

Supplemental information

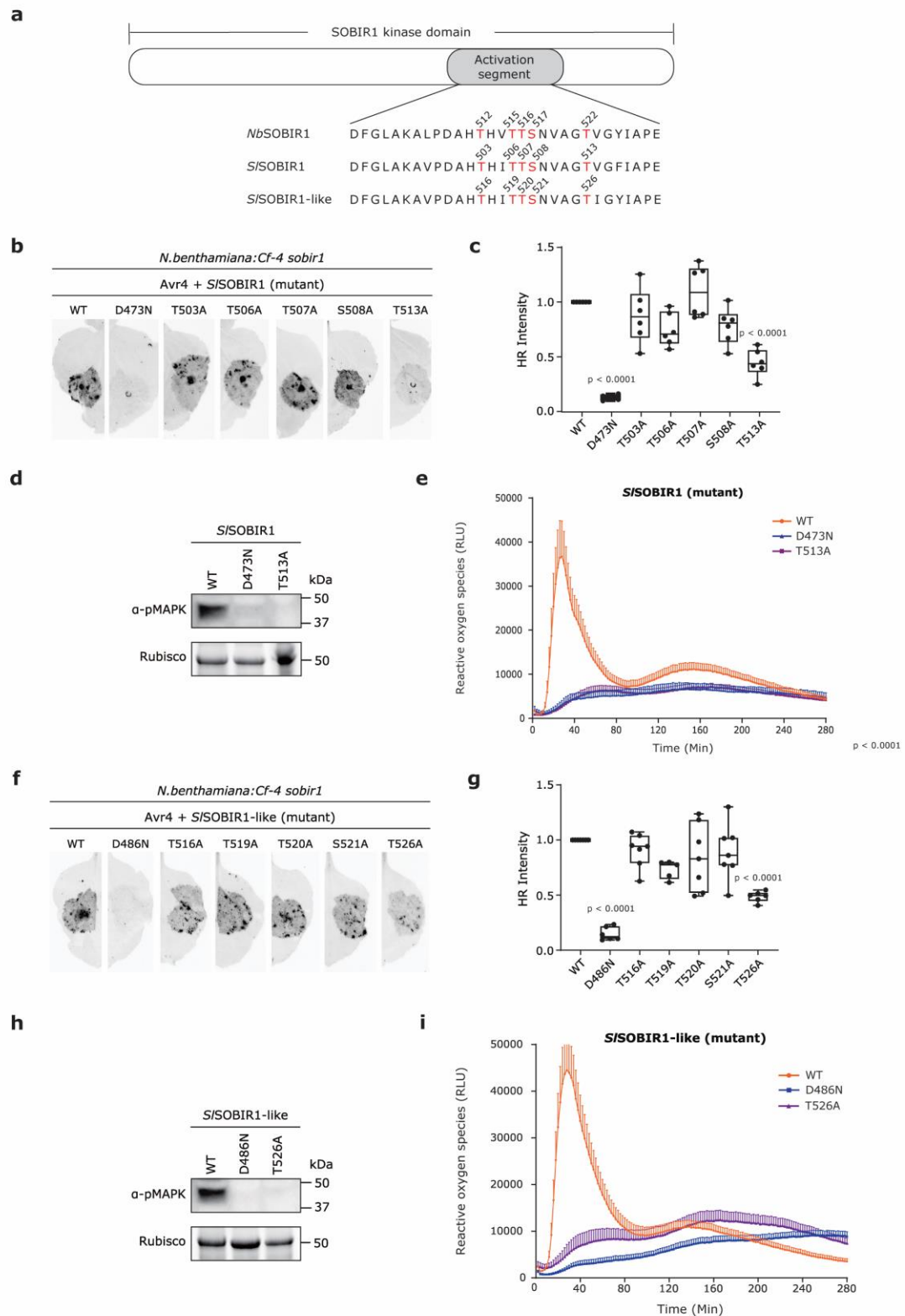
SOBIR1 homologs

Nicotiana benthamiana_Niben101Scf03816g01001.1 (<i>NbSOBIR1</i>)	DFGLAKA --L---PDAH-----	↓ ^{S12}	↓ ^{S15}	↓ ^{S16}	↓ ^{S17}	↓ ^{S22}	THVTTSNVAGTVGYIAPE
Solanum lycopersicum_Solyc06g071810.1 (<i>S/SOBIR1</i>)	DFGLAKA --V---PDAH-----						THITTSNVAGTVGFI APE
Solanum lycopersicum_Solyc03g111800.2 (<i>S/SOBIR1-like</i>)	DFGLAKA --V---PDAH-----						THITTSNVAGTIGYIAPE
Arabidopsis thaliana_AT2G31880 (<i>AtSOBIR1</i>)	DFGLAKA --M---PDAV-----						THITTSNVAGTVGYIAPE
Nicotiana tabacum_A0A1S3XP18	DFGLAKA --L---PDAH-----						THITTSNVAGTVGYIAPE
Nicotiana sylvestris_A0A1U7WH59	DFGLAKA --L---PDAH-----						THITTSNVAGTVGYIAPE
Nicotiana attenuata_A0A1J6INR1	DFGLAKA --L---PDAH-----						THITTSNVAGTVGYIAPE
Nicotiana tabacum_A0A1S3ZD83	DFGLAKA --V---PDAH-----						THVTTSNVAGTVGYIAPE
Solanum tuberosum_M1CKT4	DFGLAKA --V---PDAH-----						THITTSNVAGTMGYIAPE
Capsicum chinense_A0A2G3BFP7	DFGLAKA --V---PDAH-----						THVTTSNVAGTVGYIAPE
Capsicum baccatum_A0A2G2WM84	DFGLAKA --V---PDAH-----						THVTTSNVAGTVGYIAPE
Capsicum annuum_A0A2G2Y0L5	DFGLAKA --V---PDAH-----						THVTTSNVAGTVGYIAPE
Solanum tuberosum_M1B7X0	DFGLAKA --V---PDAH-----						THITTSNVAGTIGYIAPE
Capsicum chinense_A0A2G3CCQ8	DFGLAKA --V---PDAH-----						THITTSNVAGTIGYIAPE
Capsicum annuum_A0A1U8G6T0	DFGLAKA --V---PDAH-----						THITTSNVAGTIGYIAPE
Capsicum baccatum_A0A2G2X8C5	DFGLAKA --V---PDAH-----						THITTSNVAGTIGYIAPE
Nicotiana attenuata_A0A314KS23	DFGLAKA --I---PE ^S L-----						THVSTSHVVGTLGYIAPA
Coffea arabica_A0A6P6X9H5	DFGLAKA --M---PEAY-----						THVTTSSNVVGT LGYIAPE
Coffea arabica_A0A6P6X153	DFGLAKA --M---PEAY-----						THVTTSSNVVGT LGYIAPE
Coffea canephora_A0A068V365	DFGLAKA --M---PEAY-----						THVTTSSNVVGT LGYIAPE
Sesamum indicum_A0A6I9SRZ7	DFGLAKA --M---PDAN-----						THVTTSNVAGTVGYIAPE
Olea europaea subsp. europaea_A0A8S0R730	DFGLAKA --V---PEAN-----						THVSTSNVAGTAGYIAPE
Camellia sinensis var. sinensis_A0A4S4DBV0	DFGLAKA --V---PDAH-----						THVTTSNVAGTVGYIAPE
Rhododendron simsii_A0A834L953	DFGLAKA --V---PDQD-----						THVTTSNVAGT LGYIAPE
Vitis vinifera_F6GTQ3	DFGLAKA --V---PDAN-----						THVTTSNVAGTVGYIAPE
Tetradactylon sinense_A0A834Y9S3	DFGLAKA --V---PDAN-----						THVTTSNVAGTVGYI ^S PE
Carpinus fangiana_A0A660KKL0	DFGLAKA --M---PDAH-----						THISTSNVAGTVGYIAPE
Prunus persica_M5WTY7	DFGLAKA --V---PEYH-----						THITTSNVAGTVGYIAPE
Quercus lobata_A0A7N2MNZ2	DFGLAKA --M---PDAN-----						THITTSNVAGTVGYI ^S PE
Tetradactylon sinense_A0A834YD49	DFGLAKA --V---PDAN-----						THVTTSNVAGTVGYI ^S PE
Handroanthus impetiginosus_A0A2G9HZY5	DFGLAKA --V---PDAN-----						THVTTSNVAGTVGYIAPE
Prunus armeniaca_A0A6J5XG14	DFGLAKA --V---PEYH-----						THITTSNVAGTVGYIAPE
Actinidia chinensis var. chinensis_A0A2R6PUF8	DFGLAKA --V---PDAY-----						THVTTSNVAGTVGYIAPE
Castanea mollissima_A0A8J4REW1	DFGLAKA --M---PDAH-----						THITTSNVAGTVGYI ^S PE
Handroanthus impetiginosus_A0A2G9IBC5	DFGLAKA --V---PDAN-----						THVTTSNVAGTVGYIAPE
Quercus lobata_A0A7N2MML4	DFGLAKA --M---PDAN-----						THISTSNVAGTVGYI ^S PE
Jatropha curcas_A0A067KGN0	DFGLAKA --M---PDAQ-----						THVSTSNVAGTVGYIAPE

Other RLKs

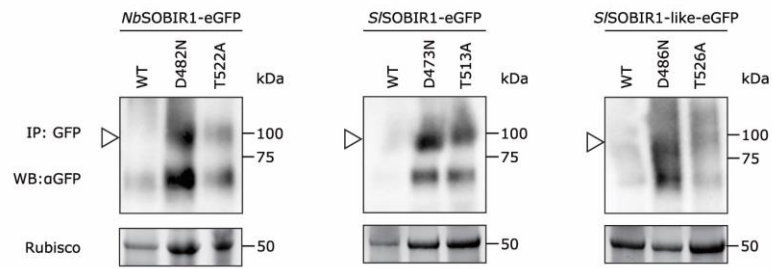
Nicotiana benthamiana_Niben101Scf02128g00022.1 (<i>NbBAK1</i>)	DFGLAKL --M---DYKD-----	THVTTA -VRGT IGHIAPE
Solanum lycopersicum_Solyc10g047140.2 (<i>S/BAK1</i>)	DFGLAKL --M---DYKD-----	THVTTA -VRGT IGHIAPE
Arabidopsis thaliana_AT4G33430 (<i>AtBAK1</i>)	DFGLAKL --M---DYKD-----	THVTTA -VRGT IGHIAPE
Arabidopsis thaliana_AT3G21630 (<i>AtCERK1</i>)	DFGLTKL --T---EVGG-----	SATRGAMGTFGYMAPE
Arabidopsis thaliana_AT5G16000 (<i>AtNIK1</i>)	DFGLAKL --L---DHQD-----	SHVTTA -VRGT VGHIAPE
Arabidopsis thaliana_AT4G39400 (<i>AtBRI1</i>)	DFGMARL --M---SAMD-----	THLSVSTLAGTPGYVPE
Arabidopsis thaliana_AT4G28490 (<i>AtHAESA</i>)	DFGIAKVGQM---SGSK-----	TPEAMSGIAGSGYIAPE
Arabidopsis thaliana_AT5G20480 (<i>AtEFR</i>)	DFGLAQL --L---YKYDRESFLNQFSSAGVVRGTIGYAAPE	
Arabidopsis thaliana_AT5G46330 (<i>AtFLS2</i>)	DFGTARI --LGFREDGSS-----	TTASTSAFEGTIGYLAPE

Supplementary Figure 1. Five potential phosphorylation sites in the activation segment of *NbSOBIR1* are highly conserved in the kinase domain of various RLKs from different plant species. Protein sequences of the SOBIR1 homologs from various plant species were obtained from the UniProt database (<https://www.uniprot.org/>), while the protein sequences of other RLKs were retrieved from TAIR (<https://www.arabidopsis.org/index.jsp>) and the Sol Genomics Network (<https://solgenomics.net/>). The alignment was visualized using JalView and only the amino acid sequences of the activation segment of the kinase domain are shown. All the putatively phosphorylatable Ser (S) and Thr (T) residues are highlighted in orange. The positions of the five Ser/Thr residues of *NbSOBIR1* that are subjected to this study are indicated on top.



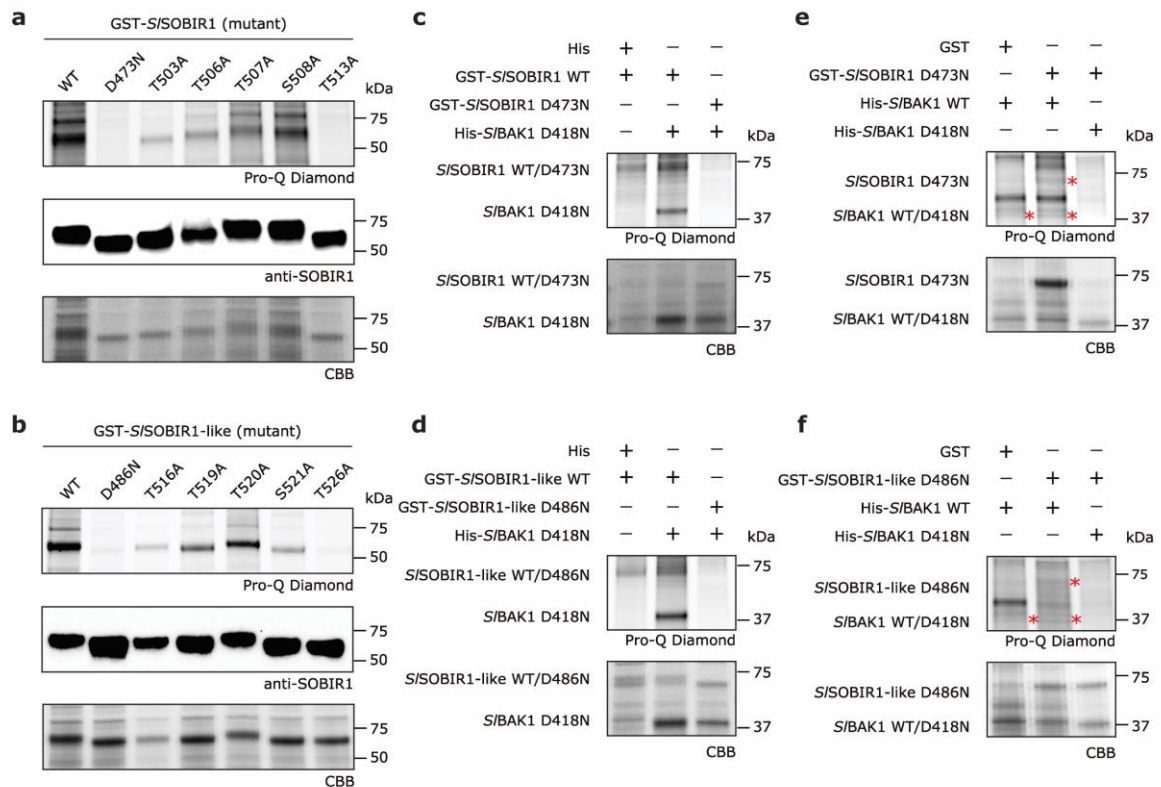
Supplementary Figure 2. The analogous residues of *NbSOBIR1* Thr522 in both tomato *SOBIR1* and *SOBIR1*-like play a crucial role in mounting *Avr4*/*Cf-4*-triggered immune responses. **a**, Schematic diagram of the kinase domain of *SOBIR1*, with the activation segment indicated. The amino acid sequences of the activation segments of *NbSOBIR1*, *S/SOBIR1* and

S/SOBIR1-like are aligned and are shown below the diagram. Conserved residues acting as potential phosphorylation sites are indicated in red. **b-e**, Mutagenesis screen of five potential phosphorylation sites in the activation segment of *S/SOBIR1* based on the *Avr4/Cf-4*-triggered HR activation (**b** and **c**), MAPK activation (**d**), and ROS burst (**e**). **f-i**, Mutagenesis screen of five potential phosphorylation sites in the activation segment of *S/SOBIR1*-like based on the *Avr4/Cf-4*-triggered HR activation (**f** and **g**), MAPK activation (**h**), and ROS burst (**i**). The development of an HR was imaged at 5 dpi by the Chemidoc (**b** and **f**), and the HR intensity was quantified by Image Lab (**c** and **g**). The HR intensity of leaves that transiently co-express *S/SOBIR1 WT* with *Avr4* (**c**) or *S/SOBIR1-like WT* with *Avr4* (**g**) was set as 1. Statistical analysis was performed with an ANOVA/Dunnett's multiple comparison test, compared with their corresponding WT. Dots indicate individual values (centre line, median; error bar, minima and maxima; n=6). **e, i**, Leaf discs of *N. benthamiana:Cf-4 sobir1* knock-out plants that transiently express individual *S/SOBIR1* and *S/SOBIR1*-like mutants, as well as their corresponding WTs and kinase-dead mutants, were treated with 0.1 μ M *Avr4* protein, followed by measuring the production of ROS. ROS production is expressed as RLUs, and the data are represented as mean + SEM (n=12). Similar to *SOBIR1 WT*, all tested *SOBIR1 Ser/Thr* mutants restored the *Avr4/Cf-4*-triggered ROS production in this complementation study, except for *S/SOBIR1 T513A* and *S/SOBIR1-like T526A*. Only the results for *S/SOBIR1 WT*, *T513A* and *D473N* are shown in (**e**), and the results for *S/SOBIR1-like WT*, *T526A*, and *D486N* are shown in (**i**). Experiments were repeated at least three times, with similar results. Source data are provided as a Source Data file.



