

Aggregation-prone TDP-43 sequesters and drives pathological transitions of free nuclear TDP-43

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Supplementary Materials

A mAvicFP1 insertion HR template

5'-

ATGGAGGCTTTGGCTCAAGCATGGATTCTAAGTCTTCTGGCTGGGGAATG**GACTACAAAGACGATGACGACAAGGAA**
CAAAAACTCATCTCAGAAAGGGATCTG**ATGAGCAAAGGAGCAGAGCTTTTCAACGGGATTGTTCCAATTCTTATTGAA**
TAAATGGTGATGTT**CACGGACACAAATTCTCTGTTAGAGGAGAGGGAGAAGGCGATGCCGATCTGGAAAAATTGA**
AATCAAATTTGTTTGCACTACTGGTACACTTCTGTGCCATGGCCACCCTTGT**CACCACATTGTGTTATGGTGTCCA**
GTGCTTCACTCGATACCCAAGACACAT**GAAACAGCATGATTTCTACAAGAGTGCCATGCCAGACGGTTACATACAAAA**
AAGAACTACAGTTTCCAAGACGACGGGCACTACAAGACACG**TGCTCAAGTCAAGTTTGAAGGTGATACCCTTGTTAA**
TAGAATCGAGTTAAAAGGTATTGATTTTAAAGAAGATG**GAAACATTCTTGAAATAAAATGGAATACA**ACTATAACTCA
CATTCTGTATACGTCTTGT**CAGACAAAGCAAACAATGGAATCAAAGTGA**ACTTCAAGATTAGACACAACCTTAAAGGT
GAAGTATACAACTTGCAGACCAGATCAACAAAATATTCTATTGGT**GATGGTCTGTTCTTCTACCAGACTATCATT**
ACCTCTCCACGCAAATAAAATAACTAAAGATCTAAT**GAAAAGAGAGACCACATGAACCTGGTCGAGTTTGTGACAG**
CTTGTGGAATCACCCATGGCATGGATGAGTTGTACAAGTAATAGACAGTGGGGTTGTGGTTGGTTGGTATAGAATGG
TGGGAATTCAAATTTTTCTA-3'

B mCherry insertion HR template

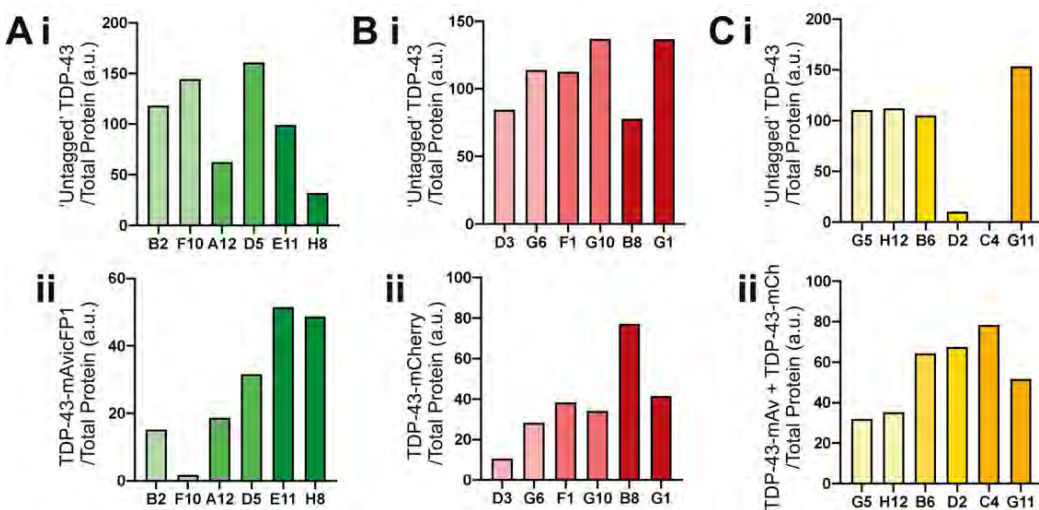
5'-

ATGGAGGCTTTGGCTCAAGCATGGATTCTAAGTCTTCTGGCTGGGGAATG**GACTACAAAGACGATGACGACAAGGAA**
CAAAAACTCATCTCAGAAAGGGATCTG**ATGGTGAGCAAGGGCGAGGAGGATAACATGGCCATCATCAAGGAGTTCAT**
CCGCTTCAAGGTGCACATGGAGGGCTCCGTGAACGGCCACAGTTCGAGATCGAGGGCGAGGGCGAGGGCCGCC
CCTACGAGGGCACCCAGACCCGCAAGCTGAAGGTGACCAAGGGTGGCCCTGCCCTTCCGCTGGACATCCTGT
CCCCTCAGTTCATGTACGGCTCCAAGGCCTACGTGAAGCACCCCGCGACATCCCCGACTACTTGAAGCTGTCTTC
CCCGAGGGCTTCAAGTGGGAGCGCGTGATGAACTTCGAGGACGGCGGCGTGGTGACCGTGACCCAGGACTCCTCC
CTGCAGGACGGCGAGTTCATCTACAAGGTGAAGCTGCGCGGCACCAACTTCCCCTCCGACGGCCCCGTAATGCAGA
AGAAGACCATGGGCTGGGAGGCCTCCTCCGAGCGGATGTACCCCGAGGACGGCGCCCTGAAGGGCGAGATCAAGC
AGAGGCTGAAGCTGAAGGACGGCGGCCACTACGACGCTGAGGTCAAGACCACCTACAAGGCCAAGAAGCCCGTGC
AGTCCCGCGCCCTACAACGTCAACATCAAGTTGGACATCACCTCCACAACGAGGACTACACCATCGTGAACA
GTACGAAACGGCGAGGGCCGCACTCCACGGCGCATGGACGAGCTGTACAAGTAATAGACAGTGGGGTTGTG
GTTGGTTGGTATAGAATGGTGGGAATTCAAATTTTTCTA-3'

C HR template PCR amplification primers

- FWD: 5'-ATGGAGGCTTTGGCTCAA-3'
- RVS: 5'-ATCTTTTAACTTAAGGG-3'

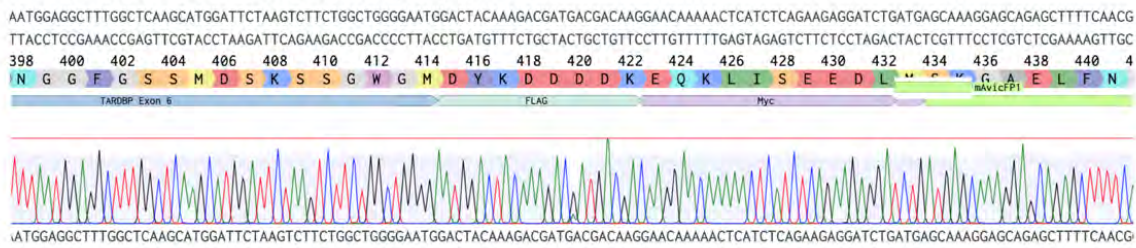
Supplementary Fig. S1 (relating to Fig. 1): *TARDBP* C-terminal fluorescent tag homologous recombination (HR) DNA templates for CRISPR/Cas9 homology-directed repair. (A) *mAvicFP1* (green) and (B) *mCherry* (red) tag sequences, preceded by *FLAG* (blue) and *MYC* (purple) linkers, flanked by homology arms (grey). (C) PCR primers for amplification of HR template DNA fragments.



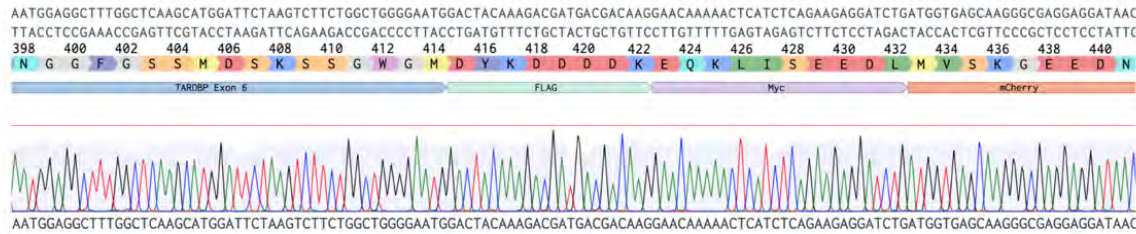
D
$$\text{TARDBP editing ratio} = \frac{\text{'Tagged' TDP-43 (72/74 kDa)}}{\text{'Tagged' + 'Untagged' TDP-43 (43 kDa)}}$$

Supplementary Fig. S2 (relating to Fig. 2): Quantification of *TARDBP* 'editing ratios' from top candidate CRISPR-tagged endogenous TDP-43 monoclonal HEK293 cell lines. Densitometry analysis for (i) 'untagged' TDP-43 (43 kDa) band, or (ii) 'CRISPR-tagged' TDP-43 (72-74 kDa) normalised to total protein signal for monoclonal (A) *TARDBP-mAvicFP1*, (B) *TARDBP-mCherry*, or (C) *TARDBP-mAv/-mCh* cell lines. (D) Formula for calculation of *TARDBP* editing ratios.

