

Figure S3

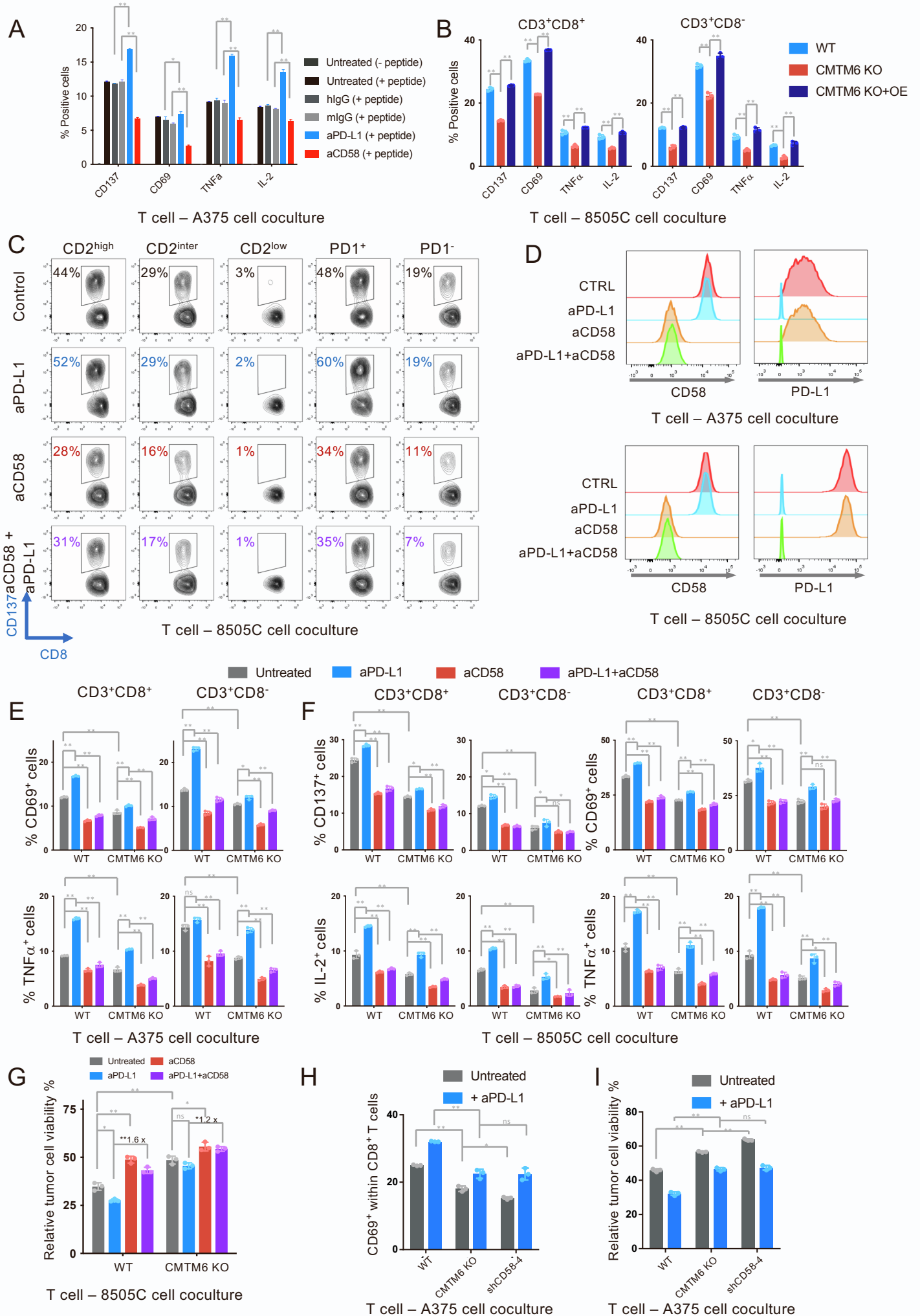


Figure S3. Influence of CMTM6, CD58, and PD-L1 on antigen-specific T cell – tumor cell interactions, Related to Figure 3

(A) Flow cytometry analysis of T cells in the T cell-tumor cell coculture system (illustrated in Figure 3A). A375 cells with (+ peptide) or without (- peptide) MART-1 peptide loading were cultured with MART-1 TCR transduced T cells. The cocultures of A375 cells with T cells were treated with blocking antibodies against CD58 (aCD58, isotype: mIgG) or PD-L1 (aPD-L1, isotype: hlgG), as indicated. Human normal IgG (hlgG) and mouse normal IgG (mIgG) served as controls. After 24 hours of coculture, expression of indicated T cell activation markers and cytokines in CD3⁺CD8⁺ cells was determined by flow cytometry. To allow detection of TNF α , and IL-2, brefeldin A was added 4 h before T cell harvesting.

(B) Effect of CMTM6 loss on T cell activation. MART-1 TCR transduced T cells were cocultured with MART-1 peptide-loaded CMTM6-wild-type, CMTM6-knockout (CMTM6 KO) or CMTM6-reconstituted (CMTM6KO+OE) 8505C cells, as indicated. After 24h of coculture, expression of indicated T cell activation markers and cytokines in CD8⁺ and CD8⁻ T cells was determined by flow cytometry. To allow detection of TNF α and IL-2, brefeldin A was added 4 h before T cell harvesting.

