SUPPLEMENTAL TABLE OF CONTENTS

Supplemental Method:

Gating strategy for flow Cytometry to confirm the expression of MSC-specific surface markers.

Using the aspect ratio intensity and area of the brightfield channel on the Amnis Flow sight we were able to visualize and gate the single cells. Thresholds on the Amnis flow sight were determined by histograms generated by single color stained positive and negative beads. Using this data, baseline thresholds were determined to run our samples. The threshold generated by the control beads was used to visualize the images of cells in a scatterplot and determine the single cells that were positively stained¹.

Supplemental Figures

Supplemental Figure 1: Mesenchymal stem cell (MSC) showing typical morphological appearance in normoxic and hypoxic conditions under the microscope as spindle-shaped, fibroblast-like cells in culture. **(A, B respectively)**

Supplemental Figure 2: Mesenchymal stem cell (MSC) characterization in normoxia conditions. **(A)** Positive MSC markers (CD73, CD90 and CD105) and **(B)** negative MSC markers (CD14, CD34, CD45).

Supplemental Figure 3: Mesenchymal stem cell (MSC) characterization in hypoxia conditions. **(A)** Positive MSC markers (CD73, CD90 and CD105) and **(B)** negative MSC markers (CD14, CD34, CD45).

Supplemental Figure 4: MSC characterization by with fluorescently conjugated antibodies against CD73, CD90, CD105. (A) . Cells strongly positive for markers CD73, CD90 and CD105.

Supplemental Figure 5: MSC Morphology and differentiation. Tri-lineage differentiation into adipocyte (FABP4), chondrocyte (Aggrecan) and osteocytes (osteocalcin) and their quantification.

Supplemental Figure 6: Normoxia HTN vs Normoxia Healthy Control MSCs. Normoxic-MSC of hypertension vs. healthy control mapped a total of 13,469 genes, with 463 significant dysregulated genes (n=306 upregulated & n=157 downregulated). Gene ontology analysis showed that upregulated genes were implicated in modulation of vasculature development, migration, response to growth factor and negative regulation of proliferation, whereas downregulated genes participated in insulin like growth factor binding proteins (A, B, respectively). Volcano plot demonstrated the distribution of differentially expressed genes, with downregulated and upregulated genes based on p-value and log₂fc (C). Heatmap showed angiogenic and inflammatory genes upregulated in HTN MSCs compared to HC MSCs and an equal dysregulation of senescence genes in MSCs of both the groups (D).

Supplemental Figure 7: Normoxia Healthy Control vs Hypoxia Healthy Control MSCs. In healthy control-MSCs, a total of 13138 genes were mapped, with 261 significant dysregulated genes (n=60 upregulated & n=201 downregulated). Gene ontology analysis showed that gene upregulated participated in modulation of angiogenesis and methylation, whereas downregulated genes were involved in cellular response to tumor necrosis factor (A, B, respectively). Volcano plot demonstrated the distribution of differentially expressed genes, with downregulated and upregulated genes based on p-value and log₂fc (C). Heatmap showed dysregulated genes significant for angiogenesis, and inflammation upregulated more in hypoxic HC MSCs compared to normoxic HC MSCs (D).

Supplemental Figure 8: Gene ontology (GO) analysis pathways (A) Biological Process (B) Molecular Functions (C) Cellular Components of Upregulated gene sets with significant changes between Hypoxia HKD and Normoxia HKD MSCs.

Supplemental Figure 9: Genes and Transcription Factor (TF) network Interactions between Normoxia HKD and Normoxia HC mRNA targets derived from STRING. Color lines represent interactions between Normoxia HKD and Normoxia HC mRNA targets according to the functional association networks. Red circles indicate common (TF) mRNA targets between Normoxia HKD MSCs and Normoxia HC MSCs.

Known Interactions

Node Color

- Colored nodes: query proteins and first shell of interactors
- White nodes: second shell of interactors

Node Content

- Empty nodes: proteins of unknown 3D structure
- Filled nodes: a 3D structure is known or predicted
- From curated databases
 Experimentally determined
 Predicted Interactions
 Gene neighborhood
 Gene fusions
 Gene co-occurrence
 Textmining
 Co-expression
 Protein homology

Supplemental Figure 10: Genes and Transcription Factor (TF) network Interactions between Normoxia HKD and Normoxia HTN mRNA targets derived from STRING. Color lines represent interactions between Normoxia HKD and Normoxia HTN mRNA targets according to the functional association networks. Red circles indicate common (TF) mRNA targets between Normoxia HKD MSCs and Normoxia HTN MSCs.

