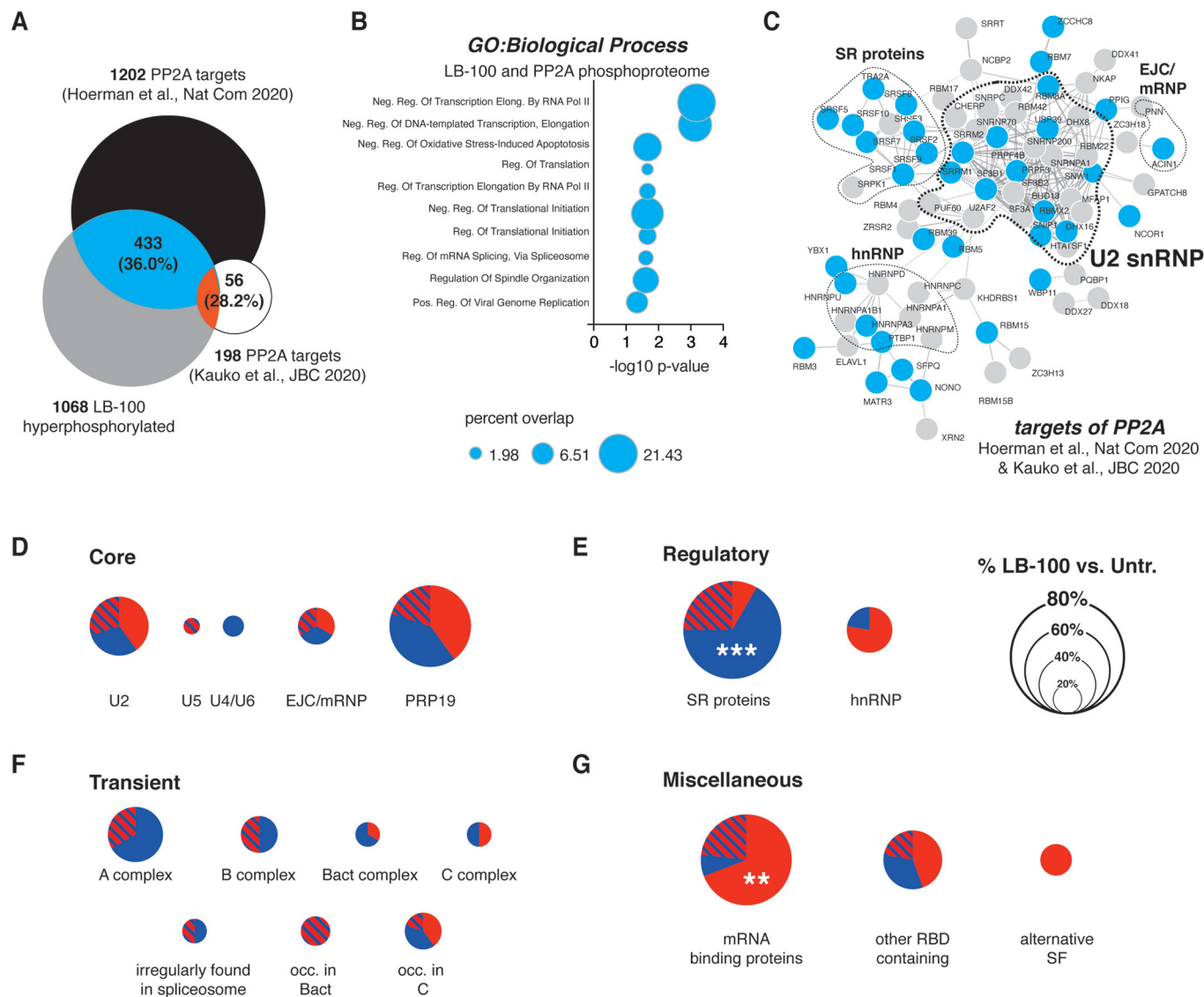
