

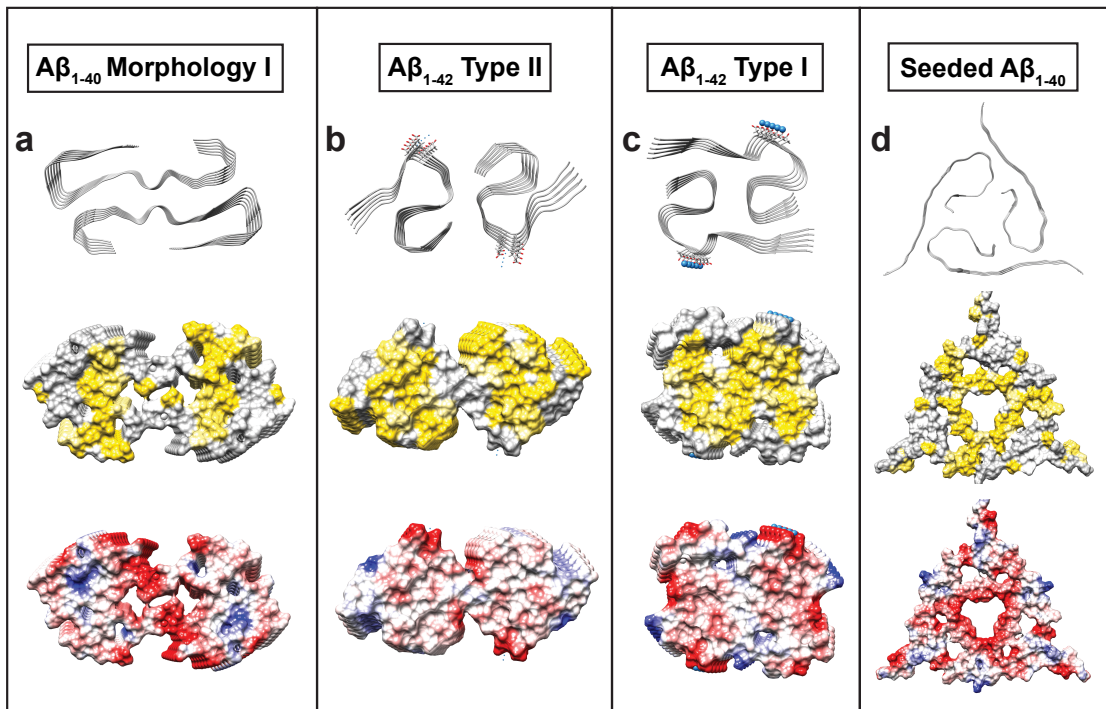
## SUPPLEMENT

### Molecular modeling of apoE in complexes with Alzheimer's amyloid- $\beta$ fibrils from human brain suggests a structural basis for apolipoprotein co-deposition with amyloids

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#### CONTENT:

Supplemental Figures 1-9, Supplemental Table 1



**Supplemental Fig. 1** Structural polymorphs of patient-based  $A\beta_{1-40}$  and  $A\beta_{1-42}$  fibrils used in the current study. Views down the fibril axis show ribbon diagrams (top) and space-filling models depicting hydrophobic (middle) and Coulombic surfaces (bottom). Color coding in these and in other space-filling models: hydrophobic – yellow, acidic – red, basic – blue.