Node Color Colored nodes: query proteins and first shell of interactors White nodes: second shell of

interactors

Node Content

- Empty nodes: proteins of unknown 3D structure
- Filled nodes: a 3D structure is known or predicted

Known Interactions From curated databases Experimentally determined Predicted Interactions Gene neighborhood Gene fusions Gene co-occurrence Textmining Co-expression Protein homology

Supplemental Figure 11: Genes and Transcription Factor (TF) network Interactions between HKD normoxia and hypoxia mRNA targets derived from STRING. Color lines represent interactions between HKD normoxia and hypoxia mRNA targets according to the functional association networks. Red circles indicate common (TF) mRNA targets between HKD normoxia and hypoxia.



Supplemental Figure S12: Inflammatory markers

Inflammatory factors under normoxic and hypoxic state in IFN- γ , IL-1 α , IL-6, IL-8, and TNF- α in HC(n=12)-, HTN(n=9)- MSCs, and HKD(n=12)- MSCs.At baseline normoxic conditions, there was no difference in inflammatory cytokines released among the groups (Fig S12A-E). However, under HPC, there was an increase of IFN- γ levels only in HC-MSCs (p-value=0.04) (Fig S12-A). Among the baseline of the HC, HTN and HKD, there was no difference (Fig S13).

Supplemental Table S1: Angiogenesis secretome markers

Under normoxic conditions the expression of the pro-angiogenic factors VEGF and EGF were similar among the groups and HPC did not exert any significant effect in these specific angiogenic markers. There was a significant increase in HC MSCs EGF and HGF (Elisa)after hypoxia. We also saw a significant decrease in HKD MSCs HGF after hypoxia.

Supplemental Table S2: Senescence markers

Under normoxic conditions the expression of P16, P21 and SA β GAL was nonsignificant but there was a trend decrease in HC MSCs SA β GAL activity after treatment with hypoxia.

Supplemental Table S3: PCR markers

At baseline normoxic conditions, there was no difference in TGF - β ,PGE, iONS,

GAPDH, IDO,IL10, IL4 among the groups.

Supplemental Table S4A:

Correlation between MSC Function (normoxia) and Age, MSC Function (hypoxia) and Age

At the baseline and in hypoxic conditions, there was no correlation found between the function of MSCs and the age of the patients.

NORMOXIA





Low Resolution 4x objective

ΗΥΡΟΧΙΑ



Low Resolution 4x objective

Fig. S1. Spindle-shaped morphology of cells isolated from adipose tissue of humans, characteristic of mesenchymal stem cells.

- A. AMSC grown to confluence in Normoxia (20% O₂)
- B. Cells grown to confluence and subjected to 48 hours of HPC (1% O₂)

AMSC, Adipose tissue derived mesenchymal stem cells; HPC, Hypoxia preconditioning, O₂, Oxygen



| Population | Count | %Gated |
|---------------|-------|--------|
| Single | 9729 | 100 |
| CD14 & Single | 1712 | 17.6 |



| Population | Count | %Gated |
|---------------|-------|--------|
| Single | 10200 | 100 |
| CD90 & Single | 10180 | 99.8 |



| Intensity_MC_Ch | | |
|-----------------|-------|--------|
| Population | Count | %Gated |
| Single | 9729 | 100 |
| CD45+ & Single | 12 | 0.12 |



| Population | Count | %Gated |
|----------------|-------|--------|
| Single | 10200 | 100 |
| CD105 & Single | 10007 | 98.1 |



| Population | Count | %Gated |
|---------------|-------|--------|
| Single | 37798 | 100 |
| CD34 & Single | 111 | 0.29 |

Fig. S2. Flowcytometry characterizing the cell surface markers confirming that isolated cells were adipose tissue derived mesenchymal stem cells in Normoxia conditions.

- A. Cells strongly positive for markers CD73, CD90 and CD105
- B. Cells negative for markers CD14, CD34 and CD45



| Population | Count | %Gated | |
|----------------|-------|--------|--|
| Single | 9878 | 100 | |
| CD73+ & Single | 8963 | 90.7 | |



1e5



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Frequ

| Intensity_MC_C | ch06 | |
|----------------|-------|--------|
| Population | Count | %Gated |
| Single | 9463 | 100 |
| CD14 & Single | 1126 | 11.9 |



