

Supplementary Information

microRNA-122 target sites in the Hepatitis C Virus RNA NS5B coding region and 3' untranslated region: function in replication and influence of RNA secondary structure

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Running head: miR-122/Ago2 binding in coding region and 3'UTR of HCV

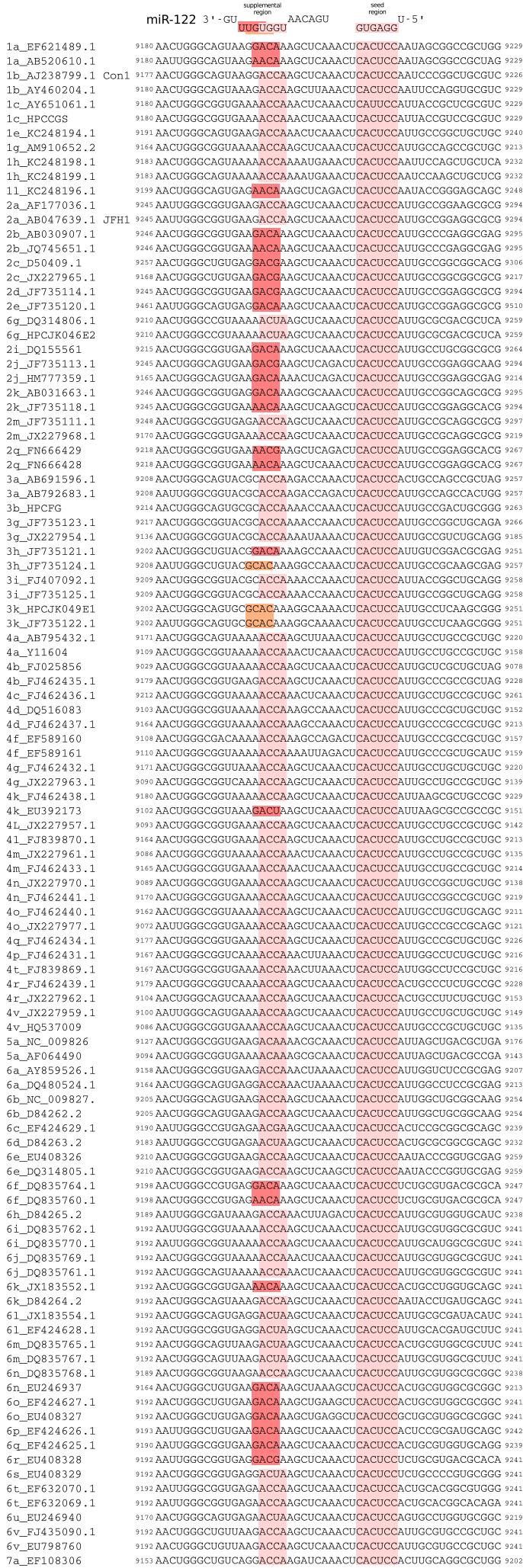
Keywords: microRNA, accessibility, Ago2, translation

Supplementary Figure S1 (next page), related to Figure 1

miR-122 and the conserved miR-122 target site 5B.2 in 106 selected HCV NS5B sequences (Fricke et al. 2015). The miR-122 sequence with the seed sequence (nts 2 – 8) and the supplementary binding region (nts 14 – 20) are marked in red. Gaps were introduced to align seed target sequences. HCV genotype and NCBI nucleotide database accession numbers are given on the left, nucleotide numbers are shown for each isolate. The miR-122 seed target site (A)CACUCC is 100 % conserved in 99 of 106 isolates, the supplementary binding site is conserved in all isolates. Below the sequences, the histogram shows the conservation of each residue.

Supplementary Figure S2 (next page), related to Figure 1

miR-122 and the conserved miR-122 target site 5B.3 in 106 selected HCV NS5B sequences (Fricke et al. 2015). The miR-122 seed sequence (nts 2 – 7) is marked in light red. Nucleotides of the supplementary binding region (involving nucleotides 14 – 20) are marked in light red, dark red or orange, respectively, as the corresponding HCV target sequences. HCV genotype and NCBI nucleotide database accession numbers are given on the left, nucleotide numbers are shown for each isolate. Both the miR-122 seed target site and the supplementary binding site are conserved in all isolates. Below the sequences, the histogram shows the conservation of each residue.



Supplementary Figure S3 (next page), related to Figure 1 and Supplementary Figure S1

Sequences with miR-122 site 5B.2. Nucleotide sequences were aligned according to amino acid sequences. Amino acids have been colored individually. Nucleotides are individually colored if more than 50 % conserved. Please note that the miR-122 seed target sequence is shifted by 2 nucleotides in some isolates (compare Supplementary Figure S1). The histogram shows the fraction of nucleotides actually used at each position relative to the number of nucleotides allowed by codon usage.

1. *1a_N_g*(0149384897)pb(ERf21489.1)
2. *1a_N_g*(025728021609)pb(A8520610.1)
3. *1b_N_g*(015420376)emb(jb)AU38999.1)
4. *1b_N_g*(0154239204)pb(j46020420.1)
5. *1c_M_g*(010233321)pb(jA632061.1)
6. *1c_M_g*(0144377)pb(jD4851.1)HCCGS
7. *1e_M_g*(015546080)pb(jC241934.1)
8. *1g_N_g*(019174856)emb(jm)AM91052.1)
9. *1h_M_g*(015546080)pb(jC241934.1)
10. *1h_M_g*(015546080)pb(jC241939.1)
11. *1h_M_g*(015546080)pb(jC241936.1)
12. *1h_N_g*(01051796)pb(jC172036.1)
13. *1j_N_g*(01322281)pb(jA80474629.1)
14. *1k_N_g*(019375141)pb(jB0300907.1)
15. *1k_M_g*(015546080)pb(jC241931.1)
16. *2v_N_g*(01485141)pb(jD5409.1)
17. *2v_N_g*(01485141)pb(jC241936.1)
18. *2d_M_g*(017977202)pb(jU735134.1)
19. *2d_M_g*(017977202)pb(jU735122.1)
20. *4g_N_g*(01369532)pb(D014808.1)
21. *4g_N_g*(01118303)pb(jD93221.1)HPC0462
22. *2z_N_g*(017975378)pb(jC153561)
23. *4z_N_g*(017975202)pb(jU735133.1)
24. *2d_M_g*(013557072)pb(jHMH773559.1)
25. *2z_N_g*(015210008)pb(jB031663.1)
26. *2w_M_g*(017977202)pb(jU735128.1)
27. *2z_M_g*(013977202)pb(jP35111.1)
28. *2z_M_g*(01395142739)pb(jX27968.1)
29. *2z_N_g*(0137447628)pb(jN664429)
30. *2z_N_g*(013744444)pb(jN664428)
31. *3z_N_g*(013220373)pb(jA691596.1)
32. *3z_N_g*(0128533052)pb(jA92683.1)
33. *3z_N_g*(017687716)pb(jD49371.1)PCG
34. *3g_M_g*(017977202)pb(jU735123.1)
35. *3g_N_g*(019514725)pb(jK27954.1)
36. *3g_M_g*(017977202)pb(jU735121.1)
37. *3z_M_g*(017977202)pb(jP735124.1)
38. *3h_M_g*(0179751082)pb(jA7092.1)
39. *3i_M_g*(017975202)pb(jU735125.1)
40. *4o_N_g*(01183032)pb(jD93211.1)HPC04951
41. *3z_M_g*(017977202)pb(jU735122.1)
42. *4x_N_g*(0139351671)pb(jA8795432.1)
43. *4x_N_g*(012324895)pb(jY13604)
44. *4y_N_g*(013994467)pb(jP035856)
45. *4y_M_g*(0139836684)pb(jUf42435.1)
46. *4y_M_g*(0139836684)pb(jP42439.1)
47. *4d_N_g*(019029367)pb(jD016003)
48. *4d_N_g*(0139836684)pb(jP42437.1)
49. *4t_N_g*(015446973)pb(jP259190.1)
50. *4t_N_g*(015449740)pb(jP509161)
51. *4y_N_g*(0139836678)pb(jP42432.1)
52. *4g_N_g*(01395142738)pb(jK27963.1)
53. *4c_M_g*(0139836684)pb(jP42439.1)
54. *4h_N_g*(012221561)pb(jE092173)
55. *4l_N_g*(0195147258)pb(jK227957.1)
56. *5h_M_g*(012456339)pb(jF838970.1)
57. *4m_N_g*(01951472458)pb(jK227961.1)
58. *4m_N_g*(0139836640)pb(jP42433.1)
59. *4m_N_g*(01395142738)pb(jK27970.1)
60. *4z_M_g*(0139836678)pb(jP42441.1)
61. *4o_N_g*(013983669)pb(jP42440.1)
62. *4o_N_g*(01395142739)pb(jK27977.1)
63. *4q_N_g*(0139836682)pb(jP42434.1)
64. *4q_N_g*(0139836678)pb(jP42431.1)
65. *4r_M_g*(013983669)pb(jP42439.1)
66. *4r_N_g*(013983669)pb(jP42439.1)
67. *4r_N_g*(01395142726)pb(jK227962.1)
68. *4v_N_g*(01395142726)pb(jK27959.1)
69. *4v_N_g*(013609119)pb(jQ373091)
70. *5a_S_g*(015739322)pb(jNC_009926)
71. *5a_N_g*(01006723)pb(jD0444490)
72. *5c_N_g*(017979939)pb(jA87952.1)
73. *5d_N_g*(013102515)pb(jD0409554.1)
74. *49_M_g*(0157732124)pb(jNC_009927.1)
75. *49_M_g*(010154546)pb(jD08262.1)
76. *4v_N_g*(0145487741)pb(jP424429.1)
77. *4d_N_g*(0111455)pb(jD08243.1)
78. *4d_N_g*(013844113)pb(jE092826)
79. *4w_N_g*(013699330)pb(jD0314805.1)
80. *4w_N_g*(013720309)pb(jD03035764.1)
81. *4t_M_g*(013720308)pb(jD03035760.1)
82. *4t_M_g*(011145478)pb(jD08245.2)
83. *4t_M_g*(013720309)pb(jD03035742.1)
84. *4t_M_g*(013720405)pb(jD03035770.1)
85. *4t_M_g*(013720404)pb(jD03035769.1)
86. *4t_M_g*(013720309)pb(jD03035761.1)
87. *4x_M_g*(0177551074)pb(jK183552.1)
88. *4x_N_g*(011145465)pb(jD08246.2)
89. *4x_N_g*(013720309)pb(jD03035741.1)
90. *4t_N_g*(014587739)pb(jF424628.1)
91. *4t_M_g*(013720309)pb(jD03035765.1)
92. *4t_M_g*(01127203599)pb(jD03035767.1)
93. *4t_M_g*(013720404)pb(jD03035768.1)
94. *4x_N_g*(013898748)pb(jU044937)
95. *4x_N_g*(014587737)pb(jF424627.1)
96. *4x_N_g*(013844113)pb(jE08327)
97. *4g_N_g*(014587735)pb(jF424626.1)
98. *4x_N_g*(013887733)pb(jF424623.1)
99. *4x_N_g*(013798864)pb(jD0409328)
100. *4t_N_g*(012443118)pb(jU0408329)
101. *4t_N_g*(013956001)pb(jD032070.1)
102. *4t_N_g*(013694601)pb(jF424626.1)
103. *4v_N_g*(015189874)pb(jD0246940)
104. *4v_N_g*(0124552249)pb(jP735090.1)
105. *4v_N_g*(020989728)pb(jU798760)
106. *4a_N_g*(012430204)pb(jF108306)

Supplementary Figure S4 (next page), related to Figure 1 and Supplementary Figure S2

Sequences with miR-122 site 5B.3. Nucleotide sequences were aligned according to amino acid sequences. Nucleotides are colored individually when more than 50 % conserved. Amino acids are colored individually. The histogram shows the fraction of nucleotides actually used at each position relative to the number of nucleotides allowed by codon usage.

