

Supplementary Figure 1. Gating Strategy.

Gating strategy used for all data in this manuscript. (a) Gating strategy for the analysis of specificity of CD8+ T cell lines for Figures 1-2. Cells are gated on lymphocytes, singlets, Live CD3^{mid-high}, CD8^{mid-high} T cells and IFNy vs TNF, with the top two quadrants added to report the percentage of IFNy⁺ of CD8⁺ T cells. (b) Gating strategy for the specificity of CD8⁺ T cell lines for Figure 2. Cells are gated on lymphocytes, singlets, Live CD3^{mid-high}, CD8^{mid-high} T cells and tetramer⁺CD8⁺ T cells (c) Gating strategy for the analysis of polyfunctionality of CD8⁺ T cell lines for Figures 3. Cells are gated on lymphocytes, singlets, Live CD3^{mid-high}, CD8^{mid-high} T cells and IFN-γ, TNF, MIP1-β, IL2 or CD107a which were subsequently analysed with Boolean gating.



Supplementary Figure 2. Summary of all intracellular cytokine staining from the initial peptide pool screening.

PBMCs of five HLA-B*18:01⁺ donors (n=5) were stimulated with 2 mM per peptide pool condition (PB1₁₇₇, NS2₁₁₁, NP₄₅, NS2₉₀, M1₅, and NS2₁₀₉) for a period of 10⁺ days. Intracellular cytokine staining was completed after 10⁺ days by restimulating cells individually at 10 mM and 2mM/peptide for peptide pool conditions. All cell line conditions incorporated a x500 positive control and a no peptide negative control.



Supplementary Figure 3. Effector function of peptide-specific CD8⁺ T cell lines.

PBMCs from three HLA-B*18:01⁺ donors (n=3) were stimulated with 10 mM of the NP₂₁₉, PB1₁₇₇, NS2₁₁₁ or M1₅ peptides individually and CD8⁺ T cell function was measured in an ICS assay (a) Representative FACS plots and (b) Summary of all cytokines produced by CD8⁺ T cell lines reported in Figures 2C and 3. The coloured dots represents a particular donor's effector response (red, SG10; blue, SG115; and purple, SG131).

















HLA-B*18:01-NP₂₁₉ HLA-B*18:01-M1₅ HLA-B*18:01-NS2₁₁₁ HLA-B*18:01-PB1₁₇₇

Supplementary Figure 4. Electron density maps for the four immunogenic peptides presented by HLA-B*18:01. Unbiased electron density (Fo-Fc, green at 3s) and final electron density (2Fo-Fc, blue at 1s) around the peptides NP₂₁₉ (\mathbf{a} , \mathbf{b}), M1₅ (\mathbf{c} , \mathbf{d}), NS2₁₁₁ (\mathbf{e} , \mathbf{f}), and PB1₁₁₇ (\mathbf{g} , \mathbf{h}), respectively.



Supplementary Figure 5. Hydrogen bonding network between the HLA-B*18:01 binding pocket (light cyan) and the peptide residue side chains of NP₂₁₉ (cyan) and M1₅ (green). (a) HLA-B*18:01 (pale cyan) presenting the NP₂₁₉ peptide (cyan), with hydrogen bond as yellow dashed lines. (b) HLA-B*18:01 (pale cyan) presenting the M1₅ peptide (green), with hydrogen bond as yellow dashed lines.



Supplementary Figure 6. Top view of pHLA-B*18:01 structures represented as surface. Top-down view of peptide (a, b) NP₂₁₉ (cyan), (c, d) M1₅ (green), (e, f) NS2₁₁₁ (yellow), and (g, h) PB1₁₁₇ (pink). The HLA-B*18:01 binding cleft (light cyan) represented as pale cyan cartoon on the top panels and as surface on the bottom panels.



Supplementary Figure 7. Distinct clonotype T cell receptor length and motif identification. PB1₁₇₇, NS2₁₁₁, M1₅ and NP₂₁₉ peptide specific T cell lines from three HLA-B*18:01+ donors were tetramer stained with the cognate peptide, single-cell sorted and subject to single-cell multiplex PCR. (a) The bar graph illustrates the length of the distinct TCR clonotype (b-c) Depicts the (b) CDR3a and (C) CRD3b motifs for the most common lengths of 14 and 15 amino acids long. Blank amino acid residues display no amino-acid preferences. Motif viewer of CDR3 was obtained via iceLogo (Colaert et al, Nat Methods 2009; 6: 786-787).