2.5

CD90

1e3 1e4 1e5

Intensity_MC_Ch02

9878

9859

Count %Gated

100

99.8

1e

nalized Frequency

N 0.5

0 -

Population

CD90 & Single

Single

-100 0 100

Intensity_MC_Ch02



| Intensity_MC_Ch03 | | | |
|-------------------|-------|--------|--|
| Population | Count | %Gated | |
| Single | 9878 | 100 | |
| CD105 & Single | 9610 | 97.3 | |



| | Inten | sity | CD |)34 |
|--|-------|------|----|-----|
|--|-------|------|----|-----|

| Population | Count | %Gated |
|---------------|-------|--------|
| Single | 28559 | 100 |
| CD34 & Single | 60 | 0.21 |

Fig. S3. Flowcytometry characterizing the cell surface markers confirming that isolated cells were adipose tissue derived mesenchymal stem cells in Hypoxia conditions.

- A. Cells strongly positive for markers CD73, CD90 and CD105
- B. Cells negative for markers CD14, CD34 and CD45

Α.



Fig. S4. A. Adipose tissue derived mesenchymal stem cells were stained with fluorescently conjugated antibodies against CD73, CD90, CD105 and the expression was analyzed by flow cytometry. The histogram in pink represents the isotype control and the histogram in orange, green, puple respectively represents the stained sample. Cells strongly positive for markers CD73, CD90 and CD105

Α.



Fig. S5. Differentiation of cells grown after isolation from adipose tissue into adipocytes (A), osteocytes (B), chondrocytes (C), and quantification of adipocytes, osteocytes, and chondrocytes (D) quantification confirming their mesenchymal origin and regenerative properties. Images captured after fluorescent staining of differentiated cells.

Figure: S6

Upregulated genes between HTN and HC (Normoxia)



GO:0071396: cellular response to lipid R-HSA-9031628: NGF-stimulated transcription GO:0001944: vasculature development GO:0042326: negative regulation of phosphorylation GO:0071345: cellular response to cytokine stimulus GO:0043408: regulation of MAPK cascade GO:0045596: negative regulation of cell differentiation GO:0009725: response to hormone hsa05200: Pathways in cancer GO:0090287: regulation of cellular response to growth factor stimulus GO:0008285: negative regulation of cell population proliferation GO:0002683: negative regulation of immune system process GO:0071363: cellular response to growth factor stimulus GO:0010720: positive regulation of cell development R-HSA-6785807: Interleukin-4 and Interleukin-13 signaling GO:0045598: regulation of fat cell differentiation GO:0032680: regulation of tumor necrosis factor production M166: PID ATF2 PATHWAY GO:0030335: positive regulation of cell migration GO:0048568: embryonic organ development

Β.

Α.



Downregulated genes between HTN and HC (Normoxia)

WP534: Glycolysis and gluconeogenesis hsa00970: AminoactRNA biosynthesis M3008: NABA ECM GLYCOPROTEINS GO:0007411L axon guidance R-HAS_381426: Regulation of IGF transport and uptake by Insulin like Growth factor Binding Proteins (IGFBPs) GO:0061061: muscle structure development GO:0001558: regulation of cell growth



ID2

Low

High



Α.





GOMF_MOLECULAR_FUNCTION_ACTIVATOR_ACTIVITY GOMF_HYDROLASE_ACTIVITY_ACTING_ON_ESTER_BONDS GOMF_SIGNALING_RECEPTOR_REGULATOR_ACTIVITY GOMF_DOUBLE_STRANDED_RNA_BINDING GOMF_DOUBLE_STRANDED_RNA_BINDING GOMF_PHOSPHOLIPASE_ACTIVITY GOMF_PHOSPHOLIPASE_A2_ACTIVITY GOMF_PHOSPHOLIPASE_A2_ACTIVITY GOMF_PHOSPHOLIPASE_A1_ACTIVITY GOMF_CXCR_CHEMOKINE_RECEPTOR_BINDING GOMF_PROTEINASE_ACTIVATED_RECEPTOR_ACTIVITY

Fig.S8. Hypoxia HKD vs Normoxia HKD MSCs. Gene ontology (GO) analysis pathways (A) Biological Process (B) Molecular Functions (C) Cellular Components of Upregulated gene sets with significant changes between Hypoxia HKD and Normoxia HKD MSCs.