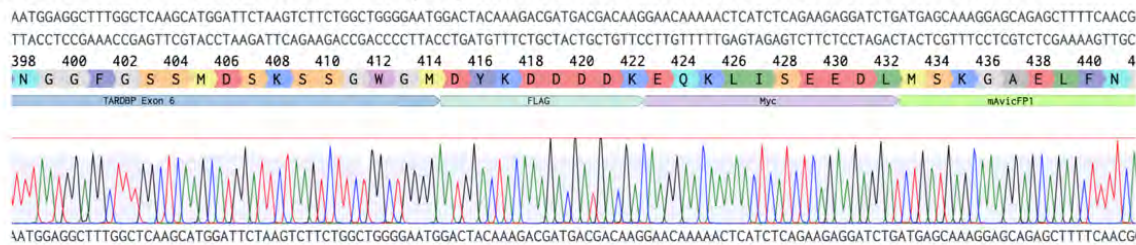
A HEK293: *TARDBP-mAvicFP1(H8)* cells



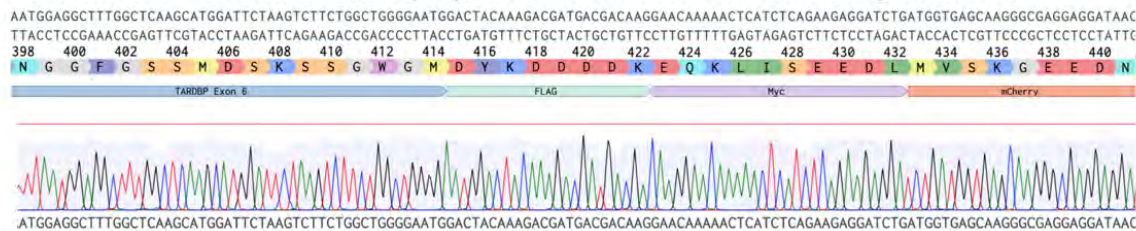
B HEK293: *TARDBP-mCherry (B8)* cells



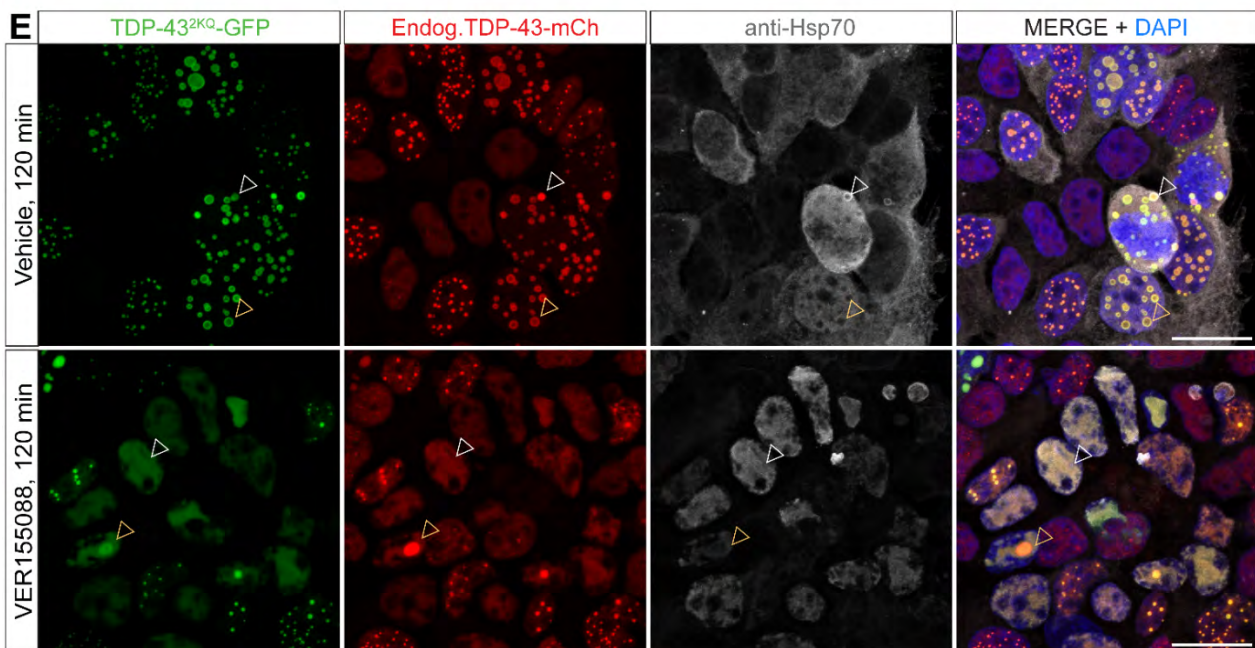
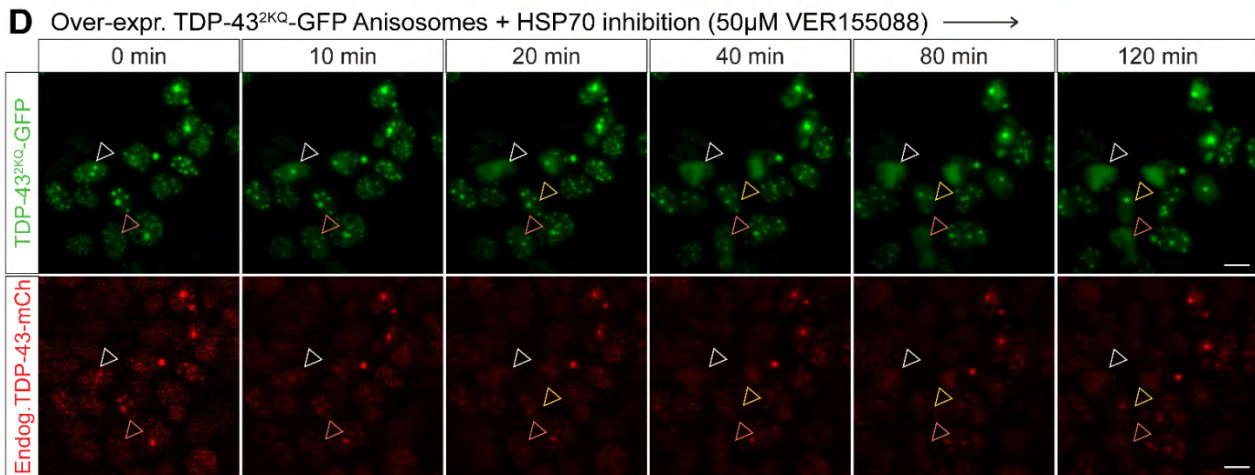
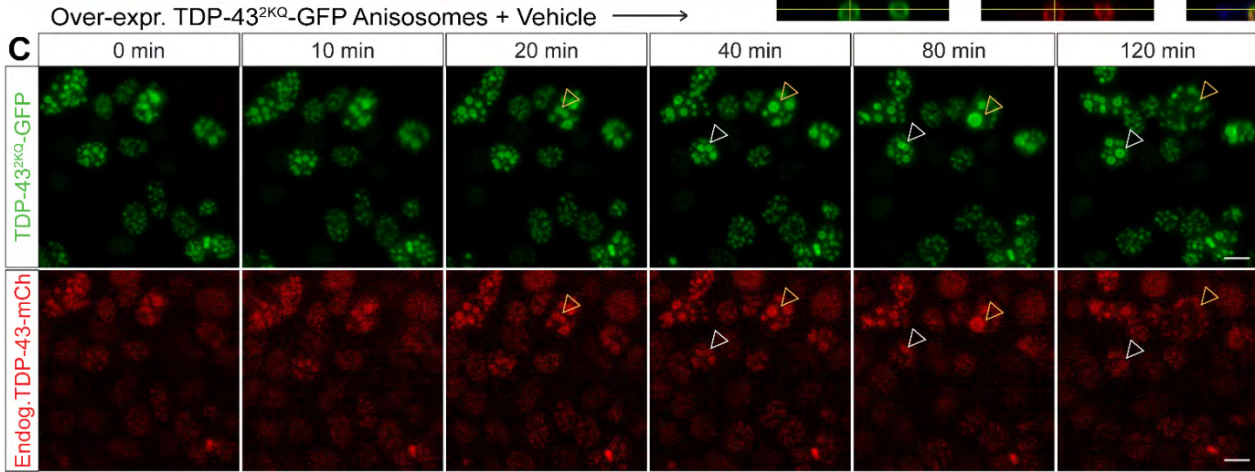
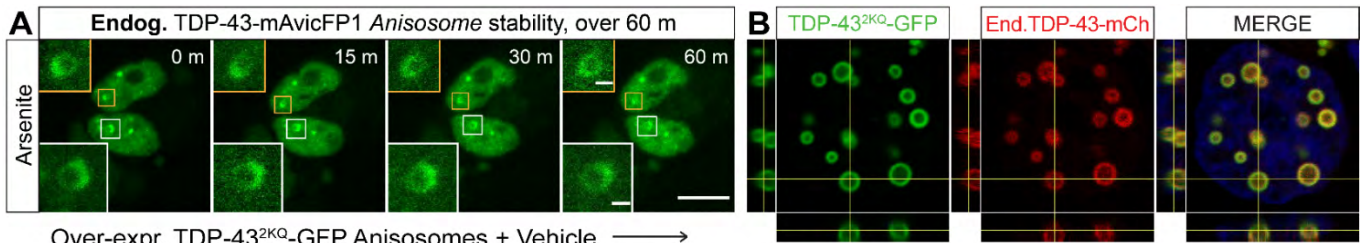
C HEK293: *TARDBP-mAv/-mCh (C4)* double knock-in cells - *mAvicFP1* insertion



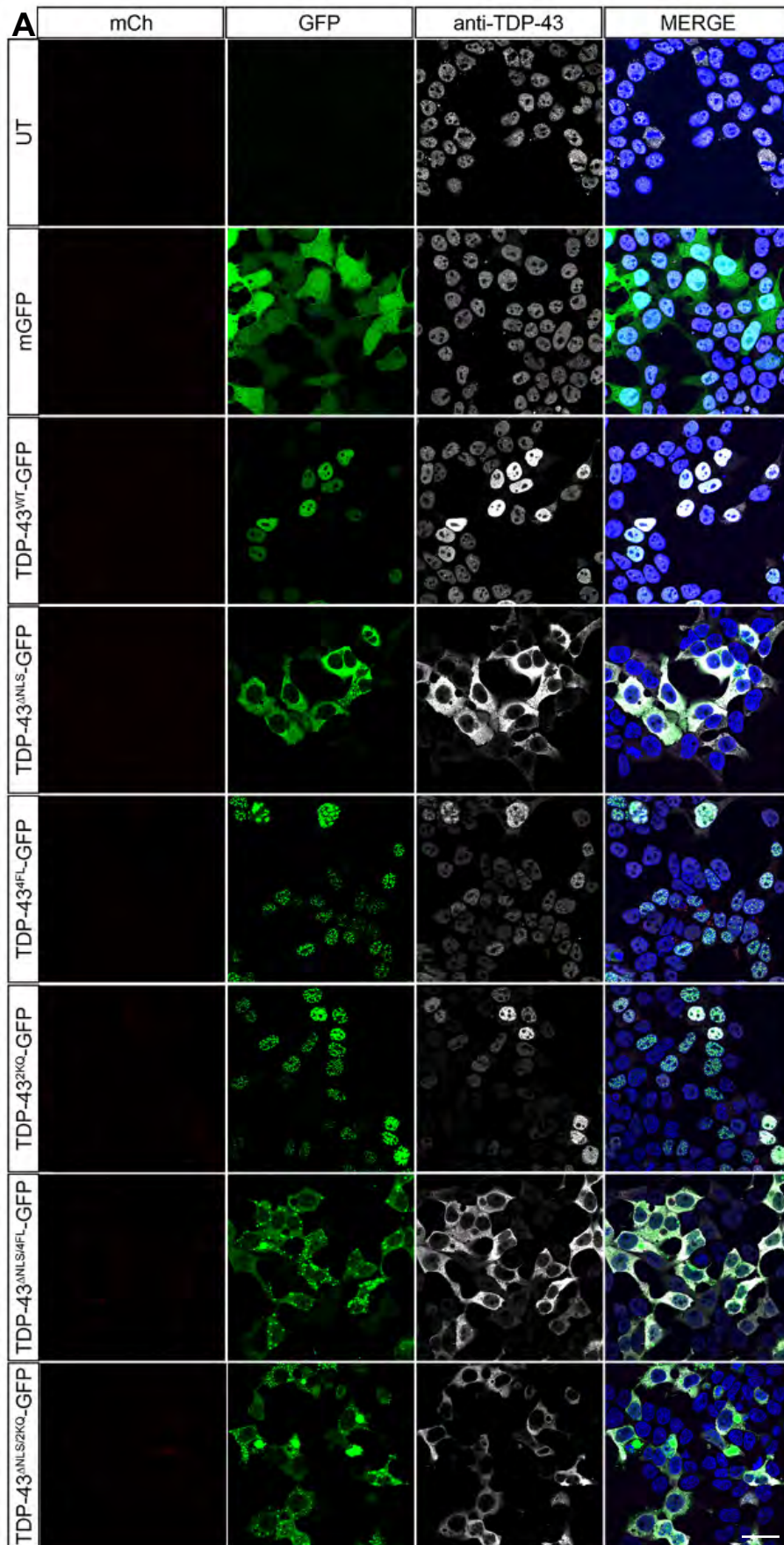
D HEK293: *TARDBP-mAv/-mCh (C4)* double knock-in cells - *mCherry* insertion



