). NaH (60%, 6 equiv., 13.5 mmol, 540 mg) was added to a solution of 89 (963 mg, 2.25 mmol, ratio of α pyranoside/furanoside = 10 : 0.7) in dry DMF (60 mL) under argon at 0 °C. After a few minutes, benzyl bromide (6 equiv., 13.5 mmol, 1.6 mL) was added dropwise. The mixture was stirred at rt for 2 h, whereupon TLC (1:2 EtOAc-hexane) showed complete conversion. The mixture was cooled to 0 °C and water was added. The phases were separated, and the aqueous phase was extracted with diethyl ether (3 ×). The combined organic phases were washed with water, dried (MgSO₄) and concentrated. The residue was purified by chromatography on silica-gel (1:23 EtOAc-hexane) to give pure 90 as a colorless oil (131 mg) followed by **90** containing residual furanoside (1.369 g, ratio α pyranoside/furanoside = 10:0.8 mixture); total yield = 1.5 g, (96%); $[\alpha]_{D}^{20}$ +70 (*c* 1.0, CHCl₃). ¹H NMR (CDCl₃, 600 MHz): δ 7.53-7.50 (m, 2 H, Ph), 7.39-7.17 (m, 18 H, Ph), 5.47 (br.s, 1 H, H-1), 4.71 (d, part A of AB system, 1 H, J = 12.4 Hz, OCH₂), 4.62 (d, part B of AB system, 1 H, OCH₂), 4.57 (d, part A of AB system, 1 H, J = 12.4 Hz, OCH₂), 4.54-4.47 (m, 3 H, OCH₂, H-5), 4.39 (d, part B of AB system, 1 H, OCH₂), 4.03-3.95 (m, 3 H, J_{5,6a} = 4.0, J_{6a,6b} = 11.0 Hz, H-6a, H-6b, H-4), 3.96 (dd, 1 H, J_{1,2} = 1.5, J_{2,3} = 4.1 Hz, H-2), 3.83 (t, 1 H, J_{2,3} = J_{3,4} = 3.4 Hz, H-3), 1.14-1.04 (m, 21 H, 3 × *i*-Pr, TIPS). ¹³C NMR (CDCl₃, 150 MHz) δ: 138,7, 138.4, 138.3, 138.1 (4 × Cq, Ph), 130.5, 128.8, 128.6, 128.5, 128.4, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 126.6 (CH, Ph), 85.6 (C-1), 78.1 (C-2), 73.7 (C-3), 72.9 (C-4), 72.6, 72.3, 72.0, 72.0, 70.2 (4 × CH₂Ph, C-5), 63.2 (C-6), 18.2, 18.1 (Me, *i*-Pr, TIPS) 12.2 (CH, *i*-Pr, TIPS). HRMS (ESI⁺): *m/z* calcd for C₄₂H₅₄O₅SSi [M + NH₄]⁺ 716.3799; found: 716.3803.

Phenyl 2,3,4-tri-*O*-benzyl-1-thio-α-D-altropyranoside (91). A 1 M solution of TBAF in THF (0.2 mL, 0.2 mmol) was added to a solution of 90 (34 mg, 0.05 mmol) in THF (1.3 mL) under argon and the solution was stirred overnight at rt. The solution was diluted with diethyl ether and washed with satd aq NH₄Cl. The aqueous layer was extracted with Et₂O (3 ×), the combined organic layers were dried (MgSO₄) and concentrated. The residue was purified by chromatography on silica gel (1:5 →1:2.5, hexane-EtOAc) to give 91 (17 mg, 64%) as a colorless oil. This protocol was also applied to a 10:0.8 mixture of α-pyranoside/furanoside (1.369 g, 1.95 mmol) which afforded 628 mg (60%) of the deprotected phenyl 1-thioaltroside. [α]_D²⁵ +103 (*c* 0.6, CHCl₃). ¹H NMR (toluene-d₈, 600 MHz): δ 7.56-7.53 (m, 2 H, Ph), 7.39-7.36 (m, 2 H, Ph), 7.23-7-20 (m, 2 H, Ph), 7.17-6.97 (m, 14 H, Ph), 5.61 (br.s, 1 H, H-1), 4.79 (dt, 1 H, J_{5,6a} = J_{5,6b} = 3.3, J_{4,5} = 9.8 Hz, H-5), 4.65 (d, part A of AB system, 1 H, J = 12.4 Hz, OCH₂), 4.56 (d, part B of AB system, 1 H, OCH₂), 4.34 (s, 2 H, OCH₂), 4.26 (d, part A of AB system, 1 H,

 $J = 12.1 \text{ Hz}, \text{ OCH}_2, 4.09-4.05 \text{ (m, 2 H, } J_{3,4} = 2.9 \text{ Hz}, \text{ H-4}, \text{ OCH}_2), 4.00 \text{ (br.d, 1 H, } J_{2,3} = 3.7 \text{ Hz}, \text{ H-2}), 3.88-3.83 \text{ (m, 2 H, H-3, H-6)}, 3.81 \text{ (dd, part B of ABX system, 1 H, } J_{5,6a} = 4.0, J_{6a,6b} = 11.7 \text{ Hz}, \text{ H-6b}). ^{13}\text{C NMR} \text{ (toluene-d}_8, 150 \text{ MHz}): } \delta 86.5 \text{ (C-1)}, 78.8 \text{ (C-2)}, 74.0 \text{ (C-3)}, 73.6, 73.1, 72.1, 71.8 \text{ (C-4, 3 × CH}_2\text{Ph}), 69.5 \text{ (C-5)}, 62.6 \text{ (C-6)}. \text{ HRMS (ESI^+): } m/z \text{ calcd for } C_{33}\text{H}_{34}\text{O}_5\text{S} \text{ [}M + \text{NH}_4\text{]}^+ 560.2465; \text{ found: } 560.2467; \text{ calcd for } [M + \text{Na}]^+ 565.2019; \text{ found: } 565.2024.$

