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Supplemental information

Lymph node dendritic cells harbor inducible

replication-competent HIV despite years

of suppressive ART

Riddhima Banga, Francesco Andrea Procopio, Erica Lana, Gregory T. Gladkov, Isabelle Roseto, Elizabeth M. Parsons, Xiaodong Lian, Marie Armani-Tourret, Maxime Bellefroid, Ce Gao, Annamaria Kauzlaric, Mathilde Foglierini, Oscar Alfageme-Abello, Susanna H.M. Sluka, Olivia Munoz, Andrea Mastrangelo, Craig Fenwick, Yannick Muller, Catherine Gerald Mkindi, Claudia Daubenberger, Matthias Cavassini, Rafael Trunfio, Sébastien Déglise, Jean-Marc Corpataux, Mauro Delorenzi, Mathias Lichterfeld, Giuseppe Pantaleo, and Matthieu Perreau



Figure. S1 (Related to Figure 1 and Figure 4): Gating strategy for sorting LN resident and migratory DCs and purity analyses. (A) Representative flow cytometry profiles of the gating strategy for sorting LN resident and migratory DCs from a representative ART treated HIV-infected individual (#136). Upon exclusion of T and B cells, myeloid DCs were identified within the CD11c⁺ fraction and distinguished into LN resident DCs (CD3⁻CD20⁻HLA-DR⁺CD45⁺CD11c⁺CCR7⁻) migratory DCs (CD3⁻CD20⁻HLAand LN $DR^+CD45^+CD11c^+CCR7^+$). (B) Representative flow cytometry profile of the gating strategy for sorting LN PD-1⁺/Tfh and PD-1⁻/non Tfh cells from a representative viremic HIV-infected individual (#1024). (C) Purity analysis of sorted LN cell populations i.e. LN resident and migratory DCs and CD4 T cells overlaid on unsorted total LN cells from three representative ART treated HIV-infected individuals. Purity for all sorting experiments was \geq 98%. (D) Detection of TREC copies in sorted CD4 T cells at various cell concentrations (5.10⁵-1 cell). (E) Detection of TREC copies in 40,000 sorted LN resident and migratory DCs from HIV uninfected individuals (N=3). Total LN cells are represented in grey (C), LN migratory DCs are depicted in violet (C and E), LN resident DCs are depicted in green (C and E) and LN CD4 T cells are depicted in blue (**C** and **D**). Red bars correspond to mean ± SEM (**E**). Statistical significance (*=P<0.05) was obtained using Spearman rank test (**D**).



Figure. S2 (Related to Figure 2): Mass cytometry gating strategy for LN mononuclear cell populations. (A) Representative example of gating strategy for LN resident and migratory DCs of an aviremic ART treated HIV-infected individual (#155). (B) Representative example of gating strategy for naïve CD4 T cells and LN Tfh cells of an aviremic ART treated HIV-infected individual (#155).