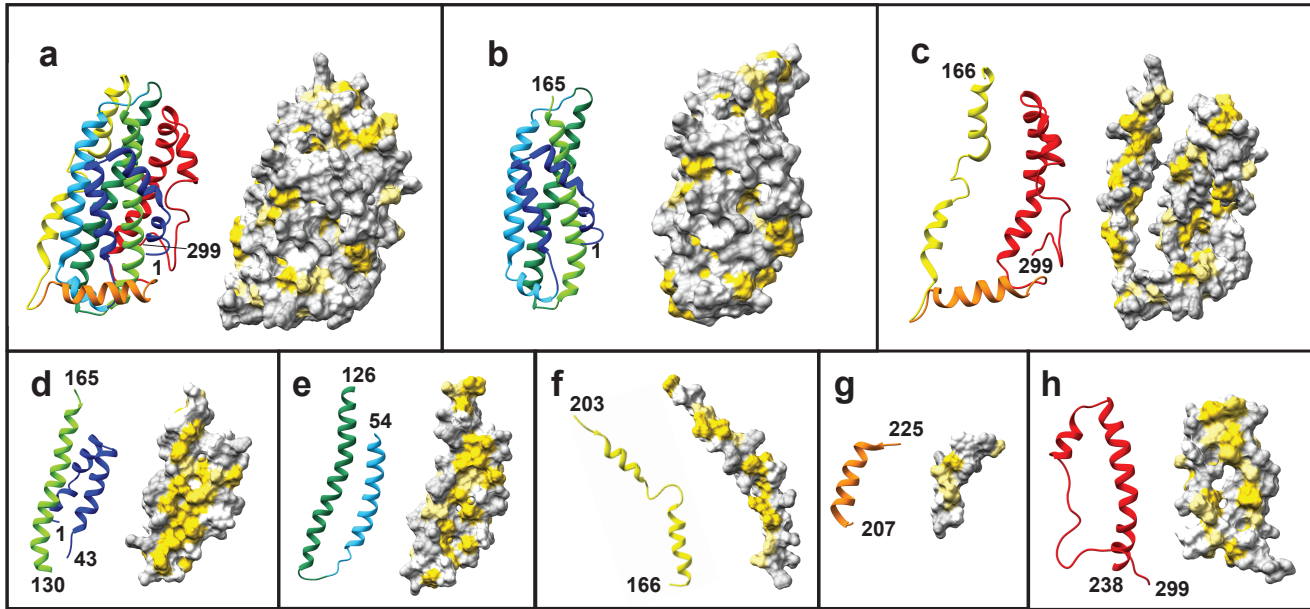
(a)  $A\beta_{1-40}$  fibril morphology I from AD vasculature, cryo-EM structure (PDB ID: 6SHS);

(b)  $A\beta_{1-42}$  type II from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4M);

(c)  $A\beta_{1-42}$  type I from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4B);

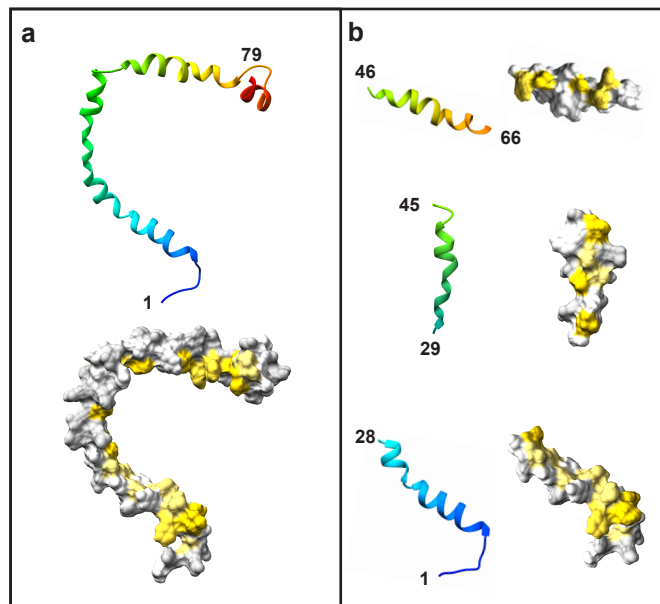
(d)  $A\beta_{1-40}$  seeded fibril with seed isolated from AD brain tissue, NMR structure (PDB ID: 2M4J).

In panels b and c, small blue spheres indicate putative metal ions bound *via* acidic ladders of E22, D23.

