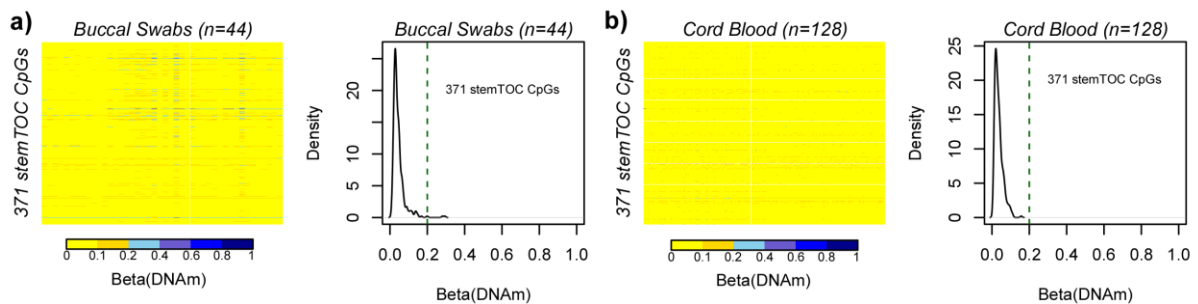


1 **Supplementary Information for “An improved**
2 **epigenetic counter to track mitotic age in normal and**
3 **precancerous tissues”**

4 **SUPPLEMENTARY FIGURES**

5

6



7

8 **Supplementary Figure 1: stemTOC CpGs retain ultra-low DNAm levels in neonatal**
9 **tissues. a) Heatmap of DNAm values (EPIC) of the 371 stemTOC CpGs in n=44 buccal swabs**
10 **from newborns ¹. Density distribution displays the average DNAm of the 371 CpGs across the**
11 **buccal swabs. b) As a) but for a cord blood Illumina 450k dataset of n=128 samples ².**

12

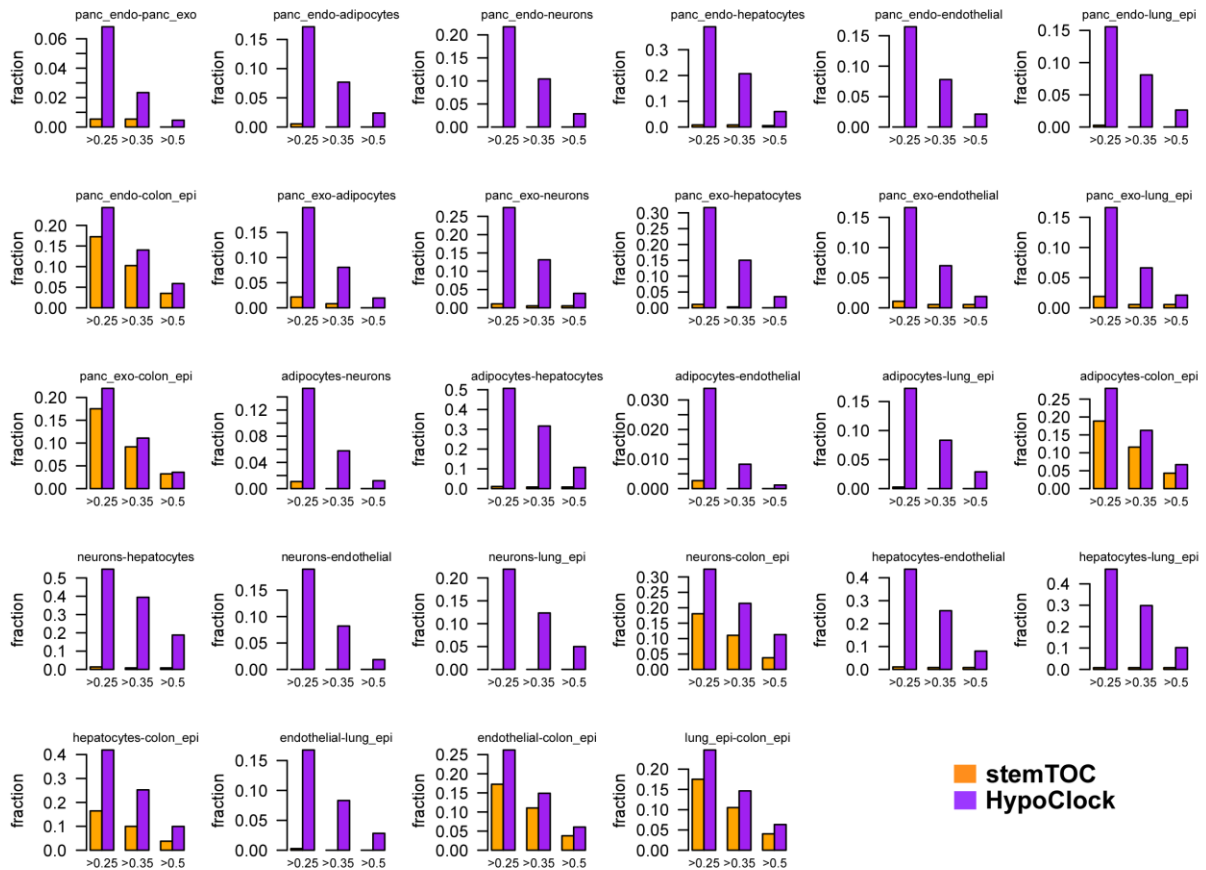


13

14 **Supplementary Figure 2: stemTOC CpGs do not display cell-type specific DNAm**
 15 **differences between age-matched sorted immune cells and between eGTEX tissue-types.**

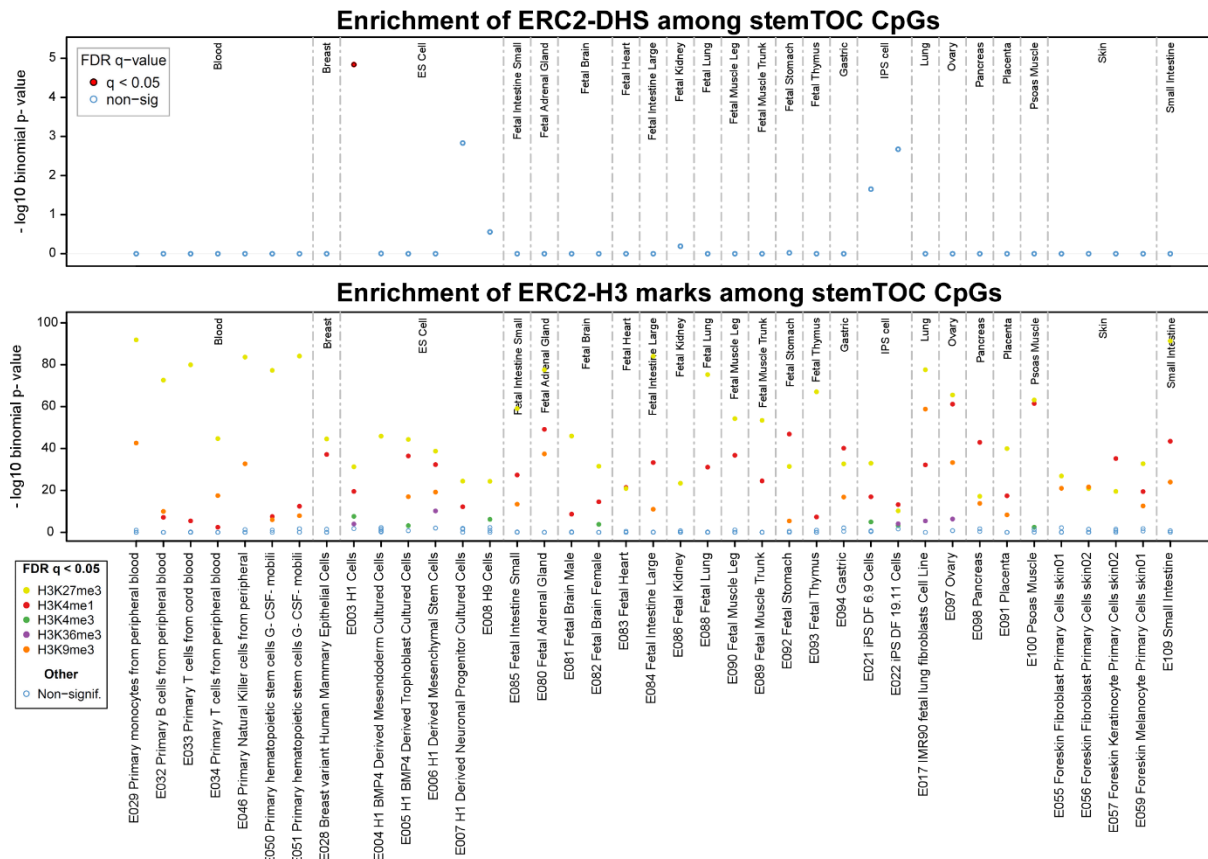
16 **a)** Barplots depicting the fraction of stemTOC (orange) and HypoClock (purple) CpGs that
 17 display absolute differences in DNAm greater than 0.25, 0.35 and 0.5, between age-matched
 18 sorted immune-cell types from Paul et al ³: B-cells (n=100), Monocytes (n=104) and CD4T-
 19 cells (n=98). **b)** As a) but for the matched sorted immune-cell types from BLUEPRINT
 20 (Monocytes n=139, Neutrophils n=139, naïve-CD4T-cells n=139) ⁴. **c)** As a) but for the age-
 21 matched tissues from eGTEX project: breast mammary tissue (n=52), colon transverse (n=224),
 22 kidney cortex (n=50), lung (n=223), skeletal muscle (n=47), ovary (n=164), prostate (n=123),
 23 testis (n=50) and whole blood (n=54)) ⁵.

24



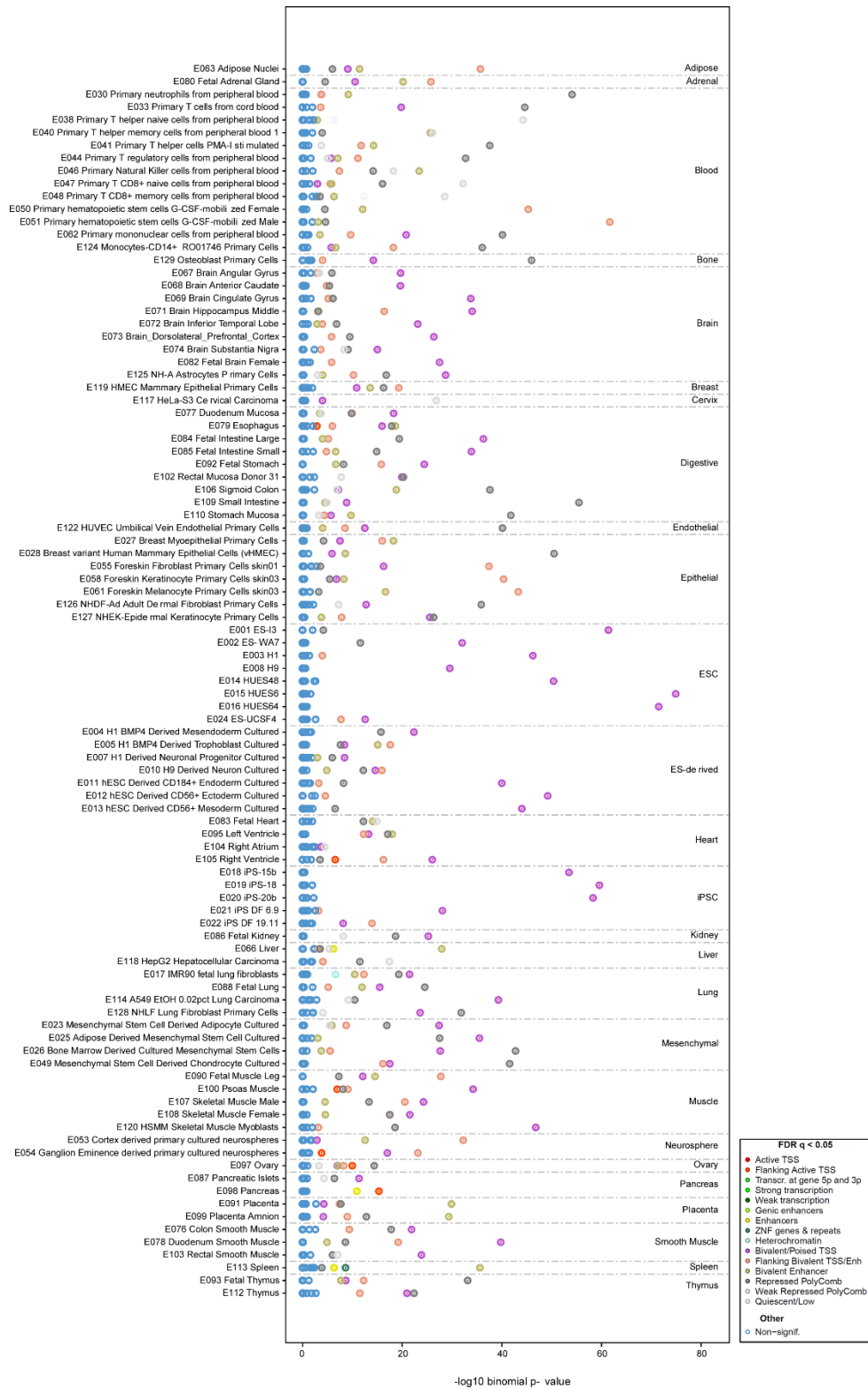
25

26 **Supplementary Figure 3: stemTOC CpGs do not display cell-type specific DNAm**
 27 **differences between age-matched sorted cells from Moss et al DNAm-atlas.** Barplots
 28 depicting the fraction of stemTOC (orange) and HypoClock (purple) CpGs that display
 29 absolute differences in DNAm greater than 0.25, 0.35 and 0.5, between age-matched sorted
 30 cell types from Moss et al ⁶: sorted pancreatic beta cells (n=4), pancreatic ductal (n=3),
 31 pancreatic acinar (n=3), adipocytes (n=3), hepatocytes (n=3), cortical neurons (n=3), leukocyte
 32 (n=1), lung-epithelial (n=3), colon-epithelial (n=3) and vascular endothelial (n=2).