LN Migratory DCs LN Resident DCs

Tfh cells



Figure. S3 (Related to Figure 2): Phenotypic characterization of LN resident and migratory DCs by mass cytometry. (A) Representative t-distributed stochastic neighbor embedding (t-SNE) plots representing the surface expression patterns of co-stimulatory molecules (CD40, OX-40 and CD86), immunoregulatory molecules (PD-L1, PD-L2 and CD155) and other molecules (CD14, CD11b, CD45, HL-DR, CD127, CD1c and CD11c) on naïve CD4 T cells, LN migratory and LN resident DCs from a representative ART treated HIVinfected individual (#161). (B) Representative mass cytometry profile of expression levels of transcription factors IRF4 and IRF8 in naïve CD4 T cells, LN Tfh cells, LN resident and LN migratory DCs isolated directly ex vivo from a representative HIV uninfected, viremic (#1009) and aviremic ART treated HIV-infected individual (#163).(C) Cumulative data representing the frequencies of LN migratory and resident DCs expressing PD-L1, PDL2, CD155, CD127, CD40 and CD86 in HIV-uninfected (N=4), viremic (N=5) and aviremic ART treated HIV-infected individuals (N=7) or the Mean signal intensity (MSI) values of IRF4 and IRF8 in naïve CD4 T cells, LN migratory and resident DCs isolated directly ex vivo from HIV uninfected (N=4), viremic (N=6) and ART treated HIV-infected individuals (N=7). Red histograms correspond to naïve CD4 T cells, green histograms correspond to LN Tfh cells, orange histograms correspond to LN migratory DCs and blue histograms correspond to LN resident DCs (B). Grey circles correspond to HIV-uninfected individuals, blue squares corresponds to viremic HIV-infected individuals and green triangles correspond to aviremic ART treated HIV-infected individuals (C). Red bars correspond to mean \pm SEM (C). Red stars indicate statistical significance (*=P<0.05) for intra-group comparisons. Green stars indicate statistical significance (*=P<0.05) for intra-group comparisons of LN migratory DCs, LN resident DCs and naïve CD4 T cells. Statistical significance (P values) was obtained using oneway ANOVA (Kruskal-Wallis test) followed by a two-tailed paired t-test.



Figure. S4 (Related to Figure 6): Levels of HIV RNA in the culture supernatants of LN CD4 T cells exposed to TLR7/8 agonist treatment. Levels of HIV production in purified LN CD4 T cell culture supernatants of ART treated HIV-infected individuals (N=5) at days 0, 5 and 14 post-exposure to TLR7/8 agonist as assessed by HIV RNA levels (copies/mL). HIV-infected individuals are color coded. Red bars correspond to mean \pm SEM (C). Red stars indicate statistical significance (*=P<0.05). Statistical significance (P values) was obtained using one-way ANOVA (Kruskal-Wallis test) followed by Wilcoxon Matched-pairs one-tailed Signed Rank test.

Supplemental Table 3 (Related to Figure 1). HIV-specific CD8 T-cell responses in ART treated HIV-infected individuals

HIV treated individuals	Peptide sequences	SFU/million		
	RDYVDRFFKTL	227		
	TPQVPLRPM	192		
#136	SPAIFQSSM	162		
	WHLGHGVSI	140		
	total SFU/million	721		
#142	YPLTFGWCY	233		
	VPLDEDFRKY	150		
#169	IEELRQHLL	140		
	total SFU/million	290		

Supplemental Table 4 (Related to Figure 4). All Flip-seq sequences obtained for viremic HIV-infected individuals

								Defective					
Patient	LN cell population	Cell count before extraction	LTRgag Count after extraction	% of cells with LTRgag	Total cells assayed	Total HIV Sequen ces	Intact	Defects in PSI	LD	LD/ Hypermut ation	LD/ Internal Inversion	PSC	Hypermutation
1024	Mig DC	3.00E+05	1.76E+03	0.6	5,684	1	1	-	-	-	-	-	-
	Res DC	2.80E+05	1.40E+03	0.5	7,146	2	-	-	1	1	-	-	-
	PD-1+/Tfh	3.00E+05	2.08E+04	6.9	5,292	9	2	0	5	2	-	-	-
	PD-1 ⁻	3.00E+05	2.64E+03	0.9	41,967	5	-	-	4	-	-	1	-
1019	Mig DC	1.96E+05	4.32E+02*	0.2	3,476	1	-	-	1	-	-	-	-
	Res DC	1.80E+05	1.80E+02*	0.1	8,322	1	1	-	-	-	-	-	-
	PD-1+/Tfh	2.70E+05	3.29E+04	12.2	1,826	4	2	-	1	1	-	-	-
	PD-1 ⁻	3.00E+05	7.81E+03	2.6	3,743	-	-	-	-	-	-	-	-
	Mig DC	3.00E+05	1.10E+03	0.4	6,600	1	1	-	-	-	-	-	-
1025	Res DC	3.00E+05	7.77E+01*	0.03	19,305	2	2	-	-	-	-	-	-
	PD-1+/Tfh	3.00E+05	4.33E+04	14.4	3,663	2	-	-	1	-	1	-	-
	PD-1	3.00E+05	2.31E+03*	0.8	1,601	-	-	-	-	-	-	-	-

Asterix (*) refers to quantifications at limit of detection. When no intact HIV-1 sequences were detected, the limit of detection was calculated as two times the number of cells analyzed without target identification. Percentage of HIV-infected cells harboring at least 1 copy of HIV was calculated as follows: LTR HIV*gag* count (copies) / cell count before extraction*100.

