nature portfolio

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Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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n/a	Confirmed
	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗴 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
	🕱 A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
	🕱 For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	$oxed{x}$ For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
×	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
,	Our web collection on statistics for biologists contains articles on many of the points above

Software and code

Policy information about availability of computer code

Data collection

Unicorn v.7, Tecan iControl for infinite 200 pro

Data analysis

ClustalO (v2.1), Unipro UGENE (v47), iTOL (v6.8.1), DADA2 (v1.26), SqueezeMeta v1.3.1, FragGeneScan v1.31, GenBank r239, eggNOG v5.0, KEGG r58.0, CAZy (as of 11/12/2023), Diamond v0.9.24.125, Pfam 33.0, HMMER3, Bowtie2, Mascot v2.7.0.1, Scaffold 4.11.1 & 5.0.1, X! Tandem v2017.2.1.4, GhostKOALA v2.0, hmmscan v3.3.2, KofamScan, MaxQuant,

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Metagenome, metatranscriptome and MAG sequence data are available from the European Nucleotide Archive (accession PRJEB52999; https://www.ebi.ac.uk/ena/browser/view/PRJEB52999). The mass spectrometry proteomic data have been deposited to the ProteomeXchange Consortium (http://proteomexchange.org) via the PRIDE partner repository83 with the dataset identifier PXD019294 (bloom 2016; http://

proteomecentral.proteomexchange.org/cgi/GetDataset?ID=PXD019294), PXD042676 (bloom 2018; http://proteomecentral.proteomexchange.org/cgi/GetDataset?ID=PXD042676), PXD042805 (bloom 2020; http://proteomecentral.proteomexchange.org/cgi/GetDataset?ID=PXD042805), PXD043390 (Single strain proteomics; http://proteomecentral.proteomexchange.org/cgi/GetDataset?ID= PXD043390). All other data supporting the findings of this study are available within the paper and its Supplementary Information and source data is provided with the paper.

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Research involving I	hiiman narticii	nante thair	data or l		matarial
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Policy information about stuand sexual orientation and re	dies with <u>human participants or human data</u> . See also policy information about <u>sex, gender (identity/presentation),</u> <u>ace, ethnicity and racism</u> .	
Reporting on sex and gender	na	
Reporting on race, ethnicity, cother socially relevant grouping		
Population characteristics	na	
Recruitment	na	
Ethics oversight	na	
Note that full information on the	e approval of the study protocol must also be provided in the manuscript.	
Field-specific	reporting	
Please select the one below	that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.	
Life sciences	Behavioural & social sciences	
For a reference copy of the documer	nt with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf	
Ecological, ev	olutionary & environmental sciences study design	
All studies must disclose on t	these points even when the disclosure is negative.	
Study description	Investigation of the microbiome of seasonal spring phytoplankton blooms in the North Sea.	
·	crobial biomass from surface water in the North Sea, representing the microbiome of the spring phytoplankton bloom around the and of Helgoland in the North Sea.	
	ampled surface water was sequentially filtered onto polycarbonate membrane filters with different pore sizes (10 micro m, 3 micro and 0.2 micro m); different pore sizes largely account for larger eukaryotes (10 micro m), smaller eukaryotes and attached acteria (3 micro m) and free-living bacteria (0.2 mircro m)	
Data collection	Metagenome, metatranscriptome, metaproteome, 16S rRNA, 18S rRNA, glycans	
Timing and spatial scale	Subsurface seawater (1 m depth) was collected at 52 time points between 2nd of March and 26th of May 2020.	
	exclusion of Metazoan reads from 18S data sets as detailed in methods to recieve an accurate depiction of relevant microalgae during the bloom	
Reproducibility	We used in all experiments at least three biological replicates.	
Randomization	not relevant, sample collection from surface water	
Blinding	not relevant, sample collection from surface water	
Did the study involve field	work? 🗶 Yes 🗌 No	
Field work, collect	ion and transport	
	Subsurface seawater (1 m depth) was collected at 52 time points between 2nd of March and 26th of May 2020 at the station Helgoland Roads near Helgoland in the southern North Sea. Since 1962 bucket water samples have been taken as part of a long-term monitoring program Helgoland Roads (54°11'N 7°54'E; DEIMS.iD: https://deims.org/1e96ef9b-0915-4661-849f-b3a72f5aa9b1)41.	
Location	Helgoland Roads (54°11'N 7°54'E)	
Access & import/export	Since 1962 bucket water samples have been taken as part of a long-term monitoring program Helgoland Roads. Membran filter	

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	samples were frozen and st	tored at -80 °C.
Disturbance	no	
Poporting fo	r specific m	enterials systems and mothods
<u> </u>	<u> </u>	aterials, systems and methods
		materials, experimental systems and methods used in many studies. Here, indicate whether each material, e not sure if a list item applies to your research, read the appropriate section before selecting a response.
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Materials & experime	ntal systems	Methods
n/a Involved in the study		n/a Involved in the study
Antibodies		▼ ChIP-seq
x Eukaryotic cell lines		Flow cytometry
Palaeontology and a	ırchaeology	MRI-based neuroimaging
Animals and other organisms		
Clinical data		
Dual use research o	fconcern	
▼ Plants		
Plants		
Seed stocks	na	
Seed Stocks	lia	
Novel plant genotypes	na	
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Authentication

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