## **Supporting Information**

## A Complete Workflow for High Throughput Human Single Skeletal Muscle Fiber Proteomics

Amanda Momenzadeh<sup>1,2,3</sup>, Yuming Jiang<sup>1,2,3</sup>, Simion Kreimer<sup>2,3</sup>, Laura E. Teigen<sup>4</sup>, Carlos S. Zepeda<sup>4</sup>, Ali Haghani<sup>2,3,4</sup>, Mitra Mastali<sup>2,3</sup>, Yang Song<sup>2,3</sup>, Alexandre Hutton<sup>1,2,3</sup>, Sarah J Parker<sup>2,3,5</sup>, Jennifer E. Van Eyk<sup>2,3,5</sup>, Christopher W. Sundberg<sup>4,6\*</sup>, Jesse G. Meyer<sup>1,2,3\*</sup>

<sup>1</sup> Department of Computational Biomedicine, Cedars Sinai Medical Center, Los Angeles, California 90069, USA

<sup>2</sup> Advanced Clinical Biosystems Research Institute, Cedars Sinai Medical Center, Los Angeles, California 90048, USA

<sup>3</sup> Smidt Heart Institute, Cedars Sinai Medical Center, Los Angeles, California 90048, USA

<sup>4</sup> Department of Physical Therapy, Marquette University, Milwaukee, Wisconsin 53233, USA <sup>5</sup> Department of Biomedical Sciences, Cedars Sinai Medical Center, Los Angeles, California 90048, USA

<sup>6</sup> Athletic and Human Performance Research Center, Marquette University, Milwaukee 53233, Wisconsin, USA

\* Correspondence to Christopher W. Sundberg <u>christopher.sundberg@marquette.edu</u> or Jesse G. Meyer <u>jesse.meyer@cshs.org</u>



Figure S1 Quality Control Metrics. (A) Total ion chromatogram (TIC) for a blank well. (B) TIC for a sample well. (C)Trapped Ion Mobility Spectrometry (TIMS) heatmap for a blank well. (D) TIMS heatmap for a sample well. (E) Mass spectrum (MS) for a blank well. (F) MS for a sample well. (G) Counts of non-zero precursors in blank compared to sample wells. (H) Summed quantities of precursors in blank compared to sample wells. (I) Counts of non-zero proteins in blank compared to sample wells.



**Figure S2. Ten most abundant proteins when quantities are computed by MaxLFQ.** Top ten proteins with highest percent of total counts in type 1 (left) and type 2A (right) fibers using MaxLFQ quantification. MYL11 is interchangeable with MYLPF and MLC2-fast.



**Figure S3. MYH4 peptide overlap with MYH2. (A)** Fraction of MYH subtypes (sorted by MYH2 from low to high) using protein quantities from iBAQ prior to removing MYH4 peptide with overlap with MYH2. **(B)** Histogram showing MYH fractions per fiber using protein quantities from iBAQ prior to removing MYH4 peptide with overlap with MYH2. **(C)** Precursors mapping to MYH4 with one peptide driving the abundance of MYH4. Peptide charge is given at the end of each sequence. **(D)** BLAST results for DEELDQKR<sup>2+</sup> peptide indicate identical sequence to MYH2 except for substitution of I for L.



**Figure S4. Volcano plot depicting all proteins.** Proteins above  $\log_{10}(\text{adjusted p-value})=2$  (equivalent to B-H adjusted p-value below 0.01) are statistically different between Leiden cluster 0 and 1. Ninety-four proteins were defined as significantly different according to the adjusted p-value cutoff of < 0.01 (see **Table S4** for  $\log_2 FC$  and adjusted p-values for each protein).



Figure S5. UMAP distribution of old and young fibers. UMAP colored by young and old fibers (left) in the same layout as Figure 4B (right).



**Figure S6. TPM1 and TPM3 quantities per fiber and correlation with fast and slow fibers. (A)** iBAQ-calculated quantity of TPM1 and TPM3 by fiber sorted by TPM3 values from low to high. **(B)** Linear regression plots visualizing relationships between MYH2, MYH7 and TPM1, TPM3 and corresponding Pearson correlation coefficients (r) for each plot.