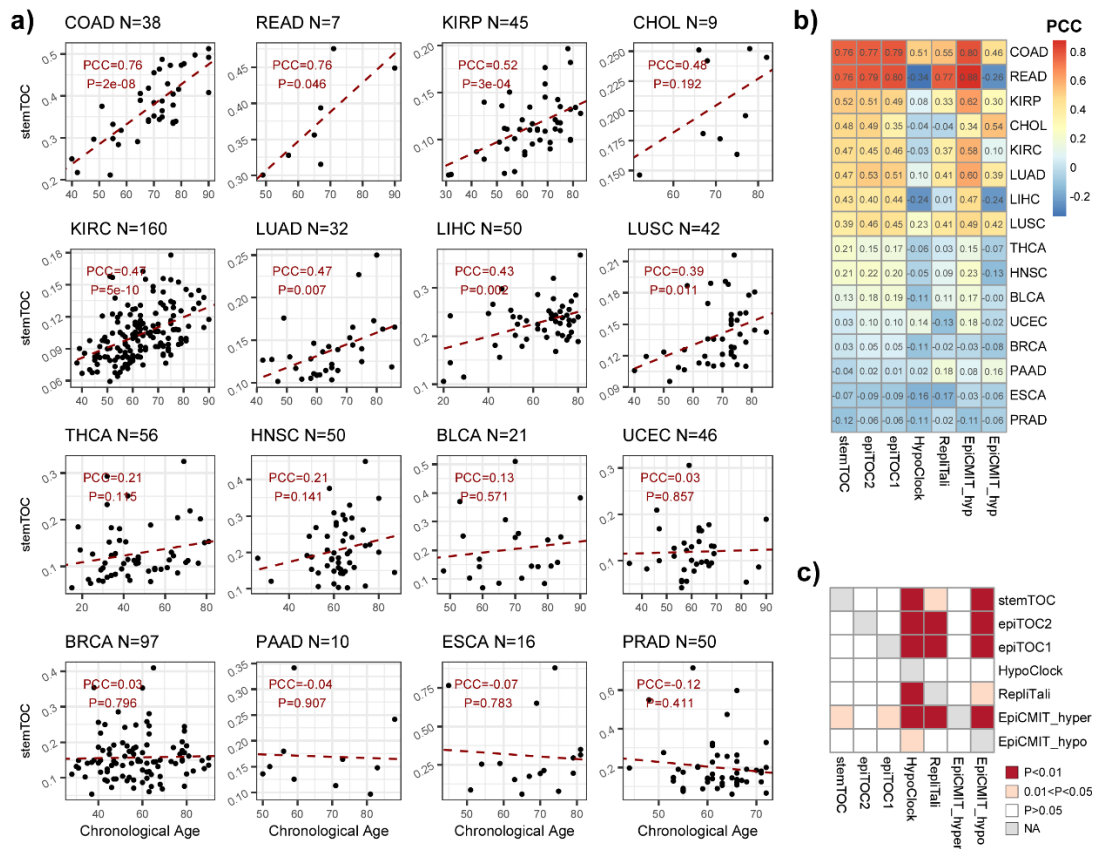
33
 34
 35
 36
 37
 38
 39
 40
 41

Supplementary Figure 4a: eFORGE2 DHS and H3 enrichment analysis of stemTOC CpGs. Top panel displays the $-\log_{10}P$ -values of enrichment (y-axis) of DHSs from the eFORGE2 Binomial test among the 371 stemTOC CpGs. The enrichment is shown for DHSs as defined by different cell-types (x-axis). Lower panel is the corresponding enrichment plot for Histone H3 marks, as indicated. In both panels, colors indicate statistical significance at $FDR < 0.05$.



42

43 **Supplementary Figure 4b: eFORGE2 chromatin state enrichment analysis of stemTOC**
 44 **CpGs.** Panel displays the $-\log_{10}P$ -values of enrichment (y-axis) of 15 chromatin states from the
 45 eFORGE2 Binomial test among the 371 stemTOC CpGs. The enrichment is shown for
 46 chromatin-states as defined by different cell-types (x-axis). Colors indicates statistical
 47 significance at $FDR < 0.05$.



49

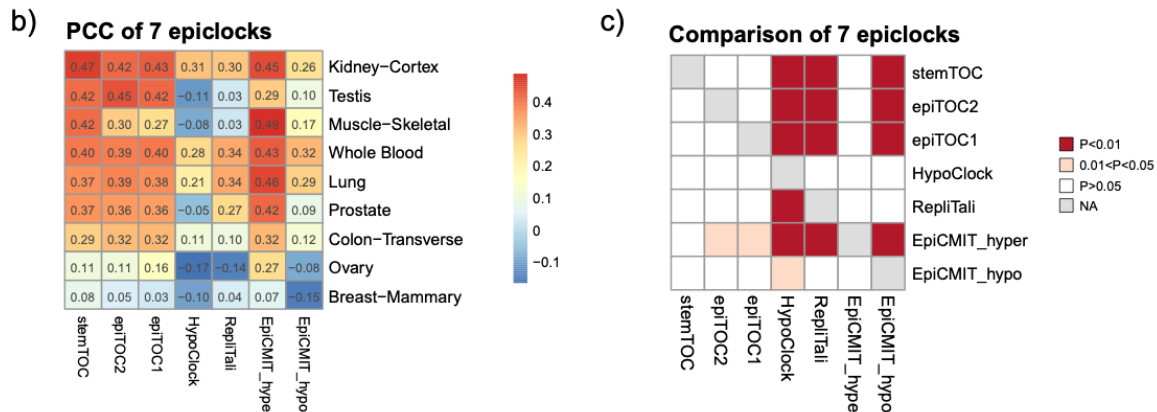
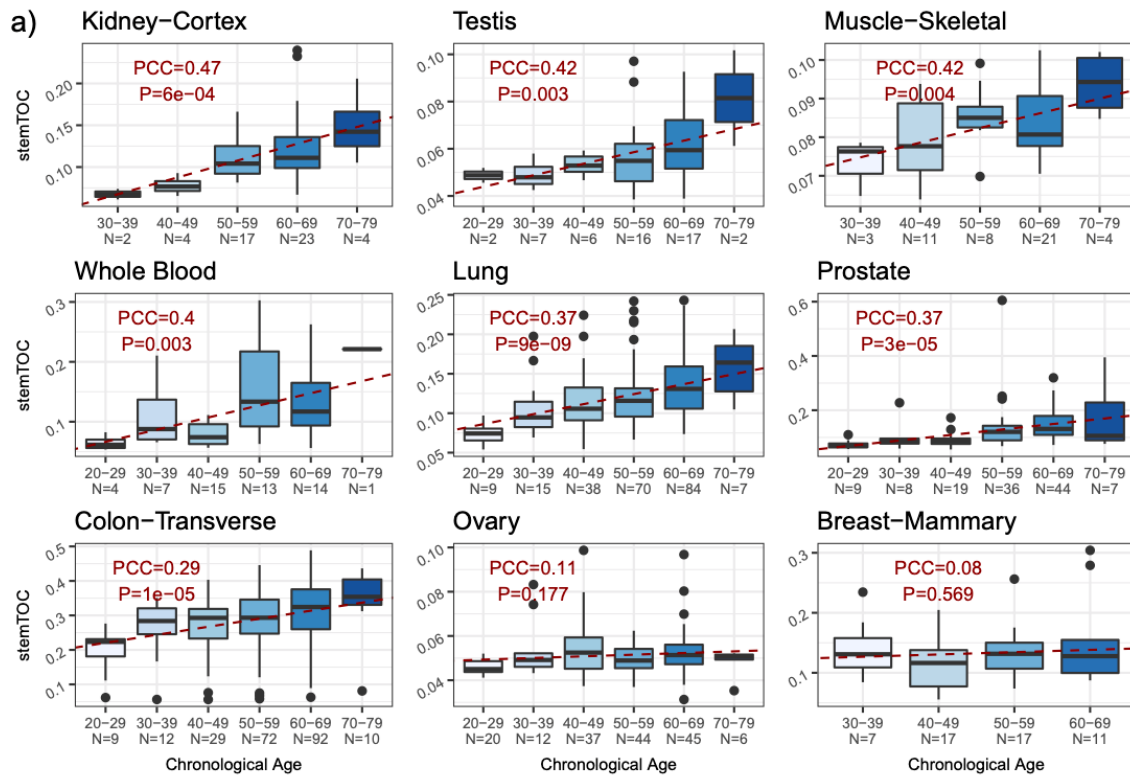
50 **Supplementary Figure 5: Correlation of mitotic-age with chronological age in normal-**
 51 **adjacent tissues from TCGA. a)** Scatterplots of the stemTOC mitotic age score (y-axis)
 52 against chronological age (x-axis) for 16 normal-adjacent tissue-types from TCGA. The
 53 number of samples is given above each plot. For each panel we provide the Pearson Correlation
 54 Coefficient (PCC) and two-tailed P-value from a linear regression. **b)** Heatmap of
 55 corresponding PCC-values for each tissue-type and for 7 different mitotic clocks, as indicated.
 56 The PCC-values are provided in the heatmap. **c)** Heatmap displaying one-tailed paired
 57 Wilcoxon rank sum test P-values, comparing clocks to each other, in how well their mitotic age
 58 correlates with chronological age. Each row indicates how well the corresponding clock's
 59 mitotic age estimate correlates with chronological age as compared to the clock specified by
 60 the column. The paired Wilcoxon test is performed over the 16 tissue-types. RepliTali results
 61 are for the probes restricted to 450k beadarrays.

62

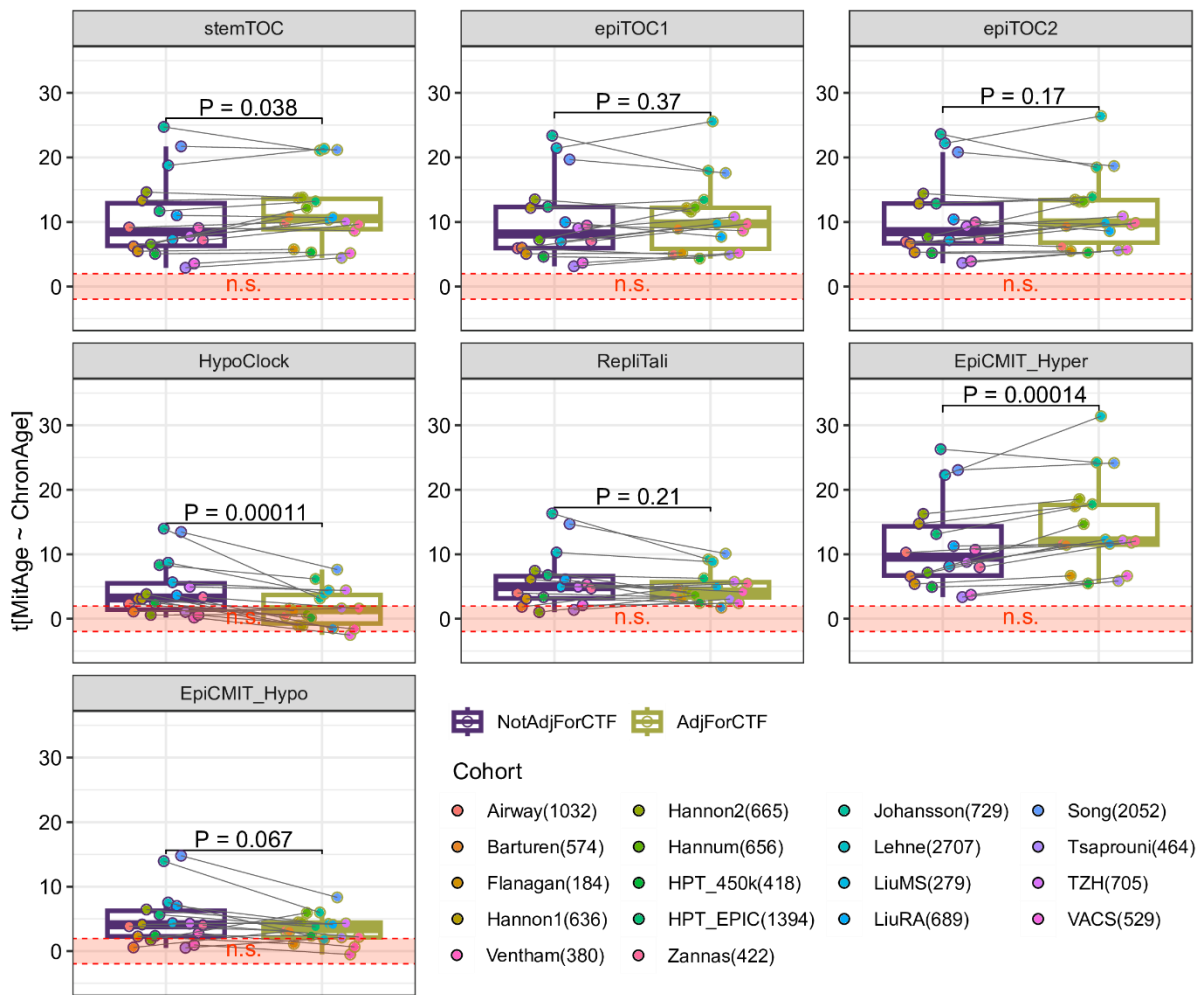
63

64

65



66
67 **Supplementary Figure 6: Correlation of mitotic-age with chronological age in normal**
68 **tissues from eGTEx. a)** Boxplots of stemTOC mitotic age score (y-axis) against chronological
69 age group (x-axis) for 9 normal tissue-types from eGTEx (EPIC data). The number of samples
70 in each age-group is indicated below the plot. For each panel we provide the Pearson
71 Correlation Coefficient (PCC) and two-tailed P-value from a linear regression. The line within
72 each box denotes the median with the box itself defining the interquartile range and whiskers
73 extending 1.5 times the interquartile range. **b)** Heatmap of corresponding PCC-values for each
74 tissue-type and for 8 different mitotic clocks, as indicated. The PCC-values are provided in the
75 heatmap. **c)** Heatmap displaying one-tailed paired Wilcoxon rank sum test P-values, comparing
76 clocks to each other, in how well their mitotic age correlates with chronological age. Each row
77 indicates how well the corresponding clock's mitotic age estimate correlates with chronological
78 age as compared to the clock specified by the column. The paired Wilcoxon test is performed
79 over the 9 tissue-types.