Phenyl 2,3,4-tri-O-benzyl-6-O-(bisphenoxy-phosphoryl)-1-thio- α -D-altropyranoside (92). A solution of 91 (245 mg, 0.45 mmol), triethylamine (0.1 mL, 0.72 mmol) and DMAP (cat. amount) in dry dichloromethane (25 mL) was cooled to 0 °C under argon, followed by addition of diphenyl phosphorochloridate (0.1 mL, 0.48 mmol). The reaction mixture was allowed to warm up to rt and was stirred overnight. The mixture was diluted with dichloromethane (20 mL) and then washed with satd aq NaHCO₃. The organic phase was dried (MgSO₄) and concentrated. The residue was purified by column chromatography on silica gel (1:4 hexane-EtOAc, containing 1% of NEt₃) to give 92 (338 mg, 97%) as a colorless oil; [α]_D²⁴+20.7 (*c* 0.5, CHCl₃). ¹H NMR (CDCl₃, 300 MHz): δ 7.50-7.42 (m, 2 H, Ph), 7.40-7.15 (m, 28 H, Ph), 5.46 (br.s, 1 H, H-1), 4.74 (ddd, 1 H, J_{5,6} = 3.0 and 5.8 Hz, H-5), 4.68 (d, part A of AB system, 1 H, J = 12.3 Hz, OCH₂), 4.61-4.49 (m, 4 H, H-6a, H-6b, OCH₂), 4.38 (d, part B of AB system, 1 H, J = 12.2 Hz, OCH₂), 4.30 (br s, 1 H, OCH₂), 3.98 (dd, 1 H, J_{2.1} = 0.9, J_{2.3} = 3.5 Hz, H-2), 3.90 (dd, 1 H, J_{3,4} = 2.9, J_{4,5} = 10.0 Hz, H-4) and 3.82 (t, 1 H, H-3). ¹³C NMR (CDCl₃, 75 MHz): δ 150.7, 137.9, 137.6 (Cq, Ph), 130.7, 129.8, 129.7, 129.7, 129.0, 128.6, 128.5, 128.5, 128.1, 128.0, 127.9, 127.8, 126.9, 125.3, 120.4, 120.3, 120.3, 120.3 (CH, Ph), 85.9 (C-1), 77.4 (C-2), 72.6, 72.4, 72.3, 72.1 (2 x CH₂Ph, C-3, C-4), 71.5 (CH₂Ph), 68.5 (C-6, J_{C-6,P} = 5.6 Hz), 67.3 (C-5, J_{C-5,P} = 8.2 Hz). ³¹P NMR (CDCl₃, 121.5 MHz) δ: -11.97

2,3,4-Tri-O-benzyl-6-O-(bisphenoxy-phosphoryl)-D-altro-pyranose (93) *N*-bromosuccinimide (NBS, 5 equiv, 2 mmol, 380 mg) was added to a solution of **92** (331 mg, 0.43 mmol) in 5:1 acetonewater (36 mL) at -11 °C in the dark. After 10 min at -11 to 0 °C, whereupon complete conversion was observed by TLC, the reaction mixture was diluted with dichloromethane (40 mL). The organic phase was washed with satd aq NaHCO₃ (10 mL) and satd aq Na₂S₂O₃ soln. (10 mL). The combined aqueous phases were extracted with dichloromethane (3 ×) and the combined organic phase was dried (MgSO₄) and concentrated. The residue was purified by column chromatography on silica gel (1:2 EtOAc-hexane) to yield **93** as colorless oil (284 mg, 98%).¹H NMR (CDCl₃, 300 MHz, α/β ratio = 1.8:1): δ 7.39-7.06 (m, Ph), 5.06 (br s, H-1 α), 5.04 (d, *J* _{1,2}(β) = 1.8 Hz, H-1 β), 4.71 (d, part A of AB system, *J* = 11.7 Hz, OCH₂), 4.65-4.23 (m, H-6a α , H-6a β , H-5 α , OCH₂- α , OCH₂- β), 4.11 (ddd, H-5 β), 3.95-3.87 (m, *J*_{3,4}(α) = 2.8, *J*_{4,5}(α) = 10.7 Hz, H-3 α , H-4 α ,), 3.78-3.71 (m, *J*_{3,4}(β) = 2.8, *J*_{4,5}(β) = 10.7 Hz, H-3 β , H-4 β), 3.60 (br d, 1 H, *J*_{2,3}(α) = 3.6 Hz, H-2 α), 3.48 (dd, 1 H, *J*_{2,3}(β) = 3.5 Hz, H-2 β). ¹³C NMR (CDCl₃, 75 MHz): δ 150.8, 150.7, 138.0, 137.8, 137.7, 137.6, 137.0 (Cq, Ph), 129.8, 129.8, 128.8, 128.8, 128.7, 128.6, 128.6, 128.5, 128.4, 128.2, 128.2, 128.1, 128.1, 128.1, 128.0, 125.3, 125.3, 120.5, 120.4, 120.4, 120.3 (CH, Ph), 93.1 (C-1 α), 92.0 (C-1 β), 76.9 (C-2 β), 74.9 (C-2 α), 74.3, 74.3, 73.6, 73.3, 72.3, 72.2, 72.2, 72.0, 71.9 (CH₂Ph, C-3, C-4, α, β), 71.3 (C-5 β, $J_{C-5,P}$ = 8.2 Hz), 68.5 (C-6 β, $J_{C-6,P}$ = 6.2 Hz), 68.4 (C-6 α, $J_{C-6,P}$ = 5.9 Hz), 66.1 (C-5 α, $J_{C-5,P}$ = 8.2 Hz). ³¹P NMR (CDCl₃, 121.5 MHz) δ: -11.87, -12.02. HRMS (ESI⁺): m/z calcd for C₃₉H₃₉O₉P [M + NH₄]⁺ 700.267; found: 700.2668.