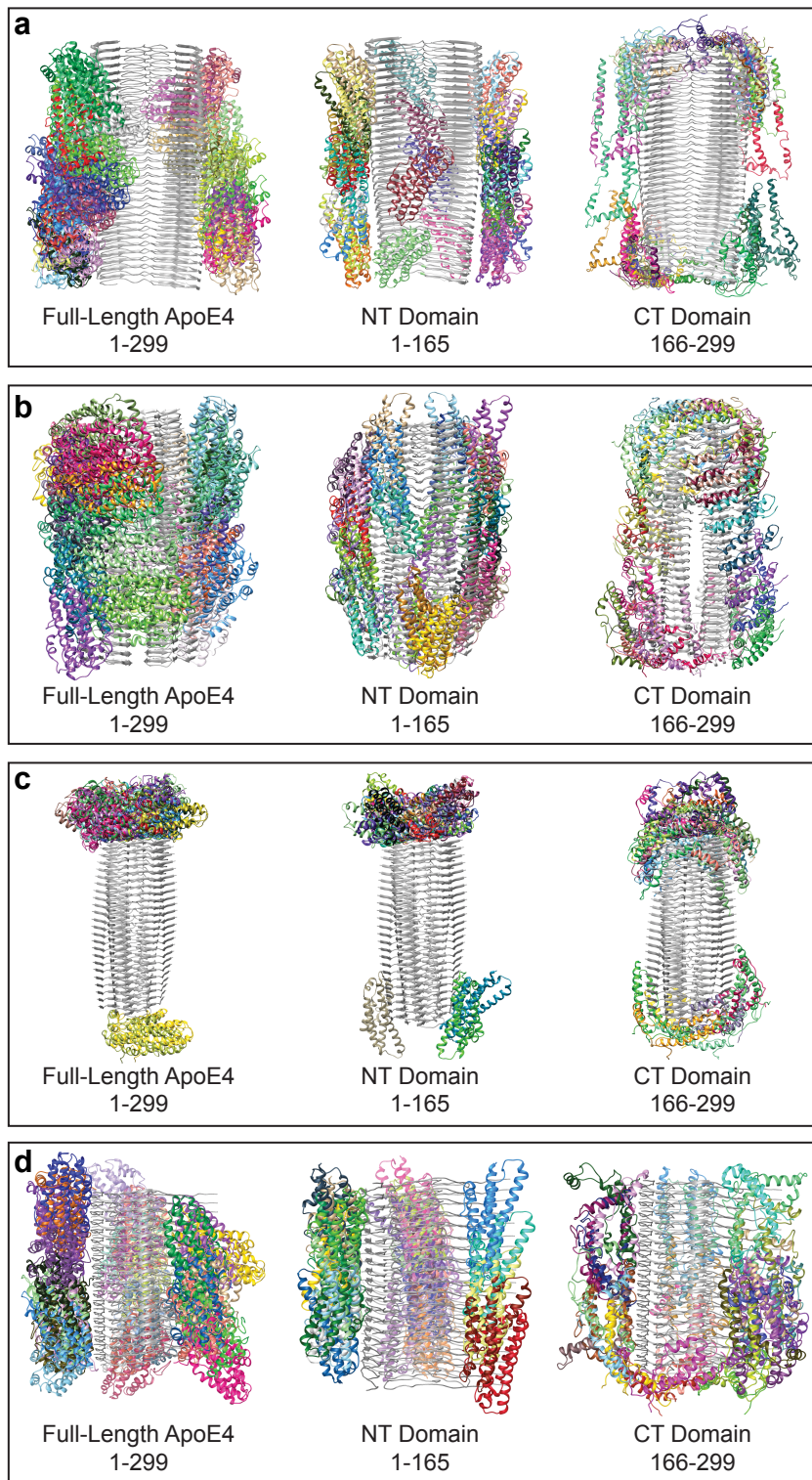


**Supplemental Fig. 2** Solution NMR structure of full-length lipid-free modified human apoE3 (PDB ID: 2L7B) and its fragments used in the current study. Helices in the ribbon diagrams are rainbow-colored N to C (blue to red). Space-filling models are oriented to show hydrophobic surfaces (in yellow). First and last residues in each fragment are shown by numbers.