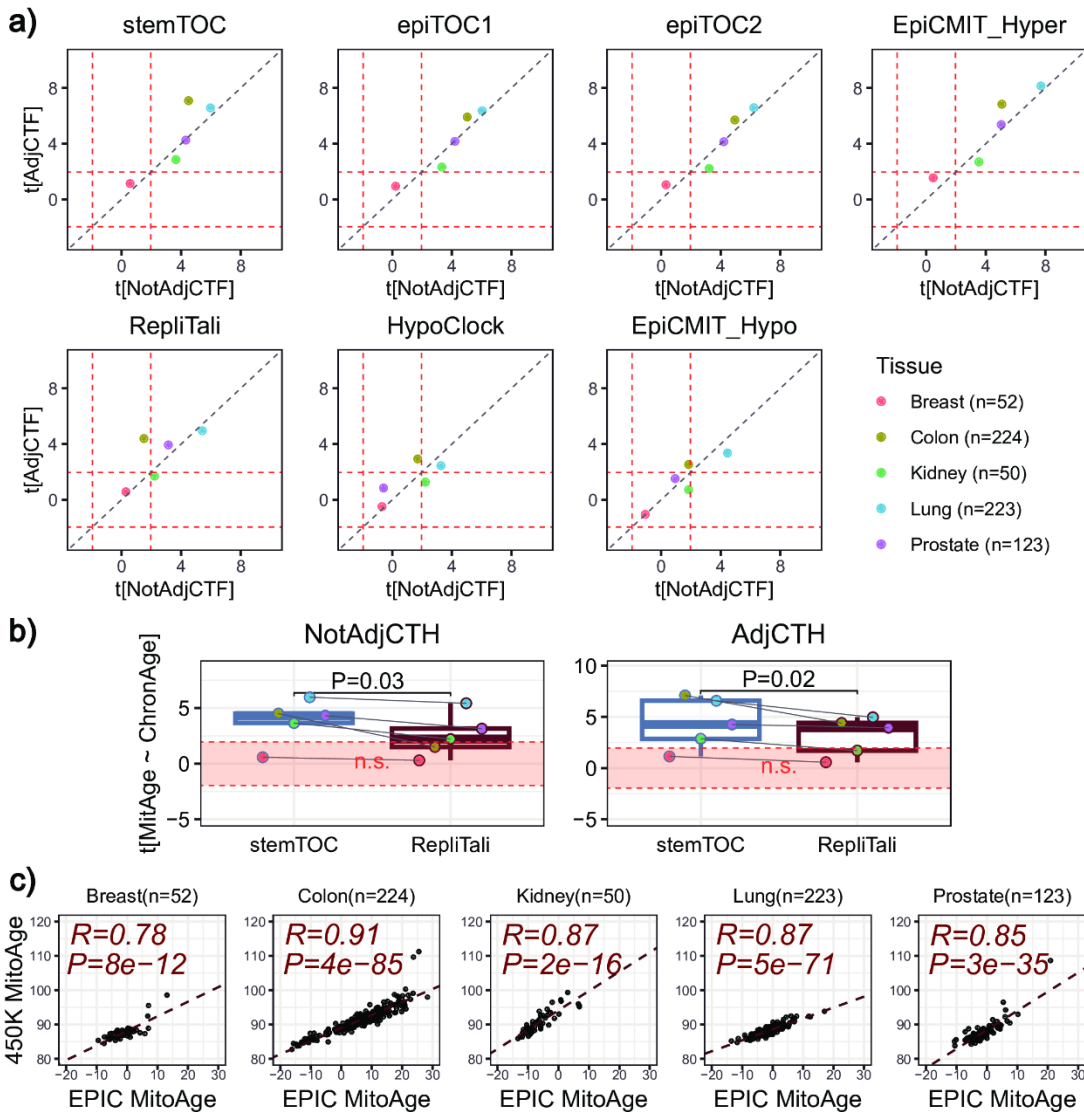
80

81 **Supplementary Figure 7: Effect of CTH on associations of mitotic age with chronological**
 82 **age in blood.** For each mitotic clock, boxplots compare the linear regression t-statistics of
 83 association between mitotic age and chronological age, not adjusting for immune cell-type
 84 fractions (NotAdjForCTF) and adjusting for 12 immune-cell type fractions (AdjForCTF). Each
 85 datapoint corresponds to one whole blood cohort, and there are a total of 18 whole blood
 86 cohorts, as indicated. The number of samples in each cohort is given in brackets after the cohort
 87 name. P-values derive from two-tailed paired Wilcoxon rank sum tests comparing the t-
 88 statistics before and after adjustment for CTFs. The line within each box denotes the median
 89 with the box itself defining the interquartile range and whiskers extending 1.5 times the
 90 interquartile range.

91

92

93



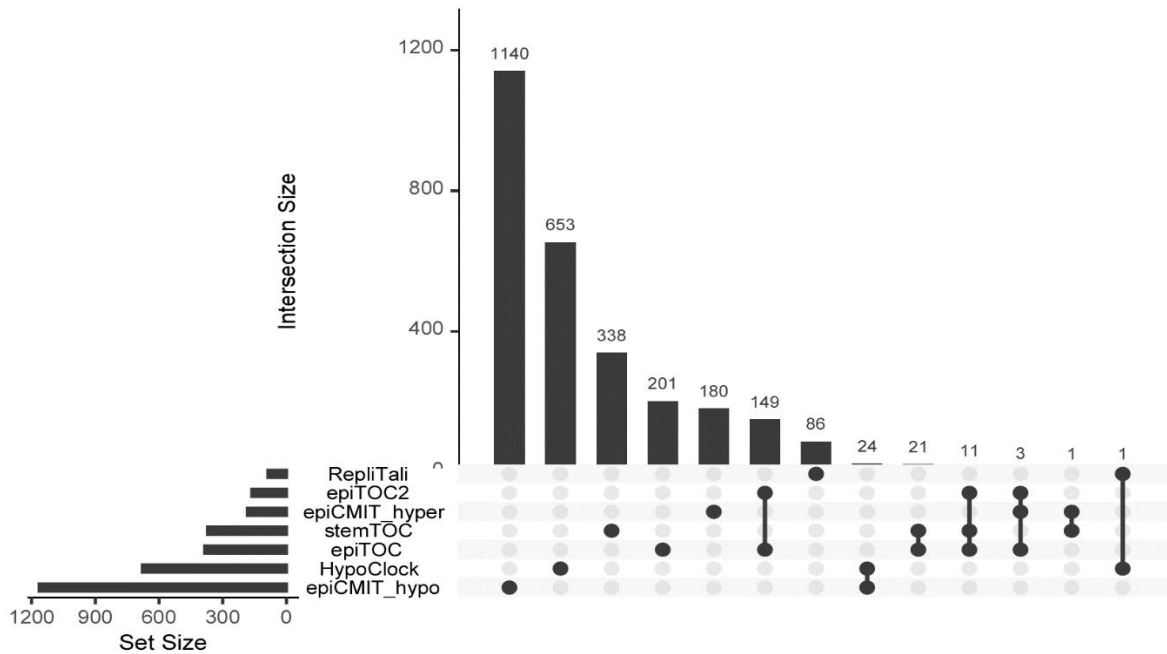
94

95 **Supplementary Figure 8: Effect of CTH on associations of mitotic age with chronological**
 96 **age in solid normal tissues from eGTEX (EPIC data).** **a)** Scatterplots of the linear regression
 97 t-statistics of association between mitotic age and chronological age, before (NotAdjCTF) and
 98 after (AdjCTF) adjustment for cell-type fractions. Each datapoint represent an eGTEX normal
 99 tissue EPIC dataset, for which the underlying cell-type fractions in the tissue could be estimated
 100 using our EpiSCORE DNAm-atlas algorithm. The red dashed-lines indicate the threshold of
 101 statistical significance ($P=0.05$). Number of samples in each dataset is given in brackets. **b)**
 102 Boxplots of the same linear regression t-statistics comparing stemTOC to RepliTali, not
 103 adjusting (left) and adjusting for CTH (right). P-values derive from a one-tailed paired t-test.
 104 The line within each box denotes the median with the box itself defining the interquartile range
 105 and whiskers extending 1.5 times the interquartile range. **c)** Scatterplots of RepliTali's Mitotic-
 106 Age in the 5 eGTEX normal tissue-datasets, computed using all available EPIC probes (x-axis)
 107 vs restricting to 450k probes only (y-axis). R-value and two-tailed correlation test P-value is
 108 given. Number of samples is given above each panel.

109

110

111



112

113 **Supplementary Figure 9: Upset plot displaying the CpG overlap of all 7 mitotic counters.**

114 Barplots to on the bottom left indicate the number of CpGs making each mitotic-counter.

115 Barplot on the top right not only indicate the number of CpGs in each counter/clock, but also

116 the overlap between various counters. Of note, if a pair of clocks display zero overlap it is not

117 shown. Thus, for instance, the overlap between RepliTali and all other clocks is zero, except

118 for HypoClock with which it shares only 1 CpG site. stemTOC has zero overlap with all

119 hypomethylated counters, has 1 CpG in common with epiCMIT-hyper, an overlap of 11 CpGs

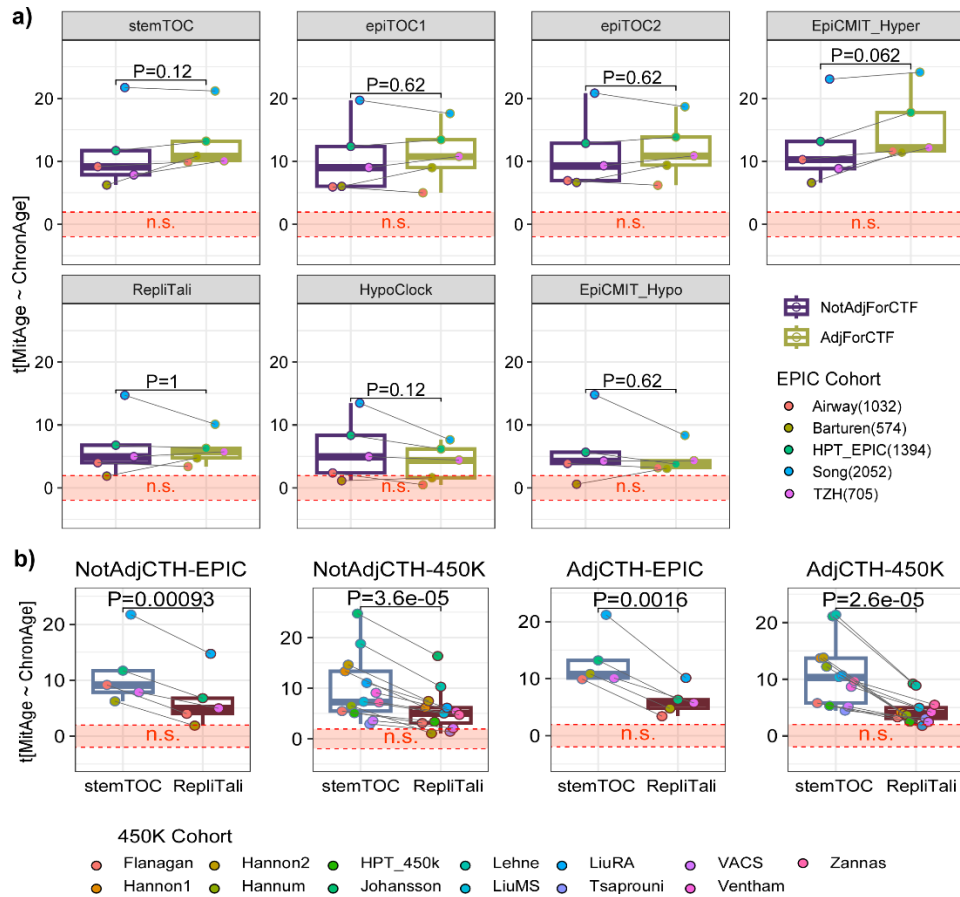
120 with epiTOC2, and an overlap of 21+11 CpGs with epiTOC.