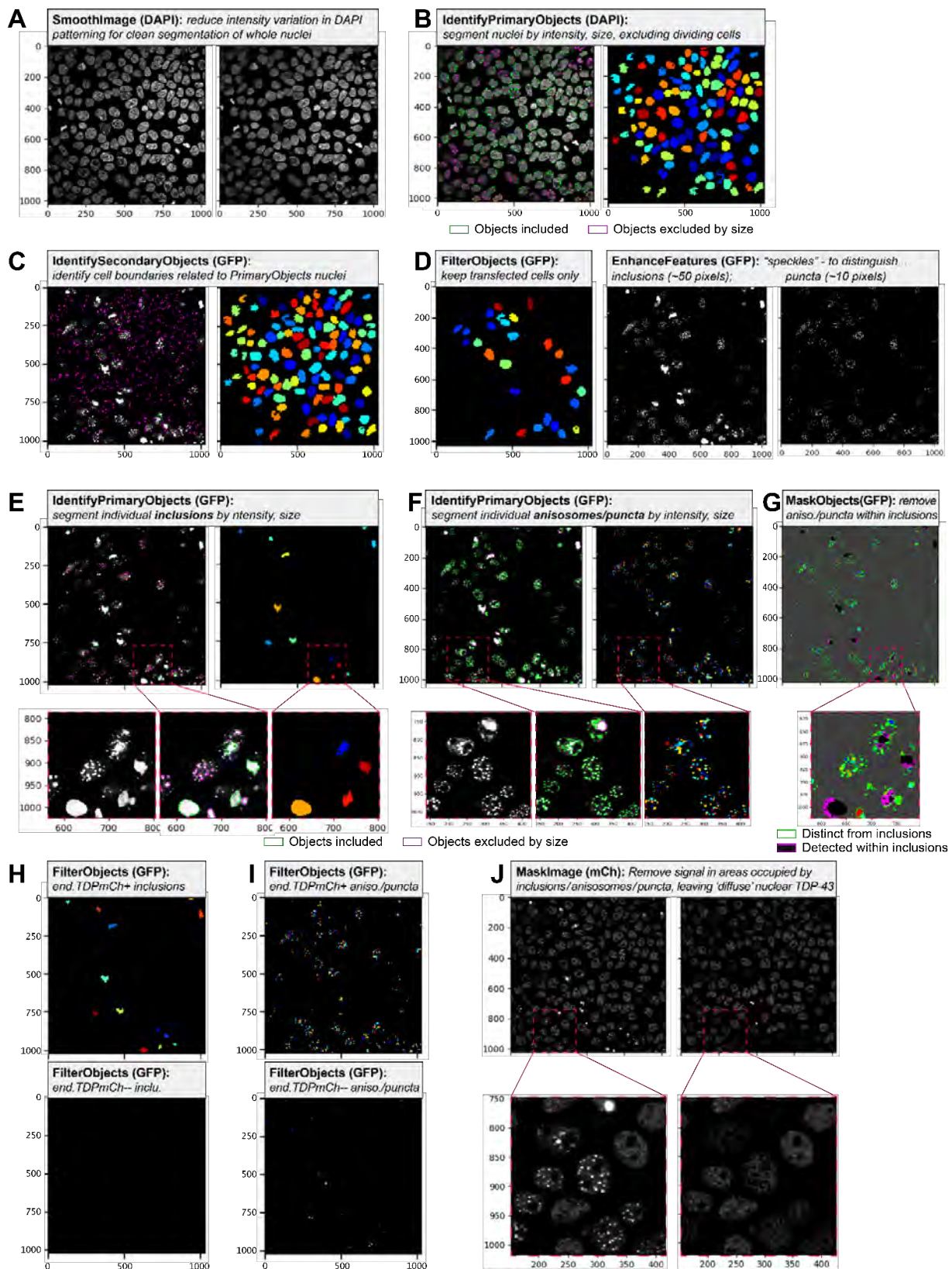
Supplementary Fig. S3 (relation to Fig. 3): Sanger sequencing indicates precise and complete homologous recombination of *mAvicFP1* or *mCherry* sequences at desired *TARDBP* C-terminal insertion sites. DNA sequences spanning *TARDBP* C-terminus, insertion site, and partially into *mAvicFP1* or *mCherry* fluorescent tags were PCR-amplified from genomic DNA extracted from top candidate CRISPR-edited (A) *TARDBP-mAvicFP1(H8)*, (B) *TARDBP-mCherry(B8)*, or (C,D) *TARDBP-mAv-mCh(C4)* monoclonal HEK293 cell lines. Top and bottom lines represent sequences amplified from forward and reverse primers, respectively. Sequence alignments performed with *Benchling*.



Supplementary Fig. S4 (relating to Fig. 4): Verification of HSP70-dependent formation anisosomes comprising endogenous under oxidative stress or with acetylation-mimicking mutation. (A) Arsenite-induced endogenous TDP-43-mAvicFP1 anisosomes under live-cell imaging were stable throughout treatment, over 60 min (scale: large panels = 10 μm , zoomed inset panels = 1 μm). **(B)** 3-dimensional orthogonal projection from confocal Z-stack images of HEK293:*TARDBP-mCh(B8)* cells demonstrates spherical shell-like structure of anisosomes formed by acetylation-mimicking mutant TDP-43^{2KQ}-GFP, which readily sequester endogenous TDP-43-mCherry. **(C)** Live-cell timelapse imaging demonstrating TDP-43^{2KQ}-GFP anisosome formation, fusion (white arrowheads), fission (yellow arrowheads) and endogenous TDP-43-mCherry recruitment under vehicle treatment for 120 min. **(D)** Timelapse of Hsp70-dependent disassembly of anisosomes (colour-coded arrowheads) following treatment with Hsp70 inhibitor, 50 μM VER155088 (scale = 10 μm). **(E)** Confocal Z-stack maximum projection of vehicle- (top panels) or Hsp70 inhibitor-treated (bottom panels) cells immunostained with anti-Hsp70 antibody, which could not penetrate anisosomes (arrowheads) to detect Hsp70 within the core of these assemblies, using a 1% Triton X-100 permeabilisation protocol. Hsp70-dependent anisosome disassembly results in mostly diffuse TDP-43 (white arrowheads), or can leave large condensed nuclear structures that still sequester endogenous TDP-43 (yellow arrowheads, scale = 20 μm).

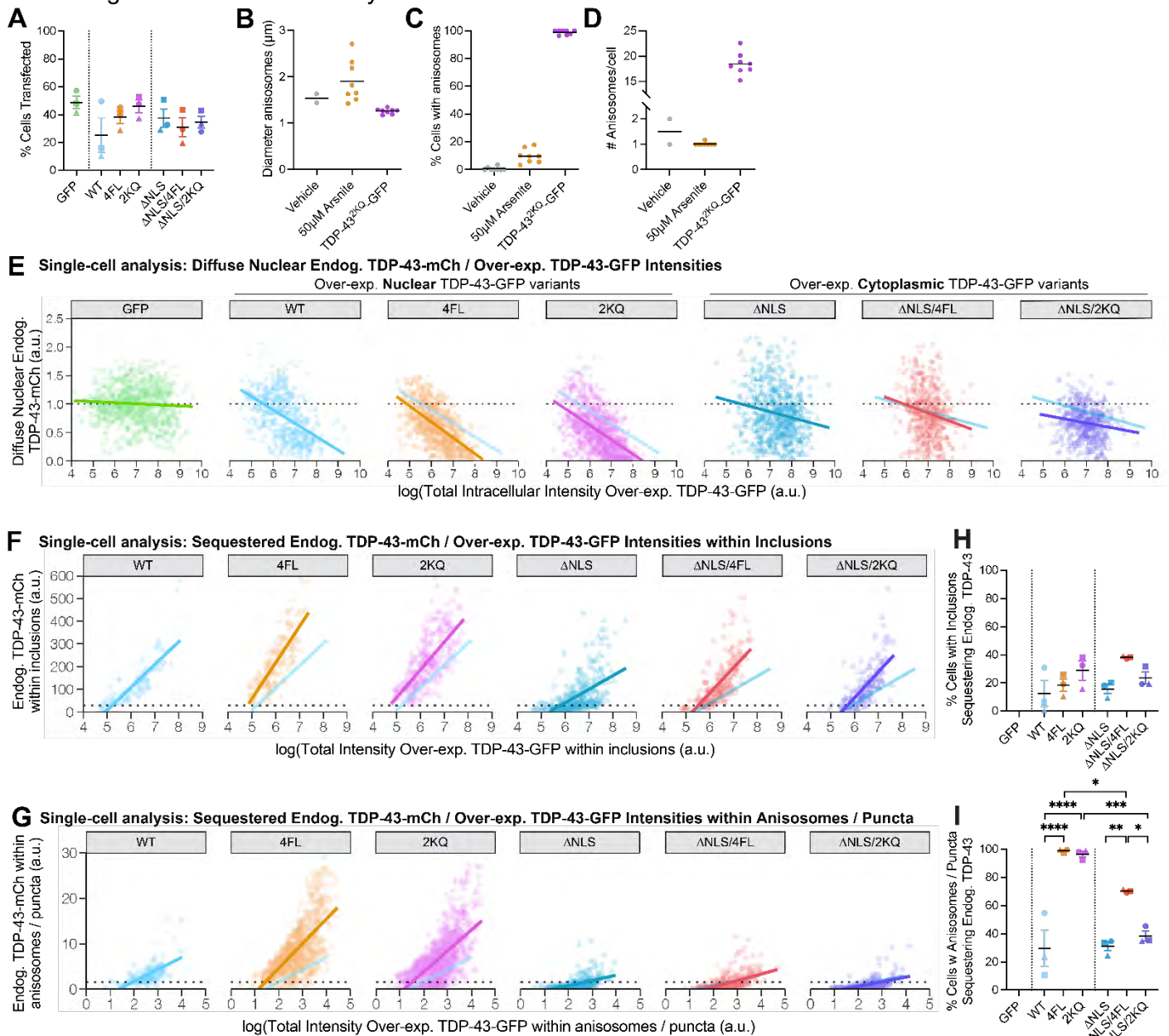


Supplementary Fig. S5 (relating to Fig. 5): Bleed-through control images of TDP-43-mCh-negative parental HEK293 cells over-expressing TDP-43-GFP variants. (A) Confocal imaging verified no bleed-through fluorescence of GFP signal into mCherry channel, used for image analysis thresholding of endogenous TDP-43-mCherry sequestration. Anti-TDP-43 antibody immunostaining is directed towards over-expressed TDP-43-GFP proteins, and does not penetrate inclusion structures (scale = 20 μ m).



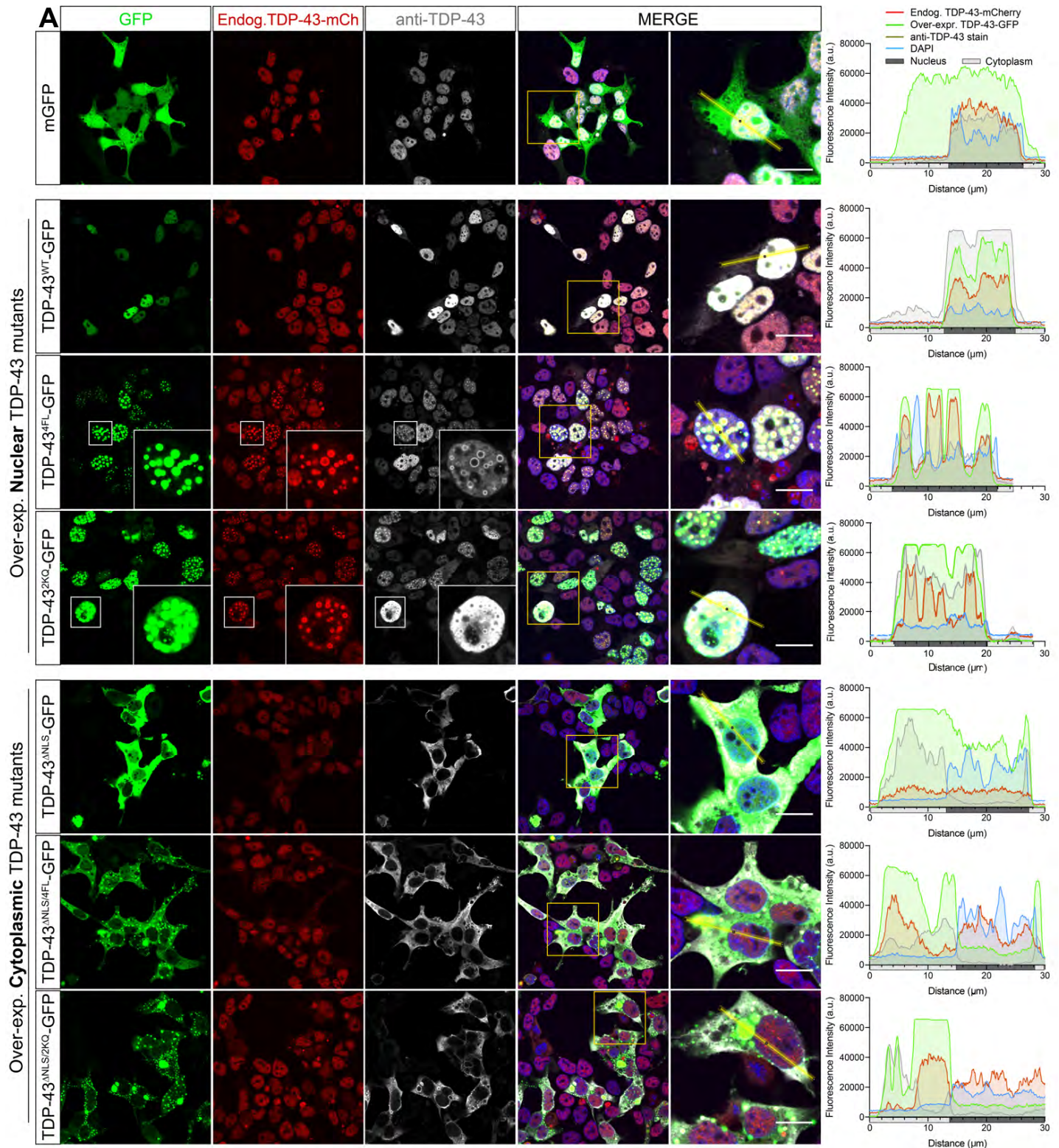
Supplementary Fig. S6 (relating to Fig. 5 & Supplementary Fig. S7): CellProfiler pipeline for unbiased and automated segmentation of transfected cells, TDP-43-GFP assemblies, and intensity/morphology measurements. (A) *SmoothImage* module over DAPI images for (B) *IdentifyPrimaryObjects* module to segment whole nuclei, filtered based on mean intensity and size to remove condensed nuclei of dividing cells. (C) *IdentifySecondaryObjects* to segment cell boundaries, anchored to primary nuclei. (D) Transfected cells filtered based on median GFP intensity. *EnhanceFeatures* module applied to GFP images to distinguish speckles of ~50 or ~10 pixels, representing 'inclusions' or 'anisomes'/puncta' respectively, segmented by (E,F) *IdentifyPrimaryObjects* modules, wherein inclusions were defined as large bright speckles with intensity above 60% of saturation point and minimum diameter of 4 μ m. Anisomes/puncta were defined as speckles with intensity above 25% of saturation point and diameter of 0.5-4 μ m. (G) *MaskObjects* module to remove smaller assemblies falsely detected by segmentation of larger