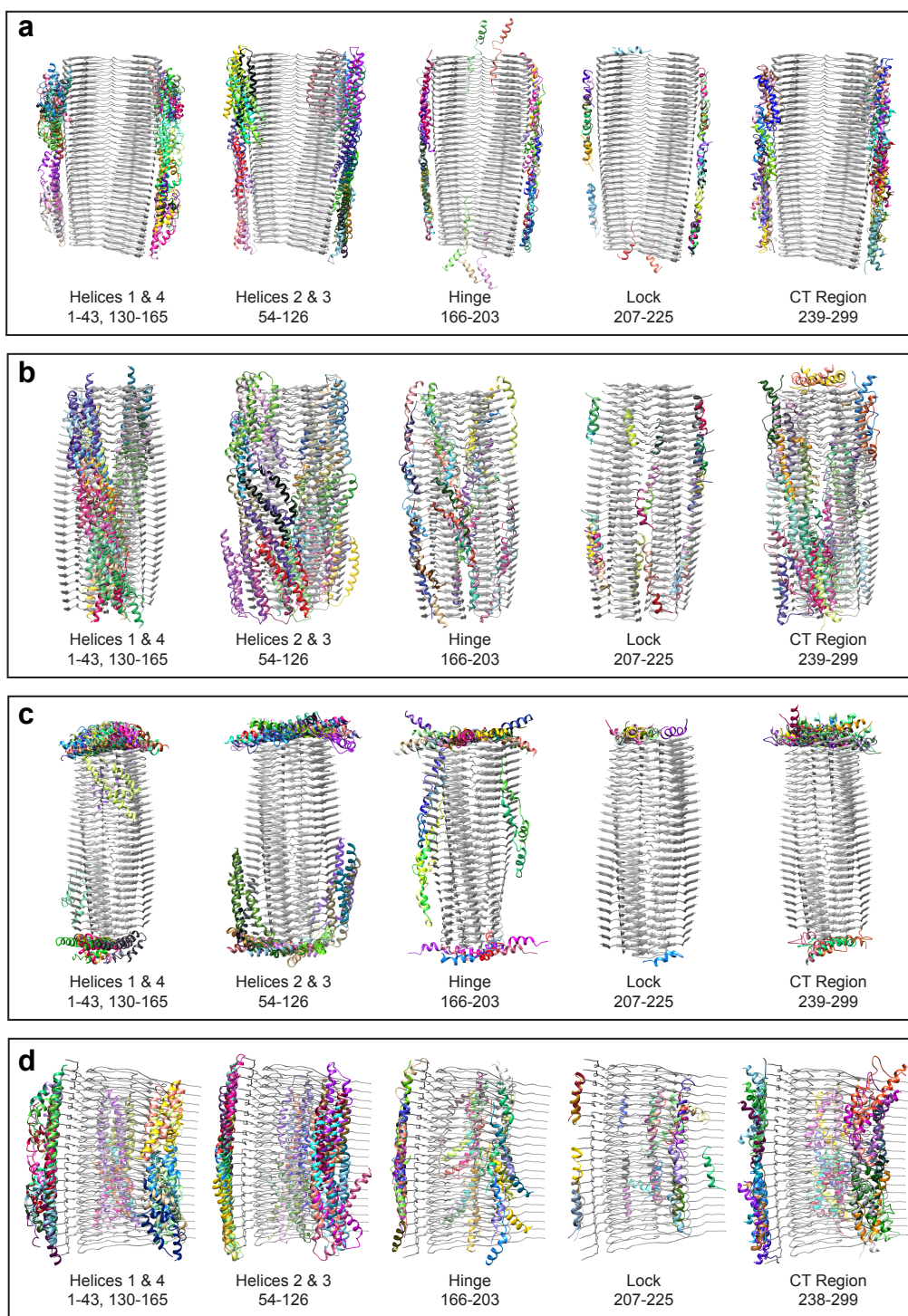
(a) Full-length apoE; (b) NTD; (c) CTD. (d-h) Segments that yielded consistent results in ClusPro docking to amyloid fibrils: (d) helices 1 and 4, (e) helices 2 and 3, (f) hinge, (g) lock, and (h) CT region.



**Supplemental Fig. 3** Solution NMR structure of full-length human apoC-III (PDB ID: 2JQ3) and its fragments used in the current study. The structure, which was determined by using the apolipoprotein bound to SDS micelles (diameter ~4.4 nm), represents the lipid-bound conformation. Ribbon diagrams are rainbow colored N to C (blue to red); space-filling models are oriented to show hydrophobic surfaces (in yellow). First and last residue numbers are indicated. (a) Full-length apoC-III. (b) NT, middle, and truncated CT fragments, which yielded consistent results in ClusPro docking to amyloid fibrils.



**Supplemental Fig. 4** Summary of docking poses for full-length apoE and its NTD and CTD fragments docked using Cluspro onto four A $\beta$  fibrils. Residues in each fragment are shown by numbers. The top-scoring docking poses for each fragment are shown; each pose is colored differently. The fibrils are:  
 (a) A $\beta$ <sub>1-40</sub> fibril morphology I from AD vasculature, cryo-EM structure (PDB ID: 6SHS);  
 (b) A $\beta$ <sub>1-42</sub> type II from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4M);  
 (c) A $\beta$ <sub>1-42</sub> type I from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4B);  
 (d) A $\beta$ <sub>1-40</sub> seeded fibril with the seed isolated from AD brain tissue, NMR structure (PDB ID: 2M4J).