121

122

123

124



125

126 **Supplementary Figure 10: Effect of CTH on associations of mitotic age with chronological**

127 **age in whole blood EPIC datasets. a)** For each mitotic clock, boxplots compare the linear

128 regression t-statistics of association between mitotic age and chronological age, not adjusting

129 for immune cell-type fractions (NotAdjForCTF) and adjusting for 12 immune-cell type

130 fractions (AdjForCTF) in each of 5 whole blood EPIC DNAm datasets, as indicated. P-values

131 derive from two-tailed paired Wilcoxon rank sum tests comparing the t-statistics before and

132 after adjustment for CTFs. Number of samples in each EPIC cohort is given in brackets after

133 the cohort name. Airway (n=1032)⁷, Barturen (n=574)⁸, HPT_EPIC (n=1394)⁹, Song (n=2502)

134 ¹⁰, TZH (n=705)¹¹. **b)** A direct comparison of stemTOC to RepliTali, stratified by technology

135 of DNAm dataset (EPIC or 450k) and by adjustment for cell-type heterogeneity (CTH)

136 (NotAdjCTH or AdjCTH). Here the P-values are from a one-tailed paired t-test. The line within

137 each box denotes the median with the box itself defining the interquartile range and whiskers

138 extending 1.5 times the interquartile range. Number of samples of 450k cohorts: Flanagan

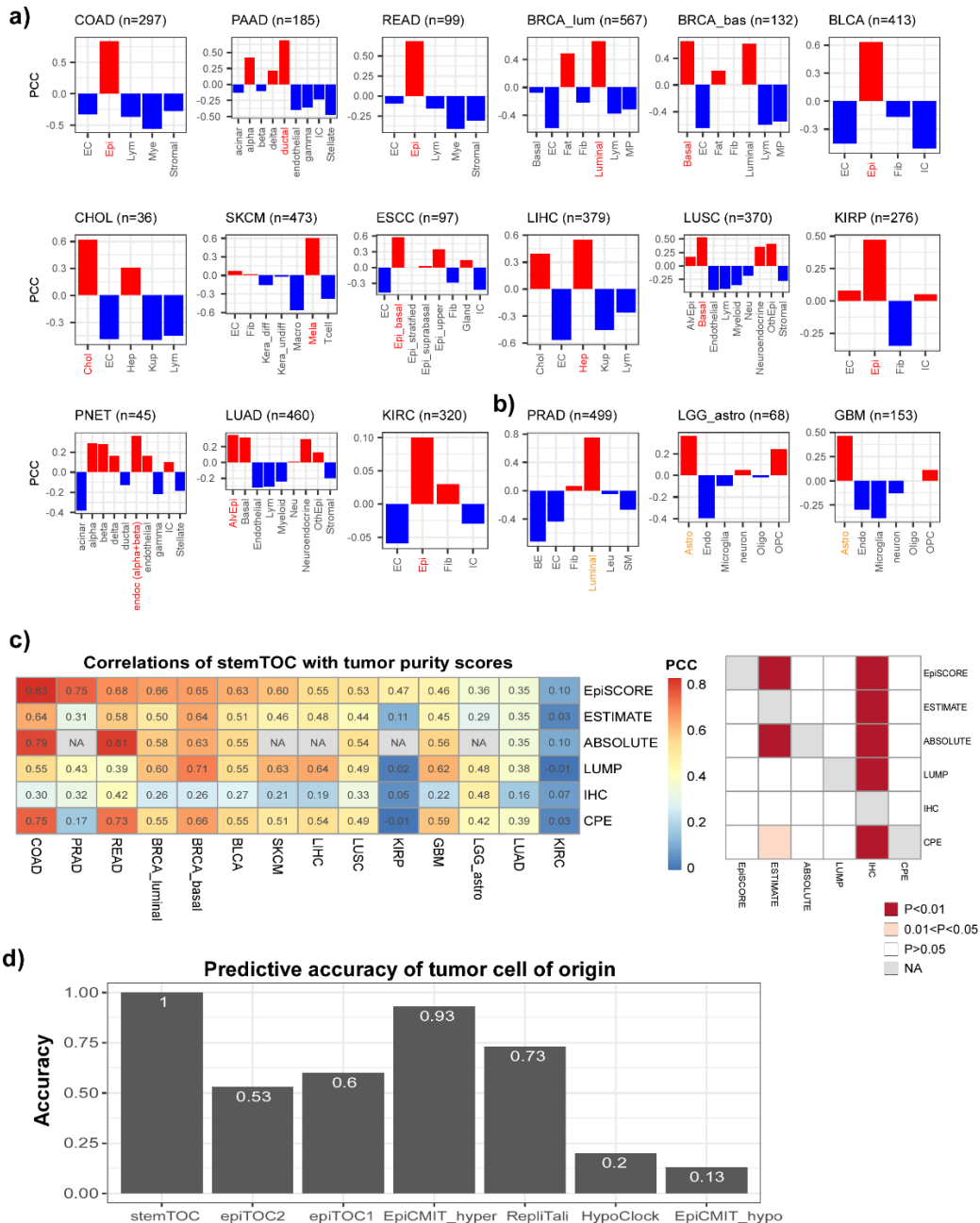
139 (n=184)¹², Hannon2 (n=665)¹³, HPT_450k (n=418)⁹, Lehne (n=2707)¹⁴, LiuRA (n=689)¹⁵,

140 VACS (n=529)¹⁶, Zannas (n=422)¹⁷, Hannon1 (n=636)¹⁸, Hannum (n=656)¹⁹, Johansson

141 (n=729)²⁰, LiuMS (n=279)²¹, Tsaprouni (n=464)²², Ventham (n=380)²³.

142

143



144

145

146

147

148

149

150

151

152

153

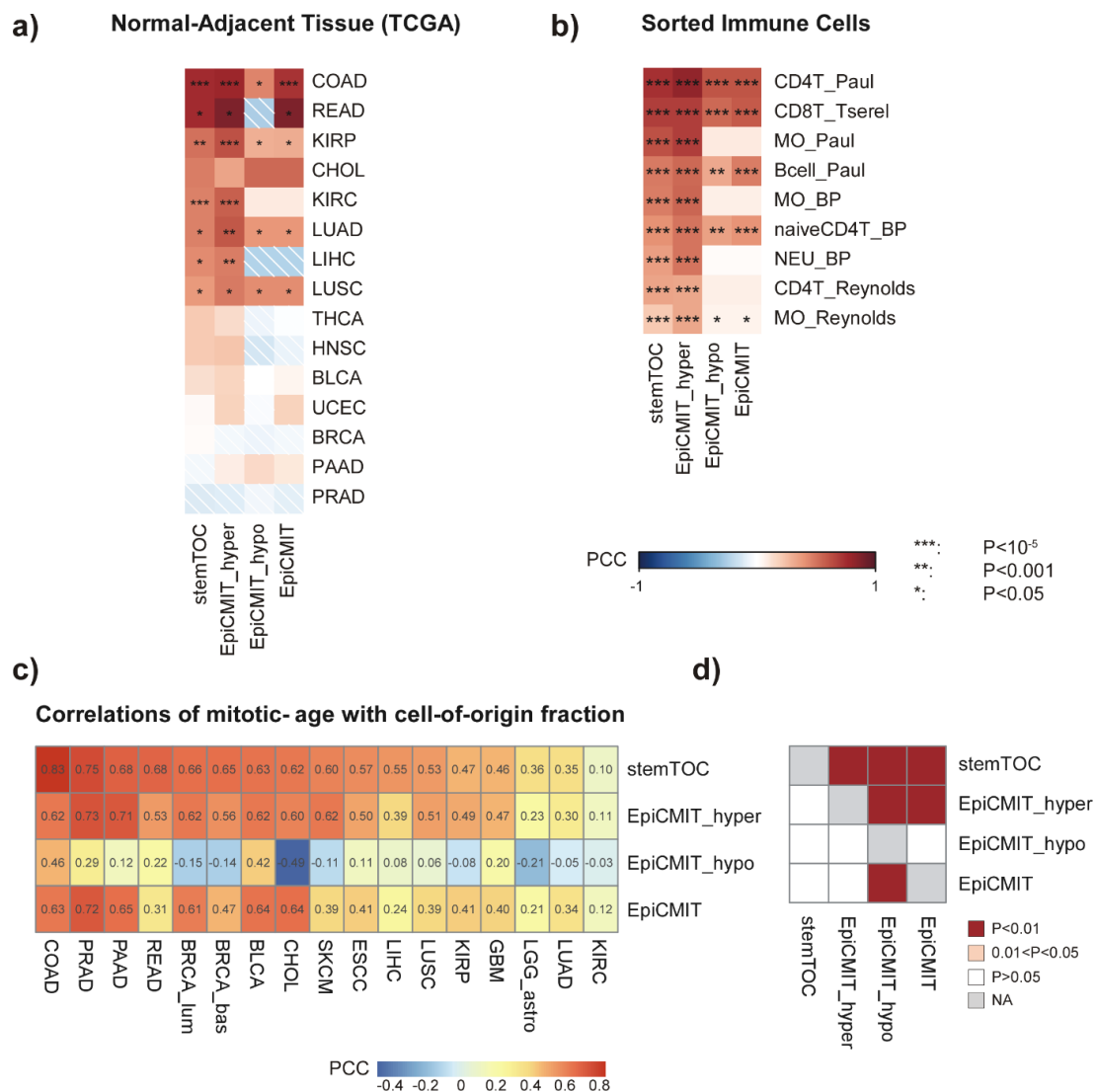
154

155

156

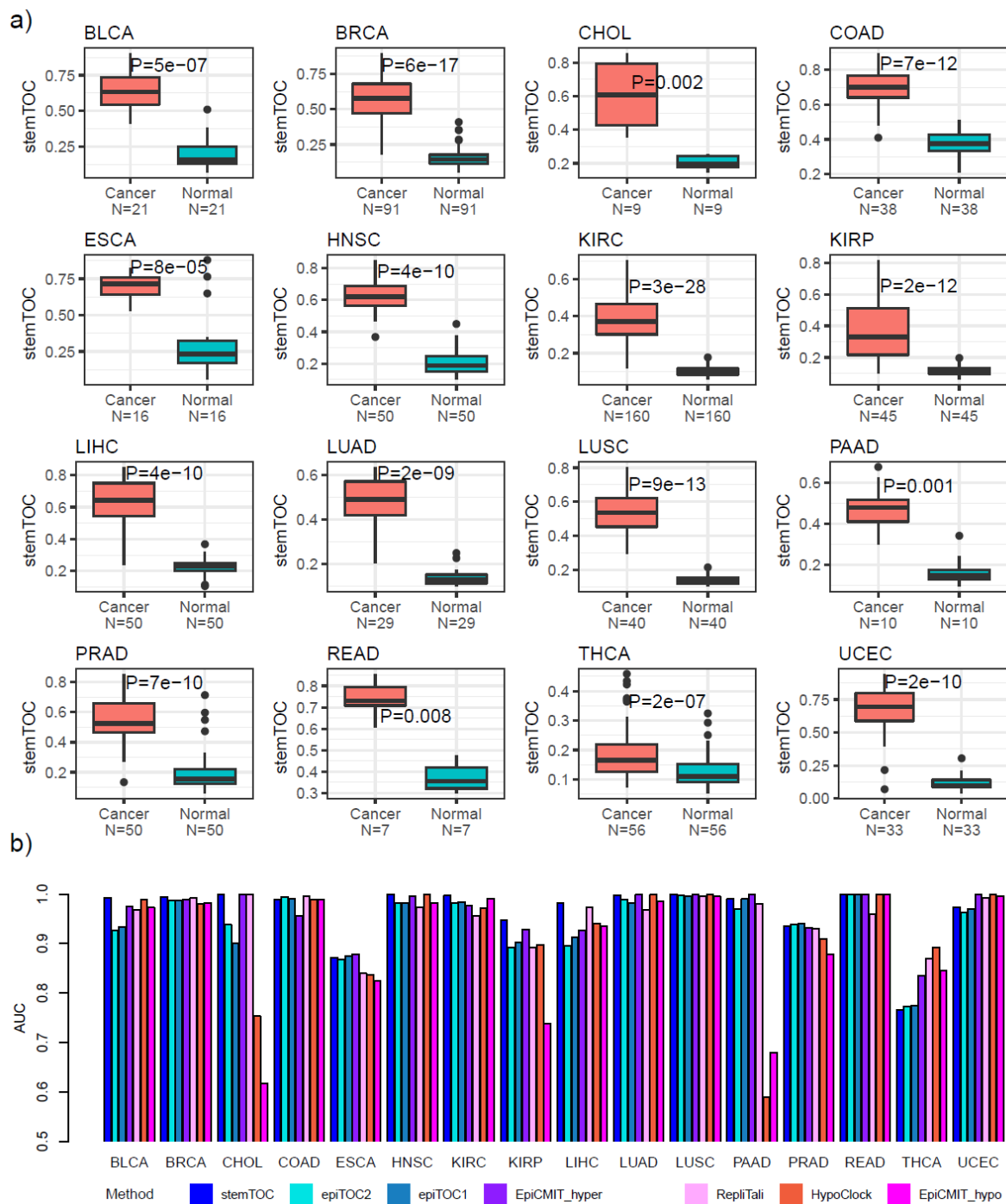
157

Supplementary Figure 11: Predicting tumor cell-of-origin. **a)** Barplots display Pearson Correlation Coefficient (PCC) between stemTOC’s mitotic age and the corresponding cell-type fraction for each of 15 cancer-types where the tumor cell-of-origin (text in red) is fairly well established. **b)** As a) but for 3 TCGA cancer-types where the tumor cell-of- origin (text in orange) is less well established. Number of samples given above each panel. **c)** Left: Heatmap of PCCs between stemTOC’s mitotic age and tumor purity indices, including EpiSCORE’s tumor cell of origin fraction, ESTIMATE, IHC, ABSOLUTE, CPE and LUMP. Right: Heatmap of one-tailed Wilcoxon rank sum test P-values comparing the PCCs obtained with each tumor purity method. A significant P-value for method “Y” on y-axis against method “X” on x-axis means significantly stronger PCCs with stemTOC’s mitotic age for method “Y” compared to method “X”. **d)** Overall prediction accuracy of 7 mitotic clocks for correctly predicting the tumor cell-of-origin, as assessed over the 15 cancer-types in a). RepliTali results are for the probes restricted to 450k beadarrays.