(C) Flow cytometry analysis of T cells that were cocultured for 18 h with 8505C cells in the absence or presence of the indicated blocking antibodies. Expression of CD137 was analyzed within CD3⁺CD8⁺ cell populations stratified by CD2 expression (CD2^{high}: top 33%, CD2^{inter}: middle 33%, CD2^{low}: bottom 33%), as well as PD-1 expression (PD-1⁺ or PD-1⁻). Representative contour plots are presented.

(D) Flow cytometry analysis of CD58 and PD-L1 staining on A375 and 8505C cells incubated with control or blocking antibodies against CD58 or PD-L1. After T cell-tumor cell coculture in the presence or absence of blocking antibodies as indicated, tumor cells were washed and stained with mixed fluorophore-conjugated anti-CD58 and anti-PD-L1 antibodies. Blocking antibodies against CD58 or PD-L1 in the coculture prevented the post-coculture staining.

(E-G) Effect of PD-L1 blockade, CD58 blockade, and CMTM6 deletion on T cell activation and tumor cell viability. MART-1 TCR transduced T cells were cocultured with MART-1 peptide-loaded tumor cells in the presence of PD-L1 blocking antibody (atezolizumab) (aPD-L1), CD58 blocking antibody (aCD58), or their combination. The coculture without antibody treatment (untreated) served as control. After 18-24h of coculture, T cell activation and tumor cell viability were analyzed.

(E-F) Flow cytometry analysis of T cells that were cocultured with CMTM6-wildtype (WT) and CMTM6-knockout (CMTM6 KO) A375 cells (E) or 8505C cells (F) in the absence or presence of the indicated blocking antibodies. Expression of CD137, CD69, IL-2, and TNF α within the CD3⁺CD8⁺ and CD3⁺CD8⁻ cell populations was depicted as indicated.

(G) Viability of CMTM6-wildtype and -knockout 8505C cells after coculture with T cells for 60 h was determined by CellTiter-Blue® Cell Viability Assay. The data presented depict relative tumor cell viability (relative to tumor cells that were cultured in the absence of T cells).

(H) Flow cytometry analysis of MART-1 TCR transduced T cells that were cocultured with MART-1 peptide-loaded CMTM6-wildtype (WT), CMTM6-knockout (CMTM6 KO), and CD58 knockdown (shCD58-4) A375 cells in the absence or presence of PD-L1 blocking antibody treatment. As Figure S1K shows, the levels of CD58 expression in CMTM6 KO and shCD58-4 A375 cells are comparable. The percentages of CD69⁺ within CD3⁺CD8⁺ cells are shown.

(I) Viability of MART-1 peptide loaded CMTM6-wildtype, CMTM6-knockout, and CD58 knockdown (shCD58-4) cells after the coculture with MART-1 TCR transduced T cells in the absence or presence of PD-L1 blocking antibody (as described in Figure S3H) was determined by CellTiter-Blue® Cell Viability Assay. The data presented depict relative tumor cell viability (relative to tumor cells that were cultured in the absence of T cells).

Data represent mean ± standard deviation of at least triplicates (A-B) (E-I) and were analyzed using two-way ANOVA (Tukey's multiple comparisons test). The statistical significance levels are denoted as follows: ns (not significant; $p \geq 0.05$), * ($p < 0.05$), and ** ($p < 0.0001$).

