

Trait-specific sensitive developmental windows: wing growth best integrates weather conditions encountered throughout the development of nestling Alpine swifts

Supporting Information and codes for the manuscript

Last compiled on 20 May 2024

Contents

Authors' list and affiliations	2
ORCIDs	2
Abstract and keywords	2
Libraries and datasets	3
Libraries	3
Reproducible environment	3
The datasets	4
Growth curves	7
Supporting Information 01 - Climwin analyses	11
Weather data	11
PCA results	12
The climwin approach	13
Base model	13
Finding the best window	14
Investigating the models	15
Wing length at 50 days	15
Sternum length at 50 days	23
Body mass at 50 days	31
Adding the variables to the dataset	41
Figure	42
Testing for segments	43
Sternum length	45
Body mass	49

Supporting Information 02: Growth rates	59
Checking the resulting growth rates	61
Growth rate models	62
Wing length	62
Sternum length	63
Figure	66
Supporting Information 03: Correlation between the size and growth	66
Figure	66
Pearson’s correlation tests	68
Supporting Information 04: Short-term consequences	69
Probability of fledging	69
Age at fledging	72
Figure	75
Session information	76

Authors’ list and affiliations

Giulia Masoero^{1,2} *, Michela Natalina Dumas², Julien G.A. Martin², Pierre Bize¹ *

¹ Swiss Ornithological Institute, Seerose 1, 6204 Sempach, Switzerland

² Department of Biology, University of Ottawa, Canada

* **Corresponding authors:** giulia.masoero@gmail.com, pierre.bize@vogelwarte.ch

ORCIDs

GM: 0000-0003-4429-7726

MND: 0000-0003-2948-8751

JGAM: 0000-0001-7726-6809

PB: 0000-0002-6759-4371

Abstract and keywords

Abstract: The size and growth patterns of nestling birds are key determinants of their survival up to fledging and long-term fitness. However, because traits such as feathers, skeleton and body mass can follow different developmental trajectories, our understanding of the impact of adverse weather on development requires insights on trait-specific sensitive developmental windows. We analysed data from nestling Alpine swifts in Switzerland measured throughout growth up to the age of 50 days (i.e. fledging between 50 and 70 days), for wing length and body mass (2693 nestlings in 25 years) and sternum length (2447 nestlings in 22 years). We show that the sensitive developmental windows for wing and sternum length corresponded to the periods of trait-specific peak growth, which span almost the whole developmental period for wings and

the first half for the sternum. Adverse weather conditions during these periods slowed down growth and reduced size. Although nestling body mass at 50 days showed the greatest inter-individual variation, this was explained by weather in the two days before measurement rather than during peak growth. Interestingly, the relationship between temperature and body mass was not linear, and the initial sharp increase in body mass associated with the increase in temperature was followed by a moderate drop on hot days, likely linked to heat stress. Nestlings experiencing adverse weather conditions during wing growth had lower survival rates up to fledging and fledged at later ages, presumably to compensate for slower wing growth. Overall, our results suggest that measures of feather growth and, to some extent, skeletal growth best capture the consequences of adverse weather conditions throughout the whole development of offspring, while body mass better reflects the short, instantaneous effects of weather conditions on their body reserves (i.e. energy depletion vs storage in unfavourable vs favourable conditions).

Keywords: Apodiformes, feathers, meteorological conditions, climate change, multi-trait, heat stress, offspring development, early-life conditions, *Tachymarptis (Apus) melba*

Journal: Ecology and Evolution

Full script, data and Supporting Information can also be found on the OSF page of the project: <https://osf.io/2ndmk/>

Libraries and datasets

Libraries

```
# data manipulation
library(dplyr)
library(lubridate)
# models
library(lme4)
library(lmerTest)
library(climwin)
library(segmented)
library(MuMIn)
# graphics
library(ggplot2)
library(ggtext)
library(scales)
# tables
library(sjPlot)
# others
library(performance)
# renv::update(prompt = FALSE) # use to update the packages within the project
# renv::snapshot(prompt = FALSE) # to update the metadata in the renv.lock about the used packages

knitr::opts_chunk$set(
  fig.path = "cache/",
  cache.path = "cache/",
  cache.extra = getRversion()
)
```

Reproducible environment

This project has been initialised using the package *renv*.

```
citation("renv")
```

```
## To cite package 'renv' in publications use:
##
##   Ushey K, Wickham H (2024). _renv: Project Environments_. R package version 1.0.7,
##   <https://CRAN.R-project.org/package=renv>.
##
## A BibTeX entry for LaTeX users is
##
##   @Manual{,
##     title = {renv: Project Environments},
##     author = {Kevin Ushey and Hadley Wickham},
##     year = {2024},
##     note = {R package version 1.0.7},
##     url = {https://CRAN.R-project.org/package=renv},
##   }
```

This package allows to upload the exact same version of each of the packages used in this project. This is done using the files in the compressed folder “renv.zip”. The file has to be decompressed to obtain “renv.lock” file, the “.Rprofile” file and the folder “renv”. The packages should then be obtained by running the function `renv::restore()` in the R console.

For more information <https://rstudio.github.io/renv/articles/renv.html>

The datasets

Study system

Data were collected between 1999 and 2023 in a Swiss population of Alpine swift. It is a long-distance migratory bird that breeds in colonies of a few to several hundred pairs in holes within cliffs or under the roofs of tall buildings. In Switzerland, Alpine swifts return to their breeding grounds from sub-Saharan Africa at the beginning of April (Meier et al. 2020) and start laying eggs between early May and June, with significant adaptive variations in laying dates depending on weather conditions (de Villemereuil et al. 2020). Females lay one clutch a year, with one to four eggs per clutch (modal clutch size is three). Both parents incubate the eggs for about 18 days and then feed their nestlings until fledging, which occurs around 55 days after hatching (range 50-76 days; (Bize et al. 2004) and this manuscript). After breeding, Alpine swifts migrate back to Africa in September (Meier et al. 2020).

Fieldwork was carried out in two Alpine swift colonies located in clock towers in the Swiss cities of Biel (60-100 breeding pairs) and Solothurn (40-55 breeding pairs), ca. 20km apart. Each year, both colonies were regularly visited to monitor egg laying and clutch size, to capture and measure adults, and to ring and measure nestlings. Nestlings were individually recognised by ringing them with numbered metal rings 10 to 15 days after hatching. Nestlings were measured regularly (usually every 5 to 10 days, on average five times in total) until fledging. At each measurement, we measured wing length with a ruler to the nearest 1 mm, sternum size with a calliper to the nearest 0.1 mm, and body mass with a digital scale to the nearest 0.1 g. The measure of sternum length provides an estimate of skeletal growth and size. Tarsus length has been commonly used in passerines, but it is difficult to measure in a repeatable way in a species with short and bulky tarsi, such as swifts. As nestlings are not ringed at hatching, the age of the nestlings in a brood is based on the hatching date of the first nestling; the last nestling is usually born on the same day or 1 day later. Therefore, measurements for a brood of three nestlings, for example, are taken when the first-hatched nestling reaches 50 days of age, the youngest one might be the same age or 1 day younger. Only nestlings that survived up to fledging were included in the statistical analyses. Sample sizes differ between traits, as wing length and body mass have been measured since 1999, while sternum length has been measured since 2003.

```

# parameters for colors
par_col_wing_50 <- "#2D708EFF"
par_col_wing_growth <- "#2D708EFF"
par_col_sternum_50 <- "#440154FF"
par_col_sternum_growth <- "#440154FF"
par_col_mass_50 <- "#56C667FF"

# functions from another script
source("script_functions.R")

# loading the datasets
data_clim <- read.csv("data/data_clim.csv",
  stringsAsFactors = TRUE, na = c("", "NA"))
)
data_am_nestlings <- read.csv("data/data_nestlings.csv",
  stringsAsFactors = TRUE, na = c("", "NA"))
)
data_am_nestlings_all <- read.csv("data/data_nestlings-all-meas.csv",
  stringsAsFactors = TRUE, na = c("", "NA"))
)
data_am_nests <- read.csv("data/data_nests.csv",
  stringsAsFactors = TRUE, na = c("", "NA"))
)

```

```

# creating the metadata
data_clim_meta <- data.frame(
  column_name = colnames(data_clim),
  description = c(
    "year of weather recording",
    "date of weather recording",
    "daily mean ambient temperatures (°C) averaged using data from five stations obtained from Swiss meteorological data",
    "daily total rainfall (degrees Celsius) averaged using data from five stations obtained from Swiss meteorological data",
    "daily mean wind (degrees Celsius) averaged using data from five stations obtained from Swiss meteorological data"
  )
)
write.csv(data_clim_meta,
  file = "data/data_clim_metadata.csv",
  row.names = FALSE
)

data_am_nestlings_meta <- data.frame(
  column_name = colnames(data_am_nestlings),
  description = c(
    "year of measurement recording",
    "colony where the nest was located",
    "code of the nest, equivalent to broodID",
    "individual ID of the nestling",
    "date of hatching of the first nestling in the brood",
    "date of measurement of the nestling",
    "age in days of the nestling at the time of measurement",
    "hour of measurement of the nestling",
    "mass at day 50",
    "wing length at day 50",
  )
)

```

```

    "sternum length at day 50",
    "age at fledging in days since hatching",
    "number of days from the 1st of May of hatching of the first nestling in the brood",
    "brood size (number of hatchlings) of the nest of the focal nestling"
  )
)
write.csv(data_am_nestlings_meta,
  file = "data/data_nestlings_metadata.csv",
  row.names = FALSE
)

data_am_nestlings_all_meta <- data.frame(
  column_name = colnames(data_am_nestlings_all),
  description = c(
    "individual ID of the nestling",
    "wing length at the time of measurement",
    "sternum length at the time of measurement",
    "mass at the time of measurement",
    "age in days of the nestling at the time of measurement"
  )
)
write.csv(data_am_nestlings_all_meta,
  file = "data/data_nestlings_all-meas_metadata.csv",
  row.names = FALSE
)

data_am_nests_meta <- data.frame(
  column_name = colnames(data_am_nests),
  description = c(
    "colony where the nest was located",
    "year of measurement recording",
    "date of hatching of the first nestling in the brood",
    "number of nestlings in the brood that did not fledge",
    "number of nestlings in the brood that fledged",
    "brood size of the nest (number of hatchlings)"
  )
)
write.csv(data_am_nests_meta,
  file = "data/data_am_nests_metadata.csv",
  row.names = FALSE
)

```

```
# handling of date variables
```

```

data_am_nestlings$date_meas <- as.Date(data_am_nestlings$date_meas)
data_am_nestlings$date_hatch <- as.Date(data_am_nestlings$date_hatch)
data_am_nestlings$date_50days <- data_am_nestlings$date_hatch + 50
data_am_nestlings$hatch_doy <- yday(data_am_nestlings$date_hatch)
data_clim$date <- as.Date(data_clim$dateYMD, "%d/%m/%y")

```

```
# scaling variables for the analyses
```

```

data_am_nestlings$day_hatch_1may_sc <-
  fun_scale(data_am_nestlings$day_hatch_1may)

```

```

data_am_nestlings$hatch_doy_sc <- fun_scale(data_am_nestlings$hatch_doy)
# age in days scaled with 50
data_am_nestlings$age_days_sc50 <- data_am_nestlings$age_days - 50

# year of measurement as factor
data_am_nestlings$year_f <- as.factor(data_am_nestlings$year)
data_am_nestlings$ring <- as.factor(data_am_nestlings$ring)

```

Creating the subsets needed for the analyses

```

data_am_nestlings_wing_50 <- subset(data_am_nestlings, !is.na(wing_50))
data_am_nestlings_sternum_50 <- subset(data_am_nestlings, !is.na(sternum_50))
data_am_nestlings_mass_50 <- subset(
  data_am_nestlings,
  !is.na(mass_50) & !is.na(day_hatch_1may)
)
# To include only the reproductive season: subset clim to 1st of May -> 10th of September
data_clim_sub <- subset(
  data_clim,
  as.numeric(strftime(data_clim$date, "%m")) %in% 5:8 |
  as.numeric(strftime(data_clim$date, "%m")) == 9 &
  as.numeric(strftime(data_clim$date, "%d")) <= 10
)

nestlings <- subset(data_am_nestlings_all, ring %in% data_am_nestlings$ring)

nestlings_n <- nestlings %>%
  group_by(ring) %>%
  mutate(n_measures = sum(!is.na(unique(mass)))) %>%
  slice(which.max(age_days))

```

Growth curves

Developmental trajectories, from hatching to fledging, of nestling Alpine swifts. Dots represent individual measurements of wing and sternum length and body mass of nestlings measured between 1999 and 2023, and smooth lines represent the average growth pattern of nestlings. The sensitive developmental windows during which weather conditions most affected the phenotype at 50 days (see results below) are highlighted with a purple background.

```

cowplot::plot_grid(
  fun_plot_growth(data_am_nestlings_all, "wing"),
  fun_plot_growth(data_am_nestlings_all, "sternum"),
  fun_plot_growth(data_am_nestlings_all, "mass") +
  xlab("Age (days)"),
  nrow = 3, align = "hv"
)

```

```

# Save the plot to the figures directory
ggsave("figures/Fig. 1 - Growth curves.png", width = 7, height = 5, dpi = 300)

```

Sample sizes for this figure:

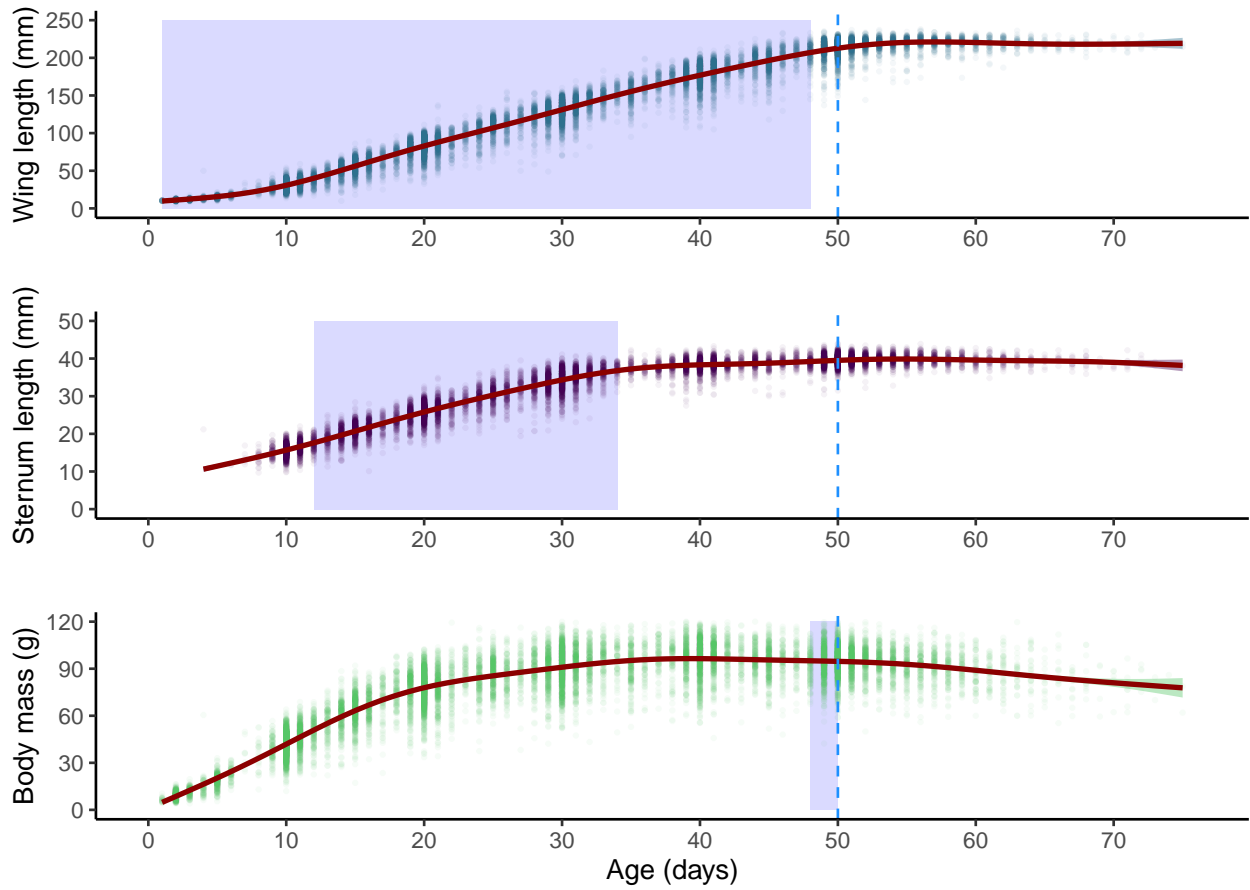


Figure 1: Fig. 1 - Growth curves for the wing, sternum and body mass in Alpine swifts nestlings. The rectangle in blue highlights the time interval used to calculate the growth rates used in the models. The vertical dashed line marks day 50 of chicks growth.


```

table_means <- data.frame(
  Trait = c("Wing", "Sternum", "Body mass"),
  No_measures = c(
    sum(
      !is.na(data_am_nestlings_all$wing)
    ),
    sum(
      !is.na(data_am_nestlings_all$sternum)
    ),
    sum(
      !is.na(data_am_nestlings_all$mass)
    )
  ),
  No_individuals = c(
    sum(
      !is.na(unique(subset(data_am_nestlings_all, !is.na(wing))$wing))
    ),
    sum(
      !is.na(unique(subset(data_am_nestlings_all, !is.na(sternum))$sternum))
    ),
    sum(
      !is.na(unique(subset(data_am_nestlings_all, !is.na(mass))$mass))
    )
  )
)

knitr::kable(table_means,
  caption =
    "Number of measures and of individuals measured for the measures wing length, sternum length and body mass",
  digits = 2
)

```

Table 1: Number of measures and of individuals measured for the measures wing length, sternum length and body mass for nestling Alpine swifts.

Trait	No_measures	No_individuals
Wing	15865	557
Sternum	12662	335
Body mass	15967	1153

```

table_means <- data.frame(
  Trait = c("Wing", "Sternum", "Body mass"),
  MeanSD = c(
    paste0(
      round(mean(data_am_nestlings_wing_50$wing_50), 2), " + ",
      round(sd(data_am_nestlings_wing_50$wing_50), 2)
    ),
    paste0(
      round(mean(data_am_nestlings_sternum_50$sternum_50), 2), " + ",
      round(sd(data_am_nestlings_sternum_50$sternum_50), 2)
    )
  )
)

```

```

),
paste0(
  round(mean(data_am_nestlings_mass_50$mass_50), 2), " + ",
  round(sd(data_am_nestlings_mass_50$mass_50), 2)
)
),
Range = c(
  paste0(
    min(data_am_nestlings_wing_50$wing_50), " - ",
    max(data_am_nestlings_wing_50$wing_50)
  ),
  paste0(
    min(data_am_nestlings_sternum_50$sternum_50), " - ",
    max(data_am_nestlings_sternum_50$sternum_50)
  ),
  paste0(
    min(data_am_nestlings_mass_50$mass_50), " - ",
    max(data_am_nestlings_mass_50$mass_50)
  )
),
C.V. = c(
  fun_cv(data_am_nestlings_wing_50$wing_50),
  fun_cv(data_am_nestlings_sternum_50$sternum_50),
  fun_cv(data_am_nestlings_mass_50$mass_50)
),
No = c(
  sum(
    !is.na(data_am_nestlings_wing_50$wing_50)
  ),
  sum(
    !is.na(data_am_nestlings_sternum_50$sternum_50)
  ),
  sum(
    !is.na(data_am_nestlings_mass_50$mass_50)
  )
)
)

```

```

knitr::kable(table_means,
caption =
  "Mean and standard deviation (SD), range (min and max), coefficient of variation (C.V.) and sample size",
  digits = 2
)

```

Table 2: Mean and standard deviation (SD), range (min and max), coefficient of variation (C.V.) and sample size in wing length, sternum length and body mass for nestling Alpine swifts.

Trait	MeanSD	Range	C.V.	No
Wing	212.46 + 11.85	151 – 237	5.58	2693
Sternum	39.48 + 1.42	30.5 – 44	3.60	2447

Trait	MeanSD	Range	C.V.	No
Body mass	94.81 + 9.51	56.3 – 127.3	10.03	2693

Supporting Information 01 - Climwin analyses

Weather data

To estimate the weather conditions during nestling development, we used meteorological data collected from five Swiss meteorological stations surrounding Biel and Solothurn (Bern-Zollikofen, Cressier, Grenchen, Koppigen, Wynau). Doing so allowed us to cover the whole foraging area of the swifts (up to 30 km in a single foraging trip) (Arn-Willi 1960) and to account for microenvironmental variations (i.e., strong weather events captured by one station only). Daily weather data were averaged across the five stations to obtain two variables: mean daily temperature (average air temperature at 2 m above ground for the whole day), daily precipitation (total rainfall for that day) and wind speed (daily mean of the wind speed scalar in m/s.). We also used a principal component analysis to calculate a daily first component (PC1) between temperature and precipitation for the meteorological data collected during the whole breeding season (May-August). PC1 explained 60% of the total variance in weather data, with factor loadings of 0.71 for the mean temperature and -0.71 for the mean precipitation. A high PC1 value, therefore, indicates warm and dry weather, whereas low values indicate cold and rainy weather.

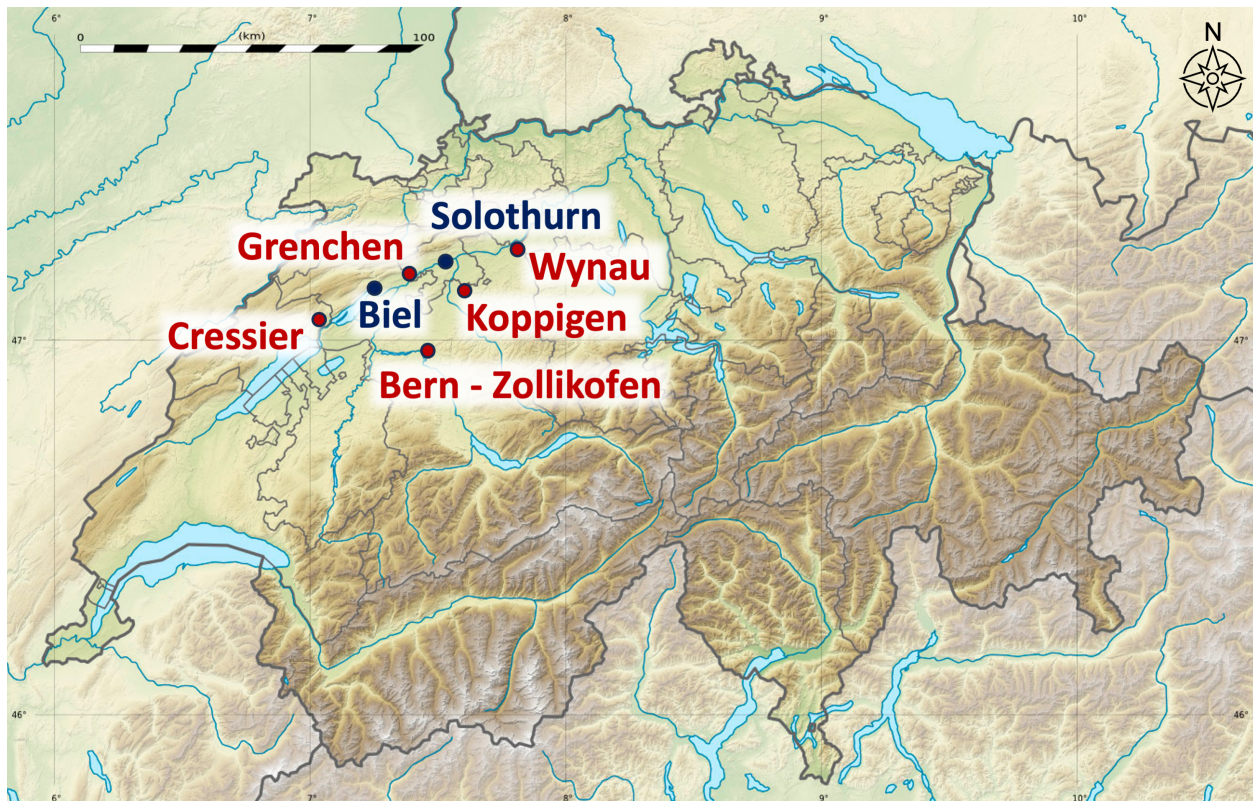


Figure 2: Map of Switzerland representing the location of the two Alpine swift colonies in Biel and Solothurn and the five meteorological stations Bern-Zollikofen, Cressier, Grenchen, Koppigen, Wynau.

[Base map] (https://commons.wikimedia.org/wiki/File:Switzerland_relief_location_map.jpg) credits to Eric Gaba [Wikimedia Commons user: Sting] (<https://commons.wikimedia.org/wiki/User:Sting>) and

Wikimedia [Commons user: NordNordWest] (<https://commons.wikimedia.org/wiki/User:NordNordWest>).
This file is licensed under the Creative Commons Attribution-Share Alike 3.0 Unported license.

PCA results

A Principal Component Analysis was run between average daily temperature and daily rain.

```
### PCA

# Principal component analyses to create a variable
# with temperature and rain together.
clim_pca1 <- princomp(data_clim_sub[, c("T.daily.mean", "Rain.daily")],
  cor = TRUE, scores = TRUE
)

summary(clim_pca1)
```

```
## Importance of components:
##                Comp.1    Comp.2
## Standard deviation    1.0879599 0.9035171
## Proportion of Variance 0.5918284 0.4081716
## Cumulative Proportion 0.5918284 1.0000000
```

```
# tr_comp_1 explains 59% of the variance
clim_pca1$loadings
```

```
##
## Loadings:
##                Comp.1 Comp.2
## T.daily.mean    0.707  0.707
## Rain.daily     -0.707  0.707
##
##                Comp.1 Comp.2
## SS loadings      1.0    1.0
## Proportion Var   0.5    0.5
## Cumulative Var   0.5    1.0
```

```
# tr_comp_1 = from cold and rainy to warm and dry day
# adding the new variable to the climate dataset
data_pca1 <- as.data.frame(clim_pca1$scores)
colnames(data_pca1) <- c("tr_comp_1", "tr_comp_2")
data_clim_sub <- cbind(data_clim_sub, data_pca1)
```

The loadings result shows that the Component 1 has positive values with warm days and dry days, and negative values cold and rainy days.