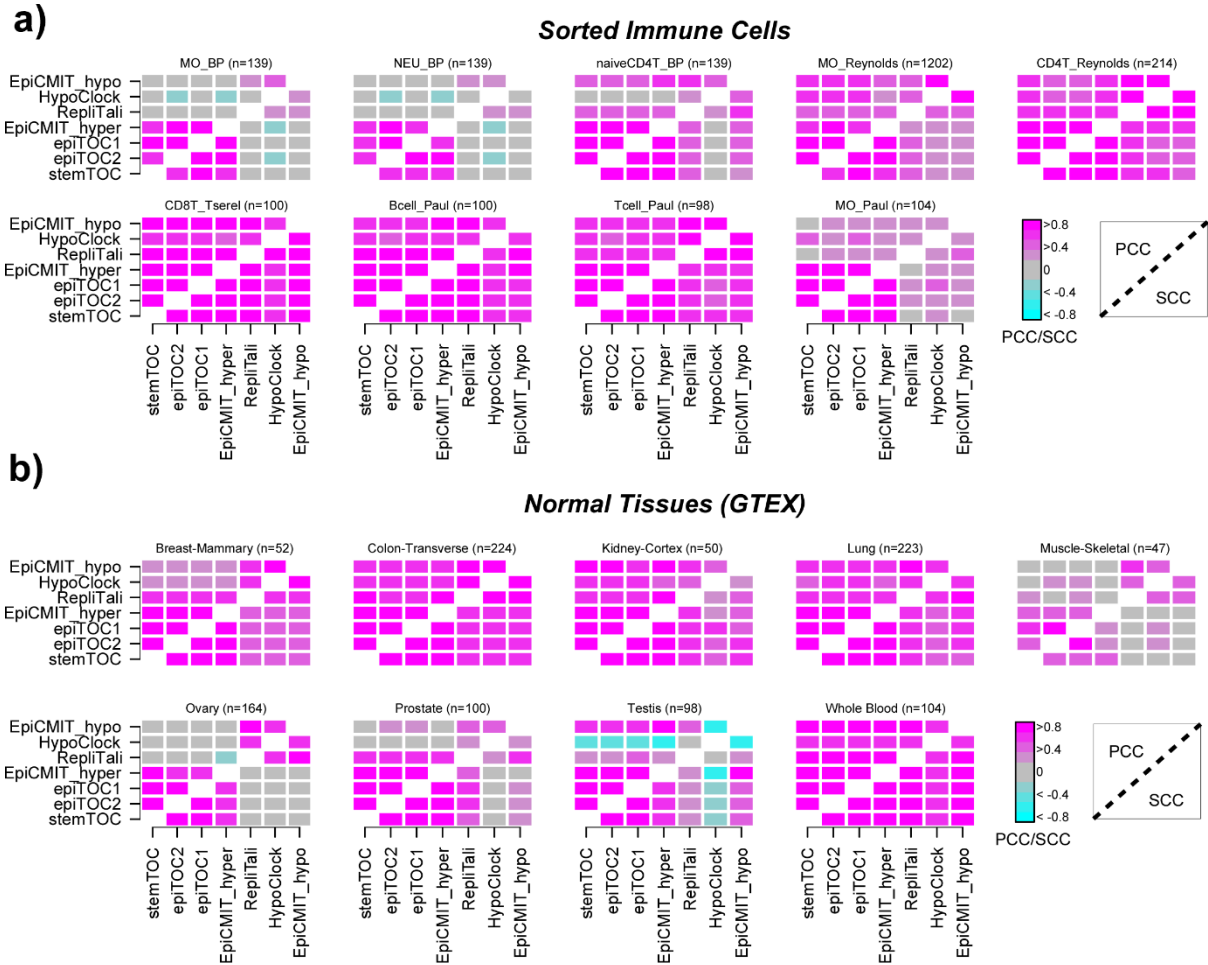
158

159 **Supplementary Figure 12: epiCMIT-hyper outperforms epiCMIT.** **a)** Pearson Correlation
 160 Coefficient (PCC) heatmap of correlations by 4 mitotic clocks (stemTOC, epiCMIT-hyper,
 161 epiCMIT-hypo, epiCMIT) with chronological in the normal-adjacent samples of TCGA cancer-
 162 types. Number of normal-adjacent samples: COAD(n=38), READ(n=7), KIRP(n=45),
 163 CHOL(n=9), KIRC(n=160), LUAD(n=32), LIHC(n=50), LUSC(n=42), THCA(n=56),
 164 HNSC(n=50), BLCA(n=21), UCEC(n=46), BRCA(n=97), PAAD(n=10), PRAD(n=50). **b)** As
 165 a) but for sorted immune-cell datasets. Sample numbers: CD4T_Paul(n=98),
 166 CD8T_Tserel(n=98), MO_Paul(n=104), Bcell_Paul(n=100), MO_BP(n=139),
 167 naiveCD4T_BP(n=139), NEU_BP(n=139), CD4T_Reynolds(n=214), MO_Reynolds(n=1202).
 168 **c)** As a) but now for correlations of the mitotic clocks with tumor cell of origin fraction (as a
 169 proxy for tumor purity). **d)** Wilcoxon rank sum paired test one-tailed P-values comparing the
 170 PCC values displayed in c) of one clock to another. Convention is that a significant P-value
 171 means that the mitotic clock labelled by the row outperforms the one displayed in the column.



172
 173
 174
 175
 176
 177
 178
 179
 180
 181
 182

Supplementary Figure 13: Mitotic counters discriminate cancer from normal-adjacent tissue. **a)** Boxplots of stemTOC's mitotic age (y-axis) using only tumor normal-adjacent pairs for TCGA cancer types with sufficient numbers of normal samples. P-value derives from a one-tailed paired Wilcoxon rank sum test. Number of samples in each groups is given below boxplot. The line within each box denotes the median with the box itself defining the interquartile range and whiskers extending 1.5 times the interquartile range. **b)** Corresponding AUC-values for all mitotic clocks. RepliTali results are for the probes restricted to 450k beadarrays.

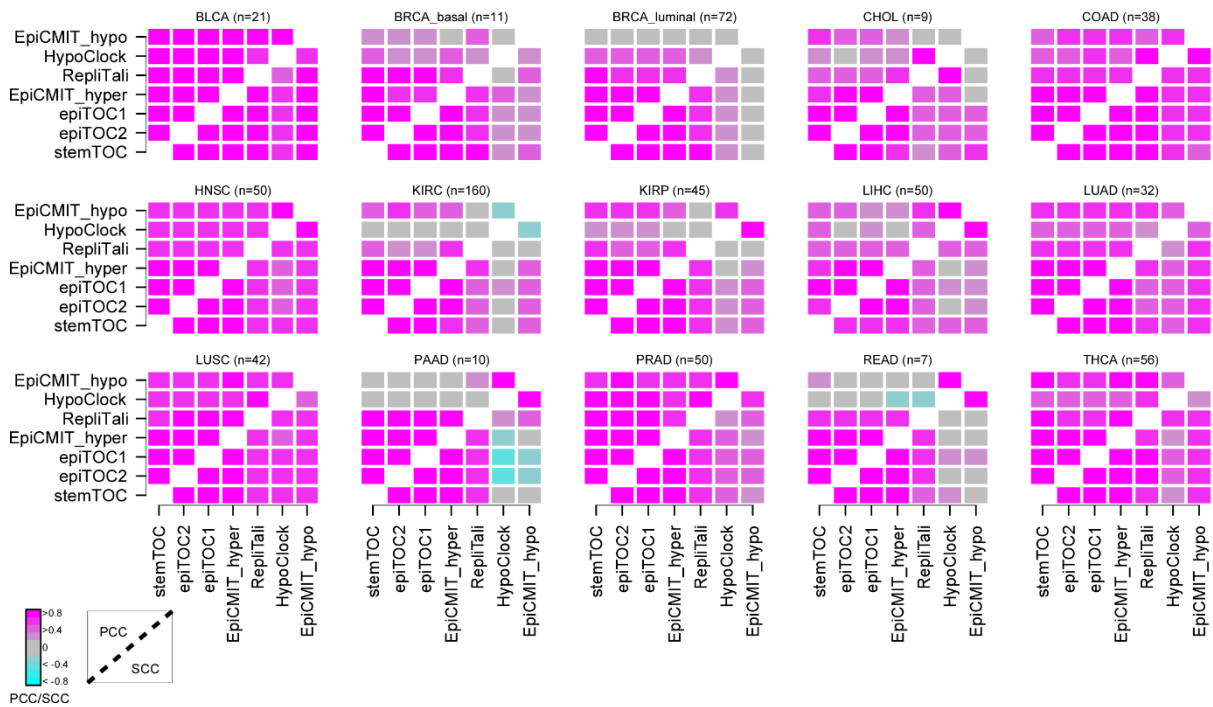


183

184 **Supplementary Figure 14: Correlation heatmaps of mitotic counters in sorted immune**
 185 **cells and normal tissues of GTEX. a)** For each dataset profiling sorted immune cells we
 186 display a heatmap of Pearson (upper triangular part, PCC) and Spearman (lower triangular part,
 187 SCC) correlation coefficients between each pair of mitotic counters. The cell-type and number
 188 of samples in each dataset are given above each heatmap. MO=monocyte, NEU=neutrophil,
 189 Tcell=CD4+ T-cell. CD4T=CD4+ T-cell, CD8T=CD8+ T-cell, naiveCD4T=naïve CD4+ T-cell.
 190 RepliTali results are for the probes restricted to 450k beadarrays. **b)** As a) but for the normal-
 191 tissue datasets from GTEX (EPIC data).

192

NormalAdj (TCGA)



193

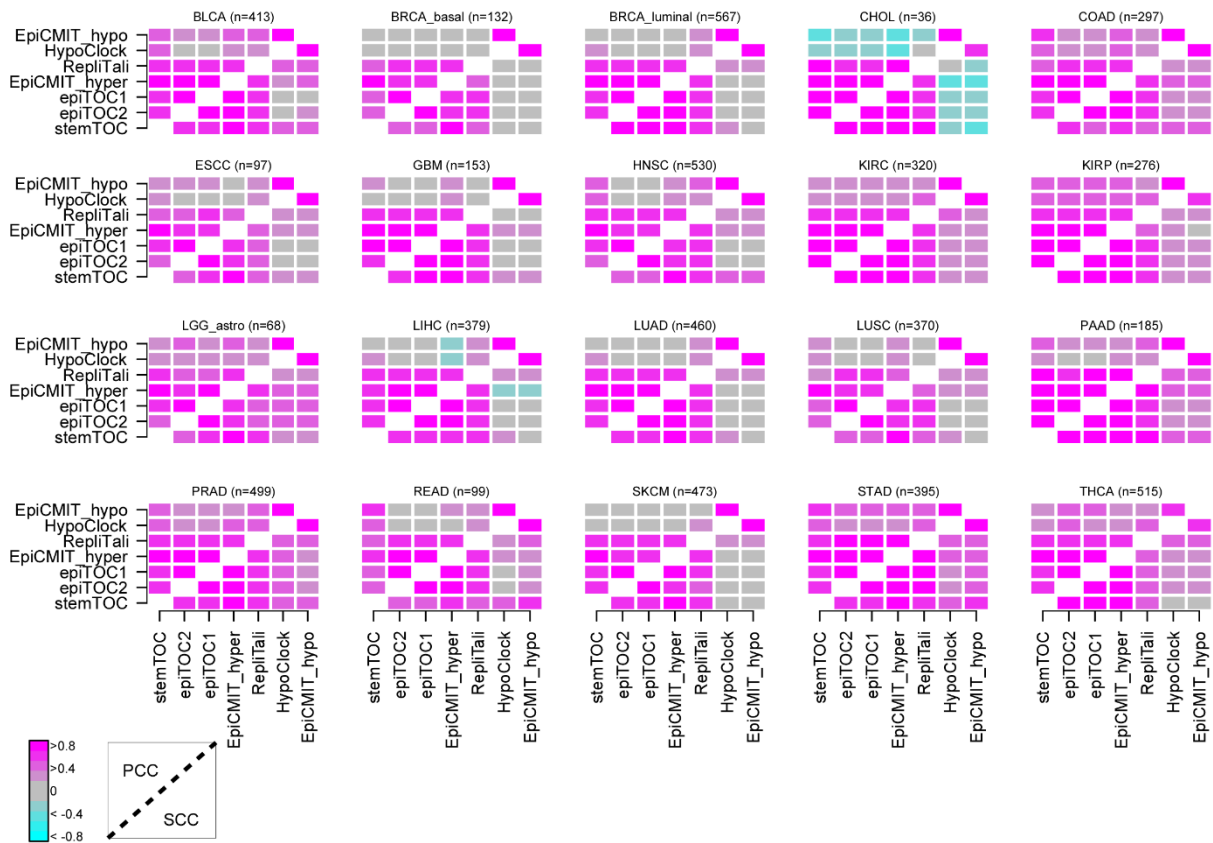
194 **Supplementary Figure 15: Correlation heatmaps of mitotic counters in the normal-**
 195 **adjacent samples from the TCGA.** For each set of normal-adjacent samples we display a
 196 heatmap of Pearson (upper triangular part, PCC) and Spearman (lower triangular part, SCC)
 197 correlation coefficients between each pair of mitotic counters. The cancer-type matched to the
 198 normal-adjacent tissue and number of normal-adjacent samples in each dataset are given above
 199 each heatmap. RepliTali results are for the probes restricted to 450k beadarrays.