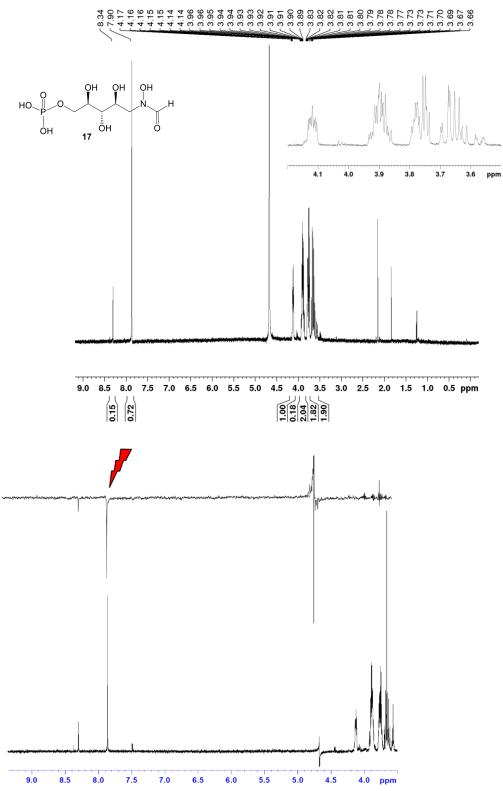
2,3,4-Tri-*O***-benzyl-6-***O***-(b**isphenoxy-phosphoryl)-**D**-altrono-1.5-lactone (**94**) A suspension of **93** (37 mg, 54 μmol) and activated 4 Å molecular sieves (150 mg) in anhydrous dichloromethane (30 mL) was stirred at room temp for 20 min. Then PCC (148 mg, 0.69 mmol) was added, and the suspension was stirred for 1 h. Within 4 h, 4 additional portions of PCC (4 x 40 mg) were added to drive the reaction to completion. EtOAc (20 mL) was added, followed by filtration over a Florisil pad. The eluate was concentrated and purified by silica gel chromatography (3:1 hexane-EtOAc, containing 0.1% of TEA) to give lactone **94** (24 mg, 65%) as a colorless oil, $[\alpha]_{p}^{22}$ -4.0 (*c* 0.5, CHCl₃). ¹H NMR (CDCl₃, 300 MHz): δ 7.38-7.10 (m, 23 H, Ph), 6.95-6.87 (m, 2 H, Ph), 4.81-4.70 (m, 2 H, H-5, H-a, Bn, *J* = 11.8 Hz. H-5, OCH₂), 4.57-4.48 (m, H-6a, H-6b, OCH₂), 4.35 (2 d, AB system, 2 H, CH₂Ph, *J* = 11.4 Hz, OCH₂), 4.13 (dd, 1 H, *J*_{3,4} = 2.4, *J*_{4,5} = 8.9 Hz, H-4), 4.06 (d, 1 H, *J*_{2,3} = 4.2 Hz, H-2), 3.87 (dd, 1 H, H-3). ¹³C NMR (CDCl₃, 75 MHz): δ 167.2 (CO), 150.6, 150.4, 137.4, 137.2 (Cq, Ph), 130.0, 130.0, 130.0, 129.8, 128.3, 128.3, 128.3, 128.0, 125.7, 125.7, 125.6, 120.8, 120.4, 120.3, 120.3, 120.3 (CH, Ph), 76.4 (C-5, *J*_{C-5,P} = 8.4 Hz), 74.9 (C-2), 73.3 (C-3), 73.2 (CH₂Ph), 72.9, 72.6 (2 x CH₂Ph), 70.6 (C-2), 67.0 (C-6, *J*_{C-5,P} = 5.4 Hz). ³¹P NMR (CDCl₃, 121.5 MHz): δ -12.18. HRMS (ESI⁺): calcd for C₃₉H₃₇O₉P [*M* + H]⁺ 681.2248; found 681.2266; calcd for [*M* + NH₄]⁺: 698.2513; found: 698.2530.