Supplementary Figure 3. Accumulation levels of *NbSOBIR1* T522A, *S/SOBIR1* T513A and *S/SOBIR1*-like T526A *in planta*, in comparison to their WT and kinase-dead (“RN”) versions.

NbSOBIR1 T522A, *S/SOBIR1* T513A and *S/SOBIR1*-like T526A, which fail to restore the *Avr4*-triggered HR in *N. benthamiana*:*Cf-4 sobir1* knock-out plants, were transiently co-expressed in *N. benthamiana*:*Cf-4 sobir1* plants with *Avr4* (both at an OD₆₀₀ of 0.8). Transient expression of their respective WT and kinase-dead D to N version, in combination with *Avr4*, was included as positive and negative controls, respectively. Leaf samples were collected at 2 dpi and total protein extracts were subjected to immuno-purification (IP) using GFP-affinity beads, followed by western blotting (WB) with αGFP antibody (upper panels). The amount of total protein that was used for the IP is reflected by the Rubisco band present in the total protein extracts loaded on SDS gel (lower panels). Arrowheads indicate the band corresponding to SOBIR1-eGFP. Note that transient expression of WT *SOBIR1* in combination with *Avr4* results in cell death, thereby causing very low WT *SOBIR1* accumulation levels.



Supplementary Figure 4. The kinase domains of SOBIR1 and BAK1 trans-phosphorylate each other *in vitro*. **a, b**, *S/OBIR1* Thr513 and *S/OBIR1*-like Thr526 are essential for their intrinsic kinase activity. The N-terminally GST-tagged cytoplasmic kinase domains of (**a**) *S/OBIR1* WT, kinase-dead mutant D473N, and five Ser/Thr-to Ala mutants, and (**b**) *S/OBIR1*-like WT, kinase-dead mutant D486N, and five Ser/Thr-to-Ala mutants were produced in *E. coli*, followed by being subjected to western blotting and *in vitro* phosphorylation assay. The phosphorylation status of the recombinant proteins was determined by using the Pro-Q Diamond stain, which specifically stains the phosphorylated proteins (top panels). The production of SOBIR1 kinase domains was confirmed by western blotting, using SOBIR1 antibodies (middle panels), and the recombinant proteins were stained by Coomassie brilliant blue (bottom panels). **c,d**, *S/OBIR1* WT and *S/OBIR1*-like WT directly phosphorylate the kinase-dead mutant of *S/BAK1* D418N. The N-terminally GST-tagged cytoplasmic domains of *S/OBIR1* WT or D473N (**c**) and *S/OBIR1*-like WT or D486N (**d**) were co-expressed with the N-terminally His-tagged cytoplasmic domain of *S/BAK1* D418N in *E. coli*. After SDS-PAGE of the *E. coli* lysates, a Pro-Q Diamond stain was employed to detect the phosphorylated recombinant proteins (top panels), while Coomassie brilliant blue was used to visualize all proteins (bottom panels). **e,f**, *S/BAK1* WT directly phosphorylates the kinase-dead mutant of *S/OBIR1* D473N and *S/OBIR1*-like D486N. The cytoplasmic domain of *S/BAK1* WT or D418N, which was fused to a His tag at its N-terminus, was co-expressed with the cytoplasmic domain of *S/OBIR1* D473N (**e**) or *S/OBIR1*-like D486N (**f**), which was fused to a GST tag at its N-terminus, in *E. coli*. The recombinant proteins were then subjected to SDS-PAGE, followed by being stained by Pro-Q Diamond stain (top panels) and Coomassie brilliant blue (bottom panels). Bands with the expected sizes are indicated with an asterisk.

Experiments were repeated at least three times with similar results, and representative pictures are shown. Source data are provided as a Source Data file.

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Nicotiana benthamiana_Niben101Scf03816g01001.1 (*NbSOBIR1*) VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAELTEEDTKALNKKMRQVKSEIQILGQIRHRNLLPLL
Solanum lycopersicum_Solyc06g071810.1 (*S/SOBIR1*) VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQSPMDAAEITEEDTKALNKKMRQVKSEIQIVGQIRHRNLLPLL
Solanum lycopersicum_Solyc03g111800.2 (*S/SOBIR1*-like) LESLELIGQGGCGKVIKAAALPGSDGKIIAVKKIIQPPKDAEELTEEDSKAMNKKMRQIKSEIKIVGQIRHRNLLPLL
Arabidopsis_thaliana_AT2G31880 (*AtSOBIR1*) LASLELIGRGGCGEVFKAAELPGSNGKIIAVKKIIQPPKDAEELTEEDSKFLNKKMRQIRSEINTVGHIRHRNLLPLL
Nicotiana tabacum_A0A1S3XP18 VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAELTEEDTKALNKKMRQVKSEIQILGQIRHRNLLPLL
Nicotiana sylvestris_A0A1U7WH59 VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAELTEEDTKALNKKMRQVKSEIQILGQIRHRNLLPLL
Nicotiana attenuata_A0A1J6INR1 VASLEMVGGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAELTEEDTKALNKKMRQVKSEIQILGQIRHRNLLPLL
Nicotiana tabacum_A0A1S3ZD83 VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAELTEEDTKALNKKMRQVKSEIQILGQIRHRNLLPLL
Solanum tuberosum_M1CKT4 VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQSPMDAAEITEEDTKALNKKMRQVKSEIQIVGQIRHRNLLPLL
Capsicum chinense_A0A2G3BFP7 VASLEMVGGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAEITEEDTKALNKKMRQVKSEIQIVGQIRHRNLLPLL
Capsicum baccatum_A0A2G2WM84 VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAEITEEDTKALNKKMRQVKSEIQIVGQIRHRNLLPLL
Capsicum annuum_A0A2G2Y0L5 VASLEMIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAEITEEDTKALNKKMRQVKSEIQIVGQIRHRNLLPLL
Solanum tuberosum_M1B7X0 LESLELIGQGGCGKVIKAAALPGSDGKIIAVKKIIQPPMDAAELTEEDSKAMSKMRQIKSEIKIVGQIRHRNLLPLL
Capsicum chinense_A0A2G3CCQ8 FESLELIGEGGCGKVIKAAELPGSNGKIIAVKKIIQPPMDAAELTEEDSKAMHKKMRQIKSEIKIVGQIRHRNLLPLL
Capsicum annuum_A0A1U8G6T0 FESLELIGEGGCGKVIKAAELPGSNGKIIAVKKIIQPPMDAAELTEEDSKAMHKKMRQIKSEIKIVGQIRHRNLLPLL
Capsicum baccatum_A0A2G2X8C5 FESLELIGEGGCGKVIKAAELPGSNGKIIAVKKIIQPPMDAAELTEEDSKAMHKKMRQIKSEIKIVGQIRHRNLLPLL
Nicotiana attenuata_A0A314KS23 LALLELIRKGGCGEVYKAAELPGSNGKIIAIIKKIIVPPKDAEELTEEDSKALNKKMRQIKSEIKIVGQIRHRNLLPLL
Coffea arabica_A0A6P6X9H5 IARLEVIGRGGCGEVYKAAELPGSNGKIIAIIKKIIQPPMDAAELTEEDSKALHKKMRQIKSEIQTVGQIRHRNLLPLL
Coffea arabica_A0A6P6X153 IARLEVIGRGGCGEVYKAAELPGSNGKIIAIIKKIIQPPMDAAELTEEDSKALHKKMRQIKSEIQTVGQIRHRNLLPLL
Coffea canephora_A0A068V365 IARLEVIGRGGCGEVYKAAELPGSNGKIIAIIKKIIQPPMDAAELTEEDSKALHKKMRQIKSEIQTVGQIRHRNLLPLL
Sesamum indicum_A0A6I9SRZ7 LASLDVIGRGGCGEVFKAVLPGSNGKIIAIIKKIIQPPMDAAELTEEDSKMMNKKMRQIRSEIQTVGQIRHRNLLPLL
Olea europaea subsp. europaea_A0A8S0R730 LNLGELVIGRGGCGEVYKATLPGTNGKIIAVKKIIQPPMDAAELTEEDSKLNNKKMRQIRSEIQTVGQIRHRNLLPLL
Camellia sinensis var. sinensis_A0A4S4DBV0 LASLELIGSGGCGIVYKAAELPGSNGKIIAIIKKIIQPPMDAAELTEEDSKLNNKKMRQIRSEIQTVGQIRHRNLLPLL
Rhododendron simsii_A0A834L953 LASLELIGRGGCGEVYKAAELPGSNGKIIAIIKKIIQPPMDAAELTEEDSKLNNKKMRQIRSEIQTVGQIRHRNLLPLL
Vitis vinifera_F6GTQ3 LASLELIGKGGCGEVYRAELPGSNGKIIAIIKKIIQPPMDAAELTEEDSKLNNKKMRQIRSEIQTVGQIRHRNLLPLL

426 429 431 469

Nicotiana benthamiana_Niben101Scf03816g01001.1 (*NbSOBIR1*) AHMPRPDCHYLVEEYMKNGSLQDILQQVTEGTRREL DWLGRHR IAVG IASGLEYLHINHNSQCI IHRDLKPANVLLDDD
Solanum lycopersicum_Solyc06g071810.1 (*S/SOBIR1*) AHMPRPDCHYLVEEYMKNGSLQDILQQVTEGTRREL DWLGRHR IAVGVAAGLEYLHINHNSQCI IHRDLKPANVLLDDD
Solanum lycopersicum_Solyc03g111800.2 (*S/SOBIR1*-like) AHMPRPDCHYLVEEYMKNGSLQDILQQVREGKREL DWSARHR IAVGVAAGLEYLHINHNSQCI IHRDLKPGNVLLDDD
Arabidopsis_thaliana_AT2G31880 (*AtSOBIR1*) AHVSRPECHYLVEEYMEKNGSLQDILTQVQAGNQLMWP ARHK IALG IAVGLEYLHMDHNPRI IHRDLKPANVLLDDD
Nicotiana tabacum_A0A1S3XP18 AHMPRPDCHYLVEEYMKNGSLQDILQQVTEGTRREL DWLGRHR IAVG IAVGLEYLHINHNSQCI IHRDLKPANVLLDDD
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Solanum tuberosum_M1CKT4 AHMPRPDCHYLVEEYMKNGSLQDILQQVTEGTRREL DWLGRHR IAVGVAAGLEYLHINHNSQCI IHRDLKPANVLLDDD
Capsicum chinense_A0A2G3BFP7 AHMPRPDCHYLVEEYMKNGSLQDILQQVTEGTRREL DWLGRHR IAVG IAVGLEYLHINHNSQCI IHRDLKPANVLLDDD
Capsicum baccatum_A0A2G2WM84 AHMPRPDCHYLVEEYMKNGSLQDILQQVTEGTRREL DWLGRHR IAVG IAVGLEYLHINHNSQCI IHRDLKPANVLLDDD
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Sesamum indicum_A0A6I9SRZ7 AHLPRPDCHYLVEEYMKNGSLQDMLQKVAAGENEL DWLGRHR IAVGVAAGLEYLHVNHTPRI IHRDLKPANVLLDDD
Olea europaea subsp. europaea_A0A8S0R730 AHLPRPDCHYLVEEYMKNGSLQDMLQKVAAGENEL DWLGRHR IAVGVAAGLEYLHVNHTPRI IHRDLKPANVLLDDD
Camellia sinensis var. sinensis_A0A4S4DBV0 AHVSRPDCHYLVEEYMKNGSLQDILNQVSGTREL DWQARHK IAVGVAAGLEYLHVNHTPRI IHRDLKPANVLLDDD
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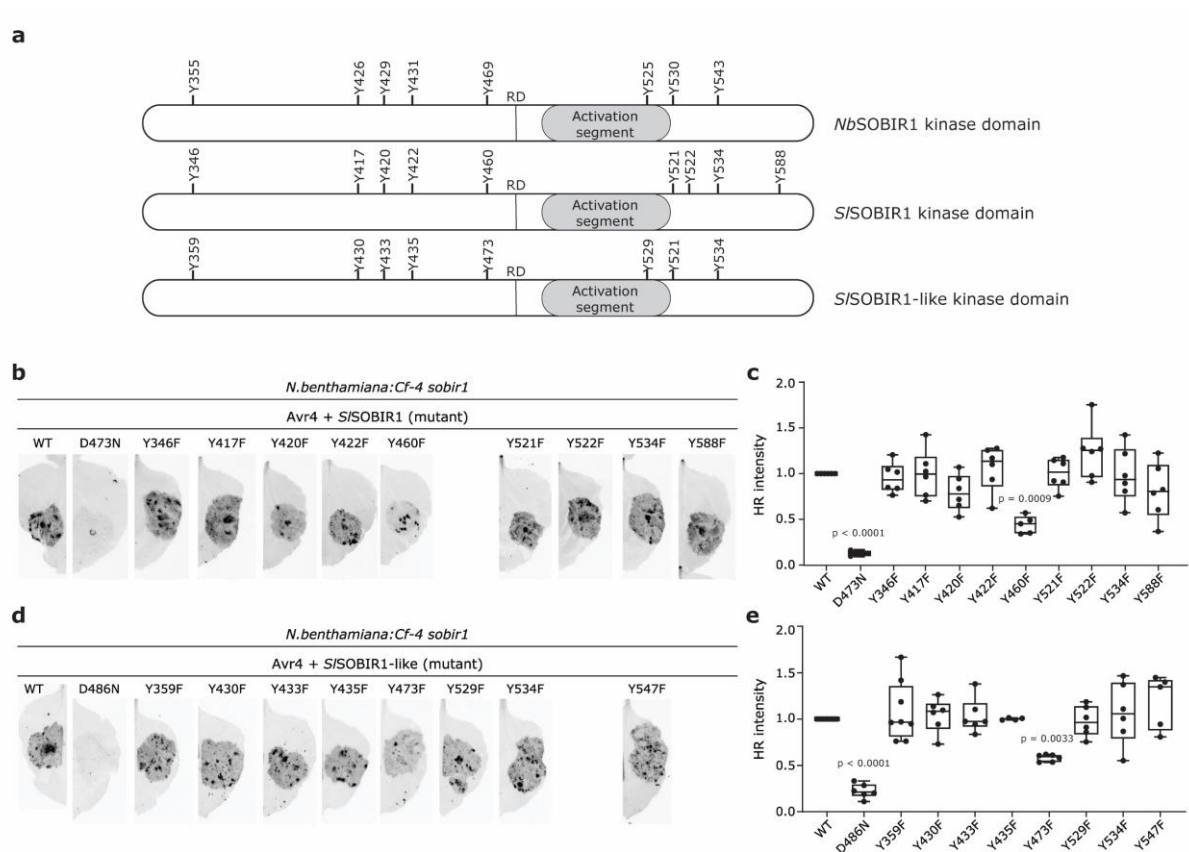
525 530 543