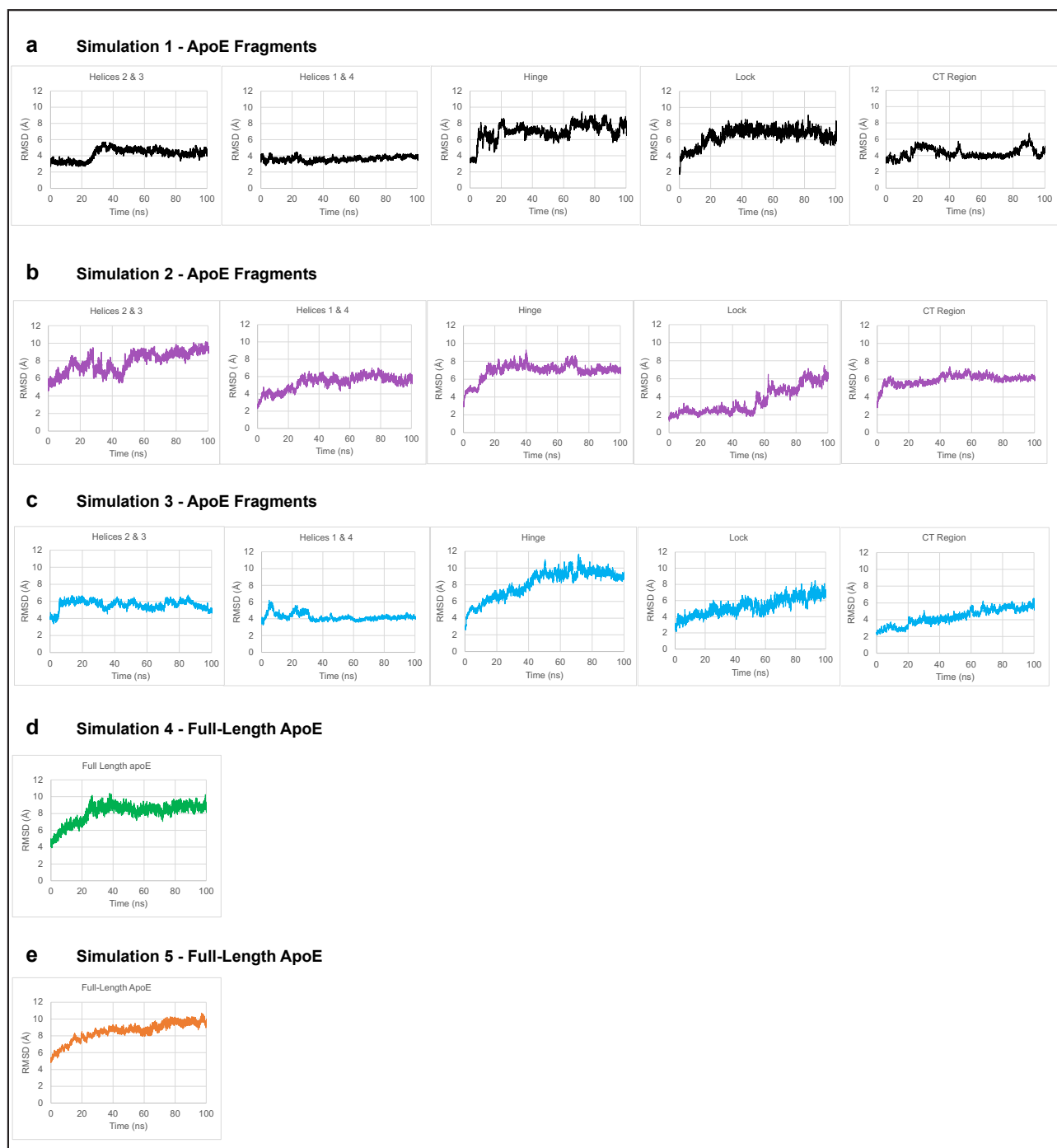
**Supplemental Fig. 5** Summary of docking poses for apoE fragments docked onto four different A $\beta$  fibrils. ApoE was split into fragments shown in supplemental Fig. 2d-h; the residues in each fragment are shown by numbers. The top-scoring docking poses for each fragment are shown; each pose is colored differently. The fibril structure is in gray. The fibrils are:

(a) A $\beta$ <sub>1-40</sub> fibril morphology I from AD vasculature, cryo-EM structure (PDB ID: 6SHS);