1. 1a_N_g|0145384897|pb|EF621489|1
 2. 1a_N_g|035286216|pb|AB922061|1
 3. 1b_N_g|05420376|pb|D123799|1
 4. 1b_N_g|018492204|pb|AB40204|1
 5. 1c_M_g|050353521|pb|AB651061|1
 6. 1e_N_g|04417780|pb|D14531|HPCCGS
 7. 1e_M_g|0525460882|pb|C241919|1
 8. 1g_N_g|018374854|pb|AB91052|2
 9. 1h_M_g|052554696|pb|C241919|1
 10. 1h_M_g|018255469|pb|KC241919|1
 11. 1l_M_g|0525546088|pb|KC241916|1
 12. 2a_N_g|01020579|pb|AP177036|1
 13. 2a_N_g|03322261|pb|AB947639|1
 14. 2b_N_g|01937541|pb|AB930907|1
 15. 2b_M_g|01886475|pb|U0745651|1
 16. 2t_N_g|01483141|pb|D010409|1
 17. 2z_N_g|019514727|pb|U0227965|1
 18. 2d_M_g|019772021|pb|U735151|1
 19. 2e_M_g|019772020|pb|U735120|1
 20. 6g_N_g|01569952|pb|U00314800|1
 21. 6g_N_g|01183030|pb|D63822|HPCX0462
 22. 2h_N_g|01785578|pb|D015561|1
 23. 2l_M_g|019772019|pb|U735111|1
 24. 2l_M_g|015570727|pb|HM77359|1
 25. 2n_N_g|01652108|pb|U0031463|1
 26. 24_M_g|019772021|pb|U735118|1
 27. 2m_M_g|019772020|pb|U735111|1
 28. 2m_N_g|019551279|pb|K237984|1
 29. 2u_h_g|013744762|pb|R666429|
 30. 2e_h_g|0137448466|pb|R666428|
 31. 3a_N_g|0143220537|pb|U048915356|1
 32. 3a_N_g|018530505|pb|AB792683|1
 33. 3b_N_g|01678770|pb|U00314801|1
 34. 3g_M_g|0197972023|pb|U735123|1
 35. 3g_N_g|019514725|pb|U0227954|1
 36. 3h_M_g|0197972023|pb|U735121|1
 37. 3h_M_g|019772023|pb|U735124|1
 38. 3l_H_g|0237510832|pb|U074092|1
 39. 3l_M_g|0197972036|pb|U735125|1
 40. 3e_N_g|01180321|pb|U01821|HPCX0401
 41. 3w_N_g|019772020|pb|U735121|1
 42. 4u_N_g|019535873|pb|AB795432|1
 43. 4h_h_g|0125249950|pb|U011604|1
 44. 4h_h_g|015994675|pb|U025805|1
 45. 4h_h_g|019893449|pb|U042439|1
 46. 4c_h_g|019893680|pb|U042438|1
 47. 4c_h_g|09029367|pb|D0516083|1
 48. 4h_h_g|019893448|pb|U042437|1
 49. 4f_h_g|0156469738|pb|F589160|1
 50. 4f_h_g|0156469740|pb|F589161|1
 51. 4h_h_g|019893667|pb|U042432|1
 52. 4g_h_g|019247270|pb|U227983|1
 53. 4h_h_g|01929893699|pb|U042433|1
 54. 4h_h_g|0193514728|pb|U227979|1
 55. 4l_N_g|012321265|pb|EU03213|1
 56. 4l_N_g|015542735|pb|U227973|1
 57. 4m_N_g|019514726|pb|U227981|1
 58. 4m_N_g|019831660|pb|U042433|1
 59. 4n_N_g|019514728|pb|U227979|1
 60. 4n_h_g|019389669|pb|U042441|1
 61. 4o_M_g|229983694|pb|U462240|1
 62. 4o_N_g|015514729|pb|U227977|1
 63. 4q_h_g|229886642|pb|U042434|1
 64. 4p_h_g|22988667|pb|U042431|1
 65. 4t_M_g|015445397|pb|U83989|1
 66. 4r_h_g|019893656|pb|U042439|1
 67. 4r_N_g|015514726|pb|U227982|1
 68. 4v_N_g|015514726|pb|U227959|1
 69. 4v_h_g|013609119|pb|H037509|1
 70. 5a_h_g|0157781210|pb|NC_009928|1
 71. 5e_h_g|0160725|pb|AB084490|1
 72. 6a_N_g|01791993|pb|U085932|1
 73. 6a_N_g|019102151|pb|U049054|1
 74. 60_H_g|0157781214|HPCX009827|1
 75. 65_N_g|015715454|pb|U08402|1
 76. 6c_N_g|014388774|pb|U0842429|1
 77. 6d_N_g|016115455|pb|U084453|1
 78. 6e_h_g|015844111|pb|EU048832|1
 79. 6e_N_g|01699530|pb|D0314805|1
 80. 6f_N_g|0113220393|pb|U0805764|1
 81. 6f_N_g|0113220385|pb|U0805760|1
 82. 6f_N_g|016115457|pb|U084453|1
 83. 6f_N_g|0113220389|pb|U0805761|1
 84. 6f_N_g|0113720405|pb|U0835770|1
 85. 6f_N_g|0113220403|pb|U0835768|1
 86. 6f_N_g|0113720387|pb|U0835761|1
 87. 6x_N_g|016755107|pb|U083555|1
 88. 6x_N_g|016115456|pb|U084642|1
 89. 6c_M_g|01973107|pb|U0244937|1
 90. 6c_M_g|014568773|pb|U083554|1
 91. 6c_M_g|013772059|pb|D0035765|1
 92. 6c_M_g|013772059|pb|D0035767|1
 93. 6c_N_g|0113220407|pb|D0035768|1
 94. 6c_N_g|0160725|pb|EU0244937|1
 95. 6c_N_g|015694773|pb|U083554|1
 96. 6c_N_g|015844111|pb|EU048832|1
 97. 6p_N_g|015687733|pb|U042426|1
 98. 6o_N_g|015687733|pb|U0424265|1
 99. 6v_h_g|018748694|pb|U0409328|1
 100. 6v_h_g|018441110|pb|U0408329|1
 101. 6v_h_g|01569010|pb|U0408320|1
 102. 6t_M_g|0156960158|pb|U0408320|1
 103. 6v_h_g|01248987|pb|U0244940|1
 104. 6v_h_g|012455239|pb|U015590|1
 105. 6v_h_g|012088722|pb|U0197870|1
 106. 7a_N_g|012432020|pb|U010806|1

Supplementary Figure S5 (next page), related to Figure 1

miR-122 and the conserved miR-122 target site S3 in 93 selected HCV NS5B sequences (Fricke et al. 2015). The miR-122 sequence with the seed sequence (nts 2 – 8) and the supplementary binding region (nts 17 – 20) are marked in red. Gaps were introduced to align seed target sequences. HCV genotype and NCBI nucleotide database accession numbers are given on the left, nucleotide numbers are shown for each isolate. The miR-122 seed target site (A)CACUCC is 100 % conserved in 85 of 94 isolates, the supplementary binding site is conserved in 92/94 isolates. Below the sequences, the histogram shows the conservation of each residue.



miR-122 3' - GU UUGUACAG U-5'

			supplemental region	seed region
1a_EF621489.1		ARG stop		
1a_AB520610.1				
1b_AJ238799.1	Con1			
1b_AY460204.1				
1c_AY651061.1				
1c_HPCCGS				
1e_KC248194.1				
1g_AM910652.2				
1h_KC248198.1				
1h_KC248199.1				
11_KC248196.1				
2a_AF177036.1				
2a_AB047639.1	JFH1			
2b_AB030907.1				
2b_JQ745651.1				
2c_D50409.1				
2c_JX227965.1				
2d_JF735114.1				
2e_JF735120.1				
6g_DQ314806.1				
6g_HPCJK046E2				
2j_JF735113.1				
2k_AB031663.1				
2k_JF735118.1				
2m_JF735111.1				
2m_JX227968.1				
2q_FN666428				
3a_AB691596.1				
3a_AB792683.1				
3b_HPCFG				
3g_JF735123.1				
3g_JX227954.1				
3h_JF735121.1				
3h_JF735124.1				
3i_FJ407092.1				
3i_JF735125.1				
3k_HPCJK049E1				
3k_JF735122.1				
4a_AB795432.1				
4a_Y11604				
4b_FJ025856				
4b_FJ462435.1				
4c_FJ462436.1				
4d_FJ462437.1				
4g_FJ462432.1				
4g_JX227963.1				
4k_FJ462438.1				
4k_EU392173				
4L_JX227957.1				
41_FJ839870.1				
4m_JX227961.1				
4m_FJ462433.1				
4n_JX227970.1				
4n_FJ462441.1				
4o_FJ462440.1				
4o_JX227977.1				
4q_FJ462434.1				
4p_FJ462431.1				
4t_FJ839869.1				
4r_FJ462439.1				
4r_JX227962.1				
4v_JX227959.1				
5a_NC_009826				
5a_AF064490				
6b_NC_009827.				
6b_D84262.2				
6c_EF424629.1				
6d_D84263.2				
6e_EU408326				
6e_DQ314805.1				
6f_DQ835764.1				
6f_DQ835760.1				
6h_D84265.2				
6i_DQ835762.1				
6i_DQ835770.1				
6j_DQ835769.1				
6j_DQ835761.1				
6k_JX183552.1				
6k_D84264.2				
6l_JX183554.1				
6l_EF424628.1				
6m_DQ835765.1				
6m_DQ835767.1				
6n_DQ835768.1				
6o_EF424627.1				
6o_EU408327				
6p_EF424626.1				
6q_EF424625.1				
6r_EU408328				
6s_EU408329				
6t_EF632070.1				
6t_EF632069.1				
6v_FJ435090.1				
6v_EU798760				

.....10.....20.....30.....40.....50.....



Supplementary Figure S6

RNAs used in binding experiments

Structures of RNAs used in binding experiments were predicted using the Vienna **RNAfold** WebServer (<http://rna.tbi.univie.ac.at/cgi-bin/RNAfold.cgi>) (Gruber et al., 2008; Lorenz et al., 2011).