Nicotiana benthamiana_Niben101Scf03816g01001.1 (*NbSOBIR1*) MEAR I ADFGLAKALPDAHTHVTTSNVAGTVGY I APEYHQTLKFTGKCDI Y SFGVVLAVLVIGKLPSEFFQHTPEMS
Solanum lycopersicum_Solyc06g071810.1 (*S/SOBIR1*) MEARVADDFGLAKAVPDAHTHITTSNVAGTVGF I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKGPSDDFFQHTSEMS
Solanum lycopersicum_Solyc03g111800.2 (*S/SOBIR1*-like) MEAR I ADFGLAKAVPDAHTHITTSNVAGTIGY I APEYHQTLKFTDKCDI Y SFGVLLGVLMGKLPSEFFQHTSEMS
Arabidopsis_thaliana_AT2G31880 (*AtSOBIR1*) MEAR I SDFGLAKAMPDAVTHITTSNVAGTVGY I APEYHQTHKFTDKCDI Y SFGVILGILVIGKLPSEFFQHTDEMS
Nicotiana tabacum_A0A1S3XP18 MEAR I ADFGLAKALPDAHTHITTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKLPSEFFQHTPEMS
Nicotiana sylvestris_A0A1U7WH59 MEAR I ADFGLAKAVPDAHTHITTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKLPSEFFQHTPEMS
Nicotiana attenuata_A0A1J6INR1 MEAR I ADFGLAKALPDAHTHITTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKLPSEFFQHTPEMS
Nicotiana tabacum_A0A1S3ZD83 MEAR I ADFGLAKAVPDAHTHVTTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKLPSEFFQHTPEMS
Solanum tuberosum_M1CKT4 MEAR I ADFGLAKAVPDAHTHITTSNVAGTMGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKGPSDEFQHTSEMS
Capsicum chinense_A0A2G3BFP7 MEPR I ADFGLAKAVPDAHTHVTTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVVVGKLPSSDDFFQHTSEMS
Capsicum baccatum_A0A2G2WM84 MEAR I ADFGLAKAVPDAHTHVTTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVIGKLPSSDDFFQHTSEMS
Capsicum annuum_A0A2G2Y0L5 MEPR I ADFGLAKAVPDAHTHVTTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVVLAVLVVVGKLPSSDDFFQHTSEMS
Solanum tuberosum_M1B7X0 MEAR I ADFGLAKAVPDAHTHITTSNVAGTIGY I APEYHQTLKFTDKCDI Y SFGVLLGVLMGKLPSEFFQNTSEMS
Capsicum chinense_A0A2G3CCQ8 LEAR I ADFGLAKAVPDAHTHITTSNVAGTIGY I APEYHQTLKFTDKCDI Y SFGVLLGVLMGKLPSEFFQNTSEMS
Capsicum annuum_A0A1U8G6T0 LEGR I ADFGLAKAVPDAHTHITTSNVAGTIGY I APEYHQTLKFTDKCDI Y SFGVLLGVLMGKLPSEFFQNTSEMS
Capsicum baccatum_A0A2G2X8C5 LEPR I ADFGLAKAVPDAHTHITTSNVAGTIGY I APEYHQTLKFTDKCDI Y SFGVLLGVLMGKLPSEFFQNTSEMS
Nicotiana attenuata_A0A314KS23 MEAR I ADFGLAKAIPESLTHVSTSHVVGLGY I APAYYQTVKFTDKCDI Y SFGVLLGVLMGKFPSEDFLQFPASGMG
Coffea arabica_A0A6P6X9H5 MEAR I ADFGLAKAMPEAYTHVTSNVVGTLY I APEYHQTLKFTDKCDI Y SFGVLLASLVMGKLPSEFFQHTDEM
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Coffea canephora_A0A068V365 MEAR I ADFGLAKAMPEAYTHVTSNVVGTLY I APEYHQTLKFTDKCDI Y SFGVLLASLVMGKLPSEFFQHTDEM
Sesamum indicum_A0A6I9SRZ7 MEAR I ADFGLAKAMPDANTHVTTSNVAGTVGY I APEYHQTFKFTDKCDI Y SFGVVLGVLMGKLPSSDDFFQHTDEL
Olea europaea subsp. europaea_A0A8S0R730 MEAR I ADFGLAKAVPEANTHVSTSNVAGTAGY I APEYHQTFKFTDKCDI Y SFGVVLGALTMGKFPSEFFQHTTEEMH
Camellia sinensis var. sinensis_A0A4S4DBV0 MEAR I ADFGLAKAVPDAHTHVTTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVLLGVLVIGKLPSSDDFFQHTSEMS
Rhododendron simsii_A0A834L953 MEAR I ADFGLAKAVPDQDTHVTTSNVAGTVGY I APEYHQTMKFTDKCDI Y SFGVLLGVLVIGRLPSDNFFQDTSEMS
Vitis vinifera_F6GTQ3 MEAR I ADFGLAKAVPDANTHVTTSNVAGTVGY I APEYHQTLKFTDKCDI Y SFGVLLGVLVVVGKLPSSDDFFQHTAEMS

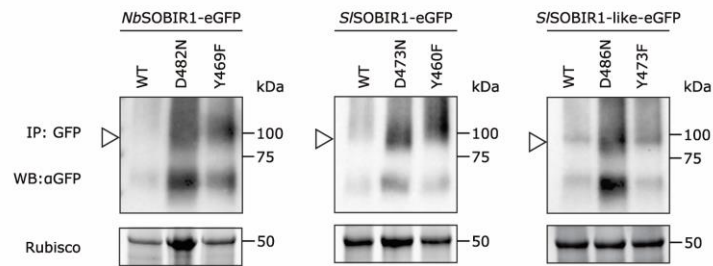
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Nicotiana benthamiana_Niben101Scf03816g01001.1 (*NbSOBIR1*) LVKWL RNVMT SEDPKRA I DSKL I GNGFEEQML LVLK I ACFCTLENPKERPNSKDVRCMLTQ I KH - - - - -
Solanum lycopersicum_Solyc06g071810.1 (*S/SOBIR1*) LVKWL RNVMT SDDPK I A I DPKL I GNGYEEQML LVLK I ACFCTLDNPKERPNSKDVRCMLTQ I KH - - - - -
Solanum lycopersicum_Solyc03g111800.2 (*S/SOBIR1*-like) LVKWMRNVMT SEDPNRA I DPKLMGNGNEEQML LVLK I ACFCTMENPKERPNSKDVRCMLMQ I KH - - - - -
Arabidopsis_thaliana_AT2G31880 (*AtSOBIR1*) L I KWMRNI I TSENPSL A I DPKLMQGGFEEQML LVLK I ACYCTLD DPKQRPNSKDVRCMLTQ I KH - - - - -
Nicotiana tabacum_A0A1S3XP18 LVKWL RNVMT SEDPKRA I DPKL I GTGFEEQML LVLK I ACFCTLENPKERPNSKDVRCMLTQ I KH - - - - -
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Capsicum chinense_A0A2G3BFP7 LVKWL RNVMT SDDPKRA I DPNL I GNGYEEQML LVLK I ACFCTMDNPKERPNSKDVRCMLTQ I KH - - - - -
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Solanum tuberosum_M1B7X0 LVKWMRNVMT SEDPNRA I DPKLMGNGNEEQML LVLK I ACFCTLENPKERPNSKDVRCMLMQ I KH - - - - -
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Coffea canephora_A0A068V365 LVLWVRNVMT SEDPKRA I DPKLLGNGYEEQML LVLK I ACFCTLENPKERPNS IDIRAML FQ I K Y EKR - - -
Sesamum indicum_A0A6I9SRZ7 LVKWMRNVMT SEDPKRA I DPKLLGNGYEEQML LVLK I ACFCTLDNPKERPNSKIDARCMML FQ I KH - - - - -
Olea europaea subsp. europaea_A0A8S0R730 LVKWMRNVMT SEDPKRA I DPRL I GNGYEEQ I L LVLK I ACFCTLDNPKERPDSKE I RCMLTQ I KH - - - - -
Camellia sinensis var. sinensis_A0A4S4DBV0 LVKWMRNVMT SEDPTRA I DPRLMGNGDHENML LVLK I ACFCTLD DPKQRPNSKDVRCML SQ I KN - - - - -
Rhododendron simsii_A0A834L953 LVKWMRNVMT SEDPNRA I DRKL I GNGHEEKML LVLK I ACFCTLEDPKQRPNSKDVRCMLDQ I TH - - - - -
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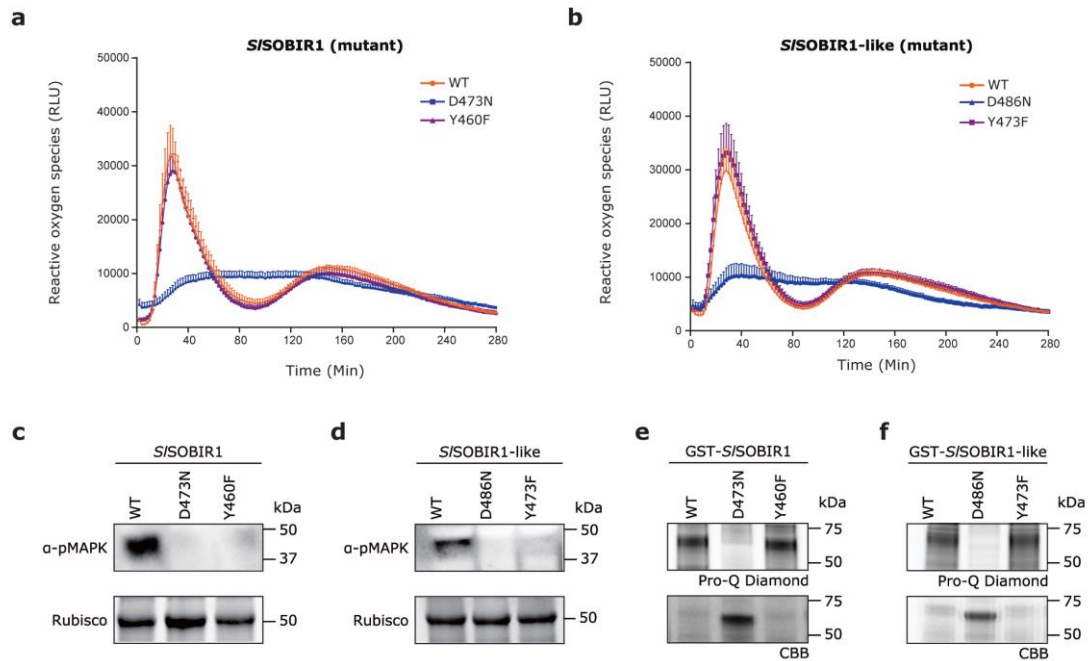
Supplementary Figure 5. Alignment of the protein sequences of the SOBIR1 kinase domain from various plant species. Amino acid sequences of the SOBIR1 homologs from various plant species were obtained from the UniProt database (<https://www.uniprot.org/>). The alignment was visualized using JalView and only the amino acid sequences of the kinase domain are shown, with all the Tyr (Y) residues highlighted in green. The Tyr residues that are subject of this study are marked with arrowheads on the top, and their position in *NbSOBIR1* is indicated on the top. The RD motif (in which a conserved arginine (R) precedes the highly conserved catalytic aspartate (D)), present in all SOBIR1 kinase domains, is indicated by a box.



Supplementary Figure 6. *S/SOBIR1* Tyr460 and *S/SOBIR1*-like Tyr473, which are the analogous residues of *NbSOBIR1* Tyr469, are essential for the Avr4/Cf-4-triggered HR in *N. benthamiana*. **a**, Schematic diagrams of the kinase domains of *NbSOBIR1*, *S/SOBIR1* and *S/SOBIR1*-like, with the location of the activation segment, the RD motif, and all Tyr residues indicated. **b-e**, Mutagenesis screen of all putative Tyr phosphorylation sites in *S/SOBIR1* (**b** and **c**) and *S/SOBIR1*-like (**d** and **e**), as shown in Figure 2 for *NbSOBIR1*, to determine their importance in immune signalling by complementation. Statistical analysis was performed with an ANOVA/Dunnett's multiple comparison test, compared with their corresponding WT. Dots indicate individual values (centre line, median; error bar, minima and maxima; n=6). Experiments were repeated at least three times, with similar results and representative pictures are shown. Source data are provided as a Source Data file.



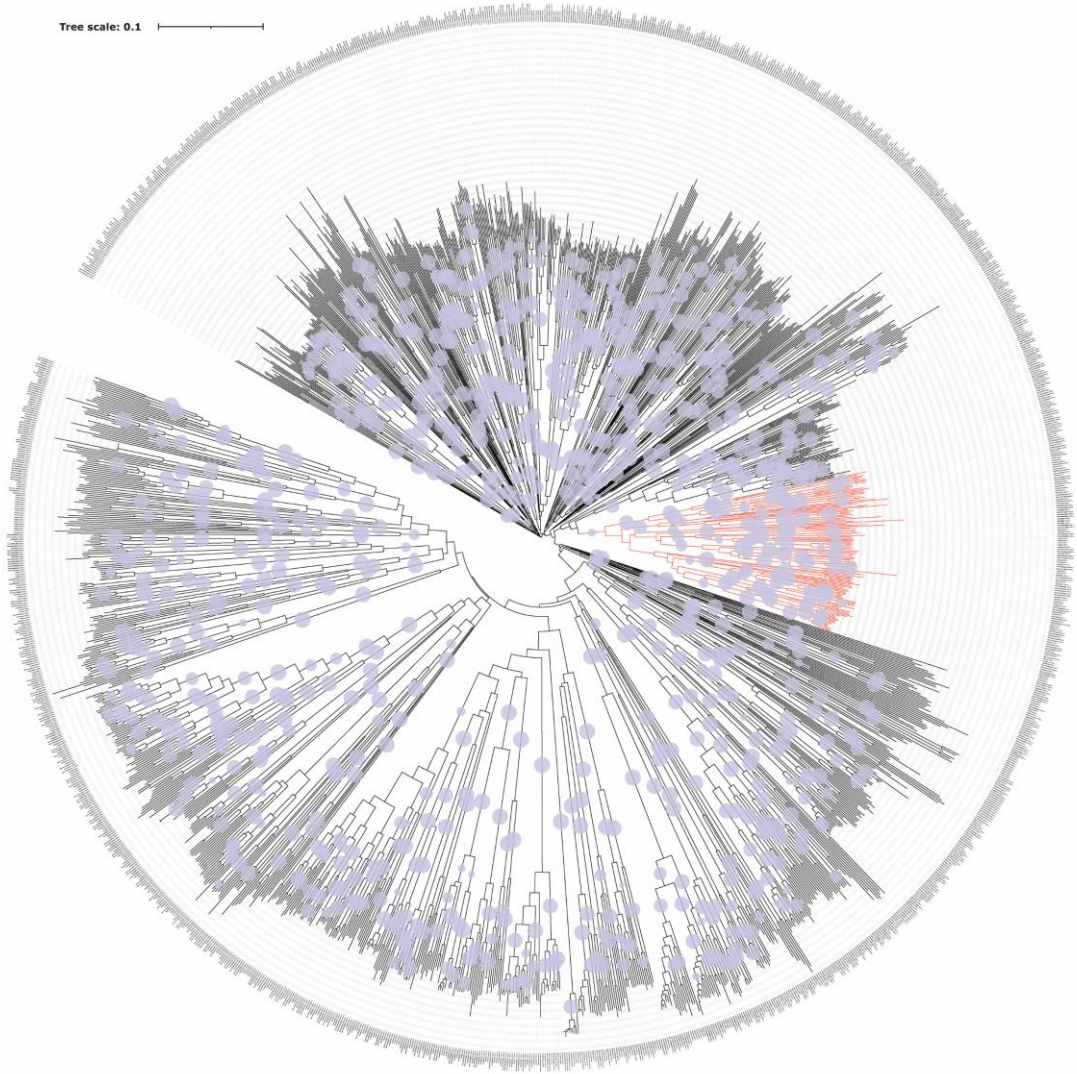
Supplementary Figure 7. Accumulation levels of *NbSOBIR1* Y469F, *S/SOBIR1* Y460F and *S/SOBIR1*-like Y473F, as well as their corresponding WT and kinase-dead (D to N) versions, *in planta*. *NbSOBIR1* Y469F, *S/SOBIR1* Y460F and *S/SOBIR1*-like Y473F, which fail to fully restore the Avr4-triggered HR in *N. benthamiana:Cf-4 sobir1* knock-out plants, were transiently expressed in *N. benthamiana:Cf-4 sobir1* knock-out plants in combination with Avr4 (both at an OD₆₀₀ of 0.8), next to their respective WT that was combined with Avr4 as a positive control, and their kinase-dead D to N version that was combined with Avr4 as a negative control. Leaf samples were collected at 2 dpi and total protein extracts were subjected to IP of the GFP-tagged SOBIR1 mutants using GFP-affinity beads, followed by WB with αGFP antibody (upper panels). The amount of total protein that was used for the IP is reflected by the Rubisco band present in the stain-free gel (lower panels). Arrowheads indicate the band representing SOBIR1-eGFP. Note that transient expression of SOBIR1 WT in combination with Avr4 triggers an HR in *N. benthamiana:Cf-4 sobir1* plants, which explains the low accumulation levels of SOBIR1 WT.