200

201

202

Cancer (TCGA)



203

PCC/SCC

204 **Supplementary Figure 16: Correlation heatmaps of mitotic counters in the tumor samples**
 205 **from the TCGA.** For each TCGA cancer-type we display a heatmap of Pearson (upper
 206 triangular part, PCC) and Spearman (lower triangular part, SCC) correlation coefficients
 207 between each pair of mitotic counters. The cancer-type and number of tumor samples in each
 208 dataset are given above each heatmap. RepliTali results are for the probes restricted to 450k
 209 beadarrays.

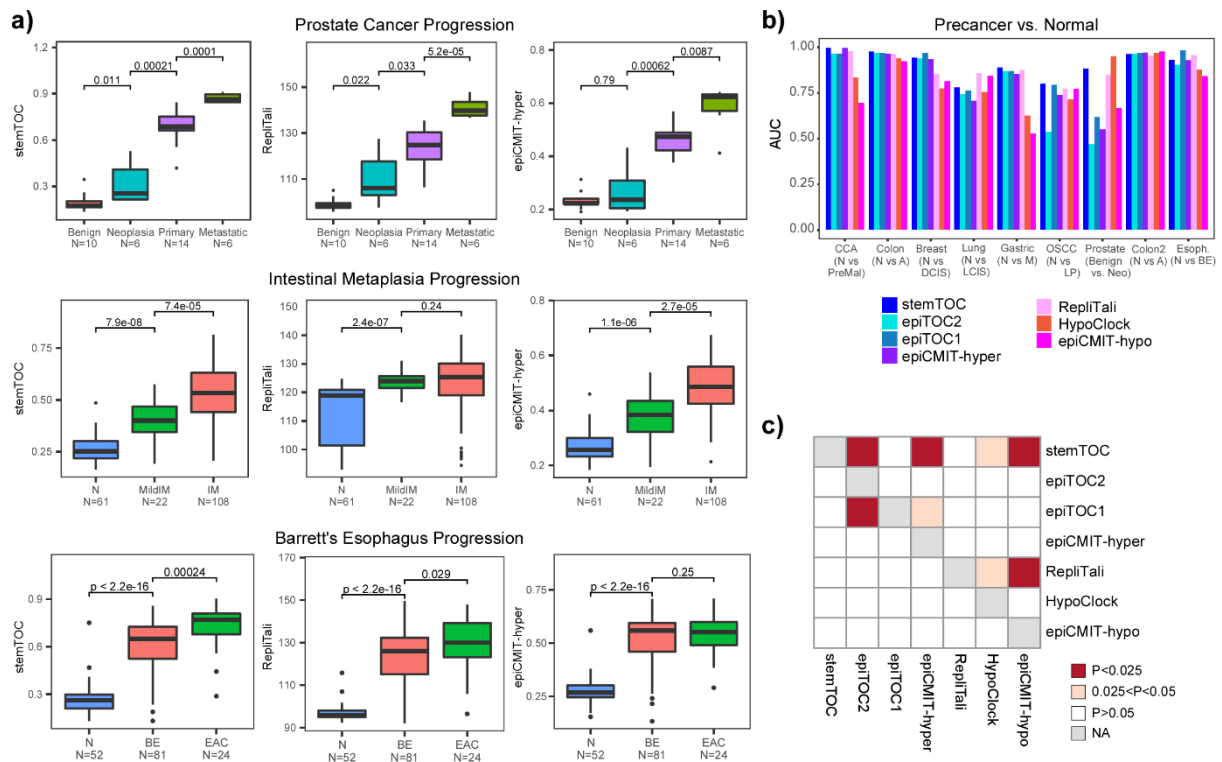
210

211

212

213

214



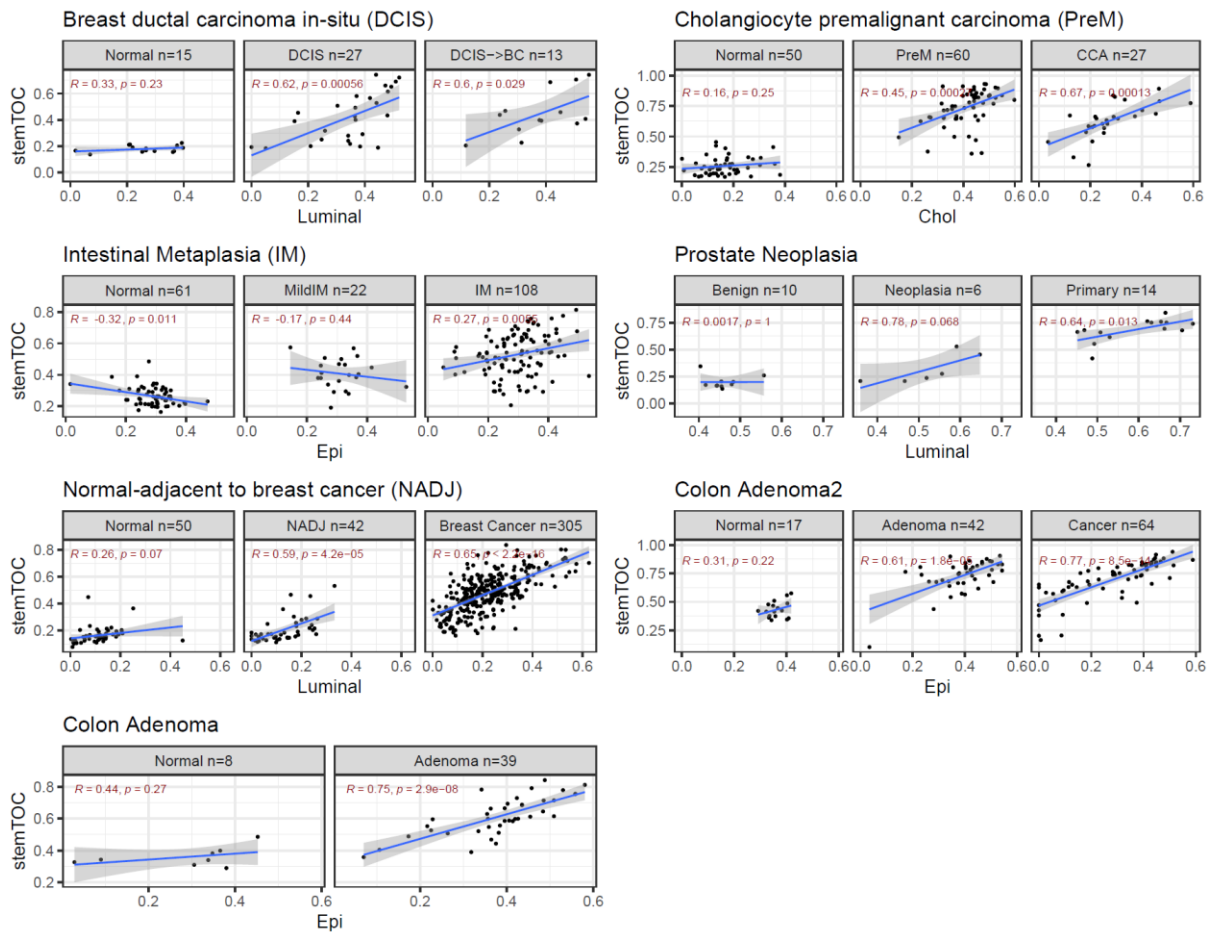
215

216 **Supplementary Figure 17: Benchmarking of stemTOC in precancerous conditions . a)**
 217 Mitotic-age estimates for stemTOC, RepliTali and epiCMT-hyper in three different DNAm
 218 datasets encompassing normal healthy tissue and precancerous lesions, including prostate
 219 cancer progression (Benign, Neoplasia, Primary and Metastasis), progression of intestinal
 220 metaplasia (N=normal, MildIM=mild intestinal metaplasia, IM=intestinal metaplasia) and
 221 esophageal adenocarcinoma (N=normal healthy squamous, BE=Barrett's Esophagus,
 222 EAC=esophageal adenocarcinoma). Number of samples in each group is indicated. Number of
 223 samples in each group is shown below boxplot. We provide one-tailed P-values from a
 224 Wilcoxon rank sum test comparing successive stages. The line within each box denotes the
 225 median with the box itself defining the interquartile range and whiskers extending 1.5 times
 226 the interquartile range. **b)** Barplot displaying the AUC from the Wilcoxon test comparing
 227 normal-healthy to normal at-risk groups across a total 9 different DNAm datasets, and for 7
 228 different mitotic clocks. Number of normal/precancerous samples: CCA (Liver preceding
 229 cholangiocarcinoma) (50/60), Colon (9/38), Breast (15/40), Lung (21/35), Gastric (61/130),
 230 OSCC (Oral tissue preceding oral squamous cell carcinoma) (18/8), Prostate (10/6), Colon2
 231 (41/42), Esophagus (52/81). **c)** Heatmap displaying one-tailed paired Wilcoxon rank sum test
 232 P-values, comparing clocks to each other, in how well their mitotic age distinguished normal-
 233 healthy from normal at-cancer-risk tissue. Each row indicates how well the corresponding
 234 clock's mitotic age estimate performs in relation to the clock specified by the column. The
 235 paired Wilcoxon test is performed over the 9 datasets. RepliTali results in 450k datasets are for
 236 the probes restricted to 450k beadarrays.

237

238

239

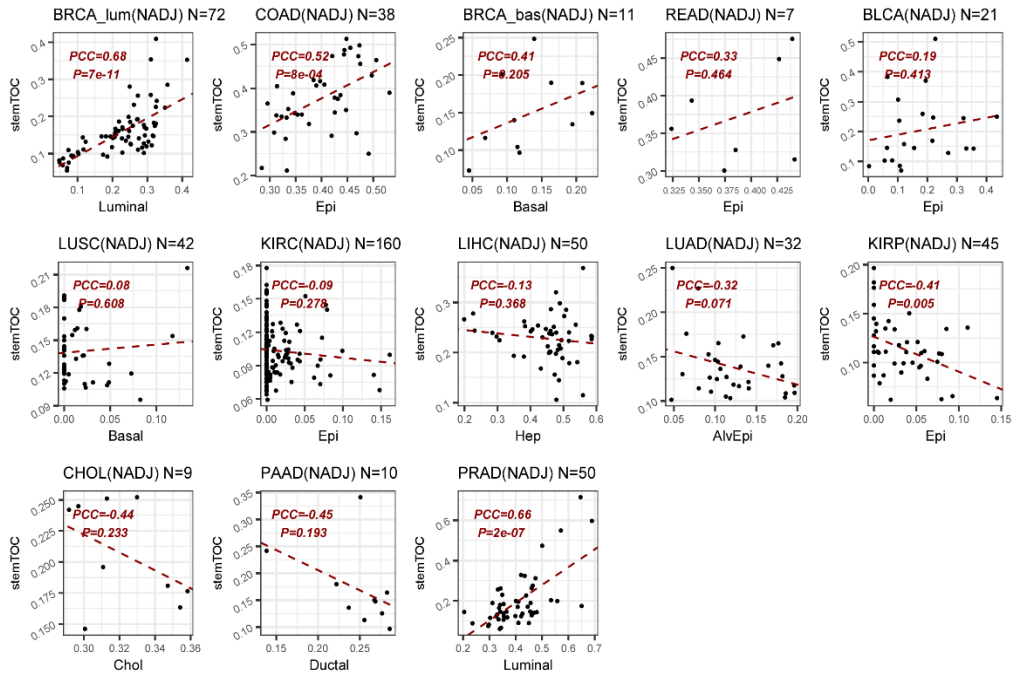


240

241 **Supplementary Figure 18: Correlations of stemTOC's mitotic age with tumor cell-of-**
 242 **origin fraction in precancerous lesions.** Scatterplots of stemTOC's mitotic age (y-axis) vs the
 243 EpiSCORE-estimated fraction of the presumed cell-of-origin for a number of Illumina
 244 EPIC/450k DNAm datasets profiling histologically normal and precancerous lesions, as
 245 indicated. In each scatterplot we give the R-value and P-value from a linear regression. The
 246 number of samples is indicated above the plot. Regression line with standard error interval is
 247 shown.