Options were set as follows: Basic options: "avoid isolated base pairs". Advanced folding options were: "no dangling end energies" with the Turner model, 2004.

The Vienna RNA structure prediction output is shown in dot-bracket form. Seed binding sites and supplemental binding sites for microRNA-122 are labelled in yellow.

The origin of the templates for PCR and those nucleotides of the PCR primers that are contained in the in vitro-transcribed RNA are listed below each sequence. Here we show only those sequences of the primers which became part of the RNA; upstream of the forward primer there was always a T7 promoter included in the primer. With most forward primers, three G residues were added to the sequence to improve T7 RNA polymerase-mediated in vitro transcription of the RNAs.

RNAs for Site 5B.2 - Secondary structure predictions

RNA_5B.2a_Con1_227_nts

GGGGUGGGAGACAGCUAGACACACUCCAGUCAAUUCCUGGCUAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAGG
AUGAUCCUGAUGACUCAUUUCCUCCAUCCUUCUAGCUCAGGAACAACUUGAAAAGCCUAGAUUGUCAGAUCUACGGGG
CCUGUUACUCCAUUGAGCCACUUGACCUACCUCAGAUCAUCAACGACUCCAUGGCCUUAGCG
((((((.....)))))).((((((((((((((....))))))).))).((((((....)))))))((((((....))))).(((((....((....))))..)))))).((((((((((....))))))).....(((((....))))..)))).(((((....((....))))..((....))))....))).(((....))))

Template: HCV genotype 1b Con1 (AJ238799.1)

Primer fwd: 5'-GGGTGGGAGACAGCTAGACACACTCCAG-3'

Primer rev: 5'-CGCTAAGGCCATGGAGTCGTTGAATGATCTGAGG-3'

RNA_5B.2a*_JFH1_267_nts

GGGAACAGUUAGACACUCCCUAUCAAUUCAUGGCUGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGUUCGCAUGGU
CCUAAUGACACACUUCCUUCUCCAUUCUCAUGGUCCAAGACACCCUGGACCAGAACCUACUUUGAGAGUAUGGAUCAGUA
UACUCCGUGAAUCCUUUUGACCUUCCAGCCAUAUUGAGAGGUUACACGGCUUGACGCCUUUCUAUGCACACAUCUC
ACCACGAACUGACGCGGGUGG
((((((.....((....)))))))....((((((((((....))))))).))).((((((....))))))).((((((....))))).((....((....))))..))).((((((((((....))))))).....))).....((....))))....((....((....)))))))

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGGAAACAGTTAGACACTCCCC-3'

Primer rev: 5'-CCACCCGCGTCAGTTGAGAGTATGTGTGC-3'

RNA_5B.2b_Con1_148_nts

GGGUGCGUGGGAGACAGCUAGACACACUCCAGUCAAUUCCUGGCUAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCA
AGGAUGAUCCUGAUGACUCAUUUCCUCCAUCCUUCUAGCUCAGGAACAACUUGAAAAGCCUA
((((((.....((....))))..)))....((((((((((....))))))).))).((((((....))))))).
((((((....((....))))..))).))).((((((((((....))))))).....))).....((....))))....((....((....)))))))

Template: HCV genotype 1b Con1 (AJ238799.1)

Primer fwd: 5'-GGGTGCGTGGGAGACAGCTAGAC-3'

Primer rev: 5'-TAGGGCTTTCAAGTTGTTCTGAG-3'

RNA_5B.2b*_JFH1_291_nts

GGGAACAGUUAGACACUCCCUAUCAAUUCAUGGCUGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGUUCGCAUGGU
CCUAAUGACACACUUCCUUCUCCAUUCUCAUGGUCCAAGACACCCUGGACCAGAACCUACUUUGAGAGUAUGGAUCAGUA
UACUCCGUGAAUCCUUUUGACCUUCCAGCCAUAUUGAGAGGUUACACGGCUUGACGCCUUUCUAUGCACACAUCUC
ACCACGAACUGACGCGGGUGGUUCAGCCUCAGAAAACUUGGGG
.....((....((....((....))))..)))....((((((((((....))))))).))).((((((....))))))).
((((((....((....))))..))).))).((((((((((....))))))).....))).....((....))))....((....((....)))))))

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGGAAACAGTTAGACACTCCCC-3'

Primer rev: 5'-CCCCAAGTTCTGAGGGCTGAAGCCACCC-3'

RNA_5B.2c_Con1_152_nts

GGGCGUGGGAGACAGCUAGACACACUCCAGUCAAUUCCUGGCUAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAG
GAUGAUCCUGAUGACUCAUUUCCUCCAUCCUUCUAGCUCAGGAACAACUUGAAAAGCCUAGAUGU
.....((....((....((....))))..)))....((((((((((....))))..))).))).((((((....))))))).
((((((....((....))))..))).))).((((((((((....))))))).....))).....((....))))....((....((....)))))))

Template: HCV genotype 1b Con1 (AJ238799.1)

Primer fwd: 5'-GGCGTGGGAGACAGCTAGACACAC-3'

Primer rev: 5'-ACAATCTAGGCCTTTCAAGTTGCC-3'

RNA_5B.2d_JFH1_160_nts

GGGCCACUCGCCGGCUGCCUGGGAAACAGUUAGACACUCCCCUAUCAAUUCAUGGCUGGGAAACAUCAUCCAGUAUGCUC
CAACCAUAUGGUUCGCAUGGUCCUAAUGACACACACUUCUCCAUUCUCAUGGUCCAAGACACCCUGGACCAGAAC
(((((((((.....((((.....((((((.....))))....))))....))))....))))....))))....
.....))))....))))....))))....))))....))))....))))....))))....))))....))))....

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGGCCACTGCCGGGCTGCCTGG-3'

Primer rev: 5'-GGTTCTGGTCCAGGGTGTC-3'

RNAs for Site 5B.2 - Alignment

CLUSTAL format alignment by MAFFT (v7.304b)

RNA_5B.2a_Con1_ -----GGGGUGGGAGACAGCUAGACACACUCC--AGUAAUUCUGGC
RNA_5B.2a_JFH1_ -----GGGAAACAGUUAG--ACACUCCCUAUCAAUUCAUGGC
RNA_5B.2b_Con1_ -----GGGUGCGUGGGAGACAGCUAGACACACUCC--AGUAAUUCUGGC
RNA_5B.2b_JFH1_ -----GGGAAACAGUUAG--ACACUCCCUAUCAAUUCAUGGC
RNA_5B.2c_Con1_ -----GGGCGUGGGAGACAGCUAGACACACUCC--AGUAAUUCUGGC
RNA_5B.2d_JFH1_ GGGCACUCGCCGGCUGCCUGGGAACAGUUAG--ACACUCCC
***** . * *** * * ***** * ***** * ***** *

RNA_5B.2a_Con1_ UAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAGGAUGAUGC
RNA_5B.2a_JFH1_ UGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGUUCGCAUGGU
RNA_5B.2b_Con1_ UAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAGGAUGAUGC
RNA_5B.2b_JFH1_ UGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGUUCGCAUGGU
RNA_5B.2c_Con1_ UAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAGGAUGAUGC
RNA_5B.2d_JFH1_ UGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGUUCGCAUGGU
* . ** * ***** * * *** * * . * *** * * *** . * *** * *

RNA_5B.2a_Con1_ UCUCUCUCCAUCCUUCUAGCUCAGAACACUUGAAAAAGCC
RNA_5B.2a_JFH1_ UCUCUCUCCAUUCUCAUGGUCCAAGACACCC
RNA_5B.2b_Con1_ UCUCUCUCCAUUCUAGCUCAGAACACUUGAAAAAGCC
RNA_5B.2b_JFH1_ UCUCUCUCCAUUCUAGCUCAGAACACCC
RNA_5B.2c_Con1_ UCUCUCUCCAUUCUAGCUCAGAACACUUGAAAAAGCC
RNA_5B.2d_JFH1_ UCUCUCUCCAUUCUAGCUCAGAACACCC
***** . * * . * *** * * *** * . *

RNA_5B.2a_Con1_ GGGCCUGUUACUCCAUUGAGCCACUUGACCUACC
RNA_5B.2a_JFH1_ GAUCAGUAUACUCCGUGAAUCCUUU
RNA_5B.2b_Con1_ -----
RNA_5B.2b_JFH1_ GAUCAGUAUACUCCGUGAAUCCUUU
RNA_5B.2c_Con1_ -----
RNA_5B.2d_JFH1_ -----

RNA_5B.2a_Con1_ UUAGCG-----
RNA_5B.2a_JFH1_ UUGACGCCUUUCUAUGCACACAUACUC
RNA_5B.2b_Con1_ -----
RNA_5B.2b_JFH1_ UUGACGCCUUUCUAUGCACACAUACUC
RNA_5B.2c_Con1_ -----
RNA_5B.2d_JFH1_ -----

RNAs for Site 5B.3 - Secondary structure predictions

RNA_5B.3a_Con1_226_nts

GGGGCUACUGUCCCAGGGGGGAGGGCUGCCACUUGUGGAAGUACCUCUACUGGGCAGUAAGGACCAAGCUAAACU
CACUCCAUUCCCCGUGCGUCCAGUUGGAUUUAUCCAGCUGGUUCUGGUACAGCGGGGAGACAUUAUCAGC
CUGUCUCGUGCCGACCCCGUGGUCAUGUGGUGCCUACUCCUACUUUCUGUAGGGUAGG
.(((((.....((((((.....((((((.....))))....))))....))))....)))).....
((((.....(((((.....((((((.....))))....))))....))))....))))....(((((((.....
.....))))....)))).....))))).....))))(((((.....))))....))))

Template: HCV genotype 1b Con1 (AJ238799.1)

Primer fwd: 5'-GGGGCTACTGTCCCAGGG-3'

Primer rev: 5'-CCTACCCCTACAGAAAGTAGGAGTAGGCAC-3'

RNA_5B.3a*_JFH1_200_nts

GGGCGGUGAAGACCAAGCUAAACU
CACUCCAUUGCCGGAGGCGGCCUACUGGACUUAUCCAGUUGGUUCACCGUCGGCGC
CGGCGGGGGCGACAUUUUCACAGCGUGUCGCGCGCCGACCCCGCUCUUACUUCGGCUACUCC
UACUUUUCGUAGGGGUAGGCCUCUUCUACUCCCCGUCGGUAGAGCGGC
(((((((.....))))....))......((((.....((((.....))))....))))....)).
.(((((((.....))))....))))....))))....((((.....))))....))))....
))))....((((.....))))....)).

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGCGGTGAAGACCAAGCTCAAACACTCCATTG-3'

Primer rev: 5'-GCCGCTCTACCGAGCGGGGAGTAGG-3'

RNA_5B.3b_JFH1_196_nts

GGGCGGUGAAGACCAAGCUAAACU
CACUCCAUUGCCGGAGGCGGCCUACUGGACUUAUCCAGUUGGUUCACCGUCGGCGC
CGGCGGGGGCGACAUUUUCACAGCGUGUCGCGCGCCGACCCCGCUCUUACUUCGGCUACUCCUACUUUCGUAGGG
GUAGGCCUCUUCUACUCCCCGUCGGUAGAG
(((((((.....))))....))......((((.....((((.....))))....))))....)).))))((
(((((((.....))))....))))((.....))))....))))....((((.....))))....))))....
))))....))))....)).