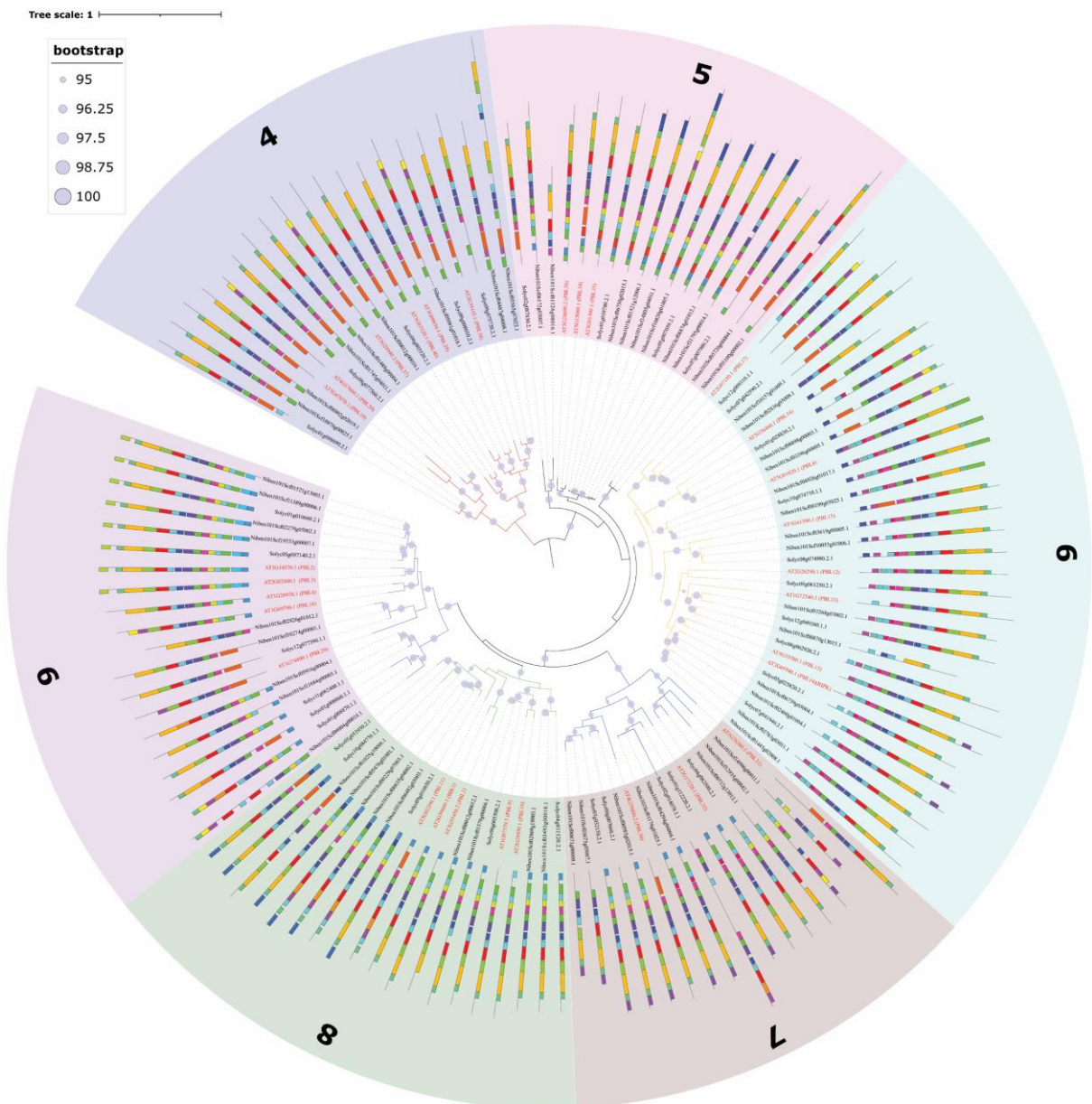


Supplementary Figure 8. *S/SOBIR1* Tyr460 and *S/SOBIR1*-like Tyr473 are required for Avr4/Cf-4-induced MAPK activation, but not for ROS accumulation and their intrinsic kinase activity. **a,b, The different Tyr mutants of *S/SOBIR1* and *S/SOBIR1*-like were transiently expressed in leaves of the *N. benthamiana*:*Cf-4 sobir1* knock-out line, with their corresponding WT as positive controls and kinase-dead mutants as negative controls. Leaf discs were taken from these plants at 24 hours after agro-infiltration, followed by adding 0.1 μ M Avr4 protein and measuring ROS accumulation over time. ROS production is expressed as RLUs, and the data are represented as mean + SEM. Similar to *SOBIR1* WT, all tested *SOBIR1* Tyr mutants restored the Avr4/Cf-4-triggered ROS production in this complementation study. Only the results for *S/SOBIR1* WT, Y460F and D473N are shown in (**a**), and results for *S/SOBIR1*-like WT, Y473F and D486N are shown in (**b**). **c, d**, *S/SOBIR1* Y460F and *S/SOBIR1*-like Y473F, as well as their corresponding WT and kinase-dead mutants, were transiently co-expressed with Avr4 in leaves of *N. benthamiana*:*Cf-4 sobir1* knock-out plants. Leaf samples were collected at 2 dpi and total protein extracts were subjected to immunoblotting with a p42/p44-erk antibody to determine the activation of downstream MAPKs by phosphorylation. **e, f**, The N-terminally GST-tagged cytoplasmic kinase domains of *S/SOBIR1* Y460F and *S/SOBIR1*-like Y473F were produced in *E. coli*, with their corresponding WT as positive controls and their kinase-dead mutants as negative controls. After SDS-PAGE of the *E. coli* lysates, the recombinant proteins were stained with Coomassie brilliant blue (lower panels), whereas the phosphorylation status of the kinase domains was determined by performing a Pro-Q Diamond stain (upper panels). Experiments were repeated at least three times and similar results were obtained. Representative pictures are shown. Source data are provided as a Source Data file.**

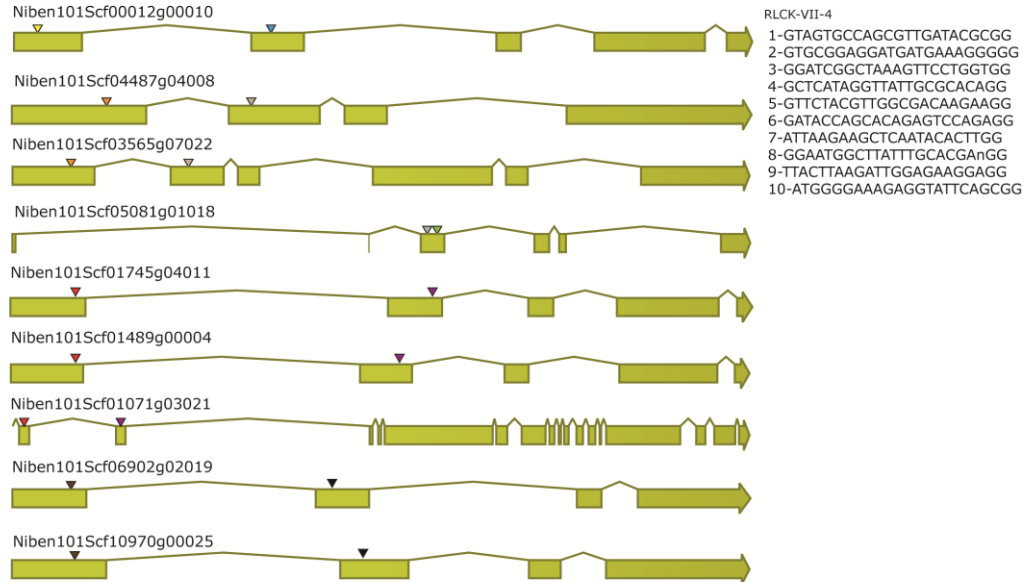
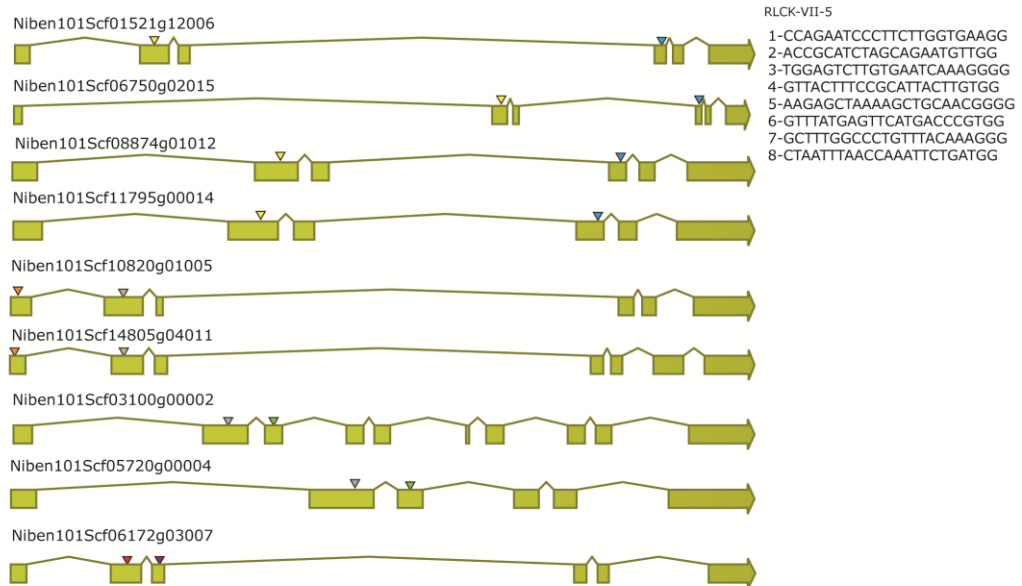
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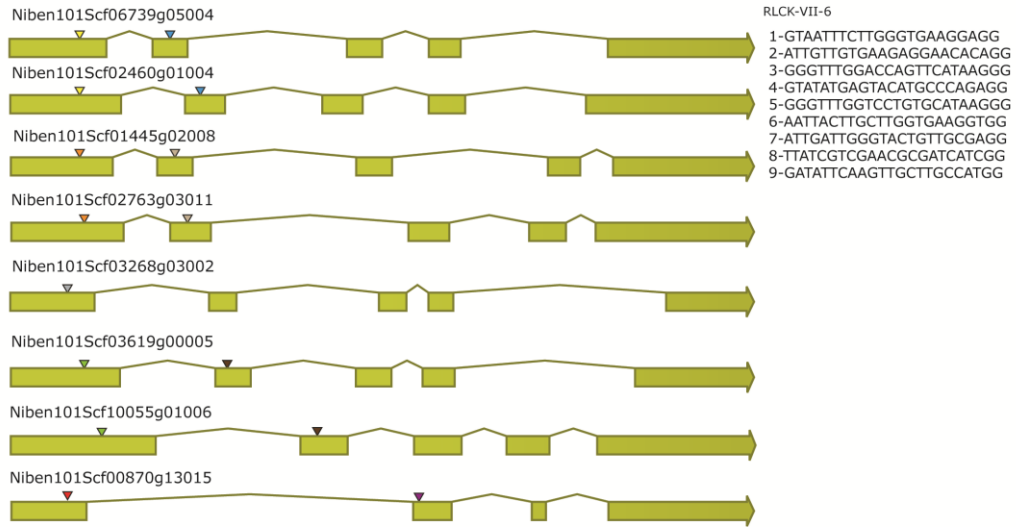


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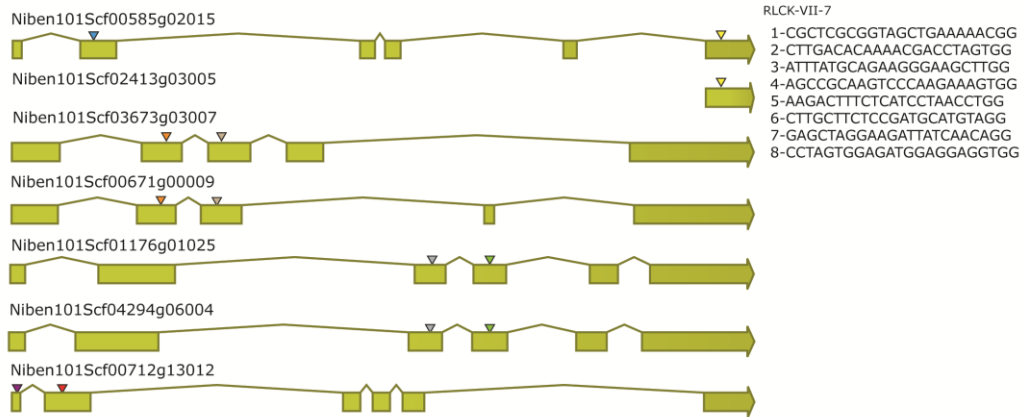
Supplementary Figure 9. Phylogenetic analysis of BIK1 homologs from Arabidopsis, tomato, and *N. benthamiana*. **a**, The amino acid sequences of only the kinase domain were extracted from all RLCK members from Arabidopsis, tomato, and *N. benthamiana* and aligned to subsequently generate a neighbour-joining phylogenetic tree, using QuickTree (Howe et al., 2002). The sub-clade of putative BIK1 homologs, which comprises 123 sequences, including *AtBIK1* (bootstrap support higher than 90%) is shown in red. **b**, Phylogenetic analysis of the RLCK-VII subfamily members from Arabidopsis (which are indicated in red), tomato, and *N. benthamiana*. Amino acid motifs identified in the complete protein sequence using MEME are shown (Bailey et al., 2009). All the members present in this tree were further assigned to six subfamilies, which are depicted in different colours. These subfamilies are referred to as subfamily 4, 5, 6, 7, 8, and 9, according to the RLCK-VII subfamilies in Arabidopsis reported previously by Rao et al. (2018).

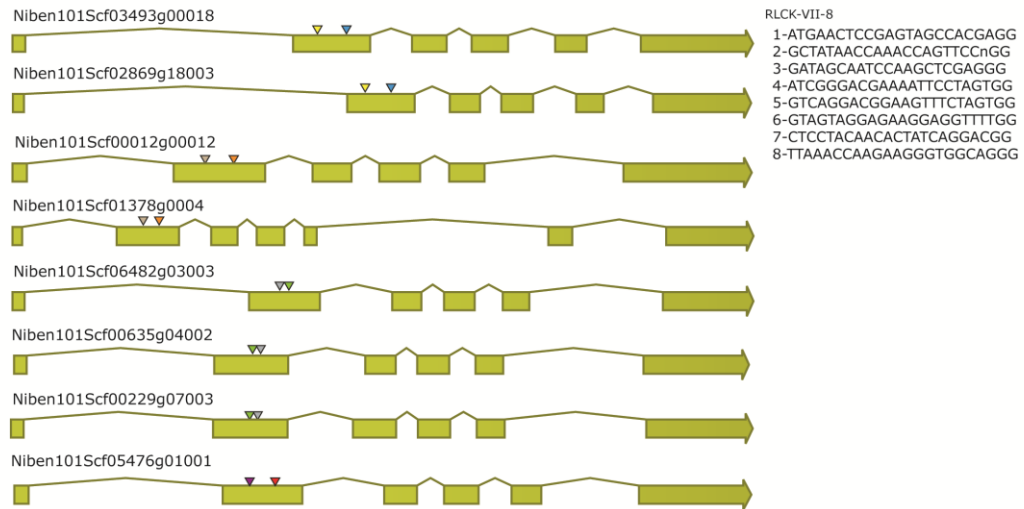
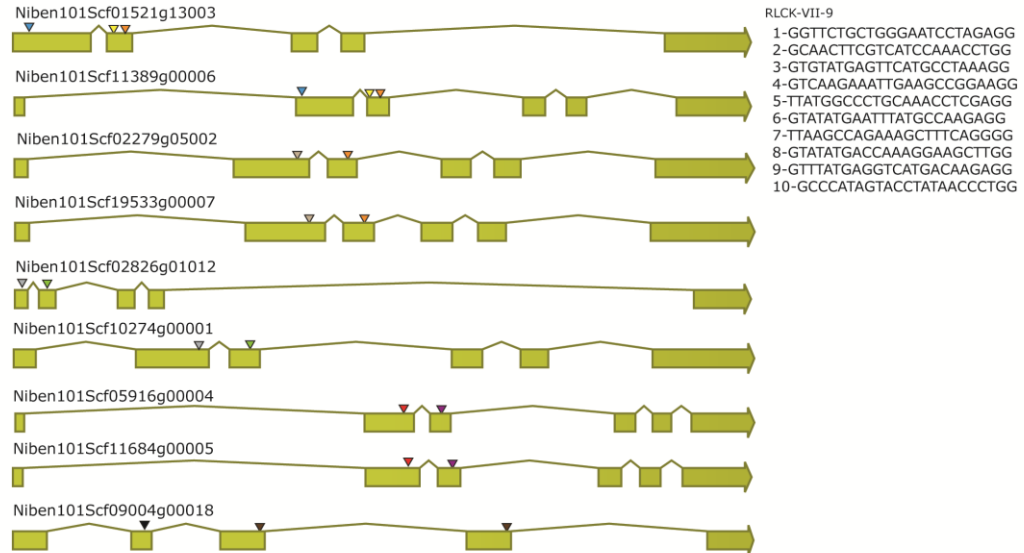
a**b****c**

