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Supplemental information

Transcriptional synergy in human aortic

endothelial cells is vulnerable to combination

p300/CBP and BET bromodomain inhibition

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Figure S1



Figure S1 (related to Figure 1): IFN γ /TNF α costimulation of HAECs induces nascent RNA production and synergistic expression of proinflammatory chemokine genes

(A) Schematic of experimental design for MIMs experiments.

(B) MIMS images of unstimulated HAECs, HAECs stimulated with IFN_Y (50ng/mL), TNF α (25ng/mL) or IFN_Y/TNF α for one hour (left) or four hours (right). At the bottom is shown MIMS images of cells with no isotope. Quantitative ³²S images reveal cellular and nuclear contours (left column). Hue saturation intensity images are used to visually represent isotope ratio measurements and in turn map ¹³C-thymidine labeling of DNA (¹³C/¹²C, middle) and ¹⁵N-uridine labeling of nascent RNA (¹⁵N/¹⁴N, right). The lower bound of the color scale (blue) is set to natural background (¹³C=1.1%; ¹⁵N=0.37%) and the upper bound of the scale is set to reveal differential labeling and expressed as % above natural background.

(C) Volcano plot of log₂(FC) vs. log₁₀(Adjusted p-value) for differential genes in IFN γ stimulated vs. Unstimulated HAECs.

(D) Volcano plot of log₂(FC) vs. log₁₀(Adjusted p-value) for differential genes in TNF α stimulated vs. Unstimulated HAECs.

(E) Volcano plot of log₂(FC) vs. log₁₀(Adjusted p-value) for differential genes in IFN γ /TNF α stimulated vs. Unstimulated HAECs.



Figure S2 (related to Figure 2): Transcriptional synergy elicited by IFN γ /TNF α signaling requires STAT1 and p65

(A) Representative photomicrographs of STAT1 (green) and p65 (red) immunofluorescence in TeloHAECs treated with vehicle or cytokines for 1 hour or 4 hours. Nuclei are indicated by DAPI staining (blue color). All samples were imaged on a Zeiss LSM880 Confocal Microscope with 10X objective. Scale bars indicate 100 μ m.

(B-E) Box and whisker plots of mean fluorescence intensity for nuclei measured in (A). 1 hour STAT and p65 in (B-C) and 4 hour STAT and p65 in (D-E). Kruskal-Wallis test was performed followed by a pairwise Wilcoxon test to determine statistical differences between treatment groups. ***p-value < 0.001; **** p-value < 0.001.

(F-I) Bar graphs of mRNA expression of *CXCL9 (F), CXCL10 (G), CXCL11 (H) or SELE (I)* in cells after cytokine stimulation (1 hour) followed by cytokine washout for 3 hours. mRNA shown as fold change compared to unstimulated baseline. One-way ANOVA was performed followed by Tukey's post-hoc analysis to determine statistical differences between treatment groups (n.s. = not significant). * p-value < 0.05; ** p-value < 0.01 *** p-value < 0.001; **** p-value < 0.0001.

(*J*) mRNA expression in HAECs treated with or without Cycloheximide (100 μ g/mL) and IFN γ /TNF α (1 hour) versus no cytokine (n=3 per condition). One-way ANOVA was performed followed by Tukey's post-hoc analysis to determine statistical differences between treatment groups (n.s. = not significant).





Figure S3 (related to Figure 3): Synergistic induction of proinflammatory genes is not dependent on saturation of IFN γ or TNF α signaling

(A) Heatmaps of *IL32*, *GUSB*, *NOL7*, *HPRT1*, *CXCL11*, *CXCL10*, *VCAM1*, *CXCL9*, *SOCS3*, and *CX3CL1* expression in response to 12 -oint dose response curves for IFN γ (left) or TNF α (right) (n=1).

(B-J) Bar plots of normalized *CXCL9* (B,E,H), *CXCL10* (C,F,I), and *CXCL11* (D,G,J) transcript counts after stimulation with 50 ng/mL IFN γ + TNF α 0.024 (B-D), 50 ng/mL IFN γ + TNF α 0.39 ng/mL (E-G), and 50 ng/mL IFN γ + TNF α 25 ng/mL (H-J). Bars indicate mean ± SD (n=4 per condition). One-way ANOVAs were performed followed by Tukey's post-hoc analysis to determine statistical differences between treatment groups. ** p ≤0.001 **** p ≤0.001

Figure S4



Figure S4 (related to Figure 5): Dual cytokine stimulation results in the formation of *de novo* and enhanced accessible elements at the *CXCL-9,-10,11* locus

(A-D) Signal alignment heatmaps for individual ATAC-seq samples.