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGCGGTGAAGACCAAGCTCAAACACTCCATTG-3'

Primer rev: 5'-CTCTACCGAGCGGGGAGTAGGAAGAGGCC-3'

RNA_5B.3c_JFH1_224_nts

GGGUCAUCUCCGUGGAGGGAAAGCGGCCGUUJCGGCCGAUACUCUAAUUGGGCGUGAAGACCAAGCUAAACU
CACC
UCCAUUGCCGGAGGCGGCCUACUGGACUUAUCCAGUUGGUUCACCGUCGGCGCCGGCGACAUUUUCACAGCGUG
UCGCGCGCCGACCCCGCUCUUACUUCGGCUACUCCUACUUUCGUAGGGUAGGC
((.((((.....((((.....))))....))))....))))....))......
.....((((.....((((.....))))....))))....))))....))......))))....
))))....))))....))......((((.....))))....))).

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGGTCATCTCCCGTGGAGG-3'

Primer rev: 5'-GCCTACCCCTACGAAAAGTAGGAGTAGGCCG-3'

RNAs for Site 5B.3 - Alignment

CLUSTAL format alignment by MAFFT (v7.304b)

RNA_5B.3a_Con1_ GGGGGCUACUGUCCCAGGGGGGAGGGCUGCACUUGUGGCAAGUACCUCUCAACUGGG
RNA_5B.3a_JFH1_ -----GGG
RNA_5B.3b_JFH1_ -----GGG
RNA_5B.3c_JFH1_ ---GGGUCAUCUCCCGUGGAGGGAAAGCGGCCGUUJCGGCCGAUAUCUCUCAAUUGGG

RNA_5B.3a_Con1_ CAGUAAGGACCAAGCUAAACUCACUCCAAUCCCGCUGCUGGUCCCCAGUUGGAUUUAUCCA
RNA_5B.3a_JFH1_ CGGUGAAGACCAAGCUAAACUCACUCCAUUGCCGGAGGCACGCCUACUGGACUUAUCCA
RNA_5B.3b_JFH1_ CGGUGAAGACCAAGCUAAACUCACUCCAUUGCCGGAGGCACGCCUACUGGACUUAUCCA
RNA_5B.3c_JFH1_ CGGUGAAGACCAAGCUAAACUCACUCCAUUGCCGGAGGCACGCCUACUGGACUUAUCCA
..*.***** * * *** * . *** * *****

RNA_5B.3a_Con1_ GCUGGUUCGUUGCUGGUUACAGCGGGGGAGACAUUAUCACAGCCUGUCUGUGCCCCGAC
RNA_5B.3a_JFH1_ GUUGGUUCACCGUCGGCGCCGGGGGGCGACAUUUUUCACAGCGUGUCGCGCGCCGAC
RNA_5B.3b_JFH1_ GUUGGUUCACCGUCGGCGCCGGGGGGCGACAUUUUUCACAGCGUGUCGCGCGCCGAC
RNA_5B.3c_JFH1_ GUUGGUUCACCGUCGGCGCCGGGGGGCGACAUUUUUCACAGCGUGUCGCGCGCCGAC
* *****. * ** * .***** * ***** * ***** * **** * *****

RNA_5B.3a_Con1_ CCCGCUGGUCAUGUGGUAGCCUACUCCUACUUUCUGUAGGGGUAGG-----
RNA_5B.3a_JFH1_ CCCGCUCAUUACUCUUUCGGCUACUCCUACUUUCGUAGGGGUAGGCCUCUCCUACUCC
RNA_5B.3b_JFH1_ CCCGCUCAUUACUCUUUCGGCUACUCCUACUUUCGUAGGGGUAGGCCUCUCCUACUCC
RNA_5B.3c_JFH1_ CCCGCUCAUUACUCUUUCGGCUACUCCUACUUUCGUAGGGGUAGGC-----
***** . * * * .***** * *****

RNA_5B.3a_Con1_ -----
RNA_5B.3a_JFH1_ CCGCUCGGUAGAGCGGC
RNA_5B.3b_JFH1_ CCGCUCGGUAGAG---
RNA_5B.3c_JFH1_ -----

RNAs for Site S3 - Secondary structure predictions

RNA_S3a_Con1_250_nts

GGGCC**GAUGAACGGGGAGCUAAACACUCCAGGCCAUAGGCCAUCCUGUUUUUUUUCCUUUUUUUUUUCUUUUUUUUUU
UU
GUCA CGGC UAGC UGUG AAAG GUCC GUG AGCC GUUG ACUG CAG AGA GUG CUG AUAC UGG CCUC UCG CAG AUA
GCUA
(((((...((.((((((.(((.....((((....))))....))))....))))....))))....
.....(((((.....))))....(((((.....))))....))))....))))....
....**

Template: HCV genotype 1b Con1 (AJ238799.1)

Primer fwd: 5'-GGCCGATGAAACGGGAGCTAACACTCCAGGC-3'

Primer rev: 5'-ACTAGTAGTACTGATCTGCAGAGAGGCC-3'

RNA_S3b_JFH1_252_nts

GGGUCCCCGUC**GGUAGAGCGGCACACACUAGGUACACUCCAUAGCUAACUGUCCUUUUUUUUUUUUUUUUUUUUUU
UU
AUCUUAGCCCUCAGUCACGGCUAGCUGUGAAAGGUCCGUGAGGCCGAUGACUGCAGAGAGUGCCUA
CUGGUCUCUGCAG
AUCAUG
.....((((((.....))))....((.((.....))))....)
.....((.....(((((.....))))...)))....))(((((.....(((((.....))))....))))....
....)).**

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-GGGTCCCCGCTCGGTAGAGCGGCACACACTAGGTACACTCCATAGCTAAC--3'

Primer rev: 5'-CATGATCTGCAGAGAGACCAGTTACGGCAC-3'

RNA_S3b*_Con1_153_nts

GGGCAGCGGGGAGACAUUAUCACAGCCUGUCGUGCCGACCCCGCUGGUCAUGUGGUCCUACUCCUACUUUCUGUA
GGGUAGGCAUCUAUCUACUCCCCAACCG**AUGAACGGGAGCUAAACACUCCAGGCCAUAGGCCAUCCUG
((.(((((((((.....))))....))))....))))....(((((.....(((((.....))))....
....))))....(((((.....))))....))....((.....(((((.....))))....))))....
....**

Template: HCV genotype 1b Con1 (AJ238799.1)

Primer fwd: 5'-GGCCAGCGGGGAGACATATATCACAGC-3'

Primer rev: 5'-CAGGATGGCCTATTGGCCTGGAG-3'

RNA_S3c_JFH1_149_nts

GGGUCCCCGUC**GGUAGAGCGGCACACACUAGGUACACUCCAUAGCUAACUGUCCUUUUUUUU
UUUCGUAGGGGUAGGCCUCUCCUACUCCCCGUC**GGUAGAGCGGCACACACUAGGUACACUCCAU
(((((.....((.....))))....))))....((.....(((((.....))))....))))....(((((.....((.....))))....))))....((.....(((((.....))))....))))....((.....(((((.....))))....))))....
....****

Template: HCV genotype 2a JFH-1 (AB047639.1)

Primer fwd: 5'-ggGTCGGCGCCGGCGGGCGA-3'

Primer rev: 5'-TATGGAGTGTACCTAGTGTGCGCTCTACCGAG-3'

RNAs for Site S3 - Alignment

CLUSTAL format alignment by MAFFT (v7.304b)

RNA_S3a_Con1_25 GGG-----
RNA_S3b_JFH1_25 GGG-----
RNA_S3b_ GGG-----CAGCGGGGAGACAUUAUCACAGCCUGUCGUGCCGACCCGCUGGU
RNA_S3c_JFH1_14 GGGUCGGCGCCGGCGGGGCACAUUUUJACAGCGUGUCGCGCCGACCCGCUCAU

RNA_S3a_Con1_25 -----CCGAU
RNA_S3b_JFH1_25 -----UCCCCGCUCGGU
RNA_S3b_ UCAUGUGGUGCUCUACUCCUACUUUCUGUAGGGUAGGCCAUCUAUCUACUCCCAACC
RNA_S3c_JFH1_14 UACUCUUCGGCCUACUCCUACUUUCGUAGGGUAGGCCUCUCCUACUCCCGCUCGGU
***.

RNA_S3a_Con1_25 GAACGGGAGCUAA-----ACACUCCAGGCCAAUAGGCCAUCCUGUUUUUUUCCUUUUU
RNA_S3b_JFH1_25 AGAGCGGCACACACUAGGUACACUCCAUAGCUACUGUUCCUUUUUUUUUUUUUUUU
RNA_S3b_ GAACGGGAGCUAA-----ACACUCCAGGCCAAUAGGCCAUCCUG-----
RNA_S3c_JFH1_14 AGAGCGGCACACACUAGGUACACUCCAU-----
. . * * * * * * * * .

RNA_S3a_Con1_25 UUU
RNA_S3b_JFH1_25 UUU
RNA_S3b_ -----
RNA_S3c_JFH1_14 -----

RNA_S3a_Con1_25 UUUUUUCCUU
RNA_S3b_JFH1_25 CUUAUUCUACUUUUUUUUUUUUUUUUUUUUUUUUUUUUUUUUUU
RNA_S3b_ -----
RNA_S3c_JFH1_14 -----

RNA_S3a_Con1_25 GGUCGUGAGCCGCUUGACUGCAGAGAGUGCUGAUACUGGCCUCUGCAGAUCAAGU
RNA_S3b_JFH1_25 GGUCGUGAGCCGCAUGACUGCAGAGAGUGCCUAACUGGCCUCUGCAGAUCAUG-
RNA_S3b_ -----
RNA_S3c_JFH1_14 -----

FASTA compatible format

>RNA_5B.2a_Con1_227_nts

GGGGUGGGAGACAGCUAGACACACUCCAGUCAAUUCCUGGCUAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAGG
AUGAUCCUGAUGACUCAUUUUCUCCAUCCUUCUAGCUCAGGAACAACUUGAAAAAGCCUAGAUUGUCAGAUCUACGGGG
CCGUUUACUCCAUUGAGCCACUUGACCUACCUCAGAUCAUCAACGACUCCAUGGCCUUAGCG

>RNA_5B.2a*_JFH1_267_nts

GGGAAACAGUUAGACACACUCCCCUAUCAAUCAUGGCUGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGGUUCGCAUGGU
CCUAAUGACACACACUUCUUCUCCAUUCUCAUGGUCCAAGACACCCUGGACCAGAACCUACUUGAGAUGUAUGGAUCAGUA
UACUCCGUGAAUCCUUUUGGACCUUCCAGCCAUAUUGAGAGGUUACACGGGCUUGACGCCUUUCUAUGCACACAUACUC
ACCACGAACUGACGCGGGUGG