The importance of the components results show that the component 1 explains 59.2% of the variation in the climatic data.

The climwin approach

The overall approach for the climwin analysis is to compare the support by the data for competing hypotheses and to formalize them into regression models (van de Pol et al., 2016).

Competing models are based upon a baseline model (called also null model, a model without weather effects) and ranked using the deltaAICc, or the difference in terms of the Akaike Information Criterion values calculated for a small sample size between the candidate model and baseline model.

Climwin presents the models using the deltaAICc value relative to the baseline model (AICc of the candidate model - AICc of the baseline model). Therefore, a model that is more supported than the baseline model will have a negative deltaAICc value. On the same hand the model with the best support from the data, usually with lowest AICc, will be shown as the model with lowest deltaAICc in the climwin output.

The baseline model was a linear mixed model with the trait (wing length, sternum length or body mass) of the nestlings at 50 days of age in relation to brood size, colony of hatching, hatching day, and exact age of measurement (range 45-55 days of age). The function slidingwin creates a candidate set of competing models testing windows of different lengths for the weather variable of interest, in this study the mean daily ambient temperature (°C).

Non-linear effects of temperature on the traits were taken into account by checking for both linear and quadratic trends. This is mentioned in the climwin output as func = lin (only linear term) func = quad (linear and quadratic terms).

For the analyses we used an individual specific time window, based on the date when the nestlings had 50 days. We then looked for windows between this reference date and 50 days before.

Base model

According to (van de Pol et al. 2016), we built a base model that includes variables that can affect the traits. As random effects, we included brood ID and year as a factor to account for non-independence among nestlings belonging to the same brood and nestlings hatched in the same year, respectively.

Baseline models

```
# double check base models are the same as in script 01

wing_50_basemod <- lme4::lmer(
  wing_50 ~
    brood_size + colony +
    hatch_doy_sc + age_days_sc50 +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_wing_50
)

sternum_50_basemod <- lme4::lmer(
  sternum_50 ~
    brood_size + colony +
    age_days_sc50 + hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_sternum_50
)

mass_50_basemod <- lme4::lmer(
```

```

mass_50 ~
  brood_size + colony +
  hatch_doy_sc + age_days_sc50 +
  (1 | year_f) + (1 | nestcode_rearing),
control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
data = data_am_nestlings_mass_50
)

```

Finding the best window

Using the function `slidingwin` allows to search for the best climatic window

```

# -----
# ----- WING -----

wing_50_sw <- slidingwin(
  baseline = wing_50_basemod,
  xvar = list(
    tr_pc1 = data_clim_sub$tr_comp_1,
    temp_mean = data_clim_sub$T.daily.mean,
    rain_mean = data_clim_sub$Rain.daily,
    wind_mean = data_clim_sub$Wind.daily.mean
  ),
  stat = c("mean"),
  func = c("lin", "quad"),
  type = "relative", # relative to the individual
  range = c(50, 0),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_wing_50$date_50days
)
save(wing_50_sw, file = "output/wing_50_sw.rda")

# -----
# ----- STERNUM -----

sternum_50_sw <- slidingwin(
  baseline = sternum_50_basemod,
  xvar = list(
    tr_pc1 = data_clim_sub$tr_comp_1,
    temp_mean = data_clim_sub$T.daily.mean,
    rain_mean = data_clim_sub$Rain.daily,
    wind_mean = data_clim_sub$Wind.daily.mean
  ),
  stat = c("mean"),
  func = c("lin", "quad"),
  type = "relative",
  range = c(50, 0),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_sternum_50$date_50days
)

```

```

save(sternum_50_sw, file = "output/sternum_50_sw.rda")

# -----
# ----- MASS -----

mass_50_sw <- slidingwin(
  baseline = mass_50_basemod,
  xvar = list(
    tr_pc1 = data_clim_sub$tr_comp_1,
    temp_mean = data_clim_sub$T.daily.mean,
    rain_mean = data_clim_sub$Rain.daily,
    wind_mean = data_clim_sub$Wind.daily.mean
  ),
  stat = c("mean"),
  func = c("lin", "quad"),
  type = "relative",
  range = c(50, 0),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_mass_50$date_50days
)
save(mass_50_sw, file = "output/mass_50_sw.rda")

```

Investigating the models

Loading the models

```

model_load <- list.files("output/", pattern = "*.rda", full.names = FALSE)
for (i in seq_len(length(model_load))) {
  assign(model_load[i], load(paste0("output/", model_load[i])))
}

```

Wing length at 50 days The linear+quadratic term for the PC1 better explains the variation in the data (deltaAICc has the lowest value), sorted by deltaAICc such that the best supported model is on top.

```

wing_50_mod_combos <- (wing_50_sw$combos)

wing_50_mod_combos[order(wing_50_mod_combos$DeltaAICc), ]

```

##	response	climate	type	stat	func	DeltaAICc	WindowOpen	WindowClose
## 5	wing_50	tr_pc1	relative	mean	quad	-54.89	50	4
## 1	wing_50	tr_pc1	relative	mean	lin	-51.78	48	2
## 2	wing_50	temp_mean	relative	mean	lin	-44.85	44	0
## 6	wing_50	temp_mean	relative	mean	quad	-44.06	46	1
## 3	wing_50	rain_mean	relative	mean	lin	-35.67	50	4
## 7	wing_50	rain_mean	relative	mean	quad	-33.13	50	4
## 8	wing_50	wind_mean	relative	mean	quad	-30.09	43	16
## 4	wing_50	wind_mean	relative	mean	lin	-20.50	50	16

```
# to save the row name of the model with the lowest DeltaAICc
n_bestmod_wing_50 <- as.numeric(rownames(
  wing_50_mod_combos[order(wing_50_mod_combos$DeltaAICc), ]
))[1]
```

The best quadratic models The 30 best windows for the linear+quadratic models sorted by deltaAICc. To investigate any other tested hypothesis we can simply replace the number on the left or in the double square brackets with the corresponding list number.

```
head(wing_50_sw[[n_bestmod_wing_50]]$Dataset, 30)
```

##	deltaAICc	WindowOpen	WindowClose	ModelBeta	Std.Error	ModelBetaQ	Std.ErrorQ	ModelBetaC	
## 1322	-54.89092	50	4	6.551631	0.3361476	11.0387223	0.4863652	NA	
## 1223	-54.84205	48	2	12.455726	0.3366066	2.7489047	0.4859876	NA	
## 1323	-54.76311	50	3	7.868213	0.3362721	9.3874591	0.4861168	NA	
## 1325	-54.45219	50	1	9.825671	0.3363533	7.5729701	0.4851552	NA	
## 1324	-54.41977	50	2	9.981766	0.3363880	6.7217760	0.4856579	NA	
## 1224	-54.32639	48	1	12.650438	0.3366280	2.9862848	0.4856233	NA	
## 1272	-54.08562	49	3	8.544611	0.3364165	8.2142865	0.4861785	NA	
## 1222	-54.03660	48	3	10.263727	0.3365971	5.4752835	0.4866291	NA	
## 1273	-53.96047	49	2	10.706399	0.3365262	5.4821138	0.4856960	NA	
## 1271	-53.87250	49	4	7.190108	0.3363290	9.8540777	0.4864658	NA	
## 1274	-53.68233	49	1	10.674798	0.3365185	6.0797486	0.4852664	NA	
## 1174	-52.90919	47	2	12.437476	0.3367720	2.1534341	0.4856307	NA	
## 1225	-52.84705	48	0	14.456870	0.3368176	0.3027216	0.4858772	NA	
## 1221	-52.84061	48	4	8.940424	0.3366223	6.9043294	0.4871025	NA	
## 1175	-52.64052	47	1	12.588455	0.3367644	2.5504995	0.4852247	NA	
## 1326	-52.30213	50	0	11.662259	0.3366616	4.8728030	0.4856115	NA	
## 1126	-51.69699	46	2	12.483527	0.3369035	1.6598150	0.4853873	NA	
## 1127	-51.66755	46	1	12.552901	0.3368703	2.2735931	0.4849586	NA	
## 1173	-51.55800	47	3	10.411740	0.3368546	4.4021966	0.4863902	NA	
## 1275	-51.55052	49	0	12.551106	0.3367897	3.2700689	0.4856945	NA	
## 1176	-51.26785	47	0	14.255534	0.3369472	0.1701031	0.4854594	NA	
## 1079	-50.76999	45	2	14.147982	0.3371526	-1.1918020	0.4852511	NA	
## 1080	-50.35263	45	1	14.108448	0.3371538	-0.4976083	0.4849790	NA	
## 1128	-50.35040	46	0	14.079579	0.3370639	0.1735787	0.4851918	NA	
## 1125	-50.23049	46	3	10.591392	0.3370209	3.6320887	0.4861570	NA	
## 1172	-50.15347	47	4	9.122739	0.3369127	5.6884318	0.4869242	NA	
## 1081	-49.36892	45	0	15.294356	0.3372935	-2.0929514	0.4851141	NA	
## 1124	-49.10548	46	4	9.264878	0.3370408	5.0356618	0.4866230	NA	
## 1078	-48.93918	45	3	12.283747	0.3373236	0.8309290	0.4859783	NA	
## 1321	-47.56793	50	5	5.641579	0.3366908	10.7798666	0.4869438	NA	
##	ModelInt	Function	Furthest	Closest	Statistics	Type K	ModWeight	sample.size	Randomised
## 1322	217.0848	quad	50	0	mean relative	0	0.084572833	25	no
## 1223	216.9736	quad	50	0	mean relative	0	0.082531069	25	no
## 1323	217.0928	quad	50	0	mean relative	0	0.079337301	25	no
## 1325	216.8889	quad	50	0	mean relative	0	0.067914088	25	no
## 1324	217.0447	quad	50	0	mean relative	0	0.066822304	25	no
## 1224	216.8182	quad	50	0	mean relative	0	0.063774173	25	no
## 1272	217.0634	quad	50	0	mean relative	0	0.056540774	25	no
## 1222	217.0684	quad	50	0	mean relative	0	0.055171766	25	no
## 1273	216.9947	quad	50	0	mean relative	0	0.053111227	25	no

##	1271	217.0835	quad	50	0	mean relative	0	0.050825589	25	no
##	1274	216.8472	quad	50	0	mean relative	0	0.046215462	25	no
##	1174	217.0212	quad	50	0	mean relative	0	0.031398094	25	no
##	1225	216.8362	quad	50	0	mean relative	0	0.030437574	25	no
##	1221	217.1366	quad	50	0	mean relative	0	0.030339628	25	no
##	1175	216.8444	quad	50	0	mean relative	0	0.027451227	25	no
##	1326	216.9188	quad	50	0	mean relative	0	0.023178328	25	no
##	1126	217.0579	quad	50	0	mean relative	0	0.017126836	25	no
##	1127	216.8582	quad	50	0	mean relative	0	0.016876543	25	no
##	1173	217.1755	quad	50	0	mean relative	0	0.015977034	25	no
##	1275	216.8741	quad	50	0	mean relative	0	0.015917366	25	no
##	1176	216.8405	quad	50	0	mean relative	0	0.013819467	25	no
##	1079	217.0713	quad	50	0	mean relative	0	0.010774101	25	no
##	1080	216.8755	quad	50	0	mean relative	0	0.008744850	25	no
##	1128	216.8407	quad	50	0	mean relative	0	0.008735114	25	no
##	1125	217.2351	quad	50	0	mean relative	0	0.008226783	25	no
##	1172	217.2726	quad	50	0	mean relative	0	0.007915981	25	no
##	1081	216.8539	quad	50	0	mean relative	0	0.005347413	25	no
##	1124	217.3224	quad	50	0	mean relative	0	0.004687464	25	no
##	1078	217.2487	quad	50	0	mean relative	0	0.004313467	25	no
##	1321	217.3428	quad	50	0	mean relative	0	0.002173012	25	no

The best windows (deltaAICc difference from the best model < 2) for the linear+quadratic models sorted by deltaAICc. All models with the lowest AICc present very comparable windows:

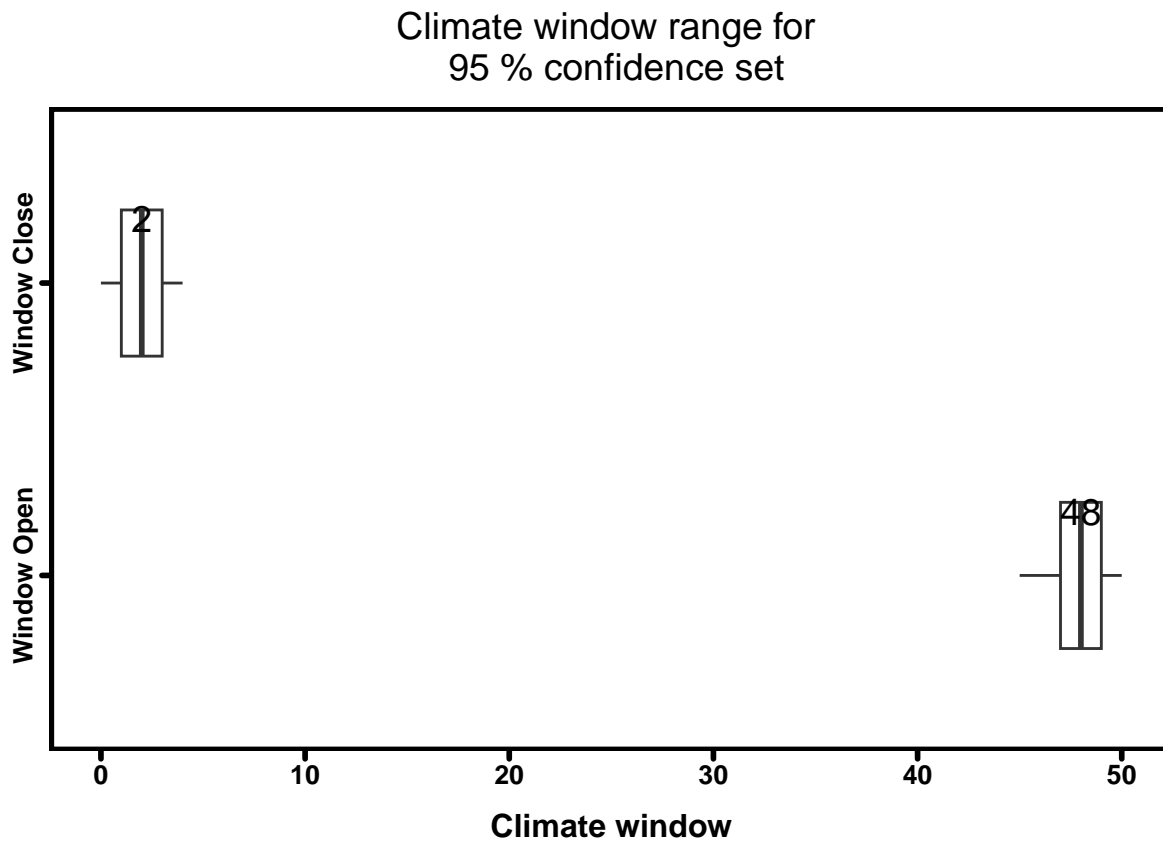
```
wing_50_bestmod <- subset(
  wing_50_sw[[n_bestmod_wing_50]]$Dataset,
  deltaAICc < wing_50_sw[[n_bestmod_wing_50]]$Dataset$deltaAICc[1] + 2
)
wing_50_bestmod
```

##	deltaAICc	WindowOpen	WindowClose	ModelBeta	Std.Error	ModelBetaQ	Std.ErrorQ	ModelBetaC		
##	1322	-54.89092	50	4	6.551631	0.3361476	11.038722	0.4863652	NA	
##	1223	-54.84205	48	2	12.455726	0.3366066	2.748905	0.4859876	NA	
##	1323	-54.76311	50	3	7.868213	0.3362721	9.387459	0.4861168	NA	
##	1325	-54.45219	50	1	9.825671	0.3363533	7.572970	0.4851552	NA	
##	1324	-54.41977	50	2	9.981766	0.3363880	6.721776	0.4856579	NA	
##	1224	-54.32639	48	1	12.650438	0.3366280	2.986285	0.4856233	NA	
##	1272	-54.08562	49	3	8.544611	0.3364165	8.214286	0.4861785	NA	
##	1222	-54.03660	48	3	10.263727	0.3365971	5.475283	0.4866291	NA	
##	1273	-53.96047	49	2	10.706399	0.3365262	5.482114	0.4856960	NA	
##	1271	-53.87250	49	4	7.190108	0.3363290	9.854078	0.4864658	NA	
##	1274	-53.68233	49	1	10.674798	0.3365185	6.079749	0.4852664	NA	
##	1174	-52.90919	47	2	12.437476	0.3367720	2.153434	0.4856307	NA	
##	ModelInt	Function	Furthest	Closest	Statistics	Type	K	ModWeight	sample.size	Randomised
##	1322	217.0848	quad	50	0	mean relative	0	0.08457283	25	no
##	1223	216.9736	quad	50	0	mean relative	0	0.08253107	25	no
##	1323	217.0928	quad	50	0	mean relative	0	0.07933730	25	no
##	1325	216.8889	quad	50	0	mean relative	0	0.06791409	25	no
##	1324	217.0447	quad	50	0	mean relative	0	0.06682230	25	no
##	1224	216.8182	quad	50	0	mean relative	0	0.06377417	25	no
##	1272	217.0634	quad	50	0	mean relative	0	0.05654077	25	no

##	1222	217.0684	quad	50	0	mean relative 0	0.05517177	25	no
##	1273	216.9947	quad	50	0	mean relative 0	0.05311123	25	no
##	1271	217.0835	quad	50	0	mean relative 0	0.05082559	25	no
##	1274	216.8472	quad	50	0	mean relative 0	0.04621546	25	no
##	1174	217.0212	quad	50	0	mean relative 0	0.03139809	25	no

Windows plot It's possible to extract the time windows of all the best supported models (i.e. multi-model inference). This panel shows the opening and closing points of the time windows that were best supported by the data, here those models that made up 95% model confidence set.

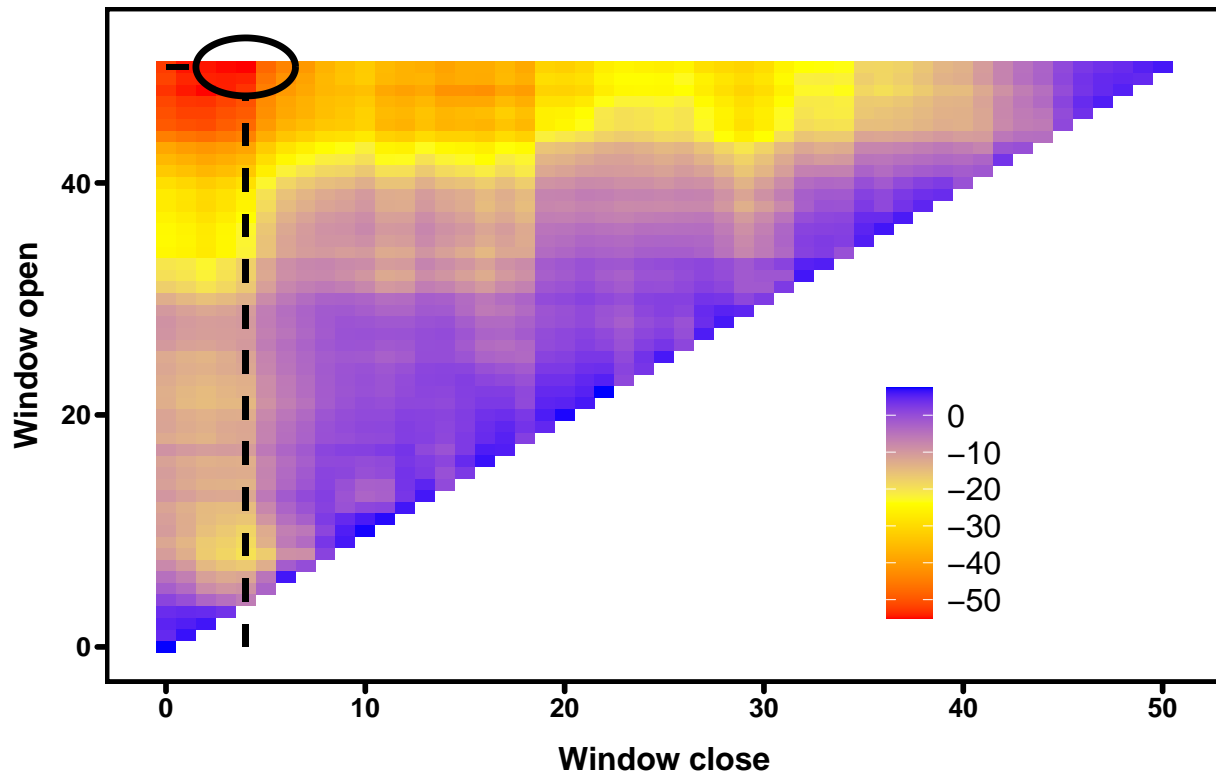
```
plotwin(wing_50_sw[[n_bestmod_wing_50]]$Dataset)
```



Delta plot The variation in deltaAICc between time windows can be better investigated using the following plot:

```
plotdelta(  
  dataset =  
    wing_50_sw[[n_bestmod_wing_50]]$Dataset, arrow = TRUE  
)
```

ΔAICc (compared to null model)

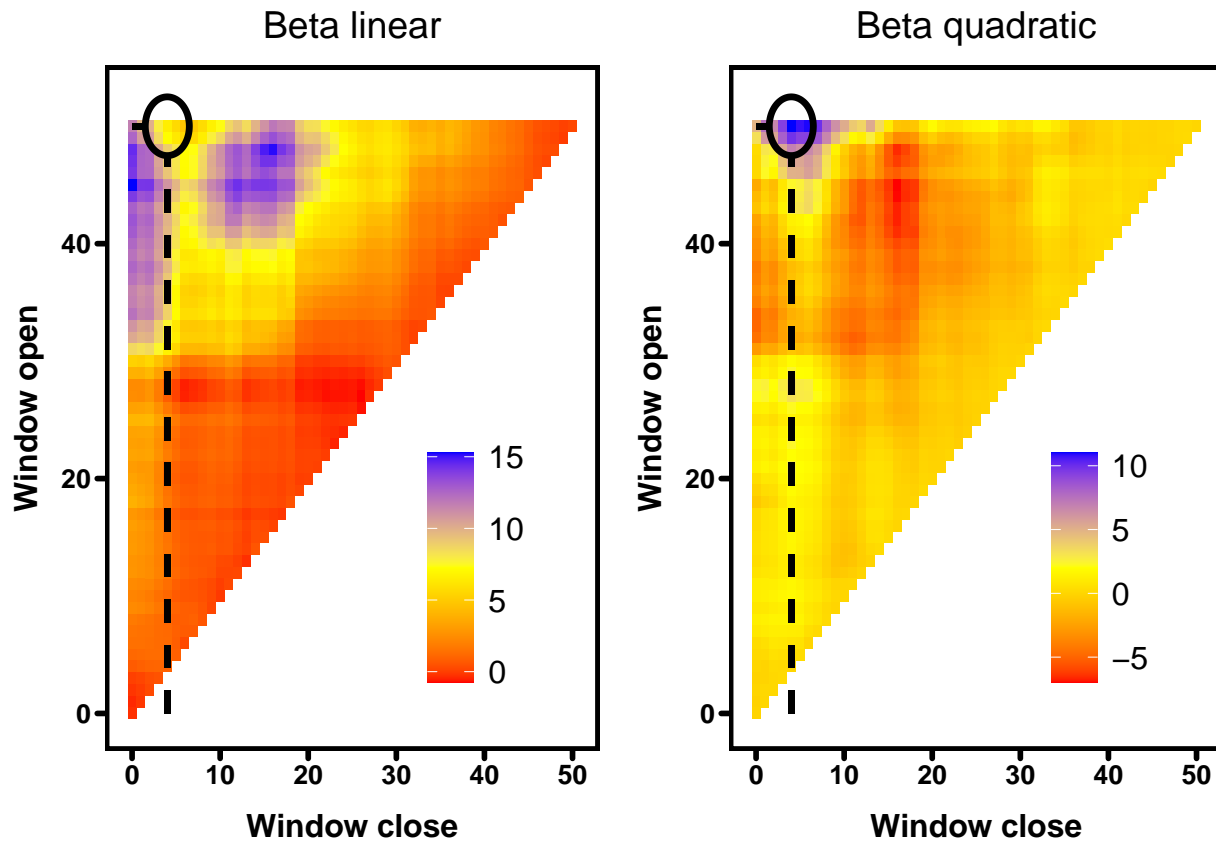


Warmer areas shows values with the lowest deltaAICc (i.e. “best models”). As explained by van de Pol et al., 2016, these deltaAICc landscapes of the different time windows shows multiple peaks (red areas) instead of a clear single peak. This can indicate the presence of multiple (e.g. possibly both long- and short-lag) weather signals within the same weather variable, but it can also occur due to collinearity or chance.

The evidence for multiple signals can be therefore investigated by adding the best supported of the weather windows to the baseline model, and re-fitting all the different time windows again: this tests whether there is still strong model support for the second best (e.g. short-lag) weather window once the other best supported (e.g. long-lag) weather window has been accounted for in the baseline model (here in the Step 2).