248

249

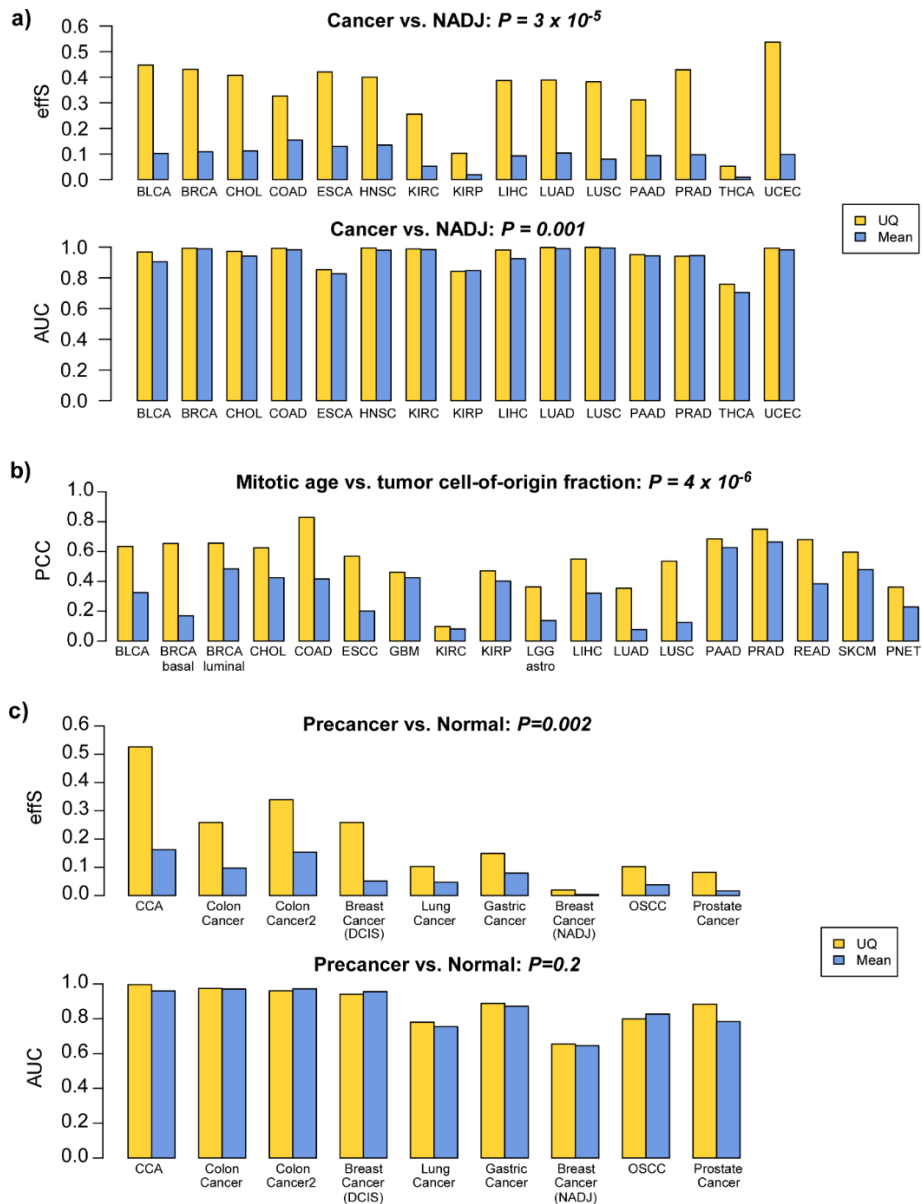


250

251 **Supplementary Figure 19: Correlations of stemTOC's mitotic age with tumor cell-of-**
 252 **origin fraction in the normal-adjacent tissues of the TCGA.** Scatterplots of stemTOC's
 253 mitotic age (y-axis) vs the EpiSCORE-estimated fraction of the presumed cell-of-origin in
 254 normal-adjacent (NADJ) tissue datasets from the TCGA. Only those with reasonable numbers
 255 of NADJ samples were included. In each scatterplot we give the R-value and P-value from a
 256 linear regression. The number of samples is indicated above the plot. Regression line is shown.

257

258



259

260

261

262

263

264

265

266

267

268

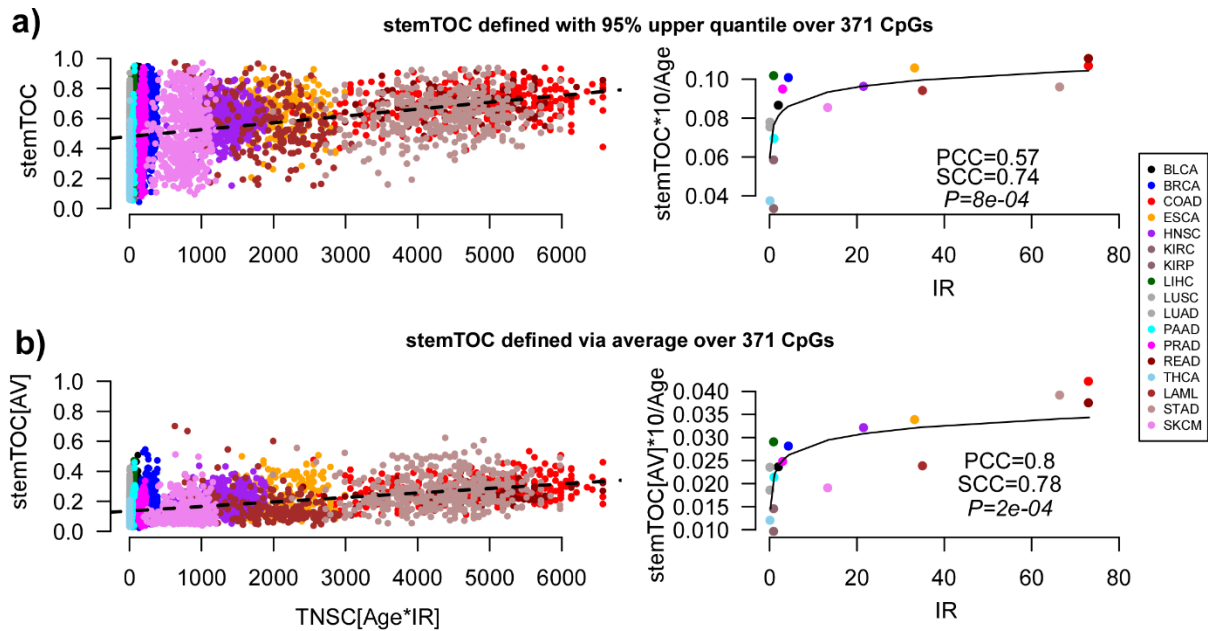
269

270

271

272

Supplementary Figure 20: Accounting for stochasticity improves associations of mitotic age. **a)** Barplots depict the effect size (effS) of the average mitotic age difference between cancer and normal-adjacent tissue for TCGA cancer-types, and with the mitotic age computed using a 95% upper quantile (UQ, stemTOC) or the mean (Mean) over the 371 mitotic-CpGs. Lower barplots display the corresponding AUCs. For both barplots, we provide the one-tailed P-value from a paired Wilcoxon test comparing UQ to mean. **b)** Barplots depicts the Pearson Correlation Coefficient (PCC) between mitotic age and tumor cell of origin fraction across TCGA cancer-types. We provide the one-tailed P-value from a paired Wilcoxon test comparing the PCCs from using UQ (stemTOC) to those using the mean. **c)** Upper barplots depict the effect size (effS) of the average mitotic age difference between normal-healthy and normal “at-cancer-risk” tissue for various cancer-types. Barplots below display the corresponding AUCs. For both sets of barplots, we provide the one-tailed P-value from a paired Wilcoxon test comparing UQ to mean.