>RNA_5B.2b_Con1_148_nts

GGGUGCGUGGGAGACAGCUAGACACACUCCAGUCAAUUCCUGGCUAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCA
AGGAUGAUCUGAUGACUCAUUUUCUCCAUCCUUCUAGCUCAGGAACAACUUGAAAAAGCCUA

>RNA_5B.2b*_JFH1_291_nts

GGGAAACAGUUAGACACACUCCCCUAUCAAUCAUGGCUGGGAAACAUCAUCCAGUAUGCUCCAACCAUAUGGGUUCGCAUGGU
CCUAAUGACACACACUUCUUCUCCAUUCUCAUGGUCCAAGACACCCUGGACCAGAACCUACUUGAGAUGUAUGGAUCAGUA
UACUCCGUGAAUCCUUUUGGACCUUCCAGCCAUAUUGAGAGGUUACACGGGCUUGACGCCUUUCUAUGCACACAUACUC
ACCACGAACUGACGCGGGUGGCUUCAGCCUCAGAAAACUUGGGG

>RNA_5B.2c_Con1_152_nts

GGGCGUGGGAGACAGCUAGACACACUCCAGUCAAUUCCUGGCUAGGCAACAUCAUCAUGUAUGCGCCCACCUUGUGGGCAAG
GAUGAUCUGAUGACUCAUUUUCUCCAUCCUUCUAGCUCAGGAACAACUUGAAAAAGCCUAGAUUGU

>RNA_5B.2d_JFH1_160_nts

GGGCCACUCGCCGGCUGCCUGGGAAACAGUUAGACACACUCCCCUAUCAAUCAUGGCUGGGAAACAUCAUCCAGUAUGCUC
CAACCAUAUGGUUCGCAUGGUCCUAUAGACACACUUCUUCUCCAUUCUCAUGGUCCAAGACACCCUGGACCAGAAC

>RNA_5B.3a_Con1_226_nts

GGGGGCUACUGUCCCAGGGGGGGAGGGCUGCCACUUGUGGCAAGUACCUUCAACUGGGCAGUAAGGACCAAGCUAAACU
CACUCCAAUCCGGCUGCGUCCAGUUGGAAUUAUCCAGCUGGUUCGUUGCUGGUACAGCGGGGGAGACAUUAUCACAGC
CUGUCUCGUGCCCGACCCCGCUGGUUCAUGUGGUGCCUACUCCUACUUUCUGUAGGGUAGG

>RNA_5B.3a*_JFH1_200_nts

GGGCGGUGAAGACCAAGCUAAACUCACUCCAUUGCCGGAGGCGGCCUACUGGACUUAUCCAGUUGGUUCACCGUCGGCGC
CGGCGGGGGCGACAUUUUCACAGCGUGUCGCGCCGACCC

>RNA_5B.3b_JFH1_196_nts

GGGCGGUGAAGACCAAGCUAAACUCACUCCAUUGCCGGAGGCGGCCUACUGGACUUAUCCAGUUGGUUCACCGUCGGCGC
CGGCGGGGGCGACAUUUUCACAGCGUGUCGCGCCGACCC

>RNA_5B.3c_JFH1_224_nts

GGGUCAUCUCCGUGGAGGGAAAGCGGCCGUUUGCGGCCGAUAUCUCAAUUGGGCGGUGAAGACCAAGCUAAACUCAC
UCCAUUGCCGGAGGCGGCCUACUGGACUUAUCCAGUUGGUUCACCGUCGGCGCCGGCGACAUUUUCACAGCGUG
UCGCGCGCCGACCCGCUCAUUACUCUUCGCCUACUCCUACUUUCGUAGGGUAGGC

>RNA_S3a_Con1_250_nts

GGGCCGAUGAACGGGGAGCUAAACACUCCAGGCCAUAGGCCAUCCUGUUUUUUUUCCUUUUUUUUUUUUUUUUUU
UU
GUACCGCUAGCUGUGAAAGGUCCGUGAGCGCUUGACUGCAGAGAGUGCUGAUACUGGCCUCUCUGCAGAUCAAGUAC
UAGU

>RNA_S3b_JFH1_252_nts

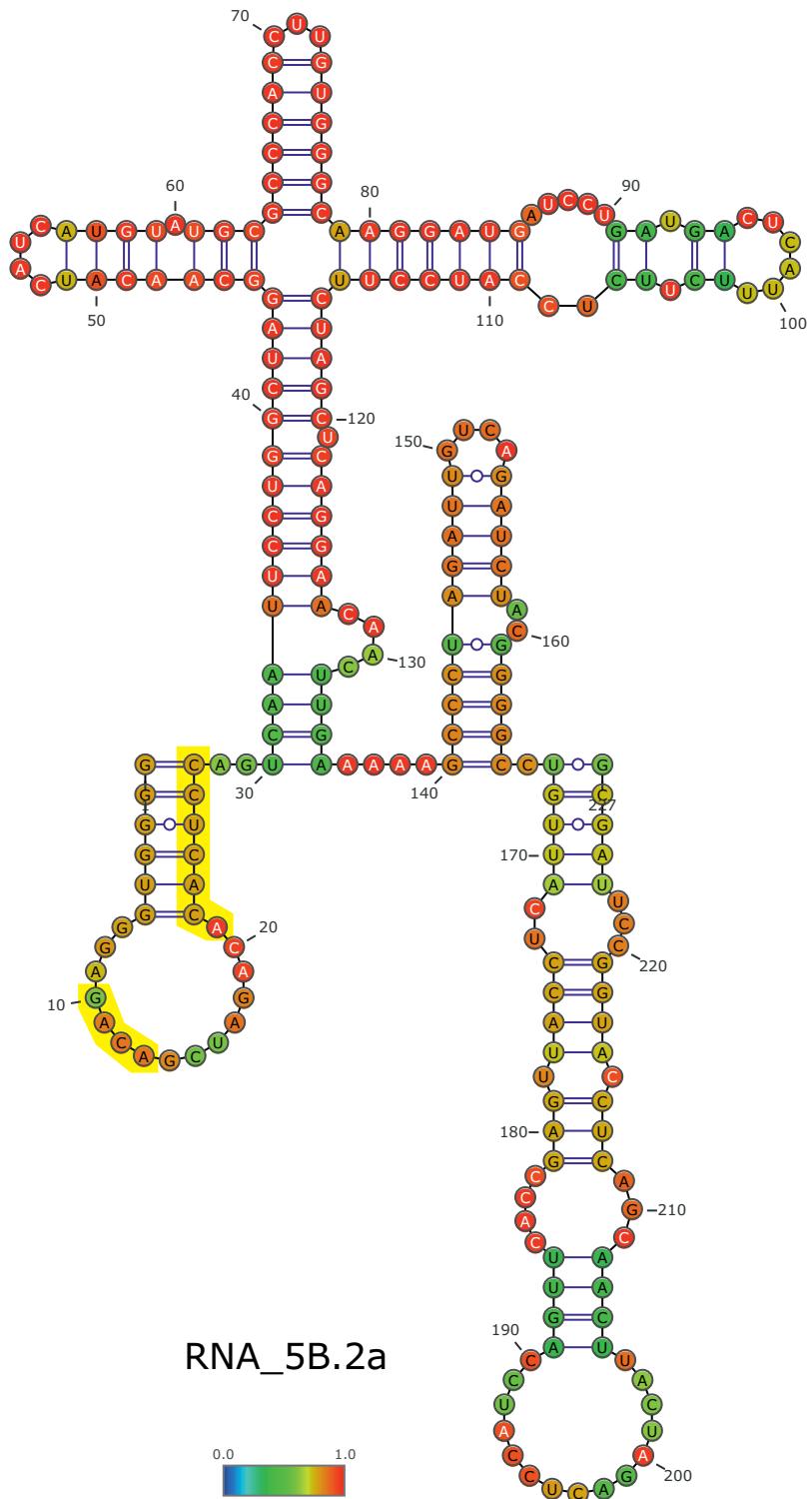
GGGUCCCCGCUCCGUAGAGCGGCACACACUAGGUACACCUUAGCUACUGUUCUUUUUUUUUUUUUUUUUUUUUU
UU
AUCUUAGCCCUAGUCACGGCUAGCUGUGAAAGGUCCGUGAGCGCAUGCAGAGAGUGCCUAACUGGUUCUCUGCAG
AUCAUG

> RNA_S3b*_Con1_153_nts

GGGCAGCGGGGGAGACAUUAUCAGCCUGUCGUGCCGACCCCGCUGGUCAUGUGGUGCCUACUCCUACUUUCUGUA
GGGUAGGCAUCUAUCUACUCCCCAACGAUGAACGGGAGCUAAACACUCCAGGCCAUAGGCCAUCCUG

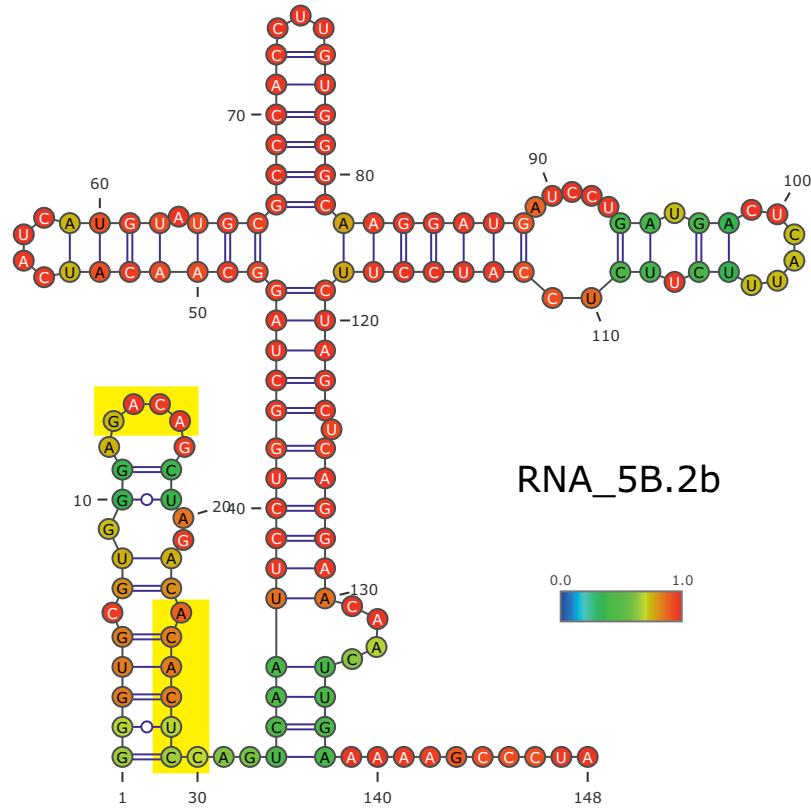
>RNA_S3c_JFH1_149_nts

GGGUAGGGUAGGCCUCUCCUACUCCCCGUCGGUAGAGCGGCACACACUAGGUACACUCCAUA
UUUCGUAGGGUAGGCCUCUCCUACUCCCCGUCGGUAGAGCGGCACACACUAGGUACACUCCAUA



