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### **Supplemental information**

#### Comprehensive proteogenomic characterization

#### of rare kidney tumors

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# Figure S1. Characterization of renal cell carcinoma proteogenomic aberration landscape reveals association between survival and copy number-based genome instability, and wGII biomarkers validation, Related to Figure 1

A. Sample availability of bulk experiments included in this study.

B. ProTrack modules include Sample Dashboard, Histology Viewer, Proteomic QC, Two-gene Correlation, Interactive Heatmap and Expression Boxplots.

C. PCA plots show distribution of samples with regard to PC1 (x axis) and PC2 (y axis) in global proteome, phosphoproteome, RNA, metabolome and glyco-enriched glycoproteome, stratified by tumor/normal condition and ccRCC/non-ccRCC cohorts, colored by TMT plexes. PTM data are represented by the site-level aggregation.

D. Boxplots show protein expression between mutated and wild type groups for kinases TSC1, TSC2, MTOR, FLCN and CCND1. P-values were calculated using Wilcoxon rank sum test.

E. Scatter plot shows ploidy plotted against wGII for non-ccRCC samples. P-values were calculated for Pearson correlation coefficient.

F. PYCR1 RNA-ISH staining in 2 high wGII non-ccRCC cases and 1 low wGII non-ccRCC case.

G. IGF2BP3 IHC staining in 2 high wGII non-ccRCC cases and 1 low wGII non-ccRCC case.



### Figure S2. Delineation of tumor transcriptomic heterogeneity, immune infiltration status and tumor cell of origin by single nuclei RNAsequencing, Related to Figure 2

A. Stacked bar plot shows the cell type composition of the different samples. Non-ccRCC tumors (except AML) showed higher tumor purity and lower immune infiltration, while in comparison 3 ccRCC samples (C3L-00606-01, C3N-00148-03, C3N-00149-02; described in the companion ccRCC study) and the 2 AML showed higher levels of immune infiltration.

B. Two-dimensional UMAP visualization of snRNA-seq data for each of the 8 tumors, colored by clusters.

C. Two-dimensional UMAP visualization of snRNA-seq data for 3 ccRCC samples from the discovery cohort, colored by cell types.

D. Two-dimensional UMAP visualization of snRNA-seq data for each of the 8 tumors, colored by tumor subclusters.

E. Two-dimensional UMAP visualization of snRNA-seq data for the RO type-1 tumor, colored by cell cycle classification predicted based on expression of phase specific markers.

F. Box plots display fold change (log2 scale) of snRNA-seq expression of genes located in selected chromosomal arms between tumor subcluster and other cells of the pRCC type-1 sample (top) and the RO type2 sample (bottom)

G. Radar plot shows the probability of cell-of-origin predicted by random forest classifier for different tumor subclusters for each of the RCC subtypes using bulk RNAseq data.

H. Heatmap shows overlaps of renal epithelial cell types identified in Lake et al. (snRNA-seq) and Zhang et al. (scRNA-seq). Color scale indicates enrichment of cell markers identified from Zhang et al dataset in Lake et al. data set.

I. Heatmap shows scores of subtype-specific gene signatures derived from top 50 most upregulated transcripts for each tumor subtype.



#### Figure S3. Phosphoproteomic changes in non ccRCC tumors, Related to Figure 3

A. Boxplots show phosphorylation intensity of the indicated phosphosites stratified by tumor subtypes. Color box on the top indicates signaling pathways the phosphorylation belongs to.

B. Boxplots show different levels of phosphosites of intermediates in mTOR pathway including DEPTOR\_S265, RPTOR\_S863, EIF4EBP1\_T70 and EIF4EBP1\_S85 between different mTOR pathway genes mutation status (TSC1, TSC2, FLCN, or mTOR mutations). P-values were calculated using Wilcoxon rank sum test.

C. Volcano plot shows differentially regulated phosphorylation sites between CCND1 mutated and wild type RO samples. Known CCND1 mutation related sites are highlighted in red. RB1 phosphorylation is downstream of CCND1.

D. Heatmap shows the log2 fold change of kinases between samples with high and low wGII in ccRCC and non-ccRCC. Significantly differentially expressed kinases (p-values less than 0.05 using Limma) are labeled. Color of the dots represents the drug discovery stage of kinases.



### Figure S4. RCC glycoproteome reflects tumor immune infiltration and angiogenesis, Related to Figure 4

A. Bar plot shows distribution of glycoforms found in phospho-enriched samples.

B. Scatter plots show glycosylation change versus its corresponding protein (fold change, glyco). Right:RO. Left: pRCC.

C. Glycosylation changes versus its corresponding protein (signed p-value, glyco). Right: RO. Left: pRCC.

D. Volcano plots show differential expressed glycoproteins in RO (left) and pRCC (right). Colored by cell-type specific expression from previous single cell RNAseq data.