inclusions. **(H)** *FilterObjects* modules to identify endogenous TDP-43-mCherry-positive or -negative inclusions or **(I)** puncta. **(J)** *MaskImage* module to distinguish sequestered from free diffuse nuclear endogenous TDP-43-mCherry.

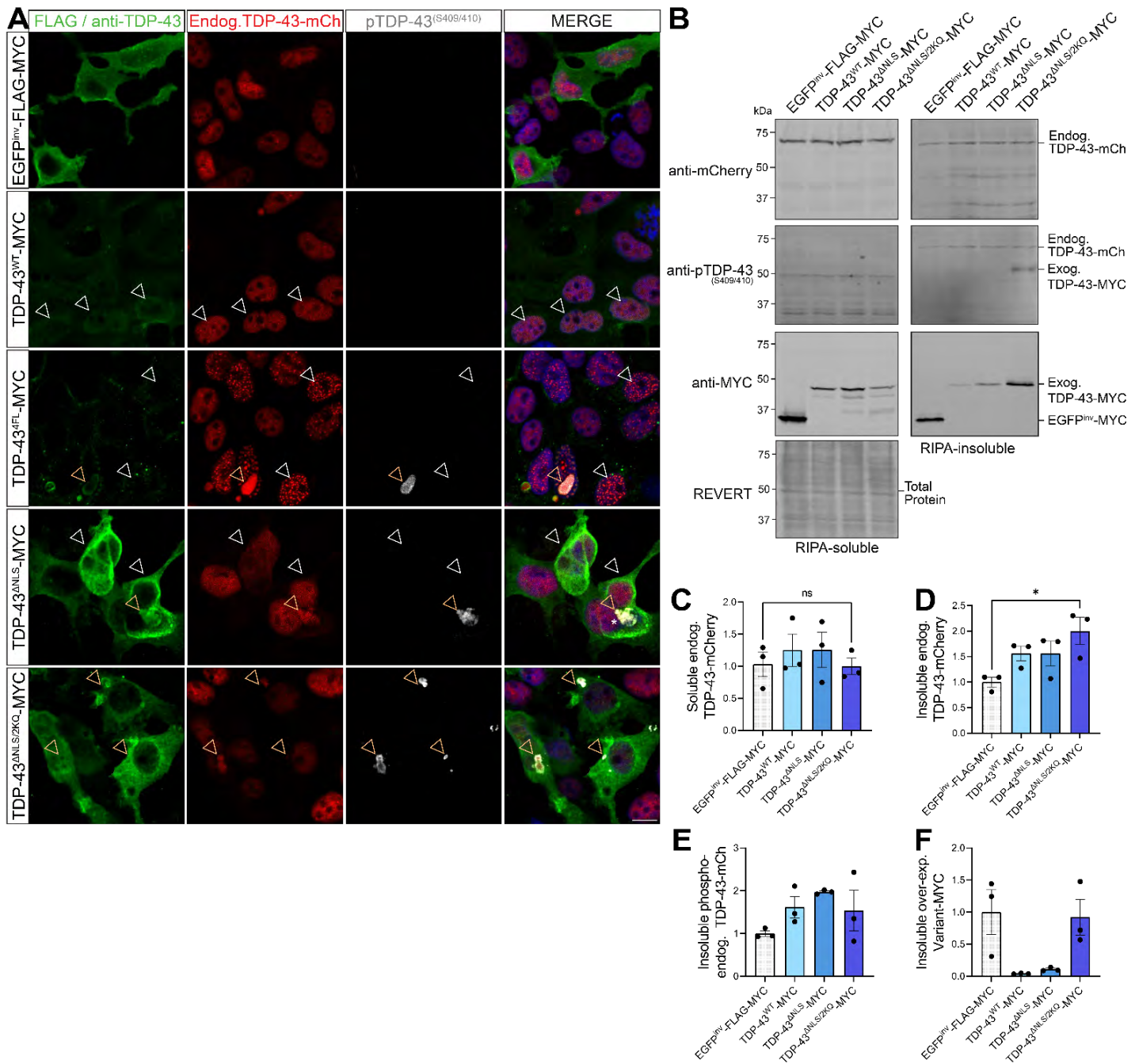


Supplementary Fig. S7 (relating to Fig. 5): Single-cell quantitative image analysis reveals concentration-dependent effect of mutant TDP-43-GFP of assemblies on sequestration and depletion of free nuclear endogenous TDP-43-mCherry. **(A)** Transfection rate for exogenous TDP-43-GFP variants measured by CellProfiler image segmentation analysis. **(B)** Diameter of anisomes, **(C)** % of cells with anisomes, and **(D)** number of anisomes per cell formed by TDP-43^{2KQ}-GFP compared to arsenite-induced endogenous TDP-43 anisomes. **(B-D)** Data points represent mean from 8 images from a single biological replicate. **(E)** Scatterplots illustrating concentration-dependent effects of over-expressed TDP-43-GFP protein levels (total intensity) on median intensity of diffuse nuclear endogenous TDP-43-mCherry (normalised to GFP-transfected controls), showing individual transfected cells. **(F)** Total intensity of endogenous TDP-43-mCherry sequestered to inclusions, or **(G)** anisomes/puncta, against total intensity of TDP-43-GFP assemblies per cell. **(E-G)** Linear regressions demonstrate concentration-dependent effects of TDP-43-GFP over-expression on endogenous TDP-43 nuclear depletion or sequestration. Comparison of WT (light blue) or ΔNLS (teal) regression lines overlaid on nuclear or cytoplasmic mutants respectively demonstrates amplified effects of 4FL and 2KQ mutations on depletion and sequestration. Data points represent individual cells segmented with CellProfiler, sampled from 8 imaging regions from each of $n = 3$ biologically independent repeats. **(H)** % Cells with inclusions or **(I)** anisomes/puncta sequestering endogenous TDP-43-mCherry. **(A,H,I)** Individual points represent mean values from 8 imaging regions of interest for each of $n = 3$ biologically independent

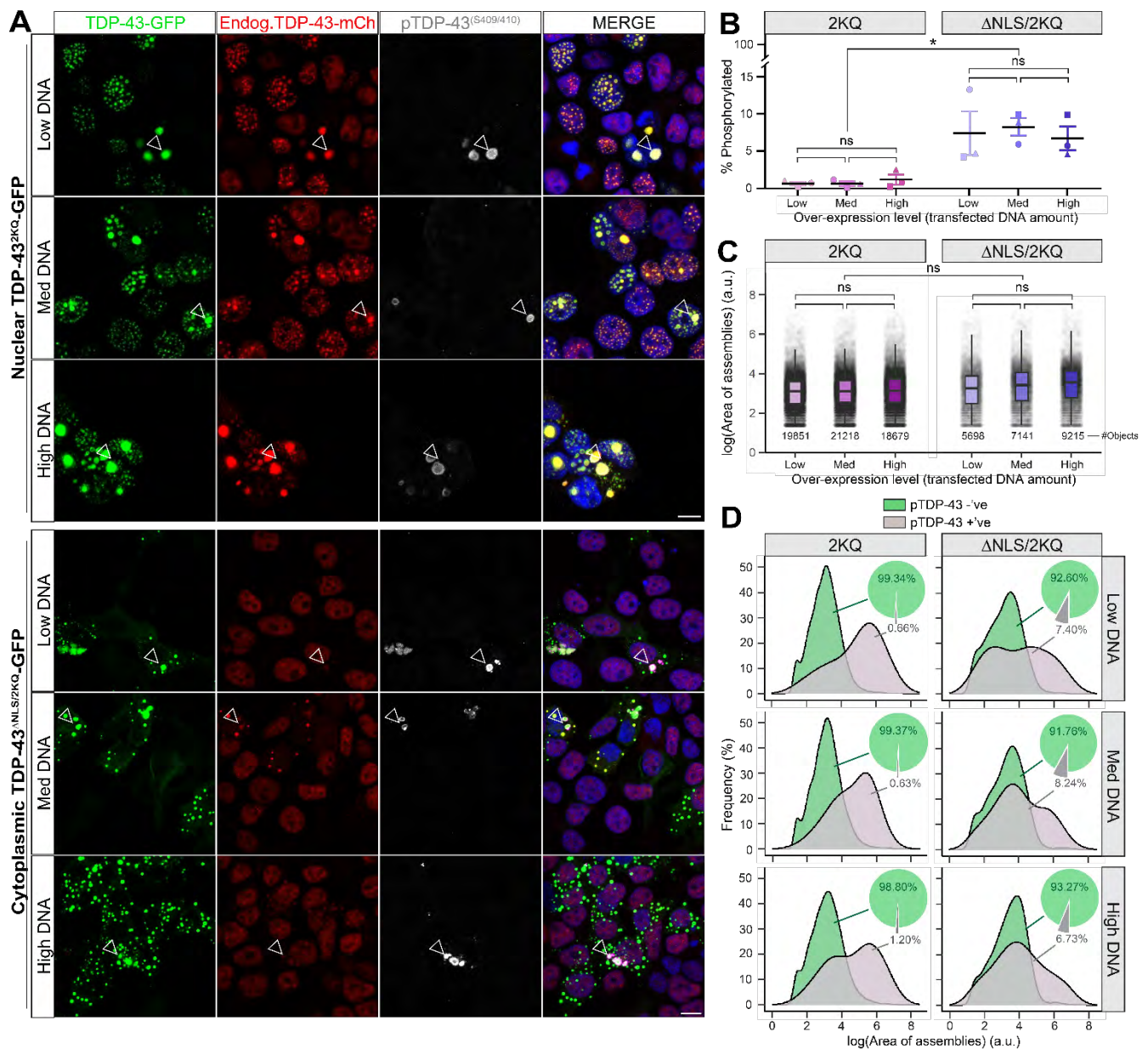
repeats. Black lines represent combined mean values, analysed by ordinary one-way ANOVA (* = $P < 0.05$; ** = $P < 0.01$; *** = $P < 0.001$; **** = $P < 0.0001$).