Beta plot This panel shows the model support (deltaAICc) for all fitted time windows tried, shown for each combination of Window open (y-axis) and Window close (x-axis). Models with the lowest deltaAICc (red) are the best supported (colours show the deltaAICc levels compared to the null model, see legend). Strongly supported windows will often be grouped together.

```
plotbetas(wing_50_sw[[n_bestmod_wing_50]]$Dataset, arrow = TRUE)
```



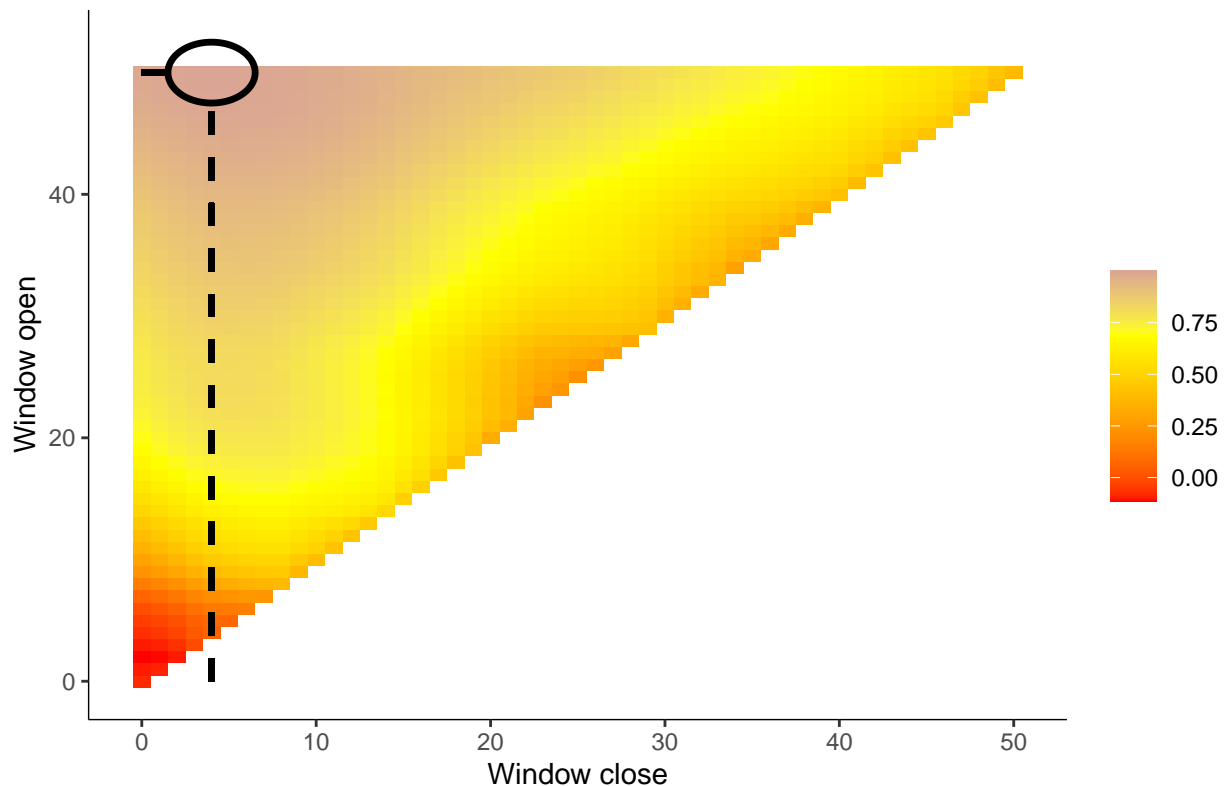
Autocollinearity Correlation between the mean temperature during the best supported time window and the mean temperature over all other time windows.

```
autocoll <- autowin(
  reference = wing_50_sw[[n_bestmod_wing_50]],
  baseline = wing_50_basemod,
  xvar = list(
    tr_pcl = data_clim_sub$tr_comp_1
  ),
  type = "relative",
  range = c(50, 0),
  stat = "mean",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_wing_50$date_50days,
  func = "quad",
  # cmissing = FALSE,
  cinterval = "day"
)

save(autocoll, file = "output/wing_50_autocoll.rda")
```

```
load(file = "output/wing_50_autocoll.rda")
plotcor(autocoll, type = "A", arrow = TRUE)
```

Correlation between single window and all other windows



Randwin Using `randwin()` to randomize the identity of the nestling, we are able to check if the window that was found before is actually important, or the relationship was just random.

```
# Performing randomization to identify
# likelihood of of signals occurring by chance

wing_50_rand1000 <- randwin(
  repeats = 1000,
  baseline = wing_50_basemod,
  xvar = list(tr_pcl = data_clim_sub$str_comp_1),
  stat = c("mean"),
  func = c("quad"),
  type = "relative", # relative to the individual
  range = c(50, 0),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_wing_50$date_50days,
  cmissing = FALSE
)
save(wing_50_rand1000, file = "output/wing_50_rand1000.rda")

load("output/wing_50_rand1000.rda")

climwin::pvalue(
  datasetrand = wing_50_rand1000[[1]],
  dataset = wing_50_sw[[n_bestmod_wing_50]]$Dataset,
```

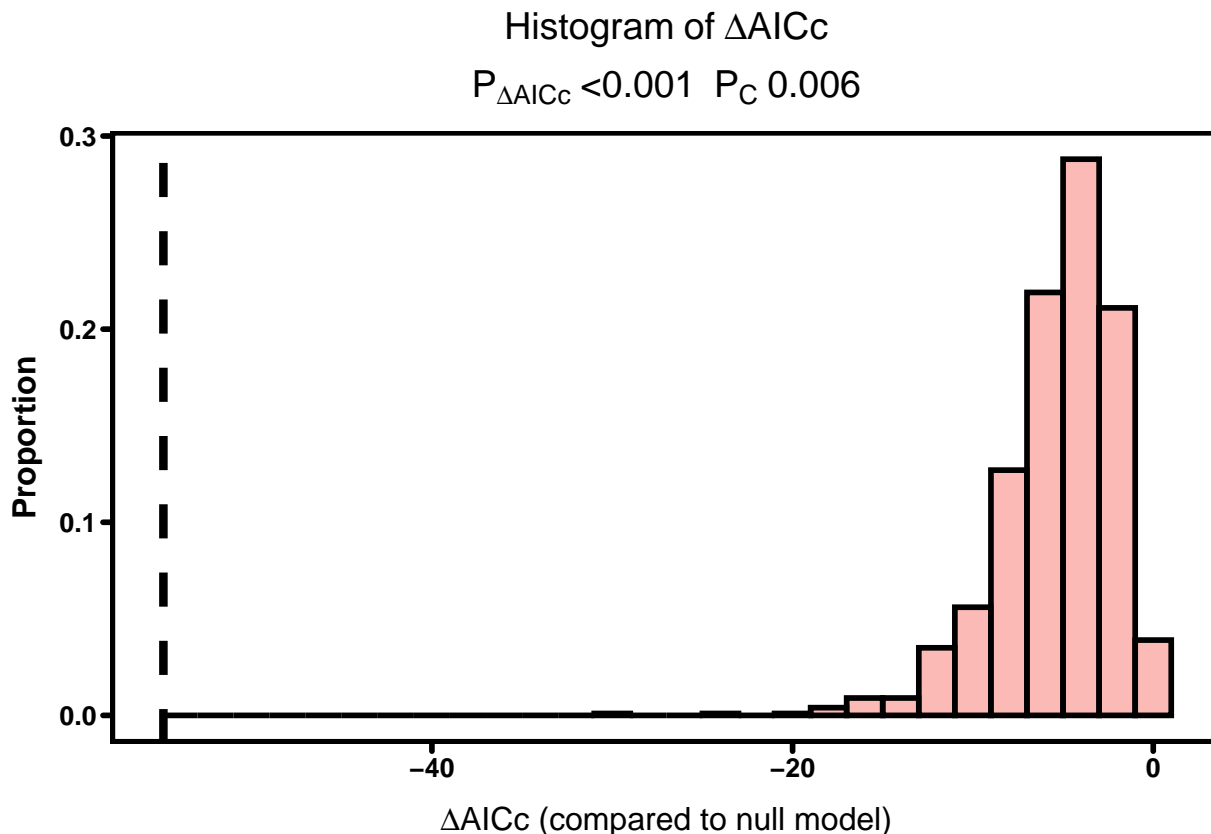
```
metric = "AIC",
sample.size = wing_50_sw[[n_bestmod_wing_50]]$Dataset$sample.size[1]
# sample size = number of years
)
```

```
## [1] "<0.001"
```

This panel shows a histogram of all $\Delta AICc$ values from all models run on the randomized (in grey) and the $\Delta AICc$ of the best model fitted on the observed data (dashed line). The best climate window in the randomized data is by definition a false positive, as the randomization procedure removes any climate signal from the data. The percentage of randomizations that generates a $\Delta AICc$ value that is at least as low as the $\Delta AICc$ value of the best model fitted to the observed data (P value above) is a measure of how likely it is that one obtained the observed $\Delta AICc$ by chance and thus whether the signal is likely to be real or not. In our case we can say that the candidate signal is very unlikely to be a false positive: $P < 0.001$.

```
climwin::plothist(
  datasetrand = wing_50_rand1000[[1]],
  dataset = wing_50_sw[[n_bestmod_wing_50]]$Dataset
)
```

```
## Warning: The dot-dot notation (`.density.`) was deprecated in ggplot2 3.4.0.
## i Please use `after_stat(density)` instead.
## i The deprecated feature was likely used in the climwin package.
## Please report the issue at <https://github.com/LiamDBailey/climwin/issues>.
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was generated.
```



Extracting the best window The best window was obtained by averaging the opening and closing dates for all the best models ($\Delta AICc < 2$). The best window opened 48.92 (mean; SD: 1) and closed 2.33 (mean; SD: 1.07) days before the 50 days measurement, which corresponds to the age of the nestling of 1 and 48 days old.

```
wing_50_bestsw <- singlewin(
  baseline = wing_50_basemod,
  xvar = list(
    tr_pc1 = data_clim_sub$tr_comp_1
  ),
  stat = c("mean"),
  func = c("quad"),
  type = "relative",
  range = c(
    round(mean(wing_50_bestmod$WindowOpen), 0),
    round(mean(wing_50_bestmod$WindowClose), 0)
  ),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_wing_50$date_50days
)
```

```
## fixed-effect model matrix is rank deficient so dropping 2 columns / coefficients
```

```
# adding the climatic variable to the wing_50 dataset
data_am_nestlings_wing_50$signal_for_wing_50 <-
  wing_50_bestsw$BestModelData$climate
```

Sternum length at 50 days The linear+quadratic term for the PC1 better explains the variation in the data ($\Delta AICc$ has the lowest value), sorted by $\Delta AICc$ such that the best supported model is on top.

```
sternum_50_mod_combos <- (sternum_50_sw$combos)

sternum_50_mod_combos[order(sternum_50_mod_combos$DeltaAICc), ]
```

##	response	climate	type	stat	func	DeltaAICc	WindowOpen	WindowClose
## 5	sternum_50	tr_pc1	relative	mean	quad	-6.92	31	16
## 1	sternum_50	tr_pc1	relative	mean	lin	-5.23	50	16
## 8	sternum_50	wind_mean	relative	mean	quad	-3.74	45	16
## 2	sternum_50	temp_mean	relative	mean	lin	-2.64	48	14
## 4	sternum_50	wind_mean	relative	mean	lin	-1.15	45	20
## 3	sternum_50	rain_mean	relative	mean	lin	0.79	29	16
## 7	sternum_50	rain_mean	relative	mean	quad	3.84	43	2
## 6	sternum_50	temp_mean	relative	mean	quad	4.93	49	14

```
n_bestmod_sternum_50 <- as.numeric(rownames(
  sternum_50_mod_combos[order(sternum_50_mod_combos$DeltaAICc), ]
))[1]
```

The best quadratic models The 30 best windows for the linear+quadratic models sorted by $\Delta AICc$. To investigate any other tested hypothesis we can simply replace the number on the left or in the double square brackets with the corresponding list number.

```
head(sternum_50_sw[[n_bestmod_sternum_50]]$Dataset, 30)
```

##	deltaAICc	WindowOpen	WindowClose	ModelBeta	Std.Error	ModelBetaQ	Std.ErrorQ	ModelBetaC		
## 512	-6.921814	31	16	0.6439868	0.04890045	-0.4618389	0.06793423	NA		
## 513	-6.567573	31	15	0.6923178	0.04888465	-0.4919496	0.06791674	NA		
## 545	-6.054905	32	16	0.6681248	0.04894310	-0.4998184	0.06789905	NA		
## 514	-5.998077	31	14	0.7277946	0.04889078	-0.5134053	0.06788579	NA		
## 546	-5.799299	32	15	0.7153519	0.04891801	-0.5325067	0.06787628	NA		
## 974	-5.526967	43	16	0.8627472	0.04896451	-0.7063225	0.06795367	NA		
## 1019	-5.476339	44	16	0.8637459	0.04896773	-0.6758509	0.06798146	NA		
## 1065	-5.469180	45	16	0.8657824	0.04898187	-0.6529619	0.06801712	NA		
## 1066	-5.062738	45	15	0.8729279	0.04899598	-0.6545456	0.06799652	NA		
## 930	-5.000570	42	16	0.8194125	0.04897776	-0.6897538	0.06795795	NA		
## 511	-5.000428	31	17	0.5647773	0.04895028	-0.4018979	0.06797507	NA		
## 1018	-4.955888	44	17	0.8356887	0.04897945	-0.6573560	0.06805801	NA		
## 975	-4.940319	43	15	0.8651537	0.04897529	-0.7012805	0.06793445	NA		
## 1064	-4.939830	45	17	0.8360772	0.04899397	-0.6333999	0.06809447	NA		
## 1020	-4.897758	44	15	0.8637132	0.04898299	-0.6678083	0.06796395	NA		
## 547	-4.855831	32	14	0.7328238	0.04892918	-0.5384602	0.06786275	NA		
## 526	-4.816052	31	2	0.9740741	0.04896630	-0.8003427	0.06818197	NA		
## 1067	-4.797222	45	14	0.8853837	0.04899490	-0.6620802	0.06800663	NA		
## 579	-4.759085	33	16	0.6867554	0.04897166	-0.4891356	0.06787970	NA		
## 1209	-4.749999	48	16	0.8035990	0.04902974	-0.5286647	0.06819112	NA		
## 973	-4.696554	43	17	0.8180629	0.04898258	-0.6667891	0.06803156	NA		
## 931	-4.631783	42	15	0.8351901	0.04897879	-0.7008244	0.06793034	NA		
## 1063	-4.623354	45	18	0.8130383	0.04899888	-0.6083582	0.06808147	NA		
## 1017	-4.572024	44	18	0.8097343	0.04898085	-0.6281466	0.06804767	NA		
## 510	-4.517666	31	18	0.5225618	0.04892357	-0.3790670	0.06793934	NA		
## 1021	-4.512650	44	14	0.8705341	0.04898466	-0.6666277	0.06796981	NA		
## 480	-4.511638	30	16	0.5494362	0.04893859	-0.3708243	0.06810227	NA		
## 944	-4.438145	42	2	1.0529507	0.04902264	-1.0107469	0.06804540	NA		
## 976	-4.436852	43	14	0.8659776	0.04897874	-0.6932581	0.06793719	NA		
## 1210	-4.414423	48	15	0.8045932	0.04903585	-0.5304052	0.06816179	NA		
##	ModelInt	Function	Furthest	Closest	Statistics	Type	K	ModWeight	sample.size	Randomised
## 512	40.19533	quad	50	0	mean relative	0	0.020872955		22	no
## 513	40.18964	quad	50	0	mean relative	0	0.017484832		22	no
## 545	40.19438	quad	50	0	mean relative	0	0.013531225		22	no
## 514	40.18390	quad	50	0	mean relative	0	0.013152155		22	no
## 546	40.18927	quad	50	0	mean relative	0	0.011907840		22	no
## 974	40.18512	quad	50	0	mean relative	0	0.010391942		22	no
## 1019	40.17877	quad	50	0	mean relative	0	0.010132185		22	no
## 1065	40.17287	quad	50	0	mean relative	0	0.010095982		22	no
## 1066	40.17256	quad	50	0	mean relative	0	0.008239309		22	no
## 930	40.19599	quad	50	0	mean relative	0	0.007987137		22	no
## 511	40.20409	quad	50	0	mean relative	0	0.007986572		22	no
## 1018	40.18291	quad	50	0	mean relative	0	0.007810675		22	no
## 975	40.18407	quad	50	0	mean relative	0	0.007750109		22	no
## 1064	40.17707	quad	50	0	mean relative	0	0.007748212		22	no
## 1020	40.17783	quad	50	0	mean relative	0	0.007586924		22	no
## 547	40.18476	quad	50	0	mean relative	0	0.007429531		22	no
## 526	40.13406	quad	50	0	mean relative	0	0.007283222		22	no
## 1067	40.16965	quad	50	0	mean relative	0	0.007214973		22	no
## 579	40.18509	quad	50	0	mean relative	0	0.007078698		22	no

##	1209	40.16699	quad	50	0	mean relative	0	0.007046612	22	no
##	973	40.18940	quad	50	0	mean relative	0	0.006860802	22	no
##	931	40.19451	quad	50	0	mean relative	0	0.006642171	22	no
##	1063	40.17907	quad	50	0	mean relative	0	0.006614238	22	no
##	1017	40.18465	quad	50	0	mean relative	0	0.006446643	22	no
##	510	40.21307	quad	50	0	mean relative	0	0.006273789	22	no
##	1021	40.17428	quad	50	0	mean relative	0	0.006258073	22	no
##	480	40.19762	quad	50	0	mean relative	0	0.006254908	22	no
##	944	40.16213	quad	50	0	mean relative	0	0.006029234	22	no
##	976	40.18070	quad	50	0	mean relative	0	0.006025337	22	no
##	1210	40.16909	quad	50	0	mean relative	0	0.005958144	22	no

The best windows (deltaAICc difference from the best model < 2) for the linear+quadratic models sorted by deltaAICc. All models with the lowest AICc present very comparable windows:

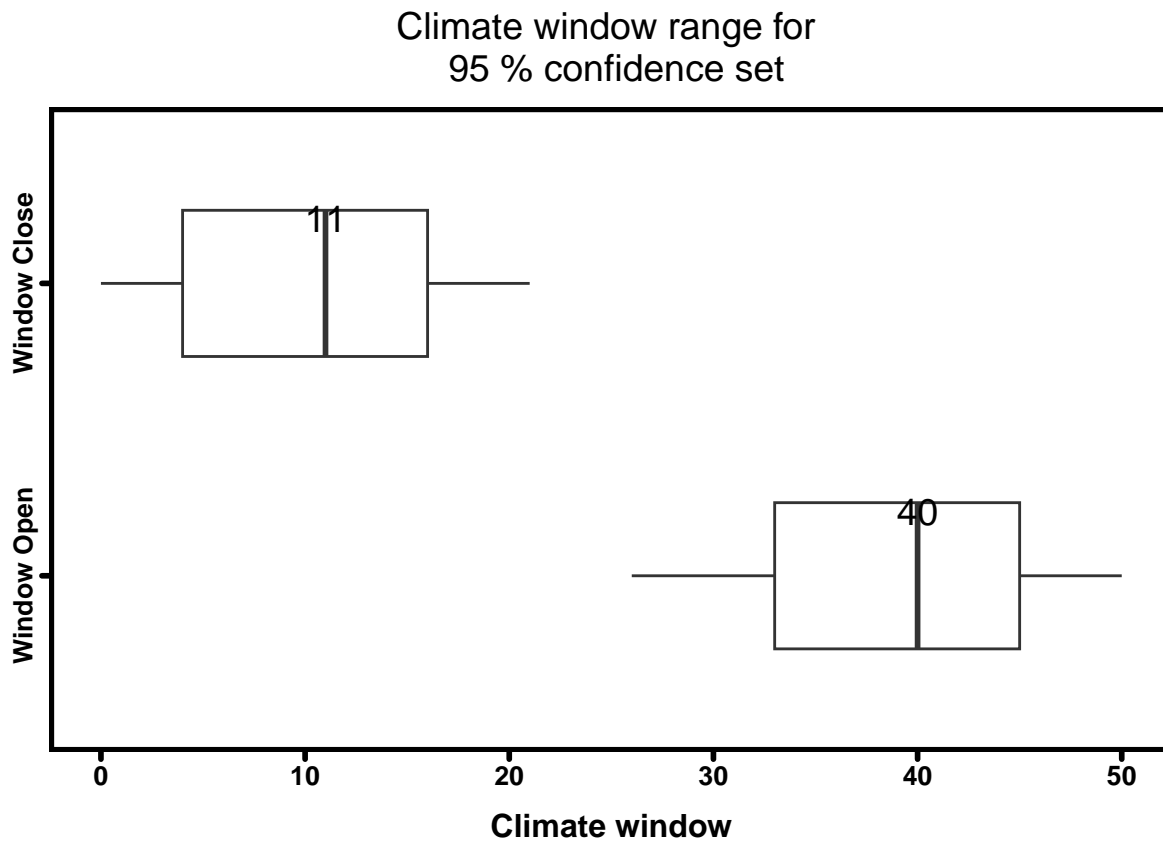
```
sternum_50_bestmod <- subset(
  sternum_50_sw[[n_bestmod_sternum_50]]$Dataset,
  deltaAICc < sternum_50_sw[[n_bestmod_sternum_50]]$Dataset$deltaAICc[1] + 2
)
sternum_50_bestmod
```

##	deltaAICc	WindowOpen	WindowClose	ModelBeta	Std.Error	ModelBetaQ	Std.ErrorQ	ModelBetaC		
##	512	-6.921814	31	16	0.6439868	0.04890045	-0.4618389	0.06793423	NA	
##	513	-6.567573	31	15	0.6923178	0.04888465	-0.4919496	0.06791674	NA	
##	545	-6.054905	32	16	0.6681248	0.04894310	-0.4998184	0.06789905	NA	
##	514	-5.998077	31	14	0.7277946	0.04889078	-0.5134053	0.06788579	NA	
##	546	-5.799299	32	15	0.7153519	0.04891801	-0.5325067	0.06787628	NA	
##	974	-5.526967	43	16	0.8627472	0.04896451	-0.7063225	0.06795367	NA	
##	1019	-5.476339	44	16	0.8637459	0.04896773	-0.6758509	0.06798146	NA	
##	1065	-5.469180	45	16	0.8657824	0.04898187	-0.6529619	0.06801712	NA	
##	1066	-5.062738	45	15	0.8729279	0.04899598	-0.6545456	0.06799652	NA	
##	930	-5.000570	42	16	0.8194125	0.04897776	-0.6897538	0.06795795	NA	
##	511	-5.000428	31	17	0.5647773	0.04895028	-0.4018979	0.06797507	NA	
##	1018	-4.955888	44	17	0.8356887	0.04897945	-0.6573560	0.06805801	NA	
##	975	-4.940319	43	15	0.8651537	0.04897529	-0.7012805	0.06793445	NA	
##	1064	-4.939830	45	17	0.8360772	0.04899397	-0.6333999	0.06809447	NA	
##	ModelInt	Function	Furthest	Closest	Statistics	Type	K	ModWeight	sample.size	Randomised
##	512	40.19533	quad	50	0	mean relative	0	0.020872955	22	no
##	513	40.18964	quad	50	0	mean relative	0	0.017484832	22	no
##	545	40.19438	quad	50	0	mean relative	0	0.013531225	22	no
##	514	40.18390	quad	50	0	mean relative	0	0.013152155	22	no
##	546	40.18927	quad	50	0	mean relative	0	0.011907840	22	no
##	974	40.18512	quad	50	0	mean relative	0	0.010391942	22	no
##	1019	40.17877	quad	50	0	mean relative	0	0.010132185	22	no
##	1065	40.17287	quad	50	0	mean relative	0	0.010095982	22	no
##	1066	40.17256	quad	50	0	mean relative	0	0.008239309	22	no
##	930	40.19599	quad	50	0	mean relative	0	0.007987137	22	no
##	511	40.20409	quad	50	0	mean relative	0	0.007986572	22	no
##	1018	40.18291	quad	50	0	mean relative	0	0.007810675	22	no
##	975	40.18407	quad	50	0	mean relative	0	0.007750109	22	no
##	1064	40.17707	quad	50	0	mean relative	0	0.007748212	22	no

Windows plot It's possible to extract the time windows of all the best supported models (i.e. multi-model inference). This panel shows the opening and closing points of the time windows that were best supported

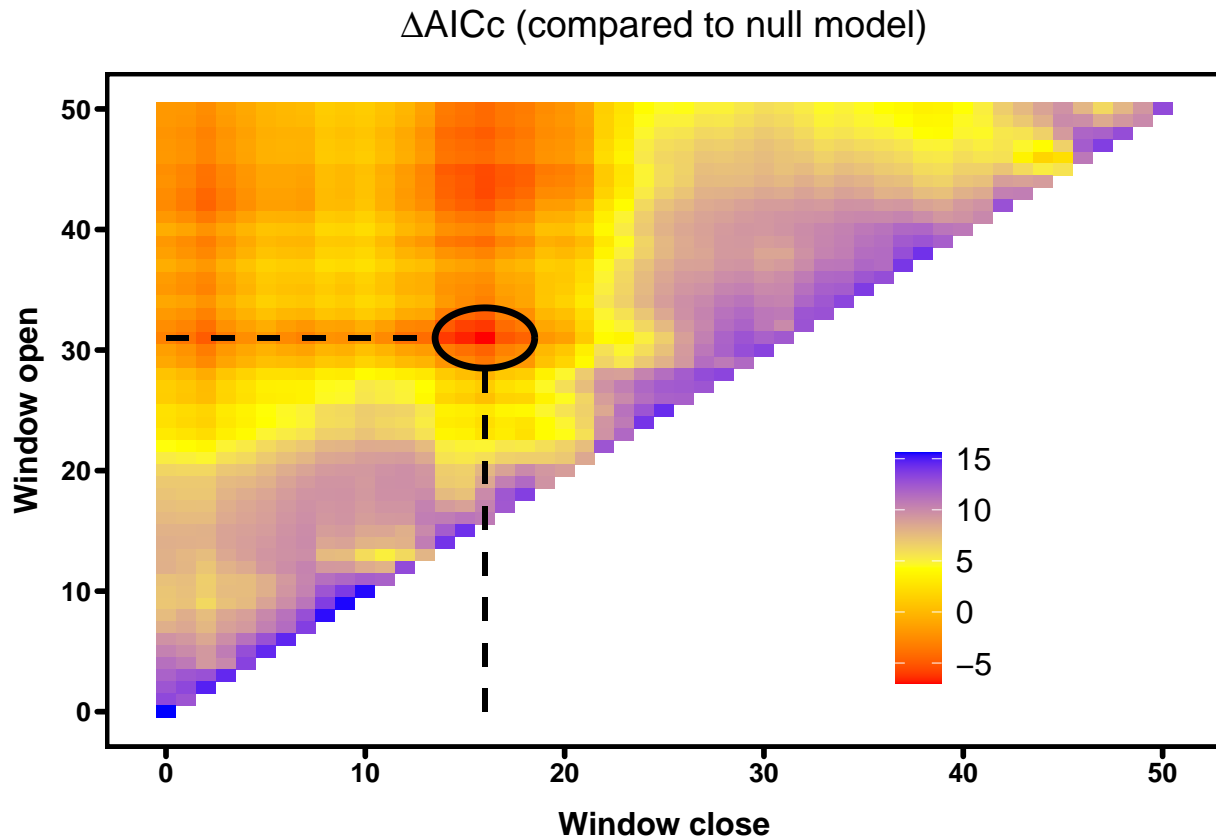
by the data, here those models that made up 95% model confidence set.

```
plotwin(sternum_50_sw[[n_bestmod_sternum_50]]$Dataset)
```