Data Collection Statistics	HLA-B*18:01-NS2 ₁₁₁	HLA-B*18:01-PB1 ₁₇₇	HLA-B*18:01-NP ₂₁₉	HLA-B*18:01-M1 ₅
Space group	P 2 ₁ 2 ₁ 2 ₁			
Cell Dimensions (a,b,c) (Å)	50.76, 81.27, 109.89	50.75, 81.63, 110.56	50.85 81.48 111.30	50.65, 81.14, 110.1
Resolution $(Å)$	46.08 - 1.15	46.12 - 1.60	46.25 - 1.40	46.02 - 1.45
Resolution (A)	(1.17 - 1.15)	(1.63 - 1.60)	(1.42 - 1.40)	(1.47 - 1.45)
Total number of observations	1182798	280508	716128	738504
	(56379)	(13853)	(34694)	(35754)
Number of unique observations	159264	59129	91186	81219
Number of unique observations	(7596)	(3078)	(4394)	(3941)
Multiplicity	7.4 (7.4)	4.7 (4.5)	7.9 (7.9)	9.1 (9.1)
Data completeness (%)	98.7 (96.5)	97 (98.5)	99.6 (98.2)	100 (100)
I/sI	13.3 (2.5)	10.3 (1.6)	15.6 (3.3)	14.9 (3.5)
Mn(I) half-set correlation CC _(1/2)	99.9 (80.0)	99.7 (63.0)	99.9 (63.9)	99.8 (70.7)
R_{pim}^{a} (%)	2.5 (29.1)	3.8 (38.9)	2.9 (27.5)	3.4 (26.1)
Refinement Statistics				
R _{factor} ^b (%)	15.24	20.16	15.37	18.60
R_{free}^{b} (%)	16.72	23.43	18.86	20.23
Rms deviations from ideality				
Bond lengths (Å)	0.004	0.007	0.009	0.001
Bond angles (°)	0.79	0.92	1.03	1.25
Ramachandran plot (%)				
Allowed region	98.42	98.64	98.68	98.68
Disallowed region	0.00	0.00	0.26	0.26
PDB code	8ROP	8RNH	8ROO	8RNG

Supplementary Table 1. Data Collection and Refinement Statistics

$$\label{eq:Rp_im} \begin{split} ^{a}R_{p.i.m} &= S_{hkl} \left[1/(N-1) \right]^{1/2} S_{i} \mid I_{hkl,\,i} - < I_{hkl} > \mid / S_{hkl} < I_{hkl} >. \\ ^{b}R_{factor} &= S_{hkl} \mid \mid F_{o} \mid - \mid F_{c} \mid \mid / S_{hkl} \mid F_{o} \mid \text{for all data except} \approx 5\% \text{ which were used for } R_{free} \text{ calculation.} \end{split}$$

Supplementary Table 2. B18/NP219 specific TCR sequences

								Number of sequence		uences
TRAV	TRAJ	CDR3a	Length	TRBV	TRBJ	CDR3β	Length	SG10	SG115	SG131
14/DV4*01	4*01	CSLREEFSGGFNKLIF	16					1		
22*01	36*01	CAPQSPWANNLFF	13					1		
				19*01/02	2-1*01	CASSIDLTSGVYNEQFF	17	1		
				19*01/02	2-5*01	CASSIDYPSGVQETQYF	17	2		
				19*01/02	2-7*01	CASSTDLATGAYEQYF	16	1		
				19*01/02	2-7*01	CASSIDLASGTTYEQYF	17	1		
				2*01	2-5*01	CASSEGSLGGGLETQYF	17	1		
				2*01/02/03	2-5*01	CASMGRDINQPQHF	14	1		
				4-1*01	1-1*01	CASSQDPGGTEAFF	14	1		
38-1*01	34*01	CAFMRDDNTDKFIF	14	4-3*01	2-7*01	CASSQDFAGGSYEQYF	16		1	
29/DV5*01	42*01	CAAGNYGGSQGNLIF	15	7-2*04	1-2*01	CASSLEAVSIHGYTF	15		1	
20*01/02	42*01	CAVGREYGGSQGNLIF	16	4-1*01	2-2*01	CASSQDKTTGELFF	14		1	
				6-5*01	1-1*01	CASSWGLEVNTEAFF	15		1	
				6-5*01	1-1*01	CASSYGLEANTEAFF	15		1	
14/DV4*03	8*01	CVMREAMNTGFOKLVF	16						1	
				4-3*01	1-2*01	CASSODRGTGAYGYTF	16		1	
14/DV4*03	42*01	CAMRPYGGSOGNLIF	15	4-3*01	2-7*01	CASSODRASGEYEOYF	16			1
29/DV5*01	49*01	CAASAONTGNOFYF	14	7-	1-1*01	CASSPSMRSGLAEAFF	16			9
				2*01/02/04						
				or 7-8*01						
29/DV5*01	49* 01	CAASAQNTGNQFYF	14							6
22*01	20*01	CAVVSNDYKLSF	12							2
13-2*01/02	37*02	CADSSNTGKLIF	12							1
l	37*02	CAEASNTGKLIF	12							1
				19*03	2-7*01	CASSWDRGTGEQYF	14			1
				4-3*01	2-1*01	CASSQDLASGTYNEQFF	17			3

		Total number of resolved	sequences	10	7	26
2*01/02/04 or 7-8*01 19*01/02	2-7*01	CASSIDLAGGSYEQYF	16			1
7-	1*01	CASSPSMRSGLAEAFF	16			1

The $\alpha\beta$ TCR repertoire of CD8⁺ T cells from SG10, SG115 and SG131 specific for the B18/NP₂₁₉ tetramer. Length refers to the number of amino acids in the CDR3.