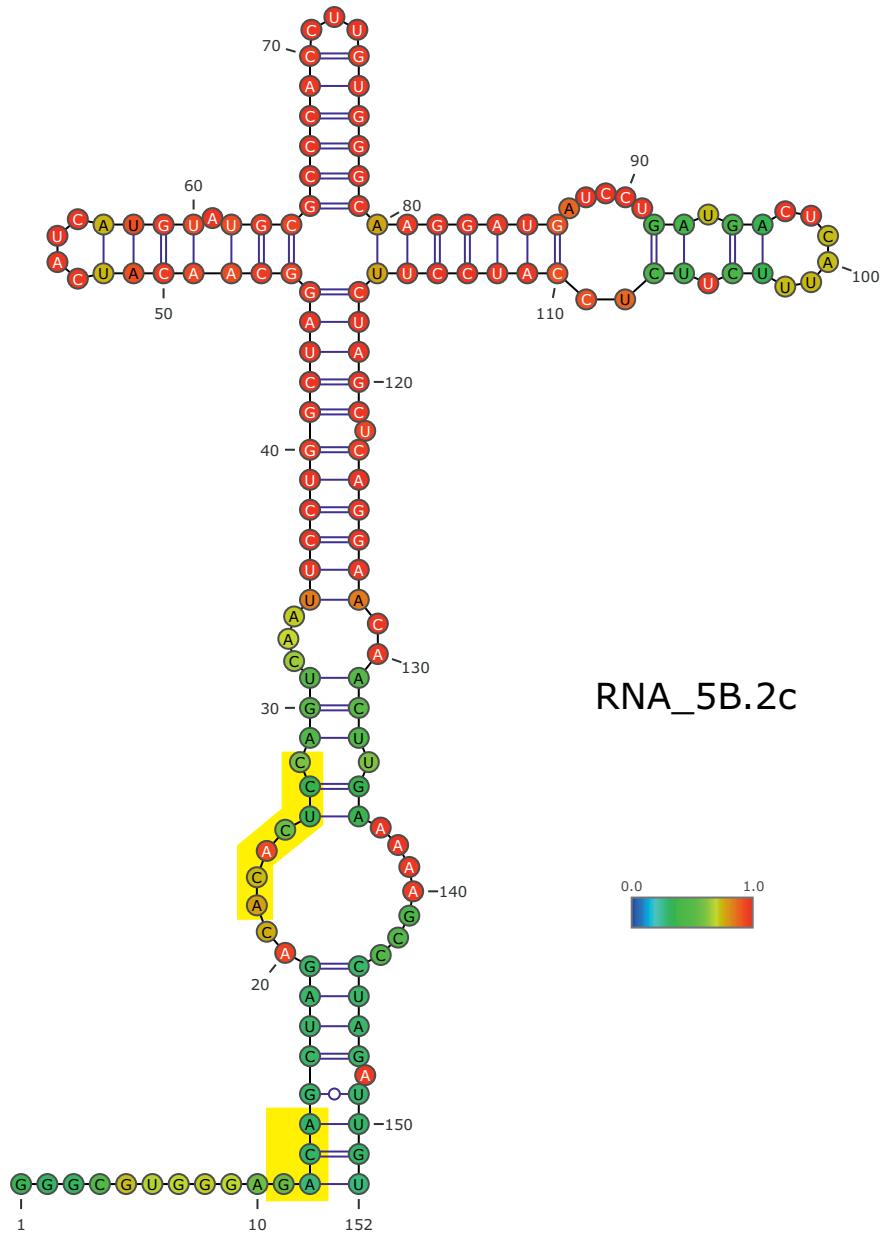
Supplementary Figure S7, related to Figure 3

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



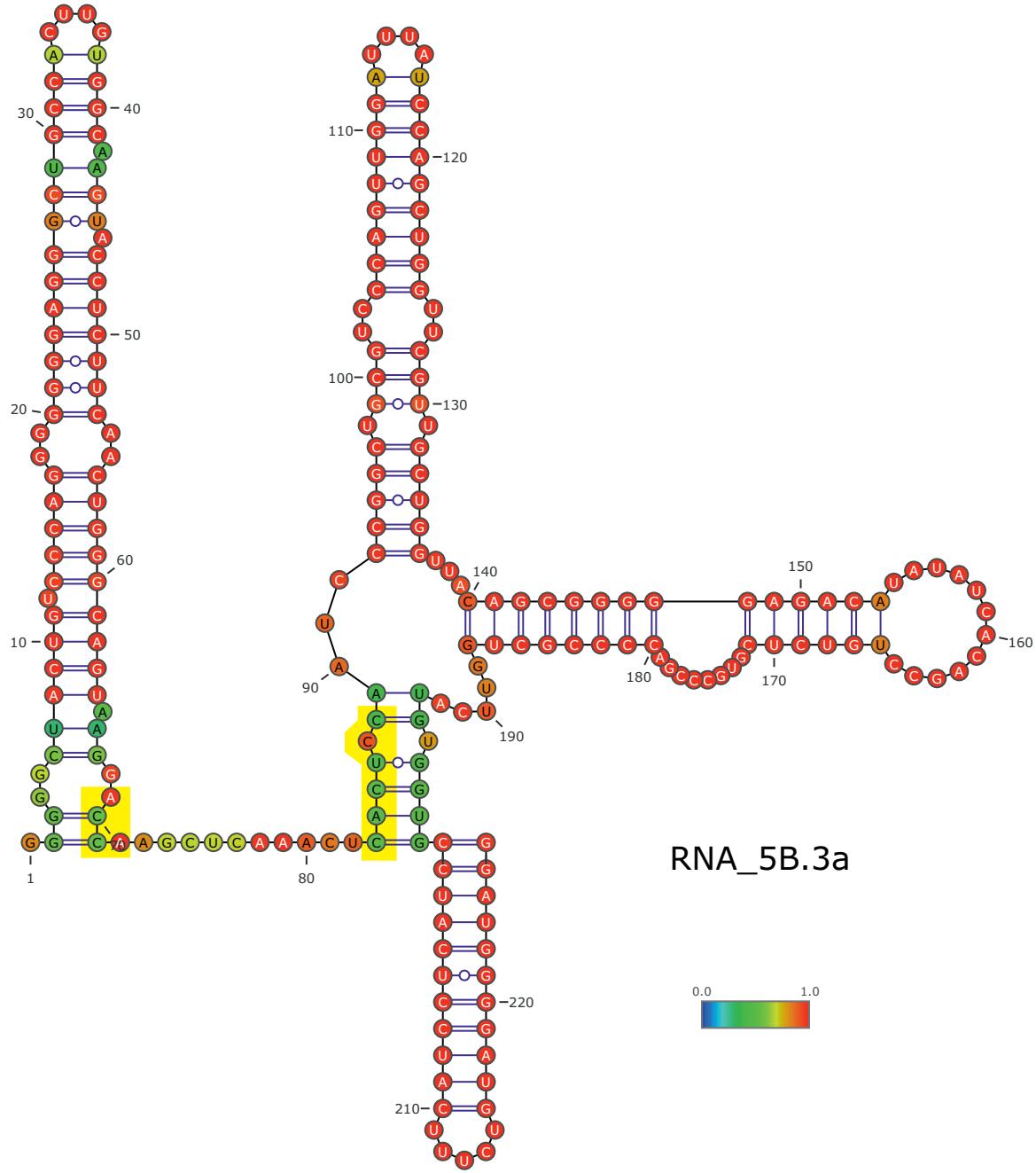
Supplementary Figure S8, related to Figure 3

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



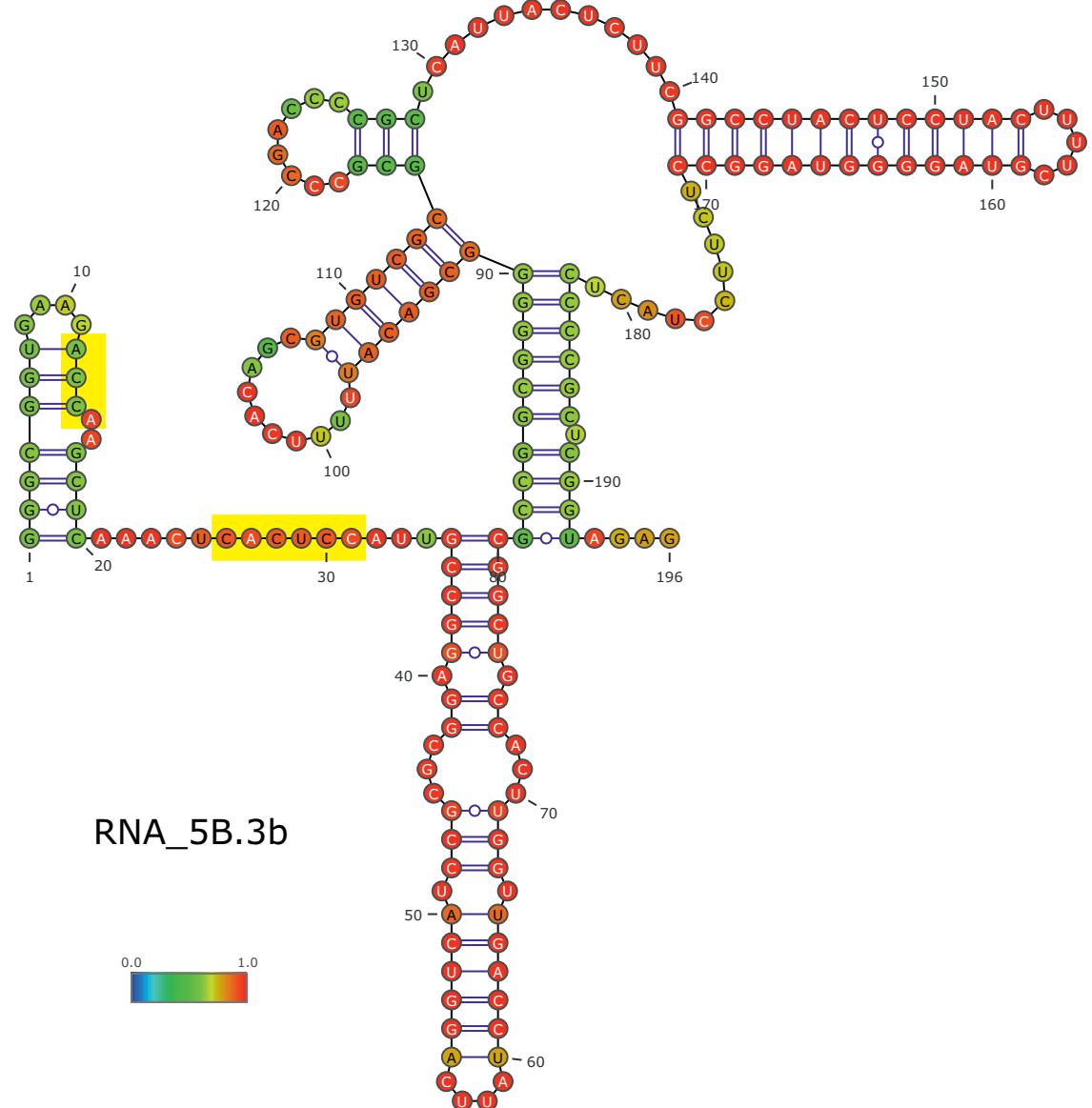
Supplementary Figure S9, related to Figure 3