Delta plot The variation in deltaAICc between time windows can be better investigated using the following plot:

```
plotdelta(  
  dataset =  
    sternum_50_sw[[n_bestmod_sternum_50]]$Dataset, arrow = TRUE  
)
```

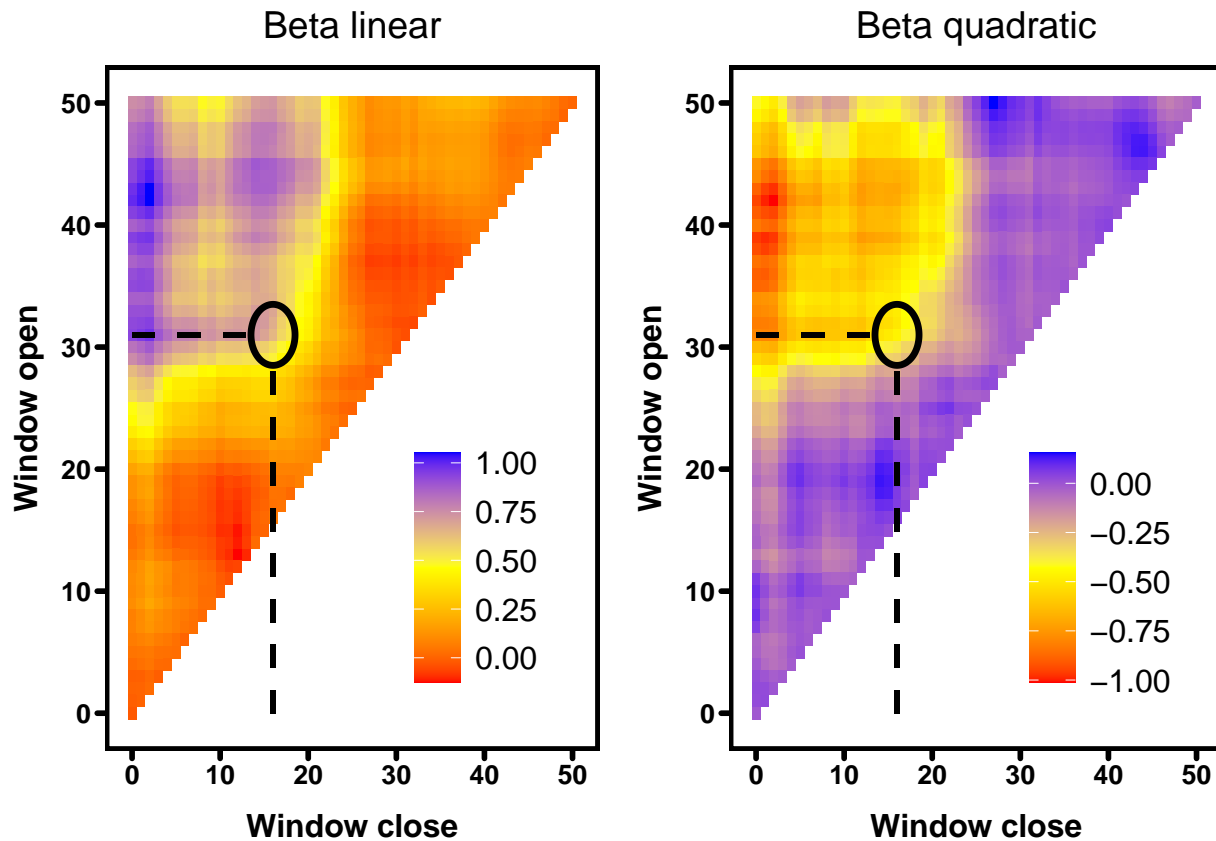


Warmer areas shows values with the lowest deltaAICc (i.e. “best models”). As explained by van de Pol et al., 2016, these deltaAICc landscapes of the different time windows shows multiple peaks (red areas) instead of a clear single peak. This can indicate the presence of multiple (e.g. possibly both long- and short-lag) weather signals within the same weather variable, but it can also occur due to collinearity or chance.

The evidence for multiple signals can be therefore investigated by adding the best supported of the weather windows to the baseline model, and re-fitting all the different time windows again: this tests whether there is still strong model support for the second best (e.g. short-lag) weather window once the other best supported (e.g. long-lag) weather window has been accounted for in the baseline model (here in the Step 2).

Beta plot This panel shows the model support (deltaAICc) for all fitted time windows tried, shown for each combination of Window open (y-axis) and Window close (x-axis). Models with the lowest deltaAICc (red) are the best supported (colours show the deltaAICc levels compared to the null model, see legend). Strongly supported windows will often be grouped together.

```
plotbetas(sternum_50_sw[[n_bestmod_sternum_50]]$Dataset, arrow = TRUE)
```



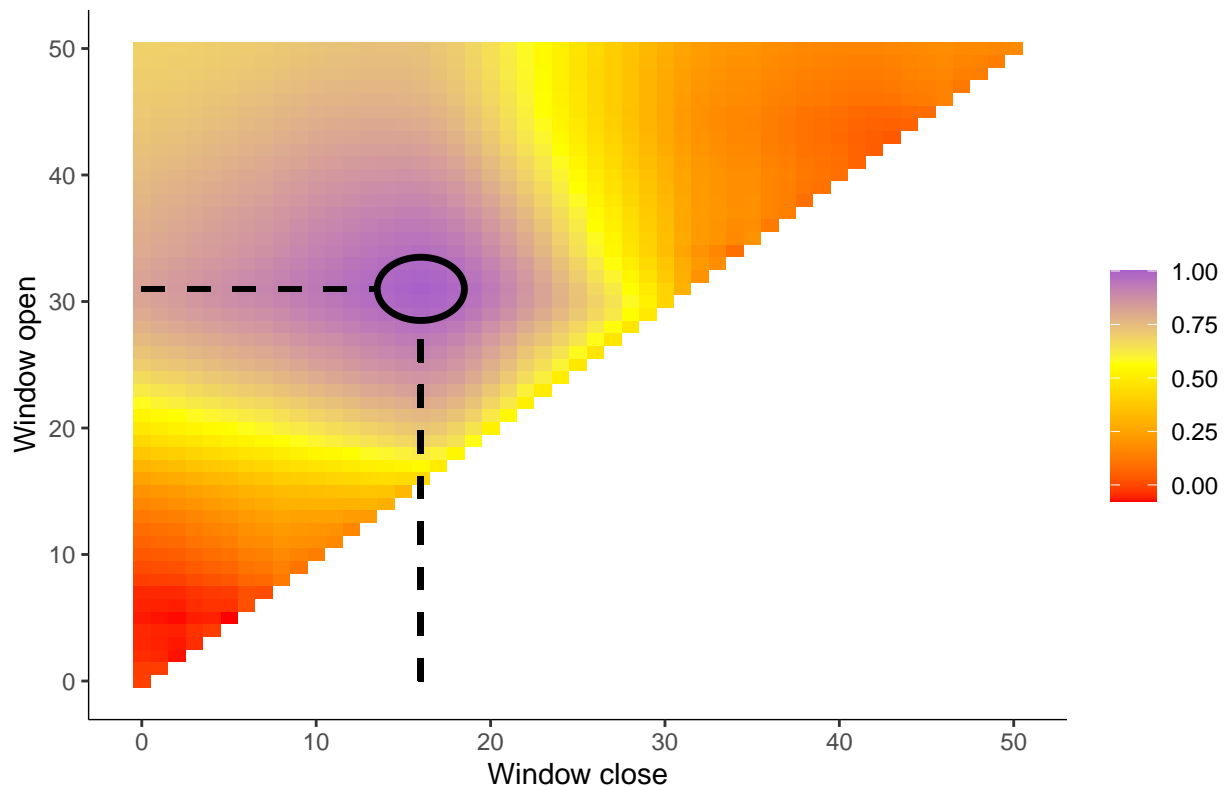
Autocollinearity Correlation between the mean temperature during the best supported time window and the mean temperature over all other time windows.

```
autocoll <- autowin(
  reference = sternum_50_sw[[n_bestmod_sternum_50]],
  baseline = sternum_50_basemod,
  xvar = list(
    tr_pcl = data_clim_sub$tr_comp_1
  ),
  type = "relative",
  range = c(50, 0),
  stat = "mean",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_sternum_50$date_50days,
  func = "quad",
  # cmissing = FALSE,
  cinterval = "day"
)

save(autocoll, file = "output/sternum_50_autocoll.rda")
```

```
load(file = "output/sternum_50_autocoll.rda")
plotcor(autocoll, type = "A", arrow = TRUE)
```

Correlation between single window and all other windows



Randwin Using `randwin` to randomize the identity of the nestling, we are able to check if the window that was found before is actually important, or the relationship was just random.

```
# Performing randomization to identify  
# likelihood of of signals occurring by chance
```

```
sternum_50_rand1000 <- randwin(  
  repeats = 1000,  
  baseline = sternum_50_basemod,  
  xvar = list(tr_pcl = data_clim_sub$str_comp_1),  
  stat = c("mean"),  
  func = c("quad"),  
  type = "relative", # relative to the individual  
  range = c(50, 0),  
  cinterval = "day",  
  cdate = data_clim_sub$date,  
  bdate = data_am_nestlings_sternum_50$date_50days,  
  cmissing = FALSE  
)  
save(sternum_50_rand1000, file = "output/sternum_50_rand1000.rda")
```

```
load("output/sternum_50_rand1000.rda")
```

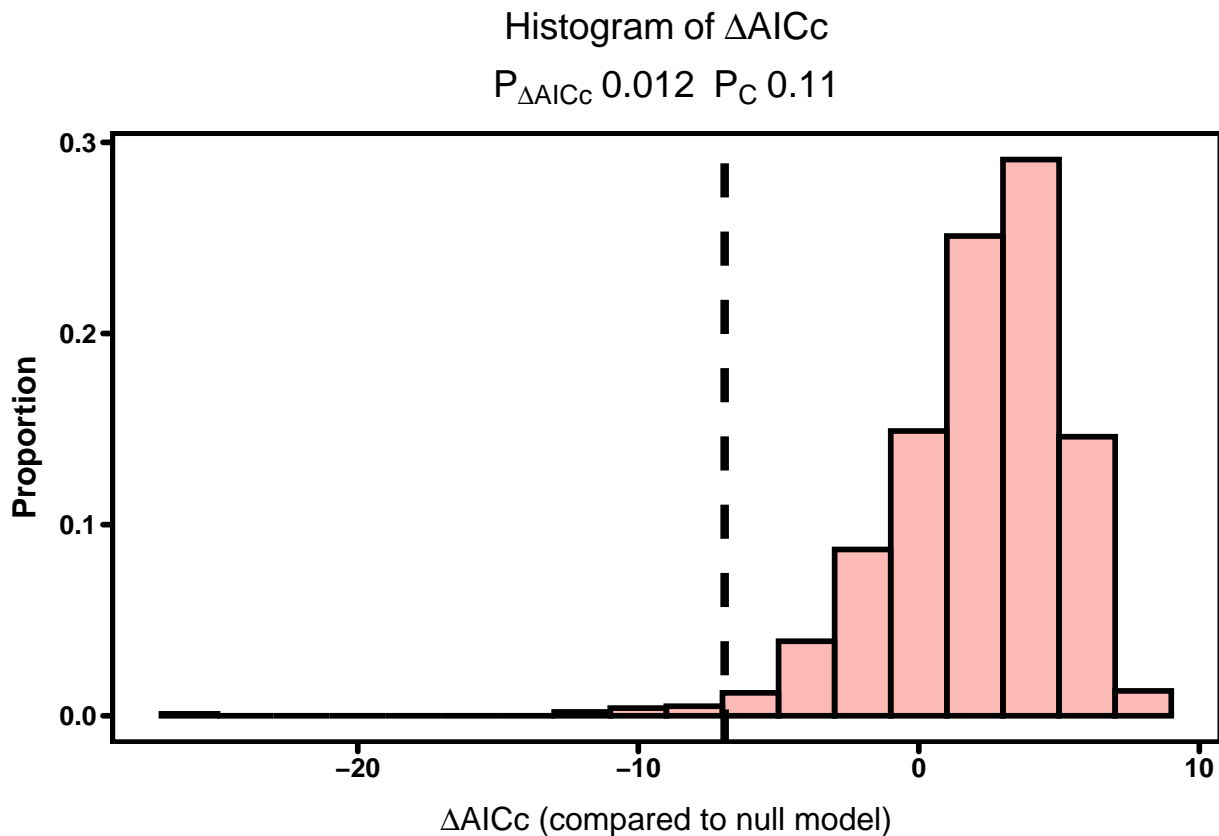
```
climwin::pvalue(  
  datasetrand = sternum_50_rand1000[[1]],  
  dataset = sternum_50_sw[[n_bestmod_sternum_50]]$Dataset,
```

```
metric = "AIC",
sample.size = sternum_50_sw[[n_bestmod_sternum_50]]$Dataset$sample.size[1]
)
```

```
## [1] 0.012
```

This panel shows a histogram of all $\Delta AICc$ values from all models run on the randomized (in grey) and the $\Delta AICc$ of the best model fitted on the observed data (dashed line). The best climate window in the randomized data is by definition a false positive, as the randomization procedure removes any climate signal from the data. The percentage of randomizations that generates a $\Delta AICc$ value that is at least as low as the $\Delta AICc$ value of the best model fitted to the observed data (P value above) is a measure of how likely it is that one obtained the observed $\Delta AICc$ by chance and thus whether the signal is likely to be real or not. In our case we can say that the candidate signal is very unlikely to be a false positive: $P < 0.05$.

```
climwin::plothist(
  datasetrand = sternum_50_rand1000[[1]],
  dataset = sternum_50_sw[[n_bestmod_sternum_50]]$Dataset
)
```



Extracting the best window The best window was obtained by averaging the opening and closing dates for all the best models ($\Delta AICc < 2$). The best window opened 38.5 (mean; SD: 6.5) and closed 15.79 (mean; SD: 0.89) days before the 50 days measurement. Which corresponds to the age of the nestling of 12 and 34 days old.

```
sternum_50_bestsw <- singlewin(
  baseline = sternum_50_basemod,
  xvar = list(
    tr_pc1 = data_clim_sub$tr_comp_1
  ),
  stat = c("mean"),
  func = c("quad"),
  type = "relative",
  range = c(
    round(mean(sternum_50_bestmod$WindowOpen), 0),
    round(mean(sternum_50_bestmod$WindowClose), 0)
  ),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_sternum_50$date_50days
)
```

```
## fixed-effect model matrix is rank deficient so dropping 2 columns / coefficients
```

```
# adding the climatic variable to the sternum_50 dataset
data_am_nestlings_sternum_50$signal_for_sternum_50 <-
  sternum_50_bestsw$BestModelData$climate
```

Body mass at 50 days The linear+quadratic term for the mean temperature better explains the variation in the data (deltaAICc has the lowest value), sorted by deltaAICc such that the best supported model is on top.

```
mass_50_mod_combos <- (mass_50_sw$combos)
mass_50_mod_combos[order(mass_50_mod_combos$DeltaAICc), ]
```

```
##  response  climate    type stat func DeltaAICc WindowOpen WindowClose
## 6  mass_50 temp_mean relative mean quad   -64.27         2         0
## 5  mass_50   tr_pc1 relative mean quad   -56.32         4         0
## 3  mass_50 rain_mean relative mean  lin   -51.49         9         0
## 7  mass_50 rain_mean relative mean quad   -46.89         9         0
## 8  mass_50 wind_mean relative mean quad   -40.15        45         8
## 1  mass_50   tr_pc1 relative mean  lin   -39.94         4         0
## 2  mass_50 temp_mean relative mean  lin   -35.18         1         0
## 4  mass_50 wind_mean relative mean  lin   -18.87        45        40
```

```
n_bestmod_mass_50 <- as.numeric(rownames(
  mass_50_mod_combos[order(mass_50_mod_combos$DeltaAICc), ]
))[1]
```

The best quadratic models The 30 best windows for the linear+quadratic models sorted by deltaAICc. To investigate any other tested hypothesis we can simply replace the number on the left or in the double square brackets with the corresponding list number.

```
head(mass_50_sw[[n_bestmod_mass_50]]$Dataset, 50)
```

##	deltaAICc	WindowOpen	WindowClose	ModelBeta	Std.Error	ModelBetaQ	Std.ErrorQ	ModelBetaC
## 6	-64.265745	2	0	8.0014807	0.3184047	-0.192959345	0.4563156	NA
## 3	-57.417441	1	0	5.9146946	0.3188554	-0.139728688	0.4574132	NA
## 2	-48.801874	1	1	5.2562216	0.3202113	-0.124370013	0.4592101	NA
## 5	-45.503437	2	1	6.5996671	0.3208174	-0.159444724	0.4595184	NA
## 1	-44.871127	0	0	4.5530382	0.3198698	-0.105871770	0.4591271	NA
## 10	-44.739372	3	0	8.0230803	0.3208091	-0.193644153	0.4599293	NA
## 121	-35.271271	15	15	-5.7236610	0.3219224	0.145600952	0.4689043	NA
## 122	-28.813650	15	14	-5.9096382	0.3225436	0.150690114	0.4691278	NA
## 15	-24.884130	4	0	6.7699591	0.3233282	-0.161733539	0.4642305	NA
## 139	-24.053360	16	14	-5.9596517	0.3232328	0.150508352	0.4713939	NA
## 9	-23.215307	3	1	5.9924116	0.3234409	-0.144702742	0.4639332	NA
## 138	-18.398435	16	15	-4.6501193	0.3239311	0.116241153	0.4718085	NA
## 4	-16.345357	2	2	4.2267672	0.3241902	-0.101840063	0.4655616	NA
## 157	-15.395147	17	14	-5.4872152	0.3243518	0.138185591	0.4732694	NA
## 140	-15.375912	16	13	-5.6176560	0.3240217	0.140428808	0.4723567	NA
## 1279	-12.905738	50	47	1.3678827	0.3240612	-0.049829258	0.4681603	NA
## 123	-12.371086	15	13	-4.9925901	0.3242315	0.125637616	0.4710788	NA
## 1278	-12.213769	50	48	1.4118977	0.3240720	-0.050341022	0.4678141	NA
## 106	-12.203046	14	14	-4.0960462	0.3242509	0.104723174	0.4697100	NA
## 1226	-12.191126	49	49	1.4532133	0.3240324	-0.050132671	0.4676623	NA
## 1280	-12.124291	50	46	1.2680288	0.3243788	-0.047418607	0.4687205	NA
## 21	-12.027305	5	0	5.7528748	0.3249174	-0.137258129	0.4679125	NA
## 1277	-11.966925	50	49	0.9682711	0.3241553	-0.037986143	0.4677597	NA
## 158	-11.823227	17	13	-5.5029344	0.3246075	0.137192885	0.4738899	NA
## 1281	-11.015150	50	45	0.8550540	0.3247216	-0.036783970	0.4691790	NA
## 176	-9.953076	18	14	-5.3554141	0.3250558	0.135516343	0.4741460	NA
## 177	-9.322745	18	13	-5.6945928	0.3250141	0.142770026	0.4746506	NA
## 1227	-9.007762	49	48	1.5543038	0.3244950	-0.052383371	0.4684170	NA
## 1276	-8.385601	50	50	0.4477120	0.3247501	-0.022686123	0.4684220	NA
## 156	-8.291613	17	15	-3.9568110	0.3251558	0.098331201	0.4731107	NA
## 1228	-8.140613	49	47	1.2620144	0.3246275	-0.044991248	0.4690207	NA
## 1282	-7.075916	50	44	0.2410714	0.3254830	-0.019691003	0.4700414	NA
## 1229	-7.037589	49	46	1.0705981	0.3249336	-0.040124275	0.4696200	NA
## 14	-6.862556	4	1	4.3699728	0.3255317	-0.103658302	0.4682194	NA
## 1230	-6.046203	49	45	0.5746021	0.3252348	-0.027306799	0.4700261	NA
## 1284	-5.702757	50	42	-0.9928712	0.3262797	0.013024044	0.4704723	NA
## 1283	-5.603272	50	43	-0.4015480	0.3259940	-0.002348656	0.4703643	NA
## 28	-5.570170	6	0	5.1919576	0.3257501	-0.124269637	0.4703250	NA
## 1285	-5.550433	50	41	-1.0187081	0.3264131	0.013189168	0.4706785	NA
## 1288	-5.370280	50	38	-4.3638939	0.3267350	0.102250124	0.4707039	NA
## 141	-5.349017	16	12	-4.5146059	0.3252159	0.111040645	0.4732118	NA
## 178	-5.335599	18	12	-5.2555336	0.3254513	0.130259952	0.4748469	NA
## 159	-5.333387	17	12	-4.6999299	0.3253386	0.115365050	0.4742245	NA
## 1287	-4.739701	50	39	-3.3248530	0.3266830	0.074979745	0.4707444	NA
## 197	-4.672331	19	13	-5.5028451	0.3256010	0.138793708	0.4753384	NA
## 1291	-4.380901	50	35	-6.3747139	0.3268649	0.155041525	0.4717930	NA
## 1286	-4.371365	50	40	-2.0213878	0.3266735	0.040255241	0.4707860	NA
## 1138	-4.186436	47	38	-5.9523265	0.3268705	0.149630331	0.4704765	NA
## 1290	-4.043183	50	36	-5.2365932	0.3269496	0.125004082	0.4714795	NA
## 965	-3.974394	43	25	13.3421156	0.3259410	-0.339959105	0.4743405	NA

##	ModelInt	Function	Furthest	Closest	Statistics	Type K	ModWeight	sample.size
## 6	18.90363	quad	50	0	mean relative	0	9.678487e-01	25
## 3	39.06261	quad	50	0	mean relative	0	3.152951e-02	25
## 2	45.93467	quad	50	0	mean relative	0	4.244931e-04	25
## 5	33.21668	quad	50	0	mean relative	0	8.158757e-05	25
## 1	52.53170	quad	50	0	mean relative	0	5.947297e-05	25
## 10	18.60605	quad	50	0	mean relative	0	5.568130e-05	25
## 121	154.78937	quad	50	0	mean relative	0	4.894829e-07	25
## 122	156.45590	quad	50	0	mean relative	0	1.938576e-08	25
## 15	30.62282	quad	50	0	mean relative	0	2.717680e-09	25
## 139	157.66145	quad	50	0	mean relative	0	1.793903e-09	25
## 9	39.20231	quad	50	0	mean relative	0	1.179827e-09	25
## 138	145.37933	quad	50	0	mean relative	0	1.061323e-10	25
## 4	57.08835	quad	50	0	mean relative	0	3.802134e-11	25
## 157	153.37041	quad	50	0	mean relative	0	2.364243e-11	25
## 140	154.98822	quad	50	0	mean relative	0	2.341613e-11	25
## 1279	91.67474	quad	50	0	mean relative	0	6.809631e-12	25
## 123	148.43010	quad	50	0	mean relative	0	5.212252e-12	25
## 1278	91.05801	quad	50	0	mean relative	0	4.817976e-12	25
## 106	138.79694	quad	50	0	mean relative	0	4.792212e-12	25
## 1226	90.34165	quad	50	0	mean relative	0	4.763736e-12	25
## 1280	92.66743	quad	50	0	mean relative	0	4.607173e-12	25
## 21	40.87098	quad	50	0	mean relative	0	4.389089e-12	25
## 1277	94.90347	quad	50	0	mean relative	0	4.258562e-12	25
## 158	154.11470	quad	50	0	mean relative	0	3.963323e-12	25
## 1281	96.53364	quad	50	0	mean relative	0	2.645987e-12	25
## 176	151.94715	quad	50	0	mean relative	0	1.555827e-12	25
## 177	155.80137	quad	50	0	mean relative	0	1.135237e-12	25
## 1227	89.25976	quad	50	0	mean relative	0	9.698152e-13	25
## 1276	99.11526	quad	50	0	mean relative	0	7.105399e-13	25
## 156	138.91248	quad	50	0	mean relative	0	6.779210e-13	25
## 1228	92.04857	quad	50	0	mean relative	0	6.286225e-13	25
## 1282	101.88165	quad	50	0	mean relative	0	3.691425e-13	25
## 1229	93.86256	quad	50	0	mean relative	0	3.621357e-13	25
## 14	54.90272	quad	50	0	mean relative	0	3.317902e-13	25
## 1230	98.48596	quad	50	0	mean relative	0	2.205944e-13	25
## 1284	113.25103	quad	50	0	mean relative	0	1.857874e-13	25
## 1283	107.68886	quad	50	0	mean relative	0	1.767719e-13	25
## 28	46.68107	quad	50	0	mean relative	0	1.738703e-13	25
## 1285	113.69355	quad	50	0	mean relative	0	1.721628e-13	25
## 1288	144.70749	quad	50	0	mean relative	0	1.573329e-13	25
## 141	144.93189	quad	50	0	mean relative	0	1.556691e-13	25
## 178	152.11223	quad	50	0	mean relative	0	1.546283e-13	25
## 159	146.94491	quad	50	0	mean relative	0	1.544573e-13	25
## 1287	134.91394	quad	50	0	mean relative	0	1.147866e-13	25
## 197	153.69299	quad	50	0	mean relative	0	1.109844e-13	25
## 1291	163.79464	quad	50	0	mean relative	0	9.593540e-14	25
## 1286	122.83998	quad	50	0	mean relative	0	9.547908e-14	25
## 1138	157.76346	quad	50	0	mean relative	0	8.704648e-14	25
## 1290	153.07979	quad	50	0	mean relative	0	8.102974e-14	25
## 965	-29.87635	quad	50	0	mean relative	0	7.829012e-14	25
##	Randomised							
## 6		no						
## 3		no						

## 2	no
## 5	no
## 1	no
## 10	no
## 121	no
## 122	no
## 15	no
## 139	no
## 9	no
## 138	no
## 4	no
## 157	no
## 140	no
## 1279	no
## 123	no
## 1278	no
## 106	no
## 1226	no
## 1280	no
## 21	no
## 1277	no
## 158	no
## 1281	no
## 176	no
## 177	no
## 1227	no
## 1276	no
## 156	no
## 1228	no
## 1282	no
## 1229	no
## 14	no
## 1230	no
## 1284	no
## 1283	no
## 28	no
## 1285	no
## 1288	no
## 141	no
## 178	no
## 159	no
## 1287	no
## 197	no
## 1291	no
## 1286	no
## 1138	no
## 1290	no
## 965	no

The best windows (deltaAICc difference from the best model < 2) for the linear+quadratic models sorted by deltaAICc. There is only one best window:

```

mass_50_bestmod <- subset(
  mass_50_sw[[n_bestmod_mass_50]]$Dataset,
  deltaAICc < mass_50_sw[[n_bestmod_mass_50]]$Dataset$deltaAICc[1] + 2
)

mass_50_bestmod

```

```

##   deltaAICc WindowOpen WindowClose ModelBeta Std.Error ModelBetaQ Std.ErrorQ ModelBetaC ModelInt
## 6 -64.26574         2         0 8.001481 0.3184047 -0.1929593 0.4563156          NA 18.90363
##   Function Furthest Closest Statistics      Type K ModWeight sample.size Randomised
## 6    quad      50      0    mean relative 0 0.9678487          25          no

```

As it can be seen here, all the other models have a deltaAICc difference from the best model much larger than 2.

```
head(mass_50_sw[[n_bestmod_mass_50]]$Dataset)
```

```

##   deltaAICc WindowOpen WindowClose ModelBeta Std.Error ModelBetaQ Std.ErrorQ ModelBetaC
## 6 -64.26574         2         0 8.001481 0.3184047 -0.1929593 0.4563156          NA
## 3 -57.41744         1         0 5.914695 0.3188554 -0.1397287 0.4574132          NA
## 2 -48.80187         1         1 5.256222 0.3202113 -0.1243700 0.4592101          NA
## 5 -45.50344         2         1 6.599667 0.3208174 -0.1594447 0.4595184          NA
## 1 -44.87113         0         0 4.553038 0.3198698 -0.1058718 0.4591271          NA
## 10 -44.73937        3         0 8.023080 0.3208091 -0.1936442 0.4599293          NA
##   ModelInt Function Furthest Closest Statistics      Type K   ModWeight sample.size Randomised
## 6 18.90363    quad      50      0    mean relative 0 9.678487e-01          25          no
## 3 39.06261    quad      50      0    mean relative 0 3.152951e-02          25          no
## 2 45.93467    quad      50      0    mean relative 0 4.244931e-04          25          no
## 5 33.21668    quad      50      0    mean relative 0 8.158757e-05          25          no
## 1 52.53170    quad      50      0    mean relative 0 5.947297e-05          25          no
## 10 18.60605    quad      50      0    mean relative 0 5.568130e-05          25          no

```

Windows plot It's possible to extract the time windows of all the best supported models (i.e. multi-model inference). This panel shows the opening and closing points of the time windows that were best supported by the data, here those models that made up 95% model confidence set.