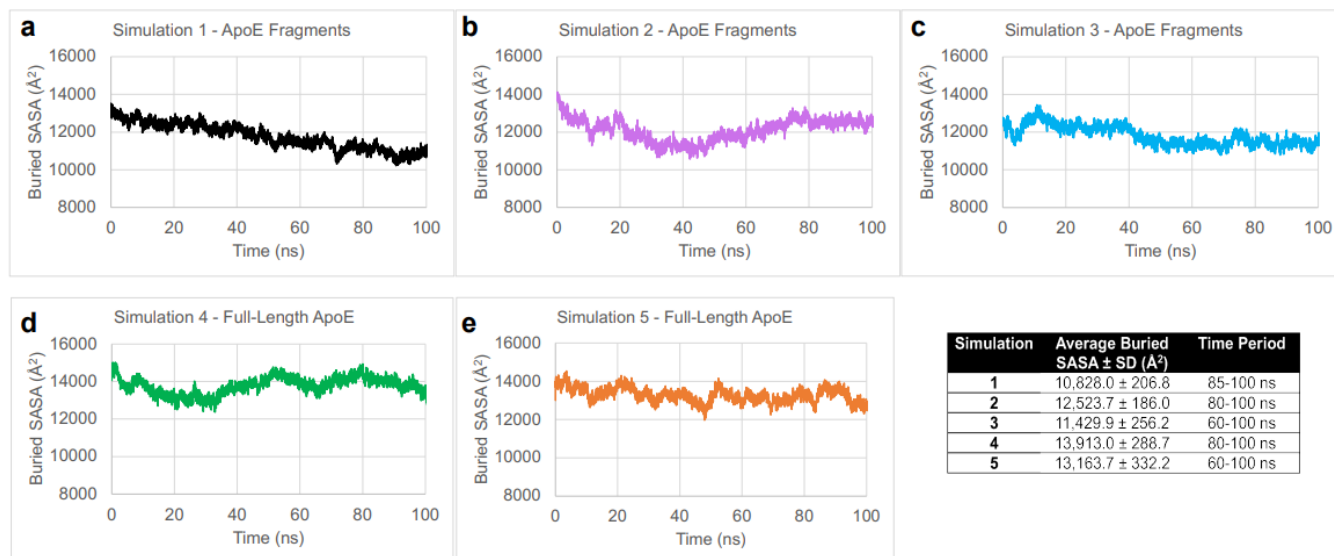
(b) A $\beta$ <sub>1-42</sub> type II from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4M);

(c) A $\beta$ <sub>1-42</sub> type I from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4B);

(d) A $\beta$ <sub>1-40</sub> seeded fibril with seed isolated from AD brain tissue, NMR structure (PDB ID: 2M4J)



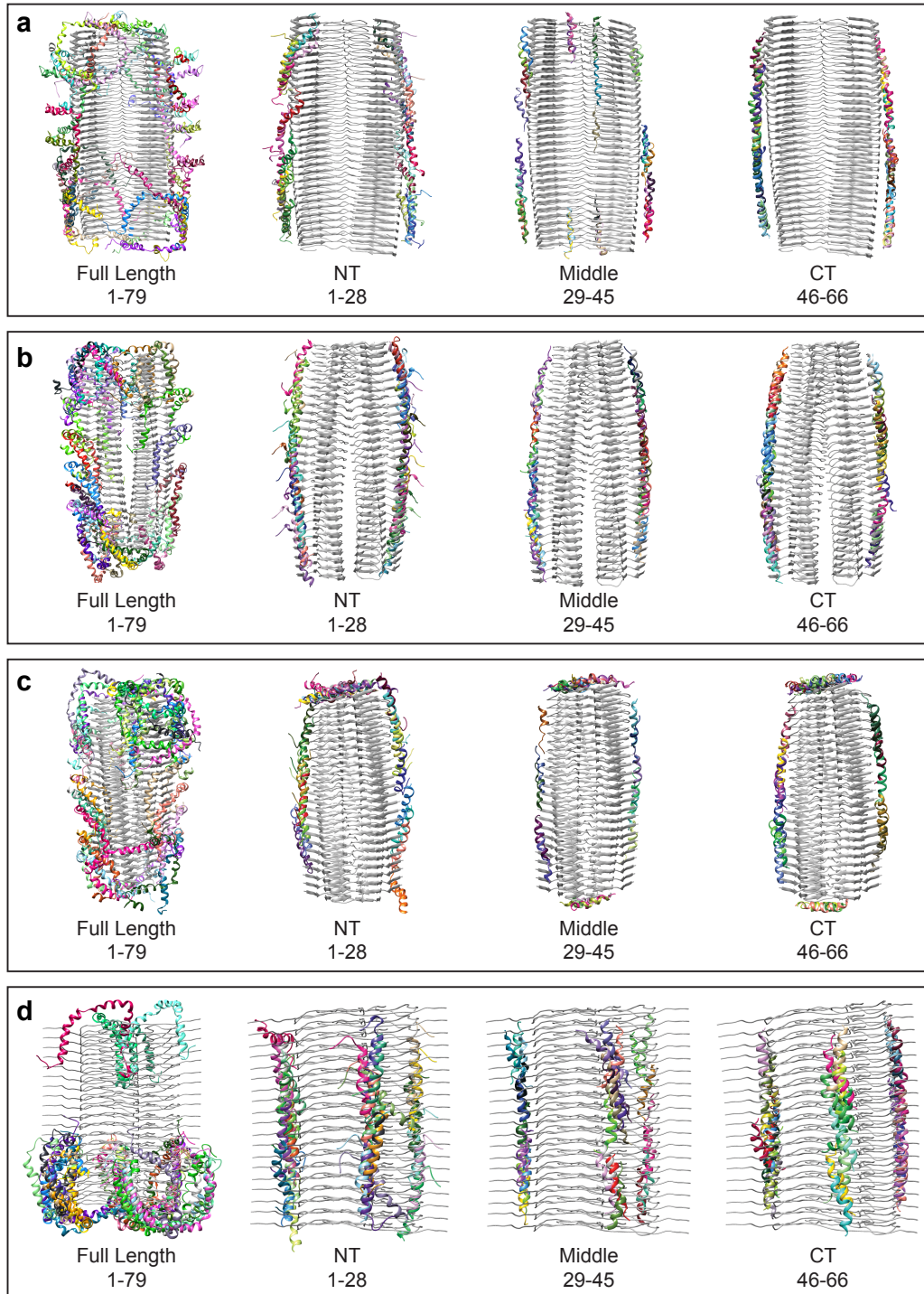
**Supplemental Fig. 6** Root mean square deviations (RMSD) for the backbone atoms in MD simulations of apoE in complex with A $\beta$ <sub>1-42</sub> type II fibril during production run time. Each model contained either 56 (a, b, d) or 64 fibril rungs (c, e). RMSD are shown for (a-c) triplicate simulations of each apoE fragment, and for (d, e) duplicate simulations of the full-length apoE model generated from these fragments as described in the text. For RMSD measurements, each frame was aligned by C $\alpha$  atoms either by fibril segments adjacent to the fragment of interest (in a-c) or the entire fibril (in d-e).