d

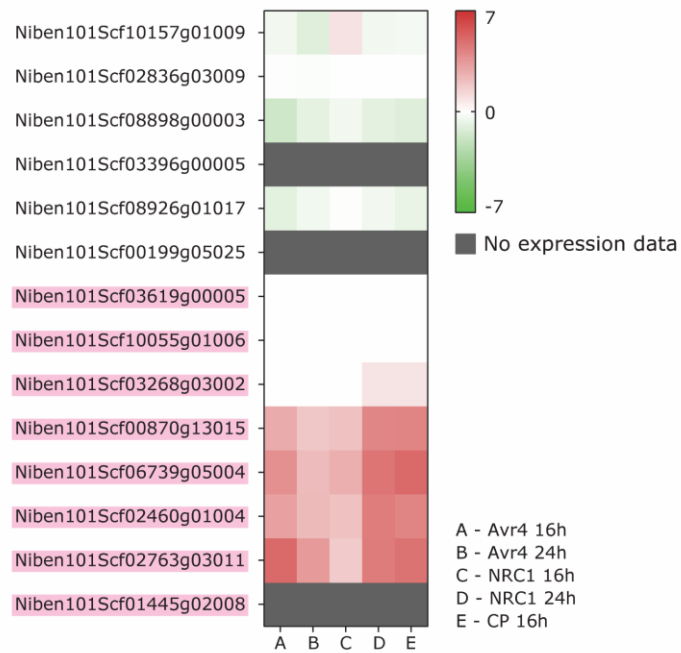


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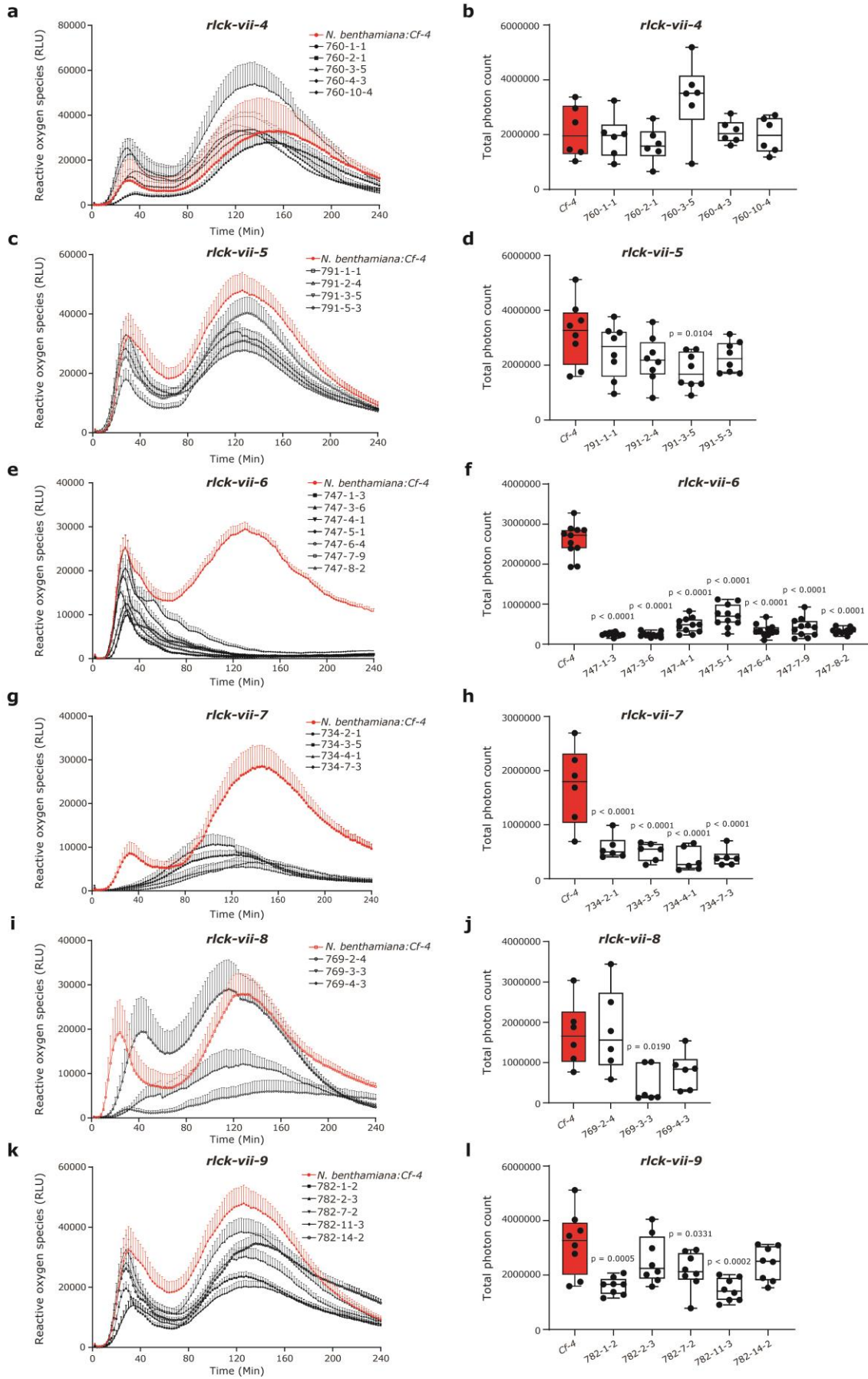


f**g****Supplementary Figure 10. Editing genes of RLCK-VII subfamilies in *N. benthamiana*:Cf-4.**

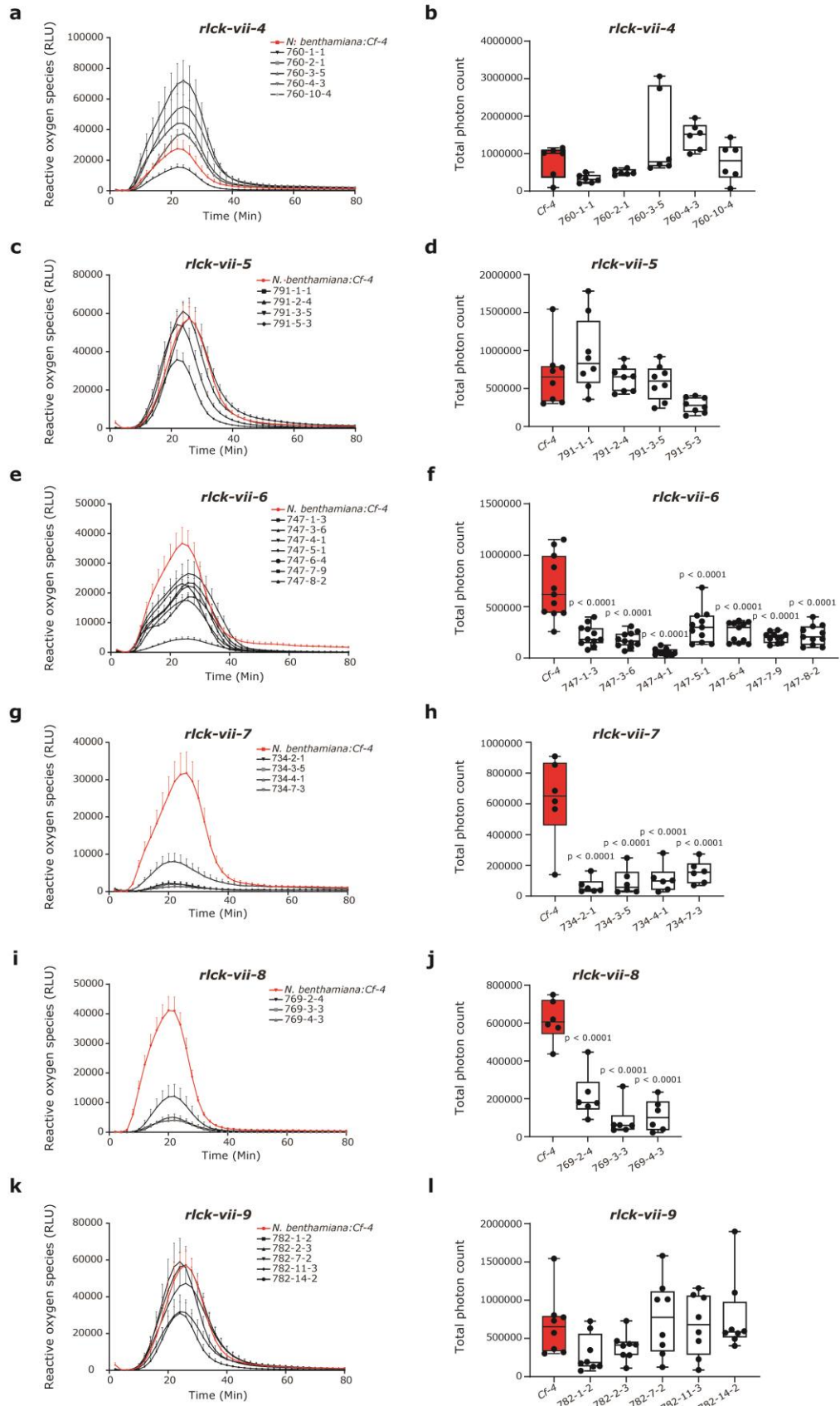
a, Scheme of the binary vector used for editing RLCK-VII genes. T-DNAs encompass the *Bar* gene (resistance to phosphinothricin; under control of the *nos* promoter), the FCY-UPP genes (p*AtUbp10* control)⁷⁸, the *Bs3* gene from pepper (*Capsicum annuum*; own promoter), the intron-optimized *zCas9i* gene (*AtRPS5a* promoter control) and up to 10 sgRNA transcriptional units driven by an *AtU6-26* promoter fragment. **b-g**, Gene models of *RLCK-VII* genes according to the solgenomics.net *N. benthamiana* genome release v1.01, with target sites indicated (coloured triangles). The colour code used for cut sites corresponds to the sgRNA array indicated in panel **a**. The target sites of the various sgRNAs are indicated for each subfamily next to the gene models.



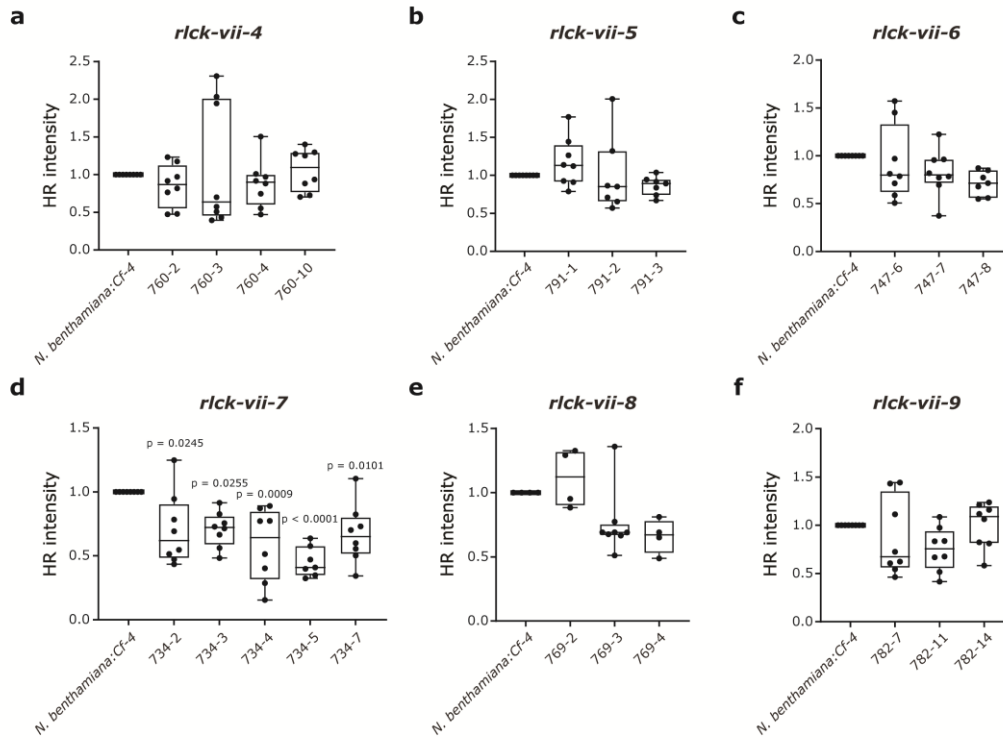
Supplementary Figure 11. Gene expression patterns of the 14 members from *N. benthamiana* RLCK-VII-6. Heat map of the relative expression (log₂) of all 14 RLCK members of subfamily 6, as determined upon transient expression of Avr4, constitutively active NRC1 (Gabiëls et al., 2007), or the coat protein (CP) of potato virus X (Tameling et al., 2010), in leaves of *N. benthamiana* plants transgenic for both *Cf-4* and *Rx*, with the latter gene mediating recognition of the CP of PVX. The RLCK-encoding genes that were selected to be knocked out are highlighted in pink. NRC1, NB-LRR PROTEIN REQUIRED FOR HR-ASSOCIATED CELL DEATH 1; CP, coat protein.



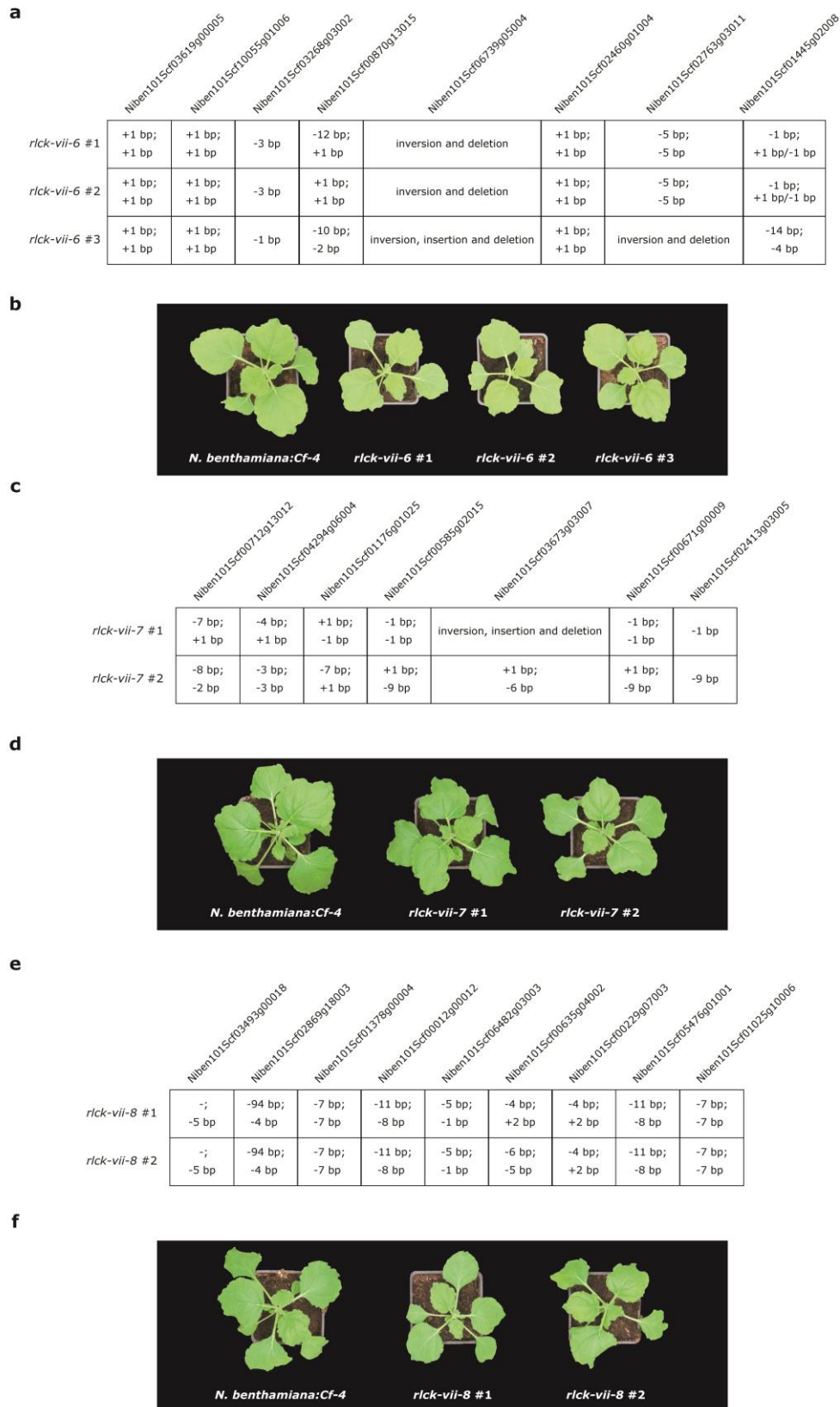
Supplementary Figure 12. Various subfamilies from RLCK-VII play an important role in the Avr4/Cf-4/SOBIR1-triggered ROS burst in *N. benthamiana*:Cf-4. Selected members from each RLCK-VII subfamily, being subfamilies 4, 5, 6, 7, 8 and 9, were targeted for knock-out in *N. benthamiana*:Cf-4 by multiplex CRISPR/Cas technology. Subsequently, ROS accumulation induced upon Avr4 protein treatment of leaf discs obtained from five individual *rlck-vii-4* knock-out transformants (**a** and **b**), four *rlck-vii-5* knock-out transformants (**c** and **d**), seven *rlck-vii-6* knock-out transformants (**e** and **f**), four *rlck-vii-7* knock-out transformants (**g** and **h**), three *rlck-vii-8* knock-out transformants (**i** and **j**) and five *rlck-vii-9* knock-out transformants (**k** and **l**), was determined. For this, leaf discs were treated with 0.1 μ M Avr4 protein, and the generation of ROS was monitored. Note that all *rlck* knock-out transformants were tested in the T1 generation. (**a**, **c**, **e**, **g**, **i**, and **k**) ROS production is expressed as RLU, and the data are represented as mean + SEM (n=8). (**b**, **d**, **f**, **h**, **j**, and **l**) Statistical analysis was performed with an ANOVA/Dunnett's multiple comparison test, compared with their corresponding WT. Dots indicate individual values (centre line, median; error bar, minima and maxima). The ROS profiles of the positive control (*N. benthamiana*:Cf-4), included in all assays, are indicated in red in all the line charts. Similar results were obtained in three replicates and data from one representative experiment are shown. Source data are provided as a Source Data file.