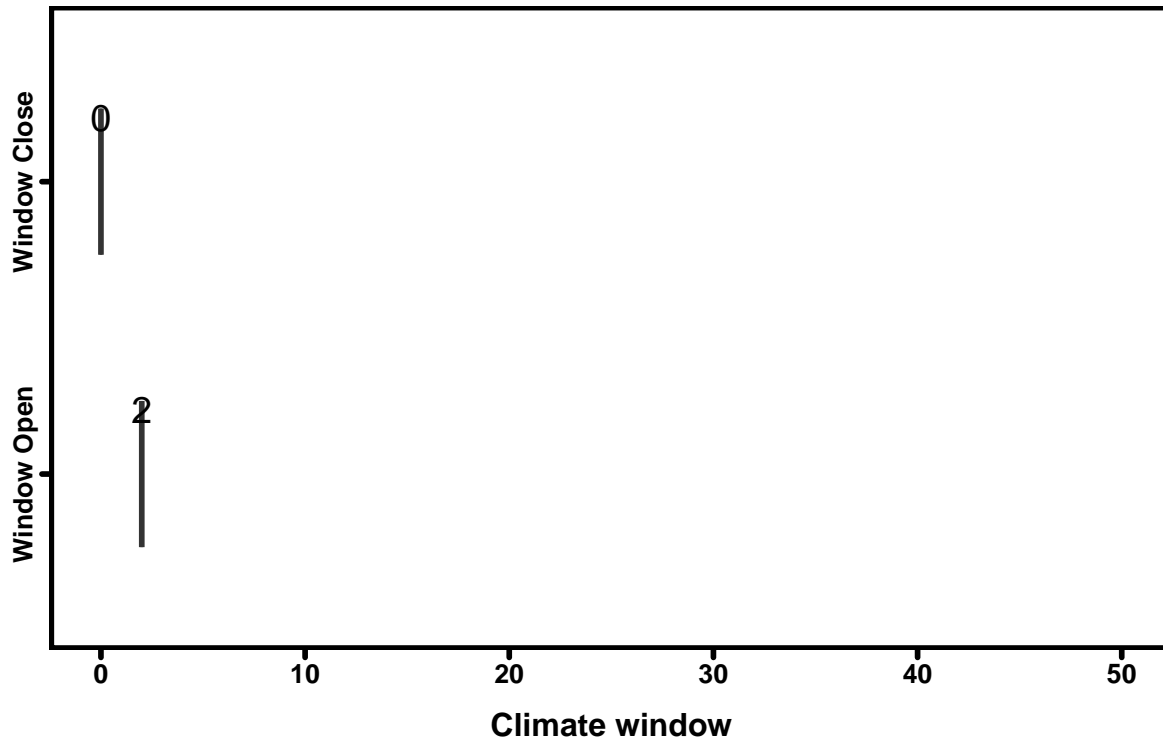
```
plotwin(mass_50_sw[[n_bestmod_mass_50]]$Dataset)
```

```

## Warning in plotwin(mass_50_sw[[n_bestmod_mass_50]]$Dataset): Top window has a weight greater
## than 0.95. Plotting single best window only.

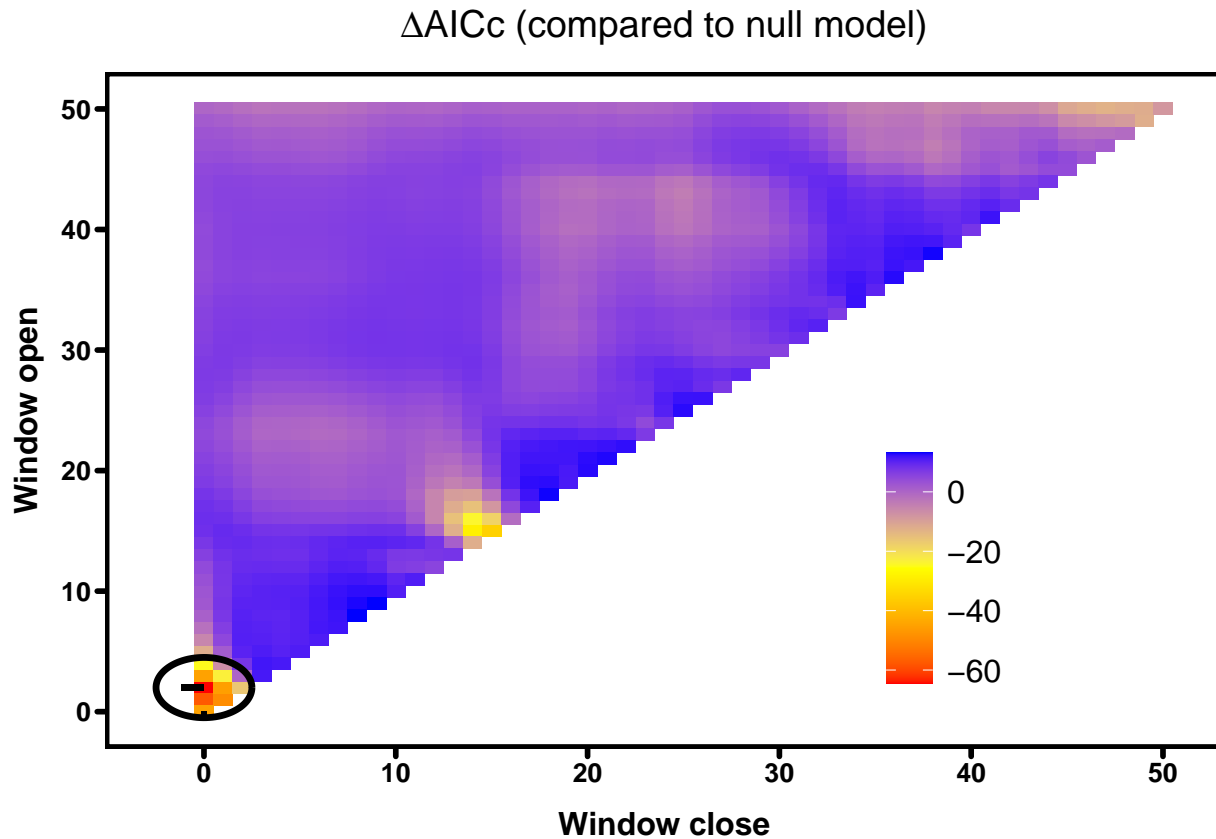
```

Climate window range for 95 % confidence set



Delta plot The variation in deltaAICc between time windows can be better investigated using the following plot:

```
plotdelta(  
  dataset =  
    mass_50_sw[[n_bestmod_mass_50]]$Dataset, arrow = TRUE  
)
```

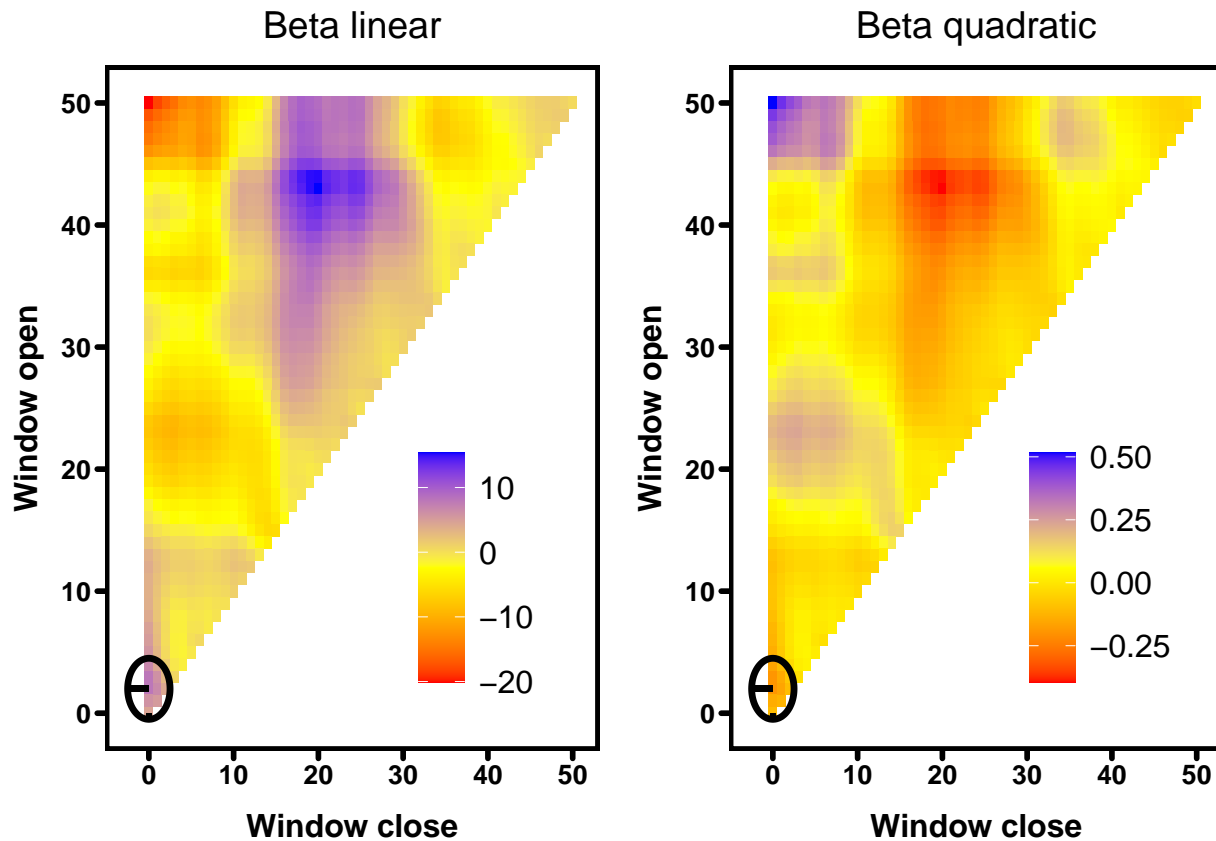


Warmer areas shows values with the lowest deltaAICc (i.e. “best models”). As explained by van de Pol et al., 2016, these deltaAICc landscapes of the different time windows shows multiple peaks (red areas) instead of a clear single peak. This can indicate the presence of multiple (e.g. possibly both long- and short-lag) weather signals within the same weather variable, but it can also occur due to collinearity or chance.

The evidence for multiple signals can be therefore investigated by adding the best supported of the weather windows to the baseline model, and re-fitting all the different time windows again: this tests whether there is still strong model support for the second best (e.g. short-lag) weather window once the other best supported (e.g. long-lag) weather window has been accounted for in the baseline model (here in the Step 2).

Beta plot This panel shows the model support (deltaAICc) for all fitted time windows tried, shown for each combination of Window open (y-axis) and Window close (x-axis). Models with the lowest deltaAICc (red) are the best supported (colours show the deltaAICc levels compared to the null model, see legend). Strongly supported windows will often be grouped together.

```
plotbetas(mass_50_sw[[n_bestmod_mass_50]]$Dataset, arrow = TRUE)
```



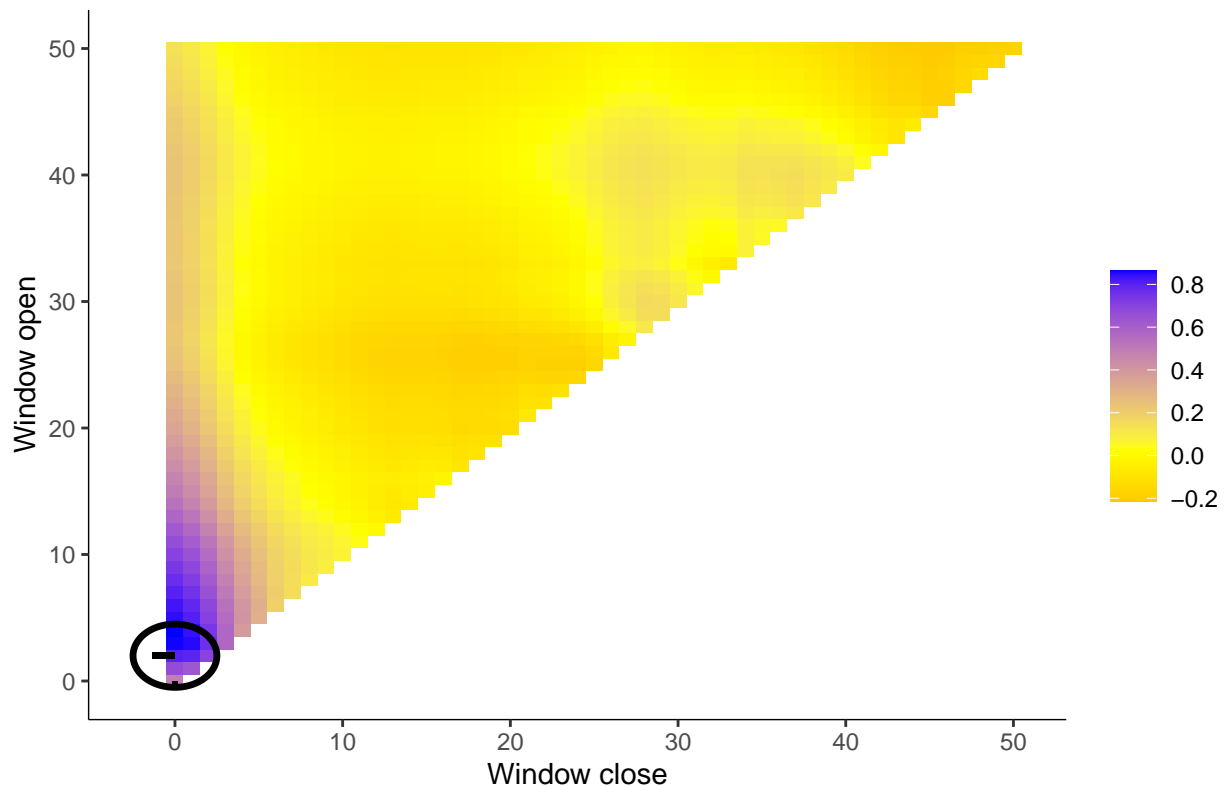
Autocollinearity Correlation between the mean temperature during the best supported time window and the mean temperature over all other time windows.

```
autocoll <- autowin(
  reference = mass_50_sw[[n_bestmod_mass_50]],
  baseline = mass_50_basemod,
  xvar = list(
    tr_pcl = data_clim_sub$tr_comp_1
  ),
  type = "relative",
  range = c(50, 0),
  stat = "mean",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_mass_50$date_50days,
  func = "quad",
  # cmissing = FALSE,
  cinterval = "day"
)

save(autocoll, file = "output/mass_50_autocoll.rda")
```

```
load(file = "output/mass_50_autocoll.rda")
plotcor(autocoll, type = "A", arrow = TRUE)
```

Correlation between single window and all other windows



Randwin Using `randwin` to randomize the identity of the nestling, we are able to check if the window that was found before is actually important, or the relationship was just random.

```
# Performing randomization to identify
# likelihood of of signals occurring by chance

mass_50_rand1000 <- randwin(
  repeats = 1000,
  baseline = mass_50_basemod,
  xvar = list(temp_mean = data_clim_sub$T.daily.mean),
  stat = c("mean"),
  func = c("quad"),
  type = "relative", # relative to the individual
  range = c(50, 0),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_mass_50$date_50days,
  cmissing = FALSE
)
save(mass_50_rand1000, file = "output/mass_50_rand1000.rda")

load("output/mass_50_rand1000.rda")

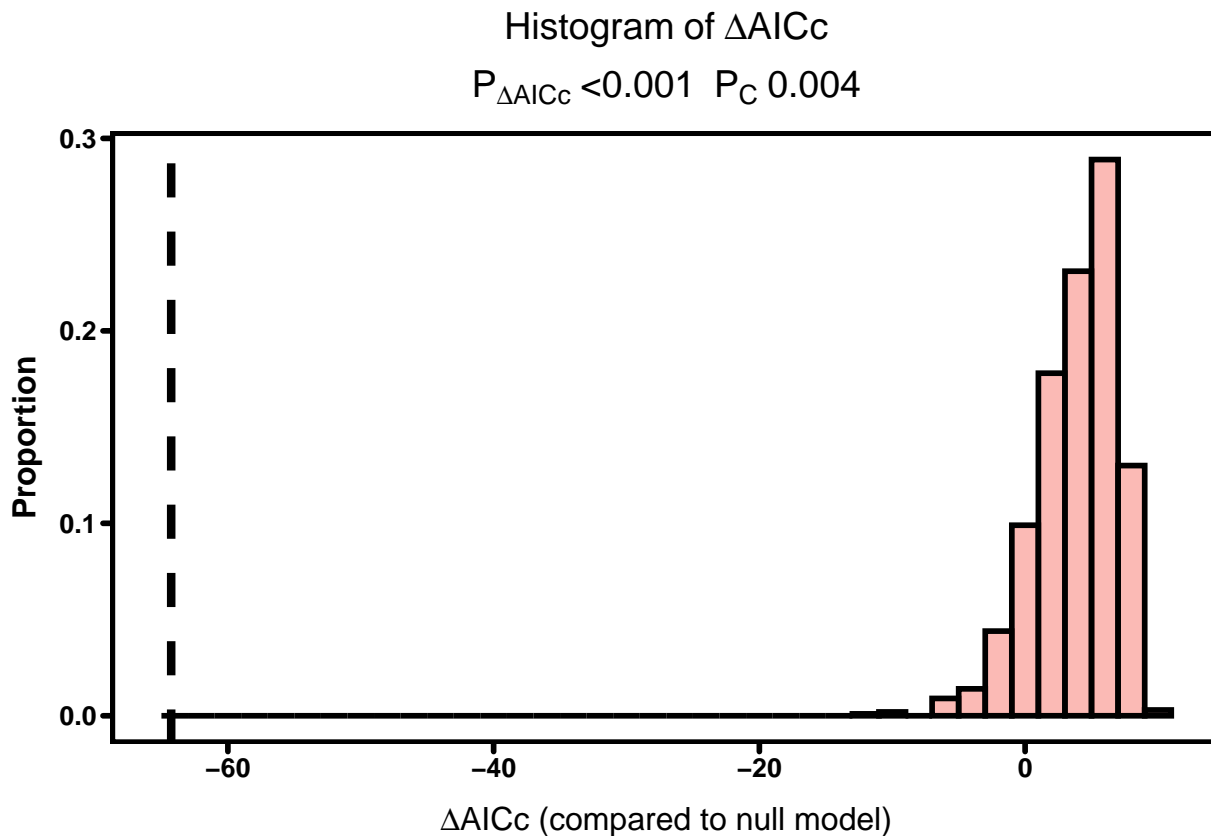
climwin::pvalue(
  datasetrand = mass_50_rand1000[[1]],
  dataset = mass_50_sw[[n_bestmod_mass_50]]$Dataset,
```

```
metric = "AIC",
sample.size = mass_50_sw[[n_bestmod_mass_50]]$Dataset$sample.size[1]
)
```

```
## [1] "<0.001"
```

This panel shows a histogram of all $\Delta AICc$ values from all models run on the randomized (in grey) and the $\Delta AICc$ of the best model fitted on the observed data (dashed line). The best climate window in the randomized data is by definition a false positive, as the randomization procedure removes any climate signal from the data. The percentage of randomizations that generates a $\Delta AICc$ value that is at least as low as the $\Delta AICc$ value of the best model fitted to the observed data (P value above) is a measure of how likely it is that one obtained the observed $\Delta AICc$ by chance and thus whether the signal is likely to be real or not. In our case we can say that the candidate signal is very unlikely to be a false positive: $P < 0.001$.

```
climwin::plothist(
  datasetrand = mass_50_rand1000[[1]],
  dataset = mass_50_sw[[n_bestmod_mass_50]]$Dataset
)
```



Extracting the best window The best window was obtained by averaging the opening and closing dates for all the best models ($\Delta AICc < 2$). In the case of the body mass there was only one top model and the window opened 2 and closed 0 days before the 50 days measurement, which corresponds to the age of the nestling of 46 and 50 days old.


```

mass_50_bestsw <- singlewin(
  baseline = mass_50_basemod,
  xvar = list(
    # tr_pc1 = data_clim_sub$str_comp_1
    temp_mean = data_clim_sub$T.daily.mean
  ),
  stat = c("mean"),
  func = c("quad"),
  type = "relative",
  range = c(
    round(mean(mass_50_bestmod$WindowOpen), 0),
    round(mean(mass_50_bestmod$WindowClose), 0)
  ),
  cinterval = "day",
  cdate = data_clim_sub$date,
  bdate = data_am_nestlings_mass_50$date_50days
)

```

```
## fixed-effect model matrix is rank deficient so dropping 2 columns / coefficients
```

```

# adding the climatic variable to the mass_50 dataset
data_am_nestlings_mass_50$signal_for_mass_50 <-
  mass_50_bestsw$BestModelData$climate

```

Adding the variables to the dataset

Extracting values for the best variable within the best window for each trait for each individual so I can use them in a glmm.

```

am_nestlings_clim <- Reduce(function(x, y) merge(x, y, all = TRUE), list(
  data_am_nestlings_wing_50,
  data_am_nestlings_sternum_50,
  data_am_nestlings_mass_50
))

```

Climatic models, quadratic effects are kept even if not significant, if present in the best model suggested by climwin.

```

wing_50_climmod <- lmer(
  wing_50 ~
    brood_size + colony +
    hatch_doy_sc + age_days_sc50 +
    signal_for_wing_50 +
    I(signal_for_wing_50^2) +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_wing_50
)

```

```

sternum_50_climmod <- lmer(
  sternum_50 ~

```

```

    brood_size + colony +
    hatch_doy_sc + age_days_sc50 +
    signal_for_sternum_50 +
    I(signal_for_sternum_50^2) +
    (1 | year_f) + (1 | nestcode_rearing),
    control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
    data = data_am_nestlings_sternum_50
)

mass_50_climmod <- lmer(
  mass_50 ~
    brood_size + colony +
    hatch_doy_sc + age_days_sc50 +
    signal_for_mass_50 +
    I(signal_for_mass_50^2) +
    (1 | year_f) + (1 | nestcode_rearing),
    control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
    data = data_am_nestlings_mass_50
)