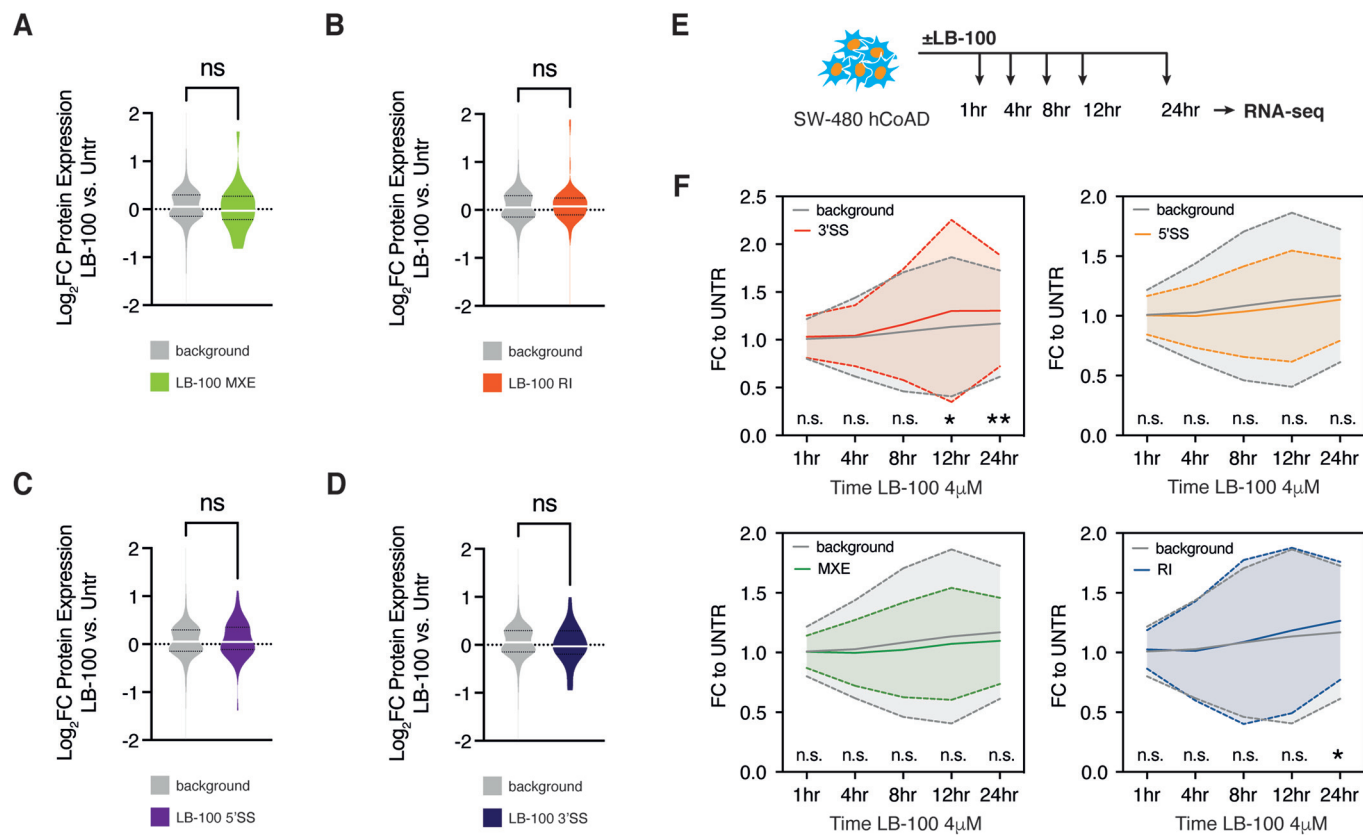