(E-G) Waterfall plots of RPKM for peaks differential in IFN γ vs unstimulated (E), TNF α vs. unstimulated (F) and IFN γ /TNF α vs. unstimulated (G).

(H, I) Box plots of peak length (top) and normalized signal (bottom) in distal elements (H) and gene bodies (I). Kruskal-Wallis tests were performed followed by pairwise Wilcoxon tests to determine statistical significance. * $p \le 0.05$ ** $p \le 0.001$





Figure S5 (related to Figure 5): Dual cytokine stimulation results in the formation of accessible elements and recruitment of STAT1 and p65 at the *CXCL-9,-10*, and *-11* locus

(A) Browser tracks of SELE and SOCS3 in cytokine treated HAECs.

(B) Box plot of ATAC Fold Change (IFN γ +TNF α vs. unstimulated) on y-axis vs. RNA-seq outlier status on x-axis. Wilcoxon test was performed followed to determine statistical differences between groups. *p-value < 0.05

(C) Pie chart of percent of outlier genes (65 total) identified in Figure 1 that have an ATAC SSAE site within specified distance to the transcriptional start site of the gene. If a gene had more than one SSAE occurrence, the SSAE closest to the gene was chosen for purposes of counting the total.

(D) Immunoblots of p65 and STAT1 in single KO cells reconstituted with 3xFLAG-p65 or 3x-FLAG-STAT1. For each blot, β actin served as a loading control.

(E) Bar plots of RT-qPCR showing chemokine gene induction in STAT1 KO cells or FLAGreconstituted p65. Results displayed as percent of maximal induction. Statistical difference tested by one-way ANOVA. n.s. indicates not significant.

(F) Immunoblot of p65 and STAT1 expression in double KO cells after reconstitution with 3xFLAGp65 or 3xFLAG-STAT1.

(G) Stacked Bar plots of differential peaks for TFs for knockout of STAT1 (left) or RELA (right) TF.

(H-J) Gene tracks of SELE (H), SOCS3 (I) or CXCL locus (J) in double KO cells with p65 or STAT1 reconstitution. Black tracks = unstimulated cells Red tracks = (+) IFN γ /TNF α . C&R = CUT&RUN. Arrows indicate sites where p65 binding has been lost in *STAT1* KO.

Figure S6





Figure S6 (related to Figure 6): Synergistically induced genes harbor transcriptional dependencies on p300/CBP and BET bromodomain-containing proteins

(A-B) STRING interaction network for STAT1 (A) and RELA/p65 (B) with 1st and 2nd shells set to 10 interactors. Co-activators and proteins involved in histone modifications are highlighted in red.

(C-E) Bar plots of secreted CXCL9 (D), CXCL10 (E), CXCL11 (F) in HAECs treated with dual cytokines and DMSO, A-485 (2000 nM), or JQ1 (500 nM) for 8 hours (n=3 per condition). Data is reported as mean \pm SD. One-way ANOVA followed by a Dunnett's test was performed to determine statistically significant changes in chemokines. **p < 0.01, ***p < 0.001, **** p < 0.0001

(F) Bar plot of absorbance at 600 nm as an indicator of cell viability as measured by an MTT assay in HAECs treated co-stimulated with IFN γ /TNF α and various concentrations of A-485 or JQ1. Bars indicate mean ± SD (n=3 per condition). A one-way ANOVA followed by Dunnett's test was performed to assess statistical significance. **p < 0.01, ***p < 0.001, **** p < 0.0001.







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Figure S7: (related to Figure 7): p300/CBP or BET bromodomain inhibition does not alter chromatin accessibility or p65 and STAT1 recruitment to chromatin

(A) Heatmap alignment plots of ATAC-seq with individual inhibitors. All samples were treated with dual cytokines.

(B) Venn diagrams of genomic distribution of STAT1 and p65 in CUT&RUN in cytokine treated cells with and without A485 (2 μ M) or JQ1 (500 nM).

(C-D) Gene tracks of ATAC-seq along with STAT1 and p65 CUT&RUN at *SELE* (C) and *SOCS3* (D) loci.

Table S1: (related to Figure 2): Fold-Change Values for WT, p65 KO and STAT1 KO HAECs in Response to IFN γ /TNF α Dual Stimulation

Fold Change By Gene	WT (STAT+/p65+)	p65 KO	STAT KO
CXCL9	2041	101	39
CXCL10	215329	1183	5935
CXCL11	205	31	6.6