Supplementary Figure 13. RLCK-VII-6, -7 and -8 from *N. benthamiana* also play a positive role in the flg22/FLS2-triggered ROS burst. ROS production, triggered upon treatment with flg22, by discs taken from leaves of *rlck-vii-4* (**a** and **b**), *rlck-vii-5* (**c** and **d**), *rlck-vii-6* (**e** and **f**), *rlck-vii-7* (**g** and **h**), *rlck-vii-8* (**i** and **j**) and *rlck-vii-9* (**k** and **l**) *N. benthamiana:Cf-4* knock-out plants from the T1 generation, was measured. For this, leaf discs were taken from the different knock-out plants, as well as from *N. benthamiana:Cf-4* (the positive control), followed by treatment with a final concentration of 0.1 μ M flg22 peptide and subsequent monitoring of the accumulation of ROS. (**a**, **c**, **e**, **g**, **i**, and **k**) ROS production is expressed as RLUs, and the data are represented as mean + SEM (n=8). (**b**, **d**, **f**, **h**, **j**, and **l**) Statistical analysis was performed with an ANOVA/Dunnett's multiple comparison test, compared with their corresponding WT. Dots indicate individual values (centre line, median; error bar, minima and maxima). The ROS traces of the positive control are indicated in red in all the line charts. All experiments were repeated at least three times and data from one representative experiment are shown. Source data are provided as a Source Data file.

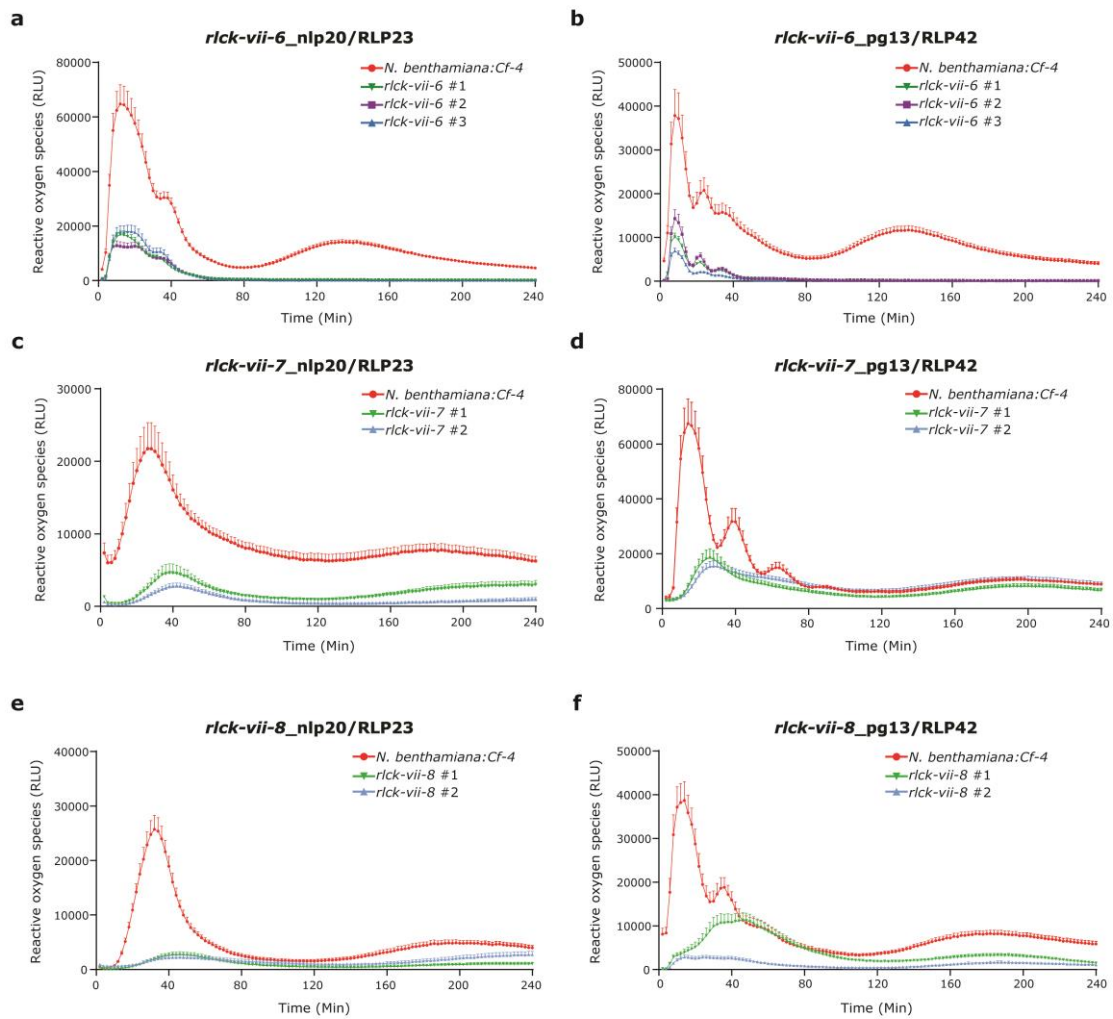


Supplementary Figure 14. RLCK-VII-7 is required for the Avr4/Cf-4-triggered HR in *N. benthamiana: Cf-4*. A solution of 5 μ M pure Avr4 protein was infiltrated in leaves of the *N. benthamiana: Cf-4* *rlck-vii-4* (a), *rlck-vii-5* (b), *rlck-vii-6* (c), *rlck-vii-7* (d), *rlck-vii-8* (e) and *rlck-vii-9* (f) transformants, and the Avr4/Cf-4-triggered HR was subsequently imaged using the ChemiDoc and quantified using Image Lab, at 2 dpi. All individual quantifications are shown as dots (n=8) and the means as lines. Statistical analysis was performed with an ANOVA/Dunnett's multiple comparison test, compared with *N. benthamiana: Cf-4*. Experiments were repeated at least three times with similar results, and representative results are shown. Source data are provided as a Source Data file.

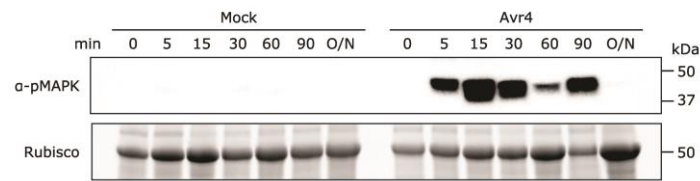


Supplementary Figure 15. Genotypes and morphological phenotypes of the different *rlck-vii* knock-out lines. Overview of the types of mutations present in all the *RLCK-VII-6* members (a),
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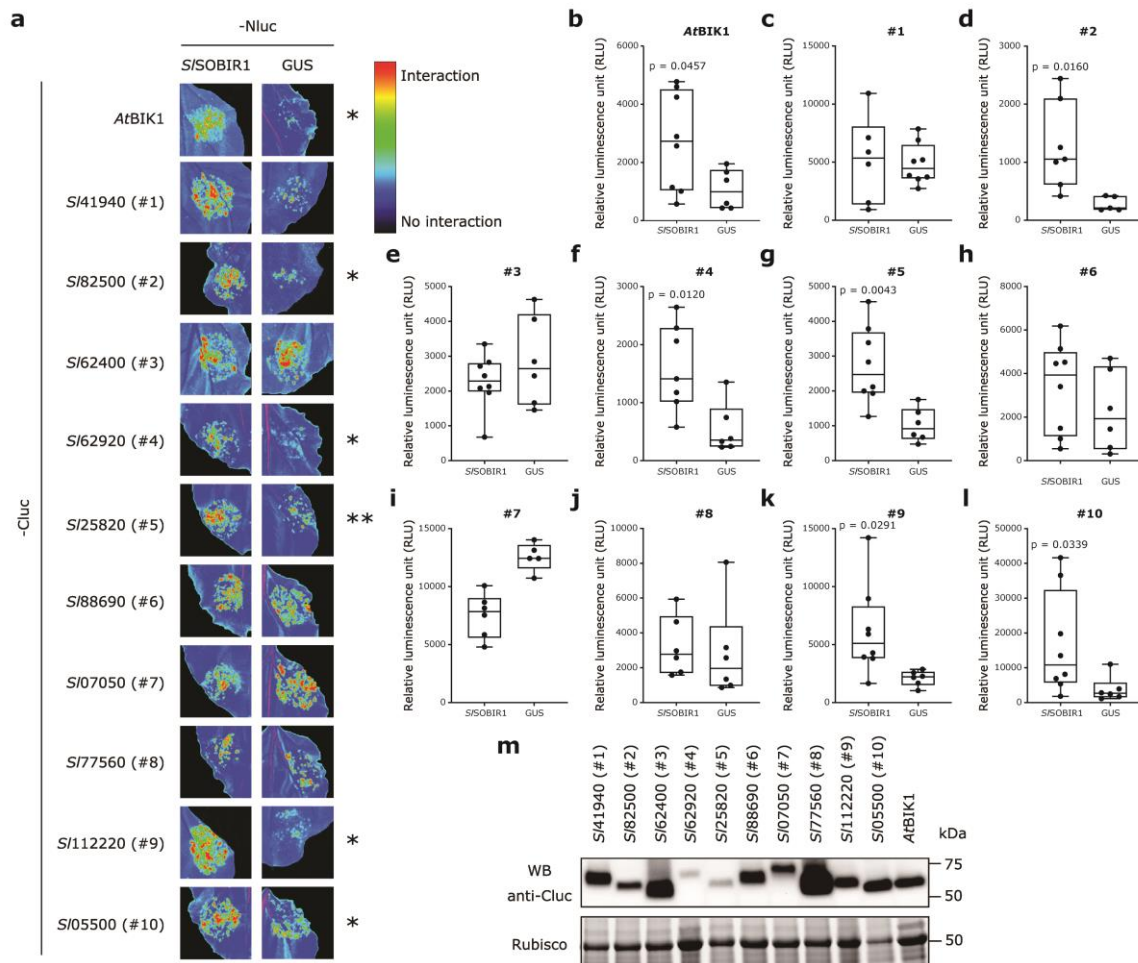
RLCK-VII-7 members (**c**), and *RLCK-VII-8* members (**e**) in the independent homozygous *N. benthamiana:Cf-4* knock-out lines. Morphological phenotypes of *N. benthamiana:Cf-4* and the three independent *rlck-vii-6* (**b**), two independent *rlck-vii-7* (**d**), and two independent *rlck-vii-8* (**f**) knock-out lines. All plants were grown in soil under the same conditions and were photographed when they were four to five weeks old.



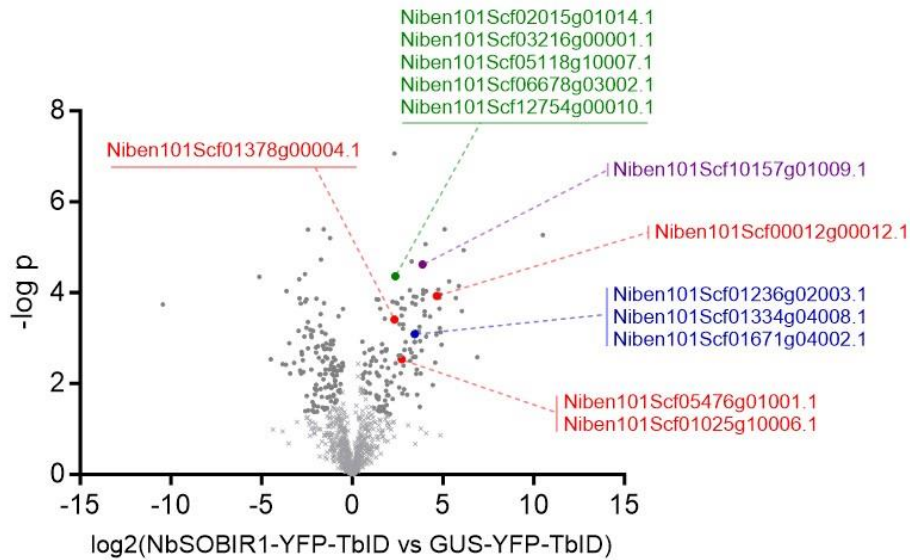
Supplementary Figure 16. Members of RLCK-VII-6, -7, and -8 differentially contribute to ROS accumulation in *N. benthamiana* induced by nlp20/RLP23 and pg13/RLP42 combinations. Leaf discs taken from independent *N. benthamiana: Cf-4 rlck-vii-6* (a and b), *rlck-vii-7* (c and d), and *rlck-vii-8* (e and f) lines, as well as from *N. benthamiana: Cf-4*, transiently expressing either RLP23 or RLP42, were treated with the corresponding elicitors at a 1 μ M concentration and the accumulation of ROS was monitored. ROS production is expressed as RLU, and the data are represented as mean + SEM (n=8). Source data are provided as a Source Data file.



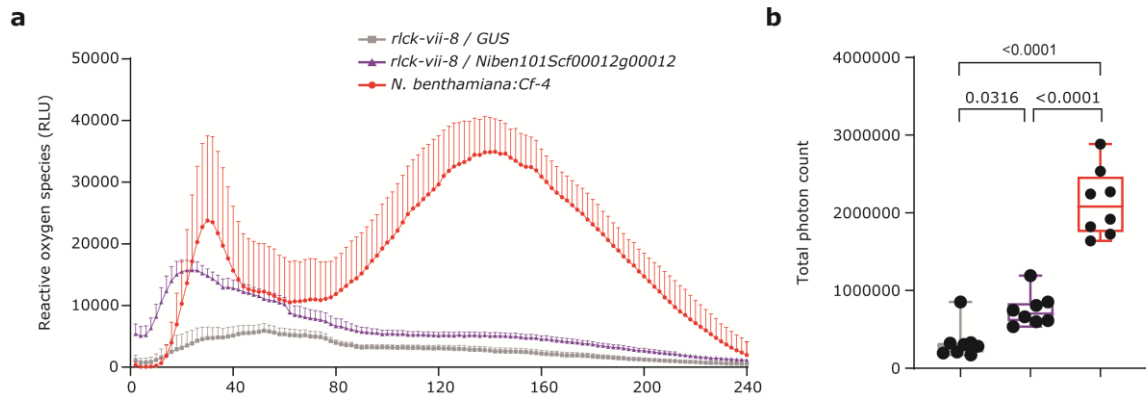
Supplementary Figure 17. The Avr4 protein triggers a swift MAPK activation in *N. benthamiana:Cf-4*. Water (mock) or 5 μ M of pure Avr4 protein was infiltrated in leaves of *N. benthamiana:Cf-4*. Leaf samples were taken at the indicated time points after Avr4 infiltration, and total protein extracts were subjected to immunoblotting using a p42/p44-erk antibody specifically detecting MAPKs that are activated by phosphorylation (α -pMAPK). Rubisco is shown as a total protein loading control. O/N, overnight. The experiment was repeated two times with similar results and a representative result is shown.



