

## **Supplementary Informations**

### **Kidney inflammaging is promoted by CCR2+ macrophages and tissue-derived micro-environmental factors**

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Cellular and Molecular Life Sciences

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## **Supplementary Methods**

### **Supplementary Tables**

**Table S1**

**Table S2**

## **Supplementary Methods**

### **Histochemistry**

Kidneys were fixed for 6h in 3.4% PFA then in 70% Ethanol and included in paraffin. Tissue sections were stained with sirius red and digitized with a Hamamatsu NanoZoomer. Stained areas and total surface from each section were determined using color-based thresholding and quantified using Fidji software as previously described (1).

### **Biochemistry**

Creatinin and blood urea nitrogen (BUN) were measured in plasma samples with ABX Pentra reagents using the Pentra C400 analyzer (HORIBA Medical).

### **References**

1. Laroumanie F, et al. (2014) CD4+ T cells promote the transition from hypertrophy to heart failure during chronic pressure overload. *Circulation* 129(21):2111-2124.

**Table S1. List of the antibodies used for DECyt analysis**

name	alternative name	clone	purchased from
<b>Set A</b>			
CD45 APC-Cy7		30-F11	Biolegend
Ly6C PB		HK1.4	Biolegend
MHCII FITC	I-A/I-E	M5/114.15.2	Biolegend
CCR2 PE		FAB5538	R&D Systems
CD14 PercPCy5.5		Sa-2-8	eBioscience
CD64 APC	Fc gamma RI	X54-5/7.1	Biolegend
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<b>Set B</b>			
Sca-1 PB	Ly6A/E	D7	Biolegend
CD31 PercPCy5.5	PECAM1	390	Biolegend
CD45 APC-Cy7		30-F11	Biolegend
CD326 FITC	EpCAM	G8.8	Biolegend
mEFSK4 PE	anti-Feeder Cells	mEF-SK4	Miltenyi Biotec
CD73 PE-Cy7	5'ecto-nucleotidase	TY/11.8	Biolegend
CD140b APC	PDGFR $\beta$	APB5	Biolegend

**Table S2. Sequences of the primers used for RT-QPCR**

gene	Primer sequences (5' -> 3')	Ref_Seq
<i>Nt5e</i>	F: CTTCATGAACATCCTGGGCT R: AACGTTCTGAGGAGGGAT	NM_011851
<i>Cdkn1c</i>	F: ATCACCAATCAGCCAGCAGAA R: CTACGCGCTATCACTGGGAAG	NM_009876.4
<i>Cdkn2c</i>	F: GGGGGACCTAGAGCAACTTAC R: CTCCGGATTCCAAGTTCA	NM_007671.2
<i>Cx3cl1</i>	F: TGGCTTGCTCATCCGCTATCAG R: CGTCTGTGCTGTGCGTCTCC	NM_009142.3
<i>Cxcl2</i>	F: CGCTGTCAATGCCTGAAG R: GGCGTCACACTCAAGCTCT	NM_009140.2
<i>Ccl2</i>	F: AACACCTGGATCGGAACCAAAT R: TACGGGTCAACTCACATTCAA	NM_011333.3
<i>Il6</i>	F: GAGGATACCACTCCAACAGACC R: AAGTGCATCATCGTTGTTCATACA	NM_031168
<i>Il1a</i>	F: TTGGTTAAATGACCTGCAACA R: GAGCGCTCACGAACAGTTG	NM_010554.4
<i>Il10</i>	F: TTCAGCCAGGTGAAGACTTTCT R: GCTTGGCAACCCAAGTAACC	NM_010548.2
<i>Tgfb1</i>	F: AGGGCTACCATGCCAACTTCT R: CCGGGTTGTGTTGGTTGAGA	NM_011577.2
<i>Vegfa</i>	F: CAGCAGATGTGAATGCAGACCAA R: CTTTCTCCGCTCTGAACAAAGGC	NM_001025250.3
<i>Cxcl12</i>	F: CTCAACACTCCAAACTGTGCC R: TTGGGCTGTTGTGCTTACTTG	NM_021704.3
<i>Cxcr4</i>	F: CCATGGAACCGATCAGTGTGA R: CAGGGTTCCCTGTTGGAGTCA	NM_009911.3
<i>Igf1</i>	F: GCTCTTCAGTTCGTGTGGAC R: AGCCTGTGGGCTTGTGAAGTA	NM_010512.4
<i>Ccr2</i>	F: CCTTGGGAATGAGTAACGTGTGA R: AATGACAGGATTAATGCAGCAGTGT	NM_009915.2
<i>Tnfa</i>	F: AGCCGATTGCTATCTCATACCA R: GGGCTCATACCAGGGTTGA	NM_013693
<i>Il1b</i>	F: CTGCACTACAGGCTCCGAGAT R: TGTTGGTTGATATTCTGTCCATTG A	NM_008361.4
<i>Il12b</i>	F: CGGACGGTTCACGTGCTC R: CACATGTCACTGCCCGAGAGT	NM_001303244.1

<i>Cdkn2a</i>	F: CCGAACTCTTCGGTCGTACCC R: CTGCTACGTGAACGTTGCCCA	NM_001040654.1
<i>Rplp0</i>	F: GCTTCATTGTGGGAGCAGAC R: ATGGTGTTCTGCCCATCAG	NM_007475.5
<i>Trp53</i>	F: TCCTGGCTGTAGGTAGCGACT R: ATCCGACTGTGACTCCTCCAT	NM_001127233.1 NM_011640.3