Supplementary 7	Fable 3.	B18/PB1 177	specific	TCR s	sequences
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							Number of sequ		uences	
TRAV	TRAJ	CDR3a	Length	TRBV	TRBJ	CDR3β	Length	SG10	SG115	SG131
1-2*01	28*01	CAVRDQYSGAGSYQLTF	17					1		
				10-3*04	2-1*01	CAISEEGQGRDEQFF	15	1		
				10-2*02	1-2*01	CASSLGTGGGYTF	13	2		
				3-2*03	2-5*01	CASSQVLVGETQYF	14	2		
				4-3*02/03	2-3*01	CASSSSGGGAGDTQYF	16	1		
				20-1*02/03	2-3*01	CSASPERGTSVGTDTQYF	18	1		
				19*01/02	1-1*01	CASSIVERAEAFF	13	1		
1-2*01	28*01	CAVRDQRAGSYQLTF	15						3	
1-2*01/03	28*01	CAVRDQRAGSYQLTF	15						12	
1-2*01/03	28*01	CAVRDQLTGAGSYQLTF	17						3	
1-2*01/03	28*01	CAMRDQRGGSHQIPL	15						1	
3*01	13*01	CAVRDSFGSGGYQKVTF	17						1	
				5-4*01	2-3*01	CASSLLTGGTDTQYF	15		2	
				19*01/03	2-3*01	RAGPGQRLWAPVF	13		1	
6*01/07	23*01	CALDTYNQGGKLIF	14	27*01	2-2*01	CASSWVPPGQGRTGELFF	18			8
6*01/07	23*01	CALDTYNQGGKLIF	14							11
				27*01	2-2*01	CASSWVPPGQGRTGELFF	18			1
						Total number of	sequences	9	23	20

The $\alpha\beta$ TCR repertoire of CD8⁺ T cells from SG10, SG115 and SG131 specific for the B18/PB1₁₇₇ tetramer. Length refers to the number of amino acids in the CDR3.

								Number of sequences		uences
TRAV	TRAJ	CDR3	Length	TRBV	TRBJ	CDR3β	Length	SG10	SG115	SG131
6*01/07	23*01	CALGGAYNQGGKLIF	15					1		
				6-4*01/02	2-2*01	CASSDSDGSGELFF	14	1		
				20-1*01	2-1*01	CSARDRQASSYNEQFF	16	2		
				27*01	1-4*01	CASSFLQGEKLFF	13	1		
				27*01	1-2*01	CASSLRPDPPYNEQFF	16	1		
				27*01	1-1*01	CASSPPGREDTEAFF	15	1		
				18*01	1-1*01	CASSPSEGLNTEAFF	15	1		
				20-1*01	1-6*02	CSARDGQGSSPLHF	14	1		
				20-1*01/04/05	2-1*01	CSANDRTSGSNYNEQFF	17	1		
				15*02	1-5*01	CATSRDRGWSNQPQHF	16	1		
				4-1*01/02	1-6*01/02	CASSQAGSLENSPLPLL	17	1		
				2*02	2-7*01	CASSTGRYYEQYF	13	1		
8-6*02	32*02	CAVTEGPYGGATNKLIF	17	4-2*01	2-3*01	CASSKGLAVDYTQYF	15		1	
				12-3/4*01	2-3*01	CASRLVSGTDTQYF	14		7	
				28*01	1-4*01	CASSFLATNEKLFF	14		1	
				20-1*04	2-2*01	CSASLERMNTGELFF	15			1
						Total number of sequence	es resolved	13	9	1

Supplementary Table 4. B18/NS2₁₁₁ specific TCR sequences

The $\alpha\beta$ TCR repertoire of CD8⁺ T cells from SG10, SG115 and SG131 specific for the B18/NS2₁₁₁ tetramer. Length refers to the number of amino acids in the CDR3.

Supplementary Table 5. B18/M15 specific TCR sequences

							Nu	umber of se	equences	
TRAV	TRAJ	CDR3	Length	TRBV	TRBJ	CDR3β	Length	SG10	SG115	SG131
20*01/02	33*01	CAVQAHDSNYQLIW	14	4-1*01/02	1-4*01	CASRRTGVVEKLFF	14	3		
20*01/02	33*01	CAVQAHDSNYQLIW	14					5		
				4-1*01/02	1-4*01	CASRRTGVVEKLFF	14	1		
				10-3*04	2-7*01	CAISAPRGERDEQYF	15	5		
				4-3*01	2-3*01	CASSQGPQVRGADTQYF	17		5	
				27*01	1-3*01	CASSLSWGPGNTIYF	15		4	
				18*01	1-2*01	CASSPTPGDPHYGYTF	16		2	
14/DV4*03	47*01	CAMREDGNKLVF	12	27*01	2-5*01	CASRSAGHQETQYF	14			1
				27*01	2-5*01	CASRSAGHQETQYF	14			2
				19*01/02	2-1*01	CASSLLAGVYNEQFF	15			1
						Total number of resolved se	14	11	4	

The $\alpha\beta$ TCR repertoire of CD8⁺ T cells from SG10, SG115 and SG131 specific for the B18/M1₅ tetramer. Length refers to the number of amino acids in the CDR3.