Figure: S9 Gene and transcription factors network interactions between HKD and HC (Normoxia)



Figure: S10 Gene and transcription factors network interactions between HKD and HTN (Normoxia)



Figure: S11 Gene and transcription factors network interactions between HKD Normoxia and HKD Hypoxia



| Table S1- Angiogenesis secretome markers at baseline normoxia | а |
|---|---|
|---|---|

| VARIABLES | HC (12) | HTN (9) | HKD (12) | P value |
|----------------------|--|--|---|---------|
| EGF | 7.94108e ⁻⁹ (5.97554e ⁻⁹ - 1.00493e ⁻⁸) | 4.98339e ⁻⁹ (3.44828e ⁻⁹ - 1.5619e ⁻⁸) | 7.19277e ⁻⁹ (5.9043e ⁻⁹ - 1.10211e ⁻⁸) | NS |
| HGF | 7.0609e ⁻⁸ (3.90828e ⁻⁸ - 2.02851e ⁻⁷) | 7.03846e- ⁸ (3.43478e ⁻⁸ - 2.77636e ⁻⁷) | 3.7796e - ⁸ (2.8515e ⁻⁸ -2.111e ⁻⁷)# | NS |
| VEGF | 0.00018 (5.88315e ⁻⁵ - 0.000262995) | 0.000338342 (8.01248e ⁻ ⁵ - 0.001057009) | 0.000119132 (4.22891e ⁻⁵ - 0.000373018) | NS |
| VEGF (at gene level) | 0.977101 (0.55706725- 1.3536075) | 1.07 (0.64-1.435) | 1.21 (0.79-1.46) | NS |
| EGF (at gene level) | 0.97 (0.705- 2.1325) | 3.06 (0.24- 5.68) | 1.344779401 (0.425334961 - 3.110220509) | NS |

Abbreviations: HC: healthy control; HTN: hypertension; HKD: Hypertensive kidney disease

| VARIABLES | HC (12) | HTN (9) | HKD (12) | P value |
|-----------|-----------------------------------|-----------------------------------|----------------------------------|---------|
| P16 | 1.063 (0.430- 1.696) | 1.160 (0.570- 1.480) | 0.866 (0.472- 2.445) | NS |
| P21 | 0.810 (0.623- 1.190) | 0.870 (0.610- 1.490) | 0.895 (0.640- 1.113) | NS |
| SA β GAL | 15765.5 (10393.500- 29550.500) | 35322.5 (25168.500- 52886.000) | 14956.5 (6672.875- 19085.000) | *0.04 |

Abbreviations: HC: healthy control; HTN: hypertension; HKD: Hypertensive kidney disease

Table S3 – PCR Markers at baseline normoxia

| VARIABLES | HC (12) | HTN (9) | HKD (12) | P value |
|-----------|-------------------------|-------------------------|-------------------------|---------|
| TGF -β | 24.214 (23.488-25.148) | 24.800 (24.441, 25.324) | 25.182 (24.504, 25.484) | NS |
| PGE | 26.827 (26.414- 27.859) | 27.621 (26.550- 28.666) | 27.020 (26.817- 27.341) | NS |
| iNOS | 36.454 (35.355- 37.295) | 38.396 (37.897- 39.677) | 36.625 (36.351- 37.391) | NS |
| IDO | 36.603 (34.912- 39.251) | 38.831 (36.902- 39.447) | 37.380 (35.873- 38.867) | NS |
| IL 10 | 38.702 (35.725- 39.716) | 35.699 (34.445- 39.221) | 36.374 (34.541- 38.427) | NS |
| IL 4 | 39.178 (37.917- 40.000) | 39.067 (37.522- 40.000) | 38.511 (36.663- 40.000) | NS |
| GAPDH | 20.156 (19.052- 21.981) | 21.375 (20.167- 22.746) | 20.847 (20.310- 21.231) | NS |

Abbreviations: HC: healthy control; HTN: hypertension; HKD: Hypertensive kidney disease; NS: non-significant

| Table S4- Correlation between age and MSC function in hypoxic and normoxic condit |
|---|
|---|

| | Age | |
|---------------|------|------|
| MSC function | r | Р |
| NX | 0.16 | 0.36 |
| Proliferation | | |
| НХ | 0.17 | 0.33 |
| Proliferation | | |

Abbreviations: MSC: mesenchymal stem cell; NX: normoxia; HX: hypoxic

REFERENCE

[1] Isik B, Thaler R, Goksu BB, Conley SM, Al-Khafaji H, Mohan A, Afarideh M, Abumoawad AM, Zhu XY, Krier JD, Saadiq IM, Tang H, Eirin A, Hickson LJ, van Wijnen AJ, Textor SC, Lerman LO, Herrmann SM: Hypoxic preconditioning induces epigenetic changes and modifies swine mesenchymal stem cell angiogenesis and senescence in experimental atherosclerotic renal artery stenosis. Stem Cell Res Ther 2021, 12:240.