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



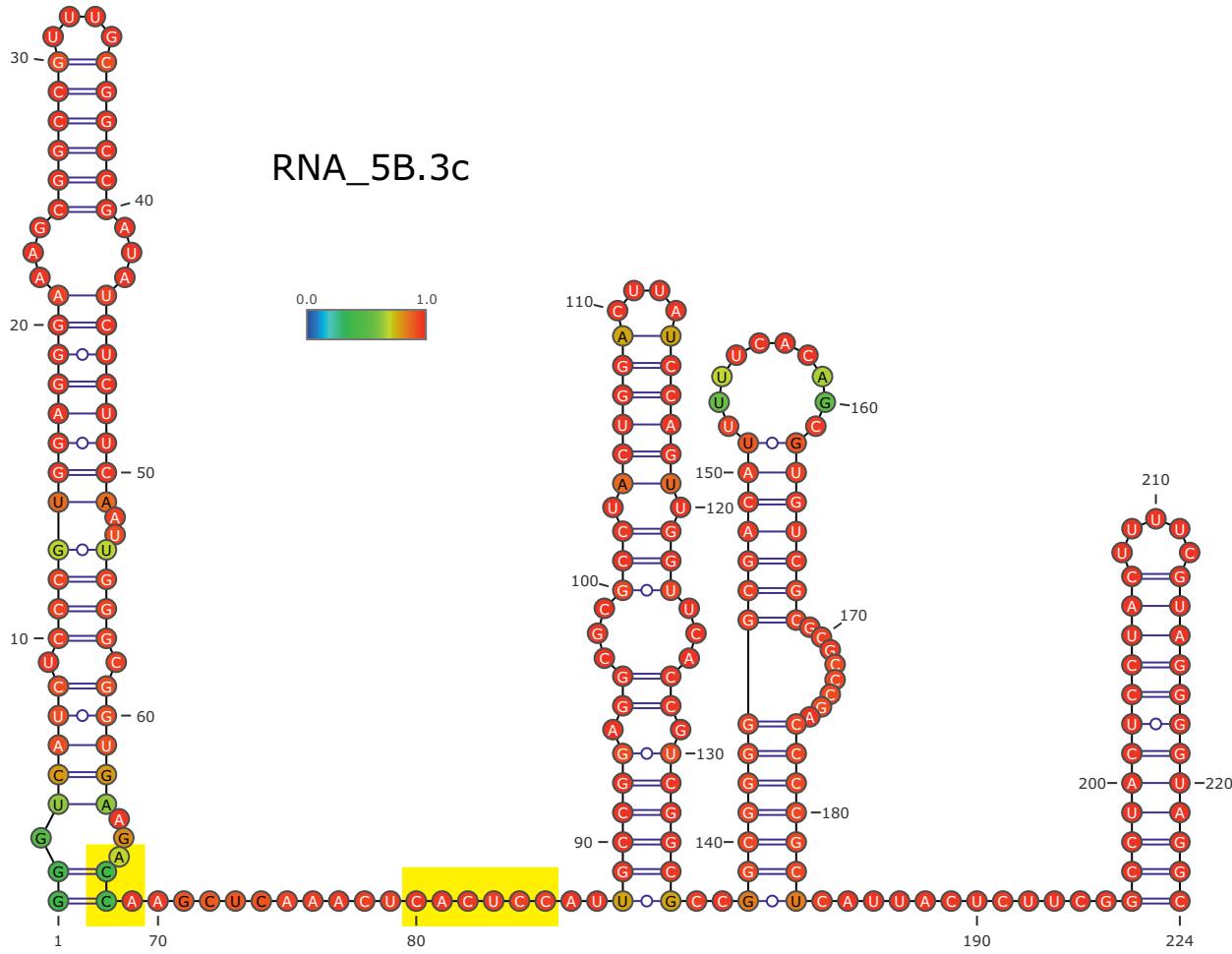
Supplementary Figure S10, related to Figure 4

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



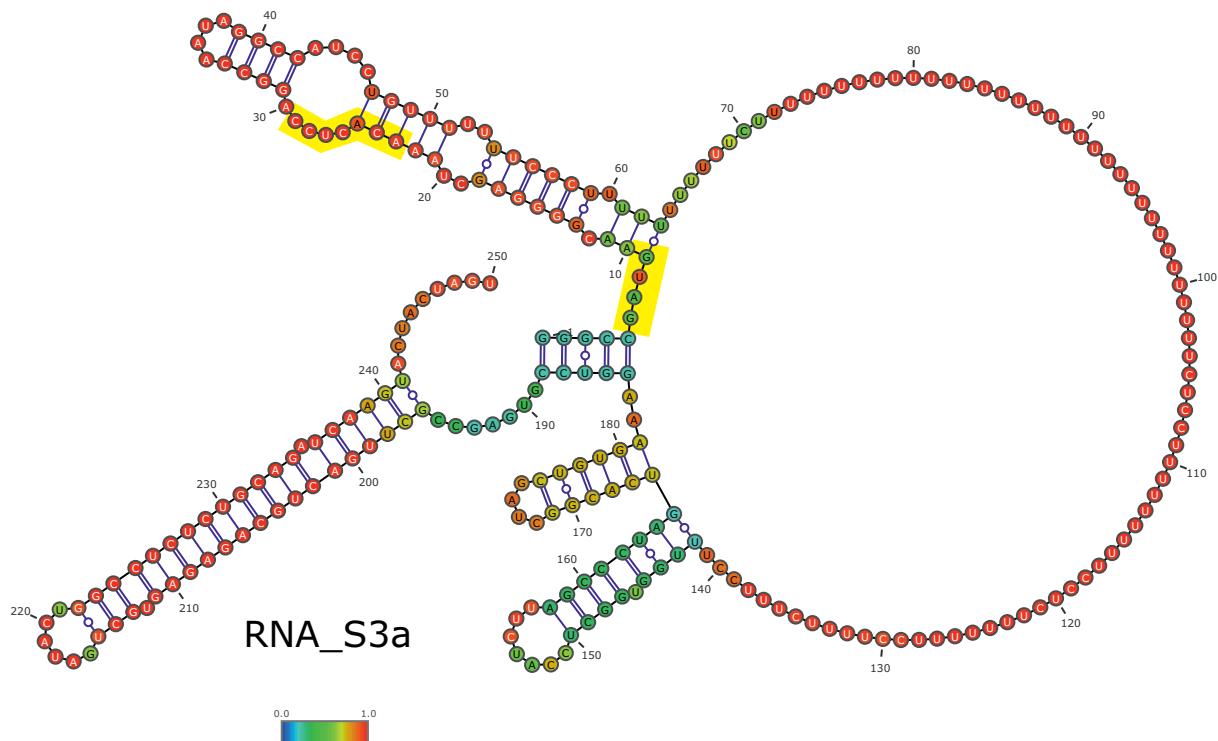
Supplementary Figure S11, related to Figure 4

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



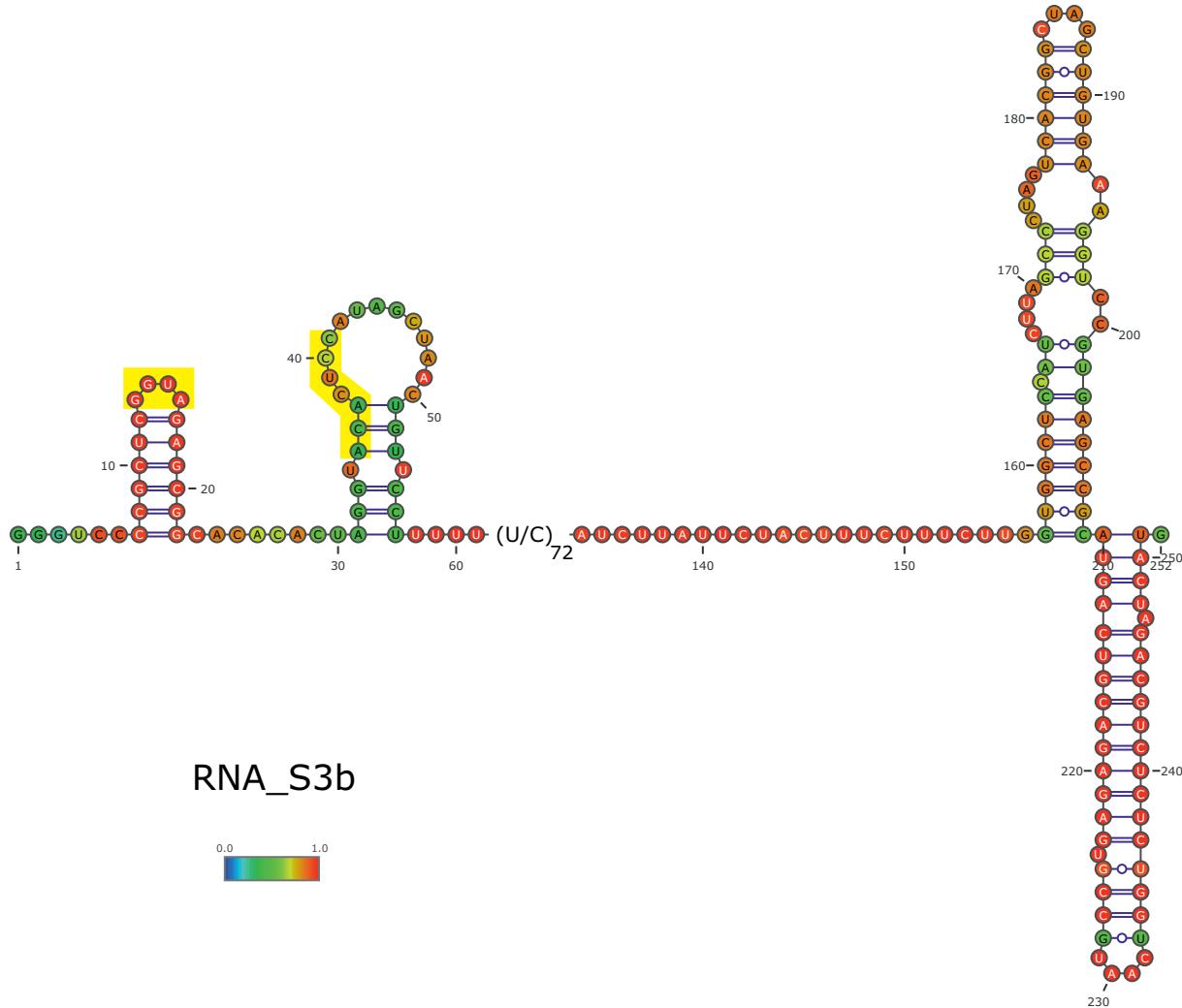
Supplementary Figure S12, related to Figure 4