D-Altrono-1.4-lactone 6-*O***-dihydrogenphosphate (95)**. A catalytic amount of 10% Pd/C was added to a solution of aldonolactone **94** (93 mg, 0.14 mmol) in anhydrous THF (6 mL). The suspension was placed under H₂ atmosphere and stirred overnight at rt, whereupon TLC showed complete conversion (EtOAc). The catalyst was filtered off, the eluate was concentrated under vacuum, the residue was dissolved in water and lyophilized. The resulting crude compound (52 mg) was dissolved in THF (4 mL), and PtO₂ (cat. amount) was added. The suspension was stirred under hydrogen atmosphere overnight, whereupon TLC (10:9:1 EtOAc-MeOH-H₂O) showed complete conversion. The catalyst was filtered off and the eluate was concentrated in vacuum. The residue was dissolved in water and extracted with diethyl ether. The aq phase was lyophilized to give a white hygroscopic solid which was subjected to gel filtration using a PD-10 Sephadex G 25 column (water as eluent). The eluate was lyophilized to give **95** as amorphous solid (32.6 mg, 90%) containing the free acid in a 3:1 ratio; data for **95**: ¹H NMR (D₂O, 600 MHz): δ 4.58 (d, 1 H, J_{2,3} = 8.6 Hz, H-2), 4.41 (t, 1 H, H-3), 4.36

S18

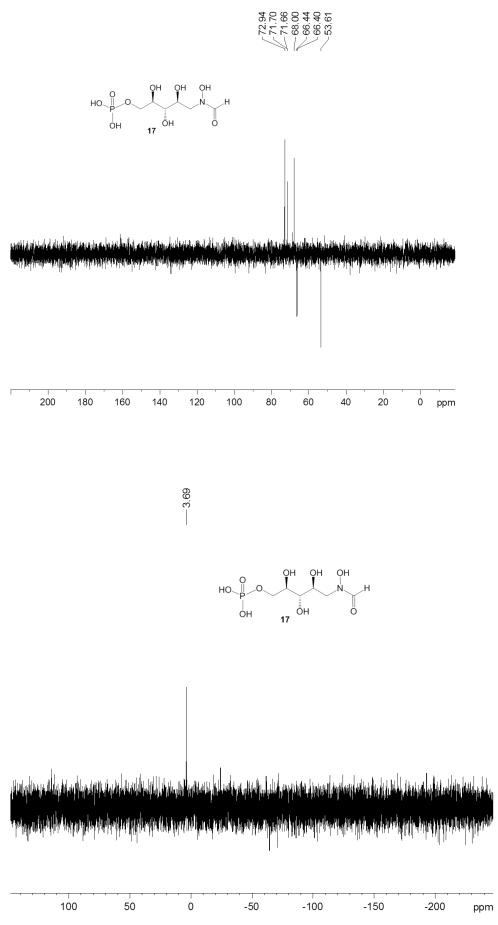
(dd, 1 H, , $J_{3,4}$ = 8.1, $J_{2,3}$ = 3.9 Hz, H-4), 4.20 (ddd, 1 H, H-5), 4.08-4.01 (m, H-6a), 3.99-3.94 (ddd, H-6 b).¹³C NMR (D₂O, 150 MHz) δ : 176.7 (C-1), 80.7 (C-4), 77.8 (C-2), 73.6 (C-3), 69.9 (d, C-5, $J_{C-5,P}$ = 7.7 Hz), 66.2 (d, C-6, $J_{C-6,P}$ = 4.9 Hz).³¹P NMR (CDCl₃, 242.9 MHz) δ : 0.27.LCMS: [M + H]⁺ 259; [M + Na]⁺ 281; [M - H]⁺ 257.

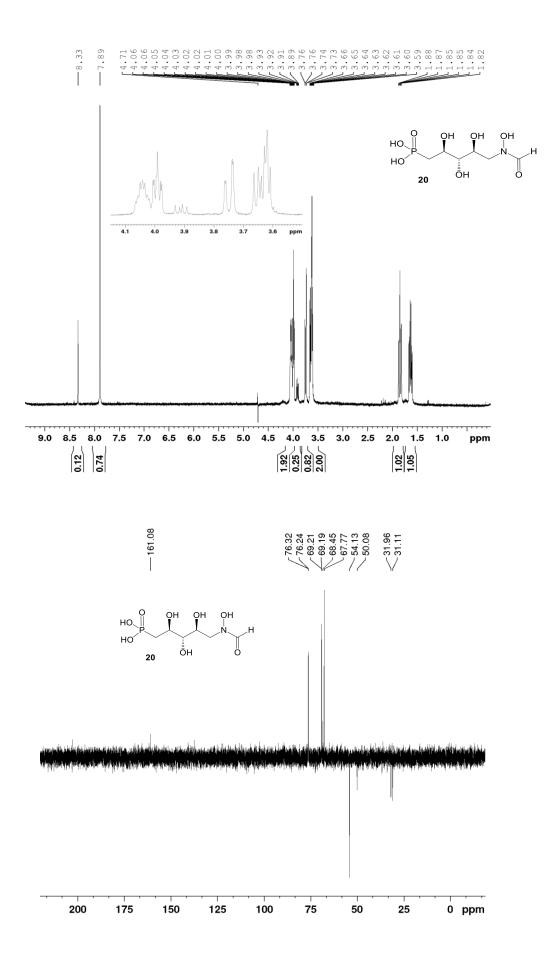
D-Altronohydroxamic acid 6-(dihydrogen phosphate) (96). A 50% aqueous hydroxylamine solution (0.4 mmol, 24 μL) was added to a solution of D-altrono-γ-lactone 6-(dihydrogen phosphate) **95** (10 mg, 39 μmol) in water (1.5 mL). The reaction mixture was stirred at rt for 4 h. The solution was concentrated under vacuum and then lyophilized. The residue was subjected to gel filtration chromatography (Bio-gel P-2) to afford **96** (10 mg, 88%). During prolonged NMR measurements, the compound slowly degraded (inter alia 6→5 phosphate migration). ¹H NMR (D₂O, 600 MHz) δ: 4.50 (d, 1 H, J_{2,3} = 1.7 Hz, H-2), 4.06-3.91 (m, 4 H, H-3, H-5, H-6a, H-6b), 3.79 (dd, 1 H, J_{4,5} = 4.7, J_{4,3} = 8.8 Hz, H-4).¹³C NMR (D₂O, 150 MHz) δ: 172.18 (C-1), 72.90 (J_{C,P} = 6.3 Hz, C-5), 72.82 (C-3), 71.54 (C-2), 71.29 (C-4), 65.54 (J_{C,P} = 4.2 Hz, C-6); ³¹P NMR (D₂O, 242.9 MHz) δ: 2.67. LCMS: calcd for C₆H₁₄NO₁₀P [*M* + H]⁺: 292.0; found: 292; [*M* - H]⁻: 290.