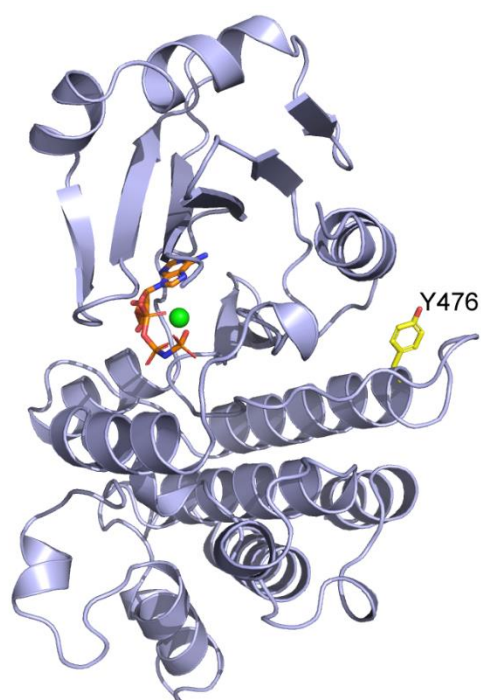
Supplementary Figure 18. A split-luciferase assay shows the interaction between several tomato RLCKs and S/SOBIR1. **a**, Cluc-tagged Arabidopsis BIK1 (*AtBIK1*) and tomato RLCKs and were transiently co-expressed with Nluc-tagged *S/SOBIR1*. Nluc-tagged GUS was included as a negative control. At 3 dpi leaves were sprayed with luciferin and luciferase activity was subsequently observed using a ChemiDoc device. Representative images of two independent experiments, each including six leaves for each interaction that was tested, are shown. Blue indicates that there is no luciferase signal, and a colouration increasing to red indicates a stronger luciferase signal, representing interaction to take place between the proteins. **b-l**, Quantification of the luminescence as shown in **a**. Dots indicate individual values (centre line, median; error bar, minima and maxima, n=6) Statistical significance was determined by Student's t-tests. **m**, Accumulation levels of the various tomato RLCKs, fused to the C-terminal part of the luciferase enzyme (Cluc). At 3 dpi an immunoprecipitation assay, followed by immunoblotting using anti-luciferase antibody (α -Cluc) was performed. *S/41940*, Solyc07G041940-Cluc; *S/82500*, Solyc04G082500-Cluc; *S/62400*, Solyc11G062400-Cluc; *S/62920*, Solyc06G062920-Cluc; *S/25820*, Solyc05G025820-Cluc; *S/88690*, Solyc01G088690-Cluc; *S/07050*, Solyc05G007050-Cluc; *S/77560*, Solyc08G077560-Cluc; *S/112220*, Solyc01G112220-Cluc; *S/05500*, Solyc06G005500-Cluc. Source data are provided as a Source Data file.



Supplementary Figure 19. Volcano plot showing the RLCKs that were identified by proximity-dependent labelling using *NbSOBIR1-YFP-TbID* as a transiently expressed bait in *N. benthamiana*. The change from grey crosses to closed circles marks the threshold for statistical significance. The RLCKs that were significantly enriched in three independent experiments in which *NbSOBIR1-YFP-TbID* (on the right) was used as a bait, when compared to *GUS-YFP-TbID* (on the left), are indicated with dots having different colours. Note that for RLCK-VII-6 (purple) and RLCK-VII-8 (red) we were able to identify individual RLCKs, with the exception of *Niben101Scf05476g01001* being detected together with *Niben101Scf01025g10006*, which is its closest paralog in the phylogeny shown in Figure S9. For RLCK-IV (green) and RLCK-VIII (blue) the determined peptide sequences did not allow the assignment of individual RLCK members.



Supplementary Figure 20. A complementation assay shows that RLCK Niben101Scf00012g00012 partially restores the ROS burst. A complementation assay with the RLCK-VII-8 family member Niben101Scf00012g00012 (*Nb00012*) was performed by transient expression, driven by the 35S promoter, in the *rlcK-vii-8* #1 knock-out line of *N. benthamiana:Cf-4*. **a**, ROS production over time upon treatment of leaf discs transiently expressing either *GUS* or *Nb00012* with Avr4, is expressed as RLUs and the data are represented as mean + SEM (n=8). *N. benthamiana:Cf-4* was taken along as a control. Note that only partial restoration of the biphasic ROS burst takes place. **b**, Total amount of ROS generated by the leaf discs, of which the profile is shown in **a**. Dots indicate individual values (centre line, median; error bar, minima and maxima, n=8) (One-way ANOVA with Tukey's test). The total photon count represents the sum of all the relative light units (RLUs) measured over a four-hour period. Letters indicate statistical groupings. Experiments were repeated three times with similar results. Source data are provided as a Source Data file.



Supplementary Figure 21. The overall structure of the inactive *AtSOBIR1* kinase domain and position of the important Tyr residues. The ribbon diagram of the SOBIR1 kinase domain is coloured in light blue. The non-hydrolysable ATP analogue AMP-PNP and Mg²⁺ are presented as an orange stick and a green sphere, respectively. *AtSOBIR1* Tyr476, which is analogous to *NbSOBIR1* Tyr469, is indicated as a yellow stick.

Supplementary Table 1. *RLCK* genes selected to be targeted by single guide (sg)RNAs.

Subfamily	Gene name	Amount of sgRNAs	Target sequence (5' - 3')	Note
4	<i>Niben101Scf10970</i> <i>g00025</i>	2	TTACTTAAGATTGGAGA AGGAGG;	
	<i>Niben101Scf06902</i> <i>g02019</i>		ATGGGGAAAGAGGTATT CAGCGG	
	<i>Niben101Scf01745</i> <i>g04011</i>	3	ATTAAGAAGCTCAATAC ACTTGG; GGAATGGCTTATTTGCA CGACGG	Not present in the phylogeny, but clustered to this subfamily according to iTAK*
	<i>Niben101Scf01489</i> <i>g00004</i>			
	<i>Niben101Scf01071</i> <i>g03021</i>			
	<i>Niben101Scf00012</i> <i>g00010</i>	2	GTAGTGCCAGCGTTGAT ACGCGG; GTGCGGAGGATGATGAA AGGGGG	
	<i>Niben101Scf03565</i> <i>g07022</i>	2	GGATCGGCTAAAGTCC TGGTGG; GCTCATAGGTTATTGCG CACAGG	
	<i>Niben101Scf04487</i> <i>g04008</i>			
<i>Niben101Scf05081</i> <i>g01018</i>		GTTCTACGTTGGCGACA AGAAGG; GATACCAGCACAGAGTC CAGAGG		
5	<i>Niben101Scf06172</i> <i>g03007</i>	2	GCTTTGGCCCTGTTTAC AAAGGG; CTAATTTAACCAAATTCT GATGG	
	<i>Niben101Scf01326</i> <i>g08016</i>			Not expressed according to QUT
	<i>Niben101Scf06750</i> <i>g02015</i>	2	CCAGAATCCCTTCTTGG TGAAGG; ACCGCATCTAGCAGAAT GTTGG	
	<i>Niben101Scf01521</i> <i>g12006</i>			
	<i>Niben101Scf08874</i> <i>g01012</i>			
	<i>Niben101Scf11795</i> <i>g00014</i>			
	<i>Niben101Scf14805</i> <i>g04011</i>	2	TGGAGTCTTGTGAATCA AAGGGG; GTTACTTTCCGCATTACT TGTGG	
	<i>Niben101Scf10820</i> <i>g01005</i>			
<i>Niben101Scf05720</i> <i>g00004</i>	2	AAGAGCTAAAAGCTGCA ACGGGG;		

	<i>Niben101Scf03100</i> <i>g00002</i>		GTTTATGAGTTCATGAC CCGTGG	
	<i>Niben101Scf10157</i> <i>g01009</i>			Not selected, see Figure S11
	<i>Niben101Scf02836</i> <i>g03009</i>			Not selected, see Figure S11
	<i>Niben101Scf08898</i> <i>g00003</i>			Not selected, see Figure S11
	<i>Niben101Scf03396</i> <i>g00005</i>			Not selected, see Figure S11
	<i>Niben101Scf08926</i> <i>g01017</i>			Not selected, see Figure S11
	<i>Niben101Scf00199</i> <i>g05025</i>			Not selected, see Figure S11
6	<i>Niben101Scf03619</i> <i>g00005</i>	2	AATTACTTGCTTGGTGA AGGTGG;	
	<i>Niben101Scf10055</i> <i>g01006</i>		ATTGATTGGGTACTGTT GCGAGG	
	<i>Niben101Scf03268</i> <i>g03002</i>	1	GGGTTTGGTCTGTGCA TAAGGG	
	<i>Niben101Scf00870</i> <i>g13015</i>	2	TTATCGTCTGAACGCGAT CATCGG; GATATTCAAGTTGCTTG CCATGG	
	<i>Niben101Scf06739</i> <i>g05004</i>	2	GTAATTTCTTGGGTGAA GGAGG;	
	<i>Niben101Scf02460</i> <i>g01004</i>		ATTGTTGTGAAGAGGAA CACAGG	
	<i>Niben101Scf02763</i> <i>g03011</i>	2	GGGTTTGGACCAGTTCA TAAGGG;	
<i>Niben101Scf01445</i> <i>g02008</i>	GTATATGAGTACATGCC CAGAGG			
	<i>Niben101Scf14996</i> <i>g00011</i>			Not expressed according to QUT
	<i>Niben101Scf12935</i> <i>g00042</i>			Not expressed according to QUT
7	<i>Niben101Scf00712</i> <i>g13012</i>	2	GAGCTAGGAAGATTATC AACAGG; CCTAGTGGAGATGGAGG AGGTGG	
	<i>Niben101Scf04294</i> <i>g06004</i>	2	AAGACTTTCTCATCCTAA CCTGG;	
	<i>Niben101Scf01176</i> <i>g01025</i>		CTTGCTTCTCCGATGCA TGTAGG	
	<i>Niben101Scf00585</i> <i>g02015</i>	2	CGCTCGCGGTAGCTGAA AAACGG; CTTGACACAAAACGACC TAGTGG	

	<i>Niben101Scf03673</i> <i>g03007</i>	2	ATTTATGCAGAAGGGAA GCTTGG;	Not present in the phylogeny, but clustered to this subfamily according to iTAK
	<i>Niben101Scf00671</i> <i>g00009</i>		AGCCGCAAGTCCCAAGA AAGTGG	
	<i>Niben101Scf02413</i> <i>g03005</i>	1	CTTGACACAAAACGACC TAGTGG	
8	<i>Niben101Scf03493</i> <i>g00018</i>	2	ATGAACTCCGAGTAGCC ACGAGG;	
	<i>Niben101Scf02869</i> <i>g18003</i>		GCTATAACCAAACCACT TCCCGG	
	<i>Niben101Scf01378</i> <i>g00004</i>	2	ATCGGGACGAAAATTCC TAGTGG;	
	<i>Niben101Scf00012</i> <i>g00012</i>		GATAGCAATCCAAGCTC GAGGG	
	<i>Niben101Scf06482</i> <i>g03003</i>	3	GTCAGGACGGAAGTTTC TAGTGG;	
	<i>Niben101Scf00635</i> <i>g04002</i>		GTAGTAGGAGAAGGAG GTTTTGG	
	<i>Niben101Scf00229</i> <i>g07003</i>			
	<i>Niben101Scf05476</i> <i>g01001</i>	2	TTAAACCAAGAAGGGTG GCAGGG;	
	<i>Niben101Scf01025</i> <i>g10006</i>		CTCCTACAACACTATCA GGACGG	
9	<i>Niben101Scf09004</i> <i>g00018</i>	2	GTTTATGAGGTCATGAC AAGAGG; GCCCATAGTACCTATAA CCCTGG	
	<i>Niben101Scf11684</i> <i>g00005</i>	2	TTAAGCCAGAAAGCTTT CAGGGG;	
	<i>Niben101Scf05916</i> <i>g00004</i>		GTATATGACCAAAGGAA GCTTGG	
	<i>Niben101Scf10274</i> <i>g00001</i>	2	TTATGGCCCTGCAAACC TCGAGG;	
	<i>Niben101Scf02826</i> <i>g01012</i>		GTATATGAATTTATGCCA AGAGG	
	<i>Niben101Scf19533</i> <i>g00007</i>	2	GTCAAGAAATTGAAGCC GGAAGG;	
	<i>Niben101Scf02279</i> <i>g05002</i>		GTGTATGAGTTCATGCC TAAAGG	
	<i>Niben101Scf11389</i> <i>g00006</i>	3	GGTTCTGCTGGGAATCC TAGAGG;	
<i>Niben101Scf01521</i> <i>g13003</i>	GCAACTTCGTCAATCCAA ACCTGG; GTGTATGAGTTCATGCC TAAAGG			

*iTAK (<http://itak.feilab.net/>); QUT (<http://www.benthgenome.qut.edu.au/>). The *RLCK* genes that are not expressed according to the QUT genome browser were not chosen to be targeted by CRISPR/Cas9. Some genes that are not present in the phylogenetic tree (Figure S9B) but were found to be clustered in a specific subfamily by using iTAK, were also selected to be knocked out.

Supplementary Table 2. Nucleotide sequences of the primers used in this study.

Primer code	Primer name	Sequence (5' - 3')
Primers used for introducing point mutations in the SOBIR1 kinase domain (the introduced mutations are indicated with a capital).		
ho113	NbSOBIR1_T512A fw*	gcactcccagatgcccatGcacatgttacgacttc
ho114	NbSOBIR1_T512A rev	gaagtcgtaacatgtgCatgggcatctgggagtg
ho115	NbSOBIR1_T515A fw	gatgccatacacatgttGcgacttcaaatgttgagg
ho116	NbSOBIR1_T515A rev	cctgcaacatttgaagtcgCaacatgtgtatgggcatc
ho117	NbSOBIR1_T516A fw	gccatacacatgttacgGcttcaaatgttgaggaaac
ho118	NbSOBIR1_T516A rev	gttcctgcaacatttgaagCcgtaacatgtgtatgggc
ho119	NbSOBIR1_S517A fw	gccatacacatgttacgactGcaaatgttgagg
ho120	NbSOBIR1_S517A rev	cctgcaacatttGagtcgtaacatgtgtatgggc
ho121	NbSOBIR1_T522A fw	cttcaaatgttgaggGctgtgggatattatgacc
ho122	NbSOBIR1_T522A rev	ggtgcaatatacccacagCtctgcaacatttgaag
ao1	SISOBIR1_T503A fw	gcagttccagatgctcatGcacatataccacttc
ao2	SISOBIR1_T503A rev	gaagtggatgatgtgCatgagcatctggaactgc
ao3	SISOBIR1_T506A fw	gctcatacacatatacGccacttcaaatgttgagg
ao4	SISOBIR1_T506A rev	ctgcaacatttgaagtgGgatgtgtatgaggc
ao5	SISOBIR1_T507A fw	caccGcttcaaatgttgaggactgttgattattg
ao6	SISOBIR1_T507A rev	caataaatccaacagttcctgcaacatttgaagCggtg
ao7	SISOBIR1_S508A fw	gctcatacacatataccactGcaaatgttgagg
ao8	SISOBIR1_S508A rev	cctgcaacatttGagtggtgatgtgtatgaggc
ao9	SISOBIR1_T513A fw	caaatgttgaggGctgttgattattg
ao10	SISOBIR1_T513A rev	gcaataaatccaacagCtctgcaacattt
ho166	SISOB-like_T516A_fw	gctgtcccagatgctcatGcacatattacaacttc
ho167	SISOB-like_T516A_rev	gaagttgtaatatgtgCatgagcatctgggacagc
ho168	SISOB-like_T519A_fw	cagatgctcatacacatattGcaacttcaaatgtggc
ho169	SISOB-like_T519A_rev	gccacatttgaagttgCaatgtgtatgagcatctg
ho170	SISOB-like_T520A_fw	gatgctcatacacatattacaGcttcaaatgtggcag
ho171	SISOB-like_T520A_rev	ctgccacatttgaagCtgaatatgtgtatgagcatc
ho172	SISOB-like_S521A_fw	ctcatacacatattacaactGcaaatgtggcaggaaac
ho173	SISOB-like_S521A_rev	gttcctgccacatttGagttgtaatatgtgtatgag
ho174	SISOB-like_T526A_fw	cttcaaatgtggcaggGctataggatacatcgctc
ho175	SISOB-like_T526A_rev	gagcgatgtatcctatagCtctgccacatttgaag
ho123	NbSOBIR1_Y355F fw	ggtggatgaggagaagtttTtagagctgagttaccggg
ho124	NbSOBIR1_Y355F rev	cccggtaactcagctctaAaaacttctccgcatccacc
ho125	NbSOBIR1_Y426F fw	ctaggccagactgccattTcttggtatatgaatatatg
ho126	NbSOBIR1_Y426F rev	catatattcatataccaagAaatggcagctggcctag
ho127	NbSOBIR1_Y429F fw	gactgccattacttggtatTtgaatatatgaaaatgg
ho128	NbSOBIR1_Y429F rev	ccatTTTTcatatattcaAataccaagtaatggcagtc
ho129	NbSOBIR1_Y431F fw	gccattacttggtatatgaatTtatgaaaaatgggagc
ho130	NbSOBIR1_Y431F rev	gctccatttttataAattcatataccaagtaatggc
ho131	NbSOBIR1_Y469F fw	gatagcttctggacttgagtTtctccatataaaccac
ho132	NbSOBIR1_Y469F rev	gtggtttatatggagaAactcaagtcagaagctatc
ho133	NbSOBIR1_Y525F fw	gcaggaaactgtgggatTtattgaccagaataaccatc
ho134	NbSOBIR1_Y525F rev	gatggattctggtgcaataAatcccacagttcctgc
ho135	NbSOBIR1_Y530F fw	gatatattgcaccagaatTccatcagacactgaag