Supplementary Fig. S8 (relating to Fig. 5): Immunostaining with anti-TDP-43 antibody preferentially binds over-expressed TDP-43-GFP variants, necessitating CRISPR-tagging for visualising endogenous TDP-43 separately from pathological forms. (A) Confocal images depict the formation of exogenous TDP-43-GFP structures in the nucleus or cytoplasm and relative localisation of endogenous TDP-43-mCherry and anti-TDP-43 (scale = 10 μ m). Areas of colocalization indicated by intensity profiles across regions marked by yellow line in right-hand magnified view panels. Anisosomes formed by 4FL and 2KQ mutant TDP-43 variants are highlighted with additional magnified inset panels, demonstrating that anti-TDP-43 antibody only labels anisosomes peripherally to TDP-43-GFP or endogenous TDP-43-mCh signals, and may not penetrate such dynamic but condensed assemblies.



Supplementary Fig. S9 (relating to Fig. 6): Over-expression of aggregation-prone mutant TDP-43 variants lacking fluorescent tags replicates sequestration and insolubility of endogenous TDP-43 driven by TDP-43-GFP variants. (A) Confocal images of HEK293: *TARDBP-mCherry(B8)* cells over-expressing TDP-43-MYC variants (white arrowheads) and assemblies that are phosphorylated and sequester endogenous TDP-43-mCherry (yellow arrowheads). **(B)** Immunoblot of RIPA-soluble and -insoluble protein fractions for mCherry, pTDP-43^{409/410}, MYC, and REVERT Total Protein. **(C)** Densitometry analysis for soluble endogenous TDP-43-mCherry, **(D)** insoluble endogenous TDP-43-mCherry, **(E)** insoluble phospho-endogenous TDP-43-mCherry, **(F)** insoluble over-expressed TDP-43-MYC variant. Data points represent means of $n = 3$ biologically independent repeats, analysed by ordinary one-way ANOVA (* = $P < 0.05$).



Supplementary Fig. S10 (relating to Fig. 6): Morphological characteristics and phosphorylation of individual mutant TDP-43-GFP assemblies, and their effects on endogenous TDP-43 sequestration, are dependent on the mutant construct and not level of over-expression. (A) Maximum intensity projection from confocal Z-stack images displaying TDP-43-GFP assemblies formed by different levels of over-expression in HEK293: *TARDBP-mCh(B8)* cells, immunostained with anti-pTDP-43^{409/410} antibody. White arrowheads indicate phosphorylated assemblies. Transfected DNA amounts “Low”, “Med”, and “High” equate with 0.25, 0.5, or 1 μ g DNA per well in a 24-well plate. (B) Percentage of 2KQ or Δ NLS/2KQ mutant assemblies containing phospho-TDP-43, for each over-expression level. Black lines represent mean values for 8 imaging regions of interest, from $n = 3$ biologically independent repeats, shown as individual points. Data analysed by ordinary one-way ANOVA ($* = P < 0.05$). (C) Boxplot distribution of log-scaled area of assemblies formed by 2KQ or Δ NLS/2KQ mutants. Each point represents individual assemblies totalling from each of $n = 3$ biologically independent repeats (number per group indicated by text below). Statistical analysis completed by ordinary one-way ANOVA, comparing overall mean area of assemblies per group, taken from individual replicate means ($ns = P > 0.05$). (D) Frequency distributions of log-scaled area of pTDP-43-positive (grey) or pTDP-43-negative (green) assemblies, for each over-expression level. Pie charts indicate percentage of all assemblies categorised into phosphorylation states.

Other Supplementary Materials

Movie1 (relating to Figure 3)

Representative FRAP acquisition of diffuse nuclear protein under physiological conditions, for endogenous TDP-43-mAvicFP1, over-expressed TDP-43^{WT}-GFP, endogenous TDP-43-mCherry, and over-expressed TDP-43^{WT}-mCherry; scale = 5 μ m.

Movie2 (relating to Figure 4)

Representative FRAP acquisition of dynamic TDP-43-mAvicFP1 protein assemblies, including physiological puncta without treatment, arsenite-induced puncta or arsenite-induced anisosomes; scale = 5 μ m.

Movie3 (relating to Figure 6)

Representative FRAP acquisition of acetylated TDP-43 assemblies sequestering endogenous TDP-43-mCherry, including cytoplasmic TDP-43 ^{Δ NLS/2KQ}-GFP inclusions (partial or complete bleaching), nuclear TDP-43^{2KQ}-GFP inclusions, or dynamic nuclear TDP-43^{2KQ}-GFP anisosomes; scale = 5 μ m.

Table1 (relating to Figure 3, 4, 6)

Fluorescence recovery after photobleaching raw and normalised data for maximal recovery, time-to-half recovery, area-under-the-curve, and mobility fraction analyses.

Table2 (relating to Figure 5)

CellProfiler unbiased quantitative image analysis pipeline output, values representing per-image means of all transfected cells sampled from 8 regions of interest, imaged at 63x magnification.

Table3 (relating to Supplementary Figure 6)

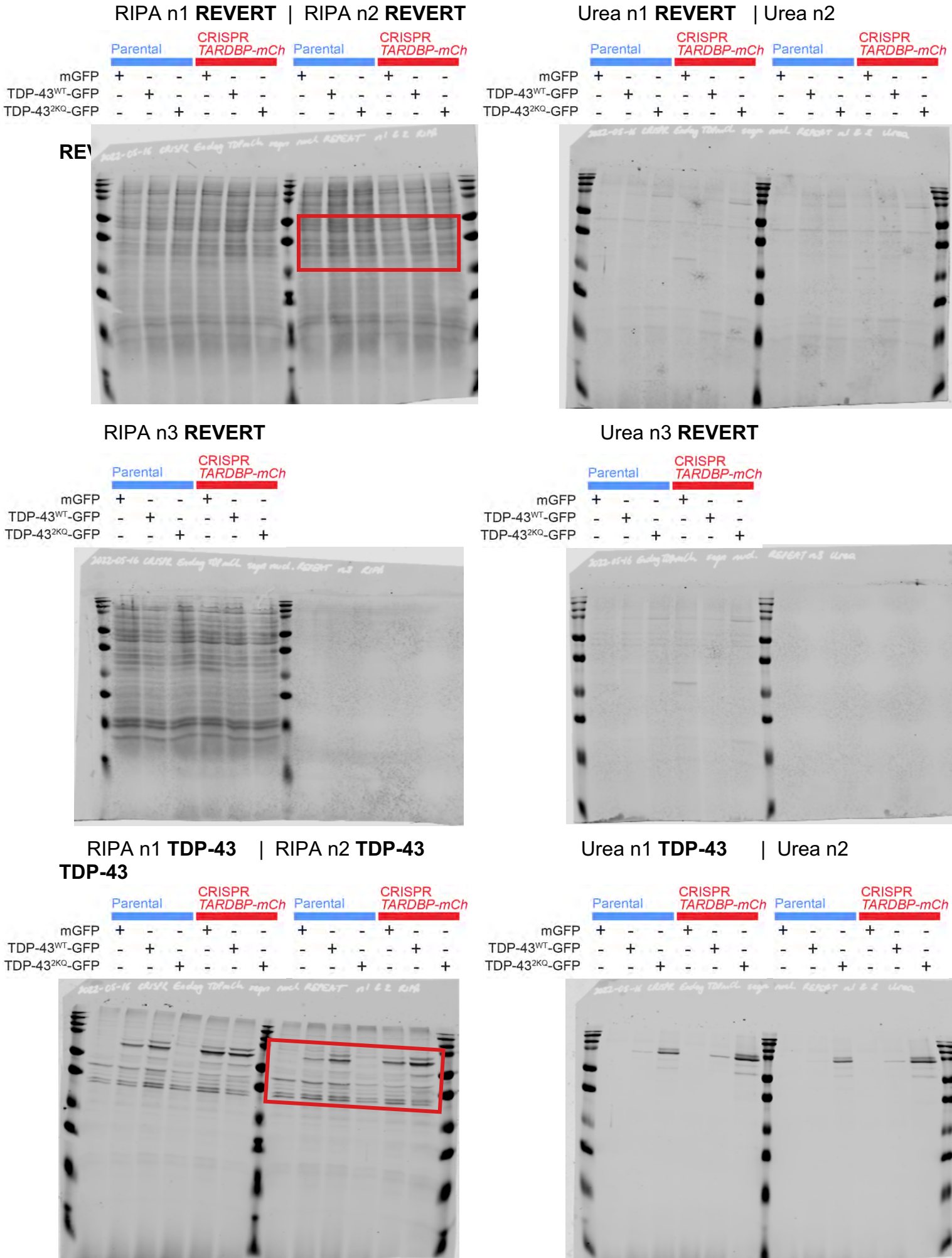
CellProfiler unbiased quantitative image analysis pipeline output, values representing per-cell values for individual transfected cells sampled from 8 regions of interest, imaged at 63x magnification.

Table4 (relating to Figure 6)

Immunoblot densitometry analysis raw and normalised values. See below immunoblotting uncropped images for reference to the representative blots and cropped regions incorporated to Figure 6.

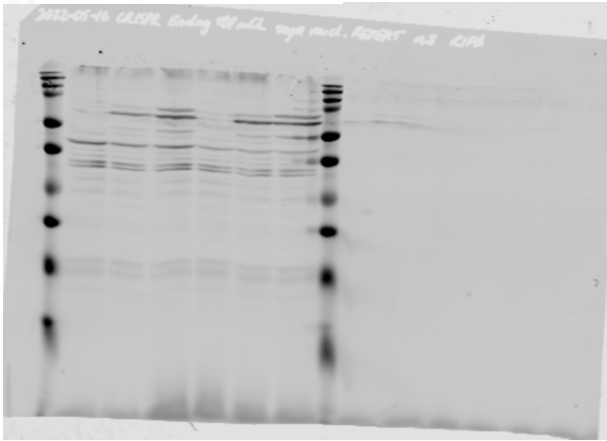
Uncropped Immunoblots

Fig. 6C: Endogenous TDP-43 is sequestered by nuclear acetylated TDP-43-GFP



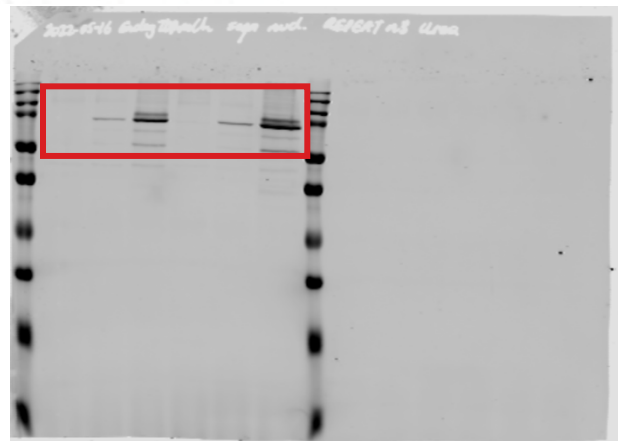
RIPA n3 TDP-43

	Parental		CRISPR TARDBP-mCh	
mGFP	+	-	+	-
TDP-43 ^{WT} -GFP	-	+	-	+
TDP-43 ^{KO} -GFP	-	-	+	-