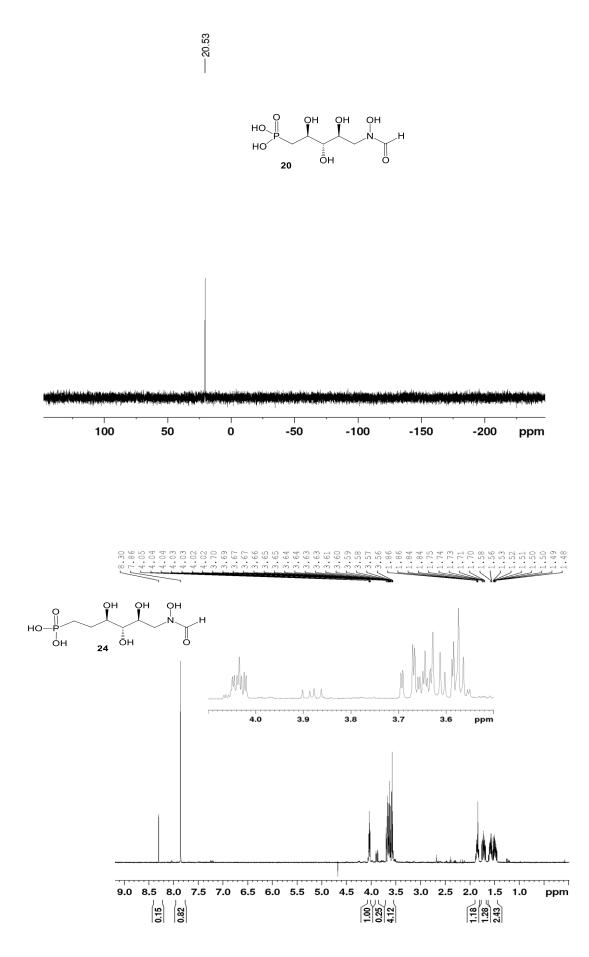


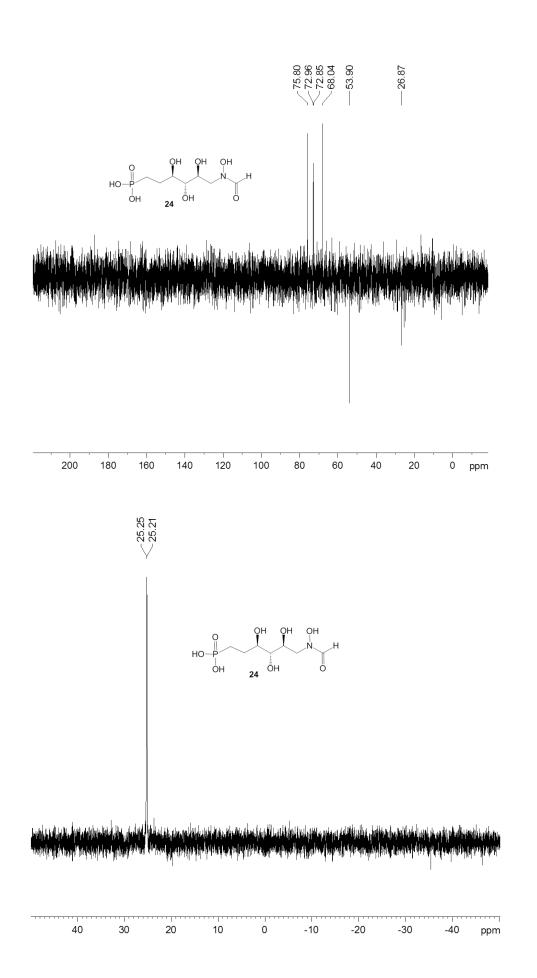
¹H, ¹³C and ³¹P NMR spectra of deprotected hydroxamate derivatives