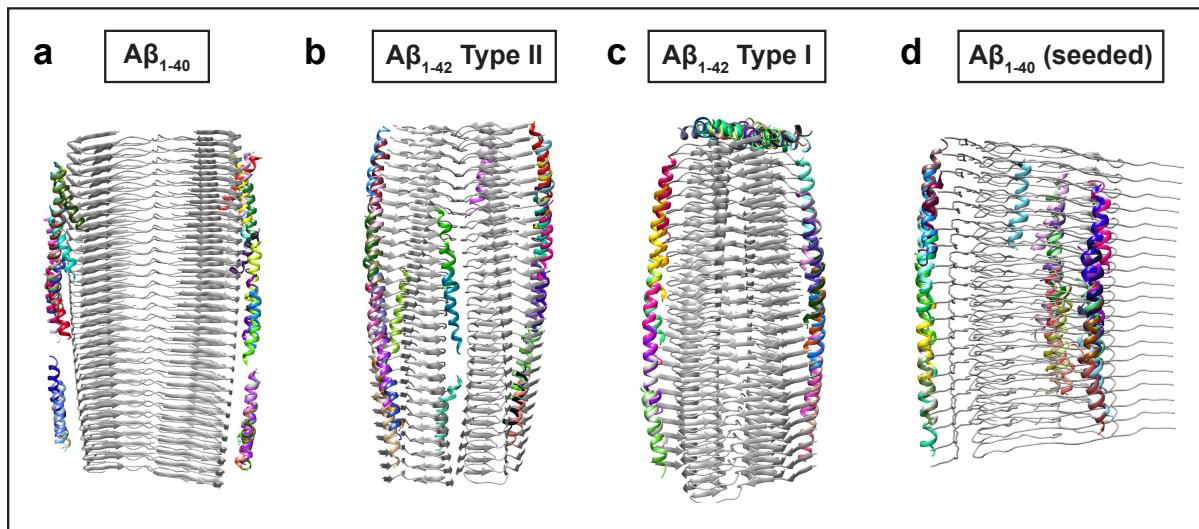
**Supplemental Fig. 7** Buried solvent-accessible surface area (SASA) between apoE and A $\beta$ <sub>1-42</sub> fibril during MD simulations. The values are shown for (a-c) triplicate simulations of apoE fragments in complex with A $\beta$ <sub>1-42</sub> type II fibril, and for (d, e) duplicate simulations of the full-length apoE – fibril model generated from these fragments. Respective models are shown in Fig 2b, d of the main manuscript. Average buried SASA for the last 20-40 ns of the production run is tabulated for each simulation.