E. Stacked bar plots summarize differentially expressed cell type specific glycoproteins (left) and proteins (right) in phospho-enriched samples.

F. Heatmap of differentially expressed cell type specific glycoprotein markers in phospho-enriched

samples. \* denotes the marker expression difference is significant (adjusted q-value < 0.05 using Limma).

G. Protein (left) and RNA (right) expression of glycosylation enzymes in kidney tumors. The expression is converted into z-score across all samples.

H. FUT8 RNA expression across kidney cancers in combined TCGA and in house MCTP cohort.

I. Volcano plot for glycosylation changes of putative FUT8 targets in pRCC type-1. Red dots are putative FUT8 target proteins collected from previous studies.

J. MET glycosylation in glyco-enriched samples.

K. MET\_N785 glycosylation in glyco-enriched samples



#### Figure S5. Metabolomic aberrations across RCC subtypes, Related to Figure 5

A. Barplot shows the top 10 metabolic pathways with the most number of compounds identified.

B. Volcano plots show upregulated compounds in pRCC type-1 (left), AML (middle) and ROs (right), with compounds of different categories colored. Only uniquely DE compounds are labeled and marked with solid circles.

C. Metabolograms depicting select metabolic pathways to show the coordinated regulation of compounds (metabolites abundance; within green border) and the corresponding enzymes (proteins abundance; within orange border) in pRCC type-1 (outer circle), AML (middle circle) and ROs (inner circle). Color represents abundance fold change in log2 scale between respective tumor class and NATs.

D. Boxplots compare fold changes of metabolites (M), proteins (P) and mRNAs (R) across tumor subtypes within each pathway. Ordered the same as panel C.



# Figure S6. Identification and validation of proteogenomic biomarkers that distinguish papillary RCC from mucinous tubular spindle cell carcinoma (MTSCC), Related to Figure 6

A. Boxplots highlight nominated pRCC type-1 mRNA markers PIGR and SOSTDC1.

B. Boxplots present the differential mRNA abundance of PIGR and SOSTDC1 across RCC subtypes among an inhouse (MCTP) and TCGA combined cohort. Asterisks mark p-value less than 0.0001 between pRCC-1, pRCC-2 and other subtypes (upper), or between pRCC-1 and all the other subtypes (lower). P-values were calculated using Wilcoxon rank sum test.

C. Volcano plot shows significant, differentially (abs(log2fc)>0.5 and p value < 0.05) regulated phosphorylation sites between missense MET mutated (n = 2) and wild type MET (n = 5) pRCC tumors. Phosphorylation sites on potential MET substrate proteins are highlighted and labeled.

D. Violin plots show distribution of gene expression fold changes between chr7 gain and no gain samples separately derived from ccRCC and non-ccRCC subgroups. P-values were calculated using Wilcoxon rank sum test.

E. Scatter plot shows the signed log2 fold change of genes's protein expression (x axis), RNA expression (y axis) derived from DE analysis comparing chromosome 7 gain with no gain samples in ccRCC (upper panel) and non-ccRCC (lower panel) subgroups. Genes on chromosome 7 are represented with solid black circles.

F. Enrichment in chromosome 7 and chromosome 17 gene sets are tested with RNA expression difference between chromosome 7 gain samples and no gain samples in ccRCC groups. P values were from GSEA.

G. Hallmark pathway enrichment analysis based on expression fold changes between chromosome 7 gain and no gain samples stratified by ccRCC and non-ccRCC subgroups with RNA expression data. Asterisk denotes p values less than 0.05 from GSEA.

H. Kinases with enriched upregulated phosphorylation (red) and downregulated phosphorylation (blue) in Chr 7 gain samples compared to Chr7 no gain samples. Chr7 kinases are colored and labeled with black.



## Figure S7. Identification and validation of proteogenomic biomarkers that distinguish Oncocytomas (RO) from Chromophobe RCC (chRCC), Related to Figure 7

A. Boxplots show the protein abundance (top) and RNA expression (bottom) of known biomarkers of RO and chRCC across different kidney tumors.

B. PyScenic analysis identifies transcriptional modules commonly enriched in RO and chRCCs (left), and those specifically enriched in chRCCs (right).

C. Boxplots show the chRCC specific marker GPNMB (left) and RO specific biomarker ADGRF5 and MAPRE3 RNA expression in different tumor subtypes.

D. Boxplots show the differential protein abundance (top) and RNA expression (bottom) (between RO type-1 from RO type-2) of THSD4 across different kidney tumors. P-values were calculated using Wilcoxon rank sum test.