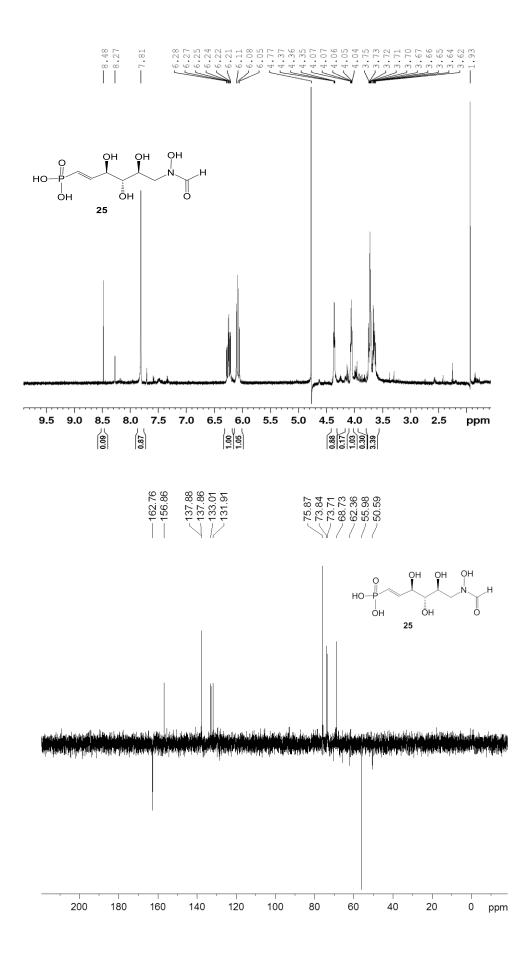
Figure S2. 1D NOe-difference spectrum of compound **17** with selective saturation of one formyl proton (top spectrum).