## Expanded View Figures



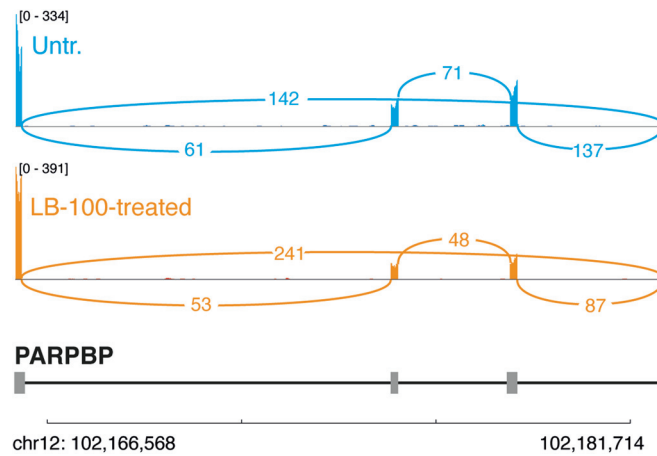
**Figure EV1. Specific splicing nodes are differentially phosphorylated in response to LB-100 treatment in colorectal adenocarcinoma.**

(A) Venn diagram shows an overlap between differentially hyperphosphorylated proteins in LB-100-treated SW-480 colon adenocarcinomas and previously reported targets of PP2A (Hoermann et al, 2020; Kauko et al, 2020). (B) The bar graph shows Gene Ontologies enriched for 34 differentially phosphorylated proteins from LB-100-treated SW-480 human colorectal carcinoma that are previously reported PP2A targets (Hoermann et al, 2020; Kauko et al, 2020). *p* values were calculated with Fisher's exact test. (C) Interaction maps of spliceosome components hyper- and hypophosphorylated in response to 12 h of LB-100 stimulation in SW-480 cells (gray) overlaid with previously reported targets of PP2A (Hoermann et al, 2020; Kauko et al, 2020) (blue). Connecting lines show interactions from STRINGdb. (D-G) Pie charts show the distribution of proteins with significantly changed (FDR <0.05 and |Log<sub>2</sub>Fold Change| >0.58) phosphorylation status within different functional groups of splicing factors. RBD RNA binding domain, SF splicing factor. Data information: In (D-G), statistical significance was calculated with hypergeometric test vs. background, \*\*\**p* < 0.001; \*\**p* < 0.01.



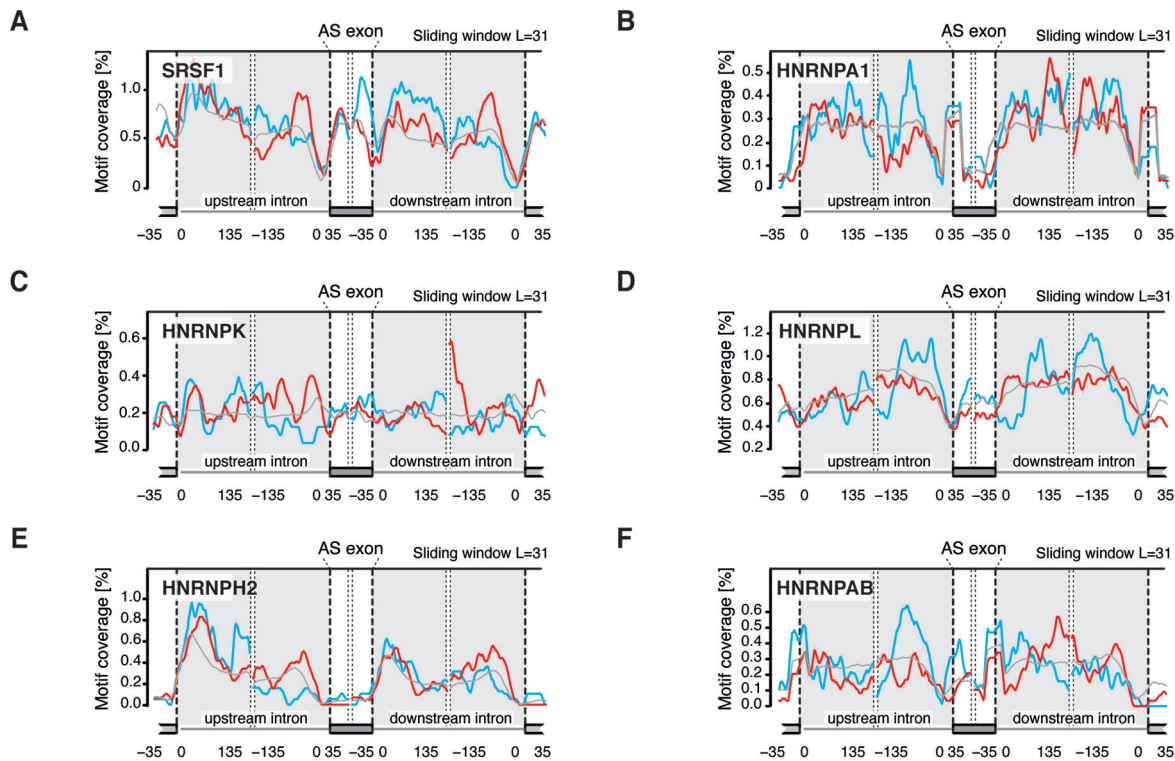
**Figure EV2. PP2A inhibition-dependent alternative splicing affects protein but not transcript levels of alternatively spliced mRNAs.**

(A–D) Violin plots show no effect on levels of proteins resulting from alternatively spliced transcripts in MXE (A), RI (B), alt5'ss (C), and alt3'ss (D) in 4 μM LB-100-treated SW-480 cells compared to control in  $n = 4$  biological replicates. (E) Schematic illustrates an experimental approach to determine time-resolved transcriptomic changes of alternatively spliced mRNAs in response to 4 μM LB-100 stimulation. (F) Graphs show time-resolved changes in alternatively spliced transcripts and mRNAs from RI, MXE, alt5'ss, and alt3'ss groups in response to LB-100 stimulation. Data information: In (F), statistical significance was calculated with one-way ANOVA,  $^{*}p < 0.01$ ;  $^{*}p < 0.05$ ; n.s. not significant. Source data are available online for this figure.