Figure S4

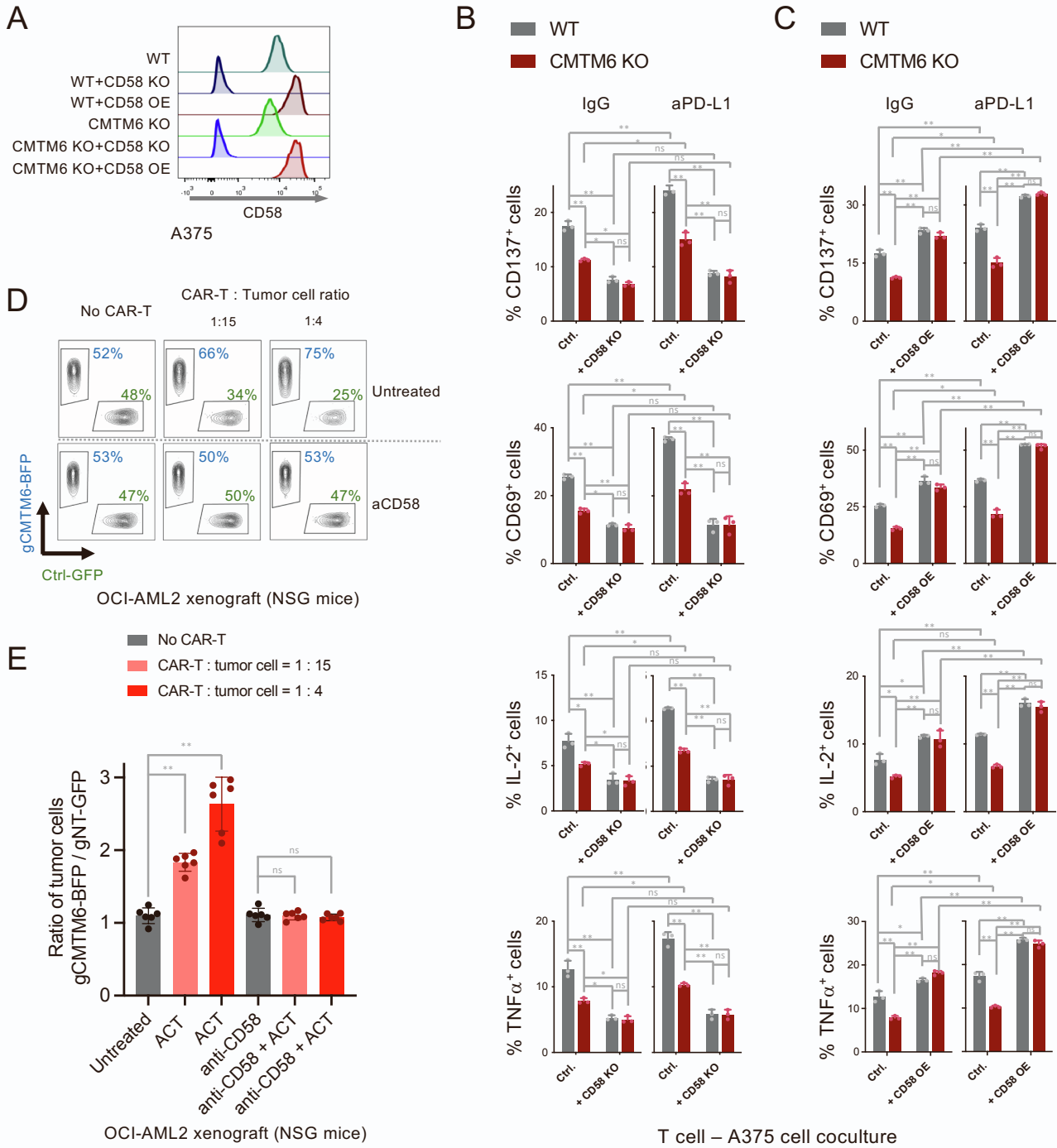


Figure S4. CMTM6 and CD58 expression in tumor cells is critically involved in antitumor T cell response, Related to Figure 3

(A) Flow cytometry analysis of CD58 expression in wildtype (WT), CD58-knockout (CD58 KO), CD58-overexpression (CD58 OE), CMTM6-knockout (CMTM6 KO), CMTM6 and CD58 double-knockout (CMTM6 KO+CD58 KO), and CMTM6-knockout plus CD58-overexpression (CMTM6 KO+CD58 OE) A375 cells.

(B-C) Effects of CD58 deletion (B) or CD58 overexpression (C), in CMTM6-proficient and -deficient tumor cells on T cell activation. MART-1 TCR transduced T cells were cocultured with A375 cells loaded with MART-1 peptide and carrying genetic modifications as indicated. The cocultures were conducted in the presence or absence of a PD-L1 blocking antibody, with normal human IgG serving as a control. Expression of CD137⁺, CD69⁺, IL-2⁺, and TNF α ⁺ cells within the CD3⁺CD8⁺ cell population was determined by flow cytometry.

(D-E) Effects of CMTM6 and CD58 on response to CAR-T treatment in an *in vivo* xenograft model of acute myeloid leukemia (AML). CMTM6-proficient and -deficient OCI-AML2 cells were individually labeled with GFP (gNT-GFP) and BFP (gCMTM6-BFP), respectively, and transplanted into NSG mice at a 1:1 ratio. The coding sequences of GFP and BFP fluorescent proteins were separately integrated into constructs containing different gRNAs: a non-targeting control gRNA for GFP and a CMTM6-targeting gRNA for BFP. Following treatment with anti-CD33 CAR-T cells at the specified CAR-T cell:tumor cell ratio for seven hours, peripheral blood was collected, and the ratio of GFP and BFP-positive AML cells was analyzed using flow cytometry. In the CD58 blockade group, an anti-CD58 antibody was administered 12 hours before the injection of AML cells.

(D) Flow cytometry analysis was performed to examine the presence of GFP (gNT) and BFP (gCMTM6)-positive AML cells remaining in the peripheral blood of the mice after CD33 CAR-T treatment, both in the presence and absence of CD58 blockade. The untreated group served as the control. Representative contour plots were generated.

(E) The ratios of BFP (gCMTM6)-positive/GFP (gNT)-positive AML cells, as described in (D), were quantified.

Data represent mean \pm standard deviation of replicates (B-C n=3; E n=6) and were analyzed using two-way (B-C) or one-way (E) ANOVA (Tukey's multiple comparisons test). The statistical significance levels are indicated as follows: ns (not significant; $p \geq 0.05$), * ($p < 0.05$), and ** ($p < 0.0001$).