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



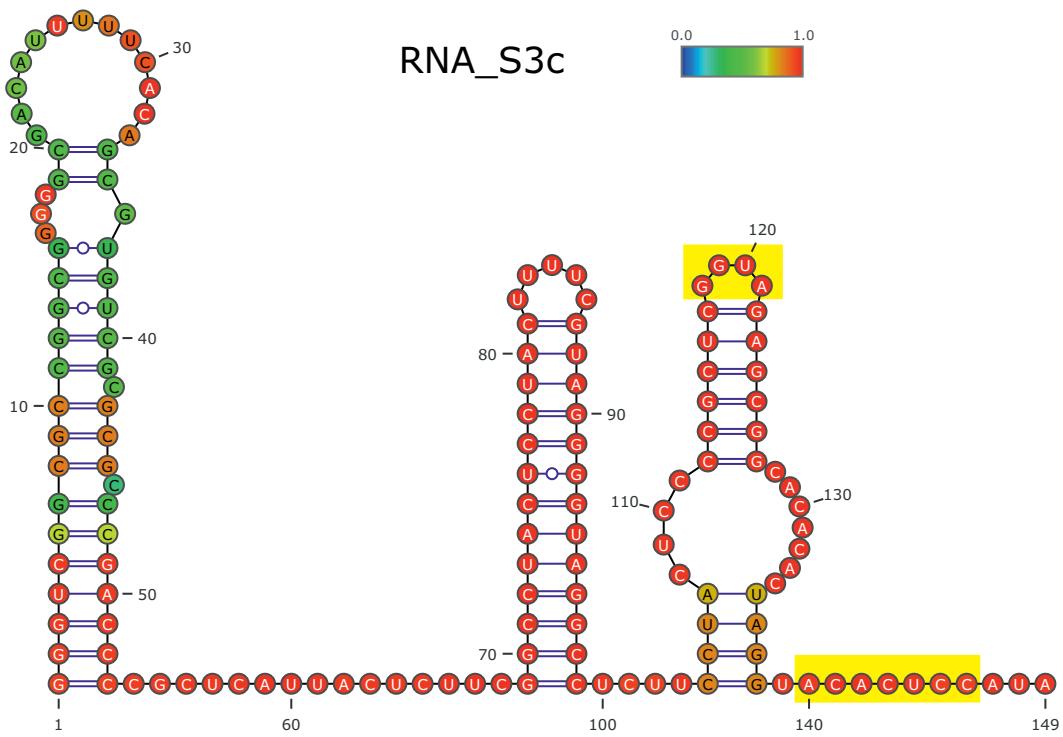
Supplementary Figure S13, related to Figure 5

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



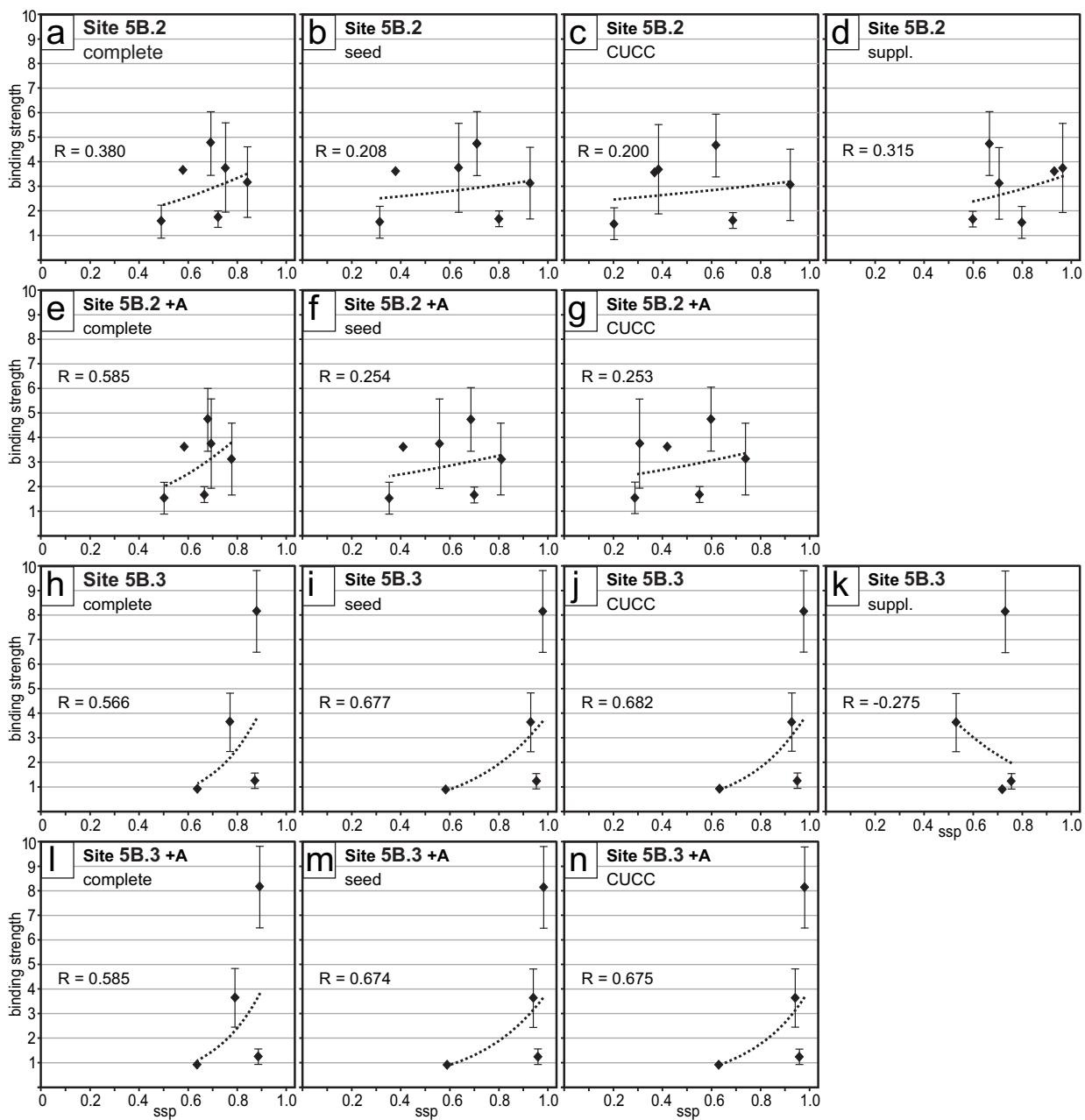
Supplementary Figure S14, related to Figure 5

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



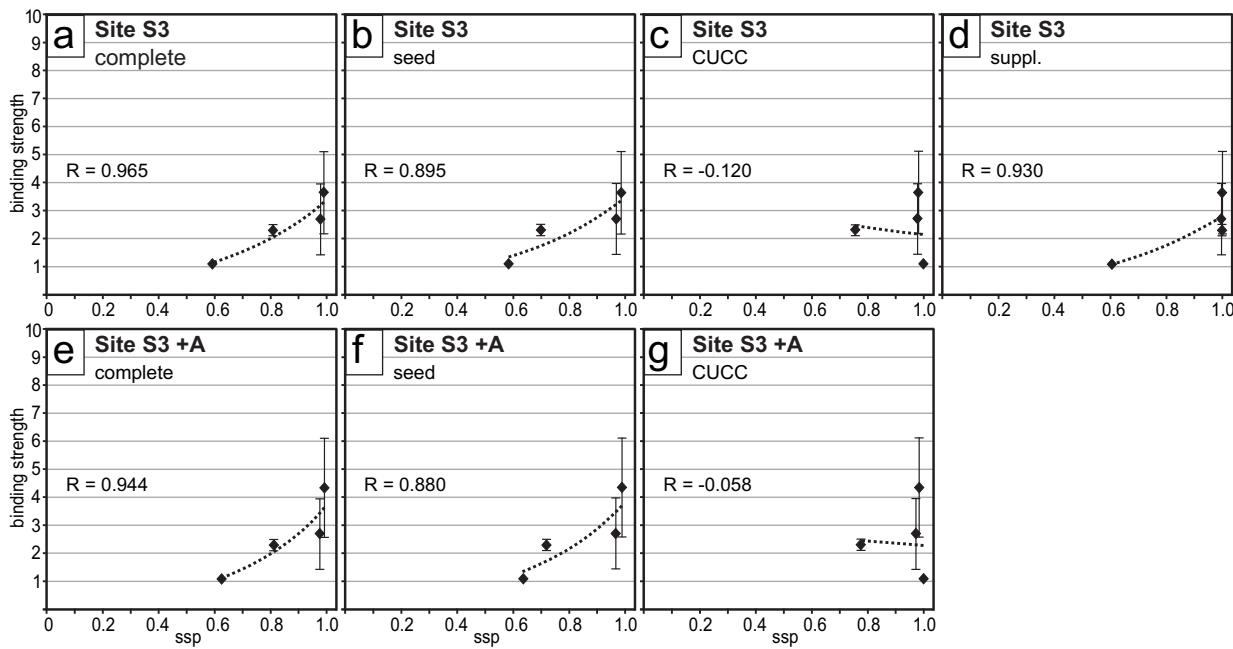
Supplementary Figure S15, related to Figure 5

Predicted target RNA minimum free energy (MFE) structures. Primers for transcription template production, RNA sequences and Vienna dot bracket outputs are shown in Supplementary Figure S6. Base pair probability (bpp) values were loaded into VARNAv3-92 (Darty et al. 2009) from the Vienna dot plot EPS file output (Gruber et al. 2008; Lorenz et al. 2011), and the bpp color code was set to Vienna style. The Vienna MFE output returns bpp values for bases predicted to pair, but single strand probabilities ($ssp = 1 - bpp$) for bases predicted to be unpaired.



Supplementary Figure S16, related to Figures 3 - 5

The intensity of miR-122/Ago2 complex binding to NS5B target sites plotted against target site accessibility. The intensity of miR-122/Ago2 complex binding to each target site was calculated from the phosphorimager scans (examples shown in Figures 2, 3 and 4) and several additional experiments using all RNAs listed in Supplementary Figure 6. Binding strength was plotted against single strand probability (ssp) values (calculated as described in Materials and Method). Error bars indicate standard deviations. "R" values are correlation coefficients for regression curves. Panels show plots against average single strand probabilities of the complete target site and separately for seed sequence, nucleotides opposite nucleotides 2 – 5 of the miRNA ("CUCC") and for the supplementary site as indicated. The A residue was additionally included in the calculations in plots labeled "+A".



Supplementary Figure S17, related to Figures 3 - 5

The intensity of miR-122/Ago2 complex binding to 3'UTR S3 target sit plotted against target site accessibility. The intensity of miR-122/Ago2 complex binding to each target site was calculated from the phosphorimager scans (examples shown in Figures 2, 3 and 4) and several additional experiments using all RNAs listed in Supplementary Figure 6. Binding strength was plotted against single strand probability (ssp) values (calculated as described in Materials and Method). Error bars indicate standard deviations. "R" values are correlation coefficients for regression curves. Panels show plots against average single strand probabilities of the complete target site and separately for seed sequence, nucleotides opposite nucleotides 2 – 5 of the miRNA ("CUCC") and for the supplementary site as indicated. The A residue was additionally included in the calculations in plots labeled "+A".

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