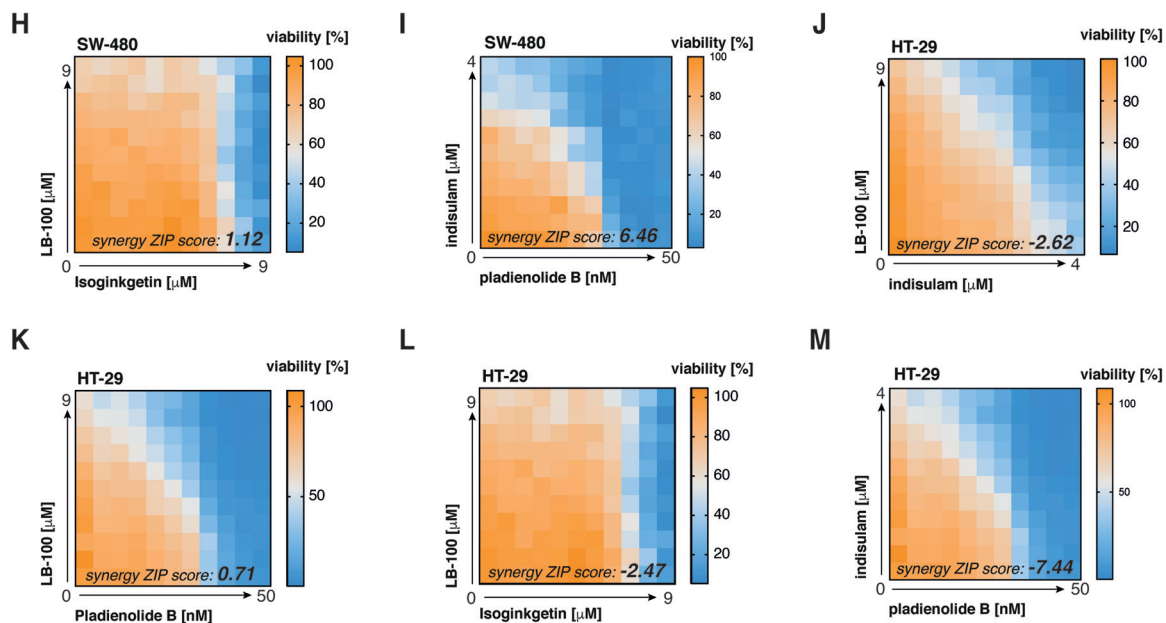
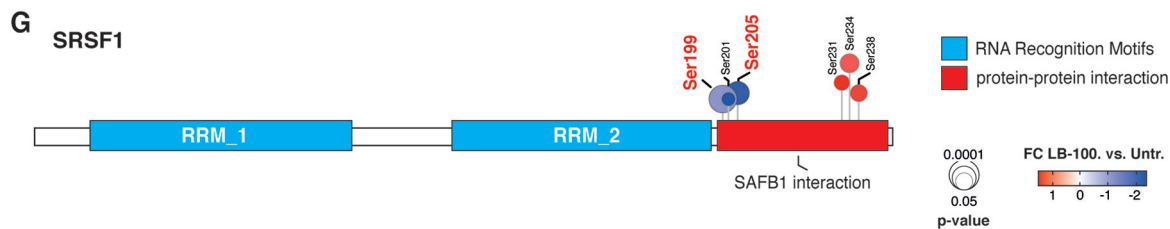


**Figure EV3. Alternative splicing events in response to LB-100 impact the fate of DNA damage-response regulators.**

An alternative splicing event within PARPBP gene and representative Sashimi plots showing differential splicing in untreated and LB-100-treated SW-480 cells.