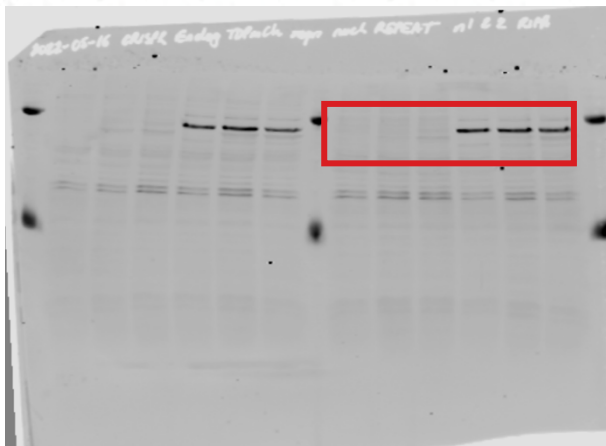
Urea n3 TDP-43

	Parental		CRISPR TARDBP-mCh	
mGFP	+	-	+	-
TDP-43 ^{WT} -GFP	-	+	-	+
TDP-43 ^{KO} -GFP	-	-	+	-



RIPA n1 MYC

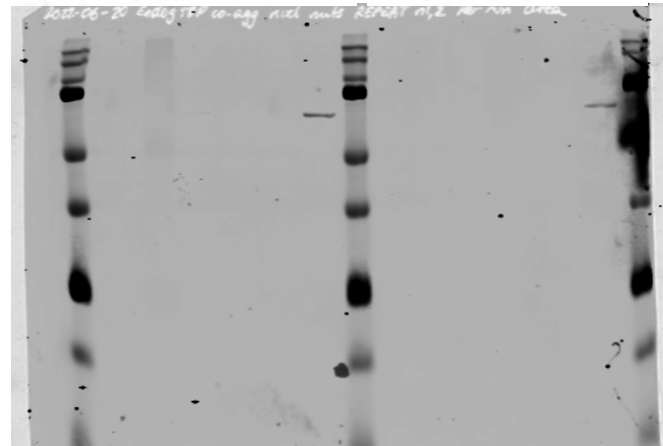
	Parental		CRISPR TARDBP-mCh		Parental		CRISPR TARDBP-mCh	
mGFP	+	-	+	-	+	-	+	-
TDP-43 ^{WT} -GFP	-	+	-	+	-	+	-	+
TDP-43 ^{KO} -GFP	-	-	+	-	-	-	+	-



RIPA n2 MYC

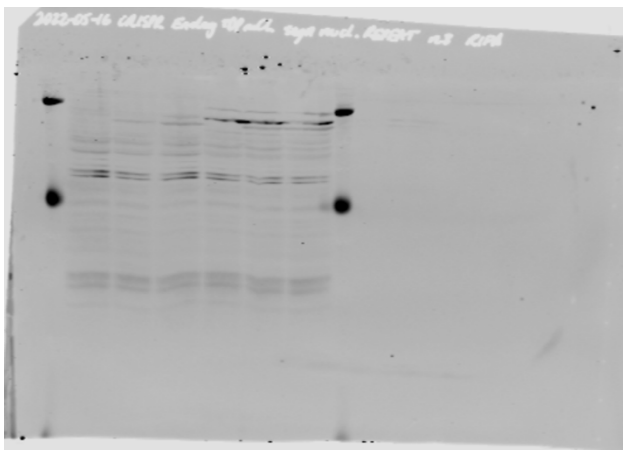
Urea n1 MYC

	Parental		CRISPR TARDBP-mCh		Parental		CRISPR TARDBP-mCh	
mGFP	+	-	+	-	+	-	+	-
TDP-43 ^{WT} -GFP	-	+	-	+	-	+	-	+
TDP-43 ^{KO} -GFP	-	-	+	-	-	-	+	-



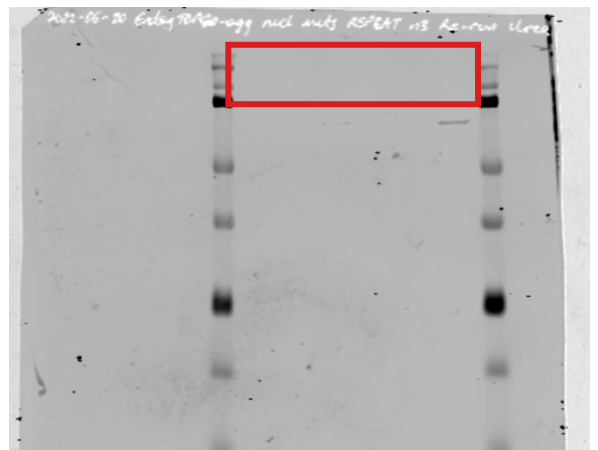
RIPA n3 MYC

	Parental		CRISPR TARDBP-mCh	
mGFP	+	-	+	-
TDP-43 ^{WT} -GFP	-	+	-	+
TDP-43 ^{KO} -GFP	-	-	+	-



Urea n3 MYC

	Parental		CRISPR TARDBP-mCh	
mGFP	+	-	+	-
TDP-43 ^{WT} -GFP	-	+	-	+
TDP-43 ^{KO} -GFP	-	-	+	-



Urea n1 pTDP

| Urea n2 pTDP

Urea n3 pTDP

	Parental				CRISPR TARDBP-mCh			
mGFP	+	-	-	+	-	-	-	+
TDP-43 ^{WT} -GFP	-	+	-	-	+	-	-	+
TDP-43 ^{2KQ} -GFP	-	-	+	-	-	+	-	+

	Parental				CRISPR TARDBP-mCh			
mGFP	+	-	-	+	-	-	-	+
TDP-43 ^{WT} -GFP	-	+	-	-	+	-	-	+
TDP-43 ^{2KQ} -GFP	-	-	+	-	-	+	-	+

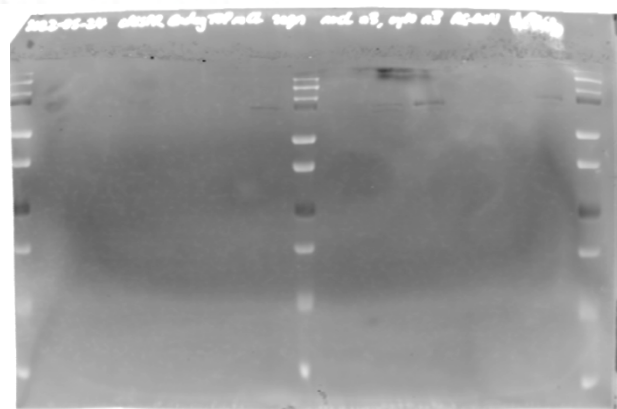


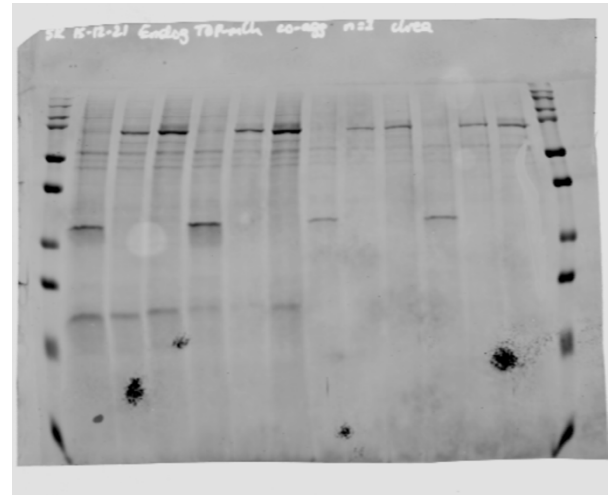
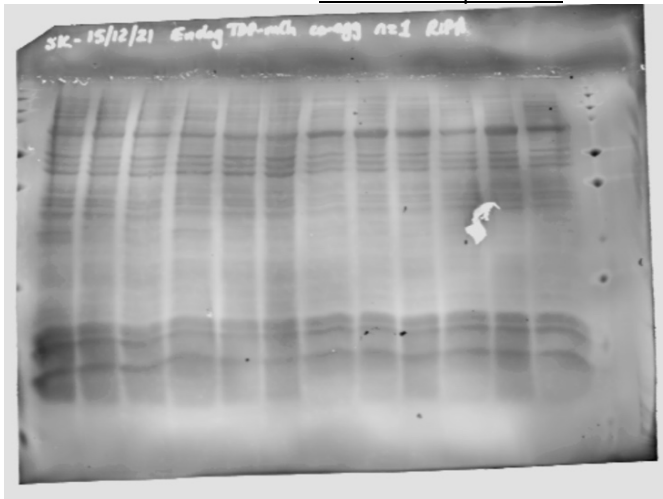
Fig. 6F: Endogenous TDP-43 is sequestered by cytoplasmic acetylated TDP-43-GFP
RIPA n1 REVERT

	Parental				CRISPR TARDBP-mCherry			
mGFP	+	-	-	+	-	-	-	+
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-	-	+
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+	-	+

Unrelated experiment

	Parental				CRISPR TARDBP-mCherry			
mGFP	+	-	-	+	-	-	-	+
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-	-	+
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+	-	+

Unrelated experiment



RIPA n2 REVERT

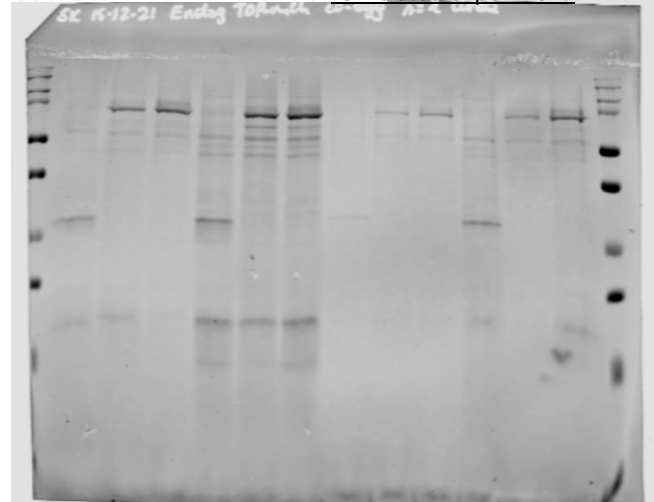
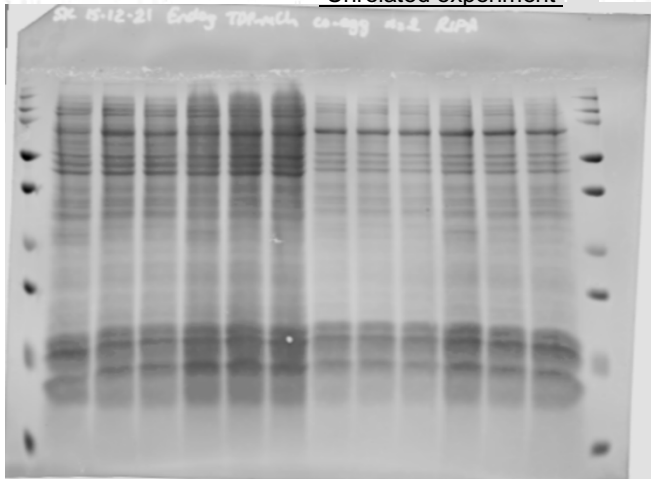
	Parental				CRISPR TARDBP-mCherry			
mGFP	+	-	-	+	-	-	-	+
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-	-	+
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+	-	+

Unrelated experiment

Urea n2 REVERT

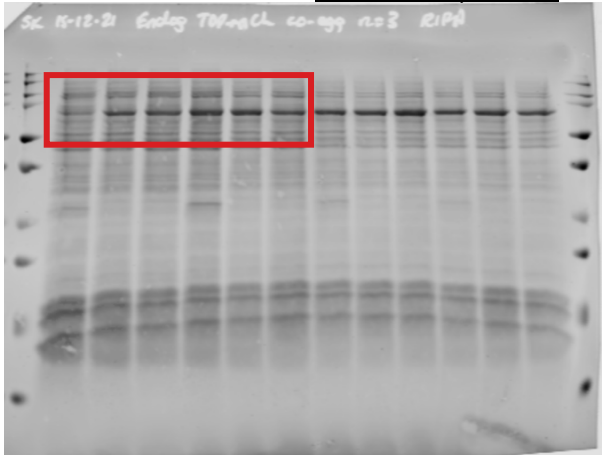
	Parental				CRISPR TARDBP-mCherry			
mGFP	+	-	-	+	-	-	-	+
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-	-	+
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+	-	+

