Study	Year	Link	Storage conditions	Storage duration for reported outcome	Visualization method	Storage effect	Relevant text
Watson 1986	1986	https://pubmed.ni	Free floating sections stored in phosphate buffered saline	6 weeks	Light microscopy after immunostaining for LHRH	Increased non-specific staining and the tissue tore and shredded easily while undergoing processing	"In contrast, while immunocytochemical labeling of neuronal elements was still detectable in tissue stored for up to 6 weeks in PBS, the tissue was typically unworkable in that it tore and was shredded easily while undergoing immunocytochemical processing. Also, non-specific staining was markedly increased following long term storage in PBS"
Morán 1992	1992	https://pubmed.ni	Storage of brain sections in 0.1 M phosphate buffer without sodium azide at 4° C	5 days	Light microscopy after immunostaining	"Good histological conditions" lost after 5 days	"Sections kept in 0.1 M phosphate buffer without sodium azide at 4°C, maintained good histological conditions for only 5 days. In 0.1 M phosphate buffer with sodium azide at 4°C, after 7 weeks the sections showed a reduction of AChE in positive varicose fibers, NFT and SP, while BChE could be demonstrated only in some SP"
Morán 1992	1992	https://pubmed.ni	Storage of brain sections in 0.1 M phosphate buffer with sodium azide at 4° C	7 weeks	Light microscopy after immunostaining	Better apparent maintenance of histology, but loss of some histochemistry after 7 weeks of storage	
Reynolds 1994	1994	https://pubmed.ni	Either 0.1M sodium phosphate or 0.1M sodium cacodylate (pH 7.4)	17 months	Confocal microscopy	No reported change in confocal fluorescence of myelinated fibers	"While good quality images of myelinated fibres can be acquired immediately following (glutaraldehyde) perfusion, maximal levels of fluorescence intensity were reached at about 2 weeks post-perfusion and appeared to be maintained for at least 15 months (the longest period investigated). This notable maintenence of fluorescence intensity permits specimens to be examined at the investigators' convenience."
Levine 2013	2013	https://pubmed.ni	0.1 M Phosphate Buffer	3 months	Light microscopy after Golgi-Cox staining	No reported changes in Golgi impregnation quality	"NHP brain sections selected for Golgi-Cox impregnation were transferred from 0.1M PB to unsubbed (50×75mm) slides and 'sandwiched' as described above for rat brain sections with the exception that tissue remained in sealed vials of 0.1M PB for up to three months while we established the optimal method for Golgi-Cox impregnation."
Beach 2015	2015	https://pubmed.ni	Sections stored in 0.1 mol/L phosphate buffer with 0.1% sodium azide, in a refrigerator at 4° C	Years	Light microscopy after immunostaining	Excellent for immunohistochemistry studies after years of storage	"Pre-cut free-floating 40 µm and 80 µm sections, as well as unsectioned fixed wet brain blocks, are stored in 0.1 mol/L phosphate buffer with 0.1% sodium azide, in a refrigerator at 4°C (sections) or at room temperature (fixed wet brain blocks). Sections and tissue stored in this manner are excellent for IHC studies even after years of storage."
Beach 2015	2015	https://pubmed.ni	Blocks stored in 0.1 mol/L phosphate buffer with 0.1% sodium azide, at room temperature	Years	Light microscopy after immunostaining	Excellent for immunohistochemistry studies after years of storage	
Sele 2020	2020	https://forschung.	0.1 M cacodylate buffer pH 7.4 or 0.1 M phosphate buffer pH 7.4 with sodium azide	Up to 14 months	Electron microscopy	No adverse effect of storage length on ultrastructure, based on quantitative ratings	"No significant differences in the ratings could be determined when the initial score was compared with the score of the stored samples. Therefore, no adverse effects regarding the duration of storage on the ultrastructure could be recognised."