ho136	NsSOBIR1_Y530F rev	cttcagtgtctgatggAattctgggtcaatatatc
ho137	NsSOBIR1_Y543F fw	cgggtaagtgatataTcagctttgggtgggtg
ho138	NsSOBIR1_Y543F rev	caccacaccaaagctgAatatcacacttaccgc
ao11	SISOBIR1_Y346F fw	gtggctgtggagaagttTtagagcagagctac
ao12	SISOBIR1_Y346F rev	gtagctctgtctcAaaacttctccacagccac
ao13	SISOBIR1_Y417F fw	gccagactgtcattTcttggctacgaatacatg
ao14	SISOBIR1_Y417F rev	catgtattcgtagaccaagAaatgacagtctggc
ao15	SISOBIR1_Y420F fw	cttggctTcgaatacatgaaaaatgggagtttacag
ao16	SISOBIR1_Y420F rev	ctgtaaactcccattttcatgtattcgAagaccaag
ao17	SISOBIR1_Y422F fw	cgaatTcatgaaaaatgggagtttacaggatattc
ao18	SISOBIR1_Y422F rev	ggatatcctgtaaactcccattttcatgAattcg
ao19	SISOBIR1_Y460F fw	gctgtgtgtctcaggtTtctccatataaaccatac
ao20	SISOBIR1_Y460F rev	gtaggtttatattggagaAactcgagaccagcagc
ao21	SISOBIR1_Y521F fw	gcaccagaatTttatcagacactgaagtttacag
ao22	SISOBIR1_Y521F rev	ctgtaaacttcaggtctgataaAattctgggtgc
ao23	SISOBIR1_Y522F fw	gcaccagaatattTcagacactgaagtttac
ao24	SISOBIR1_Y522F rev	gtaaacttcaggtctgaAaatattctgggtgc
ao25	SISOBIR1_Y534F fw	gataagtgatataTcagctttgggtgggtgc
ao26	SISOBIR1_Y534F rev	gcaccacaccaaagctgAatatcacacttatc
ao27	SISOBIR1_Y588F fw	gcttataggaaatggatTcgacgaacaaatgc
ao28	SISOBIR1_Y588F rev	gcattgttcgtcgAatccatttcctataagc
ho142	SISOB-like_Y359F_fw	cattgggcaaggtggatgtggaagttTtaaagctgc
ho143	SISOB-like_Y359F_rev	ctttccgtcactccaggaatgcagcttaAaaactttcc
ho144	SISOB-like_Y430F_fw	gccaagaccagactgccactTcttggctatgagtacatg
ho145	SISOB-like_Y430F_rev	catgtactcatagaccaagAagtggcagctctgtcttggc
ho146	SISOB-like_Y433F_fw	ctgccactacttggctTtgagtacatgaaaaatgggagc
ho147	SISOB-like_Y433F_rev	gtccccattttcatgtactcaAagaccaagtagtggcag
ho148	SISOB-like_Y435F_fw	gccactacttggctatgagtTcatgaaaaatgggagc
ho149	SISOB-like_Y435F_rev	gtccccattttcatgAactcatagaccaagtagtggc
ho150	SISOB-like_Y473F_fw	gctgtgtgactcaggtTtctccatataaatcatactcagcg
ho151	SISOB-like_Y473F_rev	cgctgagtatgatttatattggagaAactcgagtccagcagc
ho152	SISOB-like_Y529F_fw	ggcaggaactataggatTcatcgctccagaatatcatcag
ho153	SISOB-like_Y529F_rev	ctgatgatattctggagcagatgAatcctatagtctctgcc
ho154	SISOB-like_Y534F_fw	ggatacatcgctccagaatTcatcagacactgaagttcactg
ho155	SISOB-like_Y534F_rev	cagtgaacttcaggtctgatgaAattctggagcagatgatcc
ho156	SISOB-like_Y547F_fw	ctgataagtgatataTcagctttgggggtgctg
ho157	SISOB-like_Y547F_rev	cagcaccaccaaagctgAatatcacacttatcag
ho447	1004/5004_K110A_f2	ggctcagcctgtggctgtaGCgcttttgatttag
ho448	1004/5004_K110A_r2	ctaaatccaaaagcGCtacagccacaggctgagcc
ho451	13012_K127A-f2	gctgttGCaaaatcaaatcctgatagtaacaaggtctcaagg
ho452	13012_K127A-r2	gatttgattttGCaacagcaactggaattccaactccaactttag
ho449	1025_K119A-f2	gtaattgtgttGCgaaattgaattccgaaagcttacaggggttg
ho450	1025_K119A-r2	caatttcCGaacagcaattacagtgccgctgccactcc
ho453	00012_K109A-f2	gtgtgatcattgctgtgGCacggctgaatcaagaag
ho454	00012_K109A-r2	cttcttgattcagccgtGCcacagcaatgatcacac
ho472	6482_K114A-f2	gctgtgGCgaaattaaatcaagaaggtttgcaggggac
ho473	6482_K114A-r2	gatttaacttcGCcacagctatcacgattccagctccagg

Primers for genotyping the *rlck-vii-6* knock-out plants.

ho228	747_gen0_1-F_JS2419	ccaaaaccaaaggcaaaggag
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ho229	747_geno_1-R1004_JS2420	gattagccaggtagatagtgg
ho230	747_geno_1-R5004_JS2421	gactgaagttccaaaagcc
ho231	747_geno_2-F_JS2422	gtgatgaagattgcatggg
ho232	747_geno_2-R2008_JS2423	gtttacgagttattgcacgat
ho233	747_geno_2-R3011_JS2424	caccaactcccaaaaagcc
ho234	747_geno_3-F3002_JS2425	gtggtcattcaaggctagc
ho235	747_geno_3-R3002_JS2426	ttttctaaactccgtgccg
ho236	747_geno_4-F13015_JS2427	tcatcatcatagtaggctagcc
ho237	747_geno_4-R13015_JS2428	tagtcctttgcagcaccta
ho238	747_geno_5-F_JS2429	cgtaggtgtcattctcgga
ho239	747_geno_5-R1006_JS2430	tgtaacttctggggtacgt
ho240	747_geno_5-R0005_JS2431	tgcgtagcatcaaaaacacc
ho241	747_R1-13015_JS2486	tgttttctttacacacccc
ho242	747_F2-13015_JS2487	tgaatacatggcaaggggaa

Primers for genotyping the *rlck-vii-7* knock-out plants.

LHo07	734_00009F_JS2556	ccagttcttctggctacg
LHo08	734_03007F_JS2555	cctgttcttctgatctatg
LHo09	734_03007R+00009R_JS2557	caactaagcagaccttacc
LHo10	734_01025F_JS2558	tccattgttcagacacagtg
LHo11	734_06004F_JS2559	cattgctcagagatagtgg
LHo12	734_06004R+01025R_JS2560	gatgctcagaaagaagg
LHo13	734_02015F_JS2551	acaaaaggacccaaattgct
LHo14	734_02015R_JS2552	tggagtaagcgtcttctcat
LHo15	734_03005F_JS2553	ctacgtacgcctttacaga
LHo16	734_03005R_JS2554	cggtgattgctgtatcgat
LHo17	734_13012F_JS2561	tagcaacagcacaactgtt
LHo18	734_13012R_JS2562	tccttgagacctgttctact

Primers for genotyping the *rlck-vii-8* knock-out plants.

Po05	Scf00012	aaattagaacacggacaat
Po01	Scf00229_fw	gctgcaaagatagtagcaattgg
Po02	Scf00229_rv	gaagcacgcagatagggcg
Po03	Scf00635_fw	gcaattggctttgtatgtgc
Po04	Scf00635_rv	gcgacctataatagtccaaattgc
Po36	Scf1025_fw	gactagtaattcgaacagc
Po37	Scf1025_rv	tcctaactctgtatatgggg
Po09	Scf01378	aaattagacacgggcaac
Po08	Scf02819	tttcggttgaaccggtga
Po10	Scf03493	tttcggttgaactcatagc
Po11	Scf05476_fw	gtaattggtttcttggatgtatc
Po14	Scf05476_rv	gcgaatcaagtcgcctattcatca
Po15	Scf6482_fw	ggtgtcatgagaagtcagattgc
Po16	Scf6482_rv	ccaattttactacgaagtaccatgcg
Po06	Scf06482_fw	gcaaagatagtagtaattggctttgtatgtgc
Po07	Scf06482_rv	gcaaatgaatgtaatttgaactagtattacgtgc
Po12	Scf0012-1378_fw	atgttgctatctctggttt
Po13	Scf02869-3493_rv	gcctgtgttcatcttctaag

Primers used for generating *E. coli* expression constructs of the SOBIR1 kinase domain (sequences that are identical to the expression vectors are underlined).

ho201	pET-GST_fw	aagcttgcgccgcactcgag
ho202	pET-GST_rev	caggggcccctggaacagaacttc
ho203	NbSOBIR1_KD+90bp_fw	gttctgttccaggggcccctgcgaaagggaaagactgatggaac
ho204	NbSOBIR1_KD+90bp_rev	ctcgagtgcgccgcaagcttctaatagcttgatctgagttaacatac
ho215	SISOBIR1_KD+90bp_fw	gttctgttccaggggcccctgagaagaggggaataacgattcaag
ho216	SISOBIR1_KD+90bp_rev	ctcgagtgcgccgcaagcttctaatagcttgatctgagttaacatgc
ho217	SISOBIR1-like_KD+90bp_fw	gttctgttccaggggcccctgagagggatcagaaatgatccagg
ho218	SISOBIR1-like_KD+90bp_rev	ctcgagtgcgccgcaagcttttaatagcttgatctgcatcaacatgc
ho219	pET-15b_fw	cggatcctcgagcatatggctg
ho220	pET-15b_rev	gctgctaacaagcccgaaggaag
ho221	NbSO_KD+90bp_fw	gccatatactcgaggatccgcgaaagggaaagactgatggaac
ho222	NbSO_KD+90bp_rev	ctttcgggctttgttagcagcctaatagcttgatctgagttaacatac
ho223	SISO_KD+90bp_fw	gccatatactcgaggatccgagaagaggggaataacgattcaag
ho224	SISO_KD+90bp_rev	ctttcgggctttgttagcagcctaatagcttgatctgagttaacatgc
ho225	SISO-like_KD+90bp_fw	gccatatactcgaggatccgagagggatcagaaatgatccagg
ho226	SISO-like_KD+90bp_rev	ctttcgggctttgttagcagcctaatagcttgatctgcatcaacatgc
ho282	NbSERK3b_kd_fw	gccatatactcgaggatccgggacaactcaagaggtttccttg
ho283	NbSERK3b_kd_rev	ctttcgggctttgttagcagcctcatcttgccccgacaattcatc
ho286	SISERK3a_kd_fw	gccatatactcgaggatccgggacaactcaaaaggtttccttg
ho287	SISERK3a_kd_rev	ctttcgggctttgttagcagcctcatcttgccctgacaactcatc
ho326	1004-5004GST_fw	gttctgttccaggggcccctgatgaagattacatgggaatctctagttcc
ho327	1004GST_rv	ctcgagtgcgccgcaagcttagctctatggaaatcattgtgcaatg
ho333	5004GST_rv	ctcgagtgcgccgcaagcttagctctatggaaattattgcgcaatg
ho439	13012_GST-fw	gttctgttccaggggcccctgatgggaaattgtggtcctaatac
ho440	13012_GST-rev	ctcgagtgcgccgcaagcttgtagcttcgattgatggcagcatg
ho437	1025_GST-fw	gttctgttccaggggcccctgatgggaaattgttccagtaatgcgctc
ho438	1025_GST-rev	ctcgagtgcgccgcaagcttgtagctctcttgggagtgatag
ho441	00012_GST-fw	gttctgttccaggggcccctgatgggattgttggagtgctag
ho442	00012_GST-rev	ctcgagtgcgccgcaagcttttagcataaaagggagaagc
ho464	6482_GST_fw	gttctgttccaggggcccctgatgggattgtctgagcaacc
ho465	6482_GST_rev	ctcgagtgcgccgcaagcttctgcccgggagtgctcttc

Primers used for generating the split-luciferase binary constructs.

JRO03	KpnI-Solyc07g041940.2-fw	cggggtaccatgaagattggatgggaatc
JRO04	SalI-Solyc07g041940.2-rev	acgcgtcgacgagctcttgaaccattttg
JRO05	KpnI-Solyc04g082500.2-fw	cggggtaccatgggaaattgttgcc
JRO06	SalI-Solyc04g082500.2-rev	acgcgtcgacatagctacggtttttggagc
JRO07	KpnI-Solyc11g062400.1-fw	cggggtaccatgggattgtgctttg
JRO08	SalI-Solyc11g062400.1-rev	acgcgtcgactcttgaatgtgtcacatatctctt
JRO09	KpnI-Solyc06g062920.2-fw	cggggtaccatggctacttgccgaattg
JRO10	SalI-Solyc06g062920.2-rev	acgcgtcgacataagaatttagttgtttctttc
JRO11	KpnI-Solyc05g025820.2-fw	cggggtaccatgaagattacatgggaatctctag
JRO12	SalI-Solyc05g025820.2-rev	acgcgtcgacagctctatggaaatcattgtgc
JRO13	KpnI-Solyc01g088690.2-fw	cggggtaccatgaagtgttttttacttcaag
JRO14	XhoI-Solyc01g088690.2-rev	ccgctcgagtgtagttttacccttctgcatg
JRO15	KpnI-Solyc05g007050.2-fw	cggggtaccatgggattaggtggatg
JRO16	SalI-Solyc05g007050.2-rev	acgcgtcgacagttttccggtttggtt
JRO17	KpnI-Solyc08g077560.2-fw	cggggtaccatgctgaagtgttttatatattca
JRO27	SalI-Solyc08g077560.2-rev	acgcgtcgacagcctgagctaagg
JRO21	KpnI-Solyc01g112220.2-fw	cggggtaccatgggagttgtttcagttctaa

JRO22	SalI-Solyc01g112220.2-rev	acgcgtcgactgatgctcttttgggagtg
JRO23	KpnI-Solyc06g005500.2-fw	cgggggtaccatggggatatgtttgagtg
JRO24	SalI-Solyc06g005500.2-rev	acgcgtcgactttagcgtaaaggggagaag
Lo94	SISOBIR1-Kpn1-fw	cggcgcggtaccatgacttcgaatatccactttttctttatacg
Lo95	SISOBIR1-Sal1-NoStop-rev	gccgccgtcgacatgcttgatctgagttaacatgc
Lo153	GUS-Kpn1-fw	cggcgggggtaccatgttacgtcctgtagaaacc
Lo154	GUS-Sal1-rev	gaagccgtcgacttgttgcctcctgctgcgggttttc
Primers used for generating the TbID binary constructs.		
JO4	NsOBIR1-fw	ttctgaaggcatgtgcc
JO6	YFP-rev	ctcaggtagtggtgtcgg
JO8	TurboID-rev	gcagatacaggctattggg
JO9	GUS-fw	ggttatctctatgaactgtgcg
Primers used for generating the complementation construct of Niben101Scf00012g00012.1. Nucleotides in lower case letters align with the Gateway sequence.		
Po17	Niben101Scf00012g00012.1 fw	caccATGGGGATTTGTTTGAGTGCTAG
Po18	Niben101Scf00012g00012.1 fw	TTTAGCATAAAAGGAGAAGC

* fw, forward; rev, reverse.