```

```

labels <- c(
  `(Intercept)` = "Intercept",
  hatch_doy_sc = "Hatching day",
  brood_size = "Hatching brood size",
  age_days_sc50 = "Age (days)",
  colonySolothurn = "Colony [Solothurn]",
  signal_for_wing_50 = "PC1 (age: 1-48)",
  `I(signal_for_wing_50^2)` = "PC1 (age: 1-48) 2",
  signal_for_sternum_50 = "PC1 (age: 12-34)",
  `I(signal_for_sternum_50^2)` = "PC1 (age: 12-34) 2",
  signal_for_mass_50 = "Temp (age: 48-50)",
  `I(signal_for_mass_50^2)` = "Temp (age: 48-50) 2"
)

sjPlot::tab_model(
  wing_50_climmod,
  sternum_50_climmod,
  mass_50_climmod,
  file = "output/Tab_wicm_models_50.doc",
  string.est = "Estimate",
  string.se = "SE",
  show.ci = FALSE,
  show.se = TRUE,
  show.stat = FALSE,
  show.df = FALSE,
  pred.labels = labels
)

```

Figure

```

cowplot::plot_grid(
  fun_climres(wing_50_climmod, data_am_nestlings_wing_50),

```

```

fun_climres(sternum_50_climmod, data_am_nestlings_sternum_50),
fun_climres(mass_50_climmod, data_am_nestlings_mass_50),
ncol = 1, align = c("v", "h")
)

```

```

# Save the plot to the figures directory
ggsave("figures/Fig. 2 - traits-weather.png", width = 4, height = 8, dpi = 300)

```

Testing for segments

```

sternum_50_climlme <- lme(
  sternum_50 ~
    signal_for_sternum_50 +
    # I(signal_for_sternum_50^2) +
    brood_size + colony +
    hatch_doy_sc + age_days_sc50,
  random = list(~ 1 | year_f, ~ 1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_sternum_50
)

sternum_50_climmod_segmented <- segmented(sternum_50_climlme,
  seg.Z = ~signal_for_sternum_50, psi = 0.4,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_sternum_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_sternum_50 + U)
  )
)
save(sternum_50_climmod_segmented,
  file = "output/sternum_50_climmod_segmented_04.rda"
)

sternum_50_climmod_segmented <- segmented(sternum_50_climlme,
  seg.Z = ~signal_for_sternum_50, psi = 0,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_sternum_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_sternum_50 + U)
  )
)
save(sternum_50_climmod_segmented,
  file = "output/sternum_50_climmod_segmented_0.rda"
)

sternum_50_climmod_segmented <- segmented(sternum_50_climlme,
  seg.Z = ~signal_for_sternum_50, psi = 0.8,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_sternum_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_sternum_50 + U)
  )
)

```

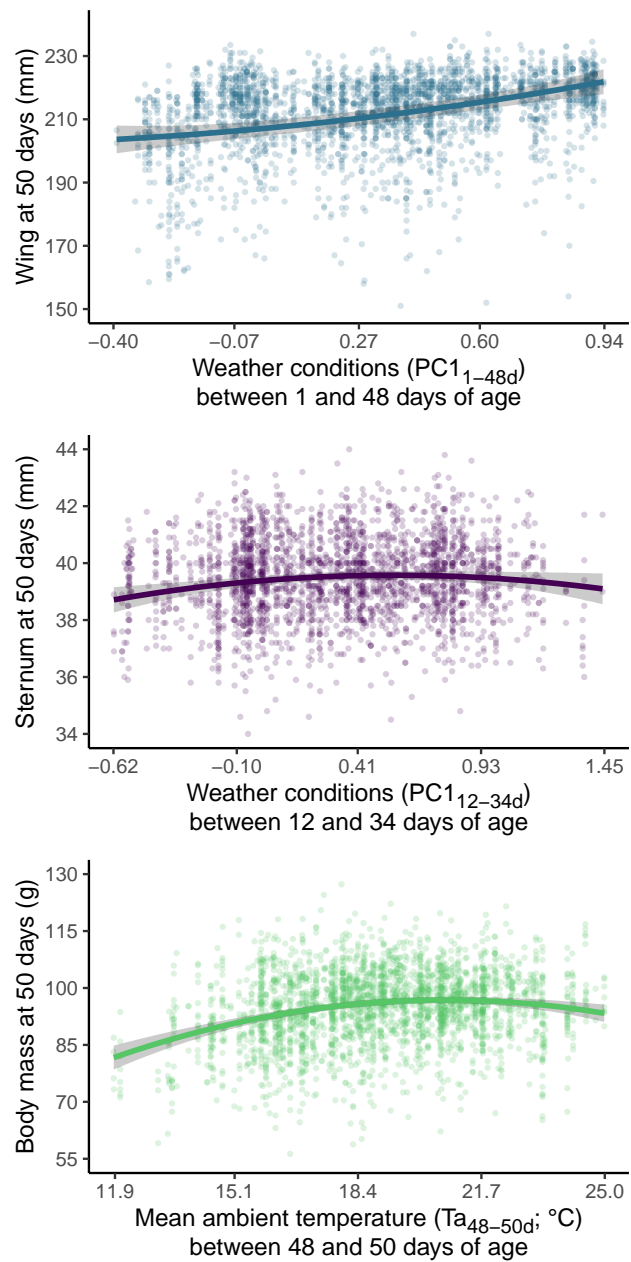


Figure 3: Fig. 2 - Variation in wing length, sternum length and body mass of 50-day-old nestling Alpine swifts in relation to weather conditions encountered earlier in their development. Weather conditions resulted from a principal component analysis (PC1) between daily average temperature and daily rain, and traits were affected by weather conditions over different time windows. Low PC1 values indicate cold and rainy days, and high values indicate sunny and warm days. Solid lines (and 95% confidence intervals) are predictions from the models presented in Table 2. Climatic windows are reported using the nestling's age as a reference.

```
save(sternum_50_climmod_segmented,
     file = "output/sternum_50_climmod_segmented_08.rda"
)
```

Sternum length Summary of the segmented analysis

psi = 0

```
load("output/sternum_50_climmod_segmented_0.rda")
summary(sternum_50_climmod_segmented)
```

```
## Segmented mixed-effects model fit by REML
##      AIC      BIC   logLik
## 8498.841 8585.837 -4234.42
## Bootstrap restarting on 10 samples; 3 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_sternum_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_sternum_50          U
## StdDev:  0.2020344          0.3586896 0.0001815599
##
## Formula: ~1 + signal_for_sternum_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_sternum_50          U Residual
## StdDev:  0.5877018          0.1656474 0.03823937  1.23168
##
## Fixed effects:
##              Value Std.Error   DF t-value p-value
## (Intercept)  41.38751  0.84875 1252  48.763  0.000
## bsh_ax      -0.34937  0.04905 1166  -7.122  0.000
## colonySolothurn  0.17642  0.06835 1166   2.581  0.010
## hatch_doy_sc   -0.22559  0.04076 1252  -5.535  0.000
## age_days_sc50  0.08825  0.02050 1252   4.305  0.000
## -- leftS:
## signal_for_sternum_50  3.42086  1.69671 1252   2.016  0.044
## -- diffS:
## U                  -3.25225  1.70482 1252  -1.908
## -- break:
## GO                  -0.37101  0.08845 1166
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -6.12336881 -0.57426371  0.01170877  0.59116152  2.82134160
##
## Number of Observations: 2447
## Number of Groups:
##              year_f nestcode_rearing %in% year_f
##              22              1191
```

```
mean(sternum_50_climmod_segmented$psi)
```

```
## [1] -0.3552971
```

```
psi = 0.4
```

```
load("output/sternum_50_climmod_segmented_04.rda")  
summary(sternum_50_climmod_segmented)
```

```
## Segmented mixed-effects model fit by REML  
##      AIC      BIC    logLik  
## 8499.856 8586.852 -4234.928  
## Bootstrap restarting on 10 samples; 3 different solution(s)  
##  
## Random effects:  
## Formula: ~1 + signal_for_sternum_50 + U | year_f  
## Structure: Diagonal  
##      (Intercept) signal_for_sternum_50      U  
## StdDev: 0.2453159      0.01219222 1.547717  
##  
## Formula: ~1 + signal_for_sternum_50 + U | nestcode_rearing %in% year_f  
## Structure: Diagonal  
##      (Intercept) signal_for_sternum_50      U Residual  
## StdDev: 0.5949472      1.927089e-05 7.278493e-09 1.231978  
##  
## Fixed effects:  
##      Value Std.Error DF t-value p-value  
## (Intercept)      40.16049 0.15477 1253 259.485 0.0000  
## bsh_ax      -0.35298 0.04909 1165 -7.190 0.0000  
## colonySolothurn      0.15740 0.06825 1165 2.306 0.0213  
## hatch_doy_sc      -0.20187 0.04117 1253 -4.903 0.0000  
## age_days_sc50      0.09706 0.02042 1253 4.752 0.0000  
## -- leftS:  
## signal_for_sternum_50 0.45642 0.15092 1253 3.024 0.0025  
## -- diffS:  
## U      -1.68317 1.25816 1165 -1.338  
## -- break:  
## GO      0.81258 0.11778 1165  
## psi.link = identity  
##  
## Standardized Within-Group Residuals:  
##      Min      Q1      Med      Q3      Max  
## -6.113264015 -0.575664046 0.009521131 0.582781653 2.815703002  
##  
## Number of Observations: 2447  
## Number of Groups:  
##      year_f nestcode_rearing %in% year_f  
##      22      1191
```

```
mean(sternum_50_climmod_segmented$psi)
```

```
## [1] 0.8377074
```

psi = 0.8

```
load("output/sternum_50_climmod_segmented_08.rda")
summary(sternum_50_climmod_segmented)
```

```
## Segmented mixed-effects model fit by REML
##      AIC      BIC    logLik
## 8499.863 8586.86 -4234.932
## Bootstrap restarting on 10 samples; 3 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_sternum_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_sternum_50      U
## StdDev: 0.2473956      0.01030922 1.330668
##
## Formula: ~1 + signal_for_sternum_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_sternum_50      U Residual
## StdDev: 0.594737      1.882626e-05 9.555731e-09 1.232142
##
## Fixed effects:
##      Value Std.Error  DF t-value p-value
## (Intercept) 40.15957 0.15500 1253 259.101 0.0000
## bsh_ax -0.35267 0.04910 1165 -7.182 0.0000
## colonySolothurn 0.15726 0.06834 1165 2.301 0.0216
## hatch_doy_sc -0.20197 0.04120 1253 -4.902 0.0000
## age_days_sc50 0.09701 0.02043 1253 4.749 0.0000
## -- leftS:
## signal_for_sternum_50 0.46091 0.15169 1253 3.038 0.0024
## -- diffS:
## U -1.60754 1.14038 1165 -1.410
## -- break:
## GO 0.87940 0.11584 1165
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -6.113739944 -0.575721368 0.009786115 0.582781417 2.815736437
##
## Number of Observations: 2447
## Number of Groups:
##      year_f nestcode_rearing %in% year_f
##      22      1191
```

```
mean(sternum_50_climmod_segmented$psi)
```

```
## [1] 0.874424
```

The threshold found can be quite different, it seems it's not possible to identify a consistent break point, thus we decided not to use the threshold approach for the sternum length.

```

mass_50_climlme <- lme(
  mass_50 ~
    signal_for_mass_50 +
    # I(signal_for_mass_50^2) +
    brood_size + colony +
    hatch_doy_sc + age_days_sc50,
  random = list(~ 1 | year_f, ~ 1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_mass_50
)

mass_50_climmod_segmented <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = 20,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)

save(mass_50_climmod_segmented, file = "output/mass_50_climmod_segmented_20.rda")

mass_50_climmod_segmented <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = 19,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)

save(mass_50_climmod_segmented, file = "output/mass_50_climmod_segmented_19.rda")

mass_50_climmod_segmented <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = 21,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)

save(mass_50_climmod_segmented, file = "output/mass_50_climmod_segmented_21.rda")

mass_50_climmod_segmented <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = 22,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)

save(mass_50_climmod_segmented, file = "output/mass_50_climmod_segmented_22.rda")

mass_50_climmod_segmented <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = 23,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),

```



```

    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)
save(mass_50_climmod_segmented, file = "output/mass_50_climmod_segmented_23.rda")

mass_50_climmod_segmented <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = 16,
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)
save(mass_50_climmod_segmented, file = "output/mass_50_climmod_segmented_16.rda")

mass_50_climmod_segmented2 <- segmented(mass_50_climlme,
  seg.Z = ~signal_for_mass_50, psi = c(17, 22),
  random = list(
    year_f = pdDiag(~ 1 + signal_for_mass_50 + U),
    nestcode_rearing = pdDiag(~ 1 + signal_for_mass_50 + U)
  )
)
save(mass_50_climmod_segmented2,
  file = "output/mass_50_climmod_segmented_2-17-22.rda"
)

```

Body mass The analysis was repeated using different starting values (ψ) to check if that was affecting the threshold found. Summary of the segmented analysis:

$\psi = 16$

```

load("output/mass_50_climmod_segmented_16.rda")
summary(mass_50_climmod_segmented)

```

```

## Segmented mixed-effects model fit by REML
##      AIC      BIC    logLik
## 19090.38 19178.82 -9530.191
## Bootstrap restarting on 10 samples; 6 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_mass_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U
## StdDev:    2.513369      0.001395197 1.266925
##
## Formula: ~1 + signal_for_mass_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U Residual
## StdDev:    5.292576      0.01473296 0.05008139 6.791907
##
## Fixed effects:
##      Value Std.Error DF t-value p-value
## (Intercept)    69.3383   3.7229 1381  18.625  0.0000
## bsh_ax         -2.4003   0.3154 1280  -7.611  0.0000

```

```

## colonySolothurn      2.5634      0.4577 1280      5.601      0.0000
## hatch_doy_sc         -1.5504      0.2994 1381     -5.179      0.0000
## age_days_sc50        0.2206      0.1344 1381      1.642      0.1009
## -- leftS:
## signal_for_mass_50   1.7136      0.2055 1381      8.338      0.0000
## -- diffS:
## U                    -2.3270      0.4935 1381     -4.715
## -- break:
## G0                   19.1590      0.2900 1381
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -3.85009835 -0.48382193  0.03584051  0.53958220  4.12865580
##
## Number of Observations: 2693
## Number of Groups:
##                year_f nestcode_rearing %in% year_f
##                25                1307

```

psi = 20

```

load("output/mass_50_climmod_segmented_20.rda")
summary(mass_50_climmod_segmented)

```

```

## Segmented mixed-effects model fit by REML
##      AIC      BIC      logLik
## 19090.37 19178.81 -9530.185
## Bootstrap restarting on 10 samples; 3 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_mass_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U
## StdDev:      2.515159      0.001408265 1.241755
##
## Formula: ~1 + signal_for_mass_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U Residual
## StdDev:      5.292365      0.01483897 0.04884255 6.792138
##
## Fixed effects:
##      Value Std.Error  DF t-value p-value
## (Intercept) 69.0893  3.7867 1381 18.245 0.0000
## bsh_ax      -2.3988  0.3154 1280 -7.606 0.0000
## colonySolothurn 2.5624  0.4577 1280  5.598 0.0000
## hatch_doy_sc   -1.5507  0.2994 1381 -5.179 0.0000
## age_days_sc50  0.2198  0.1344 1381  1.636 0.1022
## -- leftS:
## signal_for_mass_50 1.7290  0.2097 1381  8.244 0.0000
## -- diffS:
## U              -2.3166  0.4864 1381 -4.763
## -- break:

```

```

## G0          19.1349    0.2908 1381
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -3.85069006 -0.48266406  0.03708332  0.54051849  4.12827680
##
## Number of Observations: 2693
## Number of Groups:
##           year_f nestcode_rearing %in% year_f
##           25          1307

```

psi = 21

```

load("output/mass_50_climmod_segmented_21.rda")
summary(mass_50_climmod_segmented)

```

```

## Segmented mixed-effects model fit by REML
##      AIC      BIC    logLik
## 19090.4 19178.83 -9530.198
## Bootstrap restarting on 10 samples; 2 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_mass_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U
## StdDev:      2.52527      0.001413049 1.223453
##
## Formula: ~1 + signal_for_mass_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U Residual
## StdDev:      5.292029      0.01488652 0.04727261 6.792072
##
## Fixed effects:
##           Value Std.Error  DF t-value p-value
## (Intercept)  68.8372   3.8157 1381  18.041  0.0000
## bsh_ax      -2.3970   0.3154 1280  -7.600  0.0000
## colonySolothurn  2.5624   0.4578 1280   5.598  0.0000
## hatch_doy_sc   -1.5508   0.2997 1381  -5.175  0.0000
## age_days_sc50   0.2184   0.1345 1381   1.624  0.1046
## -- leftS:
## signal_for_mass_50 1.7446   0.2117 1381   8.243  0.0000
## -- diffS:
## U              -2.3077   0.4820 1381  -4.788
## -- break:
## G0          19.1047    0.2927 1381
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -3.85137691 -0.48288586  0.03695215  0.53929450  4.12791184
##
## Number of Observations: 2693

```

```
## Number of Groups:
##           year_f nestcode_rearing %in% year_f
##           25           1307
```

psi = 22

```
load("output/mass_50_climmod_segmented_22.rda")
summary(mass_50_climmod_segmented)
```

```
## Segmented mixed-effects model fit by REML
##      AIC      BIC    logLik
## 19090.4 19178.83 -9530.198
## Bootstrap restarting on 10 samples; 2 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_mass_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U
## StdDev:      2.52527      0.001413049 1.223453
##
## Formula: ~1 + signal_for_mass_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U Residual
## StdDev:      5.292029      0.01488652 0.04727261 6.792072
##
## Fixed effects:
##           Value Std.Error  DF t-value p-value
## (Intercept)  68.8372   3.8157 1381  18.041  0.0000
## bsh_ax      -2.3970   0.3154 1280  -7.600  0.0000
## colonySolothurn  2.5624   0.4578 1280   5.598  0.0000
## hatch_doy_sc   -1.5508   0.2997 1381  -5.175  0.0000
## age_days_sc50   0.2184   0.1345 1381   1.624  0.1046
## -- leftS:
## signal_for_mass_50 1.7446   0.2117 1381   8.243  0.0000
## -- diffS:
## U                -2.3077   0.4820 1381  -4.788
## -- break:
## G0                19.1047   0.2927 1381
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -3.85137692 -0.48288586 0.03695216 0.53929450 4.12791184
##
## Number of Observations: 2693
## Number of Groups:
##           year_f nestcode_rearing %in% year_f
##           25           1307
```

psi = 23

```
load("output/mass_50_climmod_segmented_23.rda")
summary(mass_50_climmod_segmented)
```

```
## Segmented mixed-effects model fit by REML
##      AIC      BIC    logLik
## 19090.37 19178.81 -9530.186
## Bootstrap restarting on 10 samples; 1 different solution(s)
##
## Random effects:
## Formula: ~1 + signal_for_mass_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U
## StdDev:    2.512856      0.001400537 1.262121
##
## Formula: ~1 + signal_for_mass_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U Residual
## StdDev:    5.292569      0.01476722 0.04996561 6.79192
##
## Fixed effects:
##      Value Std.Error  DF t-value p-value
## (Intercept)    69.2932   3.7285 1381  18.585 0.0000
## bsh_ax         -2.4001   0.3154 1280  -7.610 0.0000
## colonySolothurn 2.5631   0.4577 1280   5.600 0.0000
## hatch_doy_sc   -1.5504   0.2994 1381  -5.179 0.0000
## age_days_sc50  0.2205   0.1344 1381   1.641 0.1011
## -- leftS:
## signal_for_mass_50 1.7164   0.2059 1381   8.336 0.0000
## -- diffS:
## U                -2.3252   0.4927 1381  -4.719
## -- break:
## G0                19.1538   0.2872 1381
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -3.85021117 -0.48336447 0.03608181 0.53985955 4.12859034
##
## Number of Observations: 2693
## Number of Groups:
##      year_f nestcode_rearing %in% year_f
##      25      1307
```

psi = 19

```
load("output/mass_50_climmod_segmented_19.rda")
summary(mass_50_climmod_segmented)
```

```
## Segmented mixed-effects model fit by REML
##      AIC      BIC    logLik
## 19090.4 19178.83 -9530.198
## Bootstrap restarting on 10 samples; 2 different solution(s)
```

```

##
## Random effects:
## Formula: ~1 + signal_for_mass_50 + U | year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U
## StdDev:    2.525282      0.001413091 1.223397
##
## Formula: ~1 + signal_for_mass_50 + U | nestcode_rearing %in% year_f
## Structure: Diagonal
##      (Intercept) signal_for_mass_50      U Residual
## StdDev:    5.292029      0.01488681 0.04726972 6.792072
##
## Fixed effects:
##      Value Std.Error  DF t-value p-value
## (Intercept)    68.8366   3.8157 1381  18.040  0.0000
## bsh_ax         -2.3970   0.3154 1280  -7.600  0.0000
## colonySolithurn  2.5624   0.4578 1280   5.598  0.0000
## hatch_doy_sc    -1.5508   0.2997 1381  -5.175  0.0000
## age_days_sc50   0.2184   0.1345 1381   1.624  0.1046
## -- leftS:
## signal_for_mass_50  1.7446   0.2117 1381   8.243  0.0000
## -- diffS:
## U                -2.3077   0.4819 1381  -4.788
## -- break:
## G0                19.1046   0.2927 1381
## psi.link = identity
##
## Standardized Within-Group Residuals:
##      Min      Q1      Med      Q3      Max
## -3.85137870 -0.48288665  0.03695542  0.53929255  4.12791094
##
## Number of Observations: 2693
## Number of Groups:
##      year_f nestcode_rearing %in% year_f
##      25      1307

```

All the threshold found are very similar, this we decided to use the threshold identified by this last model. The threshold is at 19.1531402 according to the segmented analysis. The two slopes (before and after the point) are:

```

segmented_threshold <- mean(mass_50_climmod_segmented$psi)

data_am_nestlings_mass_501 <- subset(
  data_am_nestlings_mass_50,
  signal_for_mass_50 < segmented_threshold
)
data_am_nestlings_mass_502 <- subset(
  data_am_nestlings_mass_50,
  signal_for_mass_50 > segmented_threshold
)

```

Before the point

```

mass_50_climmod1 <- lmer(
  mass_50 ~
    signal_for_mass_50 +
    brood_size + colony +
    hatch_doy_sc + age_days_sc50 +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_mass_501
)
summary(mass_50_climmod1)

## Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
## Formula: mass_50 ~ signal_for_mass_50 + brood_size + colony + hatch_doy_sc +
##   age_days_sc50 + (1 | year_f) + (1 | nestcode_rearing)
## Data: data_am_nestlings_mass_501
## Control: lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
##
## REML criterion at convergence: 10315.7
##
## Scaled residuals:
##   Min       1Q   Median       3Q      Max
## -3.7567 -0.4819  0.0292  0.5206  4.0466
##
## Random effects:
##   Groups                Name            Variance Std.Dev.
## nestcode_rearing (Intercept) 32.330    5.686
## year_f            (Intercept)  7.895    2.810
## Residual                            48.426    6.959
## Number of obs: 1443, groups: nestcode_rearing, 715; year_f, 25
##
## Fixed effects:
##              Estimate Std. Error    df t value Pr(>|t|)
## (Intercept)    69.4941    4.5005 387.7888  15.442 < 2e-16 ***
## signal_for_mass_50  1.7663    0.2474 470.9707   7.139 3.58e-12 ***
## brood_size      -2.9802    0.4531 805.4378  -6.577 8.64e-11 ***
## colonySolothurn  3.2735    0.6450 655.8994   5.076 5.04e-07 ***
## hatch_doy_sc    -1.2887    0.4053 146.5579  -3.180 0.0018 **
## age_days_sc50     0.3164    0.1788 713.6018   1.770 0.0772 .
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) s___50 brd_sz clnySl htch__
## sgnl_fr__50 -0.950
## brood_size  -0.239 -0.014
## colnySlthrn -0.079 -0.007 -0.054
## hatch_dy_sc -0.197  0.134  0.138  0.094
## ag_dys_sc50 -0.009  0.049 -0.123 -0.108 -0.033

car::Anova(mass_50_climmod1)

## Analysis of Deviance Table (Type II Wald chisquare tests)

```

```
##
## Response: mass_50
##               Chisq Df Pr(>Chisq)
## signal_for_mass_50 50.9685  1 9.386e-13 ***
## brood_size         43.2537  1 4.808e-11 ***
## colony             25.7608  1 3.865e-07 ***
## hatch_doy_sc      10.1116  1 0.001473 **
## age_days_sc50     3.1329  1 0.076728 .
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

After the point

```
mass_50_climmod2 <- lmer(
  mass_50 ~
    signal_for_mass_50 +
    # I(signal_for_mass_50^2) +
    brood_size + colony +
    hatch_doy_sc + age_days_sc50 +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_mass_502
)
summary(mass_50_climmod2)
```

```
## Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
## Formula: mass_50 ~ signal_for_mass_50 + brood_size + colony + hatch_doy_sc +
##   age_days_sc50 + (1 | year_f) + (1 | nestcode_rearing)
## Data: data_am_nestlings_mass_502
## Control: lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
##
## REML criterion at convergence: 8717.9
##
## Scaled residuals:
##   Min       1Q   Median       3Q      Max
## -3.6675 -0.4777  0.0479  0.5484  2.6921
##
## Random effects:
##   Groups             Name                Variance Std.Dev.
## nestcode_rearing (Intercept) 25.133    5.013
## year_f             (Intercept)  1.622    1.274
## Residual                                42.829    6.544
## Number of obs: 1250, groups: nestcode_rearing, 597; year_f, 24
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)  110.12453   5.05255 204.70678  21.796 < 2e-16 ***
## signal_for_mass_50 -0.47951   0.23259 205.55329  -2.062 0.040504 *
## brood_size      -1.88957   0.43565 745.90016  -4.337 1.64e-05 ***
## colonySolothurn  2.25863   0.62198 442.01146   3.631 0.000315 ***
## hatch_doy_sc    -1.91266   0.36992  71.14396  -5.170 2.06e-06 ***
## age_days_sc50   -0.07131   0.20929 557.30963  -0.341 0.733429
## ---
```