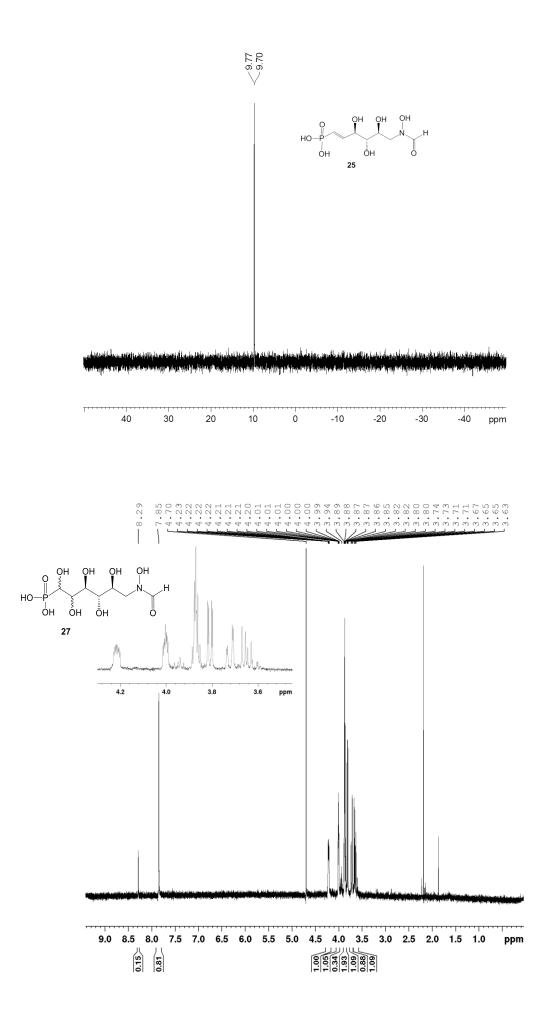




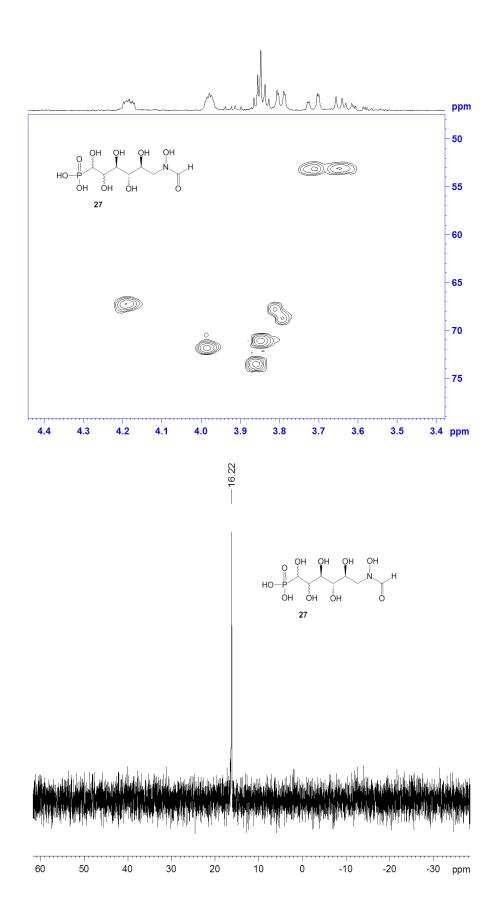


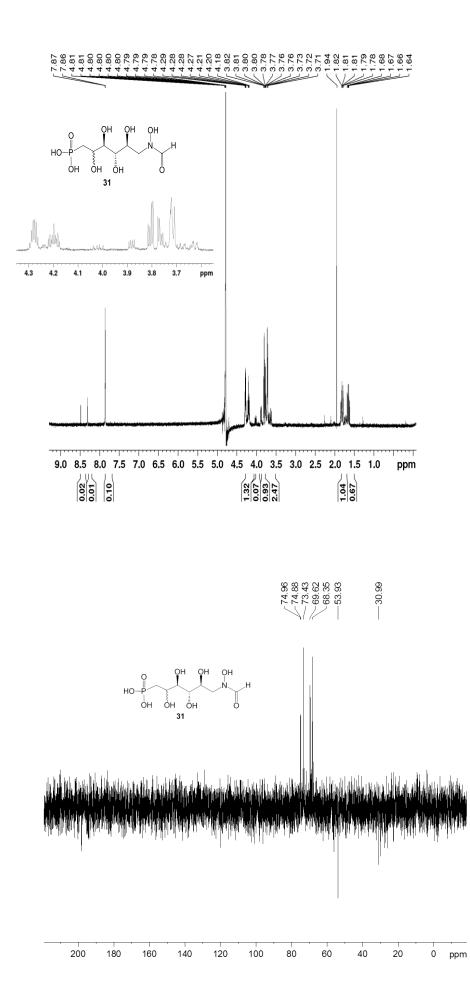


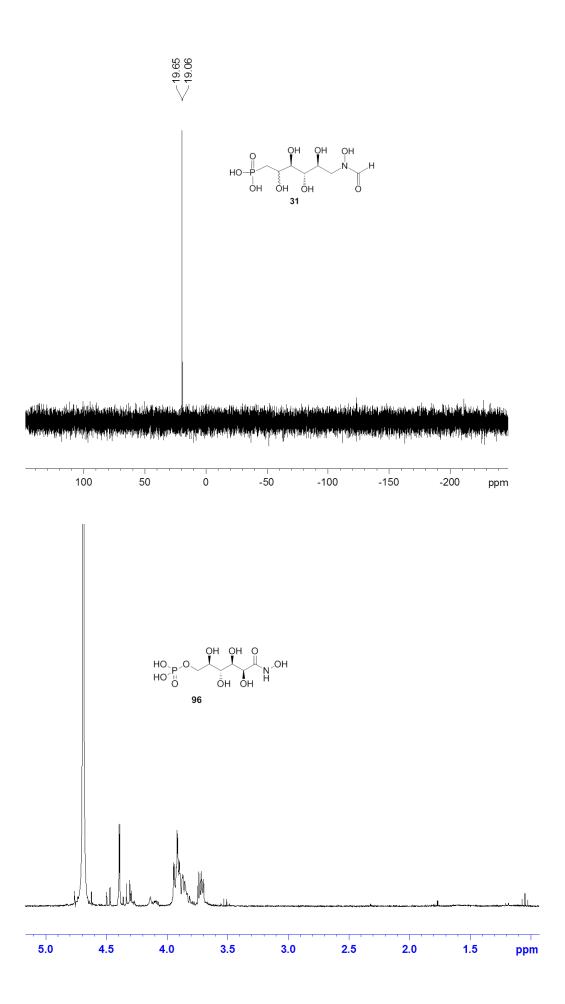
S25

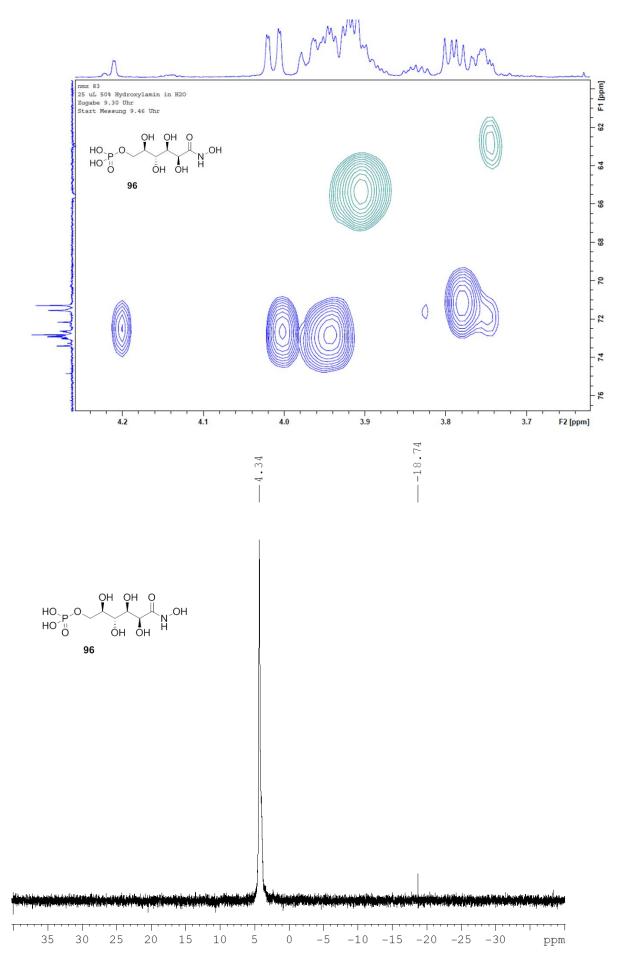


S26









The synthetic procedures for compounds **41**, **54**, **68**, **76** and **84** are fully described in the published patent WO 2014/067904 A1.

Compound **41** is described in example 4.

Compound **54** is described in example 11.

Compound **68** is described in example 15.

Compound **76** is described in example 9.

Compound **84** is described in example 12.

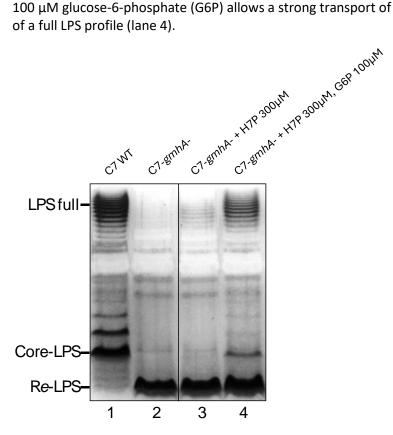
Table S1. X-Ray data	collection and	processing statistics.
Tuble D1. 11 Huy uutu	concentration und	processing stutistics.