Unrelated experiment



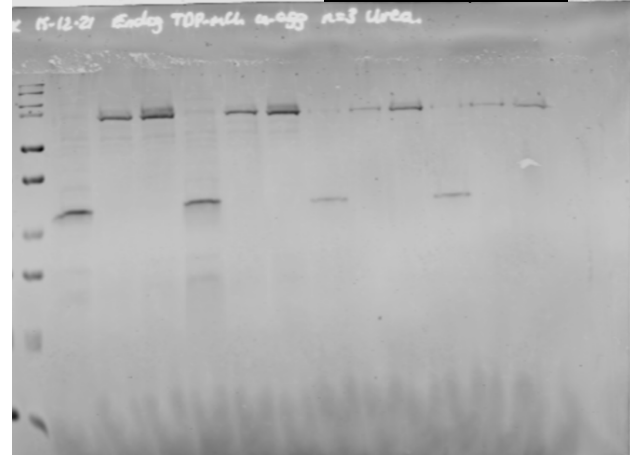
RIPA n3 REVERT

	Parental		CRISPR TARDBP-mCherry		
mGFP	+	-	+	-	
TDP-43 ^{ΔNLS} -GFP	-	+	-	+	
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	Unrelated experiment



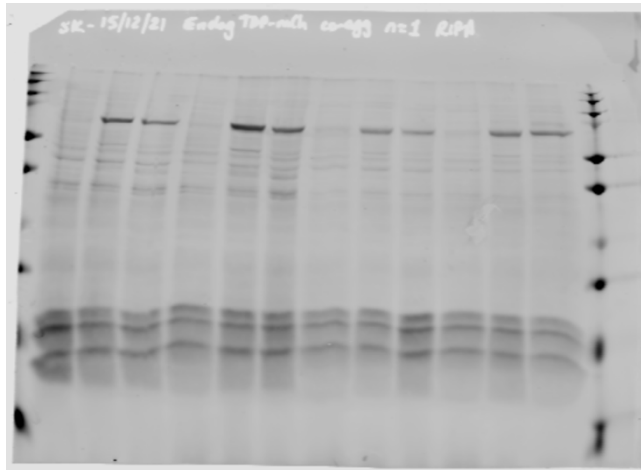
Urea n3 REVERT

	Parental		CRISPR TARDBP-mCherry		
mGFP	+	-	+	-	
TDP-43 ^{ΔNLS} -GFP	-	+	-	+	
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	Unrelated experiment



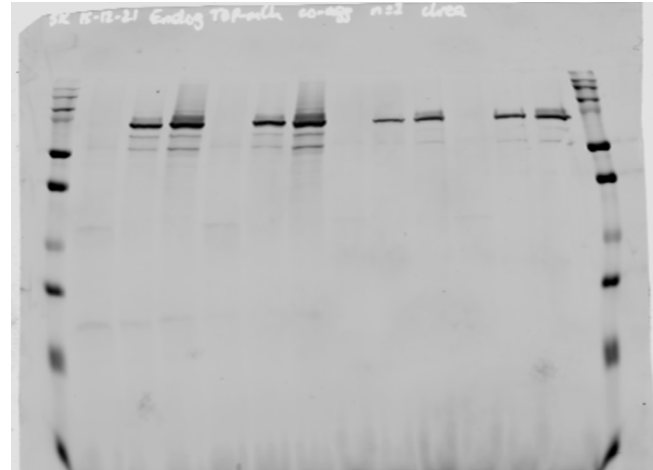
RIPA n1 TDP-43

	Parental		CRISPR TARDBP-mCherry		
mGFP	+	-	+	-	
TDP-43 ^{ΔNLS} -GFP	-	+	-	+	
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	Unrelated experiment



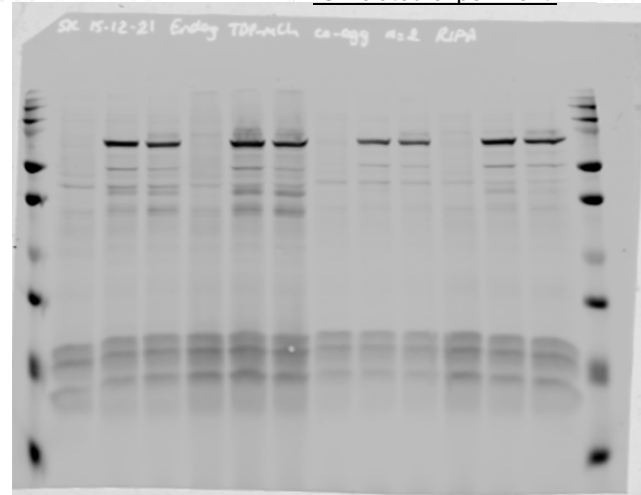
Urea n1 TDP-43

	Parental		CRISPR TARDBP-mCherry		
mGFP	+	-	+	-	
TDP-43 ^{ΔNLS} -GFP	-	+	-	+	
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	Unrelated experiment



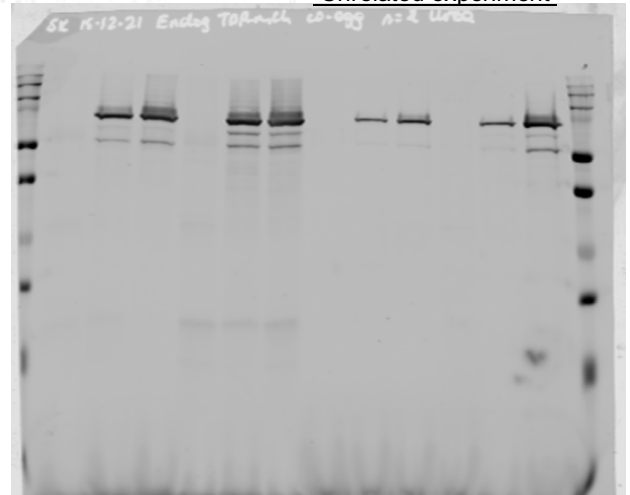
RIPA n2 total TDP-43

	Parental		CRISPR TARDBP-mCherry		
mGFP	+	-	+	-	
TDP-43 ^{ΔNLS} -GFP	-	+	-	+	
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	Unrelated experiment



Urea n2 total TDP-43

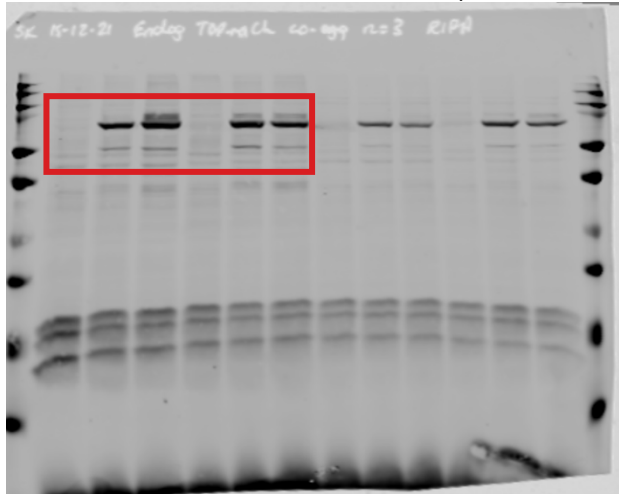
	Parental		CRISPR TARDBP-mCherry		
mGFP	+	-	+	-	
TDP-43 ^{ΔNLS} -GFP	-	+	-	+	
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	Unrelated experiment



RIPA n3 total TDP-43

	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+

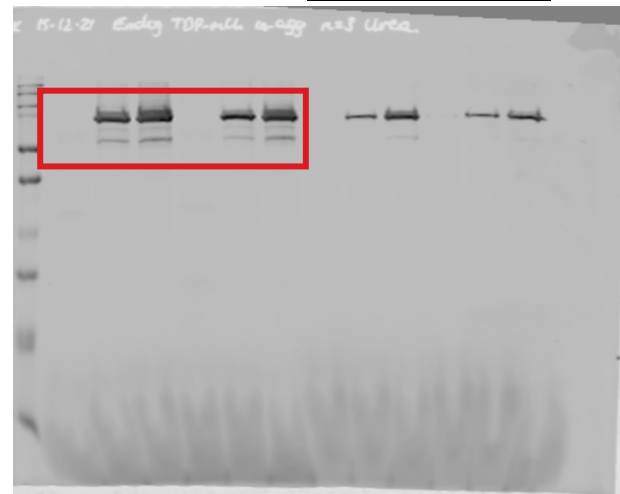
Unrelated experiment



Urea n3 total TDP-43

	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+

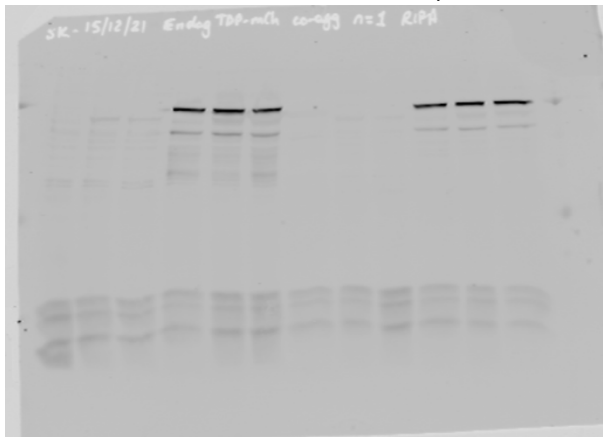
Unrelated experiment



RIPA n1 MYC

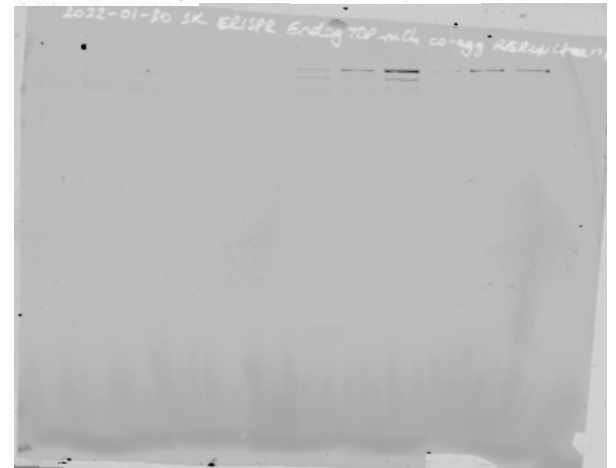
	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+

Unrelated experiment



Urea n1 MYC

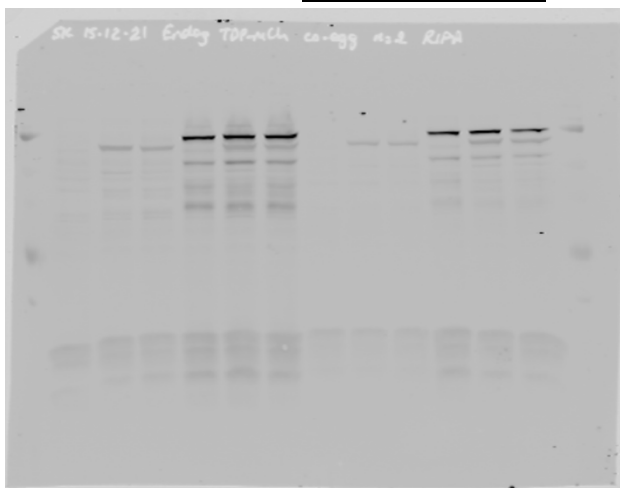
	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+