```
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##      (Intr) s___50 brd_sz clnySl htch__
## sgnl_fr__50 -0.968
## brood_size  -0.204 -0.021
## colnySlthrn -0.041 -0.029 -0.049
## hatch_dy_sc -0.044  0.034  0.066 -0.028
## ag_dys_sc50 -0.012  0.042 -0.024 -0.223  0.058
```

```
car::Anova(mass_50_climmod2)
```

```
## Analysis of Deviance Table (Type II Wald chisquare tests)
##
## Response: mass_50
##              Chisq Df Pr(>Chisq)
## signal_for_mass_50  4.2502  1  0.0392456 *
## brood_size         18.8125  1  1.442e-05 ***
## colony             13.1869  1  0.0002819 ***
## hatch_doy_sc       26.7339  1  2.335e-07 ***
## age_days_sc50      0.1161  1  0.7333002
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
clim_predict <-
  data.frame(effects::effect("signal_for_mass_50",
    mass_50_climmod,
    partial.residuals = TRUE
  ))
clim_predict_01 <-
  data.frame(effects::effect("signal_for_mass_50",
    mass_50_climmod1,
    partial.residuals = TRUE
  ))
clim_predict_02 <-
  data.frame(effects::effect("signal_for_mass_50",
    mass_50_climmod2,
    partial.residuals = TRUE
  ))

par_xmin <- round(min(data_am_nestlings_mass_50$signal_for_mass_50) - 0.005, 2)
par_xmax <- round(max(data_am_nestlings_mass_50$signal_for_mass_50) + 0.005, 2)

plot_clim_mass_50 <- ggplot(
  clim_predict,
  aes(signal_for_mass_50, fit)
) +
  geom_point(
    data = data_am_nestlings_mass_50,
    aes(
```

```

    x = signal_for_mass_50,
    y = mass_50
  ),
  col = par_col_mass_50,
  size = 1, shape = 16, alpha = 0.2
) +
geom_vline(
  xintercept = segmented_threshold,
  linetype = "dashed",
  color = "dodgerblue"
) +
geom_ribbon(
  data = clim_predict_01,
  aes(ymin = lower, ymax = upper),
  alpha = .25
) +
geom_line(
  data = clim_predict_01, aes(signal_for_mass_50, fit),
  col = "darkgreen",
  linewidth = 1.2
) +
geom_ribbon(
  data = clim_predict_02,
  aes(ymin = lower, ymax = upper),
  alpha = .25
) +
geom_line(
  data = clim_predict_02, aes(signal_for_mass_50, fit),
  col = "#20b2aa",
  linewidth = 1.2
) +
theme(
  legend.position = "none",
  panel.grid.major = element_blank(),
  panel.grid.minor = element_blank(),
  panel.background = element_blank(),
  axis.line = element_line(colour = "black"),
  axis.title.x = element_markdown()
) +
xlab(
  "Mean ambient temperature (Ta<sub>48-50d</sub>; °C)<br>between 48 and 50 days of age"
) +
ylab("Body mass at 50 days (g)") +
scale_y_continuous(
  limits = c(55, 130),
  breaks = c(seq(55, 150, 15))
) +
scale_x_continuous(
  limits = c(par_xmin, par_xmax),
  breaks = c(round(seq(
    par_xmin, par_xmax,
    (par_xmax - par_xmin) / 4
  ), 1))
), 1))

```

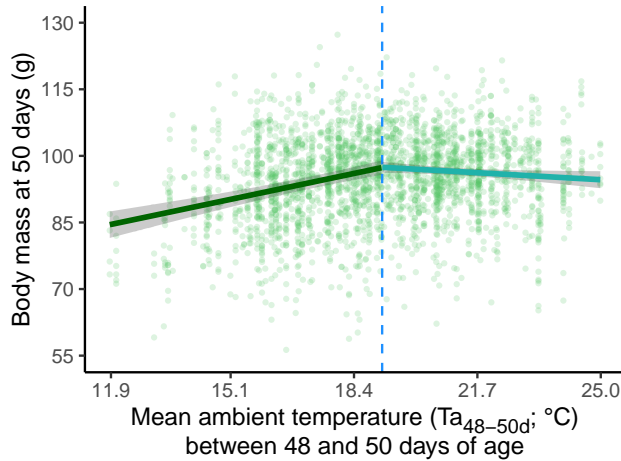


Figure 4: Fig. 3 - Variation in body mass of 50-day-old nestling Alpine swifts in relation to daily average temperature in the 2 days before the measurement. The analysis using the segmented package identified a threshold at 19.1°C. Solid lines (and 95% confidence intervals) are predictions from the models presented in Table 2. Climatic windows are reported using the nestling's age as a reference.

)

Figure

plot_clim_mass_50

```
# Save the plot to the figures directory
ggsave("figures/Fig. 3 - mass-weather_segmented.png", width = 4, height = 3, dpi = 300)
```

Supporting Information 02: Growth rates

To estimate growth rates, I extracted the slope of the linear regression between the measurements of the trait within the windows identified by the climwin analyses.

```
# Growth rates -----
am_growth_wing <- data_am_nestlings_all %>%
  subset(!is.na(wing)) %>%
  subset(age_days <= 48) %>%
  subset(age_days >= 1) %>%
  group_by(ring) %>%
  mutate(
    wing_n_measures = n()
  ) %>%
  ungroup()

am_growth_wing <- am_growth_wing %>%
  subset(!is.na(wing_n_measures) & wing_n_measures > 2) %>%
  dplyr::select(ring, wing, age_days, wing_n_measures) %>%
```

```

distinct() %>%
group_by(ring) %>%
mutate(
  wing_growth = coefficients(lm(wing ~ age_days))[2],
  wing_r2 = summary(lm(wing ~ age_days))$r.squared
) %>%
ungroup() %>%
dplyr::select(
  ring, wing_growth, wing_n_measures, wing_r2
) %>%
distinct()

```

```

## Warning: There were 6 warnings in `mutate()`.
## The first warning was:
## i In argument: `wing_r2 = summary(lm(wing ~ age_days))$r.squared`.
## i In group 61: `ring = ring_000074`.
## Caused by warning in `summary.lm()`:
## ! essentially perfect fit: summary may be unreliable
## i Run `dplyr::last_dplyr_warnings()` to see the 5 remaining warnings.

```

```

am_growth_sternum <- data_am_nestlings_all %>%
  subset(!is.na(sternum)) %>%
  subset(age_days <= 34) %>%
  subset(age_days >= 12) %>%
  group_by(ring) %>%
  mutate(
    sternum_n_measures = n()
  ) %>%
  ungroup()

am_growth_sternum <- am_growth_sternum %>%
  subset(!is.na(sternum_n_measures) & sternum_n_measures >= 2) %>%
  dplyr::select(ring, sternum, age_days, sternum_n_measures) %>%
  distinct() %>%
  group_by(ring) %>%
  mutate(
    sternum_growth = coefficients(lm(sternum ~ age_days))[2],
    sternum_r2 = summary(lm(sternum ~ age_days))$r.squared
  ) %>%
  ungroup() %>%
  dplyr::select(
    ring, sternum_growth, sternum_n_measures, sternum_r2
  ) %>%
  distinct()

```

```

## Warning: There were 3 warnings in `mutate()`.
## The first warning was:
## i In argument: `sternum_r2 = summary(lm(sternum ~ age_days))$r.squared`.
## i In group 620: `ring = ring_000987`.
## Caused by warning in `summary.lm()`:
## ! essentially perfect fit: summary may be unreliable
## i Run `dplyr::last_dplyr_warnings()` to see the 2 remaining warnings.

```

```

# adding the growth rates to the dataset
am_nestlings_clim <- merge(am_nestlings_clim,
  merge(am_growth_wing, am_growth_sternum, all = TRUE),
  all.x = TRUE
)

rm(am_growth_wing, am_growth_sternum)

# creating the subsets for the analyses without NAs
data_am_nestlings_wing_growth <- subset(
  am_nestlings_clim, !is.na(wing_growth)
)
data_am_nestlings_sternum_growth <- subset(
  am_nestlings_clim, !is.na(sternum_growth)
)

```

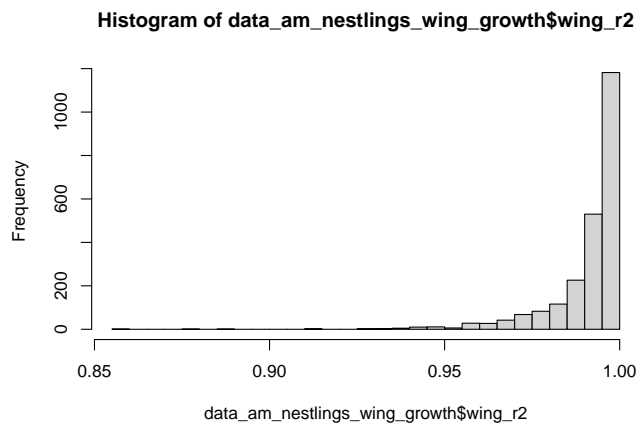
The warnings in this case just indicate that in some cases we have perfect fit, as it's quite likely with fitting a linear model using 2-3 points.

Checking the resulting growth rates

Wing growth rate

The histograms of the R squared values are plotted here.

```
hist(data_am_nestlings_wing_growth$wing_r2, breaks = 50)
```



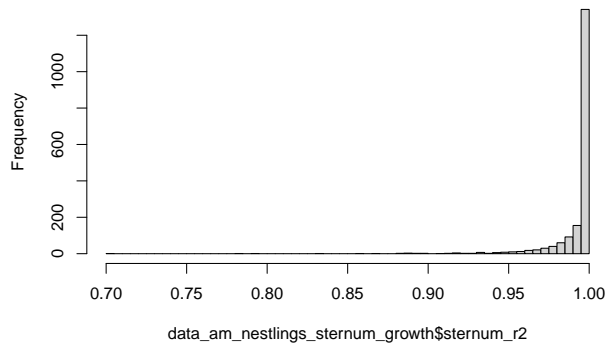
The r2 value for the wing growth rates are $0.99 + 0.01$ (mean + SD). The histogram and the average values show that the linear fit is a good approximation of the growth of the nestlings within the time window considered.

Sternum growth rate

The histograms of the R squared values are plotted here.

```
hist(data_am_nestlings_sternum_growth$sternum_r2, breaks = 50)
```

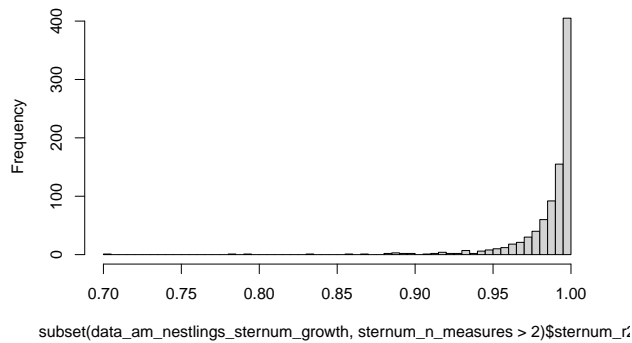
Histogram of data_am_nestlings_sternum_growth\$sternum_r2



As for the sternum we constructed the slopes for nestlings with at least 2 measurements within the window, here we show also the R squared for nestlings with at least 3 measurements within the window to check that the R squared obtained are still good.

```
hist(subset(
  data_am_nestlings_sternum_growth,
  sternum_n_measures > 2
)$sternum_r2, breaks = 50)
```

of subset(data_am_nestlings_sternum_growth, sternum_n_measures >



This shows that using a linear slope for the sternum is a good approximation even fore nestlings with only 2 measurements.

The r2 value for the sternum growth rates are $0.99 + 0.02$ (mean + SD). The histograms and the average values show that the linear fit is a good approximation of the growth of the nestlings within the time window considered.

Growth rate models

Wing length

```
wing_growth_climmod_quad <- lmer(
  wing_growth ~
  signal_for_wing_50 +
  I(signal_for_wing_50^2) +
  brood_size + colony +
  hatch_doy_sc +
  (1 | year_f) + (1 | nestcode_rearing),
```

```

control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
data = data_am_nestlings_wing_growth
)
wing_growth_climmod_lin <- lmer(
  wing_growth ~
    signal_for_wing_50 +
    brood_size + colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_wing_growth
)
MuMin::AICc(wing_growth_climmod_quad, wing_growth_climmod_lin)

```

```

##                df      AICc
## wing_growth_climmod_quad  9 1534.603
## wing_growth_climmod_lin   8 1533.470

```

The AIC value is very similar but linear is lower and more parsimonious.

Sternum length

```

sternum_growth_climmod_quad <- lmer(
  sternum_growth ~
    signal_for_sternum_50 + I(signal_for_sternum_50^2) +
    brood_size + colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_sternum_growth
)
sternum_growth_climmod_lin <- lmer(
  sternum_growth ~
    signal_for_sternum_50 +
    brood_size + colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_sternum_growth
)
MuMin::AICc(sternum_growth_climmod_quad, sternum_growth_climmod_lin)

```

```

##                df      AICc
## sternum_growth_climmod_quad  9 -2191.962
## sternum_growth_climmod_lin   8 -2197.919

```

```
# quadratic seems better
```

The linear model has a lower AIC value.

```

wing_growth_climmod <- lmer(
  wing_growth ~
    brood_size + colony +
    hatch_doy_sc +
    signal_for_wing_50 +
    (1 | year_f) + (1 | nestcode_rearing),
  control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
  data = data_am_nestlings_wing_growth
)
summary(wing_growth_climmod)

```

```

## Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
## Formula: wing_growth ~ brood_size + colony + hatch_doy_sc + signal_for_wing_50 +
##   (1 | year_f) + (1 | nestcode_rearing)
## Data: data_am_nestlings_wing_growth
## Control: lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
##
## REML criterion at convergence: 1517.4
##
## Scaled residuals:
##   Min       1Q   Median       3Q      Max
## -7.5306 -0.3636  0.0996  0.4841  4.0765
##
## Random effects:
##   Groups                Name                Variance Std.Dev.
## nestcode_rearing (Intercept) 0.05580  0.2362
## year_f            (Intercept) 0.09736  0.3120
## Residual                                0.06619  0.2573
## Number of obs: 2344, groups: nestcode_rearing, 1141; year_f, 25
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)      4.97208   0.07832  46.68061  63.484 < 2e-16 ***
## brood_size       -0.13328   0.01391 1276.31114  -9.584 < 2e-16 ***
## colonySlothurn    0.07707   0.02183 1074.50792   3.530 0.000434 ***
## hatch_doy_sc     -0.14156   0.01417 1104.04962  -9.992 < 2e-16 ***
## signal_for_wing_50 0.12872   0.08637  382.29280   1.490 0.136971
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) brd_sz clnySl htch__
## brood_size  -0.437
## colnySlthrn -0.154 -0.077
## hatch_dy_sc -0.126  0.131  0.083
## sgnl_fr__50 -0.328 -0.017 -0.062  0.110

```

```

sternum_growth_climmod <- lmer(
  sternum_growth ~
    brood_size + colony +
    hatch_doy_sc +
    signal_for_sternum_50 +
    (1 | year_f) + (1 | nestcode_rearing),

```



```

control = lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06)),
data = data_am_nestlings_sternum_growth
)
summary(sternum_growth_climmod)

## Linear mixed model fit by REML. t-tests use Satterthwaite's method ['lmerModLmerTest']
## Formula: sternum_growth ~ brood_size + colony + hatch_doy_sc + signal_for_sternum_50 +
## (1 | year_f) + (1 | nestcode_rearing)
## Data: data_am_nestlings_sternum_growth
## Control: lmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
##
## REML criterion at convergence: -2214
##
## Scaled residuals:
##   Min       1Q   Median       3Q      Max
## -5.2793 -0.4484  0.0261  0.4572 10.2427
##
## Random effects:
##   Groups             Name                Variance Std.Dev.
## nestcode_rearing (Intercept) 0.007301 0.08545
## year_f             (Intercept) 0.011989 0.10949
## Residual                                0.010893 0.10437
## Number of obs: 1824, groups: nestcode_rearing, 893; year_f, 21
##
## Fixed effects:
##              Estimate Std. Error      df t value Pr(>|t|)
## (Intercept)    9.130e-01  3.132e-02 4.855e+01  29.149 < 2e-16 ***
## brood_size     -2.782e-02  5.829e-03 1.038e+03  -4.773 2.08e-06 ***
## colonySolothurn -1.585e-03  9.607e-03 8.725e+02  -0.165 0.86897
## hatch_doy_sc   -1.810e-02  6.120e-03 8.544e+02  -2.958 0.00318 **
## signal_for_sternum_50 8.008e-02  2.714e-02 3.429e+02   2.951 0.00339 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Correlation of Fixed Effects:
##              (Intr) brd_sz clnySl htch__
## brood_size  -0.488
## colnySlthrn -0.224 -0.061
## hatch_dy_sc -0.010  0.092  0.078
## sgnl_fr__50 -0.356  0.070  0.046 -0.206

labels <- c(
  `(Intercept)` = "Intercept",
  hatch_doy_sc = "Hatching day",
  brood_size = "Hatching brood size",
  colonySolothurn = "Colony [Solothurn]",
  signal_for_wing_50 = "PC1 (age: 1-48)",
  `I(signal_for_wing_50^2)` = "PC1 (age: 1-48) 2",
  signal_for_sternum_50 = "PC1 (age: 12-34)",
  `I(signal_for_sternum_50^2)` = "PC1 (age: 12-34) 2"
)

sjPlot::tab_model(

```

```

wing_growth_climmod,
sternum_growth_climmod,
file = "output/Tab_wicm_models_growth.doc",
string.est = "Estimate",
string.se = "SE",
show.ci = FALSE,
show.se = TRUE,
show.stat = FALSE,
show.df = FALSE,
pred.labels = labels
)

```

Figure

```

cowplot::plot_grid(
  fun_climres(wing_growth_climmod, data_am_nestlings_wing_growth),
  fun_climres(sternum_growth_climmod, data_am_nestlings_sternum_growth),
  nrow = 2, align = c("v", "h")
)

```

```

# Save the plot to the figures directory
ggsave("figures/Fig. 4 - growths-weather.png", width = 4, height = 5, dpi = 300)

```

Supporting Information 03: Correlation between the size and growth

Figure

Here we show the correlation between the size at 50 days (wing, sternum and mass) and the growth rates (wing and sternum).

```

PerformanceAnalytics::chart.Correlation(am_nestlings_clim[
  ,
  c("wing_50", "wing_growth", "sternum_50", "sternum_growth", "mass_50")
], histogram = TRUE, pch = 19, method = "pearson")

```

```

## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter
## Warning in par(usr): argument 1 does not name a graphical parameter

```

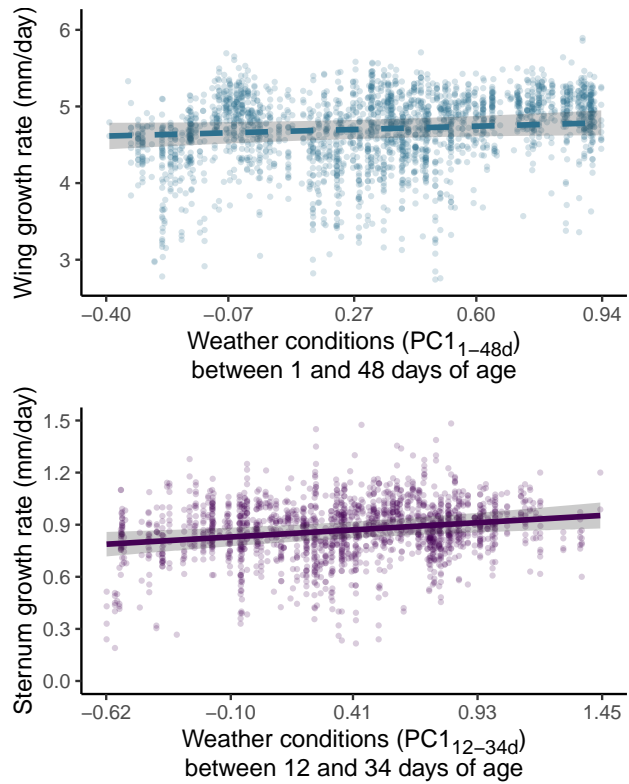
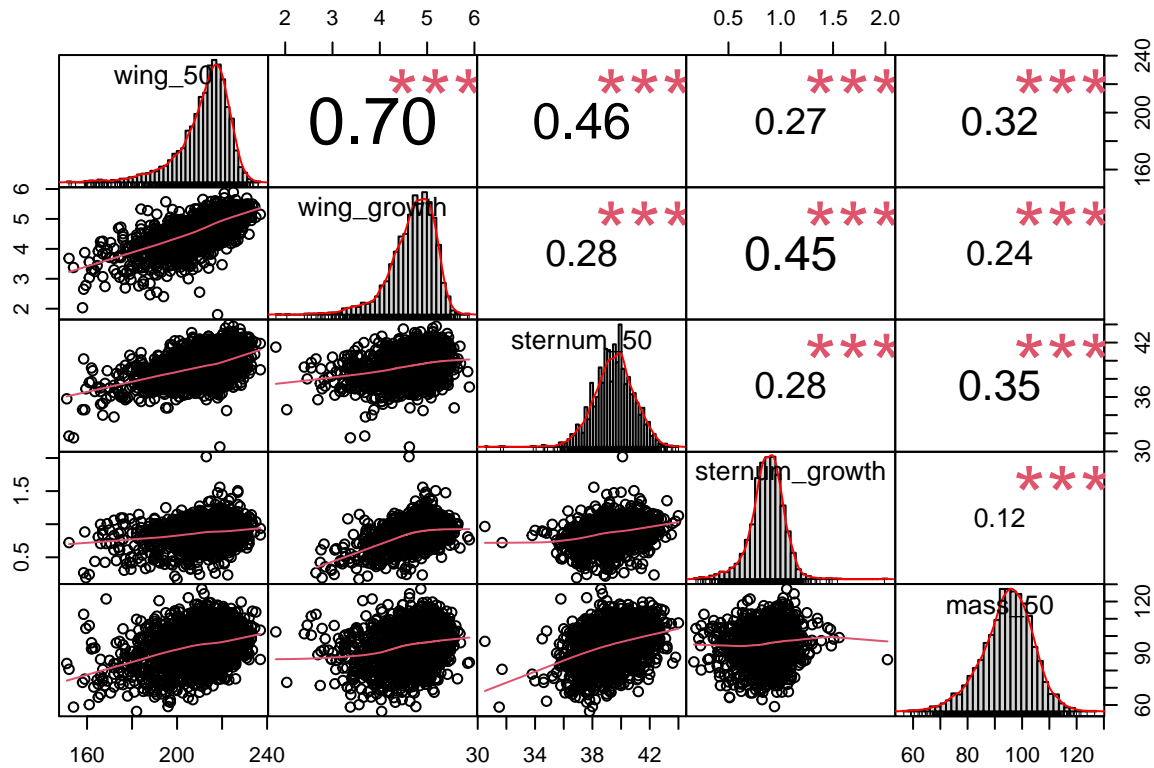


Figure 5: Fig. 4 - Variation in wing and sternum growth rates of nestling Alpine swifts in relation to weather conditions encountered during development. Weather conditions resulted from a principal component analysis (PC1) between daily average temperature and daily rain, and traits were affected by weather conditions over different time windows. PC1 has low values with cold and rainy days and high values with sunny and warm days. Solid lines (and 95% confidence intervals) are predictions from the models presented in Table 3. Climatic windows are reported using the nestling's age as a reference.



In the above plot:

- The distribution of each variable is shown on the diagonal.
- On the bottom of the diagonal: the bivariate scatter plots with a fitted line are displayed
- On the top of the diagonal: the value of the correlation (Pearson r) plus the significance level as stars
- Each significance level (p-values: 0, 0.001, 0.01, 0.05, 0.1, 1) is associated to a symbol.

Pearson's correlation tests

Wing length

```
cor.test(am_nestlings_clim$wing_50, am_nestlings_clim$wing_growth)
```

```
##
## Pearson's product-moment correlation
##
## data: am_nestlings_clim$wing_50 and am_nestlings_clim$wing_growth
## t = 47.217, df = 2342, p-value < 2.2e-16
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## 0.6770023 0.7185188
## sample estimates:
## cor
## 0.6983475
```

Sternum length

```

cor.test(am_nestlings_clim$sternum_50, am_nestlings_clim$sternum_growth)

##
## Pearson's product-moment correlation
##
## data:  am_nestlings_clim$sternum_50 and am_nestlings_clim$sternum_growth
## t = 12.544, df = 1822, p-value < 2.2e-16
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
##  0.2391484 0.3236600
## sample estimates:
##      cor
## 0.281951

```

Supporting Information 04: Short-term consequences

Probability of fledging

To investigate the number of nestlings that survived I use `cbind()` with the number of nestlings that fledged and the number of nestlings that did not fledge (number of hatchlings - number of fledglings) using the data from nests with at least 1 nestling fledging.

```

data_am_nests$hatch_date <- as.Date(data_am_nests$hatch_date)
data_am_nests$date_50days <- data_am_nests$hatch_date + 50
data_am_nests$hatch_doy_sc <- fun_scale(yday(data_am_nests$hatch_date))
data_am_nests$year_f <- data_am_nests$year

data_am_nests <- droplevels(data_am_nests)
data_am_nests$prop <- data_am_nests$fledged /
  (data_am_nests$fledged + data_am_nests$not_fledg)

data_signals <- am_nestlings_clim[c(
  "date_50days", "signal_for_wing_50",
  "signal_for_sternum_50", "signal_for_mass_50"
)] %>%
  distinct(date_50days, .keep_all = TRUE)

data_am_nests <- merge(
  data_am_nests,
  data_signals,
  all.x = TRUE, all.y = FALSE
)
data_am_nests$obs <- seq_len(nrow(data_am_nests))

```

I'm interested in comparing the weather variables that affect each of the trait, to see which is the window that best affects the probability of fledging. To be able to compare the 3 models I first create a subset that has complete cases for all 3 weather conditions.