Supplemental Table 5 (Related to Figure 4). All MIP-seq sequences obtained for viremic HIV-infected individuals

					Defective					
Patient	LN cell population	Total cells assayed	Total HIV Sequences	Intact	Defects in PSI	LD	LD/ Hypermutation	LD/ Internal Inversion	PSC	Hypermutation
1024	Mig DC	23,012	6	2	-	2	-	-	2	-
	Res DC	36,286	9	4	-	5	-	-	-	-
	PD-1+/Tfh	619	4	1	-	3	-	-	-	-
	PD-1 ⁻	0	-	-	-	-	-	-	-	-
	Mig DC	3,476	1	1	-	-	-	-	-	-
	Res DC	11,927	1	0	-	1	-	-	-	-
1019	PD-1+/Tfh	348	2	0*	-	-	-	-	-	-
	PD-1 ⁻	0	-	-	-	-	-	-	-	-
	Mig DC	5,984	1	1	-	-	-	-	-	-
1025	Res DC	26,255	0	0	-	-	-	-	-	-
	PD-1+/Tfh	228	3	0*	-	-	-	-	-	-
	PD-1 ⁻	NA	NA	NA	NA	-NA	NA	NA	NA	NA

Abbreviations: LD: Large deletions; Pre-mature stop codon. *Short sequences (<~1200 bp) were not sequenced. NA; Not applicable

Supplemental Table 6 (Related to Figure 6). Percentage of homology between HIV sequences obtained from VOA culture supernatants of LN CD4 T cells and LN DC subpopulations of treated HIV-infected individuals.

Volunteer #104									
Reverse Transcriptase (1	158 nucleotides)								
Cell populations	CD4 T cells	Resident DCs Migratory DC							
CD4 T cells		99 (6 mismatch)	100						
Resident DCs	99 (6 mismatch)		99 (6 mismatch)						
Migratory DCs	100	99 (6 mismatch)							
Protease (297 nucleotides	s)								
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		99 (3 mismatch)	100						
Resident DCs	99 (3 mismatch)	99 (3 mismat							
Migratory DCs	100 99 (3 mismatch)								
Integrase (867 nucleotide	es)								
Cell populations	CD4 T cells	Resident DCs Migratory D							
CD4 T cells		100 (2 mismatch)	100						
Resident DCs	100 (2 mismatch)	100 (2 mismatch							
Migratory DCs	100	100 (2 mismatch)							
Volunteer #109									
Reverse Transcriptase (1	158 nucleotides)								
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		NA	NA						
Resident DCs	NA		99 (8 mismatch)						
Migratory DCs	NA	99							
Protease (297 nucleotides	s)	·	·						
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		NA	NA						
Resident DCs	NA		98 (8 mismatch)						
Migratory DCs	NA	98							
Integrase (867 nucleotide	es)								
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		NA	NA						
Resident DCs	NA		100 (1 mismatch)						
Migratory DCs	Aigratory DCs NA 100								
Volunteer #133									
Reverse Transcriptase (1	158 nucleotides)								
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		100	100						
Resident DCs	100		100						
Migratory DCs	100	100							
Protease (297 nucleotides)									
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		100	100						
Resident DCs	100		100						
Migratory DCs	100	100							
Integrase (867 nucleotides)									
Cell populations	CD4 T cells	Resident DCs	Migratory DCs						
CD4 T cells		100	100						
Resident DCs	100		100						
Migratory DCs	100	100							

*NA. Not applicable