273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

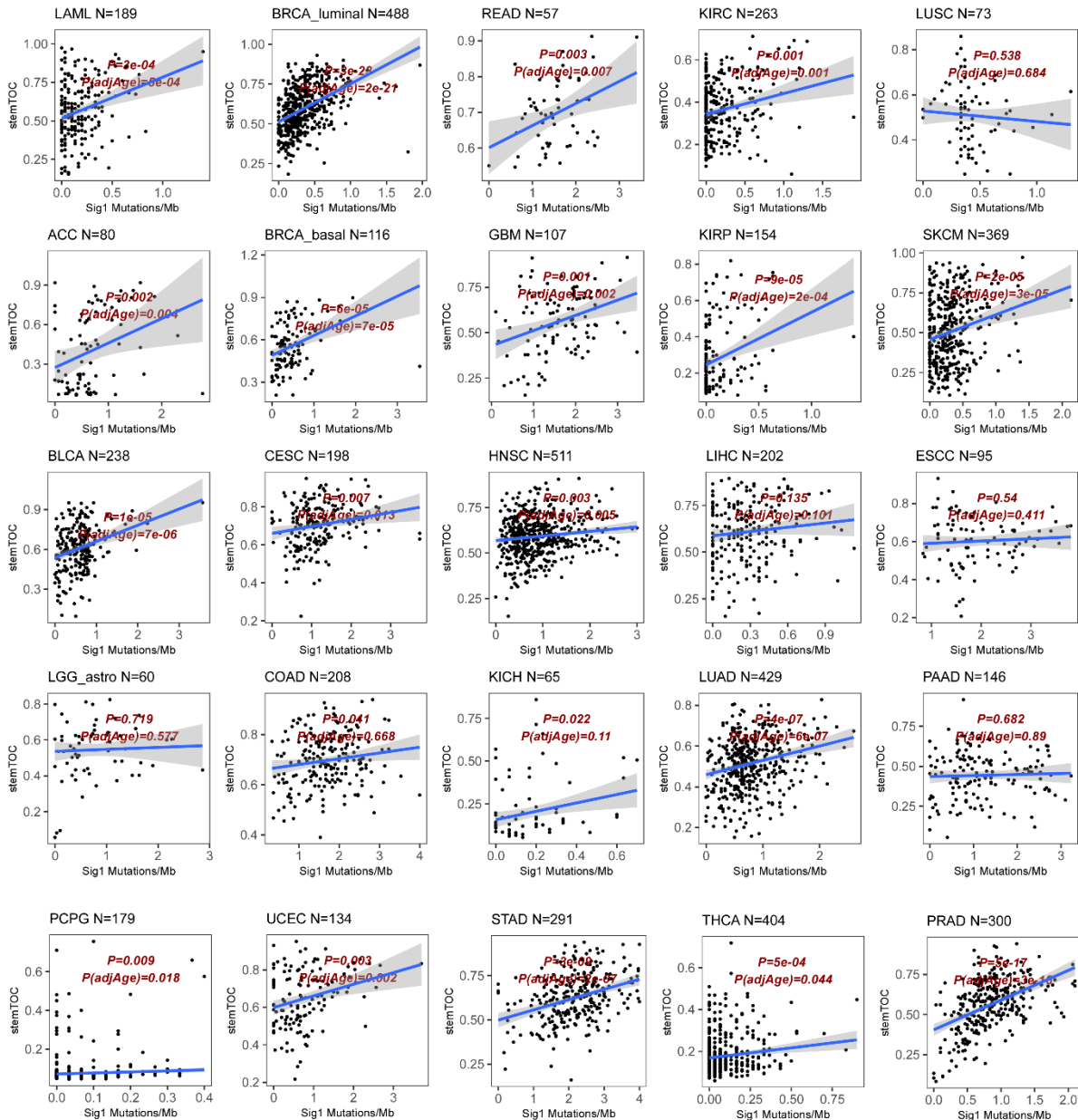
295

296

297

298

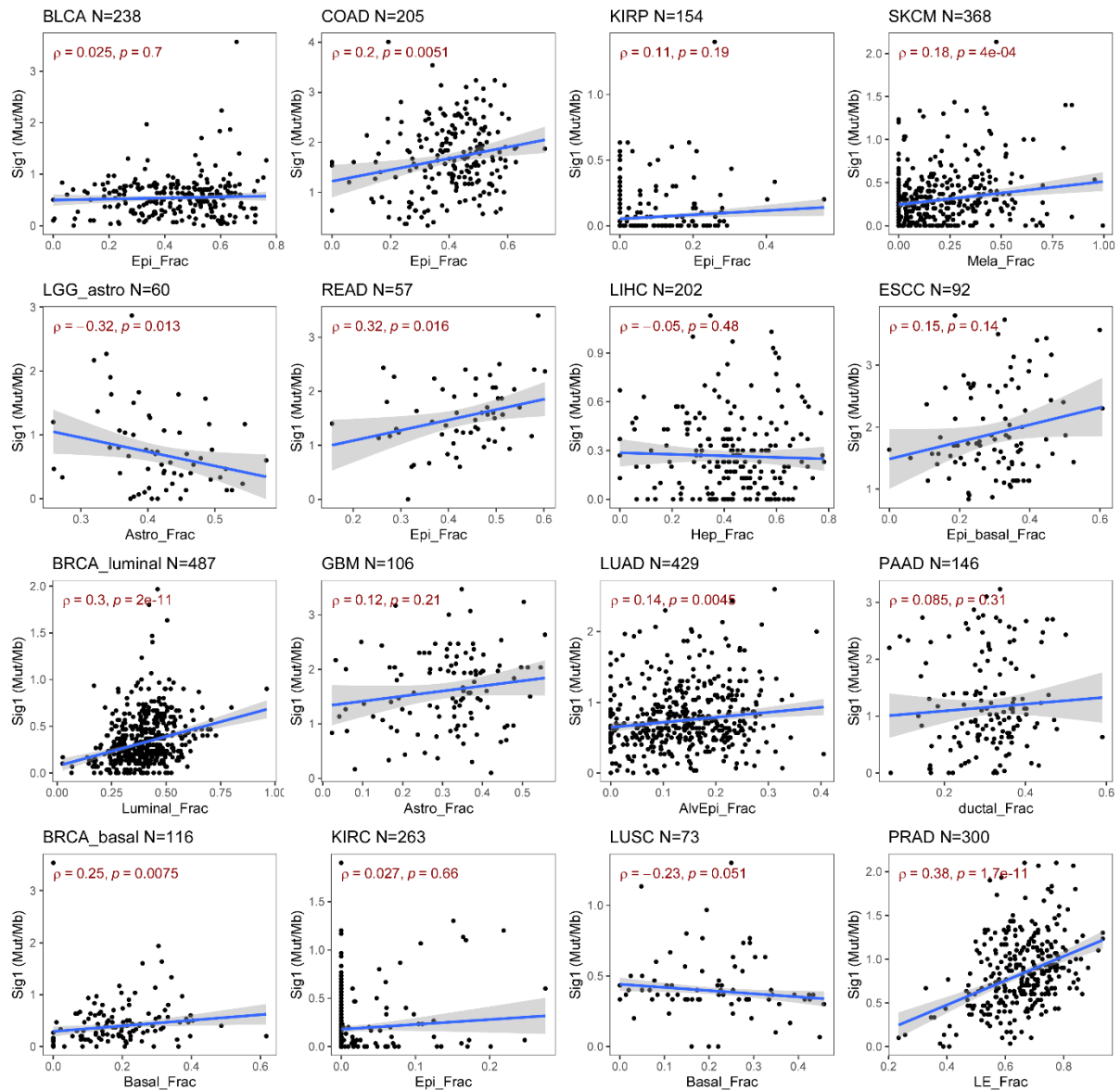
Supplementary Figure 21: Saturation effect of stemTOC is not dependent on taking an upper quantile. **a) Left:** Scatterplot of stemTOC (defined using the 95% upper quantile) (y-axis) against the estimated total number of stem-cell divisions (Age*intrinsic annual rate of stem-cell division of the corresponding normal-tissue (IR), x-axis) for >7000 cancer samples from 17 TCGA cancer-types. Only cancer-types with an IR estimate in normal tissue were used. **Right:** Median value of age-adjusted stemTOC value (multiplied by 10 to reflect change over a decade) for each cancer-type vs the annual intrinsic rate of stem-cell division of the corresponding normal tissue-type. Both Pearson (PCC) and Spearman (SCC) correlation coefficients are given. P-value tests for significance of SCC. Fitted line is that of a best fit among linear and log-linear models. **b) As a),** but defining stemTOC as the average DNAm over the 371 stemTOC CpGs. Sample sizes: BLCA(n=413), BRCA(n=699), COAD(n=297), ESCA(n=186), HNSC(n=530), KIRC(n=320), KIRP(n=276), LIHC(n=379), LUSC(n=370), LUAD(n=460), PAAD(n=185), PRAD(n=499), READ(n=99), THCA(n=515), LAML(n=194), STAD(n=395), SKCM(n=473).



300
 301 **Supplementary Figure 22: stemTOC vs MS1 in individual TCGA cancer-types.** Scatterplot
 302 of stemTOC's mitotic age vs MS1 mutational load (mutations per Mb) for 25 TCGA cancer-
 303 types, as shown. Number of cancer samples is shown above plots. In each panel, we display
 304 the P-values of a linear regression of stemTOC's mitotic age vs the mutational load, adjusting
 305 for age (P(adjAge)) and not adjusting for age (P). Regression line with standard error interval
 306 is shown.

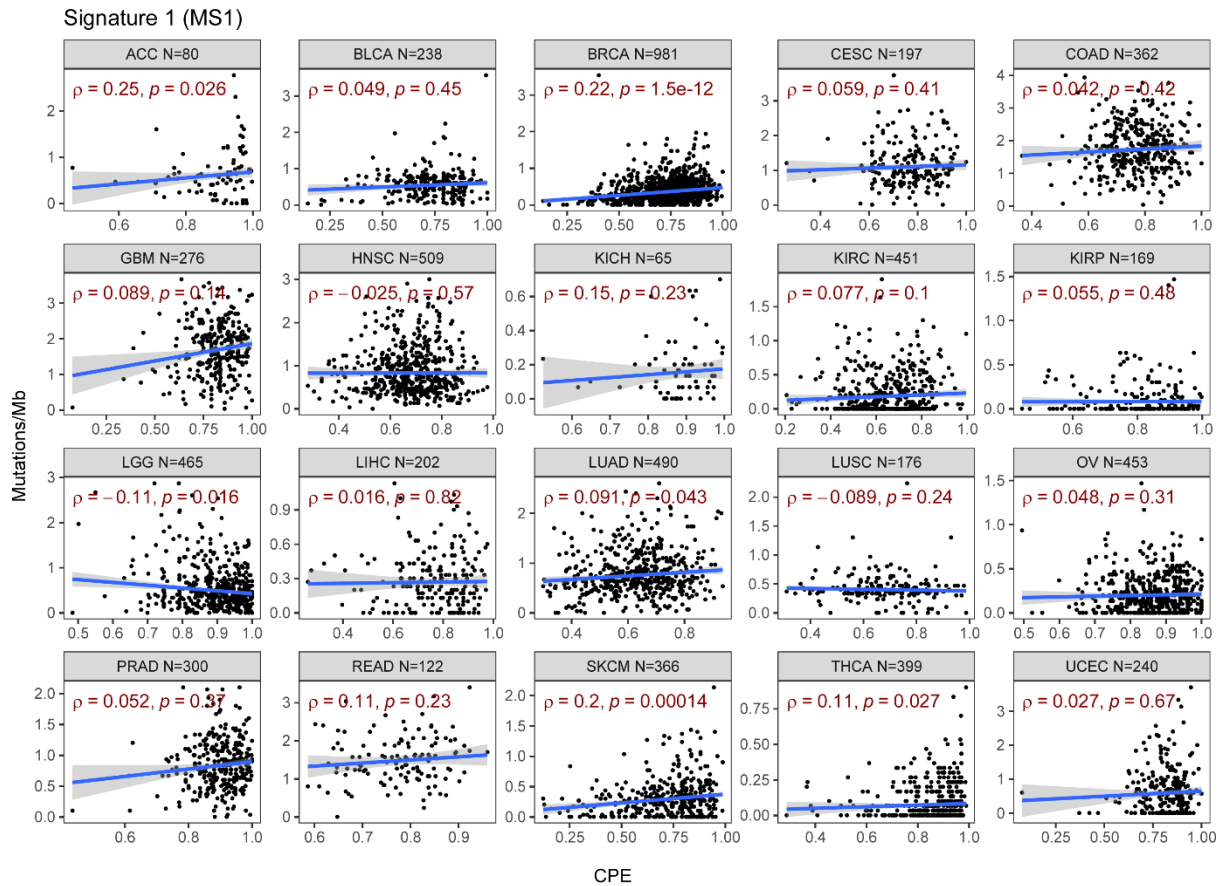
307

308



309

310 **Supplementary Figure 23: MS1 load vs tumor cell of origin fraction.** Scatterplot of MS1
 311 mutational load (mutations per Mb) vs tumor cell of origin fraction (as estimated using the
 312 EpiSCORE algorithm), for 16 TCGA cancer-types, as shown. Number of cancer samples is
 313 shown above plots. In each panel, we display Spearman's correlation coefficient and P-value.
 314 Regression line with standard error interval is shown.



315
 316 **Supplementary Figure 24: MS1 load does not generally correlate with tumor purity.**
 317 Scatterplot of MS1 mutational load (mutations per Mb) vs tumor purity (as estimated using the
 318 CPE algorithm), for 20 TCGA cancer-types, as shown. Number of cancer samples is shown
 319 above plots. In each panel, we display the Spearman Correlation Coefficient and P-value.
 320 Regression line with standard error interval is shown.

321
 322
 323
 324
 325
 326
 327
 328
 329
 330
 331
 332
 333
 334
 335

336 SUPPLEMENTARY REFERENCES

337

- 338 1. Kocher, K. *et al.* Genome-wide neonatal epigenetic changes associated with maternal exposure to the
339 COVID-19 pandemic. *BMC Med Genomics* **16**, 268 (2023).
- 340 2. Petroff, R.L. *et al.* Translational toxicoepigenetic Meta-Analyses identify homologous gene DNA
341 methylation reprogramming following developmental phthalate and lead exposure in mouse and
342 human offspring. *Environ Int* **186**, 108575 (2024).
- 343 3. Paul, D.S. *et al.* Increased DNA methylation variability in type 1 diabetes across three immune effector
344 cell types. *Nat Commun* **7**, 13555 (2016).
- 345 4. Chen, L. *et al.* Genetic Drivers of Epigenetic and Transcriptional Variation in Human Immune Cells. *Cell*
346 **167**, 1398-1414 e24 (2016).
- 347 5. Oliva, M. *et al.* DNA methylation QTL mapping across diverse human tissues provides molecular links
348 between genetic variation and complex traits. *Nat Genet* **55**, 112-122 (2023).
- 349 6. Moss, J. *et al.* Comprehensive human cell-type methylation atlas reveals origins of circulating cell-free
350 DNA in health and disease. *Nat Commun* **9**, 5068 (2018).
- 351 7. Robinson, O. *et al.* Determinants of accelerated metabolomic and epigenetic aging in a UK cohort. *Aging*
352 *Cell* **19**, e13149 (2020).
- 353 8. Barturen, G. *et al.* Whole blood DNA methylation analysis reveals respiratory environmental traits
354 involved in COVID-19 severity following SARS-CoV-2 infection. *Nat Commun* **13**, 4597 (2022).
- 355 9. Shang, L. *et al.* meQTL mapping in the GENOA study reveals genetic determinants of DNA methylation
356 in African Americans. *Nat Commun* **14**, 2711 (2023).
- 357 10. Song, N. *et al.* Persistent variations of blood DNA methylation associated with treatment exposures and
358 risk for cardiometabolic outcomes in long-term survivors of childhood cancer in the St. Jude Lifetime
359 Cohort. *Genome Med* **13**, 53 (2021).
- 360 11. You, C. *et al.* A cell-type deconvolution meta-analysis of whole blood EWAS reveals lineage-specific
361 smoking-associated DNA methylation changes. *Nat Commun* **11**, 4779 (2020).
- 362 12. Flanagan, J.M. *et al.* Temporal stability and determinants of white blood cell DNA methylation in the
363 breakthrough generations study. *Cancer Epidemiol Biomarkers Prev* **24**, 221-9 (2015).
- 364 13. Hannon, E. *et al.* DNA methylation meta-analysis reveals cellular alterations in psychosis and markers of
365 treatment-resistant schizophrenia. *Elife* **10**(2021).
- 366 14. Lehne, B. *et al.* A coherent approach for analysis of the Illumina HumanMethylation450 BeadChip
367 improves data quality and performance in epigenome-wide association studies. *Genome Biol* **16**, 37
368 (2015).
- 369 15. Liu, Y. *et al.* Epigenome-wide association data implicate DNA methylation as an intermediary of genetic
370 risk in rheumatoid arthritis. *Nat Biotechnol* **31**, 142-7 (2013).
- 371 16. Zhang, X. *et al.* Machine learning selected smoking-associated DNA methylation signatures that predict
372 HIV prognosis and mortality. *Clin Epigenetics* **10**, 155 (2018).
- 373 17. Zannas, A.S. *et al.* Epigenetic upregulation of FKBP5 by aging and stress contributes to NF-kappaB-driven
374 inflammation and cardiovascular risk. *Proc Natl Acad Sci U S A* **116**, 11370-11379 (2019).
- 375 18. Hannon, E. *et al.* An integrated genetic-epigenetic analysis of schizophrenia: evidence for co-localization
376 of genetic associations and differential DNA methylation. *Genome Biol* **17**, 176 (2016).
- 377 19. Hannum, G. *et al.* Genome-wide methylation profiles reveal quantitative views of human aging rates.
378 *Mol Cell* **49**, 359-67 (2013).
- 379 20. Johansson, A., Enroth, S. & Gyllensten, U. Continuous Aging of the Human DNA Methylome Throughout

380 the Human Lifespan. *PLoS One* **8**, e67378 (2013).

381 21. Kular, L. *et al.* DNA methylation as a mediator of HLA-DRB1*15:01 and a protective variant in multiple
382 sclerosis. *Nat Commun* **9**, 2397 (2018).

383 22. Tsaprouni, L.G. *et al.* Cigarette smoking reduces DNA methylation levels at multiple genomic loci but the
384 effect is partially reversible upon cessation. *Epigenetics* **9**, 1382-96 (2014).

385 23. Ventham, N.T. *et al.* Integrative epigenome-wide analysis demonstrates that DNA methylation may
386 mediate genetic risk in inflammatory bowel disease. *Nat Commun* **7**, 13507 (2016).

387