```

am_nest_clim_reduced <- subset(subset(
  subset(data_am_nests, !is.na(signal_for_mass_50)),
  !is.na(signal_for_wing_50)
), !is.na(signal_for_sternum_50))

am_nest_clim_wing <- subset(data_am_nests, !is.na(signal_for_wing_50))

fledg_surv_climmod_wing <- glmer(
  cbind(fledged, not_fledg) ~
  brood_size +
  colony +
  hatch_doy_sc +
  signal_for_wing_50 +
  (1 | year_f) +
  (1 | obs),
  data = am_nest_clim_reduced,
  family = binomial,
  control = glmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
)

fledg_surv_climmod_sternum <- glmer(
  cbind(fledged, not_fledg) ~
  brood_size +
  colony +
  hatch_doy_sc +
  signal_for_sternum_50 +
  (1 | year_f) +
  (1 | obs),
  data = am_nest_clim_reduced,
  family = binomial,
  control = glmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
)

fledg_surv_climmod_mass <- glmer(
  cbind(fledged, not_fledg) ~
  brood_size +
  colony +
  hatch_doy_sc +
  signal_for_mass_50 +
  (1 | year_f) +
  (1 | obs),
  data = am_nest_clim_reduced,
  family = binomial,
  control = glmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
)

MuMin::AICc(
  fledg_surv_climmod_wing,
  fledg_surv_climmod_sternum,
  fledg_surv_climmod_mass
)

```

```
##                df      AICc
```

```
## fledg_surv_climmod_wing      7 2696.659
## fledg_surv_climmod_sternum  7 2699.054
## fledg_surv_climmod_mass     7 2698.325
```

The climatic window for the sternum length is better even if the models have very similar AIC ($\Delta AIC_c < 4$). To keep a larger sample size and to be consistent with the following analysis we decided to use the one for the wing length. The window for sternum length is anyway fully included in the window for wing length.

Testing for quadratic effects

```
fledg_surv_climmod_quad <- glmer(
  cbind(fledged, not_fledged) ~
    brood_size +
    colony +
    hatch_doy_sc +
    signal_for_wing_50 +
    I(signal_for_wing_50^2) +
    (1 | year_f) +
    (1 | obs),
  data = am_nest_clim_wing,
  family = binomial,
  control = glmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
)
fledg_surv_climmod_lin <- glmer(
  cbind(fledged, not_fledged) ~
    brood_size +
    colony +
    hatch_doy_sc +
    signal_for_wing_50 +
    (1 | year_f) +
    (1 | obs),
  data = am_nest_clim_wing,
  family = binomial,
  control = glmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
)
MuMIn::AICc(fledg_surv_climmod_lin, fledg_surv_climmod_quad)
```

```
##                df    AICc
## fledg_surv_climmod_lin  7 3020.567
## fledg_surv_climmod_quad  8 3022.307
```

Not keeping the quadratic effect

Checking for overdispersion The models above showed overdispersion when I did not included an observation-level random effect. See an example of the results on overdispersion here:

```
fledg_surv_climmod_lin_overdispersed <- glmer(
  cbind(fledged, not_fledged) ~
    brood_size +
    colony +
    hatch_doy_sc +
    signal_for_wing_50 +
    (1 | year_f),
```

```

data = am_nest_clim_wing,
family = binomial,
control = glmerControl(optimizer = "bobyqa", optCtrl = list(maxfun = 1e+06))
)
performance::check_overdispersion(fledg_surv_climmod_lin_overdispersed)

```

```

## # Overdispersion test
##
## dispersion ratio = 1.178
##           p-value = 0.008

```

```
## Overdispersion detected.
```

Therefore in all the models on fledgling survival presented above I have included an observation level random effect that deals with the overdispersion of the data. See new results here:

```
performance::check_overdispersion(fledg_surv_climmod_lin)
```

```

## # Overdispersion test
##
## dispersion ratio = 1.055
##           p-value = 0.336

```

```
## No overdispersion detected.
```

Now there seem to be no overdispersion detected.

Age at fledging

For some years we have data on the age at fledging, so we can check the correlation between age at fledging, size and growth of nestlings

Correlation plot, plus I've tried some analyses, but they are VERY preliminary and need to be checked.

```

am_age_fledging <- am_nestlings_clim %>%
  subset(!duplicated(ring)) %>%
  subset(!is.na(age_fledging)) %>%
  # subset(!is.na(sternum_50)) %>%
  # subset(year < 2012) %>%
  dplyr::select(
    year, ring, age_fledging,
    brood_size, colony, hatch_doy_sc,
    date_50days,
    wing_50, wing_growth,
    sternum_50, sternum_growth,
    mass_50,
    year_f, nestcode_rearing,
    signal_for_mass_50, signal_for_wing_50,
    signal_for_sternum_50
  )

```



```

am_age_fledging <- am_age_fledging %>%
  filter(year != 2012) %>%
  droplevels()

am_age_fledging_reduced <- am_age_fledging %>%
  filter(!is.na(signal_for_mass_50)) %>%
  filter(!is.na(signal_for_wing_50)) %>%
  filter(!is.na(signal_for_sternum_50))

am_age_fledging_wing <- am_age_fledging %>%
  filter(!is.na(signal_for_wing_50))

age_fledg_climmod_wing <- lmer(
  age_fledging ~
    signal_for_wing_50 +
    brood_size +
    colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  data = am_age_fledging_reduced,
  control = lmerControl(
    optimizer = "bobyqa",
    optCtrl = list(maxfun = 1e+07)
  )
)

age_fledg_climmod_sternum <- lmer(
  age_fledging ~
    signal_for_sternum_50 +
    brood_size +
    colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  data = am_age_fledging_reduced,
  control = lmerControl(
    optimizer = "bobyqa",
    optCtrl = list(maxfun = 1e+07)
  )
)

age_fledg_climmod_mass <- lmer(
  age_fledging ~
    signal_for_mass_50 +
    brood_size +
    colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  data = am_age_fledging_reduced,
  control = lmerControl(
    optimizer = "bobyqa",
    optCtrl = list(maxfun = 1e+07)
  )
)

```

```
)
MuMIn::AICc(age_fledg_climmod_wing, age_fledg_climmod_sternum, age_fledg_climmod_mass)

##                df      AICc
## age_fledg_climmod_wing    8 2936.399
## age_fledg_climmod_sternum  8 2948.733
## age_fledg_climmod_mass    8 2940.965
```

The climatic window for the wings is the best one.

Testing for quadratic effects

```
age_fledg_climmod_quad <- lmer(
  age_fledging ~
    signal_for_wing_50 +
    I(signal_for_wing_50^2) +
    brood_size +
    colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  data = am_age_fledging_wing,
  control = lmerControl(
    optimizer = "bobyqa",
    optCtrl = list(maxfun = 1e+07)
  )
)

age_fledg_climmod_lin <- lmer(
  age_fledging ~
    signal_for_wing_50 +
    brood_size +
    colony +
    hatch_doy_sc +
    (1 | year_f) + (1 | nestcode_rearing),
  data = am_age_fledging_wing,
  control = lmerControl(
    optimizer = "bobyqa",
    optCtrl = list(maxfun = 1e+07)
  )
)
MuMIn::AICc(age_fledg_climmod_quad, age_fledg_climmod_lin)

##                df      AICc
## age_fledg_climmod_quad    9 3918.432
## age_fledg_climmod_lin    8 3920.814
```

Models practically the same, not keeping the quadratic effect

```
labels <- c(
  `(Intercept)` = "Intercept",
  hatch_doy_sc = "Hatching day",
```

```

brood_size = "Hatching brood size",
signal_for_wing_50 = "PC1 (age: 1-48)",
`I(signal_for_wing_50^2)` = "PC1 (age: 1-48) 2",
colonySolothurn = "Colony [Solothurn]"
)

sjPlot::tab_model(
  fledg_surv_climmod_lin,
  age_fledg_climmod_lin,
  file = "output/Tab_short_conseq.doc",
  string.est = "Estimate",
  string.se = "SE",
  string.pred = "Predictors",
  show.ci = FALSE,
  show.se = TRUE,
  show.stat = FALSE,
  show.df = FALSE,
  pred.labels = labels
)

```

Figure

```

eff_data <- data.frame(effects::effect("signal_for_wing_50",
  fledg_surv_climmod_lin,
  partial.residuals = TRUE
))
plot_fledg <- ggplot(eff_data, aes(signal_for_wing_50, fit)) +
  geom_ribbon(aes(ymin = lower, ymax = upper), alpha = .25) +
  geom_line(linewidth = 1) +
  theme(
    legend.position = "none",
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    panel.background = element_blank(),
    axis.line = element_line(colour = "black")
  ) +
  xlab("") +
  geom_jitter(
    data = am_nest_clim_wing,
    aes(
      x = signal_for_wing_50,
      y = prop
    ),
    height = 0.03,
    size = 1,
    shape = 19,
    alpha = 0.2
  ) +
  ylab("Nestling survival up to fledging") +
  scale_x_continuous(limits = c(-0.4, 1), breaks = seq(-0.4, 1, 0.2)) +
  scale_y_continuous(limits = c(0, 1.03), breaks = seq(0, 1, 0.2)) +
  annotate("text", x = -0.4, y = 1, label = "(a)")

```

```

eff_data <- data.frame(effects::effect("signal_for_wing_50",
  age_fledg_climmod_quad,
  partial.residuals = TRUE
))
plot_age_fl <- ggplot(eff_data, aes(signal_for_wing_50, fit)) +
  geom_ribbon(aes(ymin = lower, ymax = upper), alpha = .25) +
  geom_line(linewidth = 1) +
  theme(
    legend.position = "none",
    panel.grid.major = element_blank(),
    panel.grid.minor = element_blank(),
    panel.background = element_blank(),
    axis.line = element_line(colour = "black"),
    axis.title.x = element_markdown()
  ) +
  xlab("Weather conditions (PC1<sub>1-48d</sub><br>between 1 and 48 days of age") +
  geom_point(
    data = am_age_fledging_wing,
    aes(
      x = signal_for_wing_50,
      y = age_fledging
    ),
    size = 1,
    shape = 19,
    alpha = 0.2
  ) +
  ylab("Age at fledging (days)") +
  scale_y_continuous(limits = c(50, 76), breaks = seq(45, 90, 5)) +
  scale_x_continuous(limits = c(-0.4, 1), breaks = seq(-0.4, 1, 0.2)) +
  annotate("text", x = -0.4, y = 75, label = "(b)")

cowplot::plot_grid(plot_fledg, plot_age_fl,
  nrow = 2, align = "v"
)

```

```

# Save the plot to the figures directory
ggsave("figures/Fig. 5.png", width = 4, height = 5, dpi = 300)

```

Session information

On top of using renv, the R session information is printed here for reproducibility.

```
sessionInfo()
```

```

## R version 4.3.3 (2024-02-29)
## Platform: aarch64-apple-darwin20 (64-bit)
## Running under: macOS Sonoma 14.4.1
##
## Matrix products: default
## BLAS: /System/Library/Frameworks/Accelerate.framework/Versions/A/Frameworks/vecLib.framework/Versi

```

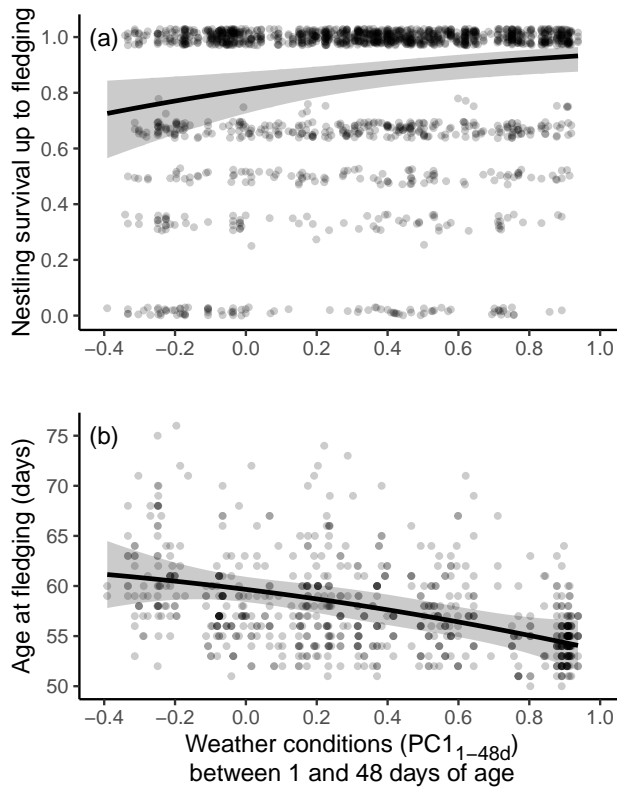


Figure 6: Fig. 5 - Variation in (a) the nestling survival (calculated as the weighted proportion of hatchlings from a nest that successfully fledge) and (b) age at fledging (in days) in relation to weather conditions encountered during development in nestling Alpine swifts. Weather conditions are the resultant of a principal component analysis between daily average temperature and daily rain in the window that best explains the variation in wing length at 50 days. Predicted lines are obtained using average values of the other continuous variables. Climatic windows are reported using the nestling's age as a reference.

```

## LAPACK: /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRlapack.dylib; LAPACK v
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## time zone: Europe/Zurich
## tzcode source: internal
##
## attached base packages:
## [1] stats      graphics  grDevices datasets  utils      methods    base
##
## other attached packages:
## [1] viridis_0.6.5      viridisLite_0.4.2  ggeffects_1.5.2    visreg_2.7.0
## [5] emmeans_1.10.1    tidyr_1.3.1        performance_0.11.0 sjPlot_2.8.15
## [9] scales_1.3.0      ggtext_0.1.2       MuMIn_1.47.5       segmented_2.0-3
## [13] nlme_3.1-164      MASS_7.3-60.0.1    climwin_1.2.3      gridExtra_2.3
## [17] ggplot2_3.5.0     lmerTest_3.1-3     lme4_1.1-35.2      Matrix_1.6-5
## [21] lubridate_1.9.3   dplyr_1.1.4
##
## loaded via a namespace (and not attached):
## [1] DBI_1.2.2          rlang_1.1.3        magrittr_2.0.3
## [4] compiler_4.3.3    mgcv_1.9-1         vctr_0.6.5
## [7] quadprog_1.5-8    stringr_1.5.1      pkgconfig_2.0.3
## [10] crayon_1.5.2      fastmap_1.1.1     backports_1.4.1
## [13] labeling_0.4.3    utf8_1.2.4         rmarkdown_2.26
## [16] markdown_1.12     nloptr_2.0.3      tinytex_0.50
## [19] purrr_1.0.2       xfun_0.43          jsonlite_1.8.8
## [22] highr_0.10        reshape_0.8.9     sjmisc_2.8.9
## [25] broom_1.0.5       R6_2.5.1          stringi_1.8.3
## [28] car_3.1-2         boot_1.3-30       numDeriv_2016.8-1.1
## [31] estimability_1.5  Rcpp_1.0.12       RcppRoll_0.3.0
## [34] knitr_1.46        modelr_0.1.11     DHARMA_0.4.6
## [37] zoo_1.8-12        splines_4.3.3     nnet_7.3-19
## [40] timechange_0.3.0 tidyselect_1.2.1  effects_4.2-2
## [43] abind_1.4-5       yaml_2.3.8        codetools_0.2-20
## [46] sjlabelled_1.2.0 lattice_0.22-6    tibble_3.2.1
## [49] plyr_1.8.9        withr_3.0.0       bayestestR_0.13.2
## [52] evaluate_0.23     PerformanceAnalytics_2.0.4 survival_3.5-8
## [55] survey_4.4-2      xts_0.13.2        xml2_1.3.6
## [58] pillar_1.9.0     carData_3.0-5     renv_1.0.7
## [61] stats4_4.3.3     insight_0.19.10   generics_0.1.3
## [64] munsell_0.5.1    commonmark_1.9.1  minqa_1.2.6
## [67] xtable_1.8-4     glue_1.7.0        tools_4.3.3
## [70] mvtnorm_1.2-4    cowplot_1.1.3     grid_4.3.3
## [73] mitools_2.4      colorspace_2.1-0  cli_3.6.2
## [76] evd_2.3-6.1     fansi_1.0.6       sjstats_0.18.2
## [79] gtable_0.3.4     digest_0.6.35     farver_2.1.1
## [82] htmltools_0.5.8.1 lifecycle_1.0.4   gridtext_0.1.5

```

References for the uploaded packages

```

packages <- c()
for (i in 1:14) {
  packages[i] <- sessionInfo()$otherPkgs[[i]]$Package
}

```

```
print(citation(packages[i]))
}
```

```
## To cite viridis/viridisLite in publications use:
```

```
##
```

```
## Simon Garnier, Noam Ross, Robert Rudis, Antônio P. Camargo, Marco Sciaini, and  
## Cédric Scherer (2024). viridis(Lite) - Colorblind-Friendly Color Maps for R. viridis  
## package version 0.6.5.
```

```
##
```

```
## A BibTeX entry for LaTeX users is
```

```
##
```

```
## @Manual{,
```

```
## title = {{viridis(Lite)} - Colorblind-Friendly Color Maps for R},
```

```
## author = {{Garnier} and {Simon} and {Ross} and {Noam} and {Rudis} and {Robert} and {Camargo} and
```

```
## year = {2024},
```

```
## note = {viridis package version 0.6.5},
```

```
## url = {https://sjmgarnier.github.io/viridis/},
```

```
## doi = {10.5281/zenodo.4679423},
```

```
## }
```

```
## To cite viridis/viridisLite in publications use:
```

```
##
```

```
## Simon Garnier, Noam Ross, Robert Rudis, Antônio P. Camargo, Marco Sciaini, and  
## Cédric Scherer (2023). viridis(Lite) - Colorblind-Friendly Color Maps for R.
```

```
## viridisLite package version 0.4.2.
```

```
##
```

```
## A BibTeX entry for LaTeX users is
```

```
##
```

```
## @Manual{,
```

```
## title = {{viridis(Lite)} - Colorblind-Friendly Color Maps for R},
```

```
## author = {{Garnier} and {Simon} and {Ross} and {Noam} and {Rudis} and {Robert} and {Camargo} and
```

```
## year = {2023},
```

```
## note = {viridisLite package version 0.4.2},
```

```
## url = {https://sjmgarnier.github.io/viridis/},
```

```
## doi = {10.5281/zenodo.4678327},
```

```
## }
```

```
## To cite package 'ggeffects' in publications use:
```

```
##
```

```
## Lüdecke D (2018). "ggeffects: Tidy Data Frames of Marginal Effects from Regression  
## Models." _Journal of Open Source Software_, *3*(26), 772. doi:10.21105/joss.00772
```

```
## <https://doi.org/10.21105/joss.00772>.
```

```
##
```

```
## A BibTeX entry for LaTeX users is
```

```
##
```

```
## @Article{,
```

```
## title = {ggeffects: Tidy Data Frames of Marginal Effects from Regression Models.},
```

```
## volume = {3},
```

```
## doi = {10.21105/joss.00772},
```

```
## number = {26},
```

```
## journal = {Journal of Open Source Software},
```

```
## author = {Daniel Lüdecke},
```

```
## year = {2018},
```

```
## pages = {772},
```

```
## }
```

```

## To cite visreg in publications use:
##
##   Breheny P and Burchett W (2017). Visualization of Regression Models Using visreg.
##   The R Journal, 9: 56-71.
##
## A BibTeX entry for LaTeX users is
##
##   @Article{,
##     author = {Patrick Breheny and Woodrow Burchett},
##     title = {Visualization of Regression Models Using visreg},
##     journal = {The R Journal},
##     year = {2017},
##     volume = {9},
##     pages = {56--71},
##     number = {2},
##   }
## To cite package 'emmeans' in publications use:
##
##   Lenth R (2024). _emmeans: Estimated Marginal Means, aka Least-Squares Means_. R
##   package version 1.10.1, <https://CRAN.R-project.org/package=emmeans>.
##
## A BibTeX entry for LaTeX users is
##
##   @Manual{,
##     title = {emmeans: Estimated Marginal Means, aka Least-Squares Means},
##     author = {Russell V. Lenth},
##     year = {2024},
##     note = {R package version 1.10.1},
##     url = {https://CRAN.R-project.org/package=emmeans},
##   }
## To cite package 'tidyr' in publications use:
##
##   Wickham H, Vaughan D, Girlich M (2024). _tidyr: Tidy Messy Data_. R package version
##   1.3.1, <https://CRAN.R-project.org/package=tidyr>.
##
## A BibTeX entry for LaTeX users is
##
##   @Manual{,
##     title = {tidyr: Tidy Messy Data},
##     author = {Hadley Wickham and Davis Vaughan and Maximilian Girlich},
##     year = {2024},
##     note = {R package version 1.3.1},
##     url = {https://CRAN.R-project.org/package=tidyr},
##   }
## To cite package 'performance' in publications use:
##
##   Lüdtke et al., (2021). performance: An R Package for Assessment, Comparison and
##   Testing of Statistical Models. Journal of Open Source Software, 6(60), 3139.
##   https://doi.org/10.21105/joss.03139
##
## A BibTeX entry for LaTeX users is
##
##   @Article{,
##     title = {{performance}: An {R} Package for Assessment, Comparison and Testing of Statistical Models}

```



```

##   author = {Daniel Lüdecke and Mattan S. Ben-Shachar and Indrajeet Patil and Philip Waggoner and D
##   year = {2021},
##   journal = {Journal of Open Source Software},
##   volume = {6},
##   number = {60},
##   pages = {3139},
##   doi = {10.21105/joss.03139},
## }
## To cite package 'sjPlot' in publications use:
##
##   Lüdecke D (2023). _sjPlot: Data Visualization for Statistics in Social Science_. R
##   package version 2.8.15, <https://CRAN.R-project.org/package=sjPlot>.
##
## A BibTeX entry for LaTeX users is
##
##   @Manual{,
##     title = {sjPlot: Data Visualization for Statistics in Social Science},
##     author = {Daniel Lüdecke},
##     year = {2023},
##     note = {R package version 2.8.15},
##     url = {https://CRAN.R-project.org/package=sjPlot},
##   }
## To cite package 'scales' in publications use:
##
##   Wickham H, Pedersen T, Seidel D (2023). _scales: Scale Functions for Visualization_.
##   R package version 1.3.0, <https://CRAN.R-project.org/package=scales>.
##
## A BibTeX entry for LaTeX users is
##
##   @Manual{,
##     title = {scales: Scale Functions for Visualization},
##     author = {Hadley Wickham and Thomas Lin Pedersen and Dana Seidel},
##     year = {2023},
##     note = {R package version 1.3.0},
##     url = {https://CRAN.R-project.org/package=scales},
##   }
## To cite package 'ggtext' in publications use:
##
##   Wilke C, Wiernik B (2022). _ggtext: Improved Text Rendering Support for 'ggplot2'_.
##   R package version 0.1.2, <https://CRAN.R-project.org/package=ggtext>.
##
## A BibTeX entry for LaTeX users is
##
##   @Manual{,
##     title = {ggtext: Improved Text Rendering Support for 'ggplot2'},
##     author = {Claus O. Wilke and Brenton M. Wiernik},
##     year = {2022},
##     note = {R package version 0.1.2},
##     url = {https://CRAN.R-project.org/package=ggtext},
##   }
## To cite package 'MuMIn' in publications use:
##
##   Bartoń K (2023). _MuMIn: Multi-Model Inference_. R package version 1.47.5,
##   <https://CRAN.R-project.org/package=MumIn>.

```

```

##
## A BibTeX entry for LaTeX users is
##
## @Manual{,
##   title = {MuMIn: Multi-Model Inference},
##   author = {Kamil Bartoń},
##   year = {2023},
##   note = {R package version 1.47.5},
##   url = {https://CRAN.R-project.org/package=MuMIn},
## }
##
## ATTENTION: This citation information has been auto-generated from the package
## DESCRIPTION file and may need manual editing, see 'help("citation")'.
## To cite segmented in publications use one or more of the following papers:
##
## Fasola S, Muggeo VMR, Kuchenhoff K. (2018). A heuristic, iterative algorithm for
## change-point detection in abrupt change models. Computational Statistics, 33,
## 997-1015.
##
## Muggeo VMR (2003). Estimating regression models with unknown break-points.
## Statistics in Medicine, 22, 3055-3071.
##
## Muggeo VMR (2008). segmented: an R Package to Fit Regression Models with Broken-Line
## Relationships. R News, 8/1, 20-25. URL https://cran.r-project.org/doc/Rnews/.
##
## Muggeo VMR (2016). Testing with a nuisance parameter present only under the
## alternative: a score-based approach with application to segmented modelling. J of
## Statistical Computation and Simulation, 86, 3059-3067.
##
## Muggeo VMR (2017). Interval estimation for the breakpoint in segmented regression: a
## smoothed score-based approach. Australian & New Zealand Journal of Statistics, 59,
## 311-322.
##
## Muggeo VMR, Atkinhs DC, Gallopp RJ, Dimidjian S (2014). Segmented mixed models with
## random changepoints: a maximum likelihood approach with application to treatment for
## depression study Statistical Modelling, 14, 293-313.
##
## To see these entries in BibTeX format, use 'print(<citation>, bibtex=TRUE)',
## 'toBibtex(.)', or set 'options(citation.bibtex.max=999)'.
## To cite package 'nlme' in publications use:
##
## Pinheiro J, Bates D, R Core Team (2023). _nlme: Linear and Nonlinear Mixed Effects
## Models_. R package version 3.1-164, <https://CRAN.R-project.org/package=nlme>.
##
## Pinheiro JC, Bates DM (2000). _Mixed-Effects Models in S and S-PLUS_. Springer, New
## York. doi:10.1007/b98882 <https://doi.org/10.1007/b98882>.
##
## To see these entries in BibTeX format, use 'print(<citation>, bibtex=TRUE)',
## 'toBibtex(.)', or set 'options(citation.bibtex.max=999)'.
## To cite the MASS package in publications use:
##
## Venables, W. N. & Ripley, B. D. (2002) Modern Applied Statistics with S. Fourth
## Edition. Springer, New York. ISBN 0-387-95457-0
##

```

```
## A BibTeX entry for LaTeX users is
##
## @Book{,
##   title = {Modern Applied Statistics with S},
##   author = {W. N. Venables and B. D. Ripley},
##   publisher = {Springer},
##   edition = {Fourth},
##   address = {New York},
##   year = {2002},
##   note = {ISBN 0-387-95457-0},
##   url = {https://www.stats.ox.ac.uk/pub/MASS4/},
## }
```