PDB Accession Code	8V4J	8V2T
Inhibitor	17	84
Wavelength (Å)	1,075	1,100
Resolution range (Å)	36.83 - 1.31 (1.357 - 1.31)	39.06 - 1.402 (1.452 - 1.402)
Space group	P 42 21 2	P 42 21 2
Cell dimensions: a, b, c (Å)	60.972 60.972 92.429	60.906 60.906 92.728
Cell dimensions: α , β , γ (°)	90 90 90	90 90 90
Total reflections	582507 (57379)	417025 (33367)
Unique reflections	42503 (4190)	34849 (3323)
Multiplicity	13.7 (13.7)	12.0 (10.0)
Completeness (%)	99.68 (100.00)	99.64 (96.54)
Mean I/sigma(I)	41.13 (8.89)	21.23 (3.22)
Wilson B-factor	10.62	13.42
R-merge	0.03807 (0.2714)	0.06467 (0.4377)
R-meas	0.03959 (0.2819)	0.06761 (0.4607)
R-pim	0.01072 (0.07575)	0.01944 (0.1396)
CC1/2	1 (0.985)	0.999 (0.946)
CC*	1 (0.996)	1 (0.986)
Reflections used in refinement	42502 (4190)	34846 (3320)
Reflections used for R-free	2125 (212)	1745 (171)
R-work	0.1607 (0.1950)	0.1568 (0.2528)
R-free	0.1725 (0.2104)	0.1633 (0.2526)
CC(work)	0.966 (0.917)	0.966 (0.896)
CC(free)	0.959 (0.901)	0.927 (0.886)
Number of non-hydrogen atoms	1698	1647
macromolecules	1424	1407
ligands	20	22
solvent	254	218
Protein residues	192	191
RMS bond length deviation (Å)	0.01	0.008
RMS bond angle deviation (°)	1.03	0.85
Ramachandran favored (%)	98.42	98.41
Ramachandran allowed (%)	1.58	1.59
Ramachandran outliers (%)	0	0
Rotamer outliers (%)	0.68	0.69
Clashscore	0,69	1,06
Average B-factor	15.14	16.5
macromolecules	12.87	14.76
ligands	12.42	17.04
solvent	28.1	27.69

Statistics for the highest-resolution shell are shown in parentheses.

Figure S3: Role of UhpT phospho-sugar transporter in the transport of D-glycero-D-manno-heptose

7-phosphate (H7P). Complementation of *E. coli* C7-*gmhA*- by the GmhA product H7P. Silver-stained LPS SDS-PAGE of *E. coli* C7 WT (lane 1) or *gmhA*- (lanes 2-4). Culture in presence of 300 μ M H7P (lane 3) restores some full length LPS, showing a limited entry of H7P into the cells. UhpT induction by 100 μ M glucose-6-phosphate (G6P) allows a strong transport of H7P, as evidenced by the restoration of a full LPS profile (lane 4).



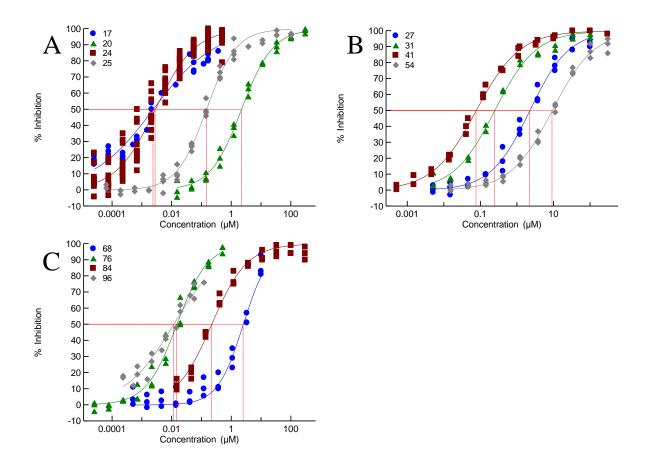


Figure S4: IC₅₀ fits of *E. coli* GmhA in biochemical inhibition assays. Red dashed lines show the fit IC₅₀ values. A: compounds **17** to **25**; B: compounds **27** to **54**; C: compounds **68** to **96**.

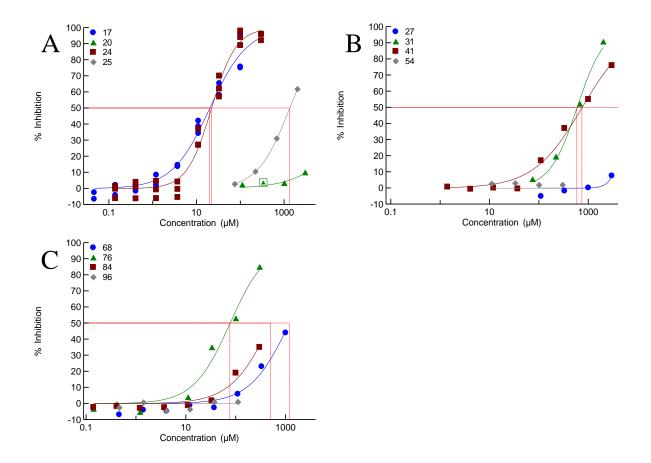


Figure S5: EC_{50} fits of LPS biosynthesis inhibition. Red dashed lines show the fit EC_{50} values. A: compounds **17** to **25**; B: compounds **27** to **54**; C: compounds **68** to **96**.