RIPA n2 MYC

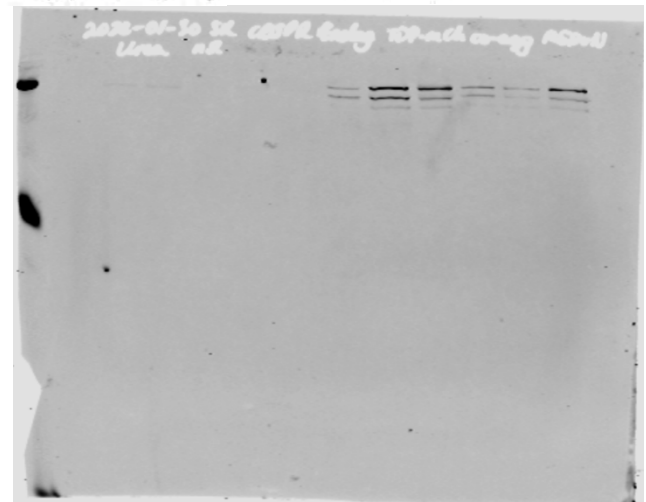
	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+

Unrelated experiment



Urea n2 MYC

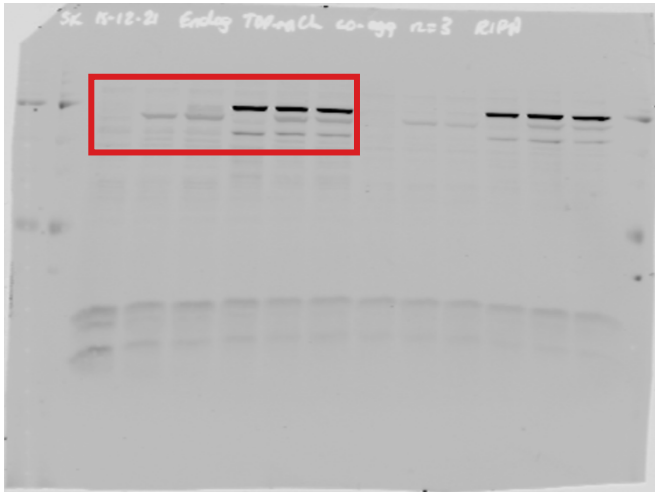
	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ANLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ANLS/2KQ} -GFP	-	-	+	-	-	+



RIPA n3 MYC

	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ΔNLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	-	+

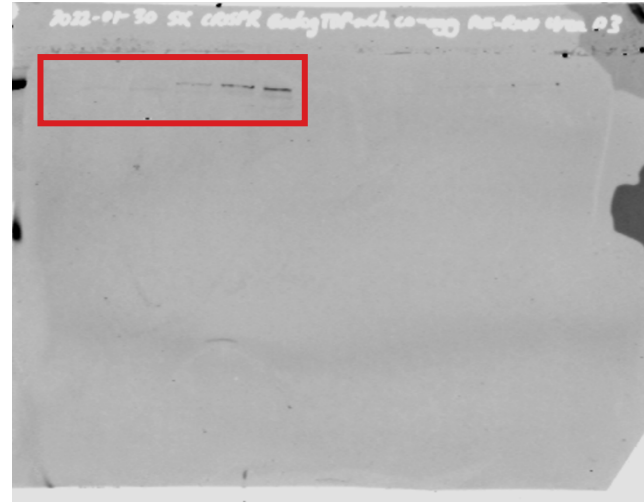
Unrelated experiment



Urea n3 MYC

	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ΔNLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	-	+

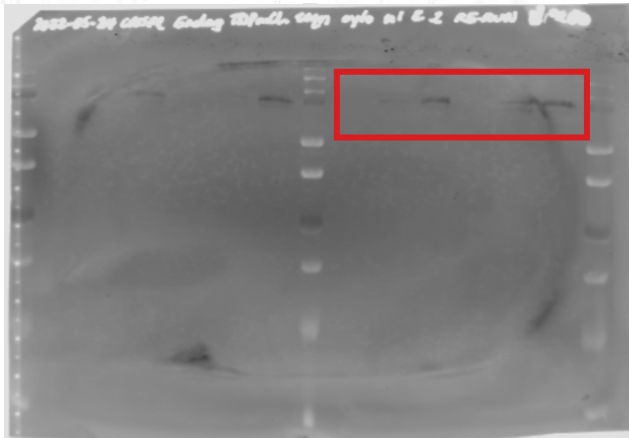
Unrelated experiment



Urea n1 pTDP

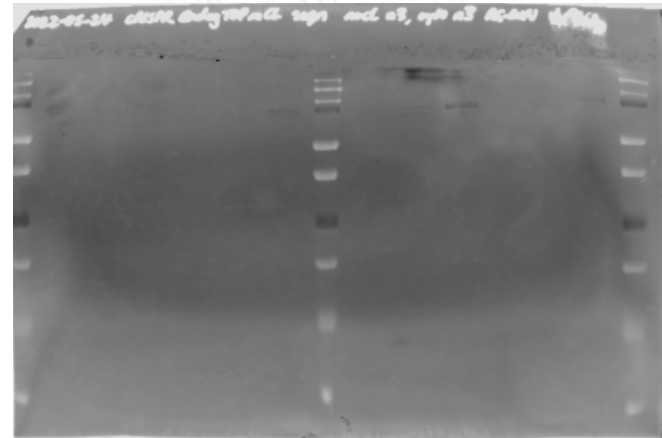
Urea n2 pTDP

	Parental			CRISPR TARDBP-mCh			Parental			CRISPR TARDBP-mCh		
mGFP	+	-	-	+	-	-	+	-	-	+	-	-
TDP-43 ^{ΔNLS} -GFP	-	+	-	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	-	+	-	-	+	-	-	+



Urea n3 pTDP

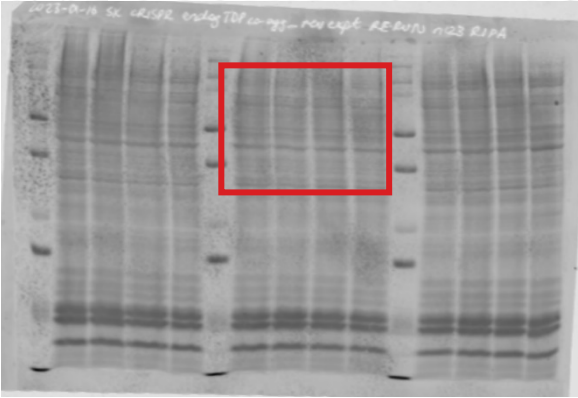
	Parental			CRISPR TARDBP-mCherry		
mGFP	+	-	-	+	-	-
TDP-43 ^{ΔNLS} -GFP	-	+	-	-	+	-
TDP-43 ^{ΔNLS/2KQ} -GFP	-	-	+	-	-	+



Supplementary Fig. S8: Endogenous TDP-43 is sequestered by cytoplasmic acetylated TDP-43

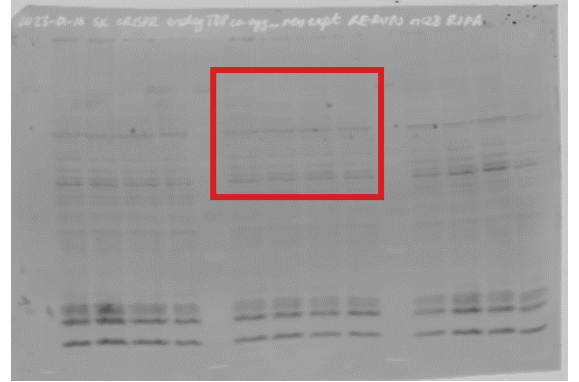
RIPA REVERT-700

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KQ} -MYC	-	-	-	-	-	+	-	-	+



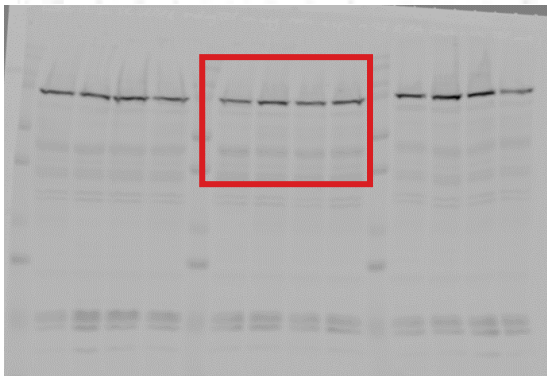
RIPA pTDP-43^(409/410)-800

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KQ} -MYC	-	-	-	-	-	+	-	-	+



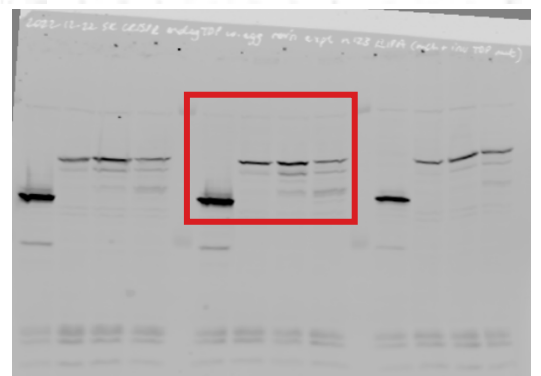
RIPA mCherry-700

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KQ} -MYC	-	-	-	-	-	+	-	-	+



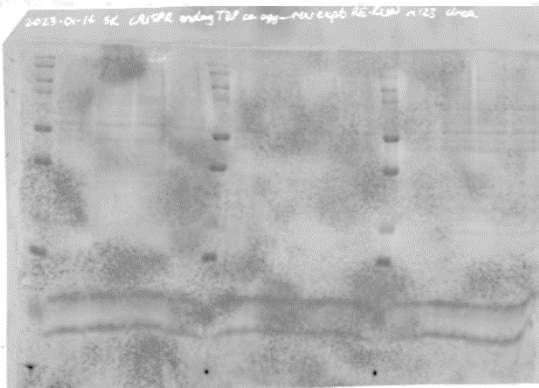
RIPA MYC-800

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KQ} -MYC	-	-	-	-	-	+	-	-	+



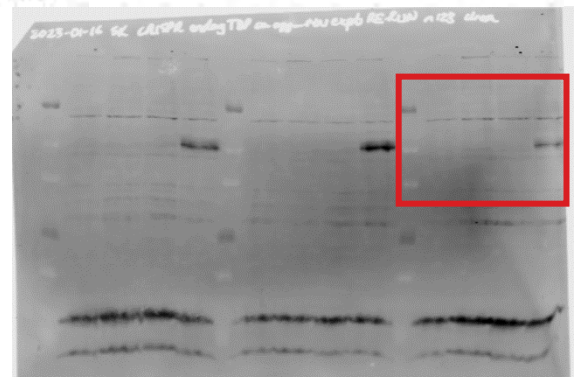
Urea REVERT-700

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KQ} -MYC	-	-	-	-	-	+	-	-	+



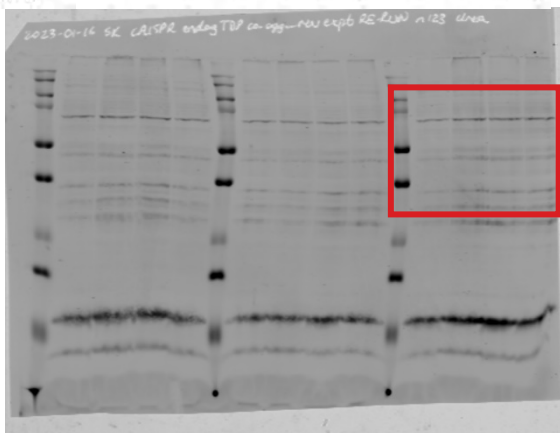
Urea pTDP-43^(409/410)-800

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KQ} -MYC	-	-	-	-	-	+	-	-	+



Urea mCherry-700

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KO} -MYC	-	-	-	-	-	-	-	-	-



Urea MYC-800

	n1			n2			n3		
EGFP ^{inv} -FLAG-MYC	+	-	-	+	-	-	+	-	-
TDP-43 ^{WT} -MYC	-	+	-	-	+	-	-	+	-
TDP-43 ^{ΔNLS} -MYC	-	-	+	-	-	+	-	-	+
TDP-43 ^{ΔNLS/2KO} -MYC	-	-	-	-	-	-	-	-	-