A $\beta$	ApoE
<i>Glu22</i>	Lys143
<i>Asp23</i>	Lys95, Gln98, Arg136, Ser139, Arg142, Lys143, Lys146, Arg147, Arg150, Arg158, Arg217, Met221, Arg224, Arg274, Gln279, Gln284, Thr289, Ser290
<i>Val24</i>	Arg147, Met221, Arg224, Gln284, Thr289, Ser290
<i>Gly25</i>	His140, Arg147, Met218, Met221, Arg224, Gln284, Ala285, Thr289, Ser290
<i>Ser26</i>	His140, Met218
<i>Val36</i>	Arg25, Leu28, Tyr36, Trp39, Gln81, Arg167
<i>Gly37</i>	Arg25, Leu28, Ala29, Tyr36, Trp39, Leu63, Gln81, Arg167, Glu168
<i>Gly38</i>	Arg25, Trp26, Leu28, Ala29, Tyr36, Trp39, Val56, Leu60, Leu63, Tyr74, Gln81, Glu168, Gly169
<i>Val39</i>	Trp26, Ala29, Leu30, Phe33, Tyr36, Leu27, Trp39, Val40, Leu60, Leu63, Met64, Thr67, Leu71, Tyr74, Leu78, Thr83, Glu87, Glu88, Leu104, Met108, Val111, Tyr118, Met125, Leu133, Leu137, Leu159, Tyr162, Gln163, Gly169, Arg172, Gly173, Ala176, Ile177, Leu198, Ala199, Arg215, Ile250, Arg251, Gln253, Ala254, Ala257, Phe265, Val294, Pro295
<i>Ile41</i>	Ala91, Arg92, Arg103, Val111, Arg114, Met125, Leu133, Leu137, His140, Leu141, Asp151, Leu155, Leu159, Tyr162, Arg172, Ala176, Arg180, Leu181, Gln187, Arg189, Val190, Ala193, Leu198, Ala199, Trp210, Gly211, Leu214, Arg215, Met218, Glu219, Gly222, Ser223, Ile250, Phe265, Ala269, Gln273, Trp276, Ala292

**Supplemental Table 1.** Contact table listing close contacts between the backbone atoms of amino acids from apoE and A $\beta$ <sub>1-42</sub> fibrils from parenchymal deposits of AD patients. Contacts within 5Å which are seen in  $\geq 75\%$  of frames are listed for simulation 3 of the model containing apoE fragments and A $\beta$ <sub>1-42</sub> type II fibril. The model, which is shown in Fig. 2b of the main manuscript, was subjected to MD simulations as described in the text. Representative simulation 3 was used to determine the contacts; RMSD and buried SASA for simulation 3 are shown in supplemental Figs. 6c and 7c.



**Supplemental Fig. 8** Summary of docking poses for full-length apoC-III and its NT, middle, and truncated CT fragments docked onto four different A $\beta$  fibrils. Residues numbers in each protein fragment are shown. Top-scoring docking poses are shown, each is colored differently. The fibrils are: (a) A $\beta$ <sub>1-40</sub> fibril morphology I from AD vasculature, cryo-EM structure (PDB ID: 6SHS); (b) A $\beta$ <sub>1-42</sub> type II from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4M); (c) A $\beta$ <sub>1-42</sub> type I from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4B); (d) A $\beta$ <sub>1-40</sub> seeded fibril with seed isolated from AD brain tissue, NMR structure (PDB ID: 2M4J).



**Supplemental Fig. 9** Summary of docking poses for the consensus sequence peptide to A $\beta$  fibrils. CSP, which is comprised of a 22-residue sequence repeat motif representing class-A amphipathic  $\alpha$ -helices found in apoA-I, apoA-IV and apoE (PLAEELRARLRAQLEELRERLG), was docked onto four different A $\beta$  fibrils. Top-scoring docking poses are shown; each is colored differently. The fibrils are: (a) A $\beta$ <sub>1-40</sub> fibril morphology I from AD vasculature, cryo-EM structure (PDB ID: 6SHS); (b) A $\beta$ <sub>1-42</sub> type II from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4M); (c) A $\beta$ <sub>1-42</sub> type I from parenchymal deposits, cryo-EM structure (PDB ID: 7Q4B); (d) A $\beta$ <sub>1-40</sub> seeded fibril with seed isolated from AD brain tissue, NMR structure (PDB ID: 2M4J).