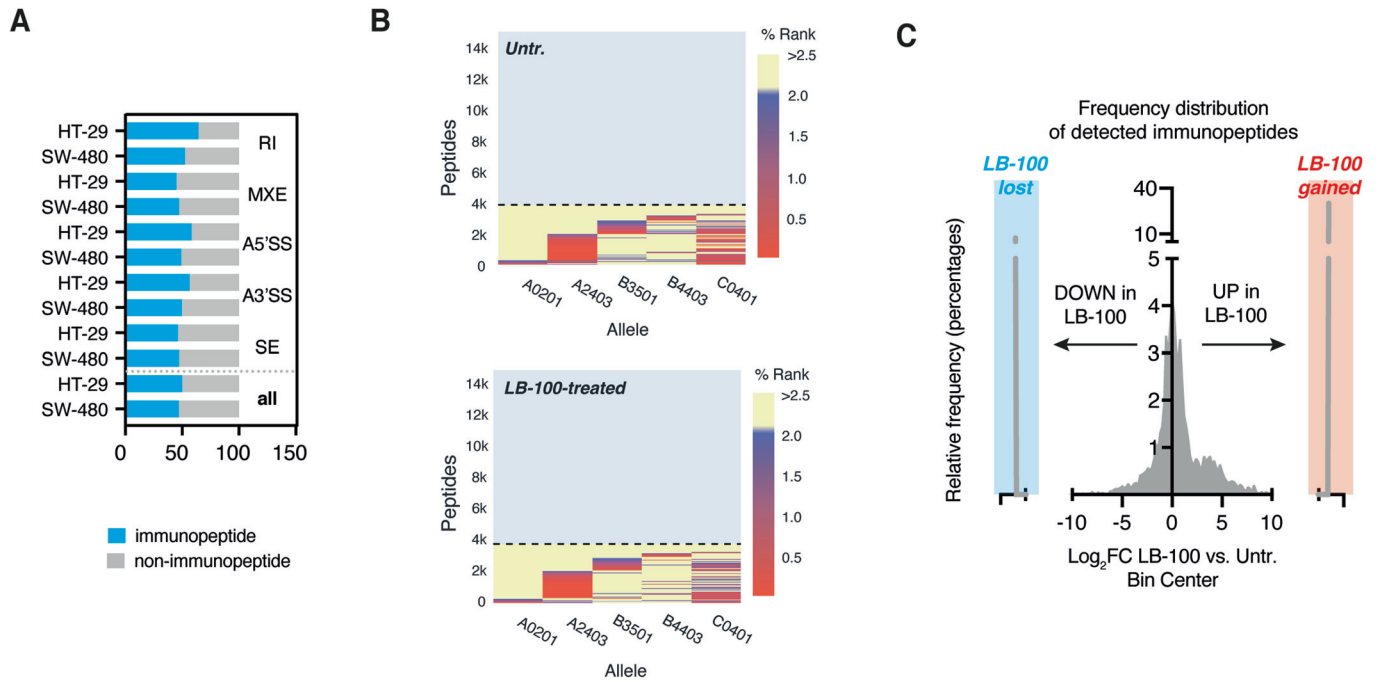


Exons with  $\Delta$ PSI in LB-100: ■ UP ■ DOWN ■ NO



**◀ Figure EV4. Impact of LB-100-driven phosphorylation changes on the function of spliceosome components.**

(A–F) Motif RNA map shows SRSF1 (A), HNRNPA1 (B), HNRNPK (C), HNRNPL (D), HNRNPH2 (E), and HNRNPAB (F) proteins binding in the proximity of excluded (DOWN), included (UP), or not affected (NO) exons in LB-100-treated as compared to untreated SW-480 colorectal adenocarcinoma cells. (G) Location of mis-phosphorylated sites across SRSF1 protein. Functionally relevant protein regions according to UniProt are shown. Significantly changing phosphosites in response to LB-100 are highlighted. (H,I) Synergy matrices show a lack of cooperativity between LB-100 and isoginkgetin (H) or indisulam and pladienolide B (I) in the SW-480 human adenocarcinoma cell line. Representative heat map for  $n = 3$  biological replicates. (J–M) Synergy matrices show a lack of cooperativity between LB-100 and indisulam (J), pladienolide B (K), isoginkgetin (L), or indisulam and pladienolide B (M) in HT-29 human adenocarcinoma cell line. Representative heat map for  $n = 3$  biological replicates. Source data are available online for this figure.



**Figure EV5. LB-100 determines neoantigen formation presented within MHC-I.**

(A) The bar graph shows an equal contribution of all the splicing events toward the potential formation of tumor neoantigens reported previously (Lu et al, 2021). (B) Heat maps show HLA haplotype representation in untreated controls (Untr.) and LB-100-stimulated HT-29 human colorectal carcinoma cells from  $n = 3$  biological replicates. (C) The plot shows the cumulative distribution of  $\log_2$  fold change in peptides presentation between LB-100-stimulated and untreated control (Untr.) HT-29 cells from  $n = 3$  biological replicates. Highlighted in red and blue are peptides detected exclusively